## Cover Page for Supplementary

## Manuscript title:

# Improvement of Sludge Dewaterability by Ultrasound-Initiated Cationic Polyacrylamide with Microblock Structure: Role of Surface-Active Monomers 

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## Scheme:

Scheme S1. Proposed reaction scheme of the synthesis of PAB and PAD.

(PAB)

Text:

Text S1. Fineman-Ross method.

$$
\begin{align*}
& \quad \mathbf{G}=\mathbf{r}_{\mathrm{AM}} \mathbf{H}-\mathbf{r}_{\mathrm{CM}} \\
& \\
& =\frac{\mathbf{G}=\frac{\mathbf{R}(\boldsymbol{\rho}-\mathbf{1})}{\rho} ; \mathbf{H}}{\rho} \tag{S1}
\end{align*}
$$

In the above formulae, $r_{A M}$ is the reactivity ratio of AM monomer in AM and cationic monomer pair, $r_{C M}$ is analogous to $r_{A M}, R$ is the molar ratio of AM and cationic monomers in the raw material before the polymerization reaction, and $\rho$ is the molar ratio of AM and cationic monomers in the polymers at low conversion. After the $G$ and $H$ of each point were obtained, the linear fitting curve relating to $G$ and $H$ could be plotted, and the $r_{A M}$ and $r_{C M}$ can be obtained through the slope and intercept of straight line.

Text S2. Kelen-Tüdö method.

$$
\begin{align*}
& \boldsymbol{\eta}=\left(\mathbf{r}_{\mathrm{AM}}+\frac{\mathbf{r}_{\mathrm{CM}}}{\boldsymbol{\delta}}\right) \xi-\frac{\mathbf{r}_{\mathrm{CM}}}{\boldsymbol{\delta}} \\
& \quad \boldsymbol{\eta}=\frac{\mathbf{G}}{\boldsymbol{\delta}+\mathbf{H}} ; \boldsymbol{\xi}=\frac{\mathbf{H}}{\boldsymbol{\delta}+\mathbf{H}} ; \boldsymbol{\delta} \\
& =\sqrt{\mathbf{H}_{\min } \times \mathbf{H}_{\max }} \tag{S2}
\end{align*}
$$

$\eta$ and $\xi$ could be obtained based on the results of Fineman-Ross method and formula S2. Then, the linear fitting curve relating to $\eta$ and $\xi$ could be plotted, and the $r_{A M}$ and $r_{C M}$ were calculated through the slope and intercept of straight line.

Text S3. Y-B-R method.

$$
\begin{gather*}
r_{A M}=\frac{A_{2} C_{1}+\mathrm{nC}_{2}}{\mathrm{~A}_{1} \mathrm{~A}_{2}-\mathrm{n}^{2}}, \quad \mathrm{r}_{\mathrm{CM}}=\frac{\mathrm{A}_{1} \mathrm{C}_{2}+\mathrm{nC}_{1}}{\mathrm{~A}_{1} \mathrm{~A}_{2}-\mathrm{n}^{2}} \\
\mathrm{~A}_{1}=\sum_{\mathrm{i}=1}^{\mathrm{n}} \frac{X_{i}^{2}}{\mathrm{Y}_{\mathrm{i}}} ; \mathrm{A}_{2}=\sum_{\mathrm{i}=1}^{\mathrm{n}} \frac{Y_{i}}{X_{i}^{2}} ; \quad C_{1}=\sum_{\mathrm{i}=1}^{\mathrm{n}} \mathrm{X}_{\mathrm{i}}\left(1-\frac{1}{Y_{\mathrm{i}}}\right) ; \mathrm{C}_{2}=\sum_{\mathrm{i}=1}^{\mathrm{n}} \frac{Y_{i}}{X_{i}}\left(\frac{1}{Y_{i}}-1\right) \tag{S3}
\end{gather*}
$$

In the above formulae, $n$ is the number of test groups; $X$ is the molar ratio of AM to cationic monomers before polymerization; and $Y$ is the molar ratio of AM units to cationic units in copolymers at low conversion.

Text S4. The composition equations of PAB and PAD.

$$
\begin{align*}
& F_{B D M D A C-P A B} \\
&= 1-\frac{-0.44 f_{B D M D A C}^{2}-0.12 f_{B D M D A C}+0.56}{0.08 f_{B D M D A C}^{2}+0.88 f_{B D M D A C}+0.56}  \tag{S4}\\
& F_{D M C-P A D}  \tag{S5}\\
&=1-\frac{-0.36 f_{D M C}^{2}-0.28 f_{D M C}+0.64}{-0.96 f_{D M D}^{2}+0.72 f_{D M C}+0.64}
\end{align*}
$$

In the above formulae, $F$ is the molar ratio of the units of one monomer to the total copolymer units, and $f$ is the molar ratio of one monomer to all material monomers before polymerization.

Text S5. The calculating formulae of sequence distribution of PAB and PAD.

$$
\begin{array}{cl}
\left(\boldsymbol{p}_{1}\right)_{x}=\boldsymbol{p}_{11}^{x-1}\left(\mathbf{1}-\boldsymbol{p}_{11}\right), & \left(\boldsymbol{p}_{2}\right)_{x}=\boldsymbol{p}_{22}^{x-1}\left(\mathbf{1}-\boldsymbol{p}_{22}\right) \\
p_{11}=\frac{r_{A M}(1-C D)}{C D}, & p_{22}=\frac{r_{C M} C D}{1-C D} \tag{S6}
\end{array}
$$

In the above formulae, $x$ is the number of monomer units in the segment. $\left(p_{1}\right)_{x}$ is the probability of generating the xAM segment which possesses $x$ successive AM monomer units, it equals to the percentage of xAM segments in all AM segments in the copolymer. $\left(p_{2}\right)_{x}$ is analogous to $\left(p_{1}\right)_{x}$; and $p_{11}$ is the probability of generating the $2 A M$ segment which possesses two successive AM monomer units. $p_{22}$ is analogous to $p_{11}$.

Text S6. Analytical methods for FCMC.
The filter cake was placed into a crucible and dried for 24 h at $105^{\circ} \mathrm{C}$ in a thermostatic drying oven. FCMC can be calculated by formulae S7:

$$
\begin{equation*}
\mathrm{FCMC}=\frac{M_{1}-M_{2}}{M_{1}-M_{0}} \tag{S7}
\end{equation*}
$$

where $M_{1}$ is the total weight of the filter cake without drying and crucible, $M_{2}$ is the total weight of the filter cake after drying and crucible, and $M_{0}$ is the weight of the crucible.

Text S7. Analytical methods for SRF.
The SRF of sludge can be calculated using formulae S8.

$$
\begin{equation*}
\mathrm{SRF}=\frac{2 b P A^{2}}{\mu C} \tag{S8}
\end{equation*}
$$

where $P$ is the filtering pressure $\left(\mathrm{N} / \mathrm{m}^{2}\right), A$ is the filtering area $\left(\mathrm{m}^{2}\right), \mu$ is the kinetic viscosity $\left(\mathrm{N} \mathrm{s} / \mathrm{m}^{2}\right), b$ is the slope of the filtration curve (S9), and $C$ is the filter cake weight per unit volume filter $\left(\mathrm{kg} / \mathrm{m}^{3}\right)$, which can be obtained by formulae 10 .

$$
\begin{equation*}
\frac{t}{v}=b v+a \tag{S9}
\end{equation*}
$$

where $t$ is the filtering time $(\mathrm{s})$, and $v$ is the filtrate volume $\left(\mathrm{m}^{3}\right)$.

$$
\begin{equation*}
\mathrm{C}=\frac{\frac{1}{C_{i}}}{100-C_{i}}-\frac{C_{f}}{100-C_{f}} \tag{S10}
\end{equation*}
$$

where $C_{i}$ is the moisture content of the initial sludge, and $C_{f}$ is the moisture content of the filter cake.

## Table:

Table S1. Fineman-Ross and Kelen-Tudos parameters for AM/BDMDAC copolymerization system initiated by ultrasound.

| No. | R | $\mathrm{\varrho}$ | G | H | $\zeta$ | $\eta$ | $\delta$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 9 | 5.22 | 7.28 | 15.52 | 0.85 | 0.40 |  |
| 2 | 4 | 2.07 | 2.07 | 7.73 | 0.75 | 0.20 |  |
| 3 | 2.33 | 1.59 | 0.86 | 3.41 | 0.56 | 0.14 |  |
| 4 | 1.5 | 1.01 | 0.01 | 2.23 | 0.46 | 0.003 | 2.67 |
| 5 | 1 | 0.65 | -0.54 | 1.54 | 0.37 | -0.13 |  |
| 6 | 0.67 | 0.4 | -1.01 | 1.12 | 0.30 | -0.27 |  |
| 7 | 0.43 | 0.3 | -1.00 | 0.62 | 0.19 | -0.31 |  |
| 8 | 0.25 | 0.14 | -1.54 | 0.45 | 0.15 | -0.50 |  |

Table S2. Fineman-Ross and Kelen-Tudos parameters for AM/DMC copolymerization system initiated by ultrasound.

| No. | R | $\mathrm{\varrho}$ | G | H | $\zeta$ | $\eta$ | $\delta$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 9 | 6.57 | 7.63 | 12.33 | 0.90 | 0.56 |  |
| 2 | 4 | 3.04 | 2.68 | 5.26 | 0.80 | 0.41 |  |
| 3 | 2.33 | 2.35 | 1.34 | 2.31 | 0.64 | 0.37 |  |
| 4 | 1.5 | 1.38 | 0.41 | 1.63 | 0.55 | 0.14 | 1.31 |
| 5 | 1 | 1.25 | 0.20 | 0.80 | 0.38 | 0.09 |  |
| 6 | 0.67 | 0.86 | -0.11 | 0.52 | 0.29 | -0.06 |  |
| 7 | 0.43 | 0.69 | -0.19 | 0.27 | 0.17 | -0.12 |  |
| 8 | 0.25 | 0.45 | -0.31 | 0.14 | 0.10 | -0.21 |  |

Table S3. The sequence distributions of monomer segments of PAB and PAD (1: AM, 2: cationic monomer) under $f_{2}=0.2$.

| x | $\left(\right.$ PPAB1 $^{\text {a }}$ | $\left(\mathrm{P}_{\text {PAB2 }}\right)_{\mathrm{x}}$ |  | (PPAd2) ${ }^{\text {a }}$ |
| :---: | :---: | :---: | :---: | :---: |
| 1 | 0.31 | 0.72 | 0.28 | 0.91 |


| 2 | 0.21 | 0.20 | 0.20 | 0.08 |
| :---: | :---: | :---: | :---: | :---: |
| 3 | 0.15 | 0.05 | 0.15 | 0.01 |
| 4 | 0.10 | 0.02 | 0.10 | 0 |
| 5 | 0.07 | 0.01 | 0.07 | 0 |
| 6 | 0.05 | 0 | 0.05 | 0 |
| 7 | 0.03 | 0 | 0.04 | 0 |
| 8 | 0.02 | 0 | 0.03 | 0 |
| 9 | 0.02 | 0 | 0.02 | 0 |
| 10 | 0.01 | 0 | 0.01 | 0 |

Table S4. The sequence distributions of monomer segments of PAB and PAD (1: AM, 2: cationic monomer) under $f_{2}=0.4$.

| x | $\left(\mathrm{P}_{\mathrm{PAB1} 1}\right)_{\mathrm{x}}$ | $\left(\mathrm{P}_{\mathrm{PAB2} 2}\right)_{x}$ | $\left(\mathrm{P}_{\mathrm{PAD1} 1}\right)_{x}$ | $\left(\mathrm{P}_{\text {PAD2 }}\right)_{\mathrm{x}}$ |
| :---: | :---: | :---: | :---: | :---: |
| 1 | 0.54 | 0.50 | 0.51 | 0.79 |
| 2 | 0.25 | 0.25 | 0.25 | 0.17 |
| 3 | 0.11 | 0.13 | 0.12 | 0.03 |
| 4 | 0.05 | 0.06 | 0.06 | 0.01 |
| 5 | 0.02 | 0.03 | 0.03 | 0 |
| 6 | 0.01 | 0.02 | 0.01 | 0 |
| 7 | 0.01 | 0.01 | 0.01 | 0 |
| 8 | 0 | 0 | 0 | 0 |
| 9 | 0 | 0 | 0 | 0 |
| 10 | 0 | 0 | 0 | 0 |

Table S5. The sequence distributions of monomer segments of PAB and PAD (1: AM, 2: cationic monomer) under $f_{2}=0.6$.

| X | $\left(\mathrm{P}_{\text {PAB1 }}\right)_{\mathrm{x}}$ | $\left(\mathrm{P}_{\text {PAB2 }}\right)_{\mathrm{x}}$ | $\left(\mathrm{PPAD1} \mathrm{x}_{\mathrm{x}}\right.$ | $\left(\mathrm{PPAD}^{2}\right)_{\mathrm{x}}$ |
| :---: | :---: | :---: | :---: | :---: |
| 1 | 0.73 | 0.31 | 0.70 | 0.63 |
| 2 | 0.20 | 0.21 | 0.21 | 0.23 |
| 3 | 0.05 | 0.15 | 0.06 | 0.09 |
| 4 | 0.02 | 0.10 | 0.02 | 0.03 |
| 5 | 0 | 0.07 | 0.01 | 0.01 |
| 6 | 0 | 0.05 | 0 | 0.01 |
| 7 | 0 | 0.03 | 0 | 0 |
| 8 | 0 | 0.02 | 0 | 0 |
| 9 | 0 | 0.02 | 0 | 0 |
| 10 | 0 | 0.01 | 0 | 0 |

Table S6. The sequence distributions of monomer segments of PAB and PAD (1: AM, 2: cationic monomer) under $f_{2}=0.8$.

| x | $\left(\mathrm{P}_{\text {PAB1 }}\right)_{\mathrm{x}}$ | $\left(\text { P }_{\text {PAB2 }}\right)_{\mathrm{x}}$ | $(\text { PPAD1 })_{\mathrm{x}}$ | $(\text { PPAD2 })_{\mathrm{x}}$ |
| :---: | :---: | :---: | :---: | :---: |
| 1 | 0.88 | 0.14 | 0.86 | 0.39 |
| 2 | 0.11 | 0.12 | 0.12 | 0.24 |
| 3 | 0.01 | 0.10 | 0.02 | 0.15 |
| 4 | 0 | 0.09 | 0 | 0.09 |
| 5 | 0 | 0.08 | 0 | 0.05 |


| 6 | 0 | 0.07 | 0 | 0.03 |
| :---: | :---: | :---: | :---: | :---: |
| 7 | 0 | 0.06 | 0 | 0.02 |
| 8 | 0 | 0.05 | 0 | 0.01 |
| 9 | 0 | 0.04 | 0 | 0.01 |
| 10 | 0 | 0.04 | 0 | 0 |

## Figure:



Figure S1. Determination of AM and DMC reactivity ratios by the Fineman-Ross method.


Figure S2. Determination of AM and DMC reactivity ratios by the Kelen-Tüdö method


Figure S3. Determination of AM and BDMDAC reactivity ratios by the Fineman-Ross method.


Figure S4. Determination of AM and BDMDAC reactivity ratios by the Kelen-Tüdö method.

