

Full Original Paper

A Sesquiterpene Quinone, 5-Epi-smenospongine, Promotes TNF- α Production in LPS-stimulated RAW 264.7 Cells

Taiko Oda ^{1,*}, Weifang Wang ², Kazuyo Ukai ², Takahiro Nakazawa ² and Masataka Mochizuki ¹

¹ Kyoritsu University of Pharmacy, Shibakoen, Minato-ku, Tokyo 105-8512, Japan

² Tohoku Pharmaceutical University, Komatsushima, Aoba-ku, Sendai 981-8558, Japan

*Author to whom correspondence should be addressed: Tel. +81-3-5400-2497, Fax +81-3-5400-2497, E-mail: oda-ti@kyoritsu-ph.ac.jp

Received: 25 September 2007 / Accepted: 29 October 2007 / Published: 30 October 2007

Abstract: Eight sesquiterpene quinones: ilimaquinone (**1**), smenospongidine (**3**), smenospongiarine (**5**), smenospongine (**7**), and their corresponding 5-epimers **2**, **4**, **6**, and **8**, isolated from the Palauan marine sponge *Hippospongia* sp., were examined regarding their effects on TNF- α production in LPS-stimulated RAW 264.7 cells. 5-Epi-smenospongine (**8**) promoted the production of TNF- α to a level three times greater than the control at 10 μ M, but compounds **1-7** did not show apparent activity. The results suggest that the *cis*-decaline ring and a primary amine in the benzoquinone ring are necessary for activity. This is the first study to report the modulation of TNF- α production by a sesquiterpene quinone.

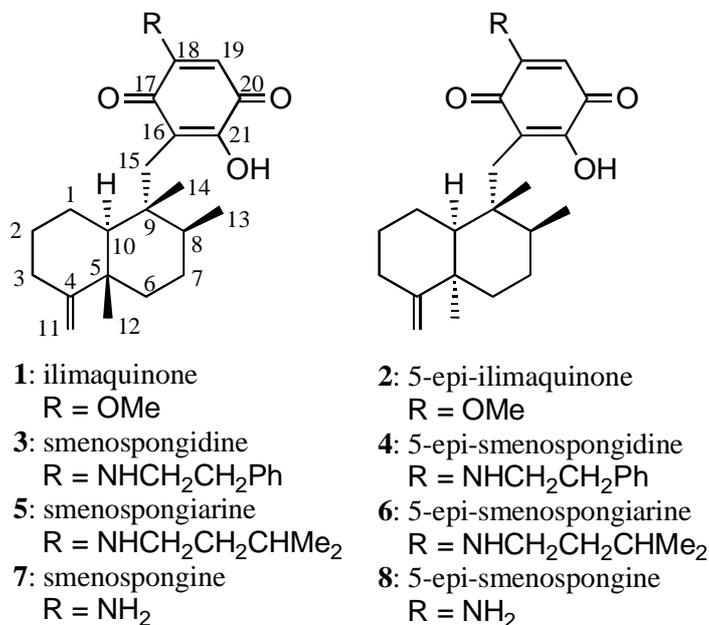
Keywords: Sesquiterpene quinone, marine sponge, *Hippospongia* sp., TNF- α production, RAW 264.7 cells

1. Introduction

Marine sponges (Porifera) are well-known as prolific producers of bioactive metabolites. A sesquiterpene quinone, ilimaquinone, was reported in 1979 [1,2], and since then, related compounds have been obtained from several genera of marine sponge [3-9]. We have isolated 11 sesquiterpene quinones: ilimaquinone (**1**) [1,2] and its 5-epimer (**2**) [3], smenospongidine (**3**) [3,4] and its 5-epimer

(4) [4], smenospongiarine (5) [4,5] and its 5-epimer (6) [4], smenospongine (7) [4,6] and its 5-epimer (8) [9], dactyloquinone B [7], 18-hydroxy-5-epihyrthiophenol [8], and pelorol [9], from *Hippospongia* sp. collected in Palau, and reported the effects of these compounds on the production of an inflammatory cytokine, interleukin-8 (IL-8), in tetradecanoyl phorbol acetate (PMA)-stimulated human promyelocytic leukemia HL-60 cells [10]. IL-8 is a member of the superfamily of C-X-C chemokines and a chemotactic factor for T-cells, neutrophils, and basophils [11]. Expression of IL-8 has been detected in a variety of human cancers and is suggested to be a factor involved in tumor progression and metastasis [12-15]. Compounds 3-8 increased the production of IL-8; therefore, we examined the effects of compounds 1-8 on the production of one of the most well-known and important inflammatory cytokines, TNF- α , in lipopolysaccharide (LPS)-stimulated murine monocytic leukemia RAW 264.7 cells.

Figure 1. Structures of compounds 1-8.



2. Materials and Methods

2.1. Test Compounds

Sesquiterpene quinones 1-8 were isolated as described in the previous paper [10]. The structures of 1-8 are shown in Fig. 1.

2.2. Cell lines and culture conditions

The RAW 264.7 cell line was obtained from the Japanese Cancer Research Resources Bank (JCRB, Kamiyoga, Tokyo, Japan). This cell line was maintained in tissue culture dishes in RPMI 1640 medium (Nissui Seiyaku, Tokyo, Japan), supplemented with 10% heat-inactivated FCS, 2 mM glutamine, 100 U/ml of penicillin G and 100 μ g/ml of streptomycin.

2.3. Detection of murine TNF- α by ELISA

TNF- α concentrations of the culture supernatants under control and various test conditions were measured by ELISA using a combination of monoclonal and polyclonal antibodies. All samples were assayed at least in duplicate. Data are presented as the mean \pm SE of three independent experiments.

2.4. Determination of cell proliferation

Cell proliferation was evaluated by enumerating the viable cells using the MTT formazan production method [16]. RAW264.7 cells (1×10^6 cells/mL) were treated with LPS (with or without test compounds) and then transferred to 96-well microtiter plates. After incubation for 24 h, 20 μ L of MTT reagent (5 mg/mL in PBS) was added to each well and further incubated for 3 h. The production of formazan was assessed by measuring optical density (OD₅₇₀ nm). Data are shown as the values relative (%) to each PMA-stimulated optical density.

3. Results and Discussion

RAW 264.7 cells are known to produce TNF- α in response to the addition of LPS, and this system is used to detect the modulating activities of compounds on TNF- α production.

5-Epi-smenospongine (**8**) stimulated the production of TNF- α to a level three times greater than the control, but compounds **1-7** did not show any apparent activity (Fig. 2).

Compounds **3**, **5**, and **7** exhibited stronger activity on the promotion of IL-8 production than the corresponding 5-epimers **4**, **6**, and **8** in PMA-stimulated HL-60 cells [10], which showed the contribution of the *trans*-decaline ring in enhancing activity. The basic functional group at C-18 increased IL-8 production, since **3-8** exhibited stronger activities than **1** and **2** [10]. It was therefore revealed that the production of TNF- α induced by **8** involves a different mechanism from that involved in IL-8 production.

Ilimaquinone (**1**) showed antibacterial, cytotoxic, anti-HIV, hemolytic, and antimetabolic activities, disruption of the Golgi apparatus, inhibition of the cytotoxicity of ricin and diphtheria toxin, and differentiation-inducing activity [5, 17-22]. Compounds **2-5** and **7** were reported to have cytotoxic, antibacterial, and hemolytic activities [4, 5, 17]. Compounds **2-4**, **7**, and **8** induced the differentiation of K562 cells into erythroblasts, as did **1** [22].

In this study, we revealed the production of TNF- α production in LPS-stimulated RAW 264.7 cells by one of the eight sesquiterpene quinones, 5-epi-smenospongine (**8**). This is the first report on the modulation of TNF- α production by sesquiterpene quinones.

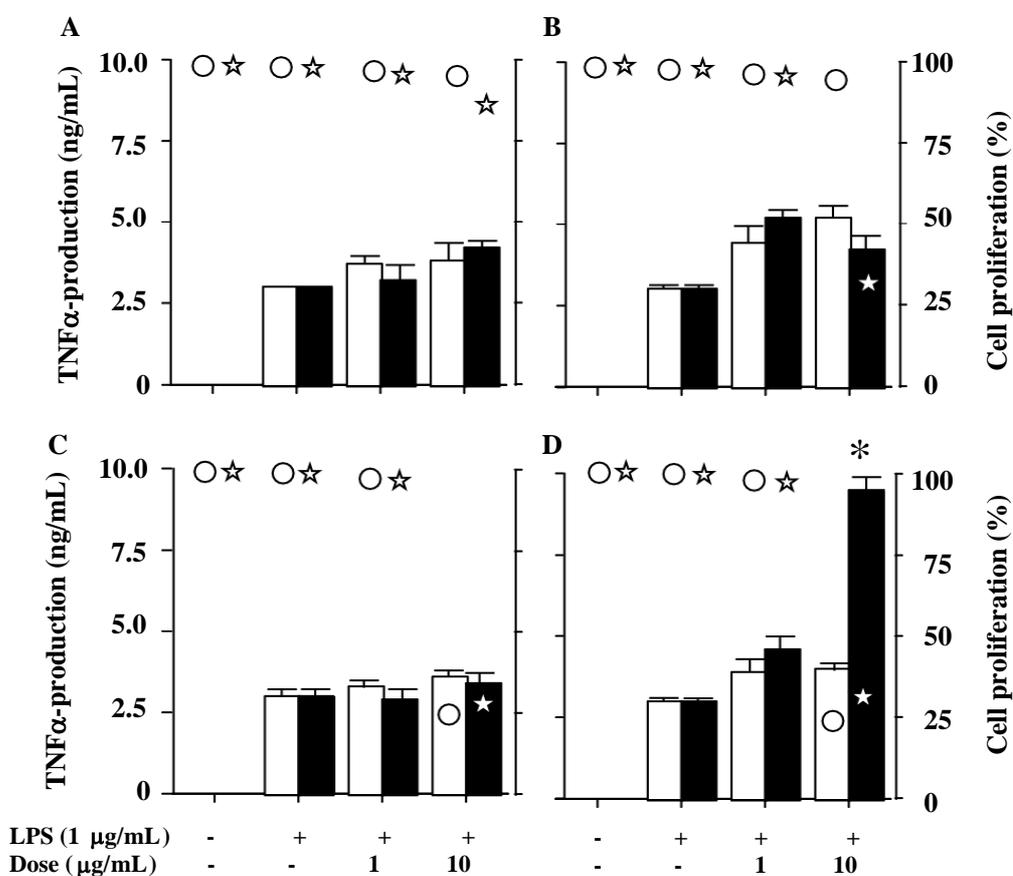
From the comparison of structures **1-8**, it is suggested that the *cis*-decaline ring and a primary amine in the benzoquinone ring are necessary for the activity of **8**.

As we previously reported, compounds **3** to **8** induced the production of IL-8 by PMA-stimulated HL-60 cells [10]. In this study, we investigated their actions on TNF- α produced by LPS-stimulated mouse macrophage cell lines. Compound **8** induced TNF- α production. However, there was no influence of the other compounds. This may have been associated with the different actions of these compounds related to variations in receptors, signal molecules, and transcription factors in the

production process of IL-8 and TNF- α as inflammatory cytokines. Therefore, the results of our experiment suggest that individual inflammatory cytokines can be controlled; these compounds should be further developed in the future.

Figure 2. Effects of compounds 1–8 on TNF- α production and cell proliferation in LPS-stimulated RAW 264.7 cells.

Panels A, B, C, and D show the results of compounds 1 and 2, 3 and 4, 5 and 6, and 7 and 8, respectively. Open and solid bars represent lower and higher numbered compounds, respectively (* $P < 0.05$). Open circle and star represent lower and higher numbered compounds, respectively.



Acknowledgements

We thank Prof. M. Namikoshi of Tohoku Pharmaceutical University for correcting our manuscript, and Ms. A. Fujita of Tokyo University of Marine Science and Technology for technical assistance.

References

1. Luibrand, R.T.; Erdman, T.R.; Vollmer, J.J.; Scheuer, P.J.; Finer, J.; Clardy, J. Ilimaquinone, a sesquiterpenoid quinone from a marine sponge. *Tetrahedron* **1979**, *35*, 609–612.

2. Capon, R.J.; MacLeod, J.K. A revision of the absolute stereochemistry of ilimaquinone. *J. Org. Chem.* **1987**, *52*, 5059–5060.
3. Carté, B.; Rose, C.B.; Faulkner, D.J. 5-*epi*-Ilimaquinone, a metabolite of the sponge *Fenestraspongia* Sp. *J. Org. Chem.* **1985**, *50*, 2785–2787.
4. Rodríguez, J.; Quiñoá, E.; Riguera, R.; Peters, B. M.; Abrell, L.M.; Crews, P. The structures and stereochemistry of cytotoxic sesquiterpene quinones from *Dactylospongia elegans*. *Tetrahedron* **1992**, *48*, 6667–6680.
5. Kondracki, M-L.; Guyot, M. Biologically active quinone and hydroquinone sesquiterpenoids from the sponge *Smenospongia* sp. *Tetrahedron* **1989**, *45*, 1995–2004.
6. Kondracki, M-L.; Guyot, M. Smenospongine: a cytotoxic and antimicrobial aminoquinone isolated from *Smenospongia* sp. *Tetrahedron Lett.* **1987**, *28*, 5815–5818.
7. Mitome, H.; Nagasawa, T.; Miyaoka, H.; Yamada, Y. van Soest, R.W. Dactyloquinones A and B, new sesquiterpenoid quinones from the Okinawan marine sponge *Dactylospongia elegans*. *J. Nat. Prod.* **2001**, *64*, 1506–1508.
8. Salmoun, M.; Devijver, C.; Daloze, D.; Braekman, J.C.; Gomez, R.; de Kluijver, M.; van Soest, R.W.M. New sesquiterpene/quinones from two sponges of the genus *Hyrtios*. *J. Nat. Prod.* **2000**, *63*, 452–456.
9. Kwak, J.H.; Schmitz, F.J.; Kelly, M. Sesquiterpene quinols/quinones from the Micronesian sponge *Petrosaspongia metachromia*. *J. Nat. Prod.* **2000**, *63*, 1153–1156.
10. Oda, T.; Wang, W.; Fujita, A.; Mochizuki, M.; Ukai, K.; Namikoshi, M. Promotion of IL-8 production in PMA-stimulated HL-60 cells by sesquiterpene quinones from a marine sponge *Hippospongia* sp. *J. Nat. Med.* **2007**, *61*, 434–437.
11. Graves, D.T.; Jiang, Y. Chemokines, a family of chemotactic cytokines. *Crit. Rev. Oral Biol. Med.* **1995**, *6*, 109–118.
12. di Celle, P.F.; Carbone, A.; Marchis, D.; Zhou, D.; Sozzani, S.; Zupo, S.; Pini, M.; Mantovani, A.; Foa, R. Cytokine gene expression in B-cell chronic lymphocytic leukemia: evidence of constitutive interleukin-8 (IL-8) mRNA expression and secretion of biologically active IL-8 protein. *Blood* **1994**, *84*, 220–228.
13. Green, A.R.; Green, V. L.; White, M.C.; Speirs, V. Expression of Cytokine messenger RNA in normal and neoplastic human breast tissue: identification of interleukin-8 as a potential regulatory factor in breast tumours. *Int. J. Cancer* **1997**, *72*, 937–941.
14. König, B.; Steinbach, F.; Janocha, B.; Drynda, A.; Stumm, M.; Philipp, C.; Allhoff, E.P.; König, W. The differential expression of proinflammatory cytokines IL-6, IL-8 and TNF-alpha in renal cell carcinoma. *Anticancer Res.* **1999**, *19*, 1519–1524.
15. Galffy, G.; Mohammed, K.A.; Dowling, P.A.; Nasreen, N.; Ward, M. J.; Antony, V.B. Interleukin 8: an autocrine growth factor for malignant mesothelioma. *Cancer Res.* **1999**, *59*, 367–371.
16. Carmichael, J.; DeGraff, W.G.; Gazdar, A.F.; Minna, J.D.; Mitchell, J.B. Evaluation of a tetrazolium-based semiautomated colorimetric assay: assessment of chemosensitivity testing. *Cancer Res.* **1987**, *47*, 936–942.
17. Prokofeva, N.G.; Utkina, N.K.; Chaikina, E.L.; Makarchenko, A.E. Biological activities of marine sesquiterpenoid quinones: structure-activity relationships in cytotoxic and hemolytic assays. *Comp. Biochem. Physiol. Bio. Mol. Biol.* **2004**, *139*, 169–173.

18. Tziveleka, L.A.; Vagias, C.; Roussis, V. Natural products with anti-HIV activity from marine organisms. *Curr. Top. Med. Chem.* **2003**, *3*, 1512–1535.
19. Pous, C.; Chabin, K.; Drechou, A.; Barbot, L.; Phung-Koskas, T.; Settegrana, C.; Bourguet-Kondracki, M.L.; Maurice, M.; Cassio, D.; Guyot, M.; Durand, G. Functional specialization of stable and dynamic microtubules in protein traffic in WIF-B cells. *J. Cell. Biol.* **1998**, *142*, 153–165.
20. Cruciani, V.; Mikalsen, S.O. Ilimaquinone inhibits gap junctional communication in a connexin isotype-specific manner. *Exp. Cell Res.* **2005**, *304*, 136–148.
21. Nambiar, M. P.; Wu, H.C. Ilimaquinone inhibits the cytotoxicities of ricin, diphtheria toxin, and other protein toxins in Vero cells. *Exp. Cell Res.* **1995**, *219*, 671–678.
22. Aoki, S.; Kong, D.; Matsui, K.; Rachmat, R.; Kobayashi, M. Sesquiterpene aminoquinones, from a marine sponge, induce erythroid differentiation in human chronic myelogenous leukemia, K562 cells. *Chem. Pharm. Bull.* **2004**, *52*, 935–937.

Samples Availability: Not available.

© 2007 by MDPI (<http://www.mdpi.org>). Reproduction is permitted for noncommercial purposes.