



Article

Interaction of Smoking and Lead Exposure among Carriers of Genetic Variants Associated with a Higher Level of Oxidative Stress Indicators

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Citation: Ho, K.-J.; Chen, T.-H.; Yang, C.-C.; Chuang, Y.-C.; Chuang, H.-Y. Interaction of Smoking and Lead Exposure among Carriers of Genetic Variants Associated with a Higher Level of Oxidative Stress Indicators. *Int. J. Environ. Res. Public Health* **2021**, *18*, 8325. <https://doi.org/10.3390/ijerph18168325>

Academic Editor: Mei-Fang Chien

Received: 15 June 2021

Accepted: 4 August 2021

Published: 6 August 2021

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Abstract: Smoking and lead (Pb) exposure increased oxidative stress in human body, and people with some gene variants may be susceptible to Pb and smoking via oxidative stress. The aim of this study is to evaluate oxidative stress by measuring thiobarbituric acid reactive substances (TBARS) and the relationship of lipid peroxidation markers in Pb workers with different gene polymorphisms (rs4673 and rs1050450) in both smokers and nonsmokers. Blood samples were collected from 267 Pb workers who received their annual health examination in the Kaohsiung Medical University Hospital. Glutathione peroxidase 1 (GPx-1) rs1050450 and cytochrome B-245 Alpha Chain (CYBA) rs4673 single-nucleotide polymorphisms (SNP) were analyzed by specific primer-probes using Real-Time PCR methods. The interaction between blood Pb and smoking increased serum levels of TBARS and the ratio of oxidative low-density lipoprotein and low-density lipoprotein (oxLDL/LDL). Analysis of workers with rs1050450 SNPs showed higher blood Pb levels in the workers with CC genotype than those with CT genotype. Smokers had significantly higher blood Pb, alanine transaminase (ALT), TBARS, and OxLDL levels than nonsmokers. TBARS increased 0.009 nmol/mL when blood Pb increased one $\mu\text{g}/\text{dL}$ in smokers compared to nonsmokers. The ratio of OxLDL/LDL increased 0.223 when blood Pb increased one $\mu\text{g}/\text{dL}$ in smokers compared to nonsmokers. TBARS levels and the ratio of OxLDL/LDL were positively correlated and interacted between blood Pb and smoking after the adjustment of confounders, suggesting that smoking cessation is an important issue in the Pb-exposed working environment.

Keywords: lead; smoke; TBARS; OxLDL; GPx-1; CYBA; single nucleotide polymorphisms



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1. Introduction

Lead (Pb) is a well-known ubiquitous environmental pollutant as well as one of the most commonly used metals in industries, especially for battery manufacturing and repair, paint manufacturing, ship demolition and construction, radiation shield manufacturing, and metal soldering [1–3]. Adverse health effects of Pb have been reported in many human organs and systems, including cardiovascular, renal, hematologic, and nervous systems. Among them, Pb-induced oxidative stress impairs cell components, such as DNA, protein, and lipids through the generation of reactive oxidative species (ROS) [4,5].

Several antioxidant enzymes and molecules have been used for the measurement of the severity of Pb-induced oxidative stress in many human and animal studies. The most commonly used include glutathione (GSH), glutathione disulfide (GSSG), glutathione peroxidase (GPx), superoxide dismutase (SOD), catalase (CAT), and malondialdehyde (MDA) [5,6]. These oxidative stress markers present the severity of lipid peroxidation, DNA damage, and depletion of cell antioxidant defense systems, which might be indicators of Pb-induced adverse health effects [7].

Oxidative stress has been reported as a major mechanism of Pb-induced toxicity [8]. Nicotinamide adenine dinucleotide/nicotinamide adenine dinucleotide phosphate (NADH/NADPH) oxidase is a key enzyme of superoxide production in the vasculature, smooth muscle cells, phagocytes, and mesangial cells [9,10], and is also implicated in a wide variety of human diseases including metabolic syndrome [11], hypertension [12], diabetes [13], renal disease [14], atherosclerosis [15], and cerebrovascular disease [16]. The cytochrome B-245 Alpha Chain (CYBA) gene (p22phox) is an essential component of NADPH oxidase located on 16q24. One of the most common single-nucleotide polymorphisms (SNP) of the CYBA gene is C242T (rs4673), resulting in a non-conservative substitution of histidine for tyrosine at codon 72 and an alteration of NAD(P)H activity by disrupting the heme-binding site [10]. The C242T polymorphism has been demonstrated to be related to multiple inflammatory diseases [17], metabolic disorders [18], and coronary artery disease [19].

In addition, glutathione peroxidase (GPX) is one of the antioxidant systems involved in the defense of ROS [20]. GPX1 is the most abundant enzyme in the GPX isoenzyme family [21]. GPX1 is a selenoprotein containing two exons within a 1.42-kb region located at chromosome 3p21 with antioxidant and anti-inflammatory functions [22,23].

Based on the dbSNP records (www.ncbi.nlm.nih.gov/snp, accessed on 8 May 2021), the polymorphic sites are widely distributed within the introns, exons, and UTR regions of the GPX1 gene so that they may affect the GPX1 function [23–25]. The rs1050450 (Pro198Leu) polymorphism is a site located within the GPX1 C-terminal region. The rs1050450 variant might be associated with cancer risk [26], might increase the risk of peripheral neuropathy in diabetes group [27], and has shown a higher prevalence of metabolic syndrome and cardiovascular diseases [24,28,29].

Low-density lipoprotein (LDL) is composed of cholesteryl ester, phospholipids, free cholesterol, triglycerides, and apolipoprotein B100 (apoB100) [30,31]. Oxidized low-density lipoprotein (OxLDL) is a complex particle derived from circulating LDL that contains lipid peroxidation and an apolipoprotein B modification, mostly from LDL, which could be used for detection of lipid oxidation [30,32]. Also, OxLDL triggered the formation of foam cells which induced the development of atherosclerosis by promoting monocyte-derived macrophages in the arterial wall and the intracellular accumulation of cholesteryl esters in these cells, which could be an indicator for evaluation of the cardiovascular disease [33].

Additionally, thiobarbituric acid reactive substances (TBARS) are also detectors for screening and monitoring lipid peroxidation [34–37]. Higher TBARS levels might increase the risk of cardiovascular diseases [38]. Schisterman EF et al. found that TBARS were the best discriminating biomarkers among reduced glutathione (GSH), Trolox equivalent antioxidant capacity (TEAC), high-density lipoprotein (HDL), uric acid, and Glutathione hydroxyperoxides peroxidase (GSHPx) when individually evaluated for coronary heart disease cases [39].

Cigarette smoking has been proved to be a source of oxidative stress [40,41]. Oxidative stress markers for smokers including F2-isoprostanes, F4-neuroprostanes, hydroxyeicosatetraenoic acid products (HETEs), 7-ketocholesterol, and 24- and 27-hydroxycholesterol were previously measured [40]. Both TBARS and OxLDL were elevated in smokers [7]. TBARS seem to be positively associated with smoking status [42].

Thus, Pb exposure and smoking impair human health by inducing oxidative damage. No investigation of the interaction of smoking and workers with Pb exposure has been performed. The aim of our research is to analyze the oxidative stress by measuring TBARS

and the relationship of lipid peroxidation markers in workers with Pb exposure with different gene polymorphisms (rs4673 and rs1050450) in both smokers and nonsmokers.

2. Materials and Methods

2.1. Study Population and Health Examination

In order to evaluate the relationship between Pb exposure and oxidative damage, we performed a cross-sectional study of 267 Pb workers who received their annual health examination in Kaohsiung Medical University Hospital. The exclusion criteria included workers who were cancer patients and taking hypercholesterolemia medicine. Anthropometric measurements including body mass index (BMI) were checked. Blood samples were obtained including lipid profile (total cholesterol, triglyceride, low-density lipoprotein [LDL], high-density lipoprotein [HDL]), fasting sugar (AC), liver function test (alanine transaminase, ALT), renal function test (serum creatinine, Cr), and oxidative stress markers (OxLDL, TBARS). The blood Pb level was measured by Graphite Furnace Atomic Absorption Spectrometry (GFAAS). All the samples were measured in the Central Laboratory in Kaohsiung Medical University Hospital. The study was approved by the IRB of Kaohsiung Medical University Hospital (KMUHIRB-E(I)-20190034) for experimentation with human subjects. In addition, a questionnaire with information on job title, medical history, working history, personal history of alcohol and cigarette consumption was collected from each participant after a full explanation of the study and informed consent signed.

2.2. Genotyping Methods for Glutathione Peroxidase-1 (GPx-1) Gene (rs1050450) and Cytochrome b Light Chain (CYBA) Gene (rs4673)

Genomic DNA was extracted from peripheral blood using FlexiGene (Qiagen, Hilden, Germany) following the manufacturer's protocol. Each real-time PCR was performed with a 10 μ L reaction volume mix fluid, containing 5 μ L Genotyping Master Mix (Applied Biosystems, Waltham, MA, USA), 3.87 μ L distilled water, 1 μ L DNA fluid (10 ng/ μ L) and 0.25 μ L primer-probe. Amplification reactions were performed using the following program for total 45 cycles: 50 $^{\circ}$ C for 2 min; 92 $^{\circ}$ C for 10 min; 95 $^{\circ}$ C for 15 s; 60 $^{\circ}$ C for 1 min. We used TaqMan Allelic Discrimination Assays (Applied Biosystems, Foster City, CA, USA) to genotype the SNPs, and the results were read with a 7300 Real-time PCR System (Life Technologies Corp., Carlsbad, CA, USA). The fluorescence level was measured with an Applied Biosystems StepOne Real-Time PCR System (Applied Biosystems, Waltham, USA). The allele frequencies were determined using ABI SDS software. Genotyping was repeated on a random 10% sample to confirm the results of the original run by the laboratory technicians, who were blinded to the original results. The estimated genotyping error rate was less than 1%.

2.3. Measurement of TBARS and Oxidized LDL

Lipid peroxidation as an indicator of oxidative damage was determined by measuring the plasma concentration of TBARS using the method of Ohkawa et al. [43]. A standard curve of TBARS was obtained by hydrolysis of 1,1,3,3-tetraethoxypropane. Activation of the antioxidative defense in response to increased oxidative damage was evaluated by measuring the plasma level of total reduced thiols, a physiological free radical scavenger. Plasma thiols were determined by direct reaction with 5,5-dithiobis (2-nitrobenzoic acid) to form 5-thio-2-nitrobenzoic acid (TNB) and then calculated from the absorbance using the extinction coefficient of TNB ($A_{412} = 13,600 \cdot M^{-1} \text{ cm}^{-1}$) [44].

Oxidized LDL concentrations were measured using a sandwich enzyme-linked immunosorbent assay (ELISA) according to the manufacturer's instructions (Mercodia oxidized LDL ELISA; Mercodia AB, Uppsala, Sweden). In brief, diluted plasma and standards were incubated in the wells of a microtiter plate coated with murine monoclonal oxidized LDL antibodies (mAb-4E6) for 2 h at room temperature. After washing 6 times to remove nonreactive plasma components, a peroxidase-conjugated antiapolipoprotein B antibody was added to the wells with the oxidized LDL bound to the solid phase. After a

second incubation for 1 h at room temperature and a washing step to remove the unbound enzyme-labeled antibody, the bound conjugate was detected by a reaction with 3,3',5,5'-tetramethylbenzidine (TMB). This reaction was stopped by adding 1 M H₂SO₄, and the optical density (OD) was measured at OD 450 nm using a microplate reader. The results were calculated using the computerized data reduction of absorbance for the standards versus the concentration using cubic spline regression [45].

2.4. Data Analysis

There are different criteria for the definition of obesity among adults over the world. The World Health Organization (WHO) defines the Asia criteria of overweight as BMI \geq 23 and obese as BMI \geq 25 [46]. Both WHO and the National Institutes of Health (NIH) have promoted BMI cutoffs of 25 for overweight and 30 for obesity [47,48]. Lifestyle and genetic differences among different race and ethnicity could explain that Chinese and other Asian groups have a higher percentage of body fat than Caucasians given the same level of BMI [49]. The Taiwan Department of Health (DOH) defined overweight as BMI \geq 24 and obese as BMI \geq 27 [50]. A cutoff value of 24 was used in our study according to the definition of overweight of the Taiwan DOH.

Some previous researchers have used the ratio of OxLDL-to-LDL because it might reflect the clinical relevance of oxidative stress more accurately [31,51]. This process was adopted from our research.

Statistical analysis was performed using statistical software package (SPSS 20.0, IBM, Armonk, New York, NY, USA). The continuous variables including blood Pb, age, body mass index (BMI), creatinine, ALT, AC sugar, TBARS, and OxLDL were presented as mean \pm SD. Proportions were used for categorical variables such as sex, BMI < 18.5, 18.5 \leq BMI < 24, BMI \geq 24, Cr > 1.5, ALT > 80, AC sugar \geq 126, rs4673, and rs1050450 SNPs. The genotype distributions for both polymorphisms were calculated using the Hardy-Weinberg equilibrium equation. The study population was divided by different genotypes of rs4673 (CC, CT, TT) and rs1050450 (CC, CT). One-way ANOVA was used to examine the differences of continuous variables and a chi-square test was used to compare categorical variables among the different types of SNPs. We used independent t tests to examine the differences of continuous variables and chi-square tests to compare categorical variables among the different types of rs1050450 because the SNP only had 2 types (CC and CT). Regression analysis was used to evaluate the association of oxidative stress markers (TBARS and OxLDL) in groups of smokers and nonsmokers, blood Pb, and SNPs after adjusting for age, sex, BMI < 18.5, BMI \geq 24 versus 18.5 \leq BMI < 24, Cr > 1.5 versus Cr \leq 1.5, ALT > 80 versus ALT \leq 80, AC sugar \geq 126 versus AC < 126. We then analyzed the interaction terms between Pb and smoke, Pb and different SNPs on the plasma oxidative stress markers (TBARS and OxLDL/LDL) as dependent variables. A two-tailed *p*-value < 0.05 was considered significant.

3. Results

Of the total 267 workers, the highest blood Pb was 58.93 mcg/dL and the lowest was 0.03 mcg/dL (office worker) with a mean 12.3 mcg/dL (SD = 13.7). 193 (72.3%) workers were male. Smokers account for 30.3% of these workers. More than half of them were overweight (BMI higher than 24) according to the Taiwan DOH definition. Less than 10% of the population has abnormal renal function, liver function, and fasting sugar. Analysis of CYBA rs4673 polymorphism showed 205 workers with CC, 58 with CT, and 4 with TT types, which was consistent with Hardy-Weinberg equilibrium (*p* = 0.96). Analysis of GPx-1 rs1050450 polymorphism revealed only 236 with CC and 31 CT types, without TT type (consistent with Hardy-Weinberg equilibrium, *p* = 0.31).

Table 1 shows that smokers had significantly higher blood Pb, BMI, and ALT levels than nonsmokers. In addition, smokers reported using more alcohol. With regard to markers of oxidative stress, TBARS and OxLDL are significantly higher in smokers. Men had a significantly higher prevalence of smokers in comparison with women.

Table 1. Descriptive data of clinical and biochemical by smoking status.

Variables	Smokers (n = 81)	Nonsmokers (n = 186)	p Value
Blood Pb (µg/dL)	22.11 ± 15.19	7.97 ± 10.42	<0.001 **
Age (years)	43.7 ± 9.7	41.1 ± 10.4	0.058
Sex (male)	80 (98.8%)	113 (60.8%)	<0.001 **
Alcohol (>3 times/week)	28 (42.4%)	13 (7.3%)	<0.001 **
BMI (kg/m ²)	25.69 ± 3.52	23.88 ± 3.71	<0.001 **
BMI < 18.5	3 (3.7%)	7 (3.8%)	0.981
BMI ≥ 24	56 (69.1%)	83 (44.6%)	0.001 **
Creatinine (mg/dL)	1.22 ± 0.14	1.15 ± 0.63	0.357
ALT (U/L)	22.85 ± 14.42	18.19 ± 16.14	0.027 *
AC sugar (mg/dL)	102.92 ± 33.41	99.54 ± 31.08	0.433
TBARS (nmol/mL)	2.289 ± 0.559	1.953 ± 0.351	<0.001 **
OxLDL (mg/dL)	59.857 ± 14.567	53.76 ± 11.252	0.001 **
LDL (mg/dL)	104.621 ± 18.304	104.457 ± 11.625	0.942
CYBA gene (rs4673)			0.625
CC	60 (74.1%)	145 (78.0%)	
CT	19 (23.5%)	39 (21.0%)	
TT	2 (2.5%)	2 (1.1%)	
GPX1 gene (rs1050450)			0.157
CC	75 (92.6%)	161 (86.6%)	
CT	6 (7.4%)	25 (13.4%)	

* $p < 0.05$, ** $p < 0.01$. p values were calculated by independent t test for continuous variables and chi-square test for categorical variables. Types of CYBA (rs4673) and GPX1 (rs1050450) in both smokers and nonsmokers groups were consistent with Hardy–Weinberg equilibrium. BMI = body mass index; ALT = alanine transaminase, AC sugar = fasting sugar; TBARS = thiobarbituric acid reactive substances; OxLDL = oxidative low-density lipoprotein; LDL = low-density lipoprotein; CYBA gene = cytochrome B-245 Alpha Chain gene; GPX1 gene = Glutathione peroxidase 1 gene.

Table 2 shows the descriptive characteristics among the different types of the CYBA rs4673 and GPx-1 rs1050450 SNPs. Rs4673 have three genotypes (CC, CT, and TT). The TT genotype has the highest blood level and the lowest oxidative stress markers, however not statistically significant. Analysis of GPx-1 rs1050450 SNP showed a statistically significant difference in blood Pb that workers with CC genotypes higher than those with CT genotypes. A comparative analysis of TBARS and OxLDL found no significant difference between CC and CT genotypes.

Table 2. Descriptive characteristics among the different types of CYBA rs4673 and GPx-1 rs1050450 SNPs.

Variables	CYBA rs4673			p Value	GPx-1 rs1050450		p Value
	CC (n = 205)	CT (n = 58)	TT (n = 4)		CC (n = 236)	CT (n = 31)	
Blood Pb (µg/dL)	11.82 ± 13.74	13.09 ± 12.75	23.04 ± 22.31	0.234	12.87 ± 14.03	7.58 ± 9.61	0.009 *
Age (years)	42.1 ± 10.1	41.6 ± 11.2	35.6 ± 4.4	0.477	42.0 ± 10.3	40.9 ± 9.8	0.556
Sex (male)	148 (72.2%)	41 (70.7%)	4 (100%)	0.448	171 (72.5%)	22 (71.0%)	0.862
Current smoking	60 (29.3%)	19 (32.8%)	2 (50%)	0.605	75 (31.8%)	6 (19.4%)	0.157
Alcohol (>3 times/week)	32 (16.9%)	9 (17.3%)	0 (0%)	0.734	38 (17.6%)	3 (10.7%)	0.36
BMI (kg/m ²)	24.43 ± 3.72	24.48 ± 3.91	24.48 ± 2.55	0.875	24.51 ± 3.81	23.83 ± 3.14	0.343
BMI < 18.5	8 (3.9%)	2 (3.4%)	0 (0%)	0.912	9 (3.8%)	1 (3.2%)	0.871
BMI ≥ 24	106 (51.7%)	32 (55.1%)	1 (25%)	0.494	123 (52.1%)	16 (51.6%)	0.958
Creatinine (mg/dL)	1.18 ± 0.60	1.15 ± 0.18	1.17 ± 0.53	0.954	1.18 ± 0.59	1.10 ± 0.17	0.438
ALT (U/L)	19.7 ± 15.1	20.1 ± 18.3	11.0 ± 6.3	0.624	19.9 ± 16.2	17.8 ± 11.7	0.501
AC sugar (mg/dL)	98.9 ± 25.1	107.3 ± 49.0	90.2 ± 5.4	0.192	101.3 ± 33.3	94.8 ± 14.0	0.308
TBARS(nmol/mL)	2.06 ± 0.46	2.06 ± 0.41	1.72 ± 0.27	0.332	2.053 ± 0.464	2.078 ± 0.344	0.776
OxLDL (mg/dL)	55.98 ± 13.02	55.04 ± 11.26	45.87 ± 10.74	0.265	55.64 ± 12.92	55.50 ± 10.53	0.955
LDL (mg/dL)	105.31 ± 13.53	102.54 ± 14.89	93.07 ± 16.5	0.105	104.00 ± 14.10	108.39 ± 12.23	0.105

* $p < 0.05$. p values were calculated by independent t test for continuous variables and chi-square test for categorical variables.

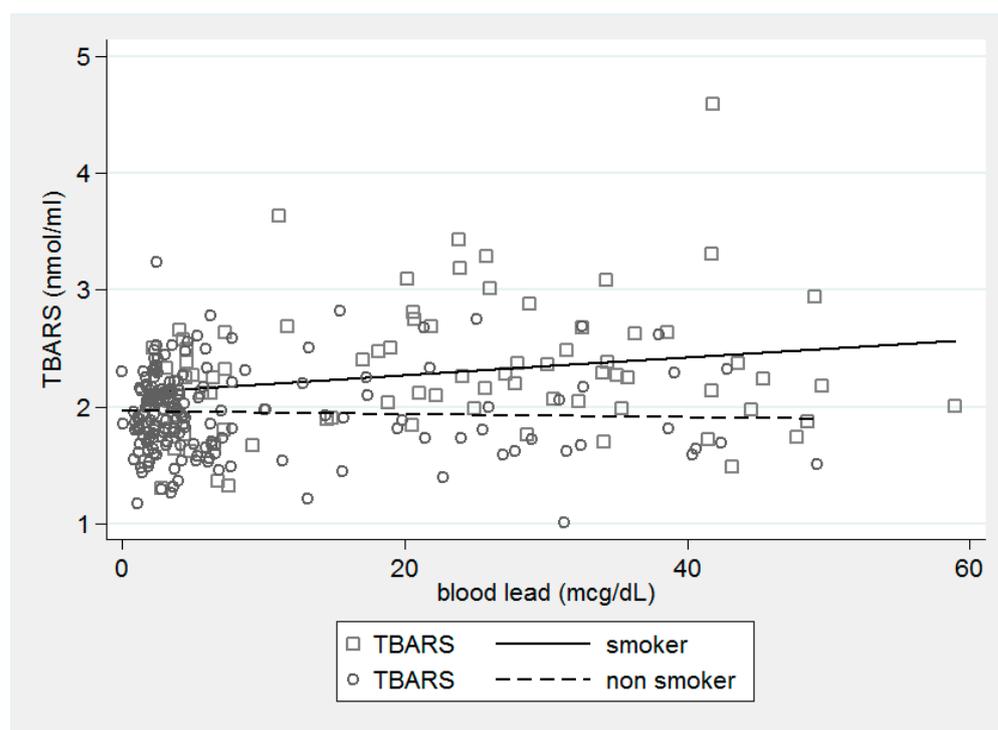
In Table 3, we present results of multiple linear regression using the markers of oxidation (TBARS and OxLDL/LDL) as dependent variables on SNPs and blood Pb after adjustment for age, sex, BMI < 18.5, BMI ≥ 24 versus 18.5 ≤ BMI < 24, Cr > 1.5 versus Cr ≤ 1.5, ALT > 80 versus ALT ≤ 80, AC sugar ≥ 126 versus AC < 126. We explored the interactive relationship of blood Pb, different SNPs, and smoking status from Models 1–5, which showed a significant positive interaction between blood Pb and smoke when TBARS and OxLDL/LDL were used as dependent variables. In addition, blood Pb alone is a positive variable in Models 2, 3, and 4 when OxLDL/LDL was the dependent variable.

Table 3. Multiple linear regression coefficients of TBARS and OxLDL/LDL in different interaction models.

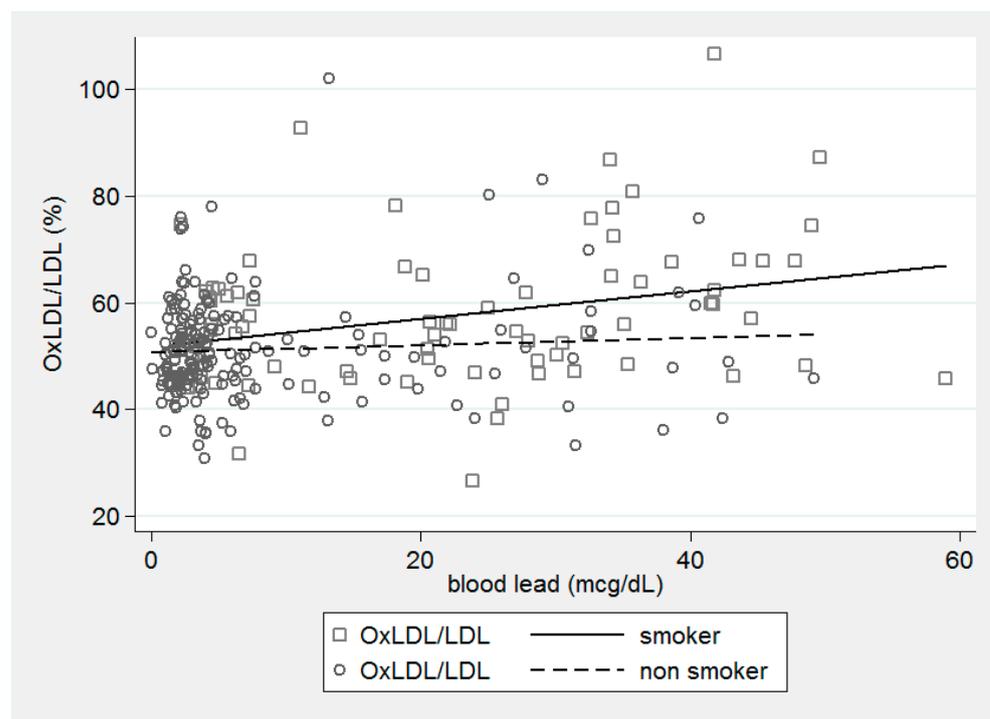
Models	TBARS β (SE)	OxLDL/LDL β (SE)
Model 1		
Blood Pb	−0.001 (0.003)	0.078 (0.08)
smoking	0.115 (0.101)	−0.704 (2.447)
Blood Pb x smoke	0.009 (0.005) *	0.223 (0.11) *
Model 2		
Blood Pb	0.004 (0.003)	0.193 (0.062) *
rs4673	0.003 (0.092)	0.336 (2.219)
Blood Pb x rs4673	−0.003 (0.005)	0 (0.114)
Model 3		
Blood Pb	0.004 (0.002)	0.212 (0.058) **
rs1050450	0.137 (0.114)	2.054 (2.746)
Blood Pb x rs1050450	−0.007 (0.009)	−0.317 (0.205)
Model 4		
Blood Pb	0.004 (0.002)	0.191 (0.057) **
rs4673	−0.055 (0.069)	0.115 (1.67)
rs1050450	0.051 (0.104)	−1.278 (2.514)
rs4673 x rs1050450	0.1 (0.193)	2.037 (4.66)
Model 5		
Blood Pb	0.004 (0.002)	0.192 (0.057) **
rs4673	−0.038 (0.067)	0.433 (1.619)
rs1050450	0.09 (0.098)	−0.559 (2.352)
Blood Pb x rs4673 x rs1050450	−0.003 (0.012)	−0.038 (0.285)

* $p < 0.05$, ** $p < 0.01$, β (SE): regression coefficient and standard error. All models were adjusted for age, sex, BMI < 18.5, BMI ≥ 24 versus 18.5 ≤ BMI < 24, Cr > 1.5 versus Cr ≤ 1.5, ALT > 80 versus ALT ≤ 80, AC sugar ≥ 126 versus AC < 126.

In Figure 1a, TBARS increased 0.009 nmol/mL when blood Pb increased one mcg/dL in smokers compared to nonsmokers. In Figure 1b, the ratio of OxLDL/LDL increased 0.223 when blood Pb increased one mcg/dL in smokers compared to nonsmokers.



(a)



(b)

Figure 1. The interaction effect of smoking on the levels of TBARS and ratio of OxLDL/LDL in Pb-exposed workers. (a) The difference between the levels of TBARS was greater for smokers than nonsmokers in Pb-exposed workers. ($p = 0.041$). (b) The difference between the ratio of OxLDL/LDL was greater for smokers than for nonsmokers. ($p = 0.044$).

4. Discussion

The major findings of this study are that smoking combined with Pb exposure had a synergistic effect that could increase serum levels of TBARS and the ratio of OxLDL/LDL after adjustment for age, sex, abnormal BMI, ALT, creatinine, and AC sugar. The previous animal studies reported an increase of serum TBARS level in Pb-exposed rats [52,53]. Meanwhile, TBARS and oxLDL levels had a positive correlation with cigarette smoking [42,54,55]. To our knowledge, there were few studies investigating the relationship between TBARS or OxLDL/LDL and the interaction between Pb and smoke. We discovered that the interaction between Pb and smoke significantly increased TBARS levels and the ratio of OxLDL/LDL, which suggested that smoking cessation is an important issue in the Pb-exposed working environment.

A study revealed nonsmoking Type II diabetic patients with macrovascular disease had a higher OxLDL/LDL ratio, with 30% of the study participants in the upper tertiles of the OxLDL/LDL ratio having a significantly higher incidence of macrovascular diseases than those in the lower tertiles of the OxLDL/LDL ratio (18% incidence of macrovascular diseases) [31]. Jeffrey et al. found that the risk of developing coronary heart disease (CHD) was significantly higher in the highest quartile of the ratio of OxLDL/LDL than the other three quartiles in male diabetic patients. The percentage of CHD in men in the upper quartiles was 35.1%, and only 18.4% in the lower quartiles [51]. It seems that the ratio of OxLDL/LDL could be a useful indicator for evaluating the development of macrovascular disease among diabetic patients. We suggest future research with different methods, and with different criteria of selection of the study participants as the present study results could be an initial guide for exploration of the relationship between workers exposed to Pb and coronary heart disease.

Smokers exposed to Pb have higher TBARS levels in our study, which led us to assess the prevalence of cardiovascular diseases (CVD) in our group as the previous study revealed an increased risk of CVD with higher TBARS levels.

GPX1 is a selenoprotein containing two exons within a 1.42-kb region located at chromosome 3p21 with antioxidant and anti-inflammatory functions [22,23]. A transition of C to T allele of the GPX1 gene (rs1050450) means the change of amino acid from proline (Pro) to leucine (Leu) at codon 198 (Pro198Leu) [56]. The T allele in the rs1050450 locus was shown to have less antioxidant capacity than the wild-type allele C of the rs1050450 locus because of less selenium-enhanced GPX1 expression [57,58]. The previous study has shown that T allele frequency in the Asian population is less than 0.2. This is consistent with our study, which showed 0.116 [59]. Other research has revealed the incidence of T allele as between 0.41 and 0.58 in the non-Asian population, while Western Europe has a lower frequency of T allele than North America and Northern Europe [60–67]. Our study did not reveal a significant effect of rs1050450 SNPs on oxidative stress markers, TBARS, or on the ratio of OxLDL/LDL, which may be due to lack of TT genotype.

The C242T polymorphism is located in exon 4 at position 214 from the ATG codon [17]. The C242T polymorphism nucleotide transition encodes a CAC→TAC codon change, thus resulting in a non-conservative substitution of Histidine-72 with tyrosine, an alteration that may modify the heme-binding site for the stability of the CYBA gene protein [68]. This replacement is expected to reduce oxidative function and to decrease the production of ROS and oxidative stress in the vasculature [69]. A previous study revealed the relationship between rs4673 polymorphism and TBARS, which showed no significant difference between the genotypes [70]. Our study showed the same results. However, the study population of that study was confined to type 2 diabetic patients, whereas our study population was focused on Pb-exposed workers without confinement to type 2 diabetes. A broader population could be studied for further analysis and we could also use different oxidative stress markers that would better stand for a standard method of oxidative stress *in vivo* in the future. Takanari et al. used OxLDL in addition to TBARS for evaluation of different genotypes of rs4673. The multiple regression analysis showed slightly significantly higher concentrations of OxLDL in those with T allele, which was

unexpected. In our study, considering the total concentrations of serum lipid status, we used the ratio of OxLDL to LDL for correction, which showed insignificant results between rs4673 SNPs and OxLDL/LDL. The influence of rs4673 might be covered by smoking status and BMI, both of which are strong oxidative resources.

The amendment to the Tobacco Hazards Prevention Act in 2009 showed smoking prevalence among men aged 18 and older in Taiwan 33.5% [71,72]. The smoking prevalence of our study is similar to the general population in Taiwan. The prevalence of overweight participants in our study is more than two-folds higher than in the general population, which was 23.9% according to Taiwan's DOH criteria in 2001 [50]. Blood Pb level is the highest, while, inversely, TBARS and OxLDL levels are the lowest in rs4673 TT polymorphism, though the numbers are not statistically significant. Data on larger populations needs to be collected for further investigation of our results. US research presented that smokers had higher blood Pb compared to nonsmokers because tobacco contained Pb [73], which is consistent with our study.

The limitation of our study is that not all variants of the GPX1 gene and CYBA gene were analyzed in this study. Other functional SNPs may interact with each other, causing different outcomes. On the other hand, we only recorded the current smoking status in this study without details about the number of packs per year. The measurement of packs per year in a future study would help us identify the dose–response relationship of quantification of smoking and oxidative stress, which would provide the precise cut-off value for initiation of smoking cessation in Pb-exposed workers. In addition, the majority of our participants were male which made it difficult to look at sex effects and also that lead levels varied between smoking and nonsmoking groups. However, this limitation wildly appeared in the research consistent of heavy workers, and might not influence our conclusion.

5. Conclusions

TBARS levels and the ratio of OxLDL/LDL were positively correlated with the interaction between blood Pb and smoking after the adjustment of confounders, suggesting that smoking cessation is an important issue in the Pb-exposed working environment.

Author Contributions: Conceptualization, K.-J.H. and H.-Y.C.; methodology, H.-Y.C.; validation, K.-J.H., Y.-C.C. and H.-Y.C.; formal analysis, K.-J.H.; investigation, T.-H.C.; resources, H.-Y.C.; data curation, C.-C.Y. and Y.-C.C.; writing—original draft preparation, K.-J.H.; writing—review and editing, H.-Y.C.; visualization, Y.-C.C.; supervision, H.-Y.C.; project administration, Y.-C.C.; funding acquisition, H.-Y.C. All authors have read and agreed to the published version of the manuscript.

Funding: This research was supported by Ministry of Science and Technology, grant number MOST108-2314-B-037-062, and Kaohsiung Medical University Hospital (KMUH109-9T05), and Kaohsiung Medical University (KMU-TC109A01-1).

Institutional Review Board Statement: The study was conducted according to the guidelines of the Declaration of Helsinki, and approved by the Institutional Review Board of Kaohsiung Medical University Hospital (KMUHIRB-E(I)-20190034).

Informed Consent Statement: Individual explaining with consent form was done for each subject.

Acknowledgments: We thank the workers and employers for their cooperation. This work was supported partially by the Research Center for Environmental Medicine, Kaohsiung Medical University, Kaohsiung City, Taiwan from The Featured Areas Research Center Program within the framework of the Higher Education Sprout Project by the Ministry of Education (MOE) in Taiwan and by Kaohsiung Medical University Research Center Grant (KMU-TC109A01-1).

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Kianoush, S.; Balali-Mood, M.; Mousavi, S.R.; Shakeri, M.T.; Dadpour, B.; Moradi, V.; Sadeghi, M. Clinical, toxicological, biochemical, and hematologic parameters in lead exposed workers of a car battery industry. *Iran. J. Med. Sci.* **2013**, *38*, 30–37.
2. Rocha, A.; Trujillo, K.A. Neurotoxicity of low-level lead exposure: History, mechanisms of action, and behavioral effects in humans and preclinical models. *Neurotoxicology* **2019**, *73*, 58–80. [[CrossRef](#)]
3. Harari, F.; Sallsten, G.; Christensson, A.; Petkovic, M.; Hedblad, B.; Forsgard, N.; Melander, O.; Nilsson, P.M.; Borné, Y.; Engström, G.; et al. Blood Lead Levels and Decreased Kidney Function in a Population-Based Cohort. *Am. J. Kidney Dis.* **2018**, *72*, 381–389. [[CrossRef](#)]
4. Obeng-Gyasi, E.; Obeng-Gyasi, B. Chronic Stress and Cardiovascular Disease among Individuals Exposed to Lead: A Pilot Study. *Diseases* **2020**, *8*, 7. [[CrossRef](#)]
5. Gurer, H.; Ercal, N. Can antioxidants be beneficial in the treatment of lead poisoning? *Free Radic. Biol. Med.* **2000**, *29*, 927–945. [[CrossRef](#)]
6. Farmand, F.; Ehdaie, A.; Roberts, C.K.; Sindhu, R.K. Lead-induced dysregulation of superoxide dismutases, catalase, glutathione peroxidase, and guanylate cyclase. *Environ. Res.* **2005**, *98*, 33–39. [[CrossRef](#)]
7. Pech-Amsellem, M.A.; Myara, I.; Storogenko, M.; Demuth, K.; Proust, A.; Moatti, N. Enhanced modifications of low-density lipoproteins (LDL) by endothelial cells from smokers: A possible mechanism of smoking-related atherosclerosis. *Cardiovasc. Res.* **1996**, *31*, 975–983. [[CrossRef](#)]
8. Flora, S.J. Arsenic-induced oxidative stress and its reversibility. *Free Radic. Biol. Med.* **2011**, *51*, 257–281. [[CrossRef](#)]
9. Ushio-Fukai, M.; Zafari, A.M.; Fukui, T.; Ishizaka, N.; Griending, K.K. p22phox is a critical component of the superoxide-generating NADH/NADPH oxidase system and regulates angiotensin II-induced hypertrophy in vascular smooth muscle cells. *J. Biol. Chem.* **1996**, *271*, 23317–23321. [[CrossRef](#)]
10. Inoue, N.; Kawashima, S.; Kanazawa, K.; Yamada, S.; Akita, H.; Yokoyama, M. Polymorphism of the NADH/NADPH oxidase p22 phox gene in patients with coronary artery disease. *Circulation* **1998**, *97*, 135–137. [[CrossRef](#)]
11. Fortunato, A.; San Jose, G.; Moreno, M.U.; Beloqui, O.; Diez, J.; Zalba, G. Phagocytic NADPH oxidase overactivity underlies oxidative stress in metabolic syndrome. *Diabetes* **2006**, *55*, 209–215. [[CrossRef](#)]
12. Fortunato, A.; Olivan, S.; Beloqui, O.; San Jose, G.; Moreno, M.U.; Diez, J.; Zalba, G. Association of increased phagocytic NADPH oxidase-dependent superoxide production with diminished nitric oxide generation in essential hypertension. *J. Hypertens.* **2004**, *22*, 2169–2175. [[CrossRef](#)]
13. Guzik, T.J.; Mussa, S.; Gastaldi, D.; Sadowski, J.; Ratnatunga, C.; Pillai, R.; Channon, K.M. Mechanisms of increased vascular superoxide production in human diabetes mellitus: Role of NAD(P)H oxidase and endothelial nitric oxide synthase. *Circulation* **2002**, *105*, 1656–1662. [[CrossRef](#)]
14. Zalba, G.; Fortuño, A.; Díez, J. Oxidative stress and atherosclerosis in early chronic kidney disease. *Nephrol. Dial. Transplant.* **2006**, *21*, 2686–2690. [[CrossRef](#)]
15. Sorescu, D.; Weiss, D.; Lassegue, B.; Clempus, R.E.; Szocs, K.; Sorescu, G.P.; Valppu, L.; Quinn, M.T.; Lambeth, J.D.; Vega, J.D.; et al. Superoxide production and expression of nox family proteins in human atherosclerosis. *Circulation* **2002**, *105*, 1429–1435. [[CrossRef](#)] [[PubMed](#)]
16. Miller, A.A.; Drummond, G.R.; Sobey, C.G. Novel isoforms of NADPH-oxidase in cerebral vascular control. *Pharmacol. Ther.* **2006**, *111*, 928–948. [[CrossRef](#)] [[PubMed](#)]
17. Dinauer, M.C.; Pierce, E.A.; Bruns, G.A.; Curnutte, J.T.; Orkin, S.H. Human neutrophil cytochrome b light chain (p22-phox). Gene structure, chromosomal location, and mutations in cytochrome-negative autosomal recessive chronic granulomatous disease. *J. Clin. Invest.* **1990**, *86*, 1729–1737. [[CrossRef](#)]
18. Matsunaga-Irie, S.; Maruyama, T.; Yamamoto, Y.; Motohashi, Y.; Hirose, H.; Shimada, A.; Murata, M.; Saruta, T. Relation between development of nephropathy and the p22phox C242T and receptor for advanced glycation end product G1704T gene polymorphisms in type 2 diabetic patients. *Diabetes Care* **2004**, *27*, 303–307. [[CrossRef](#)] [[PubMed](#)]
19. Wu, Z.; Lou, Y.; Jin, W.; Liu, Y.; Lu, L.; Chen, Q.; Xie, Y.; Lu, G. Relationship of the p22phox (CYBA) gene polymorphism C242T with risk of coronary artery disease: A meta-analysis. *PLoS ONE* **2013**, *8*, e70885. [[CrossRef](#)]
20. Men, T.; Zhang, X.; Yang, J.; Shen, B.; Li, X.; Chen, D.; Wang, J. The rs1050450 C > T polymorphism of GPX1 is associated with the risk of bladder but not prostate cancer: Evidence from a meta-analysis. *Tumor Biol.* **2014**, *35*, 269–275. [[CrossRef](#)]
21. Forgione, M.A.; Weiss, N.; Heydrick, S.; Cap, A.; Klings, E.S.; Bierl, C.; Eberhardt, R.T.; Farber, H.W.; Loscalzo, J. Cellular glutathione peroxidase deficiency and endothelial dysfunction. *Am. J. Physiol. Circ. Physiol.* **2002**, *282*, H1255–H1261. [[CrossRef](#)]
22. Hauser, F.; Rossmann, H.; Laubert-Reh, D.; Wild, P.S.; Zeller, T.; Muller, C.; Neuwirth, S.; Blankenberg, S.; Lackner, K.J. Inflammatory bowel disease (IBD) locus 12: Is glutathione peroxidase-1 (GPX1) the relevant gene? *Genes Immun.* **2015**, *16*, 571–575. [[CrossRef](#)]
23. Arthur, J.R. The glutathione peroxidases. *Cell. Mol. Life Sci.* **2001**, *57*, 1825–1835. [[CrossRef](#)]
24. Hamanishi, T.; Furuta, H.; Kato, H.; Doi, A.; Tamai, M.; Shimomura, H.; Sakagashira, S.; Nishi, M.; Sasaki, H.; Sanke, T.; et al. Functional variants in the glutathione peroxidase-1 (GPX-1) gene are associated with increased intima-media thickness of carotid arteries and risk of macrovascular diseases in Japanese type 2 diabetic patients. *Diabetes* **2004**, *53*, 2455–2460. [[CrossRef](#)]

25. Forgiione, M.A.; Cap, A.; Liao, R.; Moldovan, N.I.; Eberhardt, R.T.; Lim, C.C.; Jones, J.; Goldschmidt-Clermont, P.J.; Loscalzo, J. Heterozygous cellular glutathione peroxidase deficiency in the mouse: Abnormalities in vascular and cardiac function and structure. *Circulation* **2002**, *106*, 1154–1158. [[CrossRef](#)] [[PubMed](#)]
26. Hong, Z.; Tian, C.; Zhang, X. GPX1 gene Pro200Leu polymorphism, erythrocyte GPX activity, and cancer risk. *Mol. Biol. Rep.* **2013**, *40*, 1801–1812. [[CrossRef](#)]
27. Tang, T.S.; Prior, S.L.; Li, K.W.; Ireland, H.A.; Bain, S.C.; Hurel, S.J.; Cooper, J.A.; Humphries, S.E.; Stephens, J.W. Association between the rs1050450 glutathione peroxidase-1 (C > T) gene variant and peripheral neuropathy in two independent samples of subjects with diabetes mellitus. *Nutr. Metab. Cardiovasc. Dis.* **2012**, *22*, 417–425. [[CrossRef](#)] [[PubMed](#)]
28. Tang, N.P.; Wang, L.S.; Yang, L.; Gu, H.J.; Sun, Q.M.; Cong, R.H.; Zhou, B.; Zhu, H.J.; Wang, B. Genetic variant in glutathione peroxidase 1 gene is associated with an increased risk of coronary artery disease in a Chinese population. *Clin. Chim. Acta* **2008**, *395*, 89–93. [[CrossRef](#)]
29. Kuzuya, M.; Ando, F.; Iguchi, A.; Shimokata, H. Glutathione peroxidase 1 Pro198Leu variant contributes to the metabolic syndrome in men in a large Japanese cohort. *Am. J. Clin. Nutr.* **2008**, *87*, 1939–1944. [[CrossRef](#)] [[PubMed](#)]
30. Matsuura, E.; Hughes, G.R.; Khamashta, M.A. Oxidation of LDL and its clinical implication. *Autoimmun. Rev.* **2008**, *7*, 558–566. [[CrossRef](#)] [[PubMed](#)]
31. Girona, J.; Manzanares, J.M.; Marimon, F.; Cabre, A.; Heras, M.; Guardiola, M.; Ribalta, J.; Masana, L. Oxidized to non-oxidized lipoprotein ratios are associated with arteriosclerosis and the metabolic syndrome in diabetic patients. *Nutr. Metab. Cardiovasc. Dis.* **2008**, *18*, 380–387. [[CrossRef](#)] [[PubMed](#)]
32. Parthasarathy, S.; Raghavamenon, A.; Garelnabi, M.O.; Santanam, N. Oxidized low-density lipoprotein. *Methods Mol. Biol.* **2010**, *610*, 403–417. [[CrossRef](#)] [[PubMed](#)]
33. Steinberg, D.; Parthasarathy, S.; Carew, T.E.; Khoo, J.C.; Witztum, J.L. Beyond cholesterol. Modifications of low-density lipoprotein that increase its atherogenicity. *N. Engl. J. Med.* **1989**, *320*, 915–924. [[CrossRef](#)] [[PubMed](#)]
34. Janero, D.R. Malondialdehyde and thiobarbituric acid-reactivity as diagnostic indices of lipid peroxidation and peroxidative tissue injury. *Free Radic. Biol. Med.* **1990**, *9*, 515–540. [[CrossRef](#)]
35. Halliwell, B.; Chirico, S. Lipid peroxidation: Its mechanism, measurement, and significance. *Am. J. Clin. Nutr.* **1993**, *57*, 715S–725S. [[CrossRef](#)] [[PubMed](#)]
36. Ho, E.; Karimi Galougahi, K.; Liu, C.C.; Bhindi, R.; Figtree, G.A. Biological markers of oxidative stress: Applications to cardiovascular research and practice. *Redox Biol.* **2013**, *1*, 483–491. [[CrossRef](#)]
37. Trevisan, M.; Browne, R.; Ram, M.; Muti, P.; Freudenheim, J.; Carosella, A.M.; Armstrong, D. Correlates of markers of oxidative status in the general population. *Am. J. Epidemiol.* **2001**, *154*, 348–356. [[CrossRef](#)] [[PubMed](#)]
38. Schisterman, E.F.; Faraggi, D.; Browne, R.; Freudenheim, J.; Dorn, J.; Muti, P.; Armstrong, D.; Reiser, B.; Trevisan, M. TBARS and cardiovascular disease in a population-based sample. *J. Cardiovasc. Risk* **2001**, *8*, 219–225. [[CrossRef](#)]
39. Schisterman, E.F.; Faraggi, D.; Browne, R.; Freudenheim, J.; Dorn, J.; Muti, P.; Armstrong, D.; Reiser, B.; Trevisan, M. Minimal and best linear combination of oxidative stress and antioxidant biomarkers to discriminate cardiovascular disease. *Nutr. Metab. Cardiovasc. Dis.* **2002**, *12*, 259–266.
40. Seet, R.C.; Lee, C.Y.; Loke, W.M.; Huang, S.H.; Huang, H.; Looi, W.F.; Chew, E.S.; Quek, A.M.; Lim, E.C.; Halliwell, B. Biomarkers of oxidative damage in cigarette smokers: Which biomarkers might reflect acute versus chronic oxidative stress? *Free Radic. Biol. Med.* **2011**, *50*, 1787–1793. [[CrossRef](#)] [[PubMed](#)]
41. Morrow, J.D.; Frei, B.; Longmire, A.W.; Gaziano, J.M.; Lynch, S.M.; Shyr, Y.; Strauss, W.E.; Oates, J.A.; Roberts, L.J., 2nd. Increase in circulating products of lipid peroxidation (F2-isoprostanes) in smokers. Smoking as a cause of oxidative damage. *N. Engl. J. Med.* **1995**, *332*, 1198–1203. [[CrossRef](#)]
42. Lykkesfeldt, J. Malondialdehyde as biomarker of oxidative damage to lipids caused by smoking. *Clin. Chim. Acta* **2007**, *380*, 50–58. [[CrossRef](#)] [[PubMed](#)]
43. Ohkawa, H.; Ohishi, N.; Yagi, K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal. Biochem.* **1979**, *95*, 351–358. [[CrossRef](#)]
44. Ellman, G.; Lysko, H. A precise method for the determination of whole blood and plasma sulfhydryl groups. *Anal. Biochem.* **1979**, *93*, 98–102. [[CrossRef](#)]
45. Aviram, M.; Fuhrman, B. LDL oxidation by arterial wall macrophages depends on the oxidative status in the lipoprotein and in the cells: Role of prooxidants vs. antioxidants. *Mol. Cell. Biochem.* **1998**, *188*, 149–159. [[CrossRef](#)]
46. Pan, W.H.; Flegal, K.M.; Chang, H.Y.; Yeh, W.T.; Yeh, C.J.; Lee, W.C. Body mass index and obesity-related metabolic disorders in Taiwanese and US whites and blacks: Implications for definitions of overweight and obesity for Asians. *Am. J. Clin. Nutr.* **2004**, *79*, 31–39. [[CrossRef](#)] [[PubMed](#)]
47. World Health Organization. Obesity: Preventing and managing the global epidemic. Report of a WHO consultation. *World Health Organ. Tech. Rep. Ser.* **2000**, *894*, 1–253.
48. National Institutes of Health. Clinical Guidelines on the Identification, Evaluation, and Treatment of Overweight and Obesity in Adults—The Evidence Report. *Obes. Res.* **1998**, *6*, 51s–209s.
49. Deurenberg, P.; Yap, M.; van Staveren, W.A. Body mass index and percent body fat: A meta analysis among different ethnic groups. *Int. J. Obes.* **1998**, *22*, 1164–1171. [[CrossRef](#)]
50. Chu, N.F. Prevalence of obesity in Taiwan. *Obes. Rev.* **2005**, *6*, 271–274. [[CrossRef](#)]

51. Stephens, J.W.; Gable, D.R.; Hurel, S.J.; Miller, G.J.; Cooper, J.A.; Humphries, S.E. Increased plasma markers of oxidative stress are associated with coronary heart disease in males with diabetes mellitus and with 10-year risk in a prospective sample of males. *Clin. Chem.* **2006**, *52*, 446–452. [[CrossRef](#)] [[PubMed](#)]
52. Alya, A.; Ines, D.B.; Montassar, L.; Najoua, G.; Saloua, E.F. Oxidative stress, biochemical alterations, and hyperlipidemia in female rats induced by lead chronic toxicity during puberty and post puberty periods. *Iran. J. Basic Med Sci.* **2015**, *18*, 1034–1043. [[PubMed](#)]
53. Agrawal, S.; Flora, G.; Bhatnagar, P.; Flora, S.J. Comparative oxidative stress, metallothionein induction and organ toxicity following chronic exposure to arsenic, lead and mercury in rats. *Cell. Mol. Biol.* **2014**, *60*, 13–21. [[PubMed](#)]
54. Miller, E.R., 3rd; Appel, L.J.; Jiang, L.; Risby, T.H. Association between cigarette smoking and lipid peroxidation in a controlled feeding study. *Circulation* **1997**, *96*, 1097–1101. [[CrossRef](#)] [[PubMed](#)]
55. Panagiotakos, D.B.; Pitsavos, C.; Chrysohoou, C.; Skoumas, J.; Masoura, C.; Toutouzas, P.; Stefanadis, C. Effect of exposure to secondhand smoke on markers of inflammation: The ATTICA study. *Am. J. Med.* **2004**, *116*, 145–150. [[CrossRef](#)]
56. Forsberg, L.; de Faire, U.; Morgenstern, R. Low yield of polymorphisms from EST blast searching: Analysis of genes related to oxidative stress and verification of the P197L polymorphism in GPX1. *Hum. Mutat.* **1999**, *13*, 294–300. [[CrossRef](#)]
57. Ravn-Haren, G.; Olsen, A.; Tjønneland, A.; Dragsted, L.O.; Nexø, B.A.; Wallin, H.; Overvad, K.; Raaschou-Nielsen, O.; Vogel, U. Associations between GPX1 Pro198Leu polymorphism, erythrocyte GPX activity, alcohol consumption and breast cancer risk in a prospective cohort study. *Carcinogenesis* **2006**, *27*, 820–825. [[CrossRef](#)]
58. Hu, Y.J.; Diamond, A.M. Role of glutathione peroxidase 1 in breast cancer: Loss of heterozygosity and allelic differences in the response to selenium. *Cancer Res.* **2003**, *63*, 3347–3351.
59. Ichimura, Y.; Habuchi, T.; Tsuchiya, N.; Wang, L.; Oyama, C.; Sato, K.; Nishiyama, H.; Ogawa, O.; Kato, T. Increased risk of bladder cancer associated with a glutathione peroxidase 1 codon 198 variant. *J. Urol.* **2004**, *172*, 728–732. [[CrossRef](#)]
60. Zarbock, R.; Hendig, D.; Szliska, C.; Kleesiek, K.; Gotting, C. Pseudoxanthoma elasticum: Genetic variations in antioxidant genes are risk factors for early disease onset. *Clin. Chem.* **2007**, *53*, 1734–1740. [[CrossRef](#)]
61. Raaschou-Nielsen, O.; Sorensen, M.; Hansen, R.D.; Frederiksen, K.; Tjønneland, A.; Overvad, K.; Vogel, U. GPX1 Pro198Leu polymorphism, interactions with smoking and alcohol consumption, and risk for lung cancer. *Cancer Lett.* **2007**, *247*, 293–300. [[CrossRef](#)]
62. Mostowska, A.; Hozyasz, K.K.; Lianeri, M.; Piwowar, W.; Jagodzinski, P.P. Polymorphic variants of genes encoding main antioxidant enzymes and the risk of CL/P-affected pregnancies. *Clin. Biochem.* **2007**, *40*, 416–419. [[CrossRef](#)]
63. Choi, J.Y.; Neuhaus, M.L.; Barnett, M.; Hudson, M.; Kristal, A.R.; Thornquist, M.; King, I.B.; Goodman, G.E.; Ambrosone, C.B. Polymorphisms in oxidative stress-related genes are not associated with prostate cancer risk in heavy smokers. *Cancer Epidemiol. Biomark. Prev.* **2007**, *16*, 1115–1120. [[CrossRef](#)]
64. Lightfoot, T.J.; Skibola, C.F.; Smith, A.G.; Forrest, M.S.; Adamson, P.J.; Morgan, G.J.; Bracci, P.M.; Roman, E.; Smith, M.T.; Holly, E.A. Polymorphisms in the oxidative stress genes, superoxide dismutase, glutathione peroxidase and catalase and risk of non-Hodgkin's lymphoma. *Haematol.* **2006**, *91*, 1222–1227.
65. Knight, J.A.; Onay, U.V.; Wells, S.; Li, H.; Shi, E.J.; Andrulis, I.L.; Ozcelik, H. Genetic variants of GPX1 and SOD2 and breast cancer risk at the Ontario site of the Breast Cancer Family Registry. *Cancer Epidemiol. Biomark. Prev.* **2004**, *13*, 146–149. [[CrossRef](#)]
66. Ratnasinghe, D.; Tangrea, J.A.; Andersen, M.R.; Barrett, M.J.; Virtamo, J.; Taylor, P.R.; Albanes, D. Glutathione peroxidase codon 198 polymorphism variant increases lung cancer risk. *Cancer Res.* **2000**, *60*, 6381–6383.
67. Forsberg, L.; de Faire, U.; Marklund, S.L.; Andersson, P.M.; Stegmayr, B.; Morgenstern, R. Phenotype determination of a common Pro-Leu polymorphism in human glutathione peroxidase 1. *Blood Cells Mol. Dis.* **2000**, *26*, 423–426. [[CrossRef](#)] [[PubMed](#)]
68. Griendling, K.K.; Sorescu, D.; Ushio-Fukai, M. NAD(P)H oxidase: Role in cardiovascular biology and disease. *Circ. Res.* **2000**, *86*, 494–501. [[CrossRef](#)] [[PubMed](#)]
69. Whitehead, A.S.; FitzGerald, G.A. Twenty-first century phox: Not yet ready for widespread screening. *Circulation* **2001**, *103*, 7–9. [[CrossRef](#)]
70. Nakano, T.; Matsunaga, S.; Nagata, A.; Maruyama, T. NAD(P)H oxidase p22phox Gene C242T polymorphism and lipoprotein oxidation. *Clin. Chim. Acta* **2003**, *335*, 101–107. [[CrossRef](#)]
71. *Taiwan Tobacco Control Annual Report*; Health Promotion Administration, Ministry of Health and Welfare: Taipei, Taiwan, 2019. Available online: <https://www.hpa.gov.tw/EngPages/Detail.aspx?nodeid=1069&pid=12873> (accessed on 8 May 2021).
72. Tsai, Y.W.; Chang, L.C.; Sung, H.Y.; Hu, T.W.; Chiou, S.T. The impact of smoke-free legislation on reducing exposure to secondhand smoke: Differences across gender and socioeconomic groups. *Tob. Control.* **2015**, *24*, 62–69. [[CrossRef](#)] [[PubMed](#)]
73. Mannino, D.M.; Homa, D.M.; Matte, T.; Hernandez-Avila, M. Active and passive smoking and blood lead levels in U.S. adults: Data from the Third National Health and Nutrition Examination Survey. *Nicotine Tob. Res.* **2005**, *7*, 557–564. [[CrossRef](#)] [[PubMed](#)]