

Supporting Information

# Genome Reduction and Secondary Metabolism of the Marine Sponge-Associated Cyanobacterium *Leptothoe*

Despoina Konstantinou <sup>1</sup>, Rafael V. Popin <sup>2</sup>, David P. Fewer <sup>2</sup>, Kaarina Sivonen <sup>2</sup>, and Spyros Gkelis <sup>1,\*</sup>

<sup>1</sup> Department of Botany, School of Biology, Aristotle University of Thessaloniki, Thessaloniki GR-541 24, Greece

<sup>2</sup> Department of Microbiology, University of Helsinki, Viikinkaari 9, FI-00014 Helsinki, Finland

\* Correspondence: sgkelis@bio.auth.gr; Tel.: +30-2310-998083

## Table of Contents

**Table S1.** Number of genes in subsystems (obtained by RAST server and SEED tool) per *Leptothoe* strain.

**Table S2.** Biosynthetic gene clusters per *Leptothoe* genome (antiSMASH results)

**Table S3.** Compounds produced by *Leptothoe* cyanobacteria extracted from the CyanoMetDB (Jones et al., 2020).

**Figure S1.** Circular view of the genomes of two sponge-associated *Leptothoe* species, generated with CGview (Stothard and Wishart, 2005). Circles from interior to exterior represent GC content and GC skew. Blue circles denote the coding sequences on forward and reverse strands.

**Figure S2.** Average amino acid identity heatmap of *Leptothoe* genomes.

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**Table S1.** Number of genes in subsystems (obtained by RAST server and SEED tool) per *Leptothoe* strain.

No. of genes in subsystems	TAU-MAC 1615	TAU-MAC 1115	PCC 7375	Heron Island J	CCMR0081	CCMR0082	SIO3F4
Cofactors, Vitamins, Prosthetic Groups, Pigments	73	68	207	183	199	212	203
Biotin	6	7	12	13	11	11	22
Thiamin biosynthesis	4	5	8	8	8	8	18
Menaquinone and Phylloquinone Biosynthesis	0	8	14	6	14	14	6
Bilin Biosynthesis	8	0	13	9	16	15	12
Cobalamin synthesis	5	0	11	8	11	11	17
Chlorophyll Degradation	1	0	0	1	0	0	1
Heme and Siroheme Biosynthesis	0	8	18	17	18	21	18
Coenzyme B12 biosynthesis	0	0	18	11	15	17	0
Riboflavin, FMN, FAD	7	7	15	15	7	14	19
Pyridoxin (Vitamin B6) Biosynthesis	0	0	7	9	7	8	9
NAD and NADP	11	9	15	15	14	14	19
Folate and pterines	24	15	61	53	55	64	47
Lipoic acid metabolism	2	2	2	2	2	2	2
Coenzyme F420 synthesis	2	0	2	2	2	2	2
Coenzyme A Biosynthesis	3	5	11	14	12	11	11
Cell Wall and Capsule	24	31	32	38	43	47	63
Capsular and extracellular polysaccharides	14	23	11	20	25	25	19
Gram-Negative cell wall components	5	0	0	0	0	0	20
Recycling of Peptidoglycan Amino Acids	4	2	6	5	7	7	4
Murein Hydrolases	0	5	8	5	7	9	10
UDP-N-acetylmuramate from Fructose-6-phosphate Biosynthesis	0	0	0	6	5	5	6
Gram-Positive cell wall components	1	1	2	2	1	1	2
Virulence, Disease and Defense	21	22	44	48	45	41	51
Resistance to antibiotics and toxic compounds	17	12	32	37	32	28	37
Bacteriocins, ribosomally synthesized antibacterial peptides	0	0	2	10	0	0	0
Invasion and intracellular resistance	4	10	10	11	13	13	14
Potassium metabolism	7	11	14	17	16	15	18
Potassium homeostasis	7	11	14	17	16	15	18

Miscellaneous	10	3	17	18	17	17	17
Phages, Prophages, Transposable elements, Plasmids	1	1	8	5	6	8	4
Membrane Transport	33	16	49	63	52	48	58
ABC transporters	0	0	3	6	63	4	5
Protein translocation across cytoplasmic membrane	3	3	2	3	3	3	3
Cation transporters	4	3	8	5	5	5	4
NhaA, NhaD and Sodium-dependent phosphate transporters	0	1	2	2	1	1	2
ECF class transporters	0	0	0	8	0	0	0
Ton and Tol transport systems	10	6	13	19	14	16	2
TRAP transporters	3	3	3	4	4	3	4
Protein and nucleoprotein secretion system, Type IV pilus	13	0	17	16	18	16	21
Iron acquisition and metabolism	1	0	10	8	10	10	8
RNA Metabolism	26	23	52	60	54	49	65
RNA processing and modification	12	5	25	25	26	24	28
Transcription	13	16	23	23	21	20	34
Group II intron-associated genes	1	2	4	12	7	5	3
Nucleosides and Nucleotides	20	25	70	67	70	65	104
Pyrimidines	0	0	13	15	16	15	37
Purines	11	12	38	39	35	34	43
Ribonucleotide reduction	2	2	3	3	3	3	6
Hydantoin metabolism	4	4	9	3	10	7	9
Detoxification	3	7	7	7	6	6	9
Protein Metabolism	73	80	188	178	176	179	188
Protein folding	9	8	12	12	10	10	15
Selenoproteins	2	2	2	5	2	2	6
Protein biosynthesis	23	43	115	107	112	112	116
Protein processing and modification	25	20	27	24	24	26	25
Protein degradation	14	7	32	30	28	29	26
Cell Division and Cell Cycle	0	4	33	26	30	30	54
Motility and Chemotaxis	0	5	0	0	0	0	0
Bacterial Chemotaxis	0	5	0	0	0	0	0

Regulation and Cell signaling	23	34	49	43	50	46	44
cAMP signaling in bacteria	16	22	36	25	40	35	34
LysR-family proteins in Salmonella enterica Typhimurium	0	0	0	1	0	1	0
LysR-family proteins in Escherichia coli	0	0	0	1	0	1	0
DNA-binding regulatory proteins, strays	1	1	0	2	1	0	0
Stringent Response, (p)ppGpp metabolism	2	2	2	2	2	2	2
Programmed Cell Death and Toxin-antitoxin Systems	4	9	11	12	7	7	
<b>DNA Metabolism</b>	<b>32</b>	<b>59</b>	<b>127</b>	<b>85</b>	<b>97</b>	<b>96</b>	<b>106</b>
DNA repair	15	24	62	51	53	53	81
DNA replication	4	2	4	4	4	4	5
DNA recombination	0	0	0	0	0	0	0
CRISPs	0	7	24	11	19	13	7
DNA uptake, competence	2	2	2	2	3	2	2
Type I Restriction-Modification	4	10	12	4	5	9	2
Restriction-Modification System	4	10	12	4	5	9	2
DNA ligases	1	0	1	1	0	0	1
DNA phosphorothioation	0	0	3	2	2	2	0
DNA structural proteins, bacterial	2	4	7	6	6	4	6
<b>Fatty Acids, Lipids, and Isoprenoids</b>	<b>12</b>	<b>1</b>	<b>54</b>	<b>32</b>	<b>54</b>	<b>59</b>	<b>61</b>
Triacylglycerols	2	0	4	2	4	4	0
Phospholipids	0	0	20	0	18	20	0
Fatty acids	0	0	13	13	15	3	20
Isoprenoids	10	1	17	17	17	17	4
<b>Nitrogen Metabolism</b>	<b>13</b>	<b>16</b>	<b>23</b>	<b>21</b>	<b>23</b>	<b>23</b>	<b>25</b>
Cyanate hydrolysis	5	5	6	6	6	6	6
Ammonia assimilation	8	11	17	15	17	17	19
<b>Respiration</b>	<b>29</b>	<b>36</b>	<b>113</b>	<b>75</b>	<b>82</b>	<b>78</b>	<b>59</b>
Biotin	6	7	15	15	14	14	22
ATP synthases	9	9	13	12	10	10	0
Electron accepting reactions	3	3	50	14	21	16	18
Electron donating reactions	5	8	8	15	15	16	12
Cytochrome B6-F complex	0	5	8	9	8	8	6

Soluble cytochromes and functionally related electron carriers	10	11	18	19	19	20	18
Quinone oxidoreductase family	0	0	0	0	1	0	0
Biogenesis of cytochrome c oxidases	0	0	0	1	2	3	2
Biogenesis of c-type cytochromes	2	0	3	3	3	2	3
<b>Stress Response</b>	<b>22</b>	<b>61</b>	<b>84</b>	<b>80</b>	<b>90</b>	<b>90</b>	<b>115</b>
Oxidative stress	17	33	53	51	57	56	73
Osmotic stress	0	1	0	0	0	0	5
Detoxification	5	9	9	9	9	9	12
SigmaB stress response regulation		7	10	12	10	10	11
Bacterial hemoglobins		15	15	11	15	16	22
Commensurate regulon activation	0	0	0	0	1	1	0
Periplasmic Stress	3	3	15	4	4	4	1
<b>Amino Acids and Derivatives</b>	<b>117</b>	<b>112</b>	<b>257</b>	<b>247</b>	<b>259</b>	<b>265</b>	<b>328</b>
Glutamine, Glutamate, Aspartate and Asparagine Biosynthesis	8	12	23	21	24	23	24
Glutamate dehydrogenases	1	0	1	1	1	1	2
Glutamine synthetases	2	2	3	1	3	3	6
Histidine Biosynthesis	8	0	12	12	12	3	12
Arginine; urea cycle, polyamines	37	10	43	38	45	13	65
S-methylmethionine	0	0	1	1	1	1	1
Methionine Biosynthesis	0	0	22	23	22	23	23
Threonine and Homoserine Biosynthesis	7	9	12	11	12	12	12
Methionine Salvage		3	0	0	0	0	0
Threonine degradation		1	3	2	1	2	1
Lysine Biosynthesis DAP Pathway	0	0	9	9	8	9	11
Cysteine Biosynthesis	7	6	12	13	12	12	12
Creatine and Creatinine Degradation	4	5	7	5	9	8	9
Branched-Chain Amino Acid Biosynthesis	0	0	16	15	16	17	16
Leucine Biosynthesis	4	5	8	7	8	9	9
Aromatic amino acids and derivatives	26	34	35	40	36	37	43
Proline and 4-hydroxyproline	0	0	7	7	8	7	11
Alanine, serine, and glycine Biosynthesis	13	23	34	32	33	37	52
<b>Sulfur Metabolism</b>	<b>0</b>	<b>7</b>	<b>18</b>	<b>19</b>	<b>19</b>	<b>21</b>	<b>20</b>
<b>Phosphorus Metabolism</b>	<b>19</b>	<b>20</b>	<b>33</b>	<b>40</b>	<b>40</b>	<b>38</b>	<b>53</b>

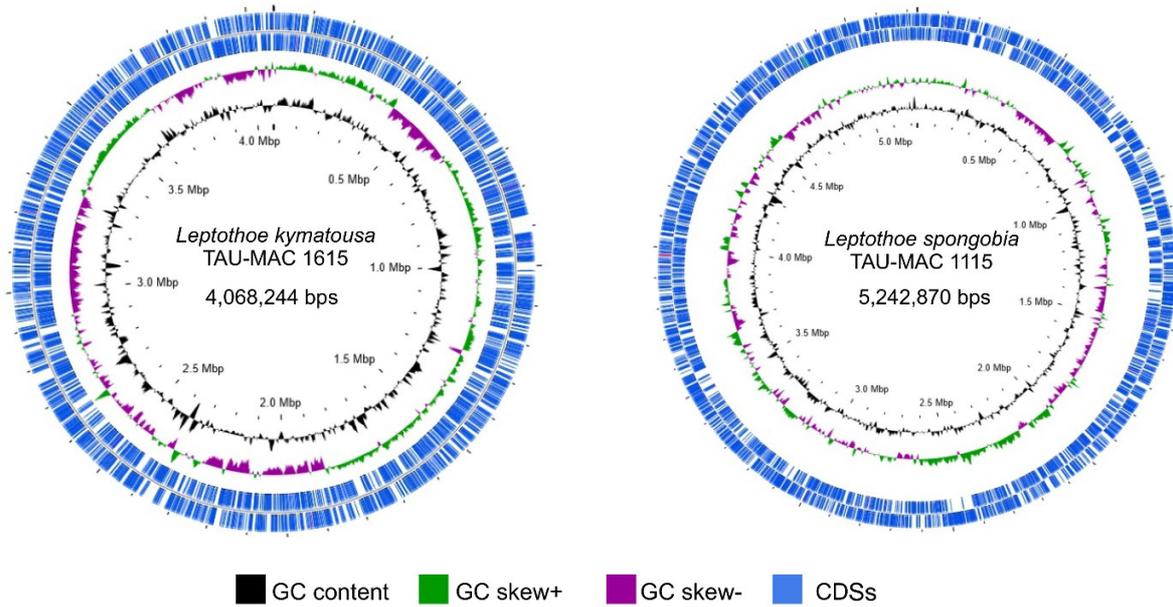
High affinity phosphate transporter and control of PHO regulon	3	3	6	12	9	9	8
Phosphate metabolism	12	13	22	23	26	24	36
Polyphosphate	4	4	5	5	5	5	9
<b>Carbohydrates</b>	<b>71</b>	<b>73</b>	<b>197</b>	<b>177</b>	<b>184</b>	<b>181</b>	<b>241</b>
Central carbohydrate metabolism	10	12	99	80	91	86	124
Aminosugars	2	2	3	3	3	3	3
Di- and oligosaccharides	5	9	14	11	12	14	10
One-carbon Metabolism	4	4	5	5	5	5	4
Organic acids	3	0	0	0	0	0	0
Fermentation	2	5	23	18	21	19	28
CO <sub>2</sub> fixation	29	20	27	32	25	28	44
Sugar alcohols	3	3	5	6	6	5	6
Polysaccharides	6	10	11	11	11	11	11
Monosaccharides	7	8	10	11	10	10	10

**Table S2.** Biosynthetic gene clusters per *Leptothoe* genome (antiSMASH results)

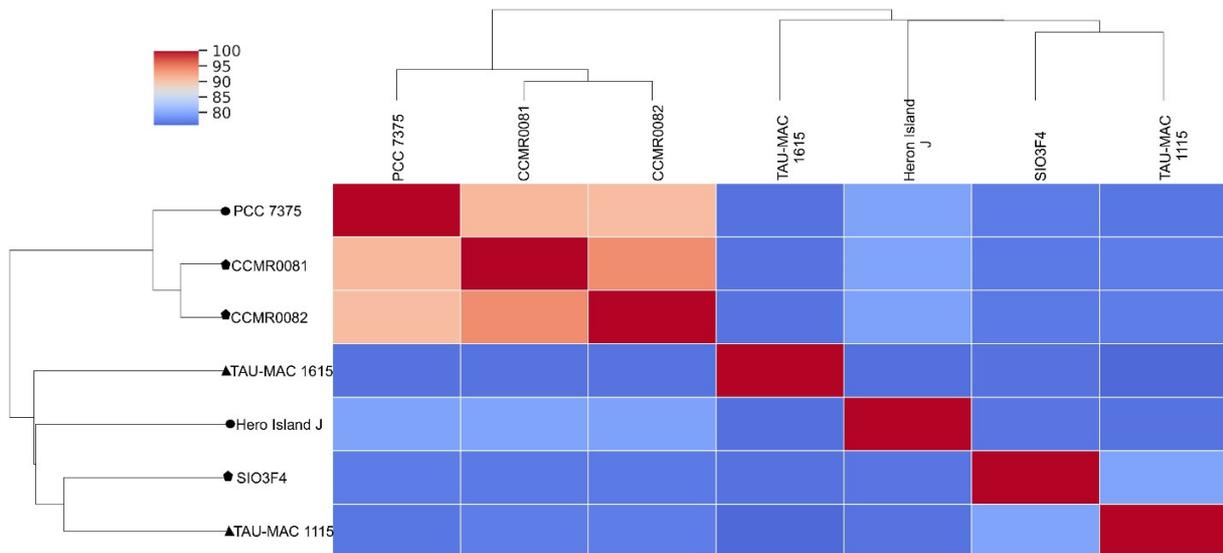
Strain	Number of genes												Total
	NR PS	NRPS-like	T1P KS	T3P KS	PKS-like	Bacteriocin	Lasso-peptide	Terpene	Cyanobactin	Ar-ylpoly-ene	Lanthipeptide	LA P	
<i>Le. spongia</i> TAU-MAC 1615	0	1	0	0	0	0	0	1	0	1	0	0	3
<i>Le. kymatousa</i> TAU-MAC 1115	1	1	2	0	0	3	2	1	0	0	1	0	11
Leptolyngbyaceae cyanobacterium CCMR0081	9	3	7	1	1	4	2	2	2	1	0	1	33
Leptolyngbyaceae cyanobacterium CCMR0082	5	2	5	2	2	5	2	2	2	1	0	1	29
<i>Leptolyngbya</i> sp. SIO3F4	3	2	3	1	0	2	0	3	0	1	0	0	15
<i>Leptolyngbya</i> sp. PCC 7375	2	3	0	1	0	2	1	2	1	1	0	0	13
<i>Leptolyngbya</i> sp. Heron Island J	4	3	3	0	0	1	2	3	0	1	0	0	17
<b>Total</b>	<b>24</b>	<b>15</b>	<b>20</b>	<b>5</b>	<b>3</b>	<b>17</b>	<b>9</b>	<b>14</b>	<b>5</b>	<b>6</b>	<b>1</b>	<b>2</b>	

**Table S3.** Compounds produced by *Leptothoe* cyanobacteria extracted from the CyanoMetDB (Jones et al., 2020).

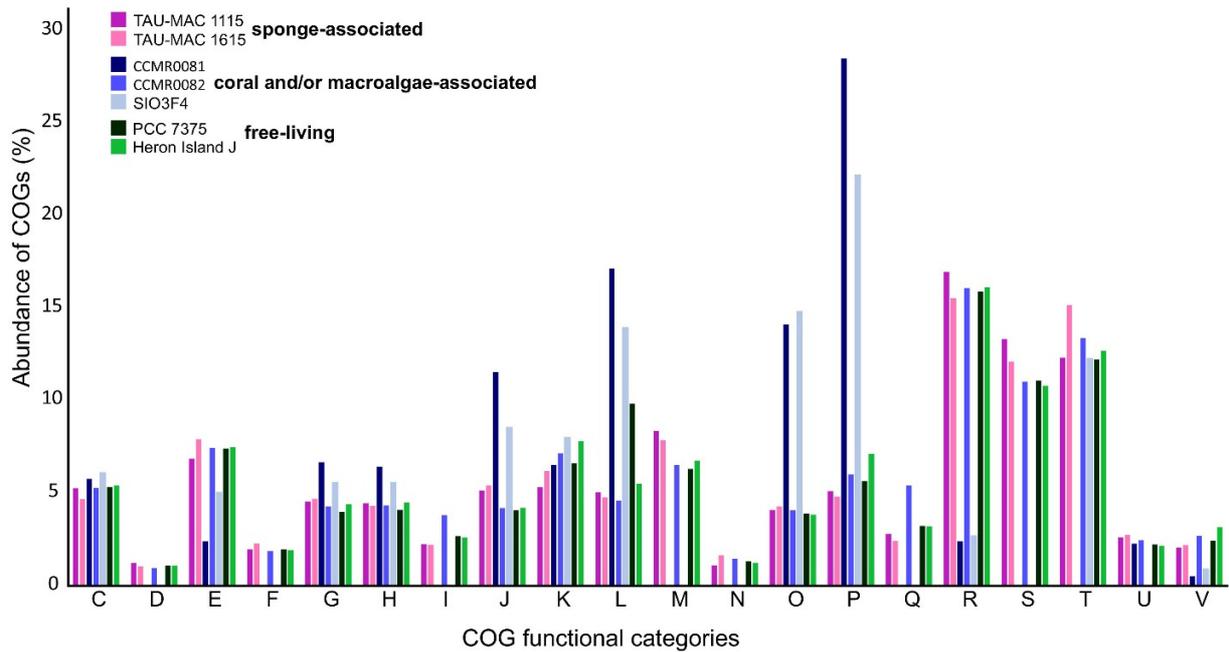
Compound name	Class of compound	Strain	Reference	DOI
Grassypeptolide E	other cyclic depsipeptide	<i>Leptolyngbya</i> sp. RSO3	Thornburg et al. 2011	<a href="https://doi.org/10.1021/np200270d">https://doi.org/10.1021/np200270d</a>
Grassypeptolide D	other cyclic depsipeptide			
Ibu-epidemethoxylyngbyastatin 3	other cyclic depsipeptide			
Dolastatin 12	other cyclic depsipeptide			
Hierridin B	other linear non-peptide	<i>Phormidium ectocarpi</i> SAG 60.90	Papendorf et al. 1998	<a href="https://doi.org/10.1016/S0031-9422(98)00440-3">https://doi.org/10.1016/S0031-9422(98)00440-3</a>
Phycocerythrobilin	other linear non-peptide	<i>Phormidium persicinum</i>	Chapman et al. 1967	<a href="https://doi.org/10.1021/ja00999a058">https://doi.org/10.1021/ja00999a058</a>



**Figure S1.** Circular view of the genomes of two sponge-associated *Leptothoe* species, generated with CGview (Stothard and Wishart, 2005). Circles from interior to exterior represent GC content and GC skew. Blue circles denote the coding sequences on forward and reverse strands.



**Figure S2.** Average amino acid identity heatmap of *Leptothoe* genomes.



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