

Supplementary table S1: Interleukins involved in West Nile virus pathogenesis.

| Cytokine | Model | Host | Expression | Source | Role in WNV pathogenesis | Reference |
|----------------|--------------------------|-------------------------------|------------------|--|--|---------------|
| IL-1 α | <i>In vitro</i> | Horse | Upregulation | PBMCs | | [1] |
| | | | No change | Blood/Brain | | [2–4] |
| | | Mouse | Upregulation | Blood/brain/ lung/ liver | Unknown | [5–8] |
| | <i>In vivo</i> | | Strain-dependent | Brain | | [9] |
| | | Human | No change | Astrocytes/ Resting M Φ / U-937/ BMDCs | Unknown | [10–14] |
| | | | Upregulation | Neural stem cells/ U373/ LAN-2/M Φ / SK-N-SH cells/ Activated M Φ / NK cells | Direct neurotoxic effects in SK-N-SH cells | [11,12,15–18] |
| IL-1 β | <i>In vitro</i> | Pig (<i>Sus domesticus</i>) | Strain-dependent | THP-1/ THP-1-derived M Φ | Unknown | [19] |
| | | | No change | BMDCs | Unknown | [13] |
| | | | | Primary BMECs and astrocytes | Mediates the ability of CXCR4 expressing lymphocytes to adhere to BMECs via CXCL12 expression. | [20] |
| | <i>In vivo</i> | Mouse | | Peritoneal exudate cells | Expression may be induced by WNV envelope protein | [21] |
| | | | Upregulation | M Φ and BMDCs | Reduces WNV replication. Protective via the modulation of type I IFN responses dependent on the IRF3. | [22] |
| | | | | BMDCs | Induced by WNV via ASC Reduces WNV replication. | [23] |
| | <i>In vitro/ in vivo</i> | Horse | | Cortical neurons | Synergizes with type I IFN to induce ISGs. | [2] |
| | | | Strain-dependent | M Φ | Unknown | [24] |
| | | | Upregulation | PBMCs | Unknown | [1] |
| <i>In vivo</i> | Horse | | Upregulation | Lymphoid tissue | Unknown | [25] |

| | | | | | |
|--------|--|---|---|--|--|
| | | | | Mediates langerhans cell migration and accumulation in the draining lymph nodes upon WNV cutaneous infection | [26] |
| | | Variable No change | Popliteal lymph node/spleen Serum | Unknown | [2,27] |
| | Mouse | Upregulation | Brain | Reduces WNV replication in the brain. Promotes CNS-Intrinsic Immune Control. Prevents synapse recovery and promotes spatial learning defects during WNV recovery | [2–4,27–29] |
| | | Strain-dependent | Brain | Unknown | [9] |
| | Rabbit (<i>Oryctolagus cuniculus</i>) | Upregulation | Draining lymph nodes | Unknown | [30] |
| IL-1ra | <i>In vivo</i> | Mouse NHP (<i>Macaca mulatta</i>) | Upregulation Upregulation | Spleen/ Lung/ kidney Brain/spinal cord | Unknown Unknown |
| IL-2 | <i>In vivo</i> | Mouse | Upregulation | Brain | Unknown |
| | <i>In vitro</i> | Mouse | Upregulation | CD4+ T | Unknown |
| IL-3 | <i>In vivo</i> | Mouse | Upregulation Downregulation | Liver/kidney Brain | Unknown Unknown |
| | <i>In vitro</i> | Human | - No change Downregulation | BMDCs BMDCs | No effect on viral replication Unknown Unknown |
| IL-4 | | Mouse | No change Downregulation | Brain Brain | Unknown Unknown |
| | <i>In vivo</i> | Rabbit (<i>Oryctolagus cuniculus</i> and <i>Sylvilagus sp</i>) | Upregulation | Lymph nodes | Unknown |
| IL-5 | <i>In vivo</i> | Mouse | No change Upregulation Downregulation | Brain Brain Brain | [27] [8] [28] |

| | | | | | |
|-----------------|----------------------------------|--|---|---|---|
| | | No change | Astrocytes/ BMDCs | Unknown | [10,35] |
| | Human | Upregulation | Microglia/ SK-N-SH/ keratinocytes | Does not interfere with WNV-induced cytotoxicity in SK-N-SH. | [10,16,38] |
| <i>In vitro</i> | Human | Strain-dependent | THP-1/ THP-1-derived MΦ | Unknown | [19] |
| | | | MΦ/H36.12j/ Renca/ | Unknown | |
| | Mouse | Upregulation | Peritoneal exudate cells/ $\gamma\delta$ T cells | | [21,24,39,40] |
| | | - | N2a/ cortical neurons | Directly responsible for neuronal apoptosis. | [41] |
| IL-6 | <i>In vitro/ in vivo</i> | Rabbit | Upregulation | PBMCs | Unknown |
| | | Mice | Upregulation | Blood/ brain | Does not impact mortality rates following lethal WNV-infection/ Production in plasma and brain mediated by Toll-like receptor 3. |
| | <i>In vivo</i> | Rabbit (<i>Oryctolagus cuniculus</i> and <i>Sylvilagus sp</i>) | Upregulation | Brain/ spleen | Unknown |
| | <i>Ex vivo</i> | Mouse | Upregulation | Spinal cord slice | Unknown |
| IL-7 | <i>In vivo</i> | Mouse | Upregulation | Brain | Unknown |
| | | | Strain-dependent | Brain | Unknown |
| | | Human | No change | BMDCs | Unknown |
| | <i>In vitro</i> | Mouse | Upregulation | Peritoneal exudate cells/ MΦ | Negatively regulates TNF- α , IL- 12/23p40 and type I interferon expression in MΦ. |
| IL-10 | | | - | Splenocytes | Negatively regulates IL-12/23p40 and type I interferon expression in splenocytes. |
| | <i>In vivo</i> | Mouse | - | - | Promotes viral replication in the periphery and in the CNS via the I.P. and footpad route, not I.C. route. Decreases IL-12/23 p40 and TNF- α and increases mortality rates |
| | | | | | [45] |

| | | | | | | |
|--------|-----------------|--|------------------|--|--|----------------|
| | | | Upregulation | Serum/spleen/ lung/ liver/ brain | Unknown | [7,8,27,28,46] |
| | | Rabbit (<i>Oryctolagus cuniculus</i> and <i>Sylvilagus sp</i>) | Upregulation | Brain/ spleen/ lymph nodes | Unknown | [37] |
| | | Horse | Upregulation | Brain | Unknown | [25] |
| | | NHP (<i>Macaca mulatta</i>) | Upregulation | Brain/spinal cord | Unknown | [31] |
| IL-11 | <i>In vivo</i> | Mouse | Upregulation | Brain | Unknown | [27] |
| | | Human | No change | BMDCs | Unknown | [14,34,35] |
| | | | Strain-dependent | THP-1/ THP-1-derived MΦ | Unknown | [19] |
| | <i>In vitro</i> | Mouse | Upregulation | Keratinocytes/ microglia | Unknown | [47,48] |
| | | Rabbit (<i>Oryctolagus cuniculus</i>) | Upregulation | PBMCs | | [42] |
| IL-12 | | | | | IL-12 p35 does not impact survival rate following WNV-challenge/ Does not interfere with brain infiltration or homing of leukocytes. | [49] |
| | <i>In vivo</i> | Mouse | - | - | | |
| | | | Upregulation | Blood/ Spleen/ lung/ liver/ brain/DCs | Produced via TLR7-MyD88 signaling during WNV infection. | [7,43,50] |
| IL-13 | <i>In vivo</i> | Mouse | No change | Brain | Unknown | [27] |
| | | | Upregulation | Serum/lung/kidney | Unknown | [7,8] |
| | | | Downregulation | Brain | Unknown | [28] |
| | | | Strain-dependent | Brain | Unknown | [8] |
| IL-15 | <i>In vitro</i> | Human | No change | BMDCs | Unknown | [35] |
| | <i>In vivo</i> | Mouse | Upregulation | Spleen/ lung/ brain | Unknown | [7,27,32] |
| | | Horse | Upregulation | Thalamus/ cerebrum | Unknown | [51] |
| IL-16 | <i>In vivo</i> | Mouse | Downregulation | Brain | Unknown | [28] |
| | <i>In vitro</i> | Human | Upregulation | PBMCs | Unknown | [52] |
| IL-17A | <i>In vivo</i> | Mouse | Upregulation | Serum/ liver/ brain | Protects mice from lethal WNV infection and facilitates CD8+T cell cytotoxicity. IL-17A production during WNV infection in mice largely | [7,32,52] |

depends on IL-23 signaling.

| | | | | | | |
|---------------|-----------------|-------|------------------|--|---|---------|
| | | | Strain-dependent | Brain | Unknown | [8] |
| IL-17B | <i>In vivo</i> | Mouse | Downregulation | Brain | Unknown | [28] |
| IL-18 | <i>In vitro</i> | Human | No change | BMDCs/ SK-N-SH | Unknown | [14,16] |
| | <i>In vivo</i> | Mouse | Upregulation | Lung/ spleen | Unknown | [7] |
| IL-20 | <i>In vivo</i> | Mouse | Downregulation | Brain | Unknown | [28] |
| IL-22 | <i>In vivo</i> | Mouse | - | - | Enhances footpad WNV-lethal infection/ Facilitates WNV Entry into the CNS/ Promotes CXCL1, CXCL5 and Cxcr2 expression | [53] |
| | | | Upregulation | Lung/kidney | Unknown | [7] |
| | | Horse | Upregulation | Peripheral blood leukocytes/ Thalamus/ cerebrum/ lymphoid tissue | Unknown | [25,51] |
| IL-23 | <i>In vitro</i> | Human | No change | BMDCs | Unknown | [14] |
| | <i>In vivo</i> | Mouse | - | - | Protects mice from lethal WNV-challenge. Mediates brain infiltration and homing of leukocytes. | [49] |
| IL-33 | <i>In vivo</i> | Mouse | Upregulation | Lung/kidney | Unknown | [7] |
| | | | Upregulation | Spleen MΦ | Unknown | [54] |

Cell lines: H36.12j; *Mus musculus* (macrophage cell line) ATCC CRL-2449™; Renca *Mus musculus* (Kidney epithelial cells line) ATCC CRL-2947™; retinal pigment epithelium cell line (ARPE-19): L929 ATCC CCL-1™; LAN-2: *Mus musculus* neuroblastoma cell line (RRID:CVCL_1829); U-937: *Homo sapiens* Lymphoma cell line CRL-1593.2™; U373: *Homo sapiens* glioblastoma astrocytoma cell-line; SK-N-SH *Homo sapiens* neuroblastoma cell line ATCC HTB-11™; THP-1: Monocytic leukemia cells ATCC TIB-202 ; N2a : *Mus musculus* (neuroblastoma cell line) ATCC CCL-131™.

Abbreviations: ASC: Apoptosis-associated speck-like protein containing C-terminal caspase recruitment domain [CARD]; BMDCs: Bone marrow-derived dendritic cells; BMECs: Brain microvasculature endothelial cells; BMECs: Brain microvascular endothelial cells; CNS: Central nervous system; DCs: Dendritic cells; RPE: retinal pigment epithelium (RPE), ISGs: Interferon-stimulated genes; I.C. : Intracranial ; I.P.: Intra-peritoneal; IFN: Interferon, IRF: IFN regulatory factor 3; MΦ: Macrophages; NHP: Non-Human primate; PBMCs: Peripheral blood mononuclear cells.

References:

1. Uddin, M.J.; Suen, W.W.; Bosco-Lauth, A.; Hartwig, A.E.; Hall, R.A.; Bowen, R.A.; Bielefeldt-Ohmann, H. Kinetics of the West Nile Virus Induced Transcripts of Selected Cytokines and Toll-like Receptors in Equine Peripheral Blood Mononuclear Cells. *Veterinary Research* **2016**, *47*, 61, doi:10.1186/s13567-016-0347-8.
2. Ramos, H.J.; Lanteri, M.C.; Blahnik, G.; Negash, A.; Suthar, M.S.; Brassil, M.M.; Sodhi, K.; Treuting, P.M.; Busch, M.P.; Norris, P.J.; et al. IL-1 β Signaling Promotes CNS-Intrinsic Immune Control of West Nile Virus Infection. *PLoS Pathogens* **2012**, *8*, e1003039, doi:10.1371/journal.ppat.1003039.
3. Garber, C.; Vasek, M.J.; Vollmer, L.L.; Sun, T.; Jiang, X.; Klein, R.S. Astrocytes Decrease Adult Neurogenesis during Virus-Induced Memory Dysfunction via Interleukin-1. *Nat Immunol* **2018**, *19*, 151–161, doi:10.1038/s41590-017-0021-y.
4. Soung, A.L.; Davé, V.A.; Garber, C.; Tycksen, E.D.; Vollmer, L.L.; Klein, R.S. IL-1 Reprogramming of Adult Neural Stem Cells Limits Neurocognitive Recovery after Viral Encephalitis by Maintaining a Proinflammatory State. *Brain, Behavior, and Immunity* **2022**, *99*, 383–396, doi:10.1016/j.bbi.2021.10.010.
5. Durrant, D.M.; Robinette, M.L.; Klein, R.S. IL-1R1 Is Required for Dendritic Cell-Mediated T Cell Reactivation within the CNS during West Nile Virus Encephalitis. *Journal of Experimental Medicine* **2013**, *210*, 503–516, doi:10.1084/jem.20121897.
6. Garcia-tapia, D.; Hassett, D.E.; Mitchell Jr, W.J.; Johnson, G.C.; Kleiboeker, S.B. West Nile Virus Encephalitis : Sequential Histopathological and Immunological Events in a Murine Model of Infection. *Journal of NeuroVirology* **2007**, *13*, 130–138, doi:10.1080/13550280601187185.
7. Peña, J.; Plante, J.A.; Carillo, A.C.; Roberts, K.K.; Smith, J.K.; Juelich, T.L.; Beasley, D.W.C.; Freiberg, A.N.; Labute, M.X.; Naraghi-Arani, P. Multiplexed Digital mRNA Profiling of the Inflammatory Response in the West Nile Swiss Webster Mouse Model. *PLoS Neglected Tropical Diseases* **2014**, *8*, e3216, doi:10.1371/journal.pntd.0003216.
8. Kumar, M.; Roe, K.; O'Connell, M.; Nerurkar, V.R. Induction of Virus-Specific Effector Immune Cell Response Limits Virus Replication and Severe Disease in Mice Infected with Non-Lethal West Nile Virus Eg101 Strain. *Journal of Neuroinflammation* **2015**, *12*, 178, doi:10.1186/s12974-015-0400-y.
9. Kumar, M.; Belcaid, M.; Nerurkar, V.R. Identification of Host Genes Leading to West Nile Virus Encephalitis in Mice Brain Using RNA-Seq Analysis. *Scientific Reports* **2016**, *6*, 26350, doi:10.1038/srep26350.

10. Cheeran, M.C.J.; Hu, S.; Sheng, W.S.; Rashid, A.; Peterson, P.K.; Lokensgard, J.R. Differential Responses of Human Brain Cells to West Nile Virus Infection. *Journal of NeuroVirology* **2005**, *11*, 512–524, doi:10.1080/13550280500384982.
11. Kong, K.; Wang, X.; Anderson, J.; Fikrig, E.; RR Montgomery West Nile Virus Attenuates Activation of Primary Human Macrophages. *Viral Immunol* **2008**, *21*, 78–82, doi:10.1089/vim.2007.0072.
12. Marle, G. Van; Antony, J.; Ostermann, H.; Dunham, C.; Hunt, T.; Halliday, W.; Maingat, F.; Urbanowski, M.D.; Hobman, T.; Peeling, J.; et al. West Nile Virus-Induced Neuroinflammation : Glial Infection and Capsid Protein-Mediated Neurovirulence. *J Virol* **2007**, *81*, 10933–10949, doi:10.1128/JVI.02422-06.
13. García-Nicolás, O.; Lewandowska, M.; Ricklin, M.E.; Summerfield, A. Monocyte-Derived Dendritic Cells as Model to Evaluate Species Tropism of Mosquito-Borne Flaviviruses. *Frontiers in Cellular and Infection Microbiology* **2019**, *9*, 5, doi:10.3389/fcimb.2019.00163.
14. Kovats, S.; Turner, S.; Simmons, A.; Powe, T.; Chakravarty, E.; Alberola-Illa, J. West Nile Virus-Infected Human Dendritic Cells Fail to Fully Activate Invariant Natural Killer T Cells. *Clinical and Experimental Immunology* **2016**, *186*, 214–226, doi:10.1111/cei.12850.
15. Riccetti, S.; Sinigaglia, A.; Desole, G.; Nowotny, N.; Trevisan, M.; Barzon, L. Modelling West Nile Virus and Usutu Virus Pathogenicity in Human Neural Stem Cells. *Viruses* **2020**, *12*, 882, doi:10.3390/v12080882.
16. Kumar, M.; Verma, S.; Nerurkar, V.R. Pro-Inflammatory Cytokines Derived from West Nile Virus (WNV)-Infected SK-N-SH Cells Mediate Neuroinflammatory Markers and Neuronal Death. *Journal of Neuroinflammation* **2010**, *7*, 73, doi:10.1186/1742-2094-7-73.
17. Qian, F.; Chung, L.; Zheng, W.; Bruno, V.; Alexander, R.P.; Wang, Z.; Wang, X.; Kurscheid, S.; Zhao, H.; Fikrig, E.; et al. Identification of Genes Critical for Resistance to Infection by West Nile Virus Using RNA-Seq Analysis. *Viruses* **2013**, *5*, 1664–1681, doi:10.3390/v5071664.
18. Yao, Y.; Strauss-albee, D.M.; Zhou, J.Q.; Malawista, A.; Garcia, M.N.; Murray, K.O.; Blish, C.A.; Montgomery, R.R. The Natural Killer Cell Response to West Nile Virus in Young and Old Individuals with or without a Prior History of Infection. *PLoS ONE* **2017**, *12*, e0172625, doi:10.1371/journal.pone.0172625.
19. Xie, G.; Luo, H.; Tian, B.; Mann, B.; Bao, X.; McBride, J.; Tesh, R.; Barrett, A.D.; Wang, T. A West Nile Virus NS4B-P38G Mutant Strain Induces Cell Intrinsic Innate Cytokine Responses in Human Monocytic and Macrophage Cells. *Vaccine* **2015**, *33*, 869–878, doi:10.1016/j.vaccine.2014.12.056.A.

20. Durrant, D.M.; Daniels, B.P.; Klein, R.S. IL-1R1 Signaling Regulates CXCL12-Mediated T Cell Localization and Fate within the CNS during West Nile Virus Encephalitis. *J Immunol* **2014**, *193*, 4095–4106, doi:10.4049/jimmunol.1401192.
21. Sapkal, G.N.; Harini, S.; Ayachit, V.M.; Fulmali, P. V; Mahamuni, S.A.; Bondre, V.P.; Gore, M.M. Neutralization Escape Variant of West Nile Virus Associated with Altered Peripheral Pathogenicity and Differential Cytokine Profile. *Virus Research* **2011**, *158*, 130–139, doi:10.1016/j.virusres.2011.03.023.
22. Aarreberg, L.D.; Wilkins, C.; Ramos, H.J.; Green, R.; Davis, M.A.; Chow, K.; Gale, M. Interleukin-1 β Signaling in Dendritic Cells Induces Antiviral Interferon Responses. *mBio* **2018**, *9*, e00342-18, doi:10.1128/mBio .00342-18.
23. Kumar, M.; Roe, K.; Orillo, B.; Muruve, D.A.; Nerurkar, V.R.; Gale, M.; Verma, S. Inflammasome Adaptor Protein Apoptosis-Associated Speck-Like Protein Containing CARD (ASC) Is Critical for the Immune Response and Survival in West Nile Virus Encephalitis. *Journal of Virology* **2013**, *87*, 3655–3667, doi:10.1128/jvi.02667-12.
24. Shirato, K.; Miyoshi, H.; Kariwa, H.; Takashima, I. The Kinetics of Proinflammatory Cytokines in Murine Peritoneal Macrophages Infected with Envelope Protein-Glycosylated or Non-Glycosylated West Nile Virus. *Virus Research* **2006**, *121*, 11–16, doi:10.1016/j.virusres.2006.03.010.
25. Bielefeldt-ohmann, H.; Bosco-lauth, A.; Hartwig, A.; Uddin, M.J.; Barcelon, J.; Suen, W.W.; Wang, W.; Hall, R.A.; Bowen, R.A. Microbial Pathogenesis Characterization of Non-Lethal West Nile Virus (WNV) Infection in Horses : Subclinical Pathology and Innate Immune Response. *Microbial Pathogenesis* **2017**, *103*, 71–79, doi:10.1016/j.micpath.2016.12.018.
26. Byrne, S.N.; Halliday, G.M.; Johnston, L.J.; King, N.J.C. Interleukin-1 β but Not Tumor Necrosis Factor Is Involved in West Nile Virus-Induced Langerhans Cell Migration from the Skin in C57BL/6 Mice. *Journal of Investigative Dermatology* **2001**, *117*, 702–709, doi:10.1046/j.0022-202x.2001.01454.x.
27. Glass, W.G.; Lim, J.K.; Cholera, R.; Pletnev, A.G.; Gao, J.L.; Murphy, P.M. Chemokine Receptor CCR5 Promotes Leukocyte Trafficking to the Brain and Survival in West Nile Virus Infection. *Journal of Experimental Medicine* **2005**, *202*, 1087–1098, doi:10.1084/jem.20042530.
28. Kumar, M.; Nerurkar, V.R. Integrated Analysis of MicroRNAs and Their Disease Related Targets in the Brain of Mice Infected with West Nile Virus. *Virology* **2014**, *0*, 143–151, doi:10.1016/j.virol.2014.01.004.

29. Lim, S.M.; van den Ham, H.J.; Oduber, M.; Martina, E.; Zaaraoui-Boutahar, F.; Roose, J.M.; van IJcken, W.F.J.; Osterhaus, A.D.M.E.; Andeweg, A.C.; Koraka, P.; et al. Transcriptomic Analyses Reveal Differential Gene Expression of Immune and Cell Death Pathways in the Brains of Mice Infected with West Nile Virus and Chikungunya Virus. *Frontiers in Microbiology* **2017**, *8*, 1556, doi:10.3389/fmicb.2017.01556.
30. Suen, W.W.; Imoda, M.; Thomas, A.W.; Nasir, N.N.B.M.; Tearnsg, N.; Wang, W.; Bielefeldt-Ohmann, H. An Acute Stress Model in New Zealand White Rabbits Exhibits Altered Immune Response to Infection with West Nile Virus. *Pathogens* **2019**, *8*, 195, doi:10.3390/pathogens8040195.
31. Maximova, O.A.; Sturdevant, D.E.; Kash, J.C.; Kanakabandi, K.; Xiao, Y.; Minai, M.; Moore, I.N.; Taubenberger, J.; Martens, C.; Cohen, J.I.; et al. Virus Infection of the Cns Disrupts the Immune-Neural-Synaptic Axis via Induction of Pleiotropic Gene Regulation of Host Responses. *eLife* **2021**, *10*, e62273, doi:10.7554/eLife.62273.
32. Clarke, P.; Leser, J.S.; Bowen, R.A.; Tylera, K.L. Virus-Induced Transcriptional Changes in the Brain Include the Differential Expression of Genes Associated with Interferon, Apoptosis, Interleukin 17 Receptor A, and Glutamate Signaling as Well as Flavivirus-Specific Upregulation of TRNA Synthetases. *mBio* **2014**, *5*, e00902-14, doi:10.1128/mBio.00902-14.
33. Brien, J.D.; Uhrlaub, J.L.; Nikolich-Zugich, J. West Nile Virus-Specific CD4 T Cells Exhibit Direct Anti-Viral Cytokine Secretion and Cytotoxicity and Are Sufficient for Antiviral Protection. *J Immunol.* **2012**, *181*, 8568–8575, doi:10.4049/jimmunol.181.12.8568.
34. Martina, B.E.E.; Koraka, P.; Doel, P. van den; Rimmelzwaan, G.F.; Haagmans, B.L.; Osterhaus, A.D.M.E. DC-SIGN Enhances Infection of Cells with Glycosylated West Nile Virus in Vitro and Virus Replication in Human Dendritic Cells Induces Production of IFN- α and TNF- α . *Virus Research* **2008**, *135*, 64–71, doi:10.1016/j.virusres.2008.02.008.
35. Zimmerman, M.G.; Bowen, J.R.; McDonald, C.E.; Pulendran, B.; Suthar, M.S. West Nile Virus Infection Blocks Inflammatory Response and T Cell Costimulatory Capacity of Human Monocyte-Derived Dendritic Cells. *J Virol* **2019**, *93*, e00664-19, doi:10.1128/JVI.00664-19.
36. Zimmerman, M.G.; Bowen, J.R.; McDonald, C.E.; Young, E.; Baric, R.S.; Pulendran, B.; Suthar, M.S. STAT5: A Target of Antagonism by Neurotropic Flaviviruses. *Journal of Virology* **2019**, *93*, e00665-19, doi:10.1128/jvi.00665-19.
37. Suen, W.W.; Uddin, M.J.; Prow, N.A.; Bowen, R.A.; Hall, R.A.; Bielefeldt-Ohmann, H. Tissue-Specific Transcription Profile of Cytokine and Chemokine Genes Associated with Flavivirus Control and Non-Lethal Neuropathogenesis in Rabbits. *Virology* **2016**, *494*, 1–14, doi:10.1016/j.virol.2016.03.026.

38. Garcia, M.; Alout, H.; Diop, F.; Damour, A.; Bengue, M.; Weill, M.; Missé, D.; Lévéque, N.; Bodet, C. Innate Immune Response of Primary Human Keratinocytes to West Nile Virus Infection and Its Modulation by Mosquito Saliva. *Front Cell Infect Microbiol* **2018**, *8*, 387, doi:10.3389/fcimb.2018.00387.
39. Saxenaa, V.; Welte, T.; Baob, X.; Xiea, G.; Wanga, J.; Higgsc, S.; Teshd, R.B.; Tian Wanga A Hamster-Derived West Nile Virus Strain Is Highly Attenuated and Induces a Differential Proinflammatory Cytokine Response in Two Murine Cell Lines. *Virus Res.* **2013**, *167*, 179–187, doi:10.1016/j.virusres.2012.04.013.A.
40. Fang, H.; Welte, T.; Zheng, X.; Chang, G.-J.J.; Holbrook, M.R.; Soong, L.; Tian Wang $\Gamma\delta$ T Cells Promote the Maturation of Dendritic Cells During West Nile Virus Infection. *FEMS Immunol Med Microbiol.* **2010**, *59*, 71–80, doi:10.1111/j.1574-695X.2010.00663.x.
41. Wang, T.; Town, T.; Alexopoulou, L.; Anderson, J.F.; Fikrig, E.; Flavell, R.A. Toll-like Receptor 3 Mediates West Nile Virus Entry into the Brain Causing Lethal Encephalitis. *Nature Medicine* **2004**, *10*, 1366–1373, doi:10.1038/nm1140.
42. Uddin, M.J.; Suen, W.W.; Prow, N.A.; Hall, R.A.; Bielefeldt-Ohmann, H. West Nile Virus Challenge Alters the Transcription Profiles of Innate Immune Genes in Rabbit Peripheral Blood Mononuclear Cells. *Frontiers in Veterinary Science* **2015**, *2*, 76, doi:10.3389/fvets.2015.00076.
43. Xiea, G.; Welte, T.; Wanga, J.; Whitemanb, M.C.; Wickerb, J.A.; Saxenaa, V.; Conga, Y.; Barretta, A.D.T.; Wang, T. A West Nile Virus NS4B-P38G Mutant Strain Induces Adaptive Immunity via TLR7-MyD88-Dependent and Independent Signaling Pathways. *Vaccine* **2013**, *31*, 4143–4151, doi:10.1016/j.vaccine.2013.06.093.A.
44. Quick, E.D.; Leser, J.S.; Clarke, P.; Tyler, K.L. Activation of Intrinsic Immune Responses and Microglial Phagocytosis in an Ex Vivo Spinal Cord Slice Culture Model of West Nile Virus Infection. *Journal of Virology* **2014**, *88*, 13005–13014, doi:10.1128/jvi.01994-14.
45. Bai, F.; Town, T.; Qian, F.; Wang, P.; Kamanaka, M.; Connolly, T.M.; Gate, D.; Montgomery, R.R.; Flavell, R.A.; Fikrig, E. IL-10 Signaling Blockade Controls Murine West Nile Virus Infection. *PLoS Pathogens* **2009**, *5*, e1000610, doi:10.1371/journal.ppat.1000610.
46. Saxena, V.; Xie, G.; Li, B.; Farris, T.; Welte, T.; Gong, B.; Boor, P.; Wu, P.; Tang, S.J.; Tesh, R.; et al. A Hamster-Derived West Nile Virus Isolate Induces Persistent Renal Infection in Mice. *PLoS Neglected Tropical Diseases* **2013**, *7*, e2275, doi:10.1371/journal.pntd.0002275.
47. Welte, T.; Reagan, K.; Fang, H.; Machain-Williams, C.; Zheng, X.; Mendell, N.; Chang, G.J.J.; Wu, P.; Blair, C.D.; Wang, T. Toll-like Receptor 7-Induced Immune Response to Cutaneous West Nile Virus Infection. *Journal of General Virology* **2009**, *90*, 2660–2668, doi:10.1099/vir.0.011783-0.

48. Luo, H.; Winkelmann, E.R.; Zhu, S.; Ru, W.; Mays, E.; Silvas, J.A.; Vollmer, L.L.; Gao, J.; Peng, B.H.; Bopp, N.E.; et al. Peli1 Facilitates Virus Replication and Promotes Neuroinflammation during West Nile Virus Infection. *Journal of Clinical Investigation* **2018**, *128*, 4980–4991, doi:10.1172/JCI99902.
49. Town, T.; Bai, F.; Wang, T.; Kaplan, A.; Qian, F.; Montgomery, R.; Anderson, J.; RA, F.; Fikrig, E. Toll-like Receptor 7 Mitigates Lethal West Nile Encephalitis via Interleukin 23-Dependent Immune Cell Infiltration and Homing. *Immunity* **2009**, *30*, 242–253, doi:10.1016/j.cortex.2009.08.003.
50. Graham, J.B.; Swarts, J.L.; Wilkins, C.; Thomas, S.; Green, R.; Sekine, A.; Voss, K.M.; Ireton, R.C.; Mooney, M.; Choonoo, G.; et al. A Mouse Model of Chronic West Nile Virus Disease. *PLoS Pathogens* **2016**, *12*, e1005996, doi:10.1371/journal.ppat.1005996.
51. Bourgeois, M.A.; Denslow, N.D.; Seino, K.S.; Barber, D.S.; Long, M.T. Gene Expression Analysis in the Thalamus and Cerebrum of Horses Experimentally Infected with West Nile Virus. *PLoS ONE* **2011**, *6*, e24371, doi:10.1371/journal.pone.0024371.
52. Acharya, D.; Wang, P.; Paul, A.; Dai, J.; Gate, D.; Lowery, J.; Stokic, D.; Leis, A.; Flavell, R.; Town, T.; et al. Interleukin-17A Promotes CD8+ T Cell Cytotoxicity To Facilitate West Nile Virus Clearance. *J Virol* **2017**, *91*, e01529-16, doi:10.1128/JVI.01529-16.
53. Wang, P.; Bai, F.; Zenewicz, L.A.; Dai, J.; Gate, D.; Cheng, G.; Yang, L.; Qian, F.; Yuan, X.; Montgomery, R.R.; et al. IL-22 Signaling Contributes to West Nile Encephalitis Pathogenesis. *PLoS ONE* **2012**, *7*, e44153, doi:10.1371/journal.pone.0044153.
54. Bryan, M.A.; Giordano, D.; Draves, K.E.; Green, R.; Gale, M.; Clark, E.A. Splenic Macrophages Are Required for Protective Innate Immunity against West Nile Virus. *PLoS ONE* **2018**, *13*, e0191690, doi:10.1371/journal.pone.0191690.

Supplementary table S2: Chemokines involved in West Nile virus pathogenesis.

| Cytokine | Mode | Host | Expression | Source | Role in WNV pathogenesis | Reference |
|-----------------------|-----------------|--------------------------------|----------------------|--|--------------------------|----------------------|
| CC- Chemokines | | | | | | |
| CCL1 | <i>In vivo</i> | Mouse | Downregulation | Brain LAN-2/ U-937/ MΦ/ No change BMDCs/ BMECs/ astrocytes | Unknown | [1] |
| | | | | | Unknown | [2–4] |
| | | Human | Upregulation | Microglia/ U373/ ARPE- 19/RPE | Unknown | [2,5,6] |
| | <i>In vitro</i> | | Strain- dependent | SK-N-SH cells. | Unknown | [7] |
| | Mouse | Upregulation | Microglia/ neurons | Unknown | [8,9] | |
| | | - | - | Mediates monocytes migration and differentiation into DC in the skin and auricular lymph node | [10] | |
| | | | | Not required for survival/ neutralization enhances survival Participates in moncytosis and monocytes trafficking to the brain | [1,9,11–20] | |
| CCL2 | <i>In vivo</i> | Mouse | Upregulation | Serum/ spleen/ lung/ liver/ brain | | |
| | | | | | | |
| | | | | Neurons/ astrocytes/ microglia/ inflammatory cells/ endothelial cells | Unknown | [21] |
| | <i>In vitro</i> | NHP <i>(Macaca mulatta)</i> | Strain- dependent | Brain | Unknown | [22] |
| | | | Upregulation | Brain/spinal cord | Unknown | [23] |
| | | Mouse | Upregulation | Spinal cord slice | Unknown | [24] |
| | | Human | No change | BMDCs | Unknown | [3] |
| CCL3 | <i>In vivo</i> | Mouse | Upregulation | Serum/ brain/spleen | Unknown | [1,9,11,12,14–16,25] |
| | | | Strain- dependent | Brain | Unknown | [22] |
| | NHP | | Upregulation | Brain/spinal cord | Unknown | [23] |

| | | | | | |
|-------------------|-----------------|---|---------------------------|--|--|
| | | | (<i>Macaca mulatta</i>) | | |
| CCL4 | <i>In vivo</i> | Mouse | Upregulation | Spinal cord slice | Unknown [24] |
| | | Human | No change | BMDCs | Unknown [3] |
| | <i>In vitro</i> | Human | Upregulation | NK cells | Unknown [26] |
| | | Mouse | Upregulation | Microglia | Unknown [8] |
| | <i>In vivo</i> | Mouse | Upregulation | Brain/ spleen/ lung | Unknown [9,11–14,16,25] |
| | | Mouse | Strain-dependent | Brain | Unknown [22] |
| | | | No change | BMDCs | Unknown [3] |
| | <i>In vitro</i> | Human | Upregulation | Astrocytes/ Microglia/ ARPE-19/RPE/ SK-N-SH/ MΦ/ BMECs | Unknown [4–7,27] |
| | | | Strain-dependent | THP-1/ THP-1-derived MΦ | Unknown [28] |
| CCL5 | <i>In vivo</i> | Mouse | Upregulation | MΦ/ neurons | Unknown [9,29] |
| | | Mouse | Upregulation | Serum/brain/spleen/ lung | Unknown [1,9,11–16,25] |
| | <i>In vitro</i> | Mouse | Upregulation | Astrogliia | Unknown [21] |
| | | Mouse | Strain-dependent | Brain | Unknown [22] |
| | | NHP (<i>Macaca mulatta</i>) | Upregulation | Brain/spinal cord | Unknown [23] |
| CCL6 ¹ | <i>In vivo</i> | Rabbit (<i>Oryctolagus cuniculus</i>) | Upregulation | Draining lymph nodes | Unknown [30] |
| | | Mouse | Upregulation | Spinal cord slice | Unknown [24] |
| | <i>In vivo</i> | Mouse | Upregulation | Brain | Unknown [13] |
| CCL7 | <i>In vivo</i> | Mouse | Upregulation | Blood/ spleen/ lung/ liver/ brain | Required for efficient moncytosis, recruitment of neutrophils and CD8+ T cells into the CNS, survival after lethal challenge and viral clearance from the brain, not blood or spleen. [1,9,11,15,17,18,20] |
| | | | Strain-dependent | Brain | Unknown [22] |

| | | | | | | |
|--------------------------|-----------------|--|----------------------------------|-------------------------------|---------|-----------|
| | <i>In vitro</i> | Human | Upregulation | MΦ | Unknown | [27] |
| | | Mouse | Upregulation | Brain | Unknown | [1] |
| CCL8 | | Mouse | Strain-dependent | Brain | Unknown | [22] |
| | <i>In vivo</i> | NHP (<i>Macaca mulatta</i>) | Upregulation | Brain/spinal cord | Unknown | [23] |
| CCL9 | <i>In vivo</i> | Mouse | Upregulation | Brain | Unknown | [1] |
| CCL11 | <i>In vivo</i> | Mouse | Upregulation | Lung/ Brain | Unknown | [11,12] |
| CCL12¹ | <i>In vivo</i> | Mouse | Upregulation | Brain | Unknown | [1,13,18] |
| CCL13 | <i>In vivo</i> | Mouse | Upregulation | Brain | Unknown | [16] |
| | | Rabbit (<i>Oryctolagus cuniculus</i>) | Downregulation | Draining lymph nodes | Unknown | [30] |
| | | Mouse | Upregulation | Lung | Unknown | [11] |
| CCL17 | <i>In vivo</i> | Rabbit (<i>Oryctolagus cuniculus</i>) | Downregulation | Draining lymph nodes | Unknown | [30] |
| CCL18 | <i>In vivo</i> | Mouse | Upregulation | Brain | Unknown | [18] |
| | | | No change | Brain | Unknown | [9] |
| CCL19 | <i>In vivo</i> | Mouse | Upregulation Strain-dependent | Spleen/ lung/ brain Brain | Unknown | [1,11] |
| | | | | | Unknown | [22] |
| CCL20 | <i>In vitro</i> | Human | Upregulation | ARPE-19/RPE/ keratinocytes | Unknown | [6,31] |
| | <i>In vivo</i> | Mouse | Downregulation | Brain | Unknown | [1] |
| | | Mouse | No change | Brain | Unknown | [9] |
| | | | Upregulation | Lung | Unknown | [11] |
| CCL21 | <i>In vivo</i> | Rabbit (<i>Oryctolagus cuniculus</i>) | Downregulation | Draining lymph nodes | Unknown | [30] |
| CCL22 | <i>In vivo</i> | Mouse | Upregulation | Lung/kidney | Unknown | [11] |
| CCL24 | <i>In vivo</i> | Mouse | Upregulation Downregulation | Spleen/ lung/ liver Brain | Unknown | [11] |
| | | | | | Unknown | [1] |

| | | | | | | |
|-------------|-----------------|--|----------------|----------------------------------|---|--------------|
| Ccr1 | <i>In vivo</i> | Mouse | Upregulation | Lung/brain | Unknown | [10, 14][14] |
| | | | | | Protects from lethal challenge and promotes viral clearance from the CNS. | |
| Ccr2 | <i>In vivo</i> | Mouse | Upregulation | Lung/brain | Required for monocytosis and accumulation of monocytes in the brain by regulating blood monocyte levels, not trafficking from blood to brain. | [1,11,18] |
| Ccr3 | <i>In vitro</i> | Human | Upregulation | MΦ | Restricts viral replication | [27] |
| | <i>In vivo</i> | Mouse | Upregulation | Brain | Unknown | [1] |
| | | Mouse | Downregulation | Brain | Unknown | [1] |
| Ccr4 | <i>In vivo</i> | Rabbit (<i>Oryctolagus cuniculus</i>) | Downregulation | Draining lymph nodes | Unknown | [30] |
| | | | - | - | Enhances survival following WNV challenge | |
| | | Mouse | | | Controls viral loads in the brain and the BBB permeability | [32] |
| Ccr5 | <i>In vivo</i> | | - | CD4+ and CD8+ T cells, NK, MΦ | Crucial for survival and viral clearance in the brain not the spleen | [14] |
| | | | Upregulation | Brain | Promotes leukocyte trafficking to the CNS | |
| | | Rabbit (<i>Oryctolagus cuniculus</i>) | Upregulation | Draining lymph nodes | Unknown | [14,18] |
| Ccr6 | <i>In vivo</i> | Mouse | Downregulation | Brain | Unknown | [1] |
| | | | | | Required for survival DC and T cell trafficking to the lymph nodes | |
| Ccr7 | <i>In vivo</i> | Mouse | - | Brain | Does not impact leukocyte accumulation or T cell priming in the spleen | [14,32][14, |
| | | | | | Limits the infiltration of WNV-infected myeloid cells into the CNS. | |
| Ccr8 | <i>In vivo</i> | Mouse | Downregulation | Brain | Unknown | [1] |
| | | Rabbit (<i>Oryctolagus</i>) | Downregulation | Draining lymph nodes | Unknown | [30] |

| | | | | | | |
|-------------------------------|-----------------|--------------------------------|-------------------|--|---|-----------------|
| | | | <i>cuniculus)</i> | | | |
| Ccr9 | <i>In vivo</i> | Mouse | Upregulation | Brain | Unknown | [1] |
| CX-Chemokines | | | | | | |
| CXCL1 | <i>In vitro</i> | Human | Upregulation | MΦ/ keratinocytes | Unknown | [31,34] |
| | | Mouse | Upregulation | Neutrophils/ MΦ | Unknown | [34] |
| | | | Upregulation | Serum/ brain | Unknown | [1,12,14,20] |
| | <i>In vivo</i> | Mouse | Strain-dependent | Brain | Unknown | [22] |
| CXCL2 | <i>In vivo</i> | NHP <i>(Macaca mulatta)</i> | Upregulation | Brain/spinal cord | Unknown | [23] |
| | | Mouse | Upregulation | Spinal cord slice | Unknown | [24] |
| | | Human | Upregulation | MΦ/ keratinocytes | Unknown | [31,34] |
| | <i>In vitro</i> | Mouse | Upregulation | Neutrophils/ MΦ | Unknown | [34] |
| CXCL3 | <i>In vivo</i> | Mouse | Upregulation | Lung/ liver/ brain | Unknown | [11–14,16,20] |
| | <i>In vivo</i> | Mouse | Upregulation | Spleen/ kidney | Unknown | [11] |
| | <i>In vivo</i> | Mouse | Upregulation | Brain | Unknown | [14] |
| | <i>In vivo</i> | Mouse | Upregulation | Spleen/ brain | Unknown | [11,12] |
| CXCL8/IL-8² | <i>In vitro</i> | Human | No change | Activated MΦ | Unknown | [35] |
| | | | Upregulation | MΦ/ SK-N-SH/ ARPE-19/RPE/ keratinocytes/ BMDCs/ plasmacytoid DCs | Not responsible for WNV-induced cell death in SK-N-SH | [6,27,31,35–37] |
| | | | Strain-dependent | SK-N-SH | Unknown | [7] |
| | <i>In vivo</i> | NHP <i>(Macaca mulatta)</i> | Upregulation | Brain/spinal cord | Unknown | [23] |
| CXCL9 | <i>In vitro</i> | Human | No change | BMDCs | Unknown | [3] |
| | | Mouse | Upregulation | Neurons | Unknown | [9] |
| | | Mouse | - | - | Not required for survival after lethal challenge | [38] |
| | <i>In vivo</i> | | Upregulation | Spleen/ lung/ liver/ brain | Unknown | [9,11–16,20] |
| | | Horse | Upregulation | Thalamus/ cerebrum | Unknown | [39] |
| | | Rabbit | Upregulation | Draining lymph nodes | Unknown | [30] |

| | | | | | |
|-------------|-------------------------|---|------------------|--|---|
| | (Oryctolagus cuniculus) | | | | |
| CXCL10/IP10 | In vitro | Human | Upregulation | Astrocytes/ Microglia/ THP-1-derived MΦ/ MΦ/ LAN-2/ PBMCs/U373/ U- 937/ ARPE-19/RPE/ SK-N- SH/ keratinocytes/ BMDCs/ Plasmacytoid DCs | Unknown [2,5- 7,27,28,31,34,37,40] |
| | | | Strain-dependent | THP-1 | Unknown [28] |
| | | Mouse | Upregulation | Neutrophils/ MΦ/ microglia | Unknown [8,29,34] |
| | | Rabbit (Oryctolagus cuniculus) | Upregulation | Draining Lymph nodes/ PBMCs | Unknown [30,41] |
| | | | Upregulation | Serum/ spleen/ lung/ liver/ brain | Required for T-lymphocyte recruitment into the CNS, control of viral infection, and survival after WNV lethal challenge [1,9,11- 16,20,25,38,42] |
| | | Mouse | | Neurons/ neutrophils/ astroglia | Unknown [21] |
| | | | Strain-dependent | Brain | Unknown [22] |
| | | Rabbit (Oryctolagus cuniculus and <i>Sylvilagus</i> <i>sp</i>) | Upregulation | Spleen/LNs/ Brain | Unknown [43] |
| | | NHP (<i>Macaca mulatta</i>) | Upregulation | Brain/spinal cord | Unknown [23] |
| | | Horse | Upregulation | Lymphoid tissue/ brain | Unknown [44] |
| CXCL11 | In vivo | Mouse | Upregulation | Spinal cord slice | Unknown [24] |
| | In vitro | Mouse | Upregulation | Neurons | Unknown [9] |
| | In vivo | Mouse | Upregulation | Brain | Unknown [1,15] |
| | | NHP | Upregulation | Brain/spinal cord | Unknown [23] |

| | | | | | | |
|--------------------------------|--|----------------------------------|-----------------------------------|---|---|---------|
| | | (<i>Macaca mulatta</i>) | | | | |
| | Rabbit (<i>Oryctolagus cuniculus</i>) | Upregulation | Draining lymph nodes | Unknown | [30] | |
| | | | | IL-1 β mediates the ability of Cxcr4 expressing lymphocytes to adhere to BMECs via CXCL12 expression on their basolateral surface | | |
| CXCL12 | <i>In vitro</i> | Mouse | - | BMECs | | [45] |
| | <i>In vivo</i> | Mouse | No change | Brain | Unknown | [20,46] |
| | | | Downregulation (β isoform) | Brain | Unknown | [45,46] |
| | | | Upregulation | Brain | Unknown | [22] |
| | <i>In vivo</i> | Mouse | Upregulation | Primary BMECs and astrocytes | CXCL12 expression at the CNS microvasculature is IL-1- mediated and regulates the efficient parenchymal entry of T lymphocytes. | [45] |
| CXCL13 | <i>In vivo</i> | Mouse | No change | Brain/spleen | Unknown | [25] |
| | | | Upregulation | Brain | Unknown | [1,15] |
| | | | Strain-dependent | Brain | Unknown | [22] |
| | | NHP (<i>Macaca mulatta</i>) | Upregulation | Brain/spinal cord | Unknown | [23] |
| CXCL14 | <i>In vivo</i> | Mouse | Upregulation | Brain | Unknown | [25] |
| | | | Strain dependent | Spleen | Unknown | [25] |
| CXCL15 | <i>In vivo</i> | Mouse | Downregulation | Brain | Unknown | [1] |
| CXCL16 | <i>In vivo</i> | Mouse | Strain-dependent | Brain | Unknown | [22] |
| CX-Chemokines receptors | | | | | | |
| Cxcr2 | <i>In vivo</i> | Mouse | - | - | Enhances viremia and death | [34] |
| Cxcr3 | <i>In vitro</i> | Human | Upregulation | MΦ | Restricts viral replication | [27] |
| | | Mouse | No change | Microglia | Unknown | [47] |

| | | | | | | |
|--|----------------|----------------------------------|-----------------------------------|-------------------------------------|--|------------|
| | | | Downregulation | Neurons | Does not influence viral replication. Leads to cell death at early time-points. Expression on neurons via TNFR1 signaling. | [47] |
| | | | - | - | Required for survival after lethal WNV-challenge and for CD8+ T cells control of WNV infection within the cerebellum. | [38] |
| | <i>In vivo</i> | Mouse | Downregulation during early phase | Neurons/ CD8+ T cells | Unknown | [47] |
| | | | Upregulation | Brain | Unknown | [14,18] |
| Cxcr4 | <i>In vivo</i> | Mouse | - | Macrophage/ microglia/ CD8+ T cells | Enhances mortality and viral loads in the brain via downregulating T cells trafficking Increases glial cells activation | [46] |
| | | | Upregulation | Lung/ brain | Unknown | [11,14,16] |
| CX3C Chemokine and its receptor | | | | | | |
| | | Mouse | Upregulation | Brain (microglia/monocytes) | Does not impact survival following WNV challenge. Monocytes (microglial precursor) recruitment to the brain. | [14] |
| CX3CL1 | <i>In vivo</i> | NHP (<i>Macaca mulatta</i>) | Upregulation | Spinal cord | Unknown | [23] |
| | | Mouse | Upregulation | - | Does not impact survival following WNV challenge. | [14,19,33] |
| Cx3cr1 | <i>In vivo</i> | NHP (<i>Macaca mulatta</i>) | Downregulation | Cerebrum/ spinal cord | Unknown | [23] |
| XC chemokine receptor | | | | | | |
| Xcr1 | <i>In vivo</i> | Mouse | Downregulation | Brain | Unknown | [1] |

¹ Present only in mice

² Present only in humans

Cell Lines: H36.12j; *Mus musculus* (macrophage cell line) ATCC CRL-2449™; Renca *Mus musculus* (Kidney epithelial cells line) ATCC CRL-2947™; retinal pigment epithelium cell line (ARPE-19): L929 ATCC CCL-1™; LAN-2: *Mus musculus* neuroblastoma cell line (RRID:CVCL_1829); U-937: *Homo sapiens* Lymphoma cell line CRL-1593.2™; U373: *Homo sapiens* glioblastoma astrocytoma cell-line; SK-N-SH *Homo sapiens* neuroblastoma cell line ATCC HTB-11™; THP-1: Monocytic leukemia cells ATCC TIB-202™.

Abbreviations: ASC: Apoptosis-associated speck-like protein containing C-terminal caspase recruitment domain [CARD]; BBB: Blood-brain barrier; BMDCs: Bone marrow-derived dendritic cells; BMECs: Brain microvasculature endothelial cells; BMECs: Brain microvascular endothelial cells; CNS: Central nervous system; DCs: Dendritic cells; NHP: Non-Human primate; RPE: retinal pigment epithelium (RPE); IFN: Interferon, IRF: IFN regulatory factor 3; MΦ: Macrophages; PBMCs: Peripheral blood mononuclear cells.

References:

1. Kumar, M.; Nerurkar, V.R. Integrated Analysis of MicroRNAs and Their Disease Related Targets in the Brain of Mice Infected with West Nile Virus. *Virology* **2014**, *0*, 143–151, doi:10.1016/j.virol.2014.01.004.
2. Marle, G. Van; Antony, J.; Ostermann, H.; Dunham, C.; Hunt, T.; Halliday, W.; Maingat, F.; Urbanowski, M.D.; Hobman, T.; Peeling, J.; et al. West Nile Virus-Induced Neuroinflammation : Glial Infection and Capsid Protein-Mediated Neurovirulence. *J Virol* **2007**, *81*, 10933–10949, doi:10.1128/JVI.02422-06.
3. Zimmerman, M.G.; Bowen, J.R.; McDonald, C.E.; Pulendran, B.; Suthar, M.S. West Nile Virus Infection Blocks Inflammatory Response and T Cell Costimulatory Capacity of Human Monocyte-Derived Dendritic Cells. *J Virol* **2019**, *93*, e00664-19, doi:10.1128/JVI.00664-19.
4. Hussmann, K.L.; Fredericksen, B.L. Differential Induction of CCL5 by Pathogenic and Non-Pathogenic Strains of West Nile Virus in Brain Endothelial Cells and Astrocytes. *Journal of General Virology* **2014**, *95*, 862–867, doi:10.1099/vir.0.060558-0.
5. Cheeran, M.C.J.; Hu, S.; Sheng, W.S.; Rashid, A.; Peterson, P.K.; Lokengard, J.R. Differential Responses of Human Brain Cells to West Nile Virus Infection. *Journal of NeuroVirology* **2005**, *11*, 512–524, doi:10.1080/13550280500384982.
6. Munoz-erazo, L.; Natoli, R.; Provis, J.M.; Madigan, M.C.; Jonathan, N.; King, C. Microarray Analysis of Gene Expression in West Nile Virus – Infected Human Retinal Pigment Epithelium. *Molecular Vision* **2012**, *18*, 730–743.
7. Huang, B.; West, N.; Vider, J.; Zhang, P.; Griffiths, R.E.; Wolvetang, E.; Burtonclay, P.; Warrilow, D. Inflammatory Responses to a Pathogenic West Nile Virus Strain. *BMC Infectious Diseases* **2019**, *19*, 912, doi:10.1186/s12879-019-4471-8.

8. Luo, H.; Winkelmann, E.R.; Zhu, S.; Ru, W.; Mays, E.; Silvas, J.A.; Vollmer, L.L.; Gao, J.; Peng, B.H.; Bopp, N.E.; et al. Peli1 Facilitates Virus Replication and Promotes Neuroinflammation during West Nile Virus Infection. *Journal of Clinical Investigation* **2018**, *128*, 4980–4991, doi:10.1172/JCI99902.
9. Klein, R.S.; Lin, E.; Zhang, B.; Luster, A.D.; Tollett, J.; Samuel, M.A.; Engle, M.; Diamond, M.S. Neuronal CXCL10 Directs CD8 + T-Cell Recruitment and Control of West Nile Virus Encephalitis . *Journal of Virology* **2005**, *79*, 11457–11466, doi:10.1128/jvi.79.17.11457-11466.2005.
10. Davison, A.M.; Nicholas J C King Accelerated Dendritic Cell Differentiation from Migrating Ly6C Lo Bone Marrow Monocytes in Early Dermal West Nile Virus Infection. *Journal of Immunology* **2011**, *186*, 2382–2396, doi:10.4049/jimmunol.1002682.
11. Peña, J.; Plante, J.A.; Carillo, A.C.; Roberts, K.K.; Smith, J.K.; Juelich, T.L.; Beasley, D.W.C.; Freiberg, A.N.; Labute, M.X.; Naraghi-Arani, P. Multiplexed Digital mRNA Profiling of the Inflammatory Response in the West Nile Swiss Webster Mouse Model. *PLoS Neglected Tropical Diseases* **2014**, *8*, e3216, doi:10.1371/journal.pntd.0003216.
12. Kumar, M.; Roe, K.; O'Connell, M.; Nerurkar, V.R. Induction of Virus-Specific Effector Immune Cell Response Limits Virus Replication and Severe Disease in Mice Infected with Non-Lethal West Nile Virus Eg101 Strain. *Journal of Neuroinflammation* **2015**, *12*, 178, doi:10.1186/s12974-015-0400-y.
13. Garcia-tapia, D.; Hassett, D.E.; Mitchell Jr, W.J.; Johnson, G.C.; Kleiboeker, S.B. West Nile Virus Encephalitis : Sequential Histopathological and Immunological Events in a Murine Model of Infection. *Journal of NeuroVirology* **2007**, *13*, 130–138, doi:10.1080/13550280601187185.
14. Glass, W.G.; Lim, J.K.; Cholera, R.; Pletnev, A.G.; Gao, J.L.; Murphy, P.M. Chemokine Receptor CCR5 Promotes Leukocyte Trafficking to the Brain and Survival in West Nile Virus Infection. *Journal of Experimental Medicine* **2005**, *202*, 1087–1098, doi:10.1084/jem.20042530.
15. Clarke, P.; Leser, J.S.; Bowen, R.A.; Tylera, K.L. Virus-Induced Transcriptional Changes in the Brain Include the Differential Expression of Genes Associated with Interferon, Apoptosis, Interleukin 17 Receptor A, and Glutamate Signaling as Well as Flavivirus-Specific Upregulation of tRNA Synthetases. *mBio* **2014**, *5*, e00902-14, doi:10.1128/mBio.00902-14.
16. Lim, S.M.; van den Ham, H.J.; Oduber, M.; Martina, E.; Zaaraoui-Boutahar, F.; Roose, J.M.; van IJcken, W.F.J.; Osterhaus, A.D.M.E.; Andeweg, A.C.; Koraka, P.; et al. Transcriptomic Analyses Reveal Differential Gene Expression of Immune and Cell Death Pathways in the Brains of Mice Infected with West Nile Virus and Chikungunya Virus. *Frontiers in Microbiology* **2017**, *8*, 1556, doi:10.3389/fmicb.2017.01556.
17. Bardina, S. v; Michlmay, D.; Hoffman, K.W.; Obara, C.J.; Sum, J.; Charo, I.F.; Lu, W.; Pletnev, A.G.; Lim, J.K. Differential Roles of Chemokines CCL2 and CCL7 in Monocytosis and Leukocyte Migration during West Nile Virus Infection. *Physiol Behav* **2015**, *195*, 4306–4318, doi:10.4049/jimmunol.1500352.
18. Lim, J.K.; Obara, C.J.; Rivollier, A.; Pletnev, A.G.; Kelsall, B.L.; Murphy, P.M. Chemokine Receptor CCR2 Is Critical for Monocyte Accumulation and Survival in West Nile Virus Encephalitis. *J Immunol* **2011**, *186*, 471–478, doi:10.4049/jimmunol.1003003.

19. Getts, D.R.; Terry, R.L.; Getts, M.T.; Marcus, M.; Rana, S.; Shrestha, B.; Radford, J.; Rooijen, N. van; Campbell, I.L.; King, N.J.C. Ly6c+ "Inflammatory Monocytes" Are Microglial Precursors Recruited in a Pathogenic Manner in West Nile Virus Encephalitis. *J. Exp. Med.* **2008**, *205*, 2319–2337, doi:10.1084/jem.20080421.
20. Michlmayr, D.; McKimmie, C.S.; Pingen, M.; Haxton, B.; Mansfield, K.; Johnson, N.; Fooks, A.R.; Graham, G.J. Defining the Chemokine Basis for Leukocyte Recruitment during Viral Encephalitis. *Journal of Virology* **2014**, *88*, 9553–9567, doi:10.1128/jvi.03421-13.
21. Vidaña, B.; Johnson, N.; Fooks, A.R.; Sánchez-Cordón, P.J.; Hicks, D.J.; Nuñez, A. West Nile Virus Spread and Differential Chemokine Response in the Central Nervous System of Mice: Role in Pathogenic Mechanisms of Encephalitis. *Transbound Emerg Dis* **2020**, *67*, 799–810, doi:10.1111/tbed.13401.
22. Kumar, M.; Belcaid, M.; Nerurkar, V.R. Identification of Host Genes Leading to West Nile Virus Encephalitis in Mice Brain Using RNA-Seq Analysis. *Scientific Reports* **2016**, *6*, 26350, doi:10.1038/srep26350.
23. Maximova, O.A.; Sturdevant, D.E.; Kash, J.C.; Kanakabandi, K.; Xiao, Y.; Minai, M.; Moore, I.N.; Taubenberger, J.; Martens, C.; Cohen, J.I.; et al. Virus Infection of the CNS Disrupts the Immune-Neural-Synaptic Axis via Induction of Pleiotropic Gene Regulation of Host Responses. *eLife* **2021**, *10*, e62273, doi:10.7554/eLife.62273.
24. Quick, E.D.; Leser, J.S.; Clarke, P.; Tyler, K.L. Activation of Intrinsic Immune Responses and Microglial Phagocytosis in an Ex Vivo Spinal Cord Slice Culture Model of West Nile Virus Infection. *Journal of Virology* **2014**, *88*, 13005–13014, doi:10.1128/jvi.01994-14.
25. Shirato, K.; Kimura, T.; Mizutani, T.; Kariwa, H.; Takashima, I. Different Chemokine Expression in Lethal and Non-Lethal Murine West Nile Virus Infection. *Journal of Medical Virology* **2004**, *74*, 507–513, doi:10.1002/jmv.20205.
26. Yao, Y.; Strauss-albee, D.M.; Zhou, J.Q.; Malawista, A.; Garcia, M.N.; Murray, K.O.; Blish, C.A.; Montgomery, R.R. The Natural Killer Cell Response to West Nile Virus in Young and Old Individuals with or without a Prior History of Infection. *PLoS ONE* **2017**, *12*, e0172625, doi:10.1371/journal.pone.0172625.
27. Qian, F.; Chung, L.; Zheng, W.; Bruno, V.; Alexander, R.P.; Wang, Z.; Wang, X.; Kurscheid, S.; Zhao, H.; Fikrig, E.; et al. Identification of Genes Critical for Resistance to Infection by West Nile Virus Using RNA-Seq Analysis. *Viruses* **2013**, *5*, 1664–1681, doi:10.3390/v5071664.
28. Xie, G.; Luo, H.; Tian, B.; Mann, B.; Bao, X.; McBride, J.; Tesh, R.; Barrett, A.D.; Wang, T. A West Nile Virus NS4B-P38G Mutant Strain Induces Cell Intrinsic Innate Cytokine Responses in Human Monocytic and Macrophage Cells. *Vaccine* **2015**, *33*, 869–878, doi:10.1016/j.vaccine.2014.12.056.A.
29. Shirato, K.; Miyoshi, H.; Kariwa, H.; Takashima, I. The Kinetics of Proinflammatory Cytokines in Murine Peritoneal Macrophages Infected with Envelope Protein-Glycosylated or Non-Glycosylated West Nile Virus. *Virus Research* **2006**, *121*, 11–16, doi:10.1016/j.virusres.2006.03.010.
30. Suen, W.W.; Imoda, M.; Thomas, A.W.; Nasir, N.N.B.M.; Tearsing, N.; Wang, W.; Bielefeldt-Ohmann, H. An Acute Stress Model in New Zealand White Rabbits Exhibits Altered Immune Response to Infection with West Nile Virus. *Pathogens* **2019**, *8*, 195, doi:10.3390/pathogens8040195.

31. Garcia, M.; Alout, H.; Diop, F.; Damour, A.; Bengue, M.; Weill, M.; Missé, D.; Lévéque, N.; Bodet, C. Innate Immune Response of Primary Human Keratinocytes to West Nile Virus Infection and Its Modulation by Mosquito Saliva. *Front Cell Infect Microbiol* **2018**, *8*, 387, doi:10.3389/fcimb.2018.00387.
32. Durrant, D.M.; Daniels, B.P.; Pasieka, T.; Dorsey, D.; Klein, R.S. CCR5 Limits Cortical Viral Loads during West Nile Virus Infection of the Central Nervous System. *Journal of Neuroinflammation* **2015**, *12*, 233, doi:10.1186/s12974-015-0447-9.
33. Bardina, S. v; Brown, J.A.; Michlmayr, D.; Hoffman, K.W. Chemokine Receptor Ccr7 Restricts Fatal West Nile Virus Encephalitis. *J Virol* **2017**, *91*, e02409-16, doi:10.1128/JVI.02409-16.
34. Bai, F.; Kong, K.; Dai, J.; Qian, F.; Zhang, L.; Brown, C.R.; Fikrig, E.; Montgomery, R.R. A Paradoxical Role for Neutrophils in the Pathogenesis of West Nile Virus. *Journal of Infectious Diseases* **2010**, *8031*, 1804–1812, doi:10.1086/657416.
35. Kong, K.; Wang, X.; Anderson, J.; Fikrig, E.; RR Montgomery West Nile Virus Attenuates Activation of Primary Human Macrophages. *Viral Immunol* **2008**, *21*, 78–82, doi:10.1089/vim.2007.0072.
36. Kumar, M.; Verma, S.; Nerurkar, V.R. Pro-Inflammatory Cytokines Derived from West Nile Virus (WNV)-Infected SK-N-SH Cells Mediate Neuroinflammatory Markers and Neuronal Death. *Journal of Neuroinflammation* **2010**, *7*, 73, doi:10.1186/1742-2094-7-73.
37. Silva, M.C.; Guerrero-plata, A.; Gilfoy, F.D.; Garofalo, R.P.; Mason, P.W. Differential Activation of Human Monocyte-Derived and Plasmacytoid Dendritic Cells by West Nile Virus Generated in Different Host Cells □. *Journal of Virology* **2007**, *81*, 13640–13648, doi:10.1128/JVI.00857-07.
38. Zhang, B.; Chan, Y.K.; Lu, B.; Diamond, M.S.; Klein, R.S. CXCR3 Mediates Region-Specific Antiviral T Cell Trafficking within the Central Nervous System during West Nile Virus Encephalitis. *The Journal of Immunology* **2008**, *180*, 2641–2649, doi:10.4049/jimmunol.180.4.2641.
39. Bourgeois, M.A.; Denslow, N.D.; Seino, K.S.; Barber, D.S.; Long, M.T. Gene Expression Analysis in the Thalamus and Cerebrum of Horses Experimentally Infected with West Nile Virus. *PLoS ONE* **2011**, *6*, e24371, doi:10.1371/journal.pone.0024371.
40. Qian, F.; Goel, G.; Meng, H.; Wang, X.; You, F.; Devine, L.; Raddassi, K.; Garcia, M.N.; Murray, K.O.; Bolen, C.R.; et al. Systems Immunology Reveals Markers of Susceptibility to West Nile Virus Infection. *Clin Vaccine Immunol* **2015**, *22*, 6–16, doi:10.1128/CVI.00508-14.
41. Uddin, M.J.; Suen, W.W.; Prow, N.A.; Hall, R.A.; Bielefeldt-Ohmann, H. West Nile Virus Challenge Alters the Transcription Profiles of Innate Immune Genes in Rabbit Peripheral Blood Mononuclear Cells. *Frontiers in Veterinary Science* **2015**, *2*, 76, doi:10.3389/fvets.2015.00076.
42. Suthar, M.S.; Brassil, M.M.; Blahnik, G.; McMillan, A.; Ramos, H.J.; Proll, S.C.; Belisle, S.E.; Katze, M.G.; Gale, M. A Systems Biology Approach Reveals That Tissue Tropism to West Nile Virus Is Regulated by Antiviral Genes and Innate Immune Cellular Processes. *PLoS Pathogens* **2013**, *9*, e1003168, doi:10.1371/journal.ppat.1003168.

43. Suen, W.W.; Uddin, M.J.; Prow, N.A.; Bowen, R.A.; Hall, R.A.; Bielefeldt-Ohmann, H. Tissue-Specific Transcription Profile of Cytokine and Chemokine Genes Associated with Flavivirus Control and Non-Lethal Neuropathogenesis in Rabbits. *Virology* **2016**, *494*, 1–14, doi:10.1016/j.virol.2016.03.026.
44. Bielefeldt-ohmann, H.; Bosco-lauth, A.; Hartwig, A.; Uddin, M.J.; Barcelon, J.; Suen, W.W.; Wang, W.; Hall, R.A.; Bowen, R.A. Microbial Pathogenesis Characterization of Non-Lethal West Nile Virus (WNV) Infection in Horses: Subclinical Pathology and Innate Immune Response. *Microbial Pathogenesis* **2017**, *103*, 71–79, doi:10.1016/j.micpath.2016.12.018.
45. Durrant, D.M.; Daniels, B.P.; Klein, R.S. IL-1R1 Signaling Regulates CXCL12-Mediated T Cell Localization and Fate within the CNS during West Nile Virus Encephalitis. *J Immunol* **2014**, *193*, 4095–4106, doi:10.4049/jimmunol.1401192.
46. McCandless, E.E.; Zhang, B.; Diamond, M.S.; Klein, R.S. CXCR4 Antagonism Increases T Cell Trafficking in the Central Nervous System and Improves Survival from West Nile Virus Encephalitis. *Proc Natl Acad Sci U S A* **2008**, *105*, 11270–11275, doi:10.1073/pnas.0800898105.
47. Zhang, B.; Patel, J.; Croyle, M.; Diamond, M.S.; Klein, R.S. TNF- α -Dependent Regulation of CXCR3 Expression Modulates Neuronal Survival during West Nile Virus Encephalitis. *Journal of Neuroimmunology* **2010**, *224*, 28–38, doi:10.1016/j.jneuroim.2010.05.003.

Supplementary table S3: Tumor necrosis factor superfamily ligands involved in West Nile virus pathogenesis.

| Cytokine | Model | Host | Expression | Source | Role in WNV pathogenesis | Reference |
|---------------|-------------------------------|-------|------------------|---|--|---------------|
| TNF- α | Human | | No change | Astrocytes/ | Unknown | [1] |
| | | | Upregulation | Microglia/ Neural stem cells/ BMDCs/ SK-N-SH/ M Φ / ARPE-19/RPE/ keratinocytes | WNV infection induces antiviral indoleamine via TNF α . Neurotoxic | [1-9] |
| | | | Strain-dependent | THP-1/ THP-1-derived M Φ | Unknown | [10] |
| | Pig (<i>Sus domesticus</i>) | | Upregulation | BMDCs | Unknown | [3] |
| | | | | | Downregulates CXCR3 expression on neurons via TNFR1 signaling. | |
| | In vitro | Mouse | Upregulation | Splenic and brain CD8+ T cell /neurons/ M Φ / DCs/ H36.12j/ Renca/ $\gamma\delta$ T cells/ microglia | Enhances neurons survival at early time-points. Reduces viral replication in M Φ and fibroblasts, not neurons and DCs. | [11-17] |
| | | | | | Not required to upregulate MHC-I in fibroblasts/ Does not activate NF- κ B during WNV infection of fibroblasts | |
| | | | Strain-dependent | Peritoneal exudate cells | Expression may be induced by WNV envelope protein | [18] |
| | | | | M Φ | Unknown | [19] |
| | In vivo | Mouse | - | Microglia/ N2a and primary cultures of murine cortical neurons | Production in microglia mediated by TLR3. Direct neuronal injury. | [20] |
| | | | | | Variable effects on viral replication and survival after WNV challenge. Controls leukocyte infiltration in the CNS/ down-regulates neuronal CXCR3 and subsequent neuronal apoptosis/ enhances BBB permeability. | |
| | | | Upregulation | Blood/liver/ spleen/ kidney/ brain | Expression positively regulated by SARM and TLR3 in the brain. Does not affect the early antibody responses and CD8+ T cells priming in the spleen after WNV infection. | [13,18,20-29] |

| | | | | |
|-----------------|--|---|---|---|
| | | | TNF-/- mice developed severe limbic seizures like wild type mice. Does not affect Langerhans cell migration and accumulation in the draining lymph nodes and viral entry into the CNS | [30–32] |
| | | Strain-dependent | Brain | Unknown [33] |
| | Rabbit (<i>Oryctolagus cuniculus</i> and <i>Sylvilagus sp.</i>) | Upregulation | PBMCs/ brain/ lymph nodes | Unknown [34,35] |
| | Horse | Upregulation | Lymphoid tissue | Unknown [36] |
| | NHP (<i>Macaca mulatta</i>) | Upregulation | Brain/spinal cord | Unknown [37] |
| <i>In vivo</i> | Mouse | Upregulation | Spinal cord slice | Unknown [38] |
| | | - | Cortical neurons | CD8+ T cells require TRAIL to optimally control WNV infection in neurons. [39] |
| <i>In vitro</i> | Mouse | Negatively correlated to replication rate | L929 | Unknown [40] |
| | | | Protected mice from lethal infection and enhanced viral clearance from the CNS. Reduced viral replication in the CNS and not in the serum or spleen when footpad- infected, not intracranially-infected. | [39] |
| | | | Did not affect trafficking of leukocytes to the brain neither adaptive B nor T cell responses induction after WNV infection. | |
| TRAIL | <i>In vivo</i> | Upregulation | Brain | Unknown [28] |
| | | Strain-dependent | Brain | Unknown [33] |
| | | Negatively correlated to replication rate | L929 | Unknown [40] |
| | Rabbit (<i>Oryctolagus cuniculus</i>) | Upregulation | Draining lymph nodes | Unknown [30] |
| <i>In vivo</i> | Mouse | Upregulation | Spinal cord slice | Unknown [38] |

| | | | | | | |
|--------------|-----------------|----------------------------------|--------------|-------------------|---|------------|
| | <i>In vitro</i> | Mouse | Upregulation | Cortical neurons | Used by CD8+ T cells to control neuronal infection. | [41] |
| FasL | <i>In vivo</i> | Mouse | Upregulation | Lung/brain | Protects mice from lethal WNV infection Required for the clearance of WNV infection from the CNS | [23,33,41] |
| | | - | - | - | Does not protect mice from lethal infection | [42] |
| | <i>In vivo</i> | NHP (<i>Macaca mulatta</i>) | Upregulation | Brain/spinal cord | Unknown | [37] |
| CD40L | <i>In vivo</i> | Mouse | - | - | Protects from lethal WNV infection | |
| | | | | | Required for efficient production of neutralizing antibodies, trafficking of CD8+ T cells into the brain, and control of WNV replication in the CNS not the serum and spleen. | [43] |
| OX40L | <i>In vitro</i> | Human | Upregulation | Brain | Unknown | [28] |
| TWEAK | <i>In vivo</i> | Mouse | Upregulation | BMDCs | Unknown | [44] |
| LIGHT | <i>In vitro</i> | Human | Upregulation | Brain | Unknown | [45] |
| | <i>In vivo</i> | Mouse | Upregulation | A172 | Unknown | [46] |
| BAFF | <i>In vivo</i> | Mouse | - | - | Not essential to control virus replication at early stages of WNV infection, but they are critical for viral clearance from sera, spleen and brain | [47] |
| | | | | | BAFF from DCs, not neutrophils, is required for protective immunity to WNV | [48] |

Cell Lines: H36.12j; *Mus musculus* macrophage cell line ATCC CRL-2449; Renca *Mus musculus* Kidney epithelial cells line ATCC CRL-2947; ARPE-19 : *Homo sapiens* Retinal pigment epithelium cell line CRL-2302; L929 : *Mus musculus* fibroblastic cell line ATCC CCL-1; LAN-2: *Mus musculus* neuroblastoma cell line (RRID:CVCL_1829); U-937: *Homo sapiens* Lymphoma cell line CRL-1593.2; U373: *Homo sapiens* glioblastoma astrocytoma cell-line; SK-N-SH *Homo sapiens* neuroblastoma cell line ATCC HTB-11; THP-1: Monocytic leukemia cells ATCC TIB-202.

Abbreviations: BAFF: B-cell activating factor; BBB: Blood-brain barrier; BMDCs: Bone marrow-derived dendritic cells; BMECs: Brain microvasculature endothelial cells; BMECs: Brain microvascular endothelial cells; CNS: Central nervous system; DCs: Dendritic cells; RPE: retinal pigment epithelium (RPE), IFN: Interferon, IRF: IFN regulatory factor 3; MΦ: Macrophages; NF-κB: Nuclear factor Kappa B; NHP: Non-Human primate; PBMCs: Peripheral blood mononuclear cells; SARM: Sterile alpha and HEAT/Armadillo motif; TLR: Toll-like receptor 3; TRAIL: TNF-related apoptosis-inducing ligand; TWEAK: TNF-related weak inducer of apoptosis.

References:

1. Cheeran, M.C.J.; Hu, S.; Sheng, W.S.; Rashid, A.; Peterson, P.K.; Lokensgard, J.R. Differential Responses of Human Brain Cells to West Nile Virus Infection. *Journal of NeuroVirology* **2005**, *11*, 512–524, doi:10.1080/13550280500384982.
2. Riccetti, S.; Sinigaglia, A.; Desole, G.; Nowotny, N.; Trevisan, M.; Barzon, L. Modelling West Nile Virus and Usutu Virus Pathogenicity in Human Neural Stem Cells. *Viruses* **2020**, *12*, 882, doi:10.3390/v12080882.
3. García-Nicolás, O.; Lewandowska, M.; Ricklin, M.E.; Summerfield, A. Monocyte-Derived Dendritic Cells as Model to Evaluate Species Tropism of Mosquito-Borne Flaviviruses. *Frontiers in Cellular and Infection Microbiology* **2019**, *9*, 5, doi:10.3389/fcimb.2019.00163.
4. Kumar, M.; Verma, S.; Nerurkar, V.R. Pro-Inflammatory Cytokines Derived from West Nile Virus (WNV)-Infected SK-N-SH Cells Mediate Neuroinflammatory Markers and Neuronal Death. *Journal of Neuroinflammation* **2010**, *7*, 73, doi:10.1186/1742-2094-7-73.
5. Yeung, A.W.S.; Wu, W.; Freewan, M.; Stocker, R.; King, N.J.C.; Thomas, S.R. Flavivirus Infection Induces Indoleamine Macrophages via Tumor Necrosis Factor and NF-Kappa B. *Journal of Leukocyte Biology* **2012**, *91*, 657–666, doi:10.1189/jlb.1011532.
6. Martina, B.E.E.; Koraka, P.; Doel, P. van den; Rimmelzwaan, G.F.; Haagmans, B.L.; Osterhaus, A.D.M.E. DC-SIGN Enhances Infection of Cells with Glycosylated West Nile Virus in Vitro and Virus Replication in Human Dendritic Cells Induces Production of IFN- α and TNF- α . *Virus Research* **2008**, *135*, 64–71, doi:10.1016/j.virusres.2008.02.008.
7. Munoz-erazo, L.; Natoli, R.; Provis, J.M.; Madigan, M.C.; Jonathan, N.; King, C. Microarray Analysis of Gene Expression in West Nile Virus – Infected Human Retinal Pigment Epithelium. *Molecular Vision* **2012**, *18*, 730–743.
8. Qian, F.; Chung, L.; Zheng, W.; Bruno, V.; Alexander, R.P.; Wang, Z.; Wang, X.; Kurscheid, S.; Zhao, H.; Fikrig, E.; et al. Identification of Genes Critical for Resistance to Infection by West Nile Virus Using RNA-Seq Analysis. *Viruses* **2013**, *5*, 1664–1681, doi:10.3390/v5071664.
9. Garcia, M.; Alout, H.; Diop, F.; Damour, A.; Bengue, M.; Weill, M.; Missé, D.; Lévéque, N.; Bodet, C. Innate Immune Response of Primary Human Keratinocytes to West Nile Virus Infection and Its Modulation by Mosquito Saliva. *Front Cell Infect Microbiol* **2018**, *8*, 387, doi:10.3389/fcimb.2018.00387.
10. Xie, G.; Luo, H.; Tian, B.; Mann, B.; Bao, X.; McBride, J.; Tesh, R.; Barrett, A.D.; Wang, T. A West Nile Virus NS4B-P38G Mutant Strain Induces Cell Intrinsic Innate Cytokine Responses in Human Monocytic and Macrophage Cells. *Vaccine* **2015**, *33*, 869–878, doi:10.1016/j.vaccine.2014.12.056.A.
11. Aguilar-valenzuela, R.; Netland, J.; Seo, Y.; Bevan, M.J.; Grakoui, A. Dynamics of Tissue-Specific CD8+ T Cell Responses during West Nile Virus Infection. *J Virol* **2018**, *92*, e00014-18, doi:10.1128/JVI.00014-18.
12. Zhang, B.; Patel, J.; Croyle, M.; Diamond, M.S.; Klein, R.S. TNF- α -Dependent Regulation of CXCR3 Expression Modulates Neuronal Survival during West Nile Virus Encephalitis. *Journal of Neuroimmunology* **2010**, *224*, 28–38, doi:10.1016/j.jneuroim.2010.05.003.

13. Shrestha, B.; Zhang, B.; Purtha, W.E.; Klein, R.S.; Diamond, M.S. Tumor Necrosis Factor Alpha Protects against Lethal West Nile Virus Infection by Promoting Trafficking of Mononuclear Leukocytes into the Central Nervous System. *Journal of Virology* **2008**, *82*, 8956–8964, doi:10.1128/jvi.01118-08.
14. Cheng, Y.; King, N.J.C.; Kesson, A.M. The Role of Tumor Necrosis Factor in Modulating Responses of Murine Embryo Fibroblasts by Flavivirus , West Nile B. *Virology* **2004**, *329*, 361–370, doi:10.1016/j.virol.2004.06.050.
15. Saxenaa, V.; Welte, T.; Baob, X.; Xiea, G.; Wangaa, J.; Higgsc, S.; Teshd, R.B.; Tian Wangaa A Hamster-Derived West Nile Virus Strain Is Highly Attenuated and Induces a Differential Proinflammatory Cytokine Response in Two Murine Cell Lines. *Virus Res.* **2013**, *167*, 179–187, doi:10.1016/j.virusres.2012.04.013.A.
16. Fang, H.; Welte, T.; Zheng, X.; Chang, G.-J.J.; Holbrook, M.R.; Soong, L.; Tian Wangaa T Cells Promote the Maturation of Dendritic Cells During West Nile Virus Infection. *FEMS Immunol Med Microbiol.* **2010**, *59*, 71–80, doi:10.1111/j.1574-695X.2010.00663.x.
17. Luo, H.; Winkelmann, E.R.; Zhu, S.; Ru, W.; Mays, E.; Silvas, J.A.; Vollmer, L.L.; Gao, J.; Peng, B.H.; Bopp, N.E.; et al. Peli1 Facilitates Virus Replication and Promotes Neuroinflammation during West Nile Virus Infection. *Journal of Clinical Investigation* **2018**, *128*, 4980–4991, doi:10.1172/JCI99902.
18. Sapkal, G.N.; Harini, S.; Ayachit, V.M.; Fulmali, P. v; Mahamuni, S.A.; Bondre, V.P.; Gore, M.M. Neutralization Escape Variant of West Nile Virus Associated with Altered Peripheral Pathogenicity and Differential Cytokine Profile. *Virus Research* **2011**, *158*, 130–139, doi:10.1016/j.virusres.2011.03.023.
19. Shirato, K.; Miyoshi, H.; Kariwa, H.; Takashima, I. The Kinetics of Proinflammatory Cytokines in Murine Peritoneal Macrophages Infected with Envelope Protein-Glycosylated or Non-Glycosylated West Nile Virus. *Virus Research* **2006**, *121*, 11–16, doi:10.1016/j.virusres.2006.03.010.
20. Wang, T.; Town, T.; Alexopoulou, L.; Anderson, J.F.; Fikrig, E.; Flavell, R.A. Toll-like Receptor 3 Mediates West Nile Virus Entry into the Brain Causing Lethal Encephalitis. *Nature Medicine* **2004**, *10*, 1366–1373, doi:10.1038/nm1140.
21. Xiea, G.; Welte, T.; Wangaa, J.; Whitemanb, M.C.; Wickerb, J.A.; Saxenaa, V.; Conga, Y.; Barretta, A.D.T.; Wang, T. A West Nile Virus NS4B-P38G Mutant Strain Induces Adaptive Immunity via TLR7-MyD88-Dependent and Independent Signaling Pathways. *Vaccine* **2013**, *31*, 4143–4151, doi:10.1016/j.vaccine.2013.06.093.A.
22. Suthar, M.S.; Brassil, M.M.; Blahnik, G.; McMillan, A.; Ramos, H.J.; Proll, S.C.; Belisle, S.E.; Katze, M.G.; Gale, M. A Systems Biology Approach Reveals That Tissue Tropism to West Nile Virus Is Regulated by Antiviral Genes and Innate Immune Cellular Processes. *PLoS Pathogens* **2013**, *9*, e1003168, doi:10.1371/journal.ppat.1003168.
23. Peña, J.; Plante, J.A.; Carillo, A.C.; Roberts, K.K.; Smith, J.K.; Juelich, T.L.; Beasley, D.W.C.; Freiberg, A.N.; Labute, M.X.; Naraghi-Arani, P. Multiplexed Digital mRNA Profiling of the Inflammatory Response in the West Nile Swiss Webster Mouse Model. *PLoS Neglected Tropical Diseases* **2014**, *8*, e3216, doi:10.1371/journal.pntd.0003216.
24. Kumar, M.; Roe, K.; O'Connell, M.; Nerurkar, V.R. Induction of Virus-Specific Effector Immune Cell Response Limits Virus Replication and Severe Disease in Mice Infected with Non-Lethal West Nile Virus Eg101 Strain. *Journal of Neuroinflammation* **2015**, *12*, 178, doi:10.1186/s12974-015-0400-y.

25. Szretter, K.J.; Samuel, M.A.; Gilfillan, S.; Fuchs, A.; Colonna, M.; Diamond, M.S. The Immune Adaptor Molecule SARM Modulates Tumor Necrosis Factor Alpha Production and Microglia Activation in the Brainstem and Restricts West Nile Virus Pathogenesis. *Journal of Virology* **2009**, *83*, 9329–9338, doi:10.1128/jvi.00836-09.
26. Garcia-tapia, D.; Hassett, D.E.; Mitchell Jr, W.J.; Johnson, G.C.; Kleiboeker, S.B. West Nile Virus Encephalitis : Sequential Histopathological and Immunological Events in a Murine Model of Infection. *Journal of NeuroVirology* **2007**, *13*, 130–138, doi:10.1080/13550280601187185.
27. Glass, W.G.; Lim, J.K.; Cholera, R.; Pletnev, A.G.; Gao, J.L.; Murphy, P.M. Chemokine Receptor CCR5 Promotes Leukocyte Trafficking to the Brain and Survival in West Nile Virus Infection. *Journal of Experimental Medicine* **2005**, *202*, 1087–1098, doi:10.1084/jem.20042530.
28. Kumar, M.; Nerurkar, V.R. Integrated Analysis of MicroRNAs and Their Disease Related Targets in the Brain of Mice Infected with West Nile Virus. *Virology* **2014**, *0*, 143–151, doi:10.1016/j.virol.2014.01.004.
29. Lim, S.M.; van den Ham, H.J.; Oduber, M.; Martina, E.; Zaaraoui-Boutahar, F.; Roose, J.M.; van IJcken, W.F.J.; Osterhaus, A.D.M.E.; Andeweg, A.C.; Koraka, P.; et al. Transcriptomic Analyses Reveal Differential Gene Expression of Immune and Cell Death Pathways in the Brains of Mice Infected with West Nile Virus and Chikungunya Virus. *Frontiers in Microbiology* **2017**, *8*, 1556, doi:10.3389/fmicb.2017.01556.
30. Getts, D.R.; Matsumoto, I.; Mu, M.; Getts, T.; Radford, J.; Shrestha, B.; Campbell, I.L.; King, N.J.C. Role of IFN- γ in an Experimental Murine Model of West Nile Virus-Induced Seizures. *J of Neurochemistry* **2007**, *103*, 1019–1030, doi:10.1111/j.1471-4159.2007.04798.x.
31. Shrestha, B.; Wang, T.; Samuel, M.A.; Whitby, K.; Craft, J.; Fikrig, E.; Diamond, M.S. Gamma Interferon Plays a Crucial Early Antiviral Role in Protection against West Nile Virus Infection. *Journal of Virology* **2006**, *80*, 5338–5348, doi:10.1128/jvi.00274-06.
32. Byrne, S.N.; Halliday, G.M.; Johnston, L.J.; King, N.J.C. Interleukin-1 β but Not Tumor Necrosis Factor Is Involved in West Nile Virus-Induced Langerhans Cell Migration from the Skin in C57BL/6 Mice. *Journal of Investigative Dermatology* **2001**, *117*, 702–709, doi:10.1046/j.0022-202x.2001.01454.x.
33. Kumar, M.; Belcaid, M.; Nerurkar, V.R. Identification of Host Genes Leading to West Nile Virus Encephalitis in Mice Brain Using RNA-Seq Analysis. *Scientific Reports* **2016**, *6*, 26350, doi:10.1038/srep26350.
34. Uddin, M.J.; Suen, W.W.; Prow, N.A.; Hall, R.A.; Bielefeldt-Ohmann, H. West Nile Virus Challenge Alters the Transcription Profiles of Innate Immune Genes in Rabbit Peripheral Blood Mononuclear Cells. *Frontiers in Veterinary Science* **2015**, *2*, 76, doi:10.3389/fvets.2015.00076.
35. Suen, W.W.; Uddin, M.J.; Prow, N.A.; Bowen, R.A.; Hall, R.A.; Bielefeldt-Ohmann, H. Tissue-Specific Transcription Profile of Cytokine and Chemokine Genes Associated with Flavivirus Control and Non-Lethal Neuropathogenesis in Rabbits. *Virology* **2016**, *494*, 1–14, doi:10.1016/j.virol.2016.03.026.

36. Bielefeldt-ohmann, H.; Bosco-lauth, A.; Hartwig, A.; Uddin, M.J.; Barcelon, J.; Suen, W.W.; Wang, W.; Hall, R.A.; Bowen, R.A. Microbial Pathogenesis Characterization of Non-Lethal West Nile Virus (WNV) Infection in Horses: Subclinical Pathology and Innate Immune Response. *Microbial Pathogenesis* **2017**, *103*, 71–79, doi:10.1016/j.micpath.2016.12.018.
37. Maximova, O.A.; Sturdevant, D.E.; Kash, J.C.; Kanakabandi, K.; Xiao, Y.; Minai, M.; Moore, I.N.; Taubenberger, J.; Martens, C.; Cohen, J.I.; et al. Virus Infection of the Cns Disrupts the Immune-Neural-Synaptic Axis via Induction of Pleiotropic Gene Regulation of Host Responses. *eLife* **2021**, *10*, e62273, doi:10.7554/eLife.62273.
38. Quick, E.D.; Leser, J.S.; Clarke, P.; Tyler, K.L. Activation of Intrinsic Immune Responses and Microglial Phagocytosis in an Ex Vivo Spinal Cord Slice Culture Model of West Nile Virus Infection. *Journal of Virology* **2014**, *88*, 13005–13014, doi:10.1128/jvi.01994-14.
39. Shrestha, B.; Pinto, A.K.; Green, S.; Bosch, I.; Diamond, M.S. CD8+ T Cells Use TRAIL To Restrict West Nile Virus Pathogenesis by Controlling Infection in Neurons. *Journal of Virology* **2012**, *86*, 8937–8948, doi:10.1128/jvi.00673-12.
40. O’Neal, J.T.; Upadhyay, A.A.; Wolabaugh, A.; Patel, N.B.; Bosinger, S.E.; Suthar, M.S. West Nile Virus-Inclusive Single-Cell RNA Sequencing Reveals Heterogeneity in the Type I Interferon Response within Single Cells. *J Virol* **2019**, *93*, e01778-18, doi:10.1128/JVI.01778-18.
41. Shrestha, B.; Diamond, M.S. Fas Ligand Interactions Contribute to CD8 + T-Cell-Mediated Control of West Nile Virus Infection in the Central Nervous System . *Journal of Virology* **2007**, *81*, 11749–11757, doi:10.1128/jvi.01136-07.
42. Arreberg, L.D.; Wilkins, C.; Ramos, H.J.; Green, R.; Davis, M.A.; Chow, K.; Gale, M. Interleukin-1 β Signaling in Dendritic Cells Induces Antiviral Interferon Responses. *mBio* **2018**, *9*, e00342-18, doi:10.1128/mBio .00342-18.
43. Sandip, S.; Lal, G. Role of Tumor Necrosis Factor Superfamily in Neuroinflammation and Autoimmunity. *Front. Immunol* **2015**, *6*, 364, doi:10.3389/fimmu.2015.00364.
44. Kovats, S.; Turner, S.; Simmons, A.; Powe, T.; Chakravarty, E.; Alberola-Illa, J. West Nile Virus-Infected Human Dendritic Cells Fail to Fully Activate Invariant Natural Killer T Cells. *Clinical and Experimental Immunology* **2016**, *186*, 214–226, doi:10.1111/cei.12850.
45. Clarke, P.; Leser, J.S.; Bowen, R.A.; Tylera, K.L. Virus-Induced Transcriptional Changes in the Brain Include the Differential Expression of Genes Associated with Interferon, Apoptosis, Interleukin 17 Receptor A, and Glutamate Signaling as Well as Flavivirus-Specific Upregulation of tRNA Synthetases. *mBio* **2014**, *5*, e00902-14, doi:10.1128/mBio.00902-14.
46. Koh, W.L.; Ng, M.L. Molecular Mechanisms of West Nile Virus Pathogenesis in Brain Cells. *Emerging Infectious Diseases* **2005**, *11*, 629–632, doi:10.3201/eid1104.041076.
47. Giordano, D.; Draves, K.E.; Young, L.B.; Roe, K.; Bryan, M.A.; Dresch, C.; Richner, J.M.; Diamond, M.S.; Gale, M.; Clark, E.A. Protection of Mice Deficient in Mature B Cells from West Nile Virus Infection by Passive and Active Immunization. *PLoS Pathog* **2017**, *13*, e1006743, doi:10.1371/journal.ppat.1006743.

48. Giordano, D.; Kuley, R.; Draves, K.E.; Roe, K.; Holder, U.; Giltay, V.; Clark, E.A. B Cell Activating Factor (BAFF) Produced by Neutrophils and Dendritic Cells Is Regulated Differently and Has Distinct Roles in Ab Responses and Protective Immunity against West Nile Virus. *J Immunol.* **2020**, *204*, 1508–1520, doi:10.4049/jimmunol.1901120.B.