

Review

Effect of Endocrine Disruptor Pesticides: A Review

Wissem Mnif ^{1,2}, Aziza Ibn Hadj Hassine ¹, Aicha Bouaziz ¹, Aghleb Bartegi ³, Olivier Thomas ⁴
and Benoit Roig ^{4,*}

¹ Laboratoire de Biochimie, Unité de Recherche 02/UR/09-01, Institut Supérieur de Biotechnologie, de Monastir, BP 74, 5019 Monastir, Tunisia; E-Mails: w_mnif@yahoo.fr (W.M.); aziza.hadjhassine@gmail.com (A.I.H.H); w_bouaziz.aicha@yahoo.fr (A.B.);

² Institut Supérieur de Biotechnologie de Sidi Thabet, Pole Technologie Sidi Thabet, 2020 Ariana, Tunisia

³ Department of Biology, Faculty of Sciences, King Faisal University, P.O. Box 1759, 31982, Al Hassa, Saudi Arabia; E-Mail: bartagi_fsm@yahoo.com

⁴ Environment and Health Research laboratory (LERES), Advanced School of Public Health (EHESP), Avenue du Professeur Léon Bernard - CS 74312, 35043 Rennes Cedex, France; E-Mail: olivier.thomas@ehesp.fr (O.T.)

* Author to whom correspondence should be addressed; E-Mail: benoit.roig@ehesp.fr; Tel.: +33-786-284-551; Fax: +33-299-022-921.

Received: 20 May 2011; in revised form: 8 June 2011 / Accepted: 9 June 2011 /

Published: 17 June 2011

Abstract: Endocrine disrupting chemicals (EDC) are compounds that alter the normal functioning of the endocrine system of both wildlife and humans. A huge number of chemicals have been identified as endocrine disruptors, among them several pesticides. Pesticides are used to kill unwanted organisms in crops, public areas, homes and gardens, and parasites in medicine. Human are exposed to pesticides due to their occupations or through dietary and environmental exposure (water, soil, air). For several years, there have been enquiries about the impact of environmental factors on the occurrence of human pathologies. This paper reviews the current knowledge of the potential impacts of endocrine disruptor pesticides on human health.

Keywords: endocrine disruptors; pesticides; biomonitoring; human effect

Abbreviations: ADI: Acceptable Daily Intake; AhR: ArylHydrocarbon Receptor; AR: Androgen Receptor; CA: Concentration Addition; CAR: Constitutive Androstane Receptor; EDC: Endocrine Disruptor Chemical; ER: Estrogen Receptor; ERR: Estrogen Related Receptor; HCH: Hexachlorocyclohexane; IA: Independent Action; LOD: Limit of Detection; PCB: PolyChloroBiphenyl; PXR: Pregnane X Receptor; WHO: World Health Organisation

1. Introduction

Since the discovery of DDT in 1939 [1], numerous pesticides (organochlorides, organophosphates, carbamates) have been developed and used extensively worldwide with few guidelines or restrictions. In industrialized countries, the Green Revolution of the 1960s significantly increased agricultural productivity by increasing the cultivated surfaces, mechanization, planting of hybrid crops with higher yields, and pest control [2]. This fight requires the massive use of pesticides, which are hazardous chemicals designed to repel or kill rodents, fungi, insects, and “weeds” that undermine intensive farming. The main effects of pesticides represent a great benefit for human health. Indeed, they help control agricultural pests (including diseases and weeds) and plant disease vectors, human and livestock disease vectors and nuisance organisms, and organisms that harm other human activities and structures (gardens, recreational areas, *etc.*). Moreover, they insure increased food production, a safe and secure food supply, and other secondary benefits [3]. However, many first generation pesticides have been found to be harmful to the environment. Some of them can persist in soils and aquatic sediments, bioconcentrate in the tissues of invertebrates and vertebrates, move up trophic chains, and affect top predators.

Rachel Carson’s book “*Silent Spring*”, published in 1962 [4], first drew attention to the hazard of the widespread extensive use of pesticides for the environment (namely birds) and also for human health. The book resulted in big modifications to the US national policy on pesticides, leading to a national ban on DDT and certain other pesticides.

Worldwide consumption of pesticides for agricultural use is constantly increasing, rising from 0.49 kg/ha in 1961 to 2 kg/ha in 2004 (see various web sources, such as for example http://ec.europa.eu/agriculture/envir/report/fr/pest_fr/report.htm#fig6; <http://faostat.fao.org/site/424/default.aspx#ancor>; [http://www.goodplanet.info/eng/Food-Agriculture/Pesticides/Pesticides/\(theme\)/266](http://www.goodplanet.info/eng/Food-Agriculture/Pesticides/Pesticides/(theme)/266)), and humans and wildlife are today continuously exposed to a number of pesticides via the environment (surface water, ground water, soil), food and drinking water [5].

The World Health Organization (WHO) has reported that roughly three million pesticide poisonings occur annually, resulting in 220,000 deaths worldwide [6]. In some cases, it has been suggested that diseases such as cancer, allergies, neurological disorders and reproductive disorders may be connected to pesticide exposure.

This article focuses on pesticides that act as endocrine disruptors, and reviews the available information about the exposure and effects of such pesticides, as well as human biomonitoring for human health risk assessment.

2. Effects of Endocrine Disruptor Pesticides

Many chemicals that have been identified as endocrine disruptors are pesticides [7-11]. About 105 substances can be listed, and most of them are shown in Table 1. Of these, 46% are insecticides, 21% herbicides and 31% fungicides; some of them were withdrawn from general use many years ago but are still found in the environment (ex. DDT and atrazine in several countries).

EDCs act mainly by interfering with natural hormones because of their strong potential to bind to estrogen or androgen receptors [12] as shown in Table 1. In particular, EDCs can bind to and activate various hormone receptors (AR, ER, AhR, PXR, CAR, ERR) and then mimic the natural hormone's action (agonist action). EDCs may also bind to these receptors without activating them. This antagonist action blocks the receptors and inhibits their action. Finally, EDCs may also interfere with the synthesis, transport, metabolism and elimination of hormones, thereby decreasing the concentration of natural hormones. For example, thyroid hormone production can be inhibited by some ten endocrine disruptor pesticides (amitrole, cyhalothrin, fipronil, ioxynil, maneb, mancozeb, pentachloronitrobenzene, prodiamine, pyrimethanil, thiazopyr, ziram, zineb, not shown in Table 1) [13-16].

At the environmental level, wildlife is particularly vulnerable to the endocrine disrupting effects of pesticides. Effects linked to endocrine disruption have been largely noted in invertebrates [17-21], reptiles [22-27], fish [28,29], birds [30-34] and mammals [35-38] as reviewed by Mnif *et al.* [39]. Most of them are linked to exposure to organochlorine pesticides (OC) and affect the reproductive function. For example, a study on *Daphnia magna* has shown that endosulfan sulphate disrupts the ecdysteroidal system (regulating processes such as molting and embryonic development) and juvenile hormone activity (regulating the sex ratio) of crustaceans [40,41]. Another example is the influence of linuron on reproductive hormone production [42], testosterone production in rats being significantly reduced after in utero exposure to linuron, whereas progesterone production was not affected [42].

At the human level, endocrine disruptor pesticides have also been shown to disrupt reproductive and sexual development, and these effects seem to depend on several factors, including gender, age, diet, and occupation.

Age is a particularly sensitive factor. Human fetuses, infants and children show greater susceptibility than adults [43-45]. Much of the damage caused by EDC occurs during gametogenesis and the early development of the fetus [45-48]. However, the effects may not become apparent until adulthood. Moreover, fetuses and infants receive greater doses due to the mobilization of maternal fat reserves during pregnancy [47-50] and breastfeeding [49,51]. Infants are extremely vulnerable to pre and postnatal exposure to endocrine disruptor pesticides, resulting in a wide range of adverse health effects including possible long-term impacts on intellectual function [52,53] and delayed effects on the central nervous system functioning [54,55].

Likewise, residential proximity to agricultural activity is a factor often described to explain developmental abnormalities in epidemiological studies of low birth weight [56], fetal death [57], and childhood cancers [58]. Additionally, a higher prevalence of cryptorchidism and hypospadias [59,60] was found in areas with extensive farming and pesticide use and in sons of women working as gardeners [61]. Recently, a relation has been reported between cryptorchidism and persistent pesticide concentration in maternal breast milk [47,62,63]. The impact of endocrine disruptor pesticides on fertility has also been discussed [64].

Table 1. Effects of different groups of endocrine disruptor pesticides and their chemical structures (adapted from [65]).

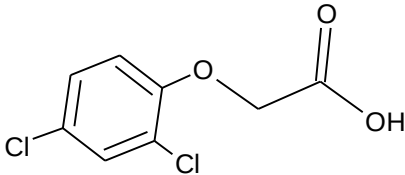
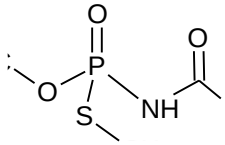
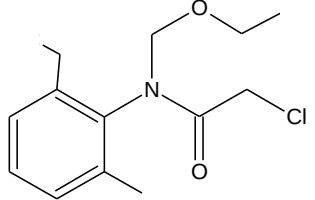
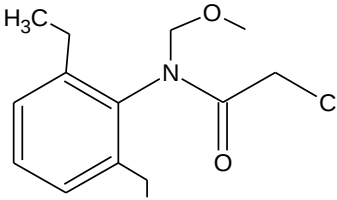
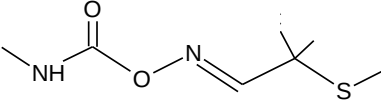
Pesticides [66]	Chemical structure	Endocrine Disruptor Effects	Biomonitoring in human samples
2,4-D (H) M(g/Mol) = 221 pKa = 2.73 logP: 2.81		Synergistic androgenic effects when combined with testosterone [67]	U: <LOD–598 ng/mL [68] S : 0.07–0.56 µg/g creatinine [69]
Acephate (I) M(g/Mol) = 183.2 pKa = 8.35 logP: –0.85		Disruption of hormone expression in the hypothalamus [70]	U: <LOD–0.26 ng/mL [68] H.S: 7.2 µg/mL [71] **
Acetochlor (H) M(g/Mol) = 269.8 pKa = n.a logP: 4.14		Interaction with uterine estrogen receptors, alteration of thyroid hormone dependant gene expression [72,73]	U: <LOD–10.9 ng/mL [68] S: 0.08–0.10 µg/g creatinine [69]
Alachlor (H) M(g/Mol) = 269.8 pKa = 0.62 logP: 3.09		Competitive binding to estrogen and progesterone receptors. Interaction with the pregnane X cellular receptor, interfering with the production of enzymes responsible for steroid hormone metabolism [13,74]	U: <LOD–305 ng/mL [68] S: 0.31–0.72 µg/g creatinine [69]
Aldicarb (I) M(g/Mol) = 190.3 pKa = n.a logP: 1.15		Inhibition of 17 beta-estradiol and progesterone activity [13,75]	

Table 1. Cont.

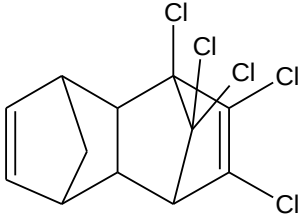
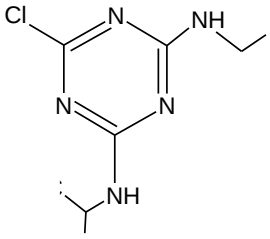
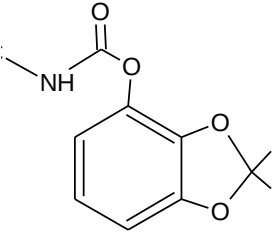
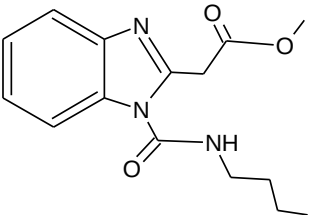
<u>Pesticides</u>	<u>Chemical structure</u>	Endocrine Disruptor <u>Effects</u>	<u>Biomonitoring in human samples</u>
Aldrin (I) M(g/Mol) = 364.9 pKa = n.a logP: 6.5		Competitive binding to androgen receptors [76]	H.S: 2.17–372 µg/L [77,78] H.M: mean 0.03 mg/L ± 0.03 [79] A.T: 25.6–137.2 ng/g lipid [78]
Atrazine (H) M(g/Mol) = 215.7 PKa = 4.14, 10.7 logP: −0.97		Androgen inhibition, weak estrogenic effect. Disruption of the hypothalamic control of lutenising hormone and prolactin levels. Induction of aromatase activity, increase estrogen production. Adrenal glands damages and reduction of steroid hormone metabolism [13,80-83]	U: <LOD–9.2 ng/mL [68,84] H.S: mean 2 pg/g [76,85] S: 0.07–0.17 µg/g creatinine [69]
Bendiocarb (I) M(g/Mol) = 223.2 pKa = 8.8 logP: 1.7		Weak estrogen effect [13]	
Benomyl (F) M(g/Mol) = 290.3 pKa = 4.48 logP: 1.4		Increase of estrogen production and aromatase activity [86]	

Table 1. Cont.

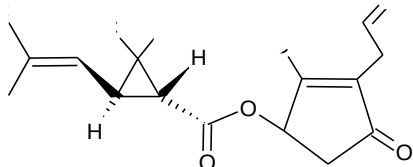
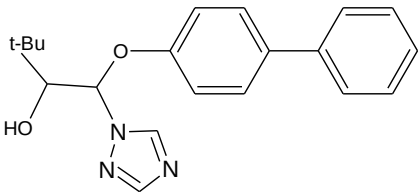
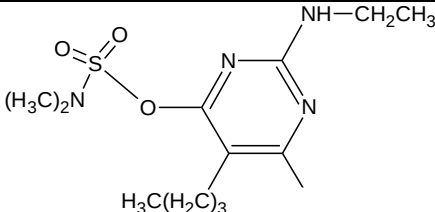
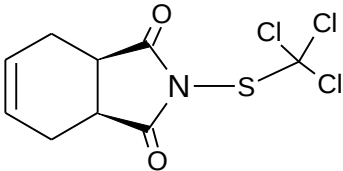
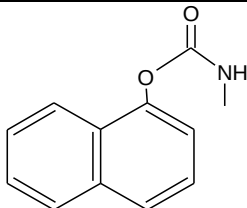
Pesticides	Chemical structure	Endocrine Disruptor Effects	Biomonitoring in human samples
Bioallethrin (I) M(g/Mol) = 302.4 pKa = n.a logP: 4.68		Inhibition of estrogen-sensitive cells proliferation [87]	M: 0.61–1.79 µg/mL [88] H: 1.08–2.74 µg/mL [88]
Bitertanol (F) M(g/Mol) = 337.4 pKa = n.a logP: 4.1		Inhibition of aromatase activity, decrease of estrogens production and increase of androgens availability [89]	
Bupirimate (F) M(g/Mol) = 316.4 pKa = 4.4 logP: 3.68		Activation of Pregnane X cellular receptor [11]	
Captan (F) M(g/Mol) = 300.6 pKa = n.a logP: 2.5		Inhibition of estrogen action [90]	
Carbaryl (I) M(g/Mol) = 201.2 pKa = 10.4 logP: 2.36		Weak estrogen effect [13]	

Table 1. Cont.

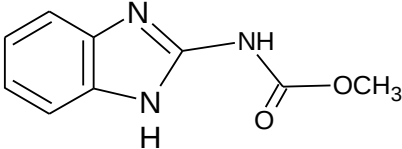
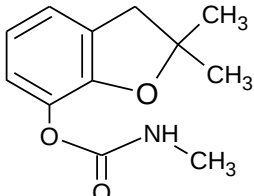
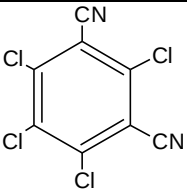
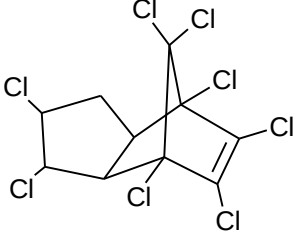
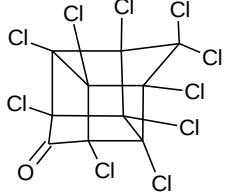
Pesticides	Chemical structure	Endocrine Disruptor Effects	Biomonitoring in human samples
Carbendazim (F) M(g/Mol) = 191.2 pKa = 4.2 logP: 1.48		Increase of estrogen production and aromatase activity [86]	
Carbofuran (I) M(g/Mol) = 221.2 pKa = n.a logP: 1.8		Increase of progesterone, cortisol and estradiol level and decrease of testosterone one [91]	M.S: 0.007–17.63 ng/g [92] U.C: 0.007–13.97 ng/g [92]
Chlorothalonil (F) M(g/Mol) = 265.9 pKa = n.a logP: 2.94		Activation of androgen-sensitive cells proliferation [93]	M.S: 0.007–25.31 ng/g [92] U.C: 0.007–25.12 ng/g [92] H.S: mean 6 pg/g [85]
Chlordane (I) M(g/Mol) = 409.8 pKa = n.a logP: 2.78		Competitive binding to androgen receptors [76] Anti-estrogenic effect, inhibition of estradiol binding [13]	M.P: 0–2.7 ng/g lipid [94] B.S: <LOD–0.9 ng/g lipid [95] H.M: 0.02–437 ng/g lipid [79,96-99] FF: 0.1–0.3 ng/L [100]
Chlordecone (I) M(g/Mol) = 490.6 pKa = n.a logP: 4.5		Binding to estrogen and androgen receptors [90,101,102]	

Table 1. Cont.

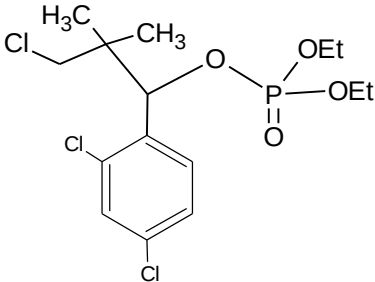
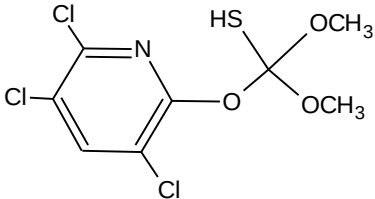
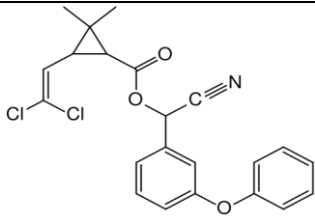
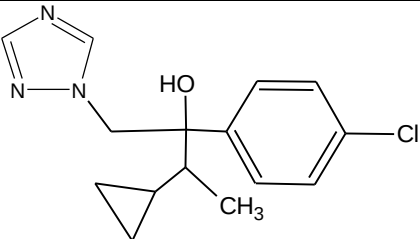
Pesticides	Chemical structure	Endocrine Disruptor Effects	Biomonitoring in human samples
Chlorfenviphos (I) M(g/Mol) = 359.6 pKa = n.a logP: 1.36		Weak estrogen effect [103]	
Chlorpyrifos methyl (I) M(g/Mol) = 322.5 pKa = n.a logP: 4		Antagonist to androgen activity [104]	U : <LOD–57.7 ng/mL* [68,84] M.S: 0.0007–10.1 ng/g [92] U.C: 0.0007–1.84 ng/g [92] H.S: mean 9 pg/g [85] H: 1.77–2.16 1.83 µg/mL [88]
Cypermethrin (I) M(g/Mol) = 416.3 pKa = n.a logP: 5.3		Estrogenic effect [105,106]	U: 0.5–100.4 µg/g * [107] M: 1.85–2.43 µg/mL [88]
Cyproconazole (F) M(g/Mol) = 291.8 pKa = n.a logP:3.09		Inhibition of aromatase activity, decrease of estrogens production and increase of androgens availability [89]	

Table 1. Cont.

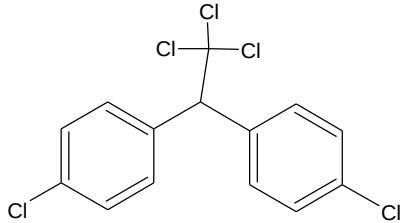
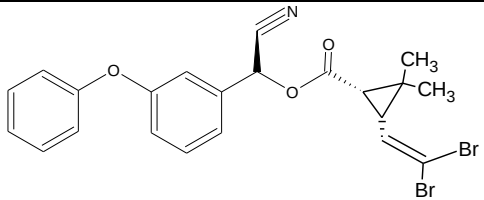
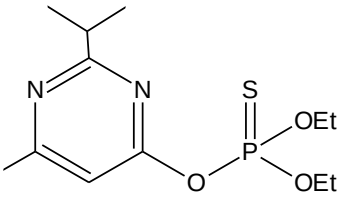
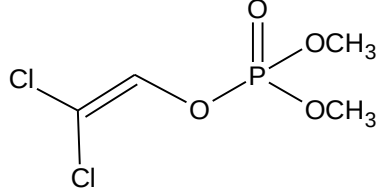
Pesticides	Chemical structure	Endocrine Disruptor Effects	Biomonitoring in human samples
DDT and metabolites (I) M(g/Mol) = 354.5 pKa = n.a logP: 6.91		Competitive binding to androgen receptors, activation of androgen-sensitive cells proliferation. Stimulation of estrogen receptor production, estrogen receptor agonist and PR antagonist [76,93,108,109]	M.P: 0.2–3588 ng/g lipid [94] B.S: <LOD–40.9 ng/g lipid [95] HM: 3.9–4700 ng/g lipid [110,96-99,111] A.F: 0.1–0.63 mg/L [112] M: 1.1–2.8 µg/mL [88] H: 0.17–0.65 µg/mL [88,113] M.B: 0–6168 ng/g lipid [110,114] H.S: 12.5–814.9 ng/mL [77] U.C: 189–3296 ng/g lipid [110]
Deltamethrin (I) M(g/Mol) = 505.2 pKa = n.a logP: 4.6		Weak estrogenic activity [8]	U: 0.5–57.7 µg/g * [107]
Diazinon (I) M(g/Mol) = 304.4 pKa = 2.6 logP: 3.69		Estrogenic effect [115]	S: 1.84–4.96 µg/g creatinine * [69] H.S: mean 2 pg/g [85]
Dichlorvos (I) M(g/Mol) = 221 pKa = n.a logP: 1.9		Weak androgen-receptor antagonist [8]	

Table 1. Cont.

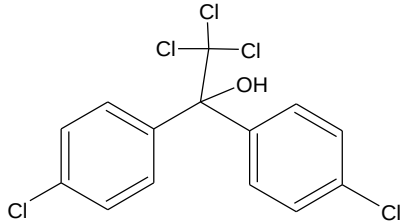
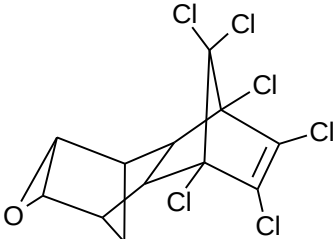
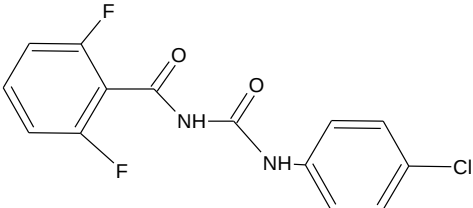
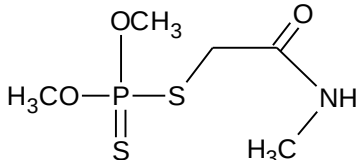
Pesticides	Chemical structure	Endocrine Disruptor Effects	Biomonitoring in human samples
Dicofol (I) M(g/Mol) = 370.5 pKa = n.a logP: 4.3		Inhibition of androgen synthesis, increase of estrogens synthesis, binding to estrogen receptor [90,83]	
Dieldrin (I) M(g/Mol) = 380.9 pKa = n.a logP: 3.7		Competitive binding to androgen receptors, estrogenic effect, stimulation of estrogen receptor production [8,76,108,116]	H.M: <0.1–64 ng/g lipid [111]
Diflubenzuron (I) M(g/Mol) = 310.7 pKa = n.a logP: 3.89		Pregnane X cellular receptor activation [11]	H.S: 1.21–356.4 µg/L [77,78] H.M: mean 0.66 mg/L ± 1.75 [79] A.T: 17.01–84.05 [78]
Dimethoate (I) M(g/Mol) = 229.3 pKa = n.a logP: 0.704		Disruption of thyroid hormones action. Increase of insulin blood concentration, decrease of luteinizing hormone blood concentration [117,118]	

Table 1. Cont.

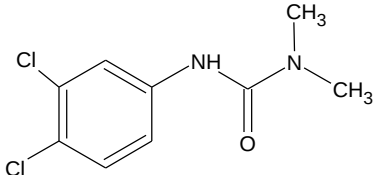
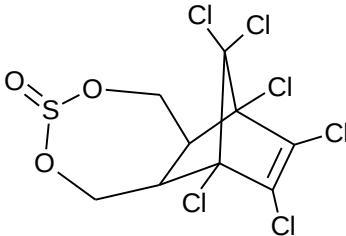
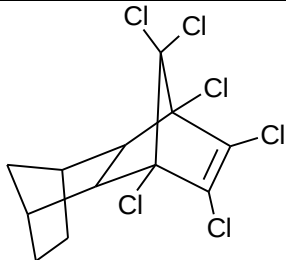
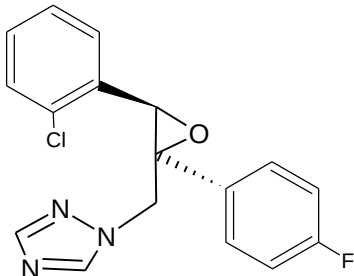
Pesticides	Chemical structure	Endocrine Disruptor Effects	Biomonitoring in human samples
Diuron (H) M(g/Mol) = 233.1 pKa = n.a logP: 2.87		Inhibition of androgens action [83]	
Endosulfan (I) M(g/Mol) = 406.9 pKa = n.a logP: 4.75		Competitive binding to androgen receptors, estrogenic effect, stimulation of estrogen receptor production, inhibition of aromatase activity [8,76,109,116]	H.S: 8.85–547.6 µg/L [77,78] A.T: 21.4–417.6 ng/g lipid [78]
Endrin (I) M(g/Mol) = 380.9 pKa = n.a logP: 3.2		Competitive binding to androgen receptors [76]	H.M: mean 0.65 mg/L ± 1.63 [79] H.S: 1.21–6.35 µg/L [78] A.T: 47.43–148.13 ng/g lipid [78]
Epoxyconazole (F) M(g/Mol) = 329.8 pKa = n.a logP: 3.3		Inhibition of aromatase activity, decrease of estrogen production and increase available androgens [89,119]	

Table 1. Cont.

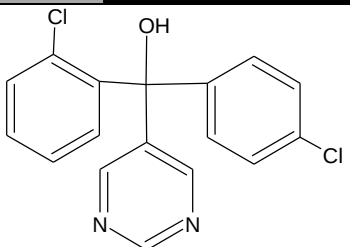
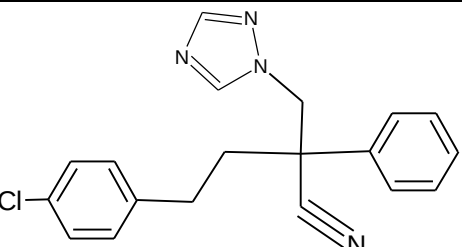
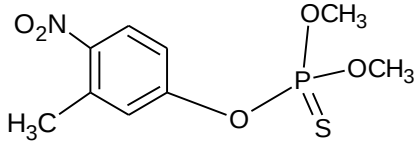
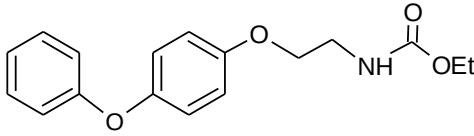
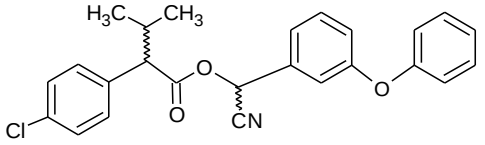
Pesticides	Chemical structure	Endocrine Disruptor Effects	Biomonitoring in human samples
Fenarimol (F) M(g/Mol) = 331.2 pKa = n.a logP: 3.69		Antagonist of androgenic action. Potential aromatase inhibition. Pregnane X cellular receptor activation [8,11,120]	
Fenbuconazole (F) M(g/Mol) = 336.8 pKa = n.a logP: 3.79		Inhibition of thyroid hormones production, Pregnane X cellular receptor activation [11,13]	
Fenitrothion (I) M(g/Mol) = 277.2 pKa = n.a logP: 3.32		Competitive binding to androgen receptor, inhibition of estrogens action [90,121]	H.S: 4.5 µg/mL [71] **
Fenoxycarb (I) M(g/Mol) = 301.3 pKa = n.a logP: 4.07		Interference with testosterone metabolism [122]	
Fenvalerate (I) M(g/Mol) = 419.9 pKa = n.a logP: 5.01		Inhibition of estrogen-sensitive cells proliferation, antagonist of the progesterone action [86,123]	

Table 1. Cont.

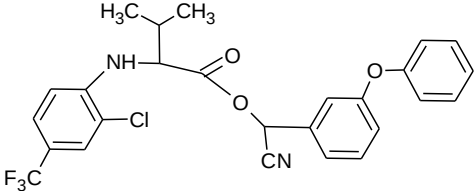
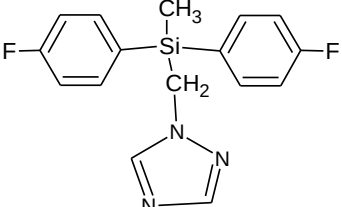
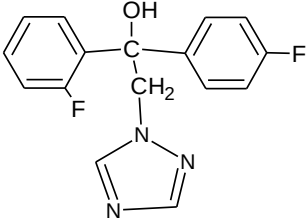
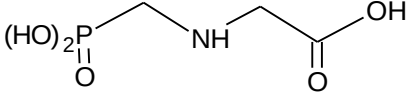
Pesticides	Chemical structure	Endocrine Disruptor Effects	Biomonitoring in human samples
Fluvalinate (I) M(g/Mol) = 502.9 pKa = n.a logP: 3.85		Binding to human sex hormone, Inhibition of progesterone production [124,125]	
Flusilazole (F) M(g/Mol) = 315.4 pKa = 2.5 logP: 3.87		Inhibition of aromatase activity, decrease of estrogens production, increase of available androgens [89]	
Flutriafol (F) M(g/Mol) = 301.3 pKa = 2.3 logP: 2.3		Weak estrogen inhibition [119]	
Glyphosphate (H) M(g/Mol) = 168.1 pKa = 0.78; 2.34; 5.96; 10.98 logP: -3.2		Disruption of aromatase activity, preventing the production of estrogens [126]	U: 1.1–2.1 ng/mL [84]

Table 1. Cont.

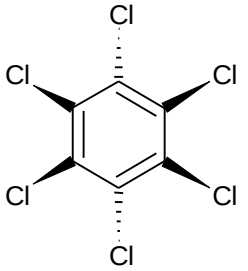
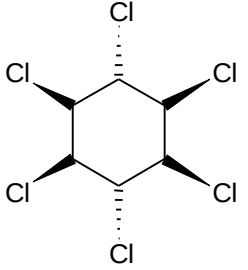
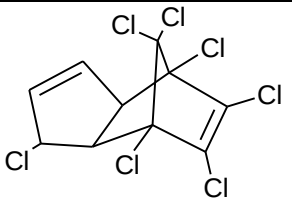
<u>Pesticides</u>	<u>Chemical structure</u>	<u>Endocrine Disruptor Effects</u>	<u>Biomonitoring in human samples</u>
HCB (F) M(g/Mol) = 284.8 pKa = n.a logP: 3.93		Severely disruption of thyroid hormone production. Enhancement of androgen action at low doses, but inhibition at high levels [127,128]	M.P: 1.6–44.3 ng/g lipid [94] B.S: 7.4–37.2 ng/g lipid [95] H.M: 0.4–472 ng/g lipid [79,96-99] H.S: 12.5–393.3 µg/L [77] H: 1.2–15.9 pg/mg [113] F.F: 0.11–0.2 ng/L [100] A.F: [112]
HCH (lindane) (I) M(g/Mol) = 290.8 pKa = n.a logP: 3.61		Reduction of oestrous cycles and luteal progesterone concentrations. Increase of insulin and estradiol blood serum concentrations, decrease thyroxine concentrations. Competitive binding to AR, ER and PR [117,129]	M.P: 0.4–2839 ng/g lipid [94] B.S: <LOD–134 ng/g lipid [95] H.M: 4.7–8700 ng/g lipid [80,110,96-99] H.S: 1.08–265.8 µg/L [77,78] H: 50.7–235 pg/mg [113] A.T: 17.44–113.31 ng/g lipid [78] M.B: 1.9–386.6 ng/g lipid [110, 114] A.F: 0.1–0.26 ng/mL [112] U.C: 4–130 ng/g lipid [110]
Heptachlor (I) M(g/Mol) = 373.3 pKa = n.a logP: 5.44		Binding to cellular estrogen and androgen receptors [130,131]	M.P: 0.2–5.2 ng/g lipid [94] B.S: <LOD–0.9 ng/g lipid [95] Human serum: 12.5–139.1 µg/L [77] H.M: mean 0.07 mg/L ± 0.34 [79]

Table 1. Cont.

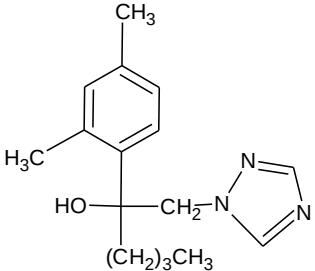
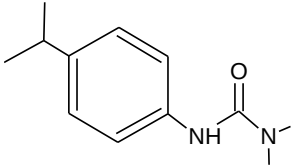
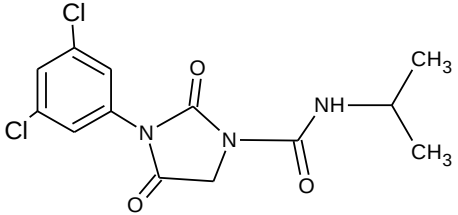
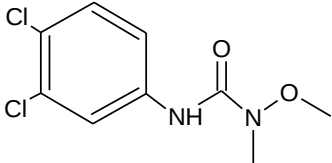
<u>Pesticides</u>	<u>Chemical structure</u>	Endocrine Disruptor <u>Effects</u>	<u>Biomonitoring in human samples</u>
Hexaconazole (F) M(g/Mol) = 314.2 pKa = 2.3 logP: 3.9		Inhibition of aromatase activity, decrease of the estrogens production and increase of available androgens [89]	
Isoproturon (H) M(g/Mol) = 206.3 pKa = n.a logP: 2.5		Pregnane X cellular receptor activation [11]	
Iprodione (F) M(g/Mol) = 330.2 pKa = n.a logP: 3.1		Increase weakly aromatase activity, and estrogen production [8]	
Linuron (H) M(g/Mol) = 249.1 pKa = n.a logP: 3		Competitive binding to androgen receptor, thyroid receptor agonist [131,132]	

Table 1. Cont.

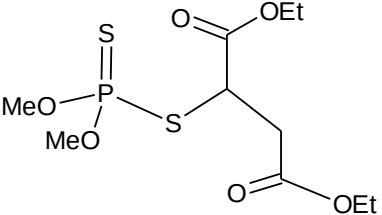
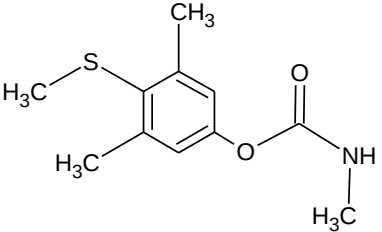
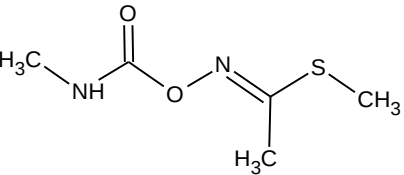
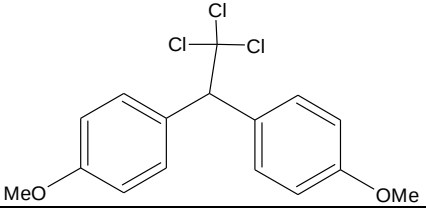
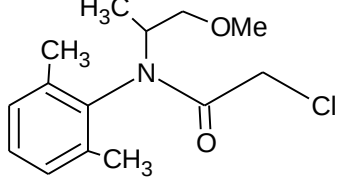
Pesticides	Chemical structure	Endocrine Disruptor Effects	Biomonitoring in human samples
Malathion (I) M(g/Mol) = 330.4 pKa = n.a logP: 2.75		Inhibition of catecholamine secretion, binding to thyroid hormone receptors [13,133]	U: <LOD–3195 ng/mL [68] * M: 2.92–5.38 µg/mL [88] H: 1.62–2.12 µg/mL [88] S: 0.37–0.92 µg/g creatinine [69]
Methiocarb (H) M(g/Mol) = 225.3 pKa = n.a logP: 3.18		Inhibition of androgen activity and increase of estrogen one [8]	
Methomyl (I) M(g/Mol) = 162.2 pKa = n.a logP: 1.24		Weak increase of aromatase activity and estrogen production [8,13]	
Methoxychlor (I) M(g/Mol) = 345.7 pKa = n.a logP: 5.83		Strong estrogenic effect. Competitive binding to androgen receptor, interaction with the pregnane X cellular receptor [13,74,76]	H.S: 0.38–0.39 µg/L [78] A.T: 29.86–155.58 ng/g lipid [78]
Metolachlor M(g/Mol) = 283.8 pKa = n.a logP: 3.4		Pregnane X cellular receptor activation [11]	U: <LOD–4.5 ng/mL [68,84] H.S: mean 2 pg/g [85] M.S: 0.007–1.96 ng/g [92] U.C: 0.007–2.37 ng/g [92] S: 0.20–0.48 µg/g creatinine [69]

Table 1. Cont.

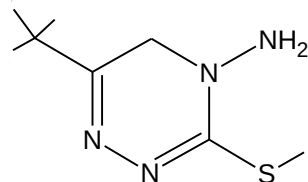
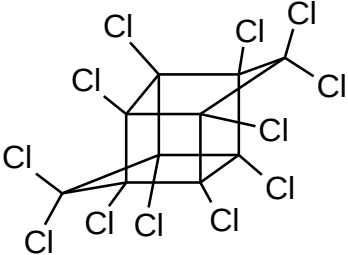
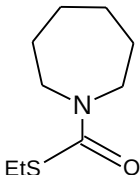
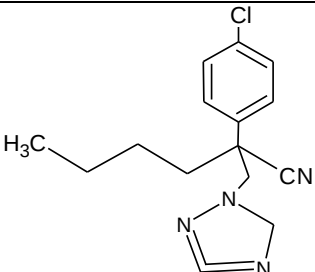
<u>Pesticides</u>	<u>Chemical structure</u>	Endocrine Disruptor <u>Effects</u>	<u>Biomonitoring in human samples</u>
Metribuzin (H) M(g/Mol) = 214.3 pKa = 0.99 logP: 1.65		Hyperthyroidism, alteration of somatotropin levels [134]	
Mirex (I) M(g/Mol) = 545.5 pKa = n.a logP: 5.28		Weak estrogen effect [13]	M.P: 0.2–1.5 ng/g lipid [94] B.S: <LOD–7.2 ng/g lipid [95] H.M: 0.2–1.7 ng/g lipid [98]
Molinate (H) M(g/Mol) = 187.3 pKa = n.a logP: 2.86		Reproductive tract damage, reduction of fertility [13]	
Myclobutanil (F) M(g/Mol) = 288.8 pKa = 2.3 logP: 2.89		Weak estrogen and androgen inhibition, Binding to estrogen and androgen receptors, aromatase inhibition [89,90,119]	

Table 1. Cont.

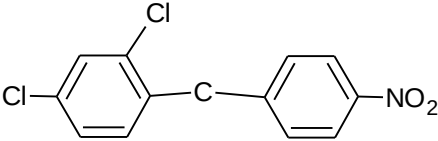
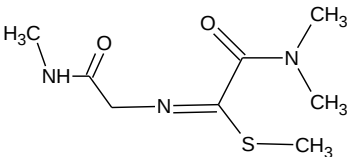
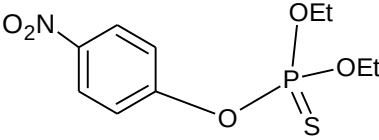
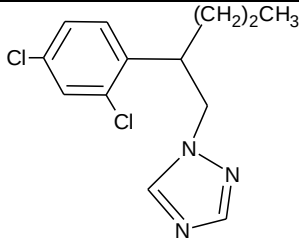
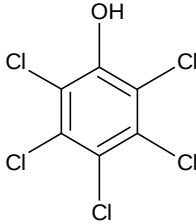
<u>Pesticides</u>	<u>Chemical structure</u>	Endocrine Disruptor <u>Effects</u>	<u>Biomonitoring in human samples</u>
Nitrofen (H) M(g/Mol) = 284.1 pKa = n.a logP: 3.4		Estrogen and androgen inhibition [90]	
Oxamyl (I) M(g/Mol) = 219.3 pKa = n.a logP: −0.44		Weak estrogen effect [13]	
Parathion (I) M(g/Mol) = 291.3 pKa = n.a logP: 3.83		Inhibition of catecholamine secretion, increase of melatonin synthesis, inhibition of gonadotrophic hormone [13]	U: <LOD–84 ng/mL * [68]
Penconazole (F) M(g/Mol) = 284.2 pKa = 1.51 logP: 3.72		Weak estrogenic effect. Inhibition of aromatase activity, decrease of estrogens production and increase androgens availability [89,119]	
Pentachlorophenol (H, F, I) M(g/Mol) = 266.3 pKa = 4.73 logP: 3.32		Weak estrogenic and anti-androgenic affect [13]	A.F : 0.15–0.54 ng/mL [135]

Table 1. Cont.

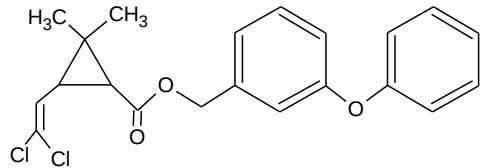
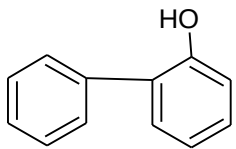
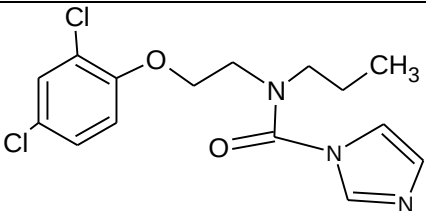
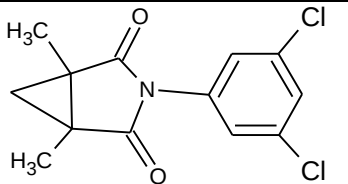
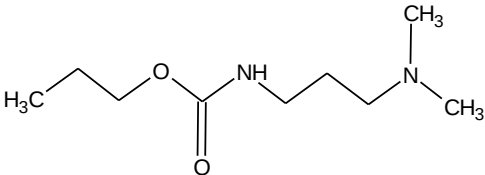
Pesticides	Chemical structure	Endocrine Disruptor Effects	Biomonitoring in human samples
Permethrin (I) M(g/Mol) = 391.3 pKa = n.a logP: 6.1		Inhibition of estrogen-sensitive cells proliferation [87,106]	U: 1–150 µg/g * [107]
Phenylphenol (F) M(g/Mol) = 170.2 pKa = 9.97 logP: 3.09		Estrogen agonist [136]	A.F: 0.1–0.17 ng/mL [135]
Prochloraz (F) M(g/Mol) = 376.7 pKa = 3.8 logP: 3.53		Activation of Pregnane X cellular receptor. Antagonist to cellular androgen and estrogen receptors, agonist to Ah receptor and inhibition of aromatase activity [8,11,120,137]	
Procymidone (F) M(g/Mol) = 284.1 pKa = n.a logP: 3.3		Competitive binding to androgen receptor [131]	
Propamocarb (F) M(g/Mol) = 188.3 pKa = 9.5 logP: 0.84		Weak increase of aromatase activity and estrogen production [8]	

Table 1. Cont.

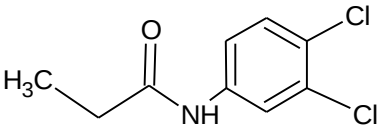
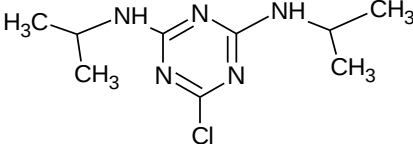
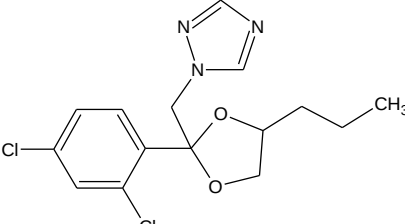
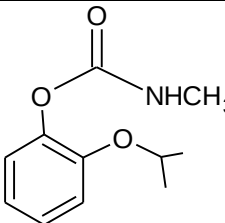
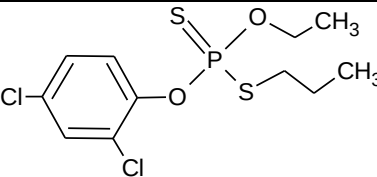
Pesticides	Chemical structure	Endocrine Disruptor Effects	Biomonitoring in human samples
Propanil (H) M(g/Mol) = 318.1 pKa = n.a logP: 2.29		Increase of cellular response to estrogen [138]	
Propazine (H) M(g/Mol) = 229.8 pKa = 1.7 logP: 3.95		Induction of aromatase activity and increase of estrogen production [81]	
Propiconazole (F) M(g/Mol) = 342.2 pKa = 1.09 logP: 3.72		Weak estrogen and aromatase activity inhibition. Decrease estrogens production and increase of androgens availability [89,119]	
Propoxur (I) M(g/Mol) = 209.2 pKa = n.a logP: 0.14		Weak estrogenic effect [13]	M: 0.24–1.50 µg/mL [88] C.B: 0.77 µg/mL [88] H: 0.22–0.42 µg/mL [88] M.B: 0.67–0.77 µg/mL [88]
Prothiophos (I) M(g/Mol) = 345.3 pKa = n.a logP: 5.67		Estrogenic effect [115]	

Table 1. Cont.

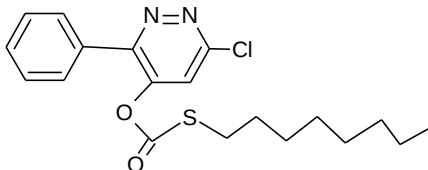
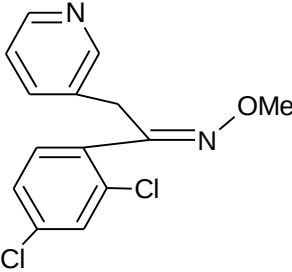
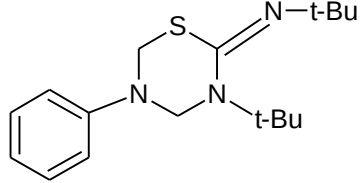
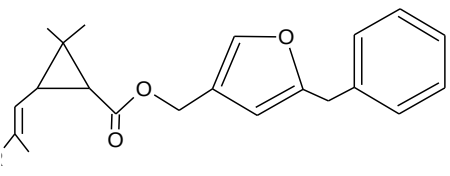
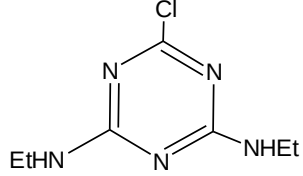
Pesticides	Chemical structure	Endocrine Disruptor Effects	Biomonitoring in human samples
Pyridate (H) M(g/Mol) = 378.9 pKa = n.a logP: 0.5		Binding to estrogen and androgen receptors [90]	
Pyrifenoxy (F) M(g/Mol) = 295.2 pKa = 4.61 logP: 3.4		Weak estrogen inhibition [119]	
Pyriproxyfen (I) M(g/Mol) = 321.4 pKa = 6.87 logP: 5.37		Estrogenic effect [115]	
Resmethrin (I) M(g/Mol) = 338.4 pKa = n.a logP: 5.43		Binding to sex hormone [124]	
Simazine (H) M(g/Mol) = 201.7 pKa = 1.62 logP: 2.3		Induction of aromatase activity, increase of estrogen production [81]	

Table 1. Cont.

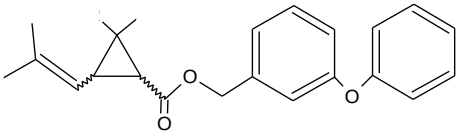
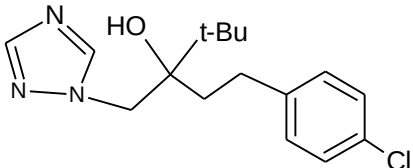
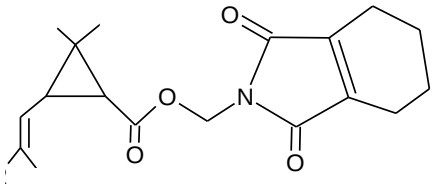
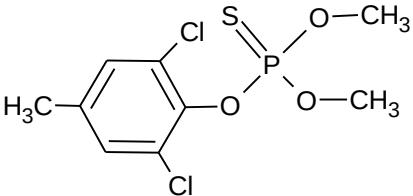
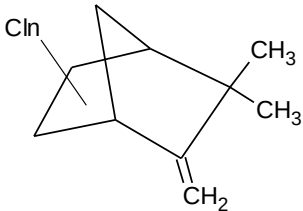
Pesticides	Chemical structure	Endocrine Disruptor Effects	Biomonitoring in human samples
Sumithrin (I) M(g/Mol) = 350.5 pKa = n.a logP: 6.01		Increase of estrogen-sensitive cells proliferation, antagonist of the progesterone action [87,123]	
Tebuconazole (F) M(g/Mol) = 307.8 pKa = n.a logP: 3.7		Inhibition of aromatase activity, decrease the estrogens production and increase androgens availability [89]	
Tetramethrin (I) M(g/Mol) = 331.4 pKa = n.a logP: 4.6		Estrogen-antagonistic effects in females only [139]	
Tolchlofos-methyl (I) M(g/Mol) = 301.1 pKa = n.a logP: 4.56		Competitive binding to cellular estrogen receptors [120]	
Toxaphene (I) M(g/Mol) = 411.8 pKa = n.a logP: 3.3		Increase of estrogen-sensitive cells proliferation. Inhibition of corticosterone synthesis in the adrenal cortex [13,116]	

Table 1. Cont.

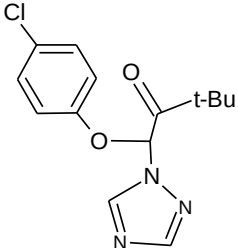
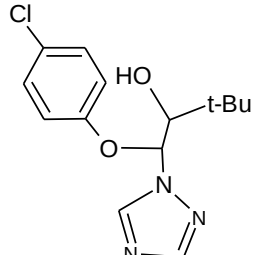
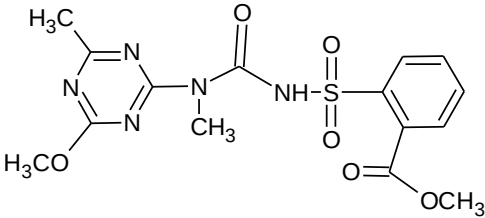
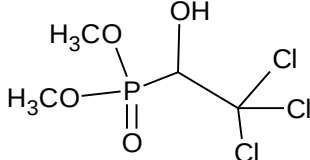
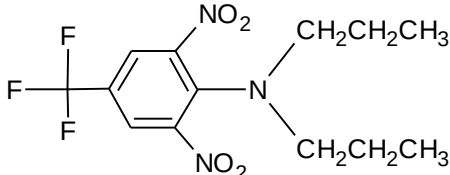
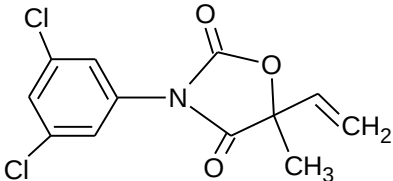
Pesticides	Chemical structure	Endocrine Disruptor Effects	Biomonitoring in human samples
Triadimefon (F) M(g/Mol) = 293.8 pKa = n.a logP: 3.18		Estrogenic effect, inhibition of aromatase activity, decrease of estrogens production and increase androgens availability [90]	
Triadimenol (F) M(g/Mol) = 295.8 pKa = n.a logP: 3.18		Estrogenic effect, inhibition of aromatase activity, decrease of estrogens production and increase androgens availability [89,90]	
Tribenuron-methyl (H) M(g/Mol) = 395.4 pKa = 4.7 logP: 0.78		Weak estrogenic effect [8]	
Trichlorfon (I) M(g/Mol) = 257.4 pKa = n.a logP: 0.43		Alteration of thyroid function [140]	
Trifluralin (H) M(g/Mol) = 335.3 pKa = n.a logP: 5.27		Interaction with pregnane X cellular receptor, interference steroid hormone metabolism [74]	M.S: 0.00–8.5 ng/g [92] U.C: 0.007–4.42 ng/g [92]

Table 1. Cont.

<u>Pesticides</u>	<u>Chemical structure</u>	Endocrine Disruptor <u>Effects</u>	<u>Biomonitoring in human samples</u>
Vinclozolin (F) M(g/Mol) = 286.1 pKa = n.a logP: 3.02		Competitive binding to androgen receptor Interactions with pregnane X cellular receptor, interference with steroid hormone metabolism. [8,74,131]	

(H): Herbicide, (F): Fungicide, (I): Insecticide, (AFA): Antifouling agent, (T): Termiticide, U: urine, S: semen, H.S: human serum, H.M human milk, M: mecomium, H = hair, A.T: adipose tissues, F.F: follicular fluid, M.P: maternal plasma, U.C: umbilical cord.

* Measured by the presence of its metabolite. ** Case of poisoning patient.

Based on the epidemiological studies since 2000, the study concluded that pesticide exposure may affect spermatogenesis leading to poor semen quality and reduced male fertility. Furthermore, an increasing number of epidemiological studies tend to link environmental exposure to pesticides and hormone-dependent cancer risks. High levels of PCBs, DDE, and DDT have been found in fat samples from women with breast cancer [141]. The risk of breast cancer is said to be four times greater in women with increased blood levels of DDE [142]. One of the latest epidemiological studies performed in Spain between 1999 and 2009 shows that among a total of 2,661 cases of breast cancer reported in the female population, 2,173 (81%) were observed in areas of high pesticide contamination [143]. Moreover, it was also suggested that women with hormone responsive breast cancer have a higher DDE body burden than women with benign breast disease [144]. Similar studies have revealed correlations between damage to the immune system and increased amounts of organochlorine residues in certain cancerous tissues [145]. Numerous other studies support the hypothesis that pesticide exposure influences the risk of breast cancer [146], but few of them are really conclusive due to some inconsistent data across the study. Further research is required to explore long-term follow-up beginning in early life, with opportunities for exposure measurement at critical periods of vulnerability. Moreover, improvements are needed in the cohort sample size and standardization of exposure assessments methods. Finally, researchers also need to consider simultaneous co-exposures to these substances and other chemicals and whether they may act in an additive, synergistic, or antagonistic manner [147].

There may also be a connection between pesticide exposure and prostate cancer. Various studies have consistently demonstrated a higher risk in agricultural populations than in the general population [148–150]. For example, pesticides (in particular DDT) were associated with a statistically significant higher rate of prostate cancer among farmers (exposed to organochloride pesticides) in a multi-site case-control study carried out in five rural areas between 1990–92 in Italy [151]. Several studies in the USA and Sweden showed that farmers and commercial pesticide applicators have a slightly and/or significantly higher rate of prostate cancer than the general population [148,152,153].

Several meta-analyses, cohort studies and case-control studies on the risk of prostate cancer in populations exposed occupationally or professionally to pesticides have been conducted in recent years [154] (and reference therein). They all showed a significantly higher risk of prostate cancer estimated at between 10 and 40%, the higher values being for professional exposure. Quite recently, a study analyzed the relationship between exposure to chlordecone (organochloride pesticide extensively used for more than 30 years in the French West Indies to control the banana root borer) and the risk of prostate cancer [155]. It showed a significant increase in the risk of prostate cancer with increasing plasma chlordecone concentrations and supported the hypothesis that exposure to environmental estrogens may increase the risk of prostate cancer.

However, in spite of these outcomes, the hypothesis that such excess risk is related to the use of pesticides has not yet been formally demonstrated. Various other factors have been suggested to explain the increase in prostate cancer in agricultural or rural populations, such as dietary issues, contact with infectious agents via livestock, dust, tobacco and chemical products [154]. Rigorous studies with larger cases that accurately and objectively estimate pesticide exposures and consider gene-environment interactions are needed to determine a potential relationship between pesticides and prostate cancer.

3. Biomonitoring for Human Exposure Assessment

Exposure to pesticides can occur via numerous pathways, including household use of pesticide products, dietary exposure to pesticide residues, and exposure to agricultural drift. Biological monitoring studies indicate that pesticide exposures are widespread in the human population. Dietary exposure comes from residues in fruits, vegetables, and from contaminated meat, fish, rice and dairy products. The European Commission [156] estimated that, in 2005, consumer intake was always below the acceptable daily intakes (ADI) for long-term exposures. Several recent studies also show the difference between EDI and ADI [157-159]. However, according to the European report, the acute reference dose (a parameter for high short-term intakes, usually in one day or one meal) was exceeded for some pesticides in different vegetables and fruits and 26.7% of samples show residues of more than one pesticide, with a significant upward trend as compared to previous years. Food intake is not the only exposure pathway for the general population. Living near sites where pesticides are used, manufactured or disposed of may significantly increase environmental exposure through inhalation and contact with air, water and soil [160-163].

Human exposure to pesticides is assessed by measuring the levels of pesticides in human samples such as breast milk, maternal blood and serum, urine and sometime umbilical cord blood. Improvements in analytical techniques have made it possible to detect pesticides and their metabolites at trace levels (from milligrams per kilogram to femtograms per kilogram in some laboratories) in almost all human samples. Table 1 reports the detection of pesticides in human samples.

Most of these studies show evidence of higher levels of pesticides in the exposed population (for example due to their occupation or geographical location) than in non-exposed control people. For example, a relation was established between employment in agriculture of Spanish women during pregnancy and serum levels of organochlorine endocrine disruptor pesticides, including DDT and isomers (despite their being banned in Spain since 1977) [164]. Also, higher levels of DDTs and HCHs were found in maternal milk and blood samples in Chinese provinces than in developed or industrialized countries [110]. Finally, higher concentrations of several pesticides were also found in urine and plasma of pregnant Israeli women compared to other populations of pregnant women in the United States and the Netherlands.

Pesticide metabolites are also monitored in human samples because they can be representative of a global contamination. This is particularly true for organophosphorus compounds. Alkyl phosphates have been reported in human samples (urine, hair) as representative of exposure to organophosphate pesticides [165-168], and some authors have observed a significant difference in the levels of total dialkyl phosphates among exposed and no exposed groups.

4. Discussion and Perspectives

Endocrine disruptor pesticides are widely used for agricultural, municipal, home and medical purposes worldwide. Humans are exposed to these compounds, and due to their toxic properties, the consequences of this exposure on human hormone-dependent pathologies are being established. Most risk assessment studies and some epidemiologic studies have looked at the exposure and toxicology of a single compound. However, two other considerations must be included: the presence of pesticide by-products and the cumulative exposure to pesticides multiresidue.

It is undisputed that in some cases, pesticide by-products can exhibit greater harmful effects than their parent compounds. For example, one study showed that, at the organism level, the only sublethal effect seen was an increase in heart rate at low concentration and a decrease at higher concentration with the use of aldicarb-sulfoxide but not with aldicarb [169]. Another study reported that the oxons of methyl-parathion, chlorpyrifos and diazinon were 15 to 10 times more toxic (to sperm DNA) than their corresponding parent compounds [170]. As another example, *in vitro* studies confirmed that 2,4-dichlorophenoxyacetic acid (2,4-D), a commonly used organophosphate herbicide promoting the proliferation of androgen-sensitive cells [171], is a known estrogen receptor ligand [172]. Vinclozolin degrades to several metabolites in the soil, in the plants and in animal organisms [173]. Two hydrolytic degradation products, 2-[(3,5-dichlorophenyl)-carbamoyl]oxy-2-methyl-3-butenic acid and 3',5"-dichloro-2-hydroxy-methylbut-3-enalide, have been identified as anti-androgenic compounds that mediate the adverse effects of vinclozolin [173].

Furthermore, the combined actions of pesticides also need also to be addressed in the risk assessment process because mixtures of these substances may cause higher toxic effects than those expected from the single compounds [174]. For example an equimolar mixture of three pesticides (deltamethrin, methiocarb, and prochloraz) suppressed androgen receptor (AR) activation *in vitro* [175]. Also, under the additional presence of simazin and tribenuronmethyl, weight changes of the adrenal gland and alterations in gene expression of AR-associated genes were observed *in vivo* in castrated testosterone-treated rats. The question of the combined effect of mixtures of contaminants has attracted the attention of the scientific community. Predictive approaches are generally based on the mathematical concepts of Concentration Addition (CA) and Independent Action (IA), both predicting the toxicity of a mixture based on the individual toxicities of the mixture components (e.g., [176] and references therein). Recently, a review showed that some other models could be useful as tools to assess combined tissue doses and to help predict potential interactions including thresholds for such effects [177].

Finally, the impact of synthetic pesticides, due in particular to an excessive use (including environmental pollution and implications to human health) have led to modifications in agricultural practices and various national and international regulations limiting their use. Further limitations and/or bans should be sought, along with alternative solutions that are safer and non-toxic to the environment and humans. One such alternative is so called “natural pesticides” that are not synthetically produced, but are derived from nature such as botanicals pesticides (pyrethrum, limonene, and many others), microbial/biological agents (microbes, parasites) and inorganic minerals (boric acid, limestone, diatomaceous earth). These solutions are generally assumed to be less toxic for human health than synthetic pesticides and could represent an interesting alternative. But their usefulness is actually questionable because some such pesticides are not potent enough to control pests but at the same time do exhibit adverse effects for human health (i.e. “natural pyrethroids”). Further studies are needed on the occurrence, fate and impact of such pesticides on the ecosystem and public health.

References

1. Mellanby, K. *The DDT Story*; British Crop Protection Council: Hampshire, UK, 1992.
2. Briggs, J. Green revolution. *Int. Encycl. Hum. Geogr.* **2009**, 634-638.

3. Cooper, J.; Dobson H. The benefits of pesticides to mankind and the environment. *Crop Prot.* **2007**, *26*, 1337-1348.
4. Carson, R. *Silent Spring*; Houghton Mifflin: Boston, MA, USA, 1962.
5. Kolpin, D.W.; Thurman, E.M.; Linhart, S.M. Finding minimal herbicide concentrations in ground water? Try looking for their degradates. *Sci. Total Environ.* **2000**, *248*, 115-122.
6. WHO. *Our Planet, Our Health*; Report of the WHO Commission on Health and Environment; WHO: Geneva, Switzerland, 1992.
7. Vinggaard, A.M.; Hnida, C.; Breinholt, V.; Larsen, J.C. Screening of selected pesticides for inhibition of CYP19 aromatase activity *in vitro*. *Toxicol. In Vitro* **2000**, *14*, 227-234.
8. Andersen, H.R.; Cook, S.J.; Waldbillig, D. Effects of currently used pesticides in assays for estrogenicity, androgenicity, and aromatase activity *in vitro*. *Toxicol. Appl. Pharmacol.* **2002**, *179*, 1-12.
9. Kojima, H.; Katsura, E.; Takeuchi, S.; Niiyama, K.; Kobayashi, K. Screening for estrogen and androgen receptor activities in 200 pesticides by *in vitro* reporter gene assays using Chinese hamster ovary cells. *Environ. Health Perspect.* **2004**, *112*, 524-531.
10. Lemaire, G.; Mnif, W.; Mauvais, P.; Balaguer, P.; Rahmani, R. Activation of alpha- and beta- estrogen receptors by persistent pesticides in reporter cell lines. *Life Sci.* **2006**, *79*, 1160-1169.
11. Lemaire, G.; Mnif, W.; Pascussi, J.M.; Pillon, A.; Rabenoelina, F.; Fenet, H.; Gomez, E.; Casellas, C.; Nicolas, J.C.; Cavaillès, V.; *et al.* Identification of new human PXR ligands among pesticides using a stable reporter cell system. *Toxicol. Sci.* **2006**, *91*, 501-509.
12. Tabb, M.M.; Blumberg, B.; New modes of action for endocrine-disrupting chemicals. *Mol. Endocrinol.* **2006**, *20*, 475-482.
13. Cocco, P. On the rumors about the silent spring. Review of the scientific evidence linking occupational and environmental pesticide exposure to endocrine disruption health effects. *Cad. Saúde Pública* **2002**, *18*, 379-402.
14. Akhtar, N.; Kayani, S.A.; Ahmad, M.M.; Shahab, M.; Insecticide-induced changes in secretory activity of the thyroid gland in rats. *J. Appl. Toxicol.* **1996**, *16*, 397-400.
15. Leghait, J.; Gayrard, V.; Picard-Hagen, N.; Camp, M.; Perdu, E.; Toutain, P.L.; Viguié C. Fipronil-induced disruption of thyroid function in rats is mediated by increased total and free thyroxine clearances concomitantly to increased activity of hepatic enzymes. *Toxicology* **2009**, *255*, 38-44.
16. Sugiyama, S.; Shimada, N.; Miyoshi, H.; Yamauchi, K. Detection of thyroid system-disrupting chemicals using *in vitro* and *in vivo* screening assays in *Xenopus laevis*. *Toxicol. Sci.* **2005**, *88*, 367-374.
17. Bryan, G.W.; Gibbs, P.E.; Hummerstone, L.G.; Burt, G.R. The decline of the gastropod *Nucella lapillus* around south-west England: Evidence for the effect of tributyltin from antifouling paints. *J. Mar. Biol. Assoc. UK* **1986**, *66*, 611-640.
18. Short, J.W.; Rice, S.D.; Brodersen, C.C.; Stickle, W.B. Occurrence of tri-N-butyltin caused imposex in the north pacific marine snail *Nucella limax* in Auke Bay; Alaska. *Mar. Biol.* **1989**, *102*, 291-297.

19. Ellis, D.V.; Pattisina, L.A. Widespread neogastropod imposex; a biological indicator of TBT contamination. *Mar. Pollut. Bull.* **1990**, *21*, 248-253.
20. Heidrich, D.D.; Steckelbroeck, S.; Klingmuller, D. Inhibition of human cytochrome P450 aromatase activity by butyltins. *Steroids* **2001**, *66*, 763-769.
21. Gooding, M.P.; Wilson, V.S.; Folmar, L.C.; Marcovich, D.T.; LeBlanc, G.A. The biocide tributyltin reduces the accumulation of testosterone as fatty acid esters in the mud snail (*Ilyanassa obsoleta*). *Environ. Health Perspect.* **2003**, *111*, 426-230.
22. Bishop, C.A.; Brooks, R.J.; Carey, J.H.; Norstrom, P.N.R.J.; Lean, D.R.S. The case for a cause-effect linkage between environmental contamination and development in egg of the common snapping turtle (*chelydra serpentina*) from Ontario. Canada. *J. Toxicol. Environ. Health* **1991**, *33*, 521-547.
23. Bishop, C.A.; Lean, D.R.S.; Brooks, R.J.; Carey, J.H.; Norstrom, P.N.R.J. Chlorinated hydrocarbons in early life stages of the common snapping turtle (*chelydra serpentina serpentina*) from a coastal wetland on lake Ontario. Canada. *Environ. Toxicol. Chem.* **1995**, *14*, 421-426.
24. Guillette, L.J.; Gross, T.S.; Masson, G.R.; Matter, J.M.; Percival, H.F.; Woodward, A.R. Developmental abnormalities of the gonad and abnormal sex hormone concentrations in juvenile alligators from contaminated and control lakes in Florida. *Environ. Health Perspect.* **1995**, *102*, 680-688.
25. Guillette, L.J.; Pickford, D.B.; Crain, D.A.; Rooney, A.A.; Percival, H.F. Reduction in penis size and plasma testosterone concentrations in juvenile alligators living in a contaminated environment. *Gen. Comp. Endocrinol.* **1996**, *101*, 32-42.
26. Guillette, L.J.; Brock, J.W.; Rooney, A.A.; Woodward, A.R. Serum concentration of various environmental contaminants and their relationship to sex steroid concentration and phallus size in juvenile American alligators. *Arch. Environ. Contam. Toxicol.* **1999**, *36*, 447-455.
27. Crain, D.A.; Guillette, L.J.; Pickford, D.B.; Rooney, A. Alterations in steroidogenesis in alligators (*alligator mississippiensis*) exposed naturally and experimentally to environmental contaminants. *Environ. Health Perspect.* **1997**, *105*, 528-533.
28. Munkittrick, K.R.; Port, C.B.; Van Der Kraak, G.J.; Smith, I.R.; Rokosh, D.A. Impact of bleach kraft mill effluent on population characteristics; liver MFO activity; and serum steroid levels of a Lake Superior white sucker (*Catostomus commersoni*). *Can. J. Fish Aquat. Sci.* **1991**, *48*, 1371-1380.
29. Purdom, C.E.; Hardiman, P.A.; Bye, V.J.; Eno, N.C.; Tyler, C.R.; Sumpter, J.P. Estrogenic effects of effluents from sewage treatment works. *Chem. Ecol.* **1994**, *8*, 275-285.
30. Fry, D.M.; Toone, C.K. DDT-induced feminization of gull embryos. *Science* **1981**, *213*, 922-924.
31. Fry, D.M.; Toone, C.K.; Speich, S.M.; Peard, R.J. Sex ratio skew and breeding patterns of gulls, demographic and toxicological considerations. *Stud. Avian Biol.* **1987**, *10*, 26-43.
32. Crisp, T.M.; Clegg, E.D.; Cooper, R.L.; Wood, W.P.; Anderson, D.G.; Baetcke, K.P.; Hoffmann, J.L.; Morrow, M.S.; Rodier, D.J.; Schaeffer, J.E.; *et al.* Environmental endocrine disruption: An effects assessment and analysis [Review]. *Environ. Health Perspect.* **1998**, *106*, 11-56.
33. Tyler, C.R.; Jobling, S.; Sumpter, J.P. Endocrine disruption in wildlife, a critical review of the evidence. *Crit. Rev. Toxicol.* **1998**, *28*, 319-361.

34. Vos, J.G.; Dybing, E.; Greim, H.A.; Ladefoged, O.; Lambré, C.; Tarazona, J.V.; Brandt, I.; Vethaak, A.D. Health effects of endocrine-disrupting chemicals on wildlife; with special reference to the European situation. *Crit. Rev. Toxicol.* **2000**, *30*, 71-133.
35. Reijnders, P.J. Reproductive failure in common seals feeding on fish from polluted coastal waters. *Nature* **1986**, *324*, 456-457.
36. Norstrom, R.J.; Muir, D.C. Chlorinated hydrocarbon contaminants in arctic marine mammals. *Sci. Total Environ.* **1994**, *154*, 107-128.
37. Facemire, C.F.; Gross, T.S.; Guillette, L.J. Reproductive impairment in the Florida panther: Nature or nurture? *Environ. Health Perspect.* **1995**, *103*, 79-86.
38. Oskam, I.C.; Ropstad, E.; Dahl, E.; Lie, E.; Derocher, A.E.; Wiig, O.; Larsen, S.; Wiger, R.; Skaare, J.U. Organochlorines affect the major androgenic hormone; testosterone; in male polar bears (*Ursus maritimus*) at Svalbard. *J. Toxicol. Environ. Health A* **2003**, *66*, 2119-2139.
39. Mnif, W.; Pillon, A.; Balaguer, P.; Bartegi, A. Endocrine xenoestrogens disrupters, molecular mechanisms and detection methods. *Therapie* **2007**, *62*, 369-386.
40. Palma, P.; Palma, V.L.; Matos, C.; Fernandes, R.M.; Bohn, A.; Soares, A.M.V.M.; Barbosa, I.R. Effects of atrazine and endosulfan sulphate on the ecdysteroid system of *Daphnia magna*. *Chemosphere* **2009**, *74*, 676-681.
41. Palma, P.; Palma, V.L.; Matos, C.; Fernandes, R.M.; Bohn, A.; Soares, A.M.V.M.; Barbosa, I.R. Assessment of the pesticides atrazine; endosulfan sulphate and chlorpyrifos for juvenoid-related endocrine activity using *Daphnia magna*. *Chemosphere* **2009**, *76*, 335-340.
42. Wilson, V.S.; Lambright, C.R.; Furr, J.R.; Howdeshell, K.L.; Gray, L.E., Jr. The Herbicide Linuron Reduces Testosterone Production from the Fetal Rat Testis During Both In Utero and In Vitro Exposures. *Toxicol. Lett.* **2009**, *186*, 73-77.
43. Birnbaum, L.S.; Fenton, S.E. Cancer and developmental exposure to endocrine disruptors. *Environ. Health Perspect.* **2003**, *111*, 389-394.
44. Goldman, L.; Falk, H.; Landrigan, P.J.; Balk, S.J.; Reigart, R.; Etzel, R.A. Environmental pediatrics and its impact on government health policy. *Pediatrics* **2004**, *113*, 1146-1157.
45. Sharpe, R.R.M. Pathways of endocrine disruption during male sexual differentiation and masculinisation. *Best. Pract. Res. Clin. Endocrinol. Metab.* **2006**, *20*, 91-110.
46. Sultan, C.; Balaguer, P.; Terouanne, B.; Georget, V.; Paris, F.; Jeandel, C.; Lumbroso, S.; Nicolas, J. Environmental xenoestrogens; antiandrogens and disorders of male sexual differentiation. *Mol. Cell. Endocrinol.* **2001**, *178*, 99-105.
47. Skakkebaek, N.N. Endocrine disruptors and testicular dysgenesis syndrome. *Horm. Res.* **2002**, *57*, 43.
48. Hardell, L.; Bavel, B.; Lindström, G.; Eriksson, M.; Carlberg, M. In utero exposure to persistent organic pollutants in relation to testicular cancer risk. *Int. J. Androl.* **2006**, *29*, 228-234.
49. Anderson, H.; Wolff, M.S. Environmental contaminants in human milk. *J. Expo. Anal. Environ. Epidemiol.* **2000**, *10*, 755-760.
50. Waliszewski, S.; Aguirre, A.A.; Infanzón, R.M.; Siliceo, J. Carry-over of persistent organochlorine pesticides through placenta to fetus. *Salud. Publica Mex.* **2000**, *42*, 384-390.
51. Przyrembel, H.; Heinrich-Hirsch, B.; Vieth, B. Exposition to and health effects of residues in human milk. *Adv. Exp. Med. Biol.* **2000**, *478*, 307-325.

52. Jacobson, J.L.; Jacobson, S.W. Intellectual impairment in children exposed to polychlorinated biphenyls in utero. *N. Engl. J. Med.* **1996**, *335*, 783-789.
53. Eskenazi, B.; Marks, A.R.; Bradman, A.; Fenster, L.; Johnson, C.; Barr, D.B. In utero exposure to dichlorodiphenyltrichloroethane (DDT) and dichlorodiphenyldichloroethylene (DDE) and neurodevelopment among young Mexican American children. *Pediatrics* **2006**, *118*, 233-241.
54. Ribas-Fito, N.; Cardo, E.; Sala, M.; Eulalia de Muga, M.; Mazon, C.; Verdu, A.; Kogevinas, M.; Grimalt, J.O.; Sunyer, J. Breastfeeding exposure to organochlorine compounds and neurodevelopment in infants. *Pediatrics* **2003**, *111*, 580-585.
55. Beard, J. Australian rural health research collaboration. DDT and human health. *Sci. Total Environ.* **2006**, *355*, 78-89.
56. Xiang, H.; Nuckols, J.R.; Stallones, L. A geographic information assessment of birth weight and crop production patterns around mother's residence. *Environ. Res. A* **2000**, *82*, 160-167.
57. Bell, E.M.; Hertz-Picciotto, I.; Beaumont, J.J. A case-control study of pesticides and fetal death due to congenital anomalies. *Epidemiology* **2001**, *12*, 148-156.
58. Reynolds, P.; Von Behren, J.; Gunier, R.B.; Goldberg, D.E.; Hertz, A.; Harnly, M.E. Childhood cancer and agricultural pesticide use, an ecologic study in California. *Environ. Health Perspect.* **2002**, *110*, 319-324.
59. Kristensen, P.; Irgens, L.M.; Andersen, A.; Bye, A.S.; Sundheim, L. Birth defects among offspring of Norwegian farmers; 1967–1991. *Epidemiology* **1997**, *8*, 537-544.
60. Carbone, P.; Giordano, F.; Nori, F.; Mantovani, A.; Taruscio, D.; Lauria, L.; Figà-Talamanca, I. Cryptorchidism and hypospadias in the Sicilian district of Ragusa and the use of pesticides. *Reprod. Toxicol.* **2006**, *22*, 8-12.
61. Weidner, I.S.; Moller, H.; Jensen, T.K.; Skakkebaek, N.E. Cryptorchidism and hypospadias in sons of gardeners and farmers. *Environ. Health Perspect.* **1998**, *106*, 793-796.
62. Skakkebaek, N.E.; Rajpert-De Meyts, E.; Main, K.M. Testicular dysgenesis syndrome, an increasingly common developmental disorder with environmental aspects. *Hum. Reprod.* **2001**, *16*, 972-978.
63. Damgaard, I.N.; Skakkebaek, N.E.; Toppari, J.; Virtanen, H.E.; Shen, H.; Schramm, K.W.; Petersen, J.H.; Jensen, T.K.; Main, K.M. Persistent pesticides in human breast milk and cryptorchidism. *Environ. Health Perspect.* **2006**, *114*, 1133-1138.
64. Roeleveld, N.; Bretveld, R. The impact of pesticides on male fertility. *Curr. Opin. Obstet. Gynecol.* **2008**, *20*, 229-233.
65. McKinlay, R.; Plant, J.A.; Bell, J.N.B.; Voulvoulis, N. Endocrine disrupting pesticides, Implications for risk assessment. *Environ. Int.* **2008**, *34*, 168-183.
66. PPDB: Pesticide Properties DataBase, University of Hertfordshire; Available online: <http://sitem.herts.ac.uk/aeru/footprint/fr/index.htm> (accessed on 12 March 2011).
67. Kim, D.O.; Lee, S.K.; Jeon, T.W.; Jin, C.H.; Hyun, S.H.; Kim, E.J.; Moon, G.I.; Kim, J.A.; Lee, E.S.; Lee, B.M.; *et al.* Role of metabolism in parathion-induced hepatotoxicity and immunotoxicity. *J. Toxicol. Environ. Health A* **2005**, *68*, 2187-2205.
68. Panuwet, P.; Prapamontol, T.; Chantara, S.; Thavornyuthikarn, P.; Montesano, M.A.; Whitehead, R., Jr.; Barr, D.B. Concentrations of urinary pesticide metabolites in small-scale farmers in Chiang Mai Province; Thailand. *Sci. Total Environ.* **2008**, *407*, 655-668.

69. Swan, S.H.; Kruse, R.L.; Liu, F.; Barr, D.B.; Drobni, E.Z.; Redmon, J.B.; Wang, C.; Brazil, C.; Overstreet, J.W.; Study for Future Families Research Group. Semen quality in relation to biomarkers of pesticide exposure. *Environ. Health Perspect.* **2003**, *111*, 1478-1484.
70. Singh, A.K. Acute effects of acephate and methamidophos and interleukin-1 on corticotropin-releasing factor (CRF) synthesis in and release from the hypothalamus *in vitro*. *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* **2002**, *132*, 9-24.
71. Inoue, S.; Saito, T.; Mase, H.; Suzuki, Y.; Takazawa, K.; Yamamoto, I.; Inokuchi, S. Rapid simultaneous determination for organophosphorus pesticides in human serum by LC-MS. *J. Pharm. Biomed. Anal.* **2007**, *44*, 258-264.
72. Rollerová E.E. Interaction of acetochlor with estrogen receptor in the rat uterus. Acetochlor—possible endocrinomodulator? *Gen. Physiol. Biophys.* **2000**, *19*, 73-84.
73. Crump, D.; Werry, K.; Veldhoen, N.; Van Aggelen, G.; Helbing, C.C. Exposure to the herbicide acetochlor alters thyroid hormone-dependent gene expression and metamorphosis in *Xenopus laevis*. *Environ Health Perspect.* **2002**, *110*, 1199-205.
74. Mikamo, E.; Harada, S.; Nishikawa, J.; Nishihara, T. Endocrine disruptors induce cytochrome P450 by affecting transcriptional regulation via pregnane X receptor. *Toxicol Appl. Pharmacol.* **2003**, *193*, 66-72.
75. Klotz, D.; Arnold, S.F.; McLachlan, J.A. Inhibition of 17 beta-estradiol and progesterone activity in human breast and endometrial cancer cells by carbamate insecticides. *Life Sci.* **1997**, *60*, 1467-1475.
76. Lemaire, G.; Terouanne, B.; Mauvais, P.; Michel, S.; Rahmani, R. Effect of organochlorine pesticides on human androgen receptor activation *in vitro*. *Toxicol. Appl. Pharmacol.* **2004**, *196*, 235-246.
77. Cruz, S.; Lino, C.; Silveira, M.I. Evaluation of organochlorine pesticide residues in human serum from an urban and two rural populations in Portugal. *Sci. Total Environ.* **2003**, *317*, 23-35.
78. Botella, B.; Crespo, J.; Rivas, A.; Cerrillo, I.; Olea-Serrano, M.F.; Olea, N. Exposure of women to organochlorine pesticides in Southern Spain. *Environ. Res.* **2004**, *96*, 34-40.
79. Nasir, K.; Bilo, Y.Y.; Al-Shuraiki, Y. Residues of chlorinated hydrocarbon insecticides in human milk of Jordanian women. *Environ. Pollut.* **1998**, *99*, 141-148.
80. Cooper, R.; Stoker, T.E.; Tyrey, L.; Goldman, J.M.; McElroy, W.K. Atrazine disrupts the hypothalamic control of pituitary-ovarian function. *Toxicol. Sci.* **2000**, *53*, 297-307.
81. Sanderson, J.T.; Seinen, W.; Giesy, J.P.; van den Berg, M. 2-Chloro-s-triazine herbicides induce aromatase (CYP19) activity in H295R human adrenocortical carcinoma cells, a novel mechanism for estrogenicity? *Toxicol. Sci.* **2000**, *54*, 121-127.
82. Hayes, T.; Tsui, M.; Hoang, A.; Haeffele, C.; Vonk, A. Atrazine-induced hermaphroditism at 0.1 ppb in American leopard frogs (*Rana pipiens*); laboratory and field evidence. *Environ. Health Perspect.* **2003**, *111*, 568-575.
83. Thibaut, R.; Porte, C. Effects of endocrine disrupters on sex steroid synthesis and metabolism pathways in fish. *J. Steroid Biochem. Mol. Biol.* **2004**, *92*, 485-494.
84. Curwin, B.D.; Hein, M.J.; Sanderson, W.T.; Striley, C.; Heederik, D.; Kromhout, H.; Reynolds, S.J.; Alavanja, M.C. Urinary Pesticide Concentrations Among Children; Mothers and Fathers Living in Farm and Non-Farm Households in Iowa. *Ann. Occup. Hyg.* **2007**, *51*, 53-65.

85. Barr, D.B.; Barr, J.R.; Maggio, V.L.; Whitehead, R.D., Jr.; Sadowski, M.A.; Whyatt, R.M.; Needham, L.L. A multi-analyte method for the quantification of contemporary pesticides in human serum and plasma using high-resolution mass spectrometry. *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* **2002**, *778*, 99-111.
86. Morinaga, H.; Yanase, T.; Nomura, M.; Okabe, T.; Goto, K.; Harada, N.; Nawata, H. A benzimidazole fungicide; benomyl; and its metabolite; carbendazim; induce aromatase activity in a human ovarian granulosa-like tumor cell line (KGN). *Endocrinology* **2004**, *145*, 1860-1869.
87. Kim, I.Y.; Shin, J.H.; Kim, H.S.; Lee, S.J.; Kang, I.H.; Kim, T.S.; Moon, H.J.; Choi, K.S.; Moon, A.; Han, S.Y. Assessing estrogenic activity of pyrethroid insecticides using *in vitro* combination assays. *J. Reprod. Dev.* **2004**, *50*, 245-255.
88. Ostrea, E.M., Jr.; Bielawski, D.M.; Posecion, N.C., Jr.; Corrion, M.; Villanueva-Uy, E.; Bernardo, R.C.; Jin, Y.; Janisse, J.J.; Ager, J.W. Combined analysis of prenatal (maternal hair and blood) and neonatal (infant hair; cord blood and meconium) matrices to detect fetal exposure to environmental pesticides. *Environ. Res.* **2009**, *10*, 116-122.
89. Trosken, E.E.; Scholz, K.; Lutz, R.W.; Volkel, W.; Zarn, J.A.; Lutz, W.K. Comparative assessment of the inhibition of recombinant human CYP19 (aromatase) by azoles used in agriculture and as drugs for humans. *Endocr. Res.* **2004**, *30*, 387-394.
90. Okubo, T.; Yokoyama, Y.; Kano, K.; Soya, Y.; Kano, I. Estimation of estrogenic and antiestrogenic activities of selected pesticides by MCF-7 cell proliferation assay. *Arch. Environ. Contam. Toxicol.* **2004**, *46*, 445-453.
91. Goad, R.; Goad, J.; Atieh, B.; Gupta, R. Carbofuran-Induced endocrine disruption in adult male rats. *Toxicol. Mech. Methods* **2004**, *14*, 233-239.
92. Barr, D.B.; Ananth, C.V.; Yan, X.; Lashley, S.; Smulian, J.C.; Ledoux, T.A.; Hore, P.; Robson, M.G. Pesticide concentrations in maternal and umbilical cord sera and their relation to birth outcomes in a population of pregnant women and newborns in New Jersey. *Sci. Total Environ.* **2010**, *408*, 790-795.
93. Tessier, D.; Matsumura, F. Increased ErbB-2 tyrosine kinase activity; MAPK phosphorylation; and cell proliferation in the prostate cancer cell line LNCaP following treatment by select pesticides. *Toxicol. Sci.* **2001**, *60*, 38-43.
94. Röllin, H.B.; Sandanger, T.M.; Hansen, L.; Channa, K.; Odland, J.Ø. Concentration of selected persistent organic pollutants in blood from delivering women in South Africa. *Sci. Total Environ.* **2009**, *408*, 146-152.
95. Kang, J.H.; Park, H.; Chang, Y.S.; Choi, J.W. Distribution of organochlorine pesticides (OCPs) and polychlorinated biphenyls (PCBs) in human serum from urban areas in Korea. *Chemosphere* **2008**, *73*, 1625-1631.
96. Schinas, V.; Leotsinidis, M.; Alexopoulos, A.; Tsapanos, V.; Kondakis, X.G. Organochlorine pesticide residues in human breast milk from southwest Greece, associations with weekly food consumption patterns of mothers. *Arch. Environ. Health* **2000**, *55*, 411-417.
97. Tanabe, S.; Kunisue, T. Persistent organic pollutants in human breast milk from Asian countries. *Environ. Pollut.* **2007**, *146*, 400-413.

98. Polder, A.; Thomsen, C.; Lindström, G.; Løken, K.B.; Skaare, J.U. Levels and temporal trends of chlorinated pesticides; polychlorinated biphenyls and brominated flame retardants in individual human breast milk samples from Northern and Southern Norway. *Chemosphere* **2008**, *73*, 14-23.
99. Devanathan, G.; Subramanian, A.; Someya, M.; Sudaryanto, A.; Isobe, T.; Takahashi, S.; Chakraborty, P.; Tanabe, S. Persistent organochlorines in human breast milk from major metropolitan cities in India. *Environ. Pollut.* **2009**, *157*, 148-154.
100. Jarrell, J.F.; Villeneuve, D.; Franklin, C.; Bartlett, S.; Wrixon, W.; Kohut, J.; Zouves, C.G. Contamination of human ovarian follicular fluid and serum by chlorinated organic compounds in three Canadian cities. *Can. Med. Assoc. J.* **1993**, *148*, 1321-1327.
101. Tapiero, H.T.; Nguyen, B.G.; Tew, K.D. Estrogens and environmental estrogens. *Biomed. Pharmacother.* **2002**, *56*, 36-44.
102. Fang, H.; Tong, W.; Branham, W.S.; Moland, C.L.; Dial, S.L.; Hong, H.; Xie, Q.; Perkins, R.; Owens, W.; Sheehan, D.M. Study of 202 natural; synthetic; and environmental chemicals for binding to the androgen receptor. *Chem. Res. Toxicol.* **2003**, *16*, 1338-1358.
103. Vinggaard, A.; Breinholt, V.; Larsen, J.C. Screening of selected pesticides for oestrogen receptor activation *in vitro*. *Food Addit. Contam.* **1999**, *16*, 533-542.
104. Kang, H.G.; Jeong, S.H.; Cho, J.H.; Kim, D.G.; Park, J.M.; Cho, M.H. Chlproprifos-methyl shows anti-androgenic activity without estrogenic activity in rats. *Toxicology* **2004**, *199*, 219-230.
105. Chen, H.; Xiao, J.; Hu, G.; Zhou, J.; Xiao, H.; Wang, X. Estrogenicity of organophosphorus and pyrethroid pesticides. *J. Toxicol. Environ. Health A* **2002**, *65*, 1419-1435.
106. McCarthy, A.R.; Thomson, B.M.; Shaw, I.C.; Abella, A.D. Estrogenicity of pyrethroid insecticide metabolites. *J. Environ. Monit.* **2006**, *8*, 197-202.
107. Hardt, J.; Angerer, J. Biological monitoring of workers after the application of insecticidal pyrethroids. *Int. Arch. Occup. Environ. Health* **2003**, *76*, 492-498.
108. Tapiero, H.T.; Nguyen, B.G.; Tew, K.D. Estrogens and environmental estrogens. *Biomed. Pharmacother.* **2002**, *56*, 36-44.
109. Bulayeva, N.N.; Watson, C.S. Xenoestrogen-induced ERK-1 and ERK-2 activation via multiple membrane-initiated signaling pathways. *Environ. Health Perspect.* **2004**, *112*, 1481-1487.
110. Qu, W.; Suri, R.P.S.; Bi, X.; Sheng, G.; Fu, J. Exposure of young mothers and newborns to organochlorine pesticides (OCPs) in Guangzhou; China. *Sci. Total Environ.* **2010**, *408*, 3133-3138.
111. Fujii, Y.; Haraguchi, K.; Harada, K.H.; Hitomi, T.; Inoue, K.; Itoh, Y.; Watanabe, T.; Takenaka, K.; Uehara, S.; Yang, H.R.; *et al.* Detection of dicofol and related pesticides in human breast milk from China; Korea and Japan. *Chemosphere* **2011**, *82*, 25-31.
112. Foster, W.; Chan, S.; Platt, L.; Hughes, C. Detection of endocrine disrupting chemicals in samples of second trimester human amniotic fluid. *J. Clin. Endocrinol. Metab.* **2000**, *85*, 2954-2957.
113. Tsatsakis, A.M.; Tzatzarakis, M.N.; Tutudaki, M. Pesticide levels in head hair samples of Cretan population as an indicator of present and past exposure. *Forensic Sci. Int.* **2008**, *176*, 67-71.
114. Pathak, R.; Ahmed, R.S.; Tripathi, A.K.; Guleria, K.; Sharma, C.S.; Makhijani, S.D.; Banerjee, B.D. Maternal and cord blood levels of organochlorine pesticides, Association with preterm labor. *Clin. Biochem.* **2009**, *42*, 746-749.

115. Manabe, M.; Kanda, S.; Fukunaga, K.; Tsubura, A.; Nishiyama, T. Evaluation of the estrogenic activities of some pesticides and their combinations using MtT/Se cell proliferation assay. *Int. J. Hyg. Environ. Health* **2006**, *209*, 413-421.
116. Soto, A.M.; Chung, K.L.; Sonnenschein, C. The pesticides endosulfan; toxaphene; and dieldrin have estrogenic effects on human estrogen-sensitive cells. *Environ. Health Perspect.* **1994**, *102*, 380-383.
117. Rawlings, N.C.; Cook, S.J.; Waldbillig, D. Effects of the pesticides carbofuran; chlorpyrifos; dimethoate; lindane; triallate; trifluralin; 2;4-D; and pentachlorophenol on the metabolic endocrine and reproductive endocrine system in ewes. *J. Toxicol. Environ. Health A* **1998**, *54*, 21-36.
118. Mahjoubi-Samet, A.; Hamadi, F.; Soussia, L.; Fadhel, G.; Zeghal, N. Dimethoate effects on thyroid function in suckling rats. *Ann. Endocrinol.* **2005**, *66*, 96-104.
119. Hurst, M.R.R.; Sheahan, D.A. The potential for estrogenic effects of pesticides in headwater streams in the UK. *Sci. Total Environ.* **2003**, *301*, 87-96.
120. Grunfeld, H.T.; Bonefeld-Jorgensen, E.C. Effect of *in vitro* estrogenic pesticides on human estrogen receptor alpha and beta mRNA levels. *Toxicol. Lett.* **2004**, *151*, 467-480.
121. Tamura, H.; Yoshikawa, H.; Gaido, K.W.; Ross, S.M.; DeLisle, R.K.; Welsh, W.J.; Richard, A.M. Interaction of organophosphate pesticides and related compounds with the androgen receptor. *Environ. Health Perspect.* **2003**, *111*, 545-552.
122. Verslycke, T. Testosterone and energy metabolism in the estuarine mysid *Neomysis integer* (Crustacea, Mysidacea) following exposure to endocrine disruptors. *Environ. Toxicol. Chem.* **2004**, *23*, 1289-1296.
123. Garey, J.; Wolff, M.S. Estrogenic and antiprogestagenic activities of pyrethroid insecticides. *Biochem. Biophys. Res. Commun.* **1998**, *251*, 855-859.
124. Eil, C.C.; Nisula, B.C. The binding properties of pyrethroids to human skin fibroblast androgen receptors and to sex hormone binding globulin. *J. Steroid Biochem.* **1990**, *35*, 409-414.
125. Chen, H.Y.; Liu, R.; He, J.; Song, L.; Bian, Q.; Xu, L.; Zhou, J.; Xiao, H.; Dai, G.; Chang, H.C.; Wang, X. Effects of fenvalerate on progesterone production in cultured rat granulosa cells. *Reprod. Toxicol.* **2005**, *20*, 195-202.
126. Richard, S.; Moslemi, S.; Sipahutar, H.; Benachour, N.; Seralini, G-E. Differential effects of glyphosate and roundup on human placental cells and aromatase. *Environ. Health Perspect.* **2005**, *113*, 716-720.
127. Ralph, J.J.; Orgebin-Crist, M.C.; Lareyre, J.J.; Nelson, C.C. Disruption of androgen regulation in the prostate by the environmental contaminant hexachlorobenzene. *Environ. Health Perspect.* **2003**, *111*, 461-466.
128. Verreault, J.; Skaare, J.U.; Jenssen, B.M.; Gabrielsen, G.W. Effects of organochlorine contaminants on thyroid hormone levels in Arctic breeding glaucous gulls; *Larus hyperboreus*. *Environ. Health Perspect.* **2004**, *112*, 532-537.
129. Beard, A.B.; Rawlings, N.C. Thyroid function and effects on reproduction in ewes exposed to the organochlorine pesticides lindane or pentachlorophenol (PCP) from conception. *J. Toxicol. Environ. Health A* **1999**, *58*, 509-530.

130. Oduma, J.A.; Wango, E.O.; Oduor-Okelo, D.; Makawiti, D.W.; Odongo, H. In vivo and *In vitro* effects of graded doses of the pesticide heptachlor on female sex steroid production in rats. *Comp. Biochem. Physiol. C Pharmacol. Toxicol. Endocrinol.* **1995**, *111*, 191-196.
131. Fang, H.; Tong, W.; Branham, W.S.; Moland, C.L.; Dial, S.L.; Hong, H.; Xie, Q.; Perkins, R.; Owens, W.; Sheehan, D.M. Study of 202 natural; synthetic; and environmental chemicals for binding to the androgen receptor. *Chem. Res. Toxicol.* **2003**, *16*, 1338-1358.
132. Schmutzler, C.; Gotthardt, I.; Hofmann, P.J.; Radovic, B.; Kovacs, G.; Stemmler, L.; Nobis, I.; Bacinski, A.; Mentrup, B.; Ambrugger, P.; *et al.* Endocrine Disruptors and the Thyroid Gland—A Combined *in Vitro* and *in Vivo* Analysis of Potential New Biomarkers. *Environ. Health Perspect.* **2007**, *115*, 77-83.
133. Ishihara, A.; Nishiyama, N.; Sugiyama, S.; Yamauchi, K. The effect of endocrine disrupting chemicals on thyroid hormone binding to Japanese quail transthyretin and thyroid hormone receptor. *Gen. Comp. Endocrinol.* **2003**, *134*, 36-43.
134. Porter, W.; Green, S.M.; Debbink, N.L.; Carlson, I. Groundwater pesticides, interactive effects of low concentrations of carbamates aldicarb and methomyl and the triazine metribuzin on thyroxine and somatotropin levels in white rats. *J. Toxicol. Environ. Health* **1993**, *40*, 15-34.
135. Bradman, A.; Barr, D.B.; Henn, B.G.C.; Drumheller, T.; Curry, C.; Eskenazi, B. Measurement of pesticides and other toxicants in amniotic fluid as a potential biomarker of prenatal exposure: A validation study. *Environ. Health Perspect.* **2003**, *111*, 1779-1782.
136. Sonnenschein, C.; Soto, A.M. An updated review of environmental estrogen and androgen mimics and antagonists. *J. Steroid Biochem. Mol. Biol.* **1998**, *65*, 143-150.
137. Vinggaard, A.M.; Hass, U.; Dalgaard, M.; Andersen, H.R.; Bonefeld-Jørgensen, E.; Christiansen, S.; Laier, P.; Poulsen, M.E. Prochloraz, an imidazole fungicide with multiple mechanisms of action. *Int. J. Androl.* **2006**, *29*, 186-192.
138. Salazar, K.D.; Schafer, R.; Barnett, J.B.; Miller, M.R. Evidence for a novel endocrine disruptor, the pesticide propanil requires the ovaries and steroid synthesis to enhance humoral immunity. *Toxicol. Sci.* **2006**, *93*, 62-74.
139. Kim, S.S.; Kwack, S.J.; Lee, R.D.; Lim, K.J.; Rhee, G.S.; Seok, J.H.; Kim, B.H.; Won, Y.H.; Lee, G.S.; Jeung, E.B.; *et al.* Assessment of estrogenic and androgenic activities of tetramethrin *in vitro* and *in vivo* assays. *J. Toxicol. Environ. Health A* **2005**, *68*, 2277-2289.
140. Nicolau, G.G. Circadian rhythms of RNA; DNA and protein in the rat thyroid; adrenal and testis in chronic pesticide exposure. III. Effects of the insecticides (dichlorvos and trichlorphon). *Physiologie* **1983**, *20*, 93-101.
141. Falck, F.; Ricci, A.; Wolff, M.S.; Godbold, J.; Deckers, P. Pesticides and polychlorinated biphenyl residues in human breast lipids and their relation to breast cancer. *Arch. Environ. Health.* **1992**, *47*, 143-146.
142. Davis, D.L.; Bradlow, H.L.; Wolff, M.; Woodruff, T.; Hoel, D.G.; Anton-Culver, H. Hypothesis, xenoestrogens as preventable causes of breast cancer. *Environ. Health Perspect.* **1993**, *101*, 372-377.
143. Parron, T.; Alarcon, R.; Requena, M.D.M.; Hernandez, A. Increased breast cancer risk in women with environmental exposure to pesticides. *Toxicol. Lett.* **2010**, *196*, S180.

144. Dewailly, E.; Nantel, A.; Weber, J.P.; Meyer, F. High levels of PCBs in breast of Inuit women from arctic Quebec. *Bull. Environ. Contam. Toxicol.* **1989**, *4*, 641-646.
145. Krieger, N.; Wolff, M.S.; Hiatt, R.A.; Rivera, M.; Vogelmann, J.; Orentreich, N. Breast cancer and serum organochlorines, a prospective study among white; black; and Asian women. *J. Natl. Cancer Inst.* **1994**, *86*, 589-599.
146. Cohn, B.A. Developmental and environmental origins of breast cancer: DDT as a case study. *Reprod. Toxicol.* **2011**, *31*, 302-311.
147. Meeker, J.D. Exposure to environmental endocrine disrupting compounds and men's health. *Maturitas* **2010**, *66*, 236-241.
148. Alavanja, M.C.; Samanic, C.; Dosemeci, M.; Lubin, J.; Tarone, R.; Lynch, C.F.; Knott, C.; Thomas, K.; Hoppin, J.A.; Barker, J.; Coble, J.; Sandler, D.P.; Blair, A. Use of agricultural pesticides and prostate cancer risk in the Agricultural Health Study cohort. *Am. J. Epidemiol.* **2003**, *157*, 800-814.
149. Prins, G.S. Endocrine disruptors and prostate cancer risk. *Endocr. Relat. Cancer* **2008**, *15*, 649-656.
150. Diamanti-Kandarakis, E.; Bourguignon, J.P.; Giudice, L.C.; Hauser, R.; Prins, G.S.; Soto, A.M.; Zoeller, R.T.; Gore, A.C. Endocrine-disrupting chemicals; An Endocrine Society scientific statement. *Endocr. Rev.* **2009**, *30*, 293-342.
151. Settimi, L.; Masina, A.; Andron, A.; Axelson, O. Prostate cancer and exposure to pesticides in agricultural settings. *Int. J. Cancer* **2003**, *104*, 458-461.
152. Alavanja, M.C.; Sandler, D.P.; Lynch, C.F.; Knott, C.; Lubin, J.H.; Tarone, R.; Thomas, K.; Dosemeci, M.; Barker, J.; Hoppin, J.A.; *et al.* Cancer incidence in the agricultural health study. *Scand. J. Work Environ. Health.* **2005**, *31*, 39-45.
153. Dich, J.; Wiklund, K. Prostate cancer in pesticide applicators in Swedish agriculture. *Prostate* **1998**, *34*, 100-112.
154. Ndong, J.R.; Blanchet, P.; Multigner, L. Pesticides et cancer de la prostate: données épidémiologiques. *Bull. Cancer* **2009**, *96*, 171-180.
155. Multigner, L.; Ndong, J.R.; Giusti, A.; Romana, M.; Delacroix-Maillard, H.; Cordier, S.; Jégou, B.; Thome, J.P.; Blanchet, P. Chlordecone exposure and risk of prostate cancer. *J. Clin. Oncol.* **2010**, *28*, 3457-3462.
156. European Commission. *Monitoring of Pesticide Residues in Products of Plant Origin in the European Union, Norway, Iceland and Liechtenstein*; European Commission: Brussels, Belgium, 2007.
157. Iñigo-Núñez, S.; Herreros, M.A.; Encinas T.; Gonzalez-Bulnes, A. Estimated daily intake of pesticides and xenoestrogenic exposure by fruit consumption in the female population from a Mediterranean country (Spain). *Food Control* **2010**, *21*, 471-477.
158. Osman, K.A.; Al-Humaid, A.I.; Al-Rehiyani, S.M.; Al-Redhaiman, K.N. Estimated daily intake of pesticide residues exposure by vegetables grown in greenhouses in al-qassim region; Saudi Arabia. *Food Control* **2010**, doi:10.1016/j.foodcont.2010.11.031.
159. Fromberg, A.; Granby, K.; Højgård, A.; Fagt, S.; Larsen, J.C. Estimation of dietary intake of PCB and organochlorine pesticides for children and adults. *Food Chem.* **2011**, *125*, 1179-1187.
160. Peakall, D.B. Review of books on endocrine disruptors. *Ecotoxicology* **2000**, *9*, 137-144.

161. Curl, C.L.; Fenske, R.A.; Kissel, J.C.; Shirai, J.H.; Moate, T.F.; Griffith, W.; Coronado, G.; Thompson, B. Evaluation of take-home organophosphorus pesticide exposure among agricultural workers and their children. *Environ. Health Perspect.* **2002**, *110*, A787-A792.
162. Whyatt, R.M.; Camann, D.; Perera, F.P.; Rauh, V.A.; Tang, D.; Kinney, P.L.; Garfinkel, R.; Andrews, H.; Hoepner, L.; Barr, D.B. Biomarkers in assessing residential insecticide exposures during pregnancy and effects on fetal growth. *Toxicol. Appl. Pharmacol.* **2005**, *206*, 246-254.
163. Mantovani, A.; Maranghi, F.; La Rocca, C.; Tiboni, G.M.; Clementi, M. The role of toxicology to characterize biomarkers for agrochemicals with potential endocrine activities. *Reprod. Toxicol.* **2008**, *26*, 1-7.
164. Carreno, J.; Rivas, A.; Granada, A.; Lopez-Espinosa, M.J.; Mariscal, M.; Olea, N.; Olea-Serrano, F. Exposure of young men to organochlorine pesticides in Southern Spain. *Environ. Res.* **2007**, *103*, 55-61.
165. Berman, T.; Hochner-Celnikier, D.; Boyd Barr, D.; Needham, L.L.; Amitai, Y.; Wormser, U.; Richter, E. Pesticide exposure among pregnant women in Jerusalem, Israel: Results of a pilot study. *Environ. Int.* **2011**, *37*, 198-203.
166. Duggan, A.; Charnley, G.; Chen, W.; Chukwudebe, A.; Hawk, R.; Krieger, R.I.; Ross, J.; Yarborough, C. Di-alkyl phosphate biomonitoring data; assessing cumulative exposure to organophosphate pesticides. *Regul. Toxicol. Pharmacol.* **2003**, *37*, 382-395.
167. Payne-Sturges, D.; Cohen, J.; Castorina, R.; Axelrad, D.A.; Woodruff, T.J. Evaluating cumulative organophosphorus pesticide body burden of children, a national case study. *Environ. Sci. Technol.* **2009**, *43*, 7924-7930.
168. Tsatsakis, A.M.; Barbounis, M.G. Kavalakis, M.; Kokkinakis, M.; Terzi, I.; Tzatzarakis, M.N. Determination of dialkyl phosphates in human hair for the biomonitoring of exposure to organophosphate pesticides. *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* **2010**, *878*, 1246-1252.
169. Kuster, E.; Altenburger, R. Suborganismic and organismic effects of aldicarb and its metabolite aldicarb-sulfoxide to the zebrafish embryo (*Danio rerio*). *Chemosphere* **2007**, *68*, 751-760.
170. Salazar-Arredondo, E.; Solís-Heredia, M.; Rojas-García, E.; Hernández-Ochoa, I.; Quintanilla-Vega, B. Sperm chromatin alteration and DNA damage by methyl-parathion; chlorpyrifos and diazinon and their oxon metabolites in human spermatozoa. *Reprod. Toxicol.* **2008**, *25*, 455-460.
171. Kim, H.J.; Park, Y.I.; Dong, M.S. Effects of 2,4-D and DCP on the DHT-induced androgenic action in human prostate cancer cells. *Toxicol. Sci.* **2005**, *88*, 52-59.
172. Blair, R.M.; Fang, H.; Branham, W.S.; Hass, B.S.; Dial, S.L.; Moland, C.L.; Tong, W.; Shi, L.; Perkins, R.; Sheehan, D.M. The estrogen receptor relative binding affinities of 188 natural and xenochemicals: Structural diversity of ligands. *Toxicol. Sci.* **2000**, *54*, 138-153.
173. Kelce, W.R.; Monosson, E.; Gamcsik, M.P.; Law, S.C.; Gray, L.E. Environmental hormone disruptors, evidence that vinclozolin developmental toxicity is mediated by antiandrogenic metabolites. *Toxicol. Appl. Pharmacol.* **1994**, *126*, 276-285.

174. Larsen, J.C.; Binderup, M.L.; Dalgaard, M.; Dragsted, L.O.; Hossaini, A.; Ladefoged, O.; Lam, H.R.; Madsen, C.; Meyer, O.; Rasmussen, E.S.; *et al.* Combined Actions and Interactions of Chemicals in Mixtures. The Toxicological Effects of Exposure to Mixtures of Industrial and Environmental Chemicals; FødevareRapport 12; Larsen, J.C., Ed.; Danish Veterinary and Food Administration: Søborg, Denmark, 2003.
175. Birkhoj, M.; Nellemann, C.; Jarfelt, K.; Jacobsen, H.; Andersen, H.R.; Dalgaard, M.; Vinggaard, A.M. The combined antiandrogenic effects of five commonly used pesticides. *Toxicol. Appl. Pharmacol.* **2004**, *201*, 10-20.
176. Altenburger, R.; Greco, W.R. Extrapolation concepts for dealing with multiple contamination in environmental risk assessment. *Integr. Environ. Assess. Manage.* **2009**, *5*, 62-68.
177. Reffstrup, T.K.; Larsen, J.C.; Meyer, O. Risk assessment of mixtures of pesticides. Current approaches and future strategies. *Regul. Toxicol. Pharmacol.* **2010**, *56*, 174-192.

© 2011 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (<http://creativecommons.org/licenses/by/3.0/>).