

Supplementary Information

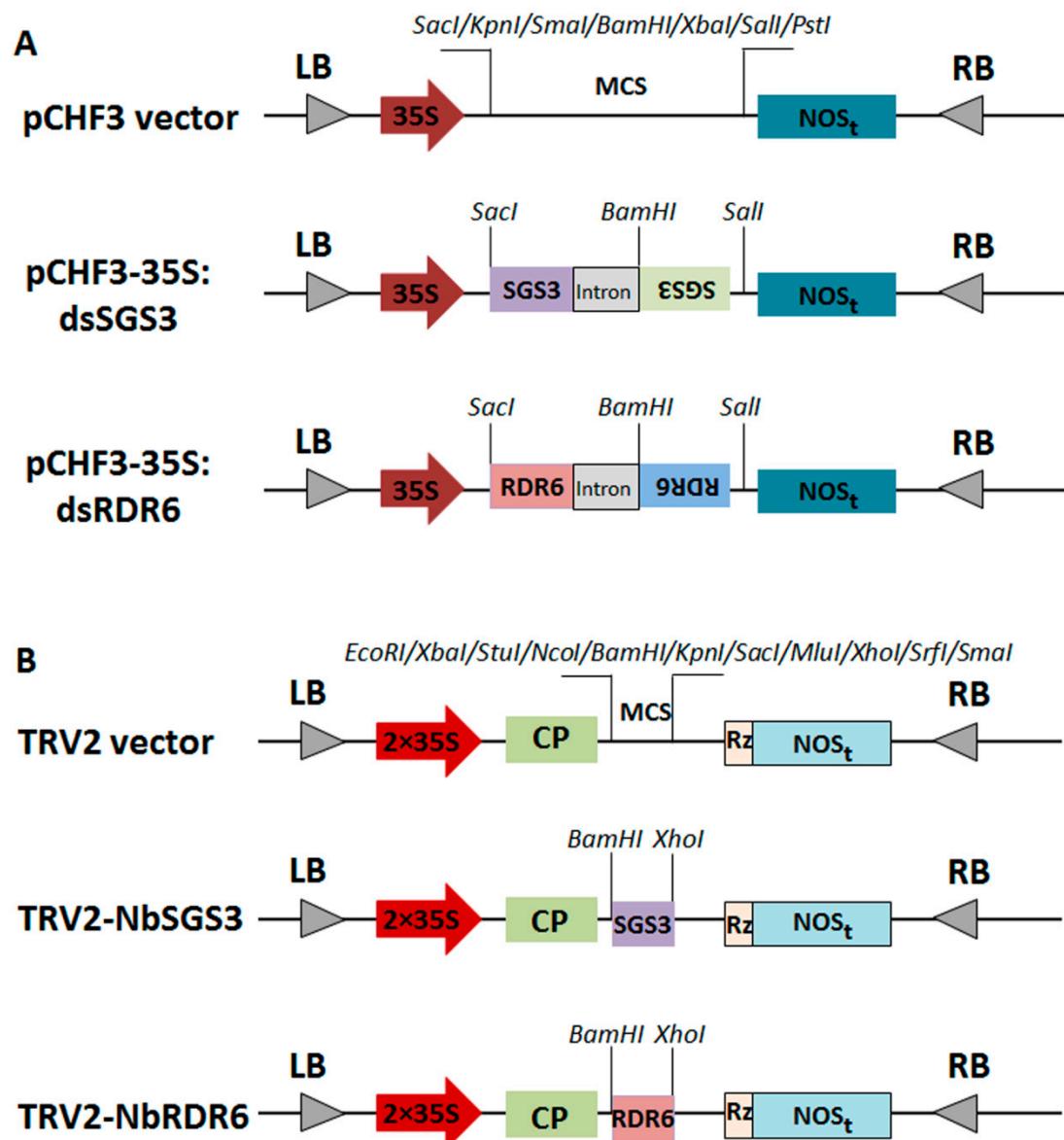


Figure S1. The schematic diagrams of constructs used in this study. (A) Schematic representation of RNAi constructs containing an inverted repeat sequence of *NbSGS3* or *NbRDR6* separated by an *Arabidopsis* intron. LB: left border, 35S: 35S promoter of cauliflower mosaic virus (CaMV), MCS: multiple cloning sites, NOS_t: termination sequence of the nopaline synthase gene; RB: right border. (B) Schematic representation of TRV-VIGS constructs carrying a partial fragment of *NbSGS3* or *NbRDR6* sequences. 2×35S: two tandem 35S promoters, CP: coat protein, Rz: ribozyme.

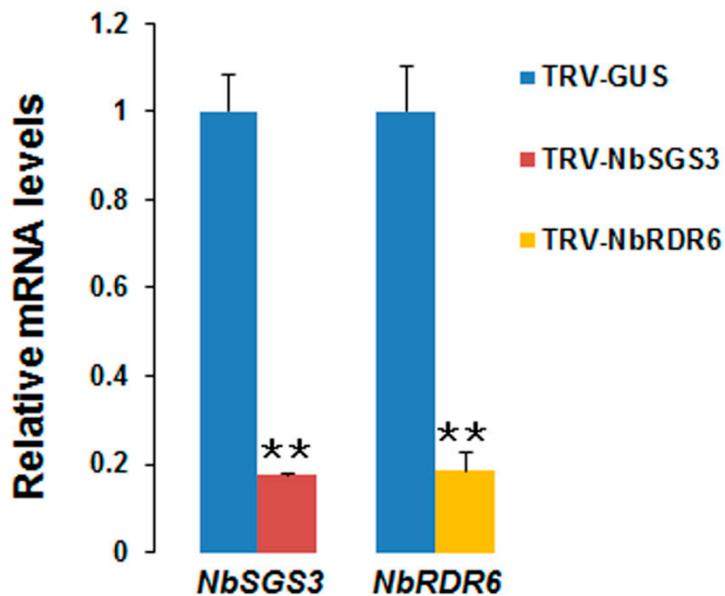


Figure S2. The silencing efficiency of *NbSGS3* or *NbRDR6* in newly emerged leaves at 7 days post infiltration. *NbSGS3* mRNA levels were reduced by approximate 83% in the plants infiltrated with *NbSGS3*-silencing vectors when compared to the plants infiltrated with TRV-GUS (control). *NbRDR6* mRNA levels were reduced by approximate 81% in the plants infiltrated with *NbRDR6*-silencing vectors when compared to the control. The mRNA level of *NbSGS3* or *NbRDR6* in TRV-GUS infected plants was arbitrarily set to 1. Values represent means relative to the *NbSGS3* or *NbRDR6* levels in TRV-GUS infected plants. Double asterisks indicate $P < 0.01$ between the two treatments (Student's *t* test).

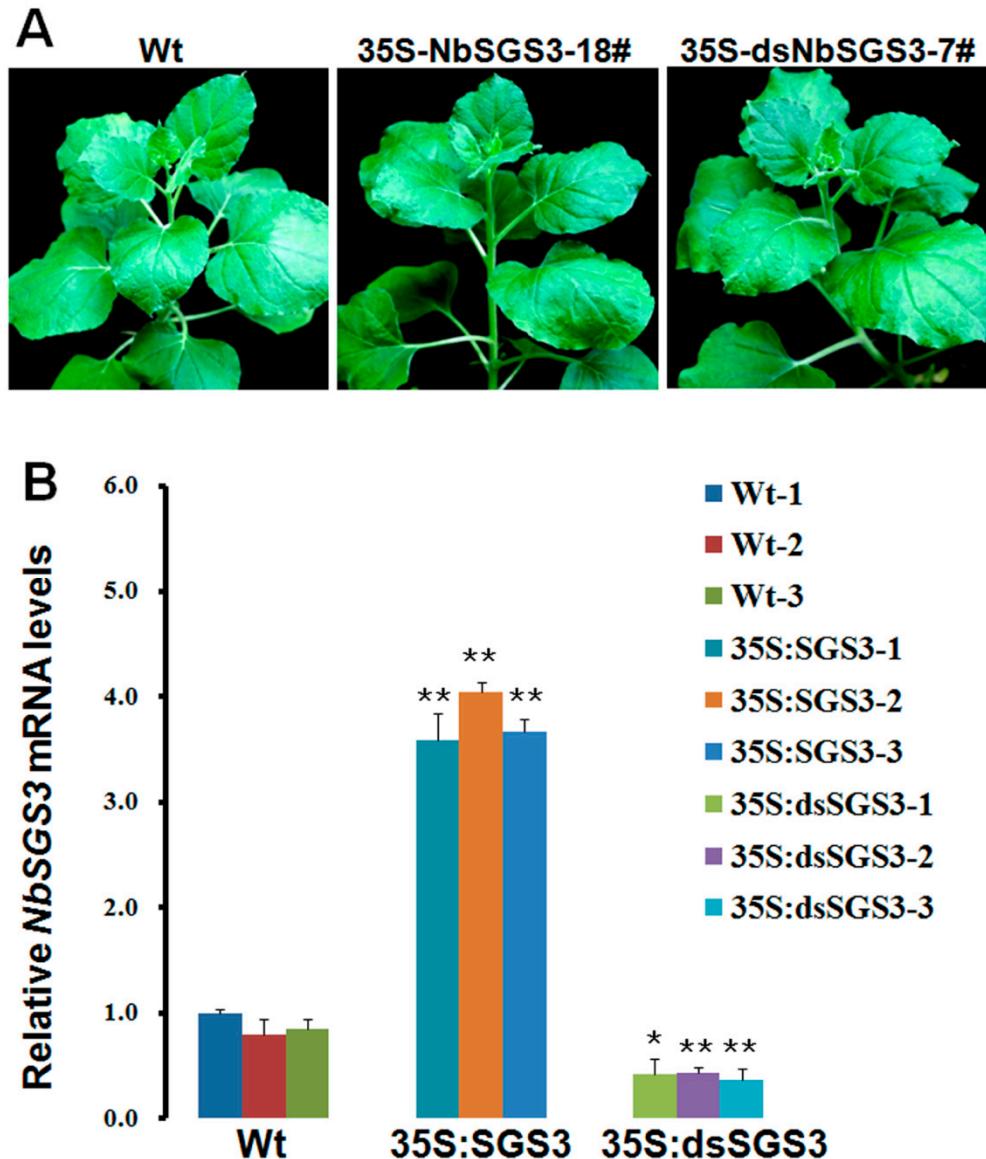


Figure S3. Ectopic expression or knockdown of *NbSGS3* in *N. benthamiana*. (A) Comparison of phenotypes in 40-day-old transgenic *N. benthamiana* plants overexpressing *NbSGS3* (35S-NbSGS3-18#) or silenced for *NbSGS3* (35S:dsNbSGS3-7#) and wild type (Wt) plants. (B) mRNA levels of *NbSGS3* were analyzed by RT-qPCR in 35S:SGS3 and 35S:dsSGS3 transgenic lines at the 6-7 leaf stage in the T1 generation. Three individual plants per phenotype (T1 generation) were used for each of the measurements. Values represent the mean \pm standard deviation (SD). Student's *t* test was performed to compare differences between Wt plants and the transgenic lines, a single asterisk indicates a significant difference ($p < 0.05$) and double asterisks indicate a highly significant difference ($p < 0.01$).

Table1. Primers used in plasmid construction and other experiments in this study.

Primer Name	Primer Sequence (5'-3')	Purpose
NbSGS3:Flag-F	<u>CGGGATCCATGAGTTCAAGCAAAGGGGTTG</u>	pCHF3-NbSGS3: Flag, transient expression and transgene analysis
NbSGS3:Flag-R	<u>GCGTCGACTTGAATTGCTCTGGGAG</u>	
A-NbSGS3-F	TCCGAGCTGAGATGAGGTATACCAAGGAAG	pCHF3-dsNbSGS3, transient expression, transgene
A-NbSGS3-Intro-R	ATCAGACTTACAACGCTCTCACACGATGATCC	pCHF3-dsNbSGS3, transient expression, transgene
B-NbSGS3-Intro-F	GGATCATCGTGTGAGGACGTTGAAGTCTGAT	pCHF3-dsNbSGS3, transient expression, transgene
B-Intron-R	<u>CGGGATCCGCTATCTGCTGGTCCAATC</u>	pCHF3-dsNbSGS3, transient expression, transgene
C-NbSGS3-F	<u>CGGGATCCCTCACACGATGATCCTCAACC</u>	pCHF3-dsNbSGS3, transient expression, transgene
C-NbSGS3-R	<u>GCGTCGACGAGATGAGGTATACCAAGGAAG</u>	pCHF3-dsNbRDR6
A-HpRDR6/Sac I/F	TCCGAGCTCGAAGAACAAAGTTGGCAAGTATGG	
A-HpRDR6+Intro/R	ATCAGACTTACAACGCTCAACCAAAGTCATC	
B-HpRDR6+Intron/F	GATGACTTGTGGTGGACGTTGAAGTCTGAT	
C-HpRDR6/BamH I/F	<u>CGGGATCCCAACACAAAGTCATTTAC</u>	
C-HpRDR6/Sal I/R	<u>GCGTCGACGAAGAACAAAGTTGGCAAGTATGG</u>	
Y10A-F	ATGTGGGATCCTCTGCTCAACGGAGTTTC	To make probe for the detection of TYLCCNV in Southern blot
Y10A-R	CATCCTCAGACCTTGCGTTCTTAAGAG'	
BETA01	GTAGGTACCACTACGCTACGCAGCAGCC	To make probe for the detection of TYLCCNB in Southern blot
BETA02	AGTGGTACCTACCCCTCCCAGGGGTACAC	
TLCYnV-F	ATGCCTCGTCTTAATTCAATTC	To make probe for the detection of TLCYnV in Southern blot
TLCYnV-R	TCAACTCTCCGTCTGCTGG	
TbCSV-F	ATCGATCTGGAAAATTCCATGATC	To make probe for the detection of TbCSV in Southern blot
TbCSV-R	GGATCCCACATAGTGCAGGAG	
q-25S-rRNA-F	ATAACCGCATCAGGTCTCCA	As an internal control for relative quantitative genomic PCR analysis
q-25S-rRNA-R	CCGAAGTTACGGATCCATT	
q-10A-F	TTAGAGATCGTCGCTAGTGG	Relative quantitative PCR analysis of TYLCCNV
q-10A-F	GCTCCTACAAGCATATTGTCC	
q-10b-F	ATACATCATACTCATCCCCTACATCTA	Relative quantitative PCR analysis of TYLCCNB
q-10b-R	ATTATCCACCATTGACTTCAACATT	
q-TLCYnV-F	GGTGCCTGCCGTCTGAACATTG	Relative quantitative PCR analysis of TLCYnV
q-TLCYnV-R	CCAGTATGAGATACATCATGACGGGC	
q-TbCSV-F	TACGCCGCCGTCTCAACTTCGAC	Relative quantitative PCR analysis of TbCSV
q-TbCSV-R	CTTACCTATATGCTGAATGTCATGTC	
35S-F	ACATGGTGGAGCACGACACG	PCR screening transgenic plants with CaMV 35S promoter
35S-R	GAGGAAGGGTCTTGCAGAAGG	
TRV2-NbSGS3-F	<u>CGGGATCCTCCAGCAAAGCAGATGATGG</u>	TRV2-NbSGS3, the silencing of NbSGS3
TRV2-NbSGS3-R	<u>CCGCTCGAGCATCTCGCGTTGCTCCTGC</u>	
TRV2-NbRDR6-F	<u>CGGGATCCGAAGAACAAAGTTGGCAAG</u>	TRV2-NbRDR6, the silencing of NbRDR6
TRV2-NbRDR6-R	<u>CCGCTCGAGCATCTGAAATTAAAGGGC</u>	
NbGADPH-q-F	GCAGTGAACGACCCATTATCTC	Relative qRT-PCR analysis of NbGADPH
NbGADPH -q-R	AACCTTCTGGCACCCACCC	
NbSGS3-q-F	GGGCTCATATTAGGAAAGAACCC	Relative qRT-PCR analysis of NbSGS3
NbSGS3-q-R	GTGAAAAGTATTTGCATGCCCG	
NbRDR6-q-F	CAGATTGAAAACAAAAATCCACCATG	Relative qRT-PCR analysis of NbRDR6
NbRDR6-q-R	ATTTTGAACACCGCTCTGCC	
NbSu-q-F	5'-GCTCTACACCCATTGTCTTCG-3'	Relative qRT-PCR analysis of NbSu
Nb-Su-q-R	5'-CCCTATACCCATTATCATCAC-3'	