

Supplementary materials

Synthesis of Alpha Olefins: Catalytic Decarbonylation of Carboxylic Acids and Vegetable Oil Deodorizer Distillate (VODD)

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1. General Considerations

Unless otherwise noted, all reagents were purchased from commercial suppliers and used without purification [1]. Toluene and THF were freshly distilled from sodium and sodium benzophenone ketyl under nitrogen. All decarbonylation reactions were performed in resealable screw cap Schlenk tube (approx. 15 mL volume) in the presence of Teflon-coated magnetic stirrer bar (4 mm × 10 mm). 2-(2-Bromophenyl)naphthalene and Ligands **L3** – **L9** were purchased from commercial suppliers. New bottle of *n*-butyllithium was used (*Note*: since the concentration of *n*-BuLi may vary, we recommend performing a titration prior to use). Thin layer chromatography was performed on pre-coated silica gel 60 F₂₅₄ plates. Silica gel (Merck, 70-230 and 230-400 mesh) was used for column chromatography. Melting points were recorded on an uncorrected Büchi Melting Point B-545 instrument. NMR spectra were recorded on a Bruker spectrometer (400 MHz for ¹H, 100 MHz for ¹³C and 162 MHz for ³¹P). Spectra were referenced internally to the residual proton resonance in CDCl₃ (δ 7.26 ppm) as the internal standard. Chemical shifts (δ) were reported as part per million (ppm) in δ scale downfield from TMS. ¹³C NMR spectra were referenced to CDCl₃ (δ 77.0 ppm, the middle peak). ³¹P NMR spectra were referenced to 85% H₃PO₄ externally. Coupling constants (*J*) were reported in Hertz (Hz). Mass spectra (EI-MS and ES-MS) were recorded on a HP 5989B Mass Spectrometer. High-resolution mass spectra (HRMS) were obtained on a Bruker APEX 47e FTICR mass spectrometer (ESI-MS). GC-MS analysis was conducted on a HP 5973 GCD system using a HP5MS column (30 m × 0.25 mm). The products described in GC yield were accorded to the authentic samples/dodecane calibration standard from Agilent 6890A GC-FID system. X-ray crystal structure was determined by Bruker D8 Venture. All yields reported refer to isolated yield of compounds estimated to be greater than 95% purity as determined by capillary gas chromatography (GC) or ¹H NMR. Compounds described in the literature were characterized by comparison of their ¹H and/or ¹³C NMR spectra to the previously reported data. The procedures in this section are representative, and thus the yields may differ from those reported in tables.

2. Determination of free fatty acid % (FFA%) in canola oil deodorizer distillate by titrimetric method

0.5–2.0 g of samples (based on the expected acidity) were weighed into Erlenmeyer flasks. Neutralized 2-propanol (50 mL) was added. The flask was placed on a hot plate and the temperature was regulated to about 40 °C. The sample was shaken gently while being titrated against standard KOH (0.1 M) to the first permanent pink colour using phenolphthalein as indicator. The colour must persist for 30 s.

$$\text{FFA\% (in terms of oleic acid)} = \frac{(V-B) \times N \times 28.2}{m}$$

V = volume of potassium hydroxide used in sample.

B = volume of potassium hydroxide used in blank sample.

N = Normality of potassium hydroxide

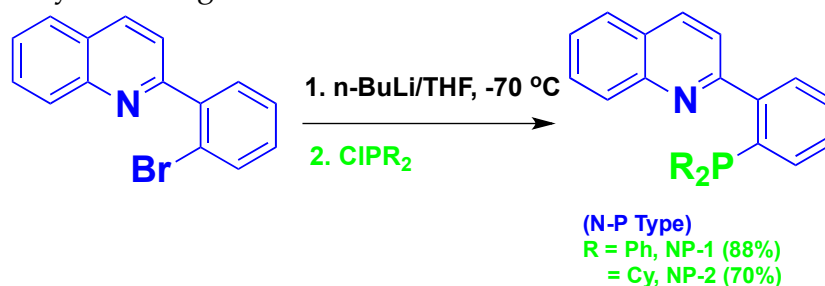
m = weight of sample, in g.

3. Determination of the composition of fatty acids in canola oil deodorizer distillate by GC method

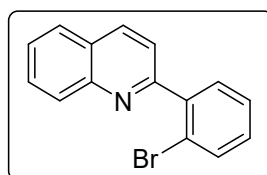
0.1 mL canola oil deodorizer distillate was placed into 10 mL centrifugal tubes to which 2 mL of H₂SO₄–MeOH solution (1 v/v%) was added. The mixture was heated at 70 °C for 1 h. After cooling to room temperature, 3 mL of n-hexane and 2 mL of distilled water were added and mixed thoroughly. The extracts were removed and injected for GC analysis. Peaks are identified against fatty acid methyl esters (FAMES) standard purchased from sigma Aldrich (Supelco® 37 Component FAME Mix). Analysis of FAME products by GC–FID analysis was carried out using an Agilent 6890A. Operating conditions: temperature (°C): injector, 225; detector, 285; initial temp, 100 (hold 4 min); ramp, 3°C/min; final temp 240; hold 15 min; carrier gas, helium; flow rate, 0.75 mL/min; linear velocity, 18 cm/s; split ratio, 200:1. Capillary column.—SP2560 100 m x 0.25 mm with 0.20 µm.

4. Preparation of quinoline-scaffold monophosphine (NP-1, NP-2) and naphthalene-scaffold monophosphine (CP-1, CP-2) ligands

Synthetic pathway for NP ligands

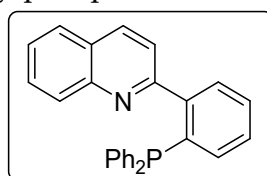


2-(2-Bromophenyl)quinoline



Synthesis of 2-(2-Bromophenyl)quinoline was according to literature procedure.^[2] ^1H NMR (400 MHz, CDCl_3) δ 7.30 (td, $J = 1.6\text{Hz}$, 7.6Hz , 1H), 7.45 (t, $J = 7.5\text{Hz}$, 1H), 7.58 (t, $J = 7.5\text{Hz}$, 1H), 7.65 (dd, $J = 1.5\text{Hz}$, 7.6Hz , 1H), 7.71 (d, $J = 8.8\text{Hz}$, 2H), 7.75 (t, $J = 7.6\text{Hz}$, 1H), 7.87 (d, $J = 8.2\text{Hz}$, 1H) 8.21 (t, $J = 8.3\text{Hz}$, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 121.8, 126.4, 126.8, 127.1, 127.5, 127.7, 129.5, 129.7, 130.0, 131.6, 133.2, 135.7, 141.5, 147.8, 158.6; MS (EI): m/z (relative intensity) 283.1 (M^+ , 50), 204.2 (100), 176.2 (25), 102.1 (32)

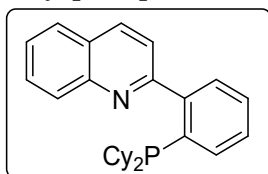
(2-(Quinoline-2-yl)phenyl)diphenylphosphine, **NP-1**



2-(2-Bromophenyl)quinoline (0.849 g, 3.0 mmol) was dissolved in freshly distilled THF (20 mL) at room temperature under a nitrogen atmosphere. The solution was cooled to $-78\text{ }^{\circ}\text{C}$ in dry ice/acetone bath. Titrated $n\text{-BuLi}$ (3.3 mmol) was added dropwise by syringe. After the reaction mixture was stirred for 30 min at $-78\text{ }^{\circ}\text{C}$, chlorodiphenylphosphine (0.66 mL, 3.3 mmol) in THF (5 mL) was added. The reaction was allowed to warm to room temperature and stirred overnight. Solvent was removed under reduced pressure. After

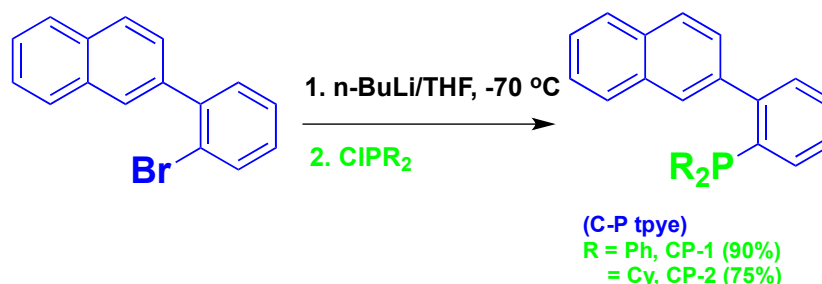
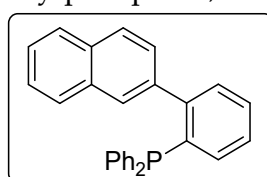
the solvent was removed under vacuum, the product was successively washed with cold MeOH/EtOH mixture. The product was then dried under vacuum. Pale yellow solid of (2-(Quinoline-2-yl)phenyl)diphenylphosphine **NP-1** (1.03 g, 88 %) were obtained. Melting point. 162.6-163.3 °C; ^1H NMR (400 MHz, CD_2Cl_2) δ 7.21 (dd, $J=3.4\text{Hz}$, 7.9Hz , 1H), 7.31-7.42 (m, 11H), 7.55 (t, $J=8.0\text{Hz}$, 2H), 7.59-7.67 (m, 2H), 7.76 (d, $J=8.3\text{Hz}$, 1H), 7.83-7.88 (m, 2H), 8.21 (d, $J=8.3\text{Hz}$, 1H); ^{13}C NMR (100 MHz, CD_2Cl_2) δ 121.3, 126.4, 126.6, 127.4, 128.1 (d, $J=4.0\text{Hz}$), 128.2, 128.5, 128.6, 128.9, 129.3 (d, $J=4.0\text{Hz}$), 129.4, 133.8 (d, $J=19.8\text{Hz}$), 135.4, 135.9, 137.1 (d, $J=18.8\text{Hz}$), 139.4 (d, $J=11.1\text{Hz}$), 145.7 (d, $J=23.3\text{Hz}$), 146.9, 158.3; ^{31}P NMR (162 MHz, CD_2Cl_2) δ -10.86; MS (EI): m/z (relative intensity) 389.3 (M^+ , 23), 312.2 (100), 235.2 (35), 194.7 (7); HRMS: calcd. for $\text{C}_{27}\text{H}_{21}\text{NP}^+$: 390.1412, found 390.1403.

(2-(Quinoline-2-yl)phenyl)dicyclohexylphosphine, **NP-2**

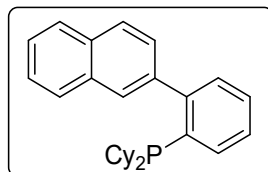


2-(2-Bromophenyl)quinoline (0.849 g, 3.0 mmol) was dissolved in freshly distilled THF (20 mL) at room temperature under a nitrogen atmosphere. The solution was cooled to -78 °C in dry ice/acetone bath. Titrated *n*-BuLi (3.3 mmol) was added dropwise by syringe. After the reaction mixture was stirred for 30 min at -78 °C, chlorodicyclohexylphosphine (0.72 mL, 3.3 mmol) in THF (5 mL) was added. The reaction was allowed to warm to room temperature and stirred overnight. Solvent was removed under reduced pressure. After the solvent was removed under vacuum, the product was successively washed with cold MeOH/EtOH mixture. The product was then dried under vacuum. White solid of (2-(Quinoline-2-yl)phenyl)dicyclohexylphosphine **NP-2** (0.84 g, 70 %) were obtained. Melting point. 136.6-137.2 °C; ^1H NMR (400 MHz, CD_2Cl_2) δ 1.09-1.32 (m, 10H), 1.63-1.76 (m, 10H), 1.99 (td, $J=3.0\text{Hz}$, 12.0Hz , 2H), 7.48-7.52 (m, 2H), 7.56-7.62 (m, 3H), 7.70-7.73 (m, 1H), 7.76 (td, $J=1.3\text{Hz}$, 7.2Hz , 1H), 7.91 (d, $J=8.2\text{Hz}$, 1H), 8.12 (d, $J=8.2\text{Hz}$, 1H), 8.18 (d, $J=8.2\text{Hz}$, 1H); ^{13}C NMR (100 MHz, CD_2Cl_2) δ 26.4, 27.1 (d, $J=7.0\text{Hz}$), 27.2 (d, $J=4.3\text{Hz}$), 29.7 (d, $J=10.5\text{Hz}$), 30.3 (d, $J=17.5\text{Hz}$), 34.5 (d, $J=13.4\text{Hz}$), 124.3 (d, $J=6.6\text{Hz}$), 126.2, 126.7, 127.5 (d, $J=2.7\text{Hz}$), 128.3, 129.3 (d, $J=16.6\text{Hz}$), 129.8 (d, $J=5.0\text{Hz}$), 132.9 (d, $J=2.0\text{Hz}$), 134.2, 134.7 (d, $J=23.6\text{Hz}$), 147.6, 149.3 (d, $J=27.3\text{Hz}$), 161.0 (d, $J=4.9\text{Hz}$); ^{31}P NMR (162 MHz, CD_2Cl_2) δ -11.01; MS (EI): m/z (relative intensity) 401.4 (M^+ , 2), 318.3 (100), 235.2 (43). HRMS: calcd. for $\text{C}_{27}\text{H}_{33}\text{NP}^+$: 402.2351, found 402.2342.

Synthetic pathway for CP ligands

(2-(Naphthalene-2-yl)phenyl)diphenylphosphine, **CP-1**^[3]

2-(2-Bromophenyl)naphthalene (0.849 g, 3.0 mmol) was dissolved in freshly distilled THF (20 mL) at room temperature under a nitrogen atmosphere. The solution was cooled to -78 °C in dry ice/acetone bath. Titrated *n*-BuLi (3.3 mmol) was added dropwise by syringe. After the reaction mixture was stirred for 30 min at -78 °C, chlorodiphenylphosphine (0.66 mL, 3.3 mmol) in THF (5 mL) was added. The reaction was allowed to warm to room temperature and stirred overnight. Solvent was removed under reduced pressure. After the solvent was removed under vacuum, the product was successively washed with cold MeOH/EtOH mixture. The product was then dried under vacuum. White solid of (2-(Naphthalene-2-yl)phenyl)diphenylphosphine **CP-1** (1.05 g, 90 %) were obtained. ¹H NMR (400 MHz, CD₂Cl₂) δ 7.14 (dd, *J* = 3.8 Hz, 7.6 Hz, 1H), 7.22-7.26 (m, 4H), 7.32-7.34 (m, 7H), 7.4-7.51 (m, 5H), 7.57 (s, 1H), 7.63 (d, *J* = 7.6 Hz, 1H), 7.78 (d, *J* = 8.2 Hz, 1H), 7.86 (d, *J* = 8.2 Hz, 1H); ¹³C NMR (100 MHz, CD₂Cl₂) δ 125.8 (d, *J* = 10.0 Hz), 126.8, 127.3, 127.4, 127.8, 127.9 (d, *J* = 2.3 Hz), 128.2, 128.3, 128.4, 128.6, 130 (d, *J* = 5.0 Hz), 132.5 (d, *J* = 29.0 Hz), 133.6, 133.8, 134.1, 137.6, 137.7, 139.2, 148.1 (d, *J* = 27.0 Hz); ³¹P NMR (162 MHz, CD₂Cl₂) δ -13.70; MS (EI): *m/z* (relative intensity) 387.3 (*M*⁺, 100), 309.2 (5), 233.2 (16).

(2-(Naphthalene-2-yl)phenyl)dicyclohexylphosphine, **CP-2**

2-(2-Bromophenyl)naphthalene (0.849 g, 3.0 mmol) was dissolved in freshly distilled THF (20 mL) at room temperature under a nitrogen atmosphere. The solution was cooled to -78 °C in dry ice/acetone bath. Titrated *n*-BuLi (3.3 mmol) was added dropwise by syringe. After the reaction mixture was stirred for 30 min at -78 °C, chlorodicyclohexylphosphine (0.72 mL, 3.3 mmol) in THF (5 mL) was added. The reaction was allowed to warm to room temperature and stirred overnight. Solvent was removed under reduced pressure. After the solvent was removed under vacuum, the product was successively washed with cold MeOH/EtOH mixture. The product was then dried under vacuum. White solid of (2-(Naphthalene-2-yl)phenyl)dicyclohexylphosphine **CP-2** (0.90 g, 75 %) were obtained. Melting point. 128.6-129.8 °C; ¹H NMR (400 MHz, CD₂Cl₂) δ 1.07-1.34 (m, 10H), 1.56-1.72 (m, 10H), 1.90 (t, *J* = 11.4 Hz, 2H), 7.39-7.46 (m, 3H), 7.49-7.55 (m, 3H), 7.69-7.71 (m, 1H), 7.76 (s, 1H), 7.86-7.93 (m, 3H); ¹³C NMR (100 MHz, CD₂Cl₂) δ 26.4, 27.1 (d, *J* = 4.0 Hz), 27.2, 29.4 (d, *J* = 9.3 Hz), 30.6 (d, *J* = 17.5 Hz), 34.9 (d, *J* = 17.4 Hz), 125.8 (d, *J* = 21.9 Hz), 126.2, 126.6, 127.7 (d, *J* = 33.0 Hz), 128.2, 129.0 (d, *J* = 3.0 Hz), 129.6 (d, *J* = 4.9 Hz), 130.2 (d, *J* = 4.9 Hz), 132.1, 132.9, 133.0 (d, *J* = 4.0 Hz), 134.5 (d, *J* = 22.9 Hz), 140.8 (d, *J* = 7.1 Hz), 150.4 (d, *J* = 30.3 Hz); ³¹P NMR (162 MHz, CD₂Cl₂) δ -10.19; MS (EI): *m/z* (relative intensity) 399.4 (M⁺, 100), 317.3 (60), 233.2 (87), 202.2 (16). HRMS: calcd. for C₂₈H₃₄P⁺: 401.2398, found 401.2395.

5. Determination of fatty acid profile of canola oil deodorizer distillate by GC-FID

Figure S1: Determination of fatty acid profile of canola oil deodorizer by H₂SO₄ catalyzed transesterification

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 Sample Name: H2SO4
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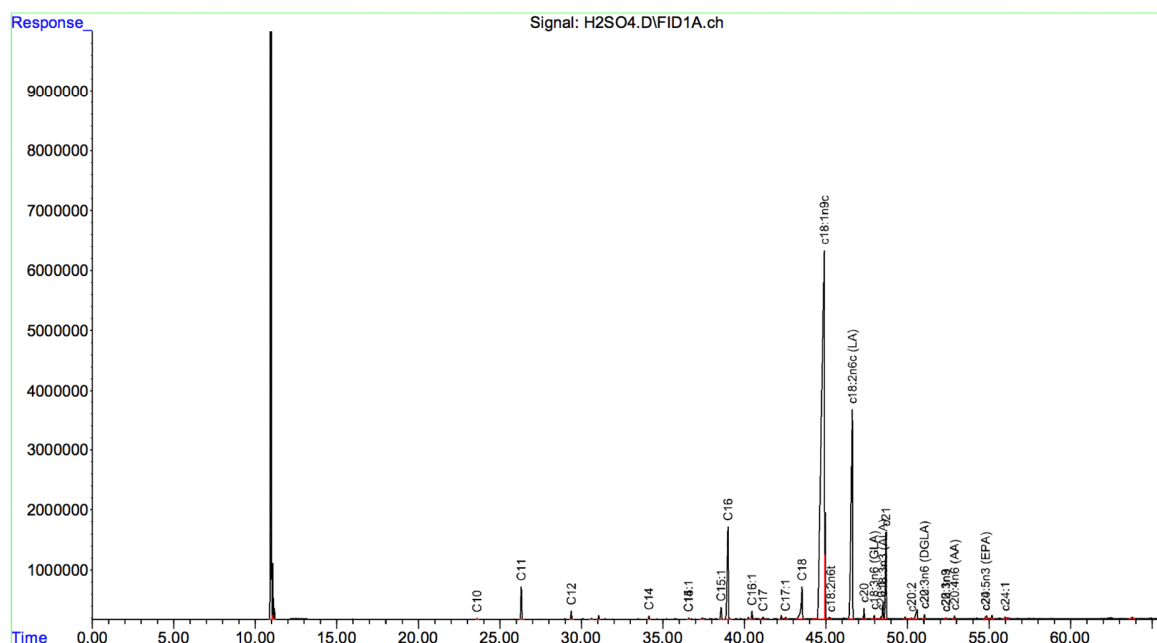


Table S1: Fatty acids composition of canola oil deodorizer distillate determined by GC-FID

	H ₂ SO ₄ catalyzed transesterification		
Fatty acid	Weight of individual fatty acids, mg)	Weight of fatty acids in terms of carbon chain number, mg	Wt% of fatty acids, %
C11	9.36	C11: 9.36	1.05
C12	2.22	C12: 2.22	0.25
C14	0.66	C14: 0.66	0.07
C15	0.23	C15: 3.34	0.38
C15:1	3.11		
C16	25.37	C16: 26.9	3.02
C16:1	1.53		
C17	0.44	C17: 0.73	0.08
C17:1	0.29		
C18	10.25	C18: 372.52	41.86
C18:1 oleic acid	261.94		
C18:2	77.03		
C18:3	3.04		
C18:3	20.26		
C20	1.74	C20: 6.22	0.7
C20:1	2.78		
C20:2	0.20		
C20:3n6	0.82		
C20:3n3	0.24		
C20:4n6	0.44		
Total weight of fatty acid, mg	424.94		
Weight of sample, mg	890.00		
wt% of fatty acid, %	47.75		

Figure S2: Olefin product determined by GC-FID

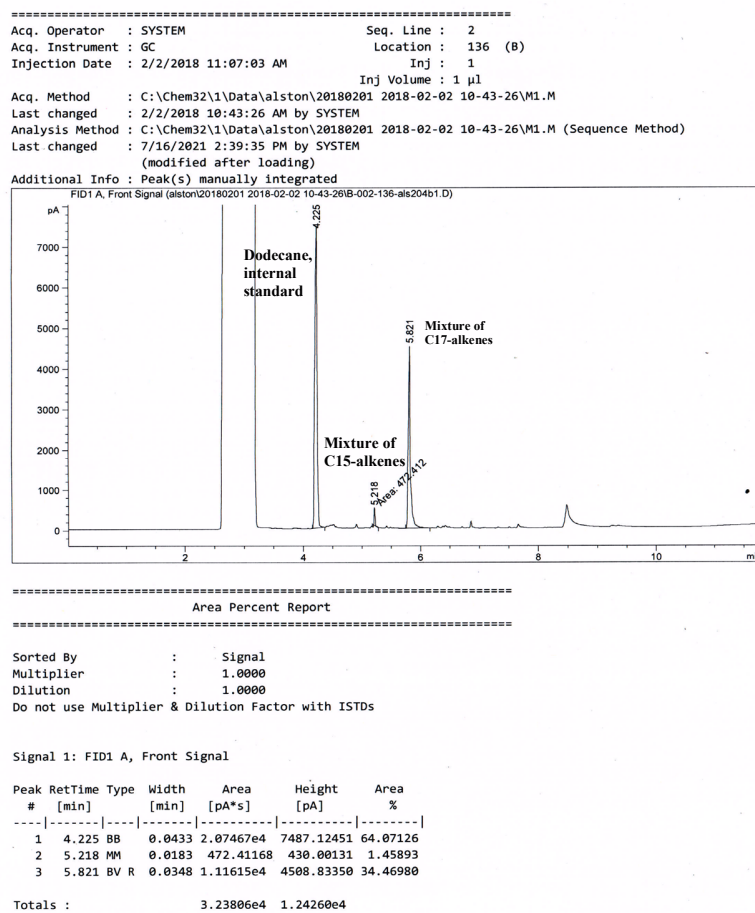


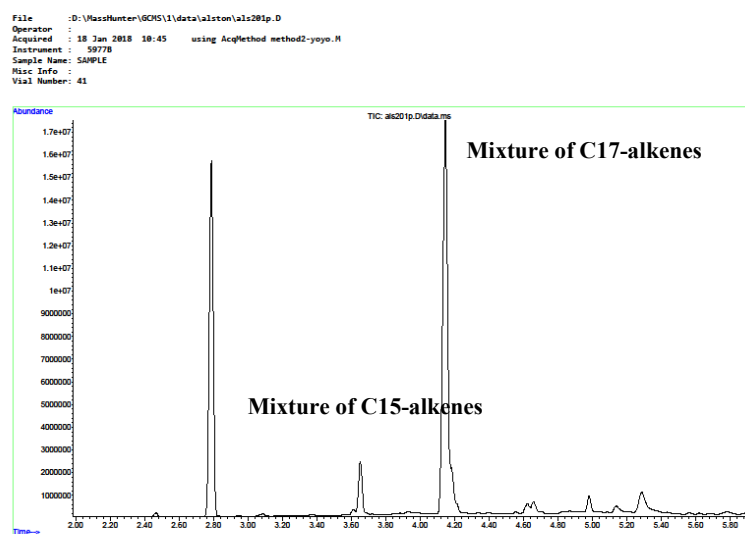
Figure S3: Olefin product determined by GC-MS

Figure S4: GCMS spectrum of mixture of C15-alkenes at retention time 3.6 min.

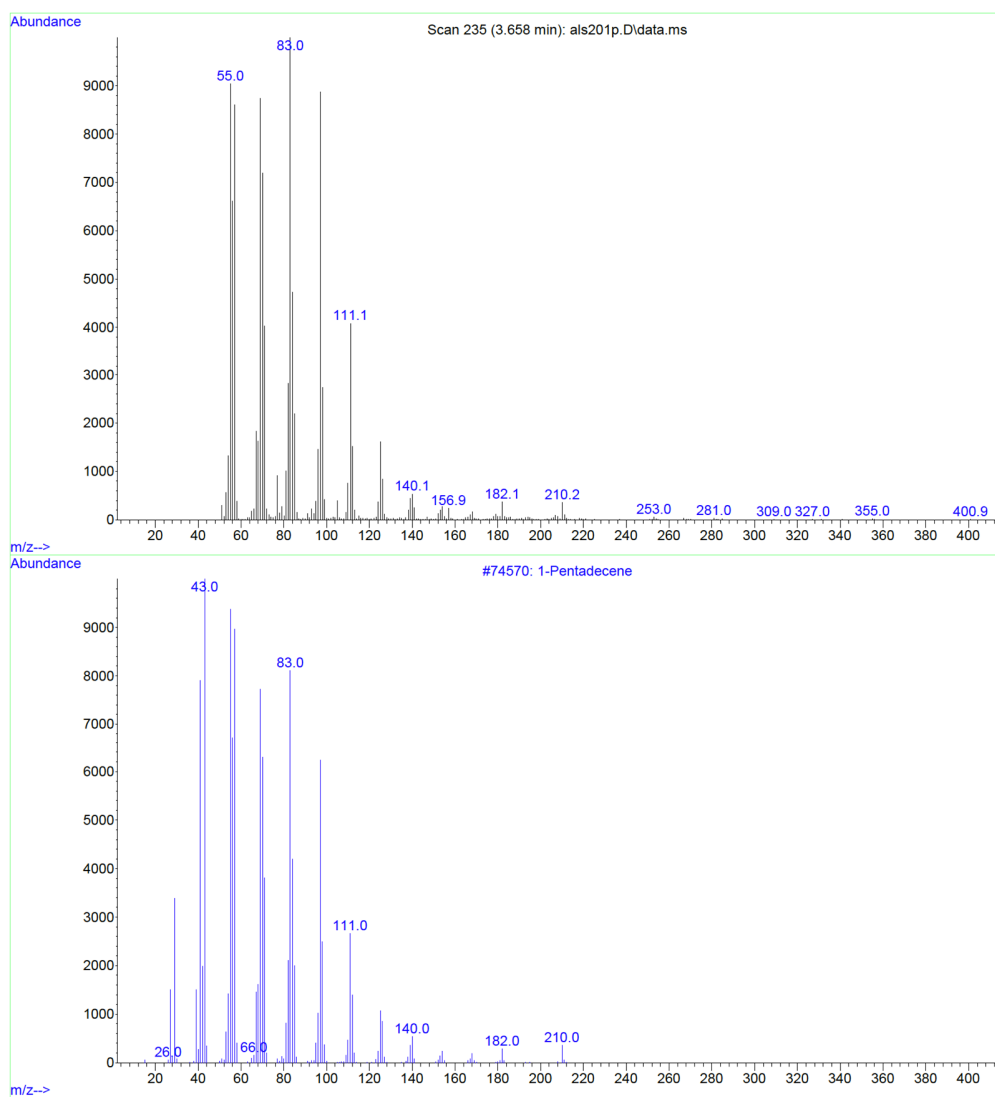
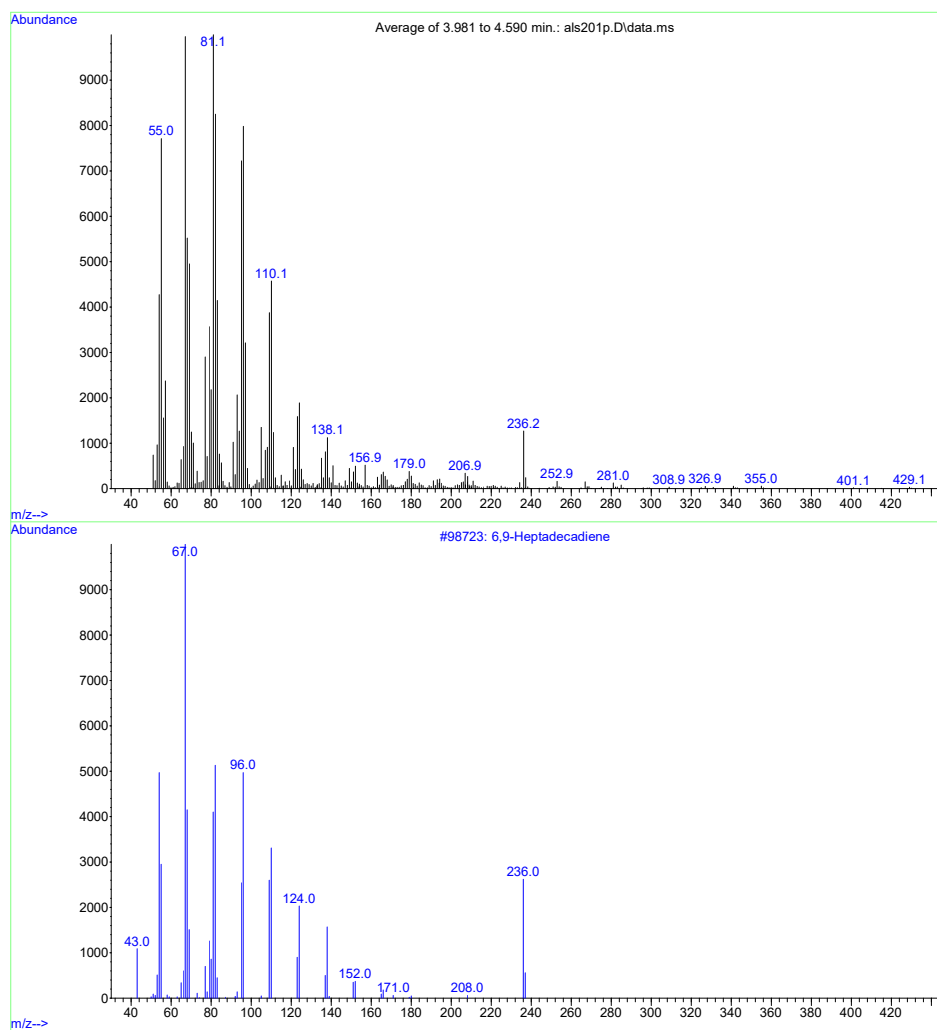


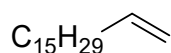
Figure S5: GCMS spectrum of mixture of C17-alkenes at retention time 4.5 min

File :D:\MassHunter\GCMS\1\data\alston\als201p.D
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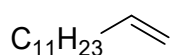
6. Characterization data for alkene products

(Z)-Heptadeca-1,8-diene (Table 2, **2a**)⁴



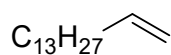
Eluents (Hexane, R_f 0.7) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 5.76-5.86 (m, 1H), 5.34-5.37 (m, 2H), 4.92-5.02 (m, 2H), 2.03-2.06 (m, 6H), 1.28-1.39 (m, 18H), 0.89 (t, J = 7.9Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 139.0, 129.9, 129.7, 114.1, 33.7, 32.4, 31.9, 29.7, 29.6, 29.5, 29.3, 28.8, 28.7, 27.2, 27.1, 22.6, 14.0; MS (EI): m/z (relative intensity) 236.4 (M^+ , 20), 138.2 (11), 124.2 (22), 109.2 (36), 96.2 (96), 81.2 (100), 67.2 (97), 55.2 (98).

1-Tridecene (Table 2, **2b**)⁵

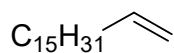


Eluents (Hexane, R_f 0.7) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 5.77-5.87 (m, 1H), 5.00 (dd, J = 1.8Hz, 17.6Hz, 1H), 4.93 (d, J = 10.7Hz, 1H), 2.05 (q, J = 7.9Hz, 2H), 1.27 (s, 18H), 0.89 (t, J = 7.9Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 139.2, 114.0, 33.8, 31.9, 29.7, 29.68, 29.66, 29.5, 29.3, 29.2, 28.9, 22.7, 14.1; MS (EI): m/z (relative intensity) 182.4 (M^+ , 9), 125.3 (10), 111.3 (35), 97.2 (80), 83.2 (96), 69.2 (95), 55.2 (100).

1-Pentadecene (Table 2, **2c**)⁵

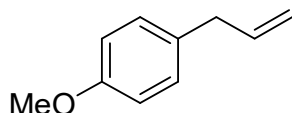


Eluents (Hexane, R_f 0.7) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 5.78-5.85 (m, 1H), 5.01 (dd, J = 1.9Hz, 17.7Hz, 1H), 4.92 (d, J = 10.0Hz, 1H), 2.05 (q, J = 7.4Hz, 2H), 1.27 (s, 22H), 0.89 (t, J = 7.0Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 139.2, 114.0, 33.8, 31.9, 29.7, 29.6, 29.5, 29.4, 29.2, 29.0, 22.7, 14.1; MS (EI): m/z (relative intensity) 210.4 (M^+ , 7), 182.4 (5), 140.3 (7), 125.3 (17), 111.3 (46), 97.2 (95), 83.2 (100), 69.2 (87), 55.2 (93).

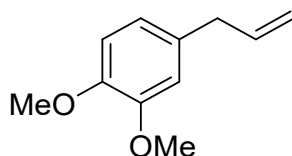
1-Heptadecene (Table 2, **2d**)⁵

Eluents (Hexane, $R_f = 0.7$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 5.77-5.87 (m, 1H), 5.01 (dd, $J = 1.9\text{Hz}$, 17.1Hz , 1H), 4.93 (d, $J = 10.0\text{Hz}$, 1H), 2.05 (q, $J = 7.0\text{Hz}$, 2H), 1.27 (s, 26H), 0.89 (t, $J = 7.2\text{Hz}$, 3H);

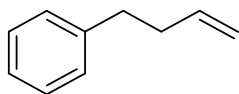
^{13}C NMR (100 MHz, CDCl_3) δ 139.2, 114.0, 33.8, 31.9, 29.7, 29.6, 29.5, 29.4, 29.2, 29.0, 22.7, 14.1; MS (EI): m/z (relative intensity) 238.5 (M^+ , 7), 210.4 (5), 125.3 (28), 111.3 (57), 97.2 (100), 83.2 (98), 69.2 (78), 55.2 (85).

1-Allyl-4-methoxybenzene (Table 2, **2e**)⁶

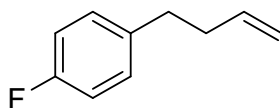
Eluents (Hexane: ethyl acetate 10:1, $R_f = 0.6$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 7.11 (d, $J = 8.5\text{Hz}$, 2H), 6.85 (d, $J = 8.6\text{Hz}$, 2H), 5.91-6.01 (m, 1H), 5.04-5.0 (m, 2H), 3.79 (s, 3H), 3.34 (d, $J = 6.6\text{Hz}$, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 157.9, 137.8, 132.0, 129.4, 115.3, 113.8, 55.2, 39.3; MS (EI): m/z (relative intensity) 148.2 (M^+ , 100), 133.1 (23), 121.1 (38), 105.1 (21), 91.1 (21), 77.1 (23).

1-Allyl-3,4-dimethoxybenzene (Table 2, **2f**)⁷

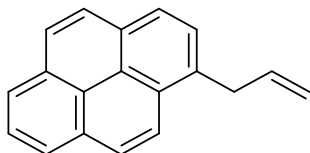
Eluents (Hexane: ethyl acetate 10:1, $R_f = 0.3$) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 6.78-6.81 (m, 1H), 6.72 (d, $J = 7.6\text{Hz}$, 2H), 5.91-5.99 (m, 1H), 5.05-5.10 (m, 2H), 3.86 (s, 3H), 3.85 (s, 3H), 3.33 (d, $J = 6.6\text{Hz}$, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 148.8, 147.3, 137.6, 134.4, 120.3, 115.5, 111.8, 111.2, 55.8, 55.7, 39.7; MS (EI): m/z (relative intensity) 178.2 (M^+ , 100), 163.1 (30), 147.1 (30), 103.1 (25), 91.1 (25), 77.1 (20).

4-Phenyl-1-butene (Table 2, **2g**)⁸

Eluents (Hexane, R_f = 0.6) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 7.28-7.32 (m, 2H), 7.20-7.22 (m, 3H), 5.83-5.93 (m, 1H), 5.06 (dd, J = 1.8Hz, 17.3Hz, 1H), 4.99 (d, J = 10.2Hz, 1H), 2.73 (t, J = 7.6Hz, 2H), 2.40 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 141.8, 138.0, 128.4, 128.2, 125.7, 114.8, 35.4, 35.3; MS (EI): m/z (relative intensity) 132.1 (M^+ , 20), 91.1 (100).

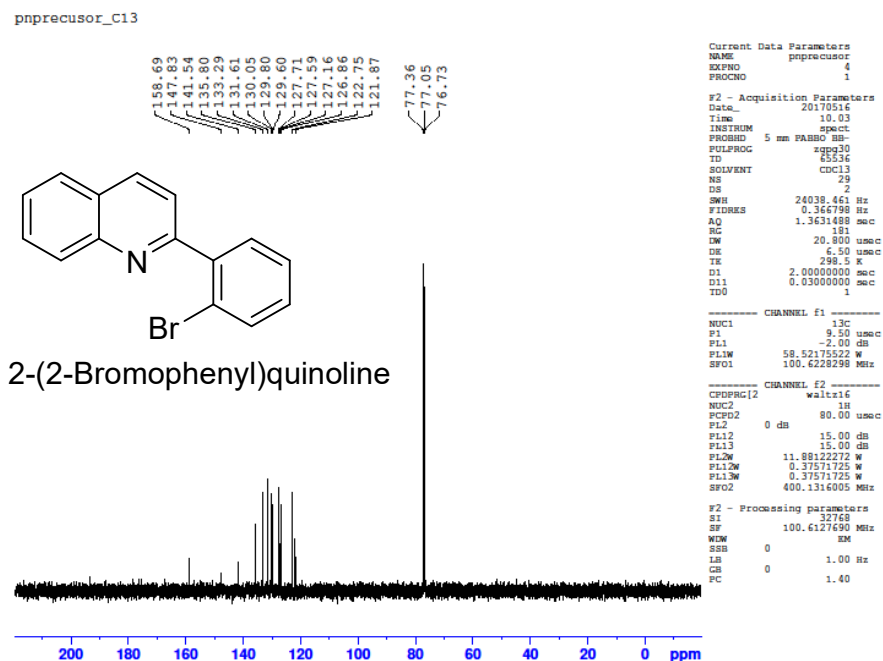
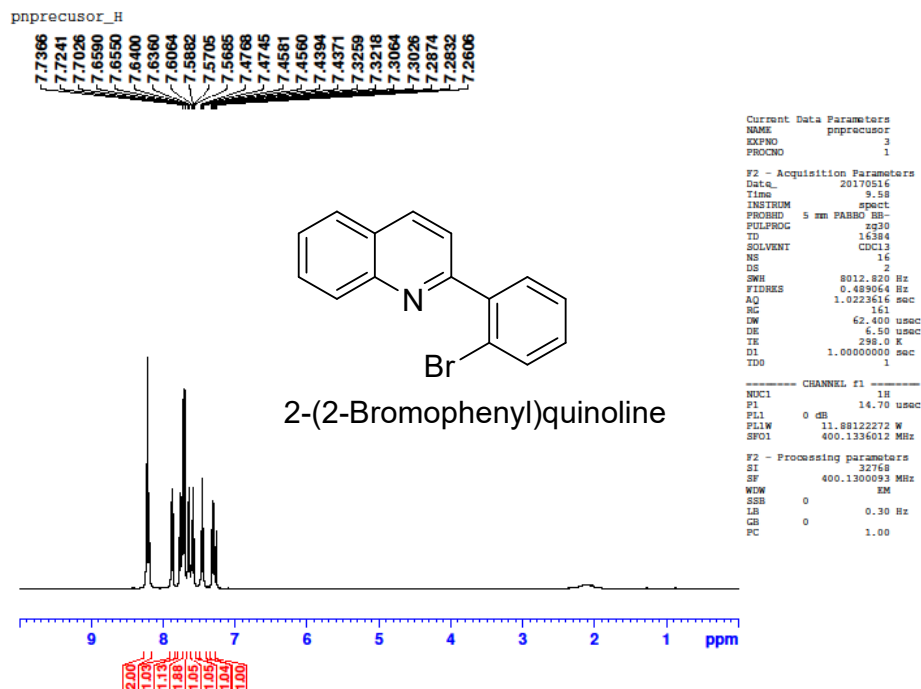
4-(4-Fluorophenyl)-1-butene (Table 2, **2h**)⁹

Eluents (Hexane, R_f = 0.5) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 7.11-7.14 (m, 2H), 6.96 (t, J = 8.6Hz, 2H), 5.78-5.88 (m, 1H), 4.97-5.05 (m, 2H), 2.68 (t, J = 7.7Hz, 2H), 2.34 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 161.2 (d, J = 243.0Hz), 137.7, 137.3 (d, J = 3.0Hz), 129.6 (d, J = 7.9Hz), 115.0 (d, J = 4.3Hz), 114.8, 35.5, 34.5; MS (EI): m/z (relative intensity) 150.2 (M^+ , 14), 109.2 (100), 83.2 (7).

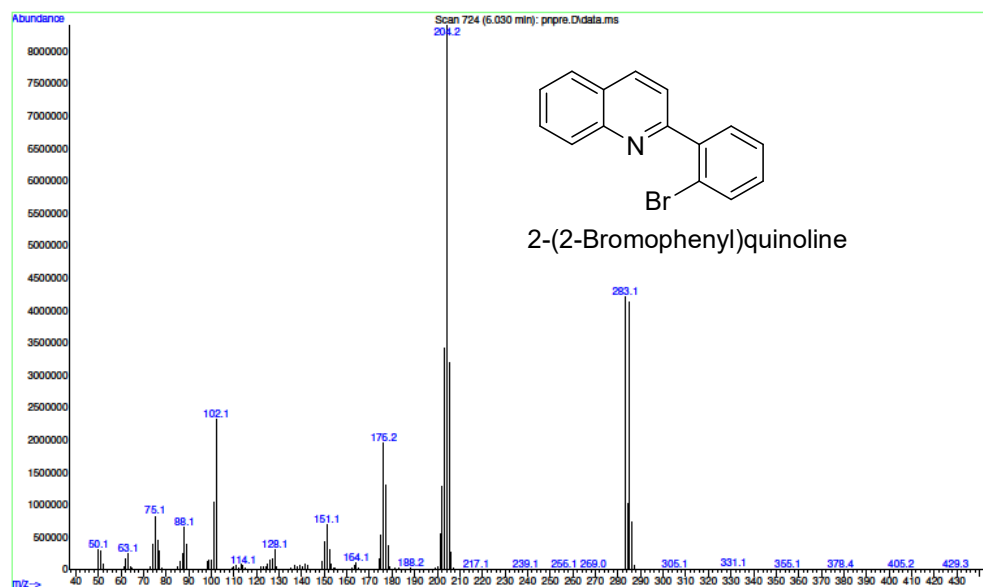
1-Allyl-pyrene (Table 2, **2i**)¹⁰

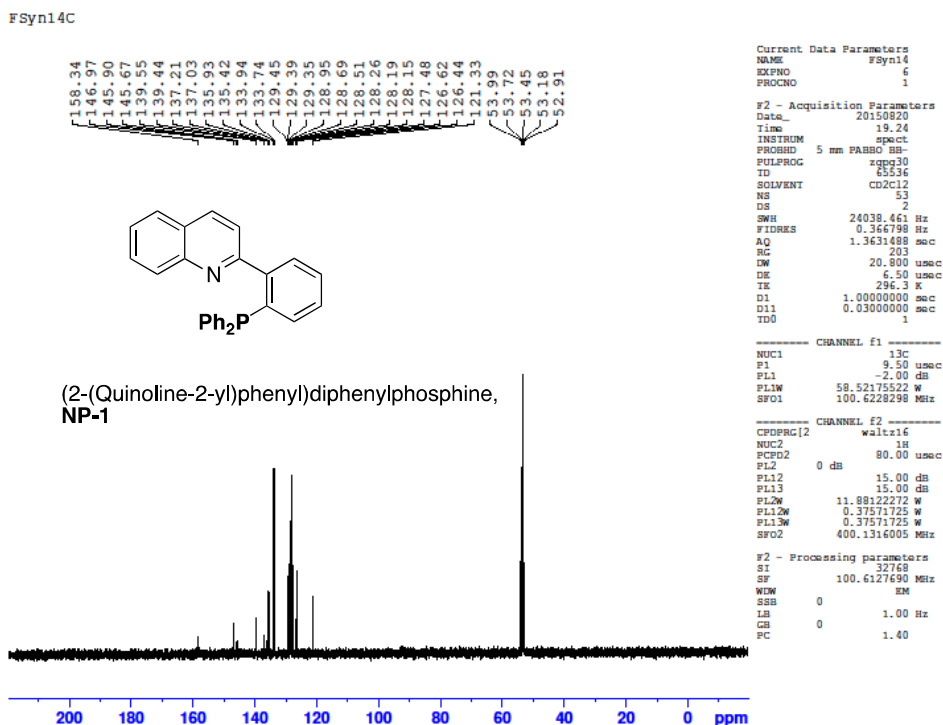
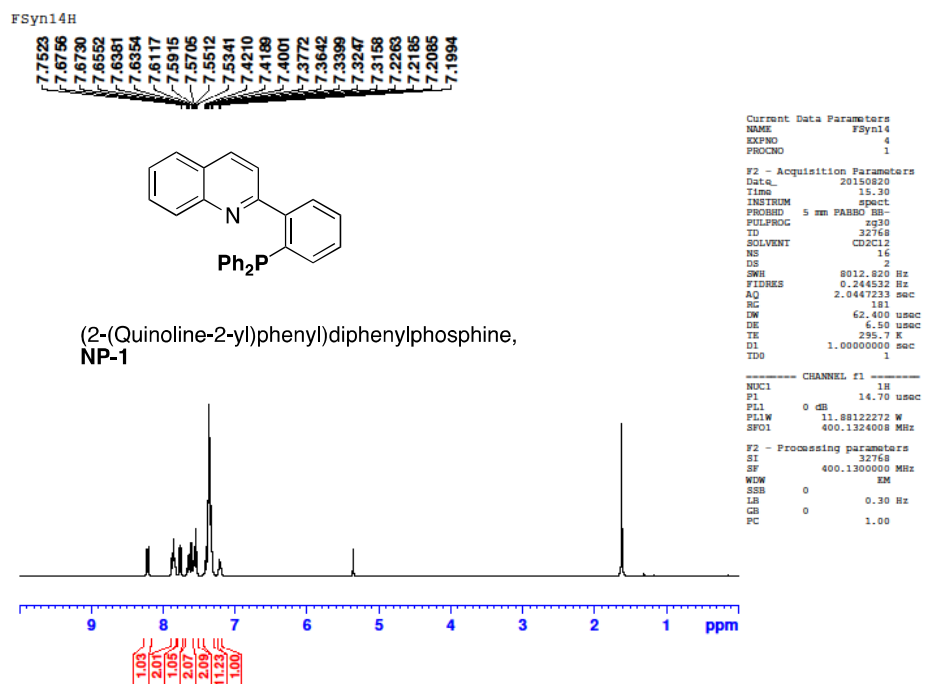
Eluents (Hexane, R_f = 0.5) was used for flash column chromatography. ^1H NMR (400 MHz, CDCl_3) δ 8.03-8.30 (m, 8H), 7.92 (d, J = 7.5Hz, 1H), 6.26-6.36 (m, 1H), 5.17-5.27 (m, 2H), 4.15 (d, J = 6.6Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 137.2, 133.7, 131.2, 130.7, 129.9, 128.8,

127.38, 127.35, 127.2, 127.1, 126.6, 125.6, 124.9, 124.8, 124.7, 123.4, 116.0, 37.5; MS (EI): m/z (relative intensity) 242.2 (M^+ , 100), 227.2(29), 215.2 (33), 119.9 (20).

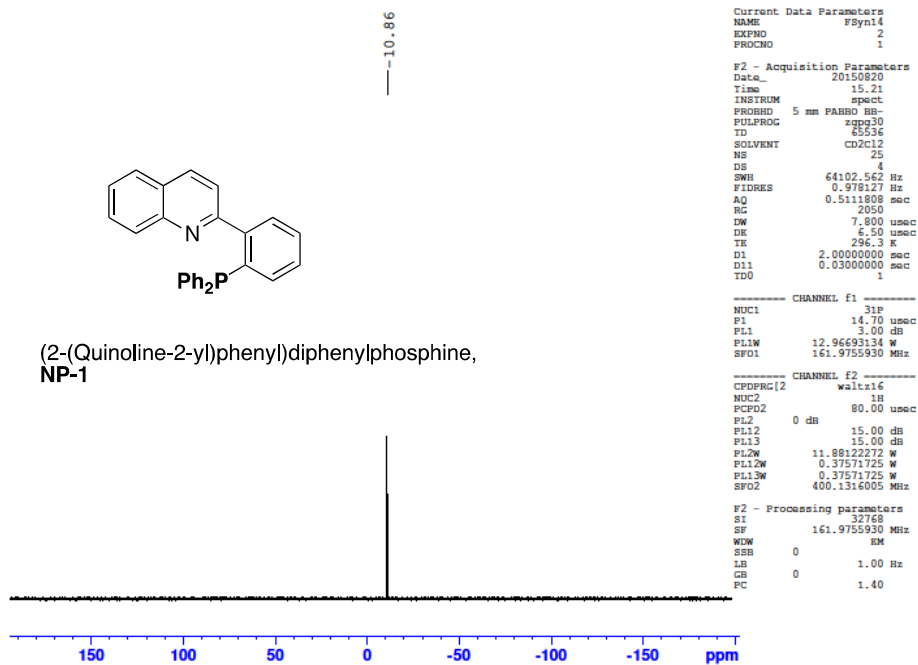
7. ^1H , ^{13}C , ^{31}P NMR, MS and HRMS spectra

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Sample Name: pnpre
Misc Info :
Vial Number: 5

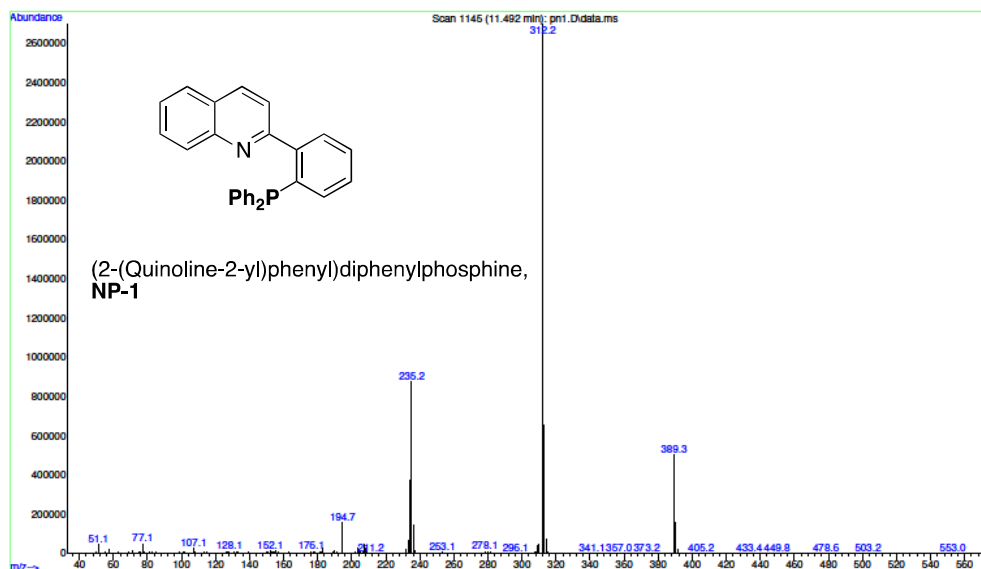


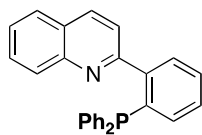
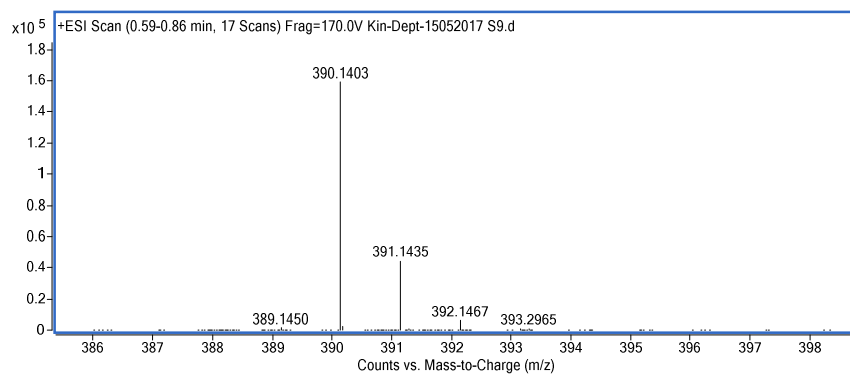


FSyn14P



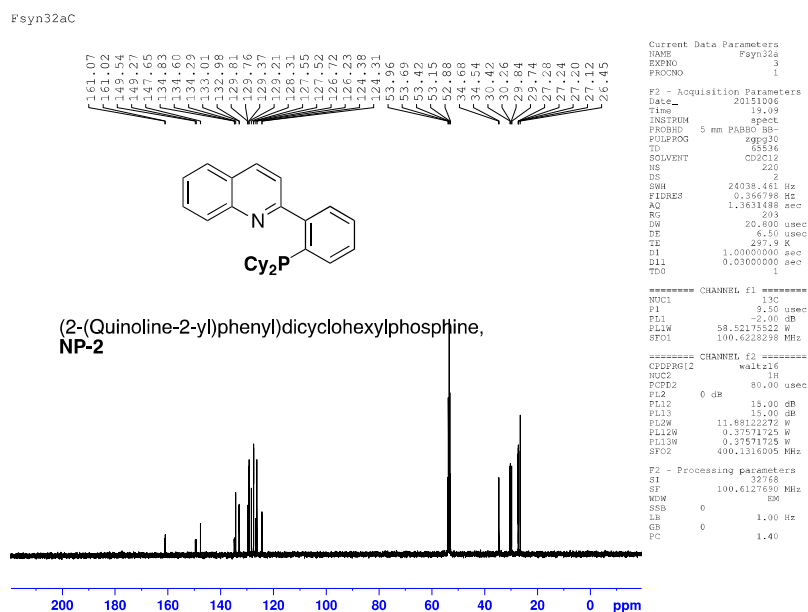
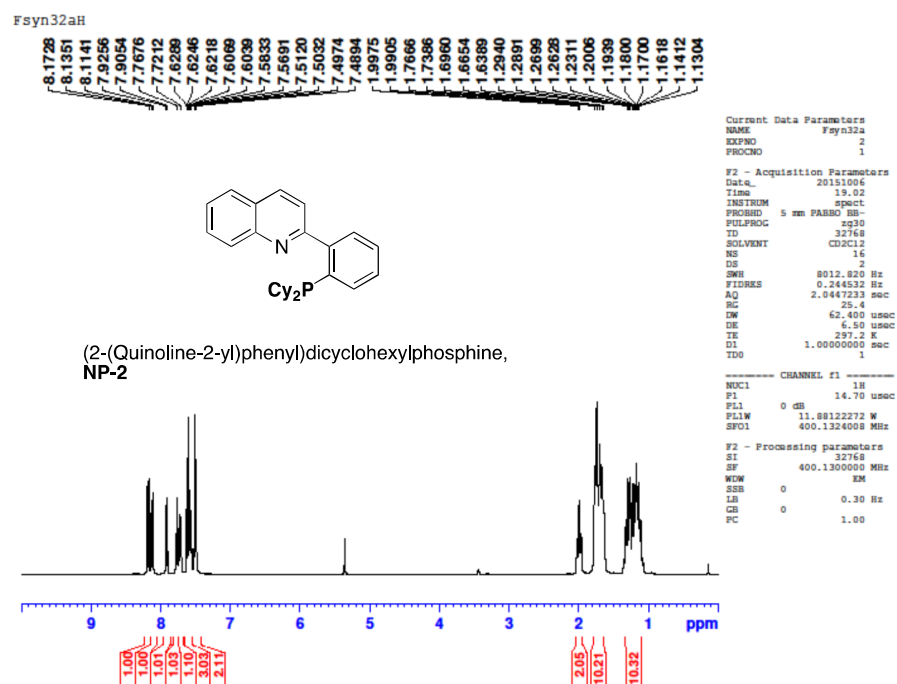
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 Instrument : 5977
 Sample Name: pn1
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 Vial Number: 7



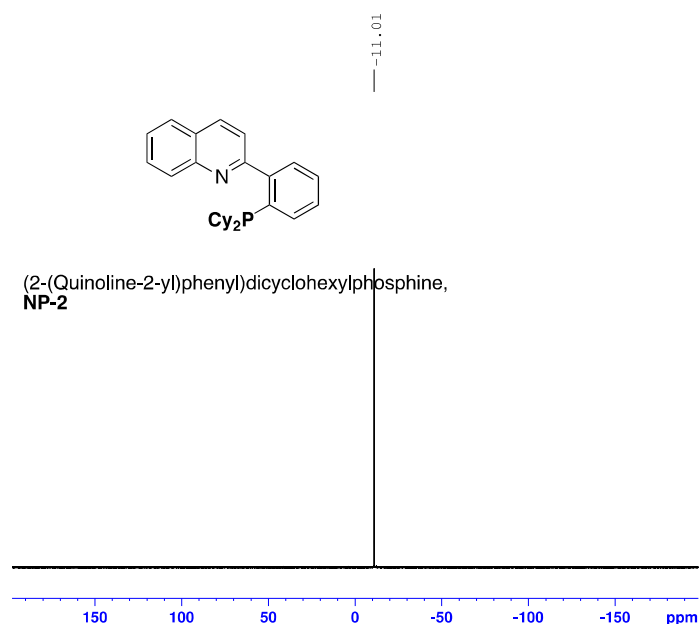


(2-(Quinoline-2-yl)phenyl)diphenylphosphine,
NP-1

Mass	Calc. Mass	mDa	PPM	Formula
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Fsyn32aP



Current Data Parameters

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PROCNO	1

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FIDRES	0.978127 Hz
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RG	2050
DW	7.800 usec
DE	6.50 usec
TE	297.0 K
D1	2.00000000 sec
D11	0.03000000 sec
TDO	1

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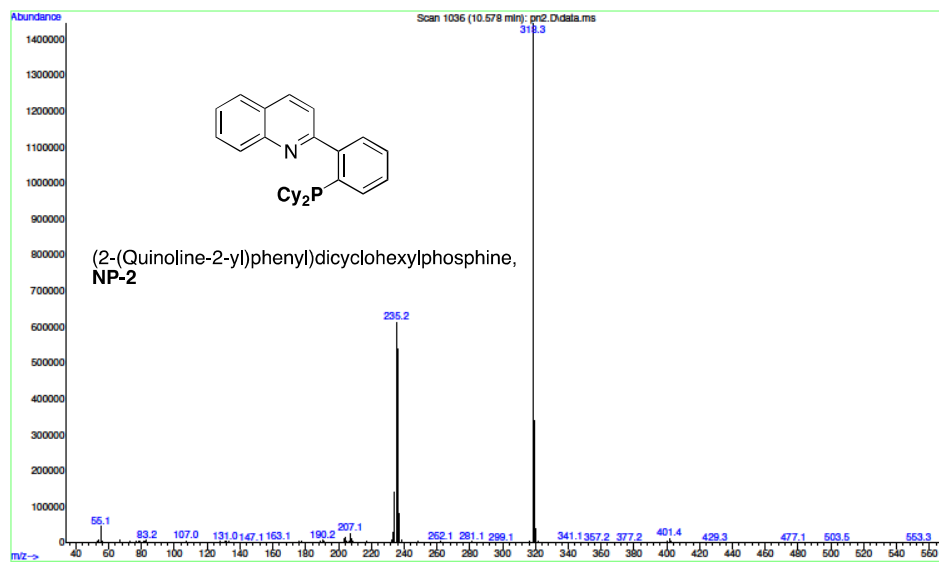
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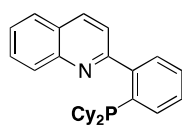
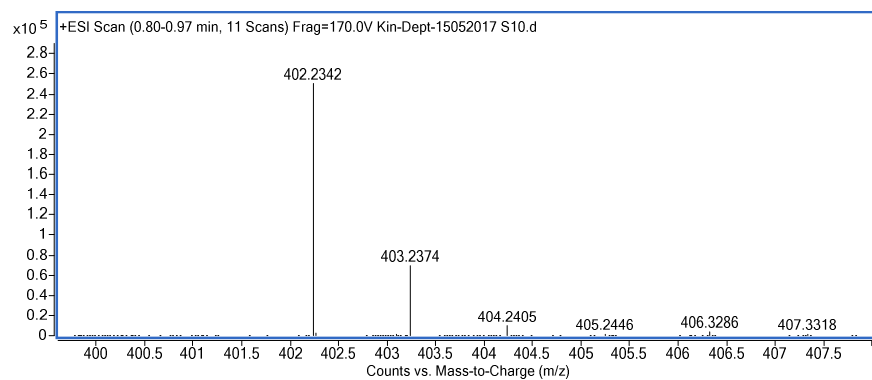
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NUC2	1H
P2P2	80.00 usec
PL2	0 dB
PL12	15.00 dB
PL13	15.00 dB
PL2W	11.88122272 W
PL12W	0.37571725 W
PL13W	0.37571725 W
SFO2	400.1316005 MHz

F2 - Processing parameters

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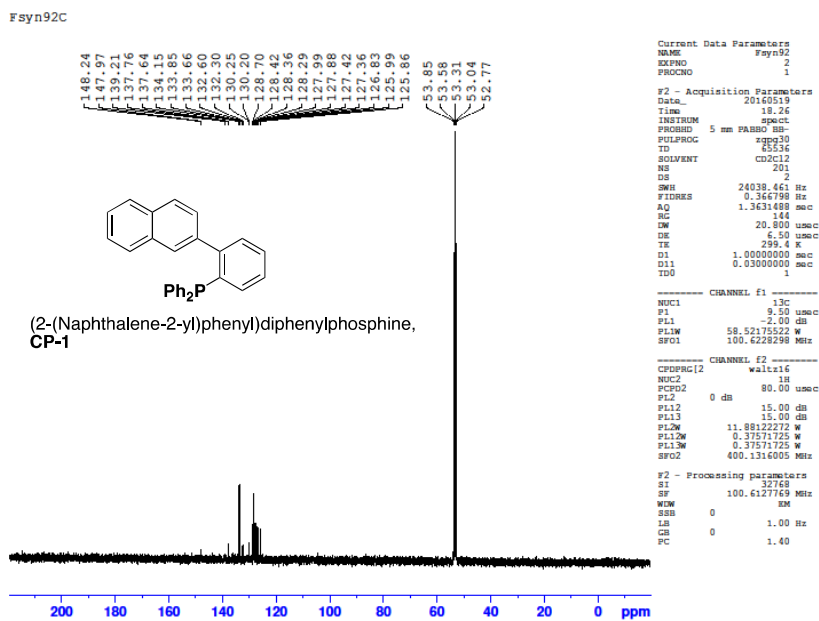
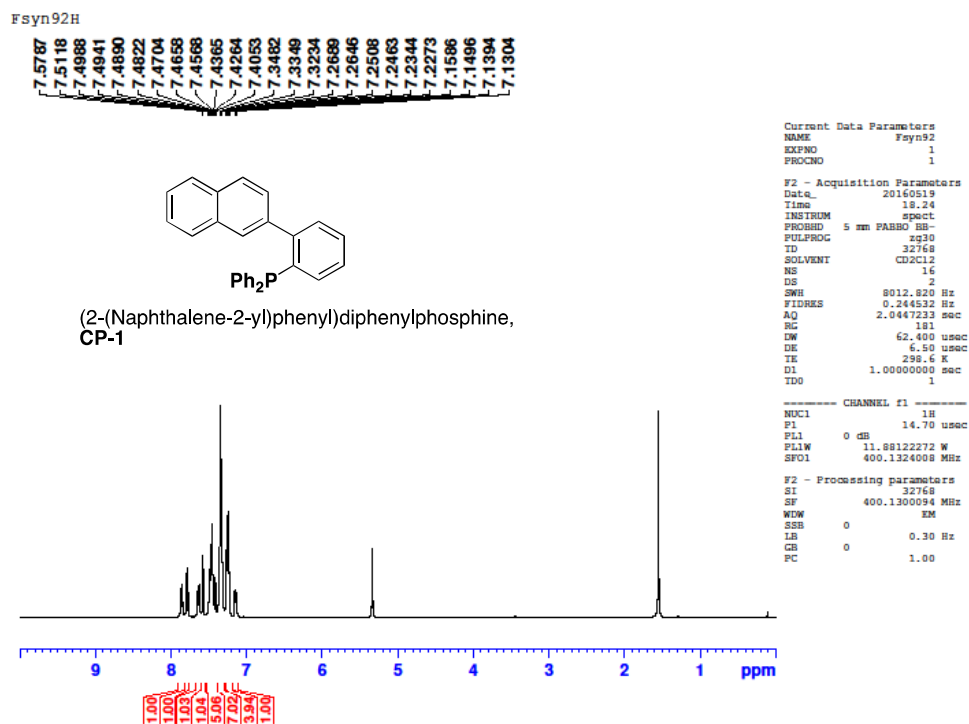
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 Instrument : 5977
 Sample Name: pn2
 Misc Info :
 Vial Number: 8



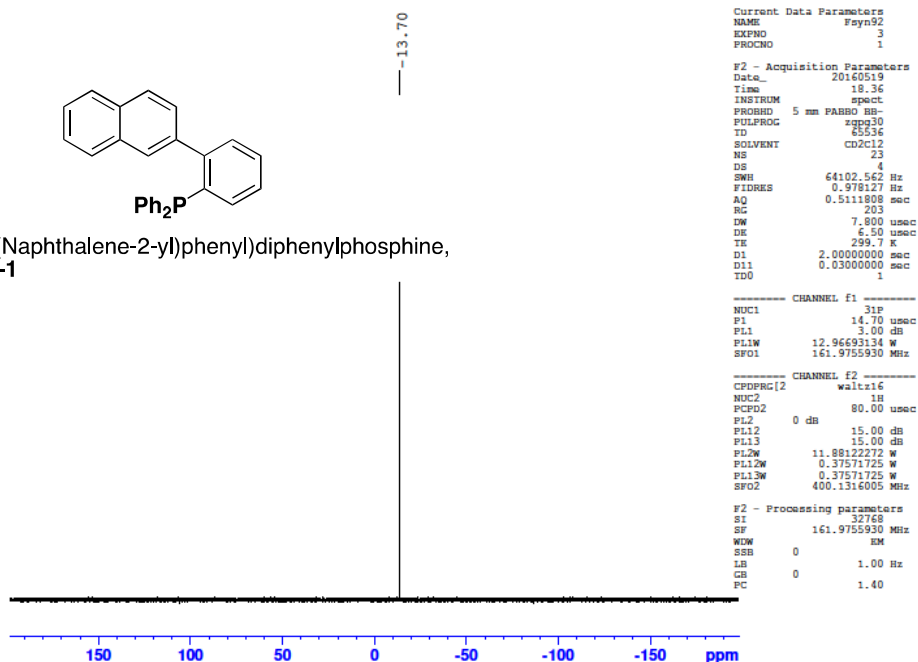
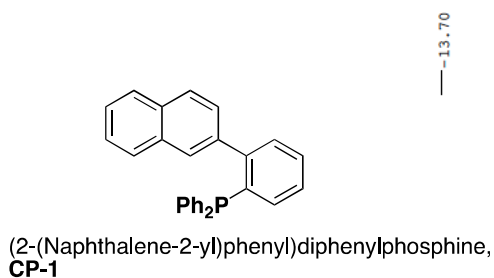


(2-(Quinoline-2-yl)phenyl)dicyclohexylphosphine,
NP-2

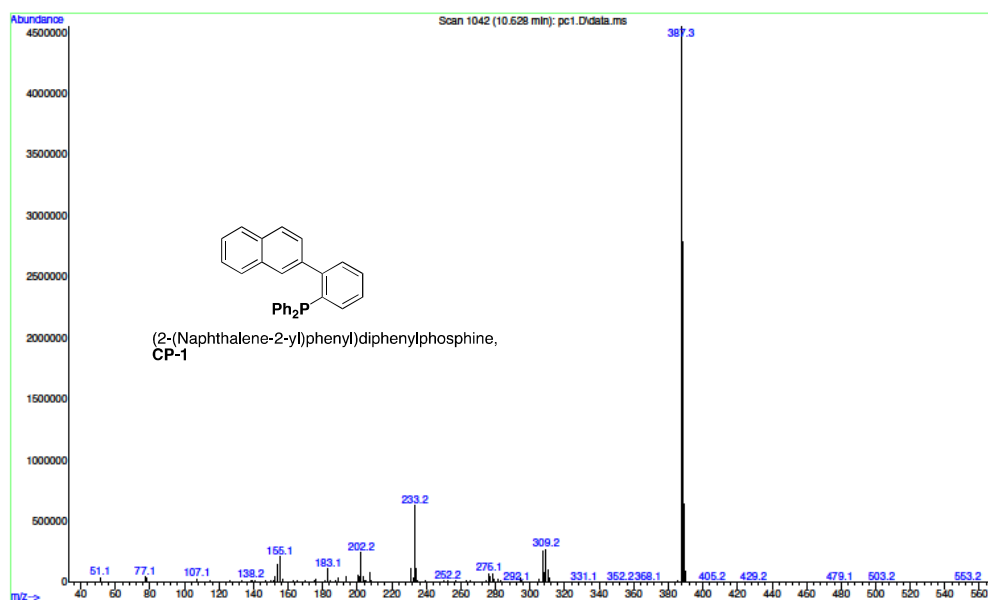
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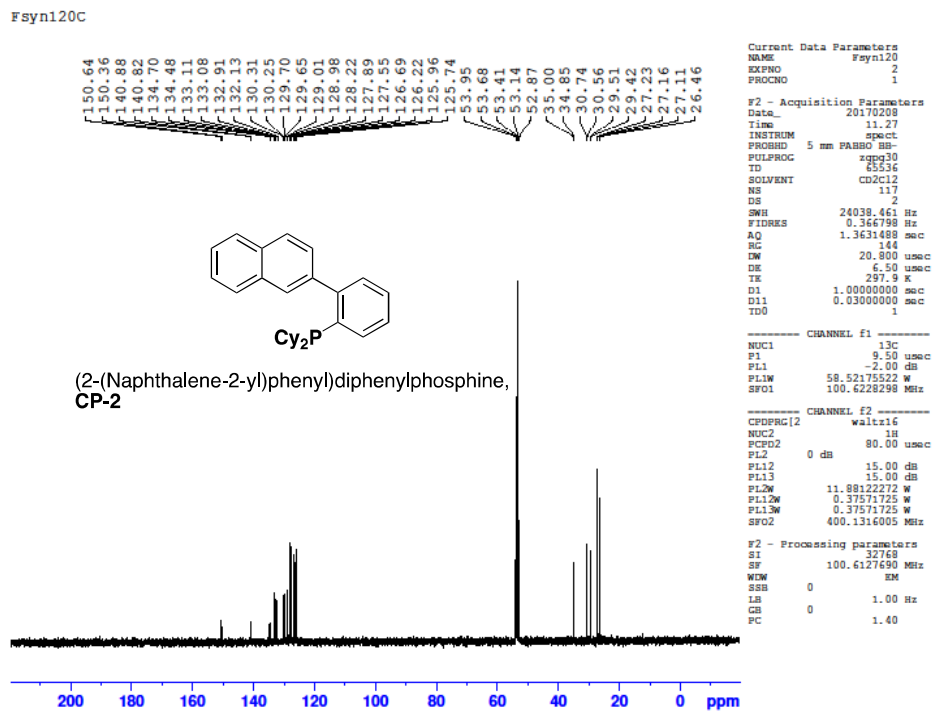
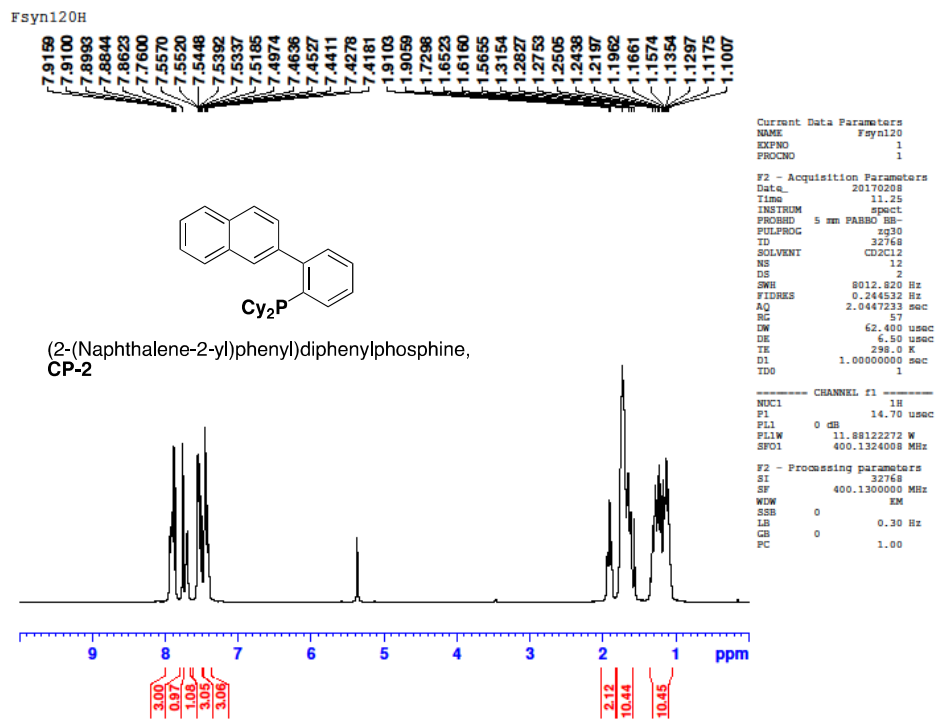


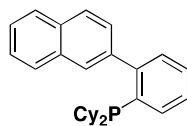
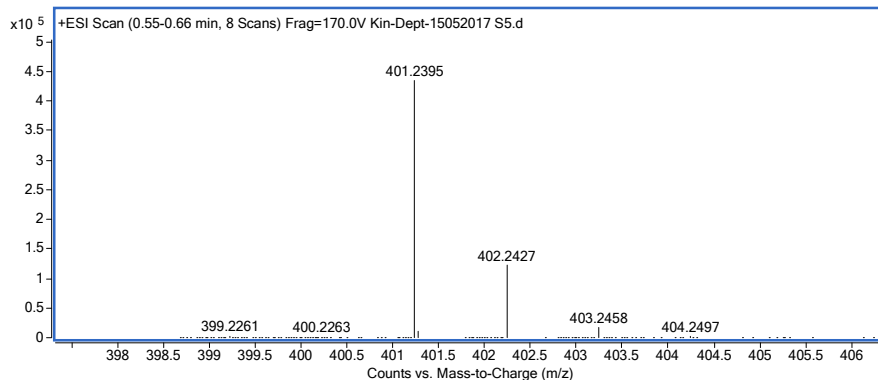
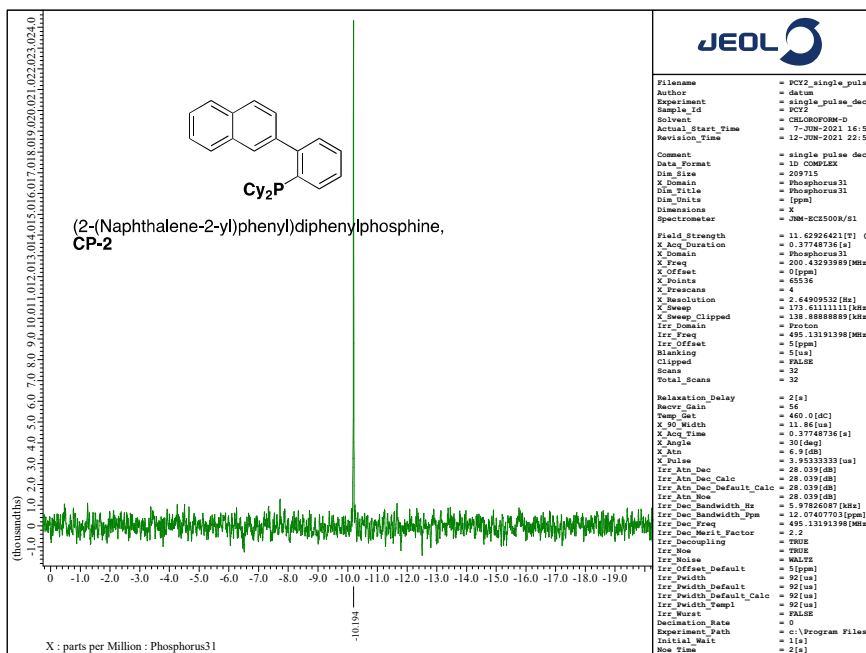
Fsyn92P



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Instrument : 5977
Sample Name: pc1
Misc Info :
Vial Number: 9



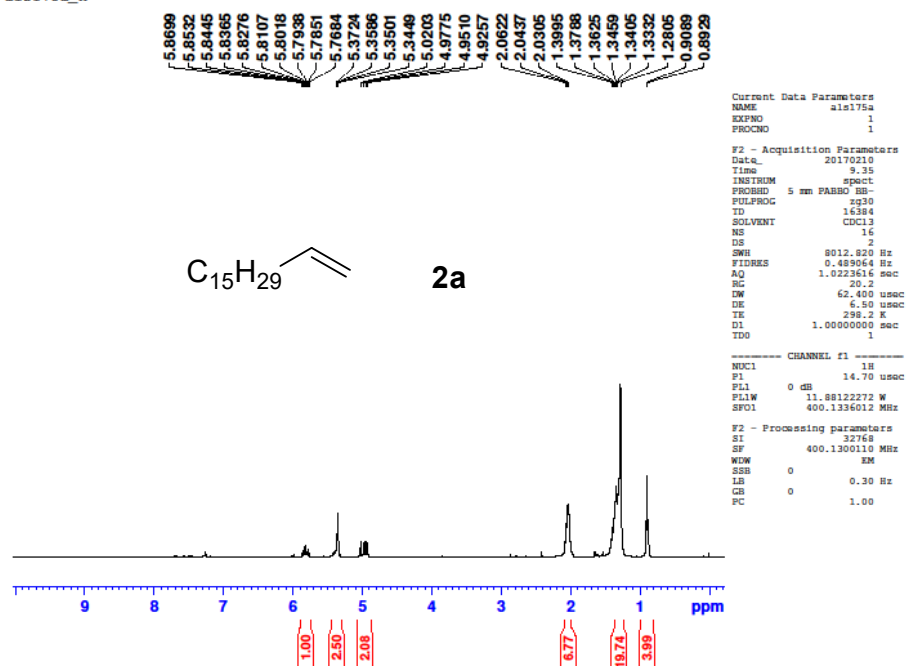




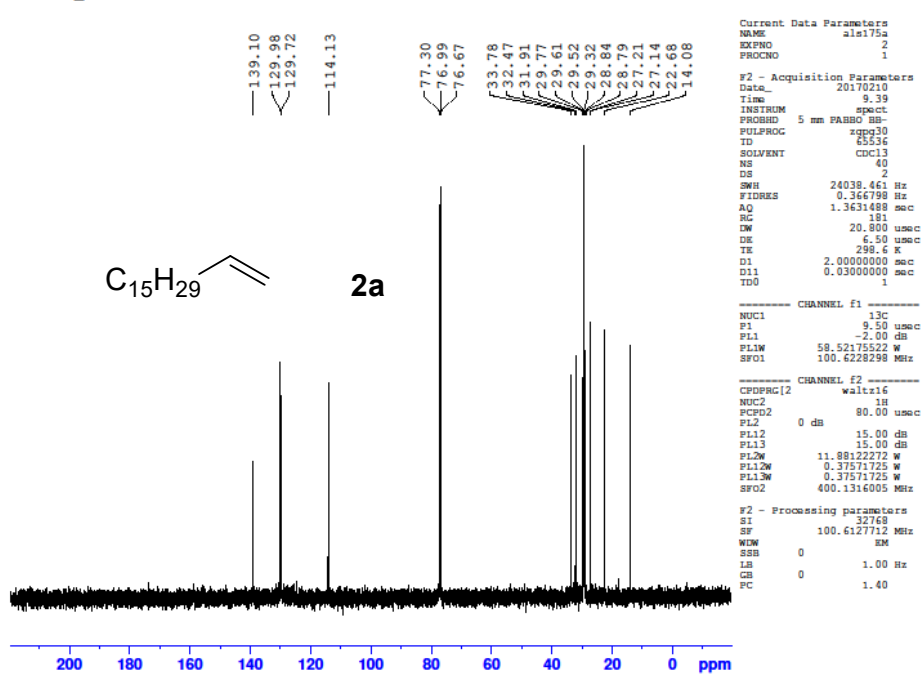
(2-(Naphthalene-2-yl)phenyl)diphenylphosphine, CP-2

Mass	Calc. Mass	mDa	PPM	Formula
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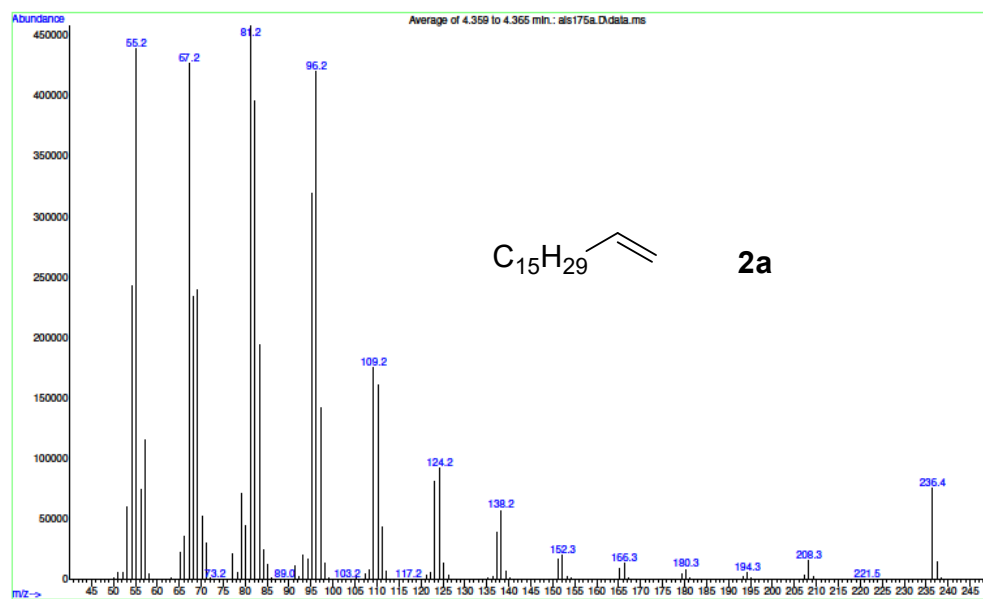
als175a_H

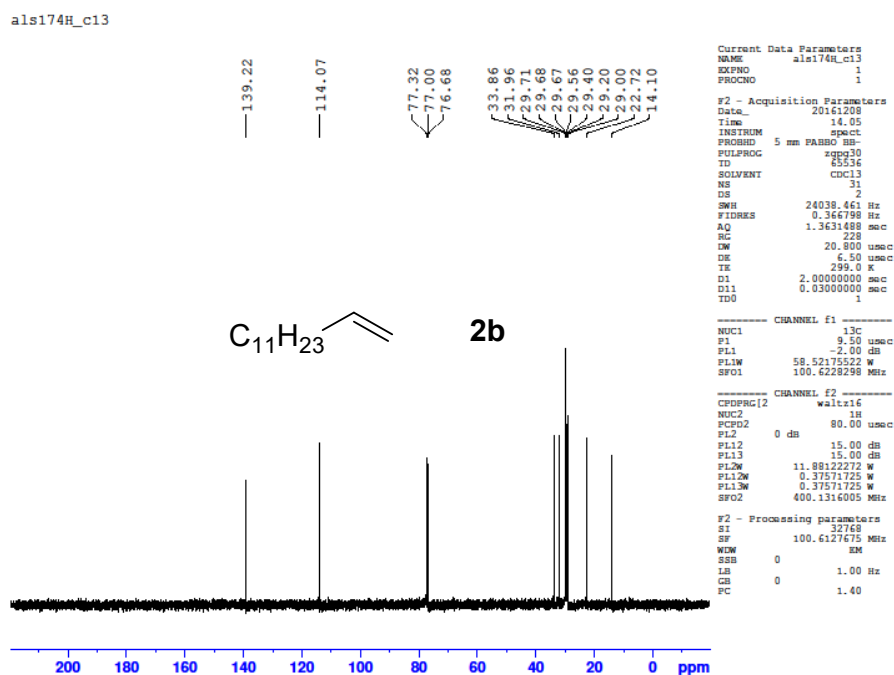
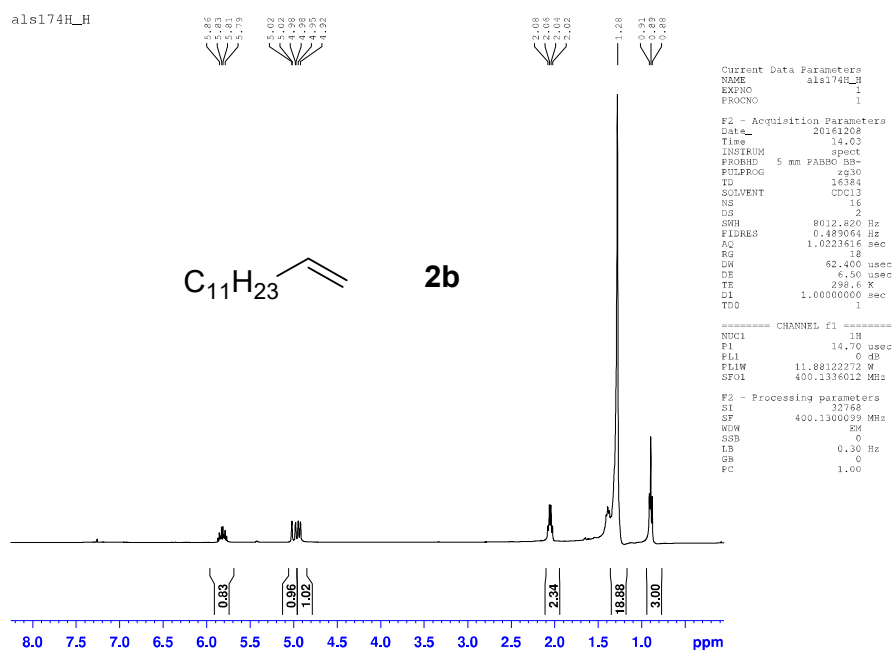


als175a_C13

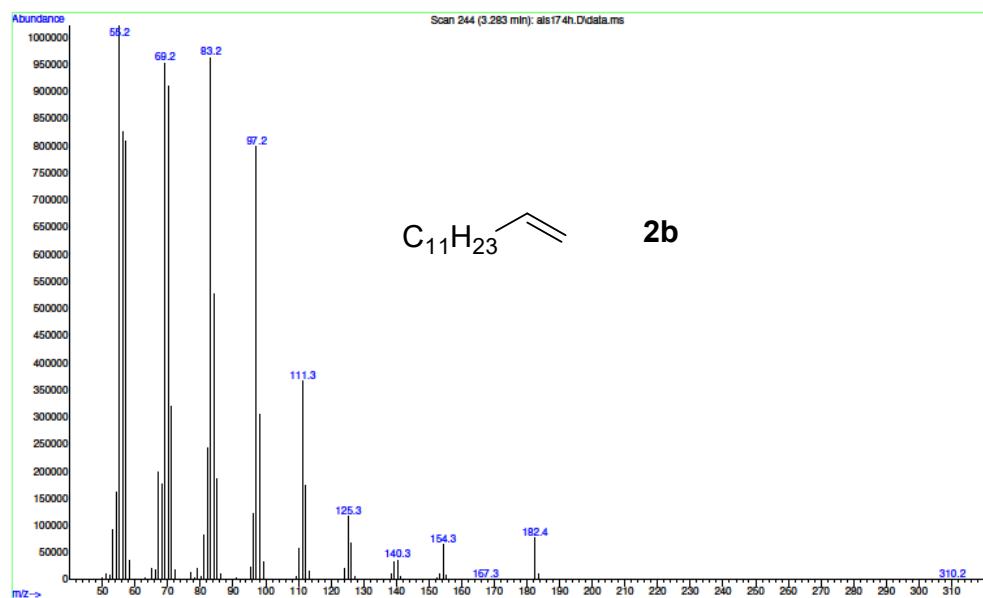


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Instrument : 5977
Sample Name: als175a
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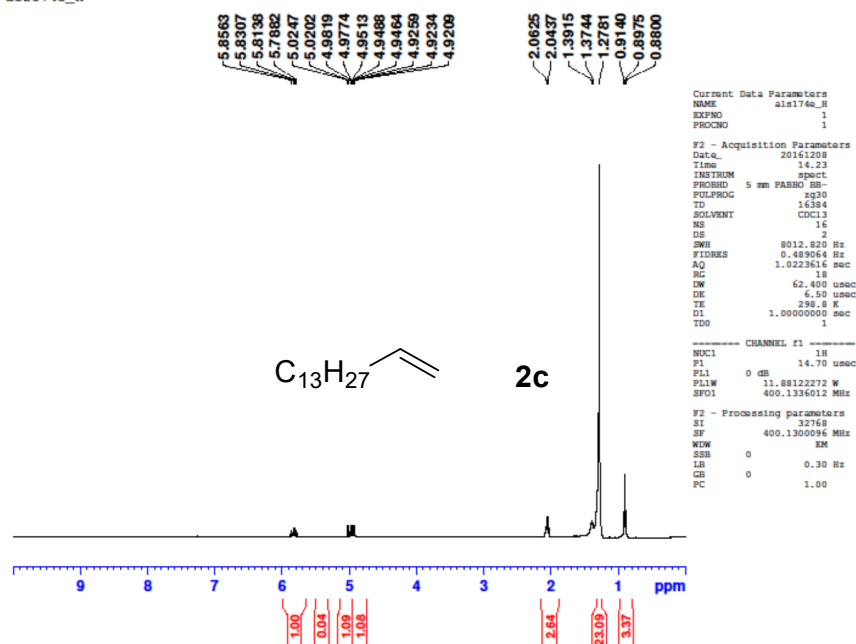




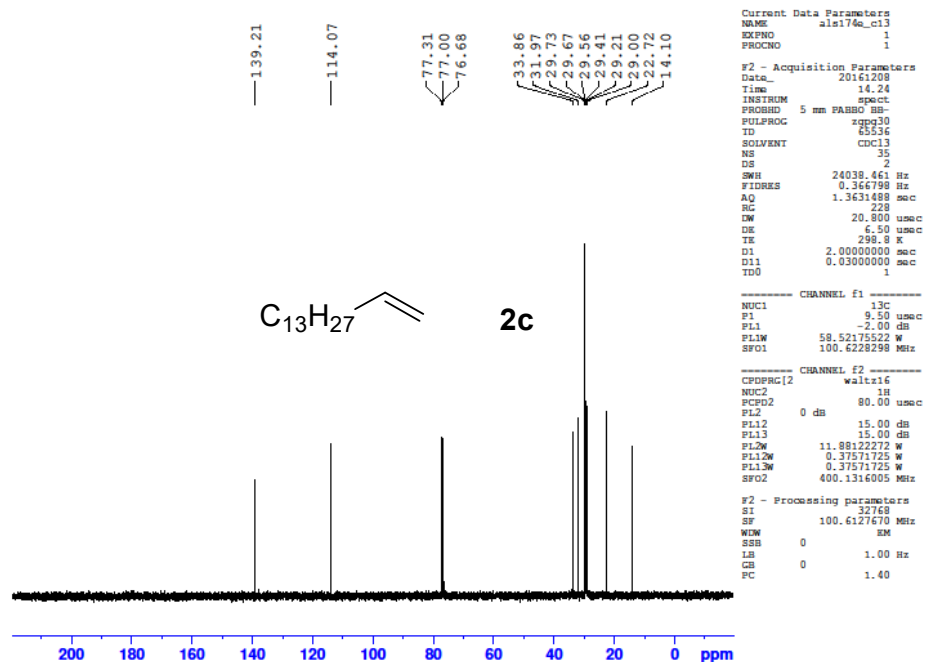
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Instrument : 5977
Sample Name: als174h
Misc Info :
Vial Number: 8



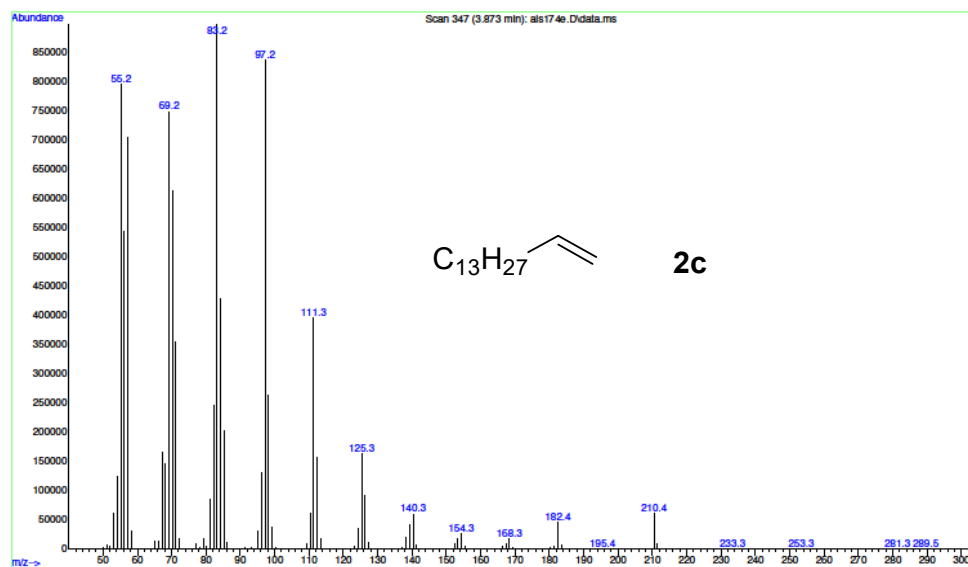
als174e_H



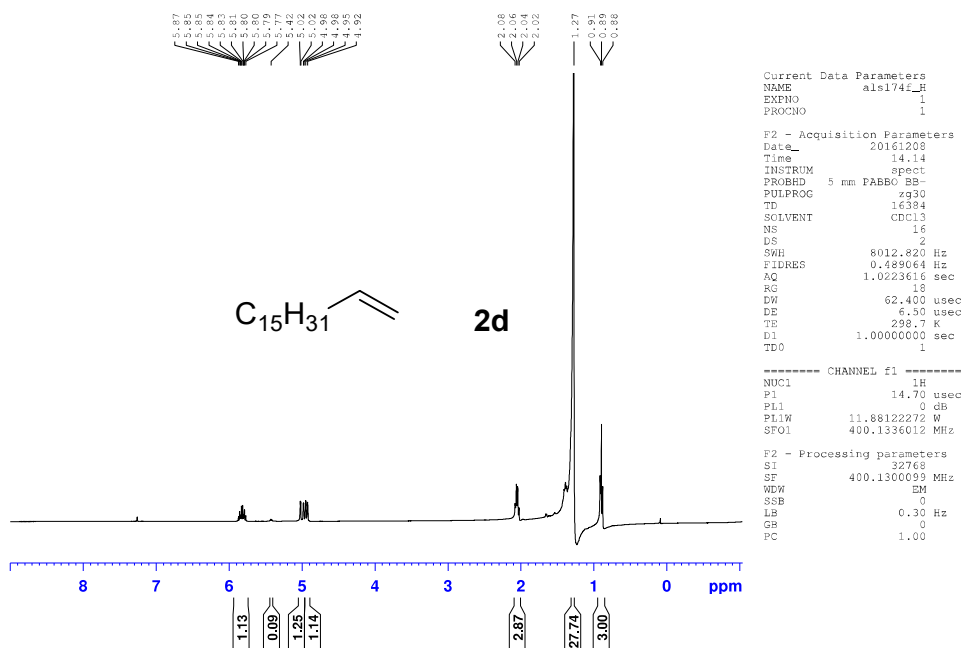
als174e_c13



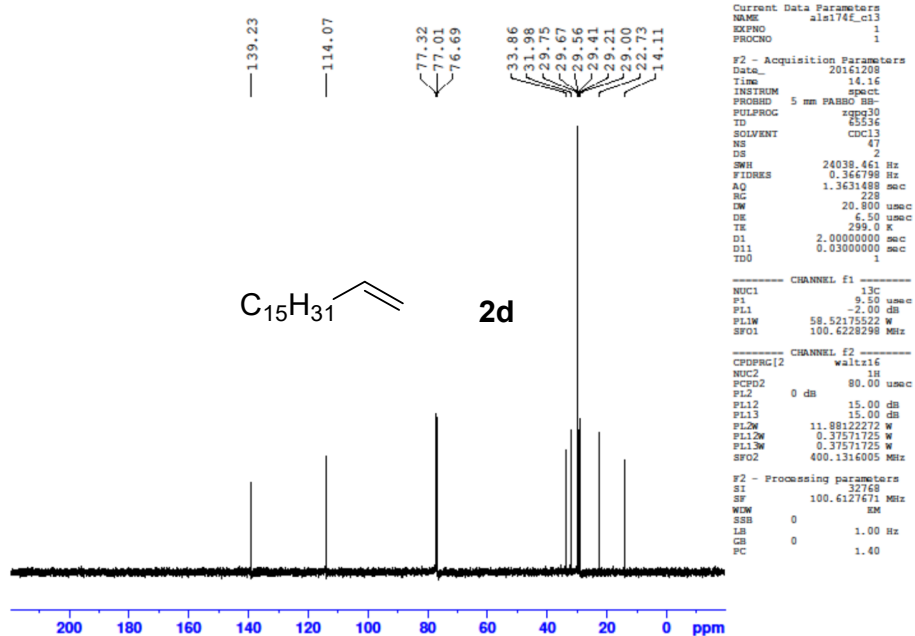
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Instrument : 5977
Sample Name: als174e
Misc Info :
Vial Number: 6



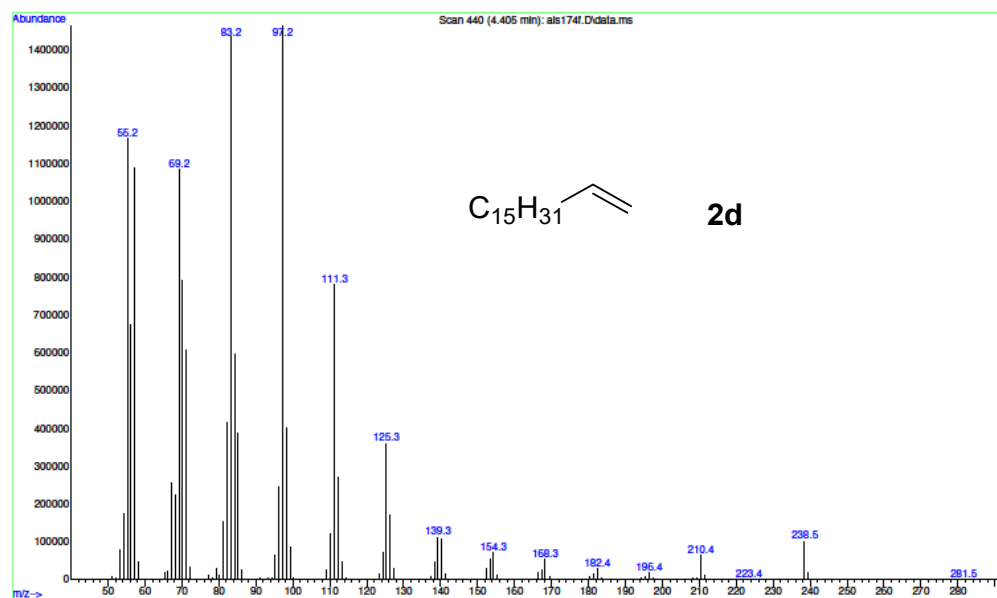
als174f_H



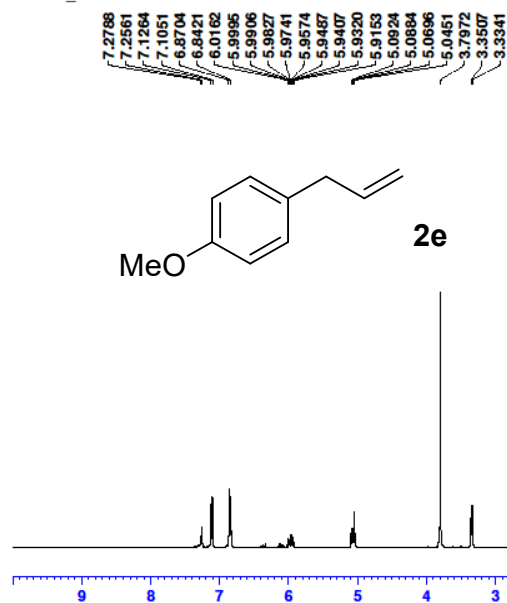
als174f_c13



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Instrument : 5977
Sample Name: als174f
Misc Info :
Vial Number: 7



als184e_H



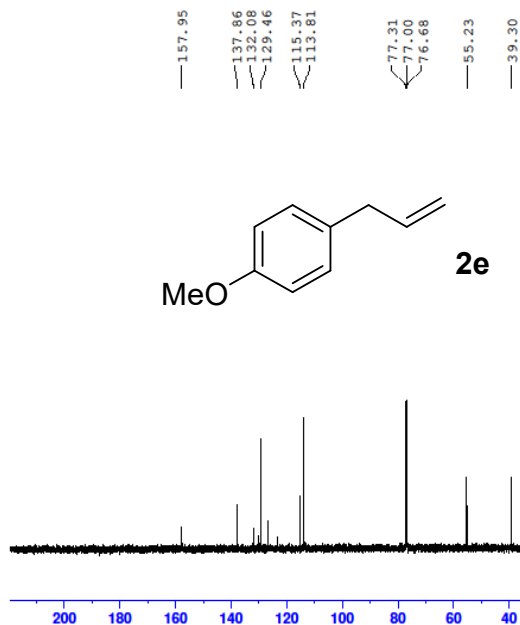
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 PROCNO 1

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 FIDRES 0.489064 Hz
 AQ 1.0223616 sec
 RG 80.6
 DW 62.400 usec
 DE 6.50 usec
 TE 297.9 K
 D1 1.00000000 sec
 TD 1

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 PL1 0 dB
 PL1W 11.88122272 W
 SFO1 400.1336012 MHz

F2 - Processing parameters
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 SF 400.1300113 MHz
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 SSB 0
 LB 0.30 Hz
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 PC 1.00

als184e_13C



Current Data Parameters
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 PROCNO 1

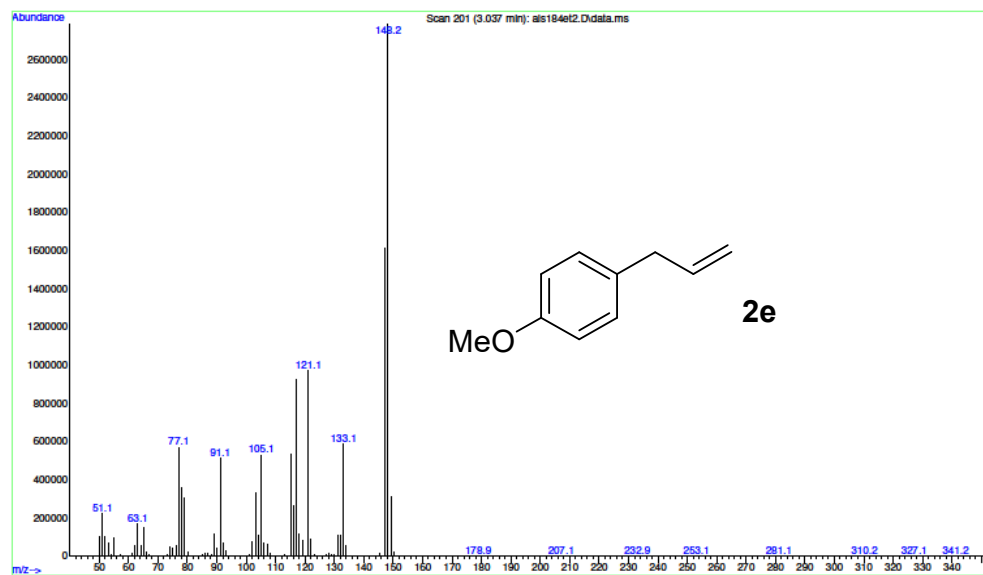
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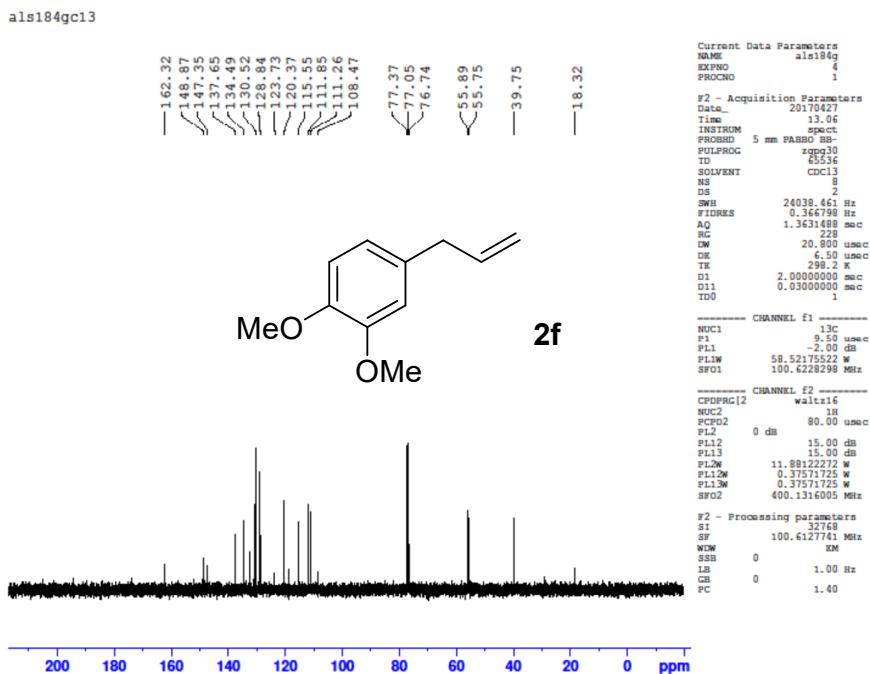
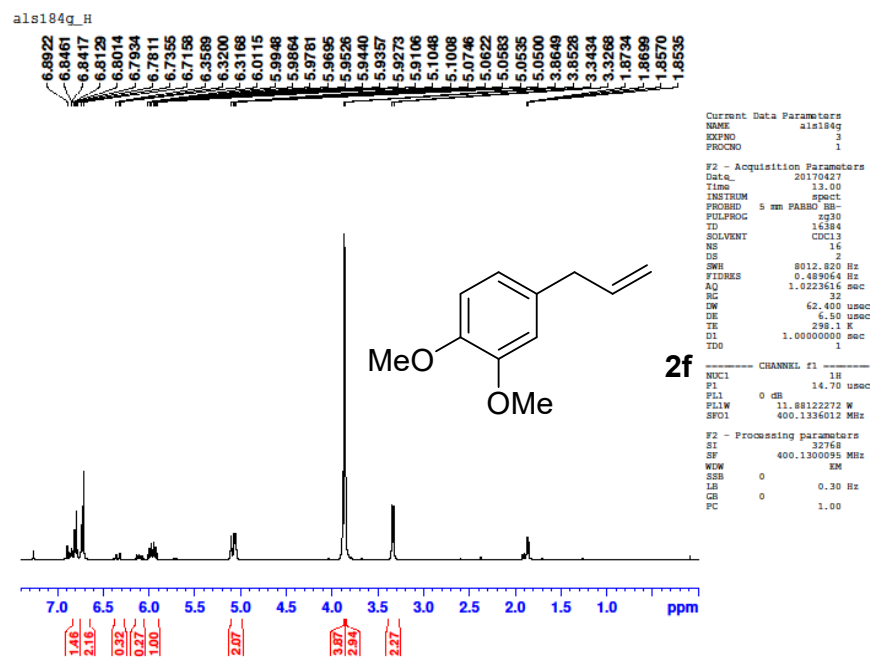
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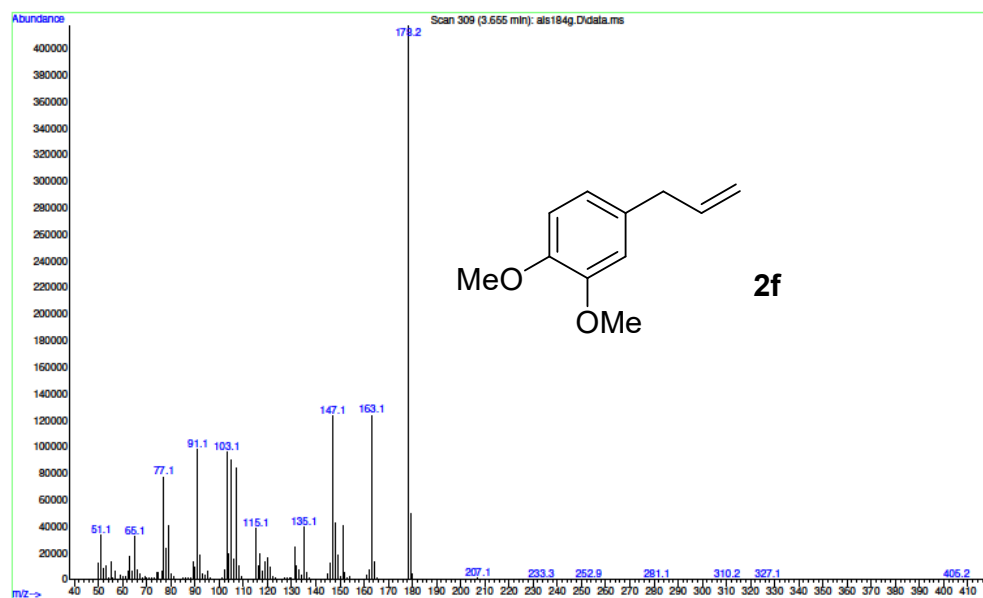
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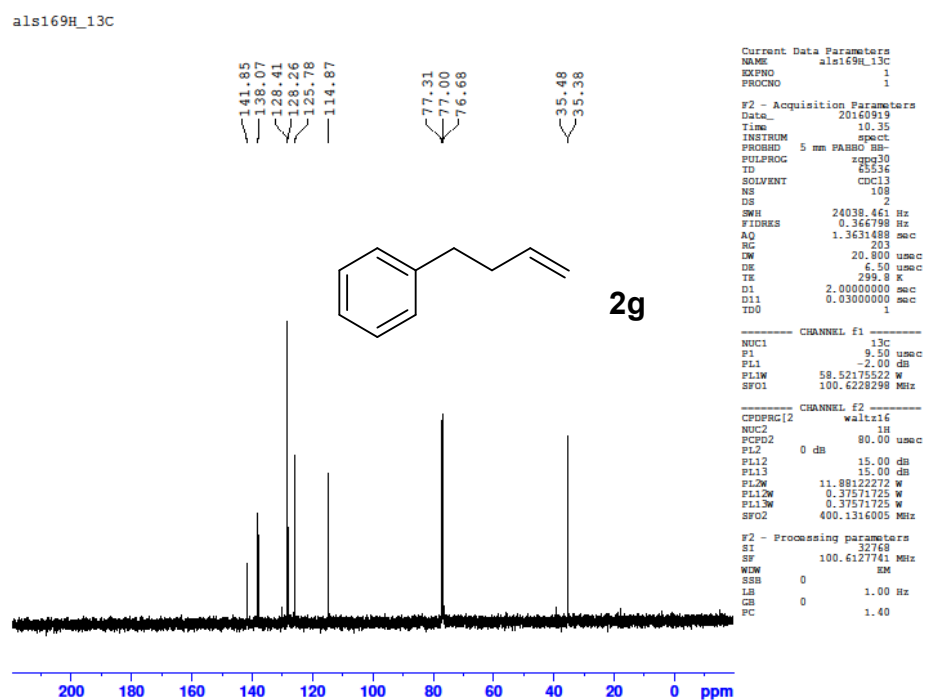
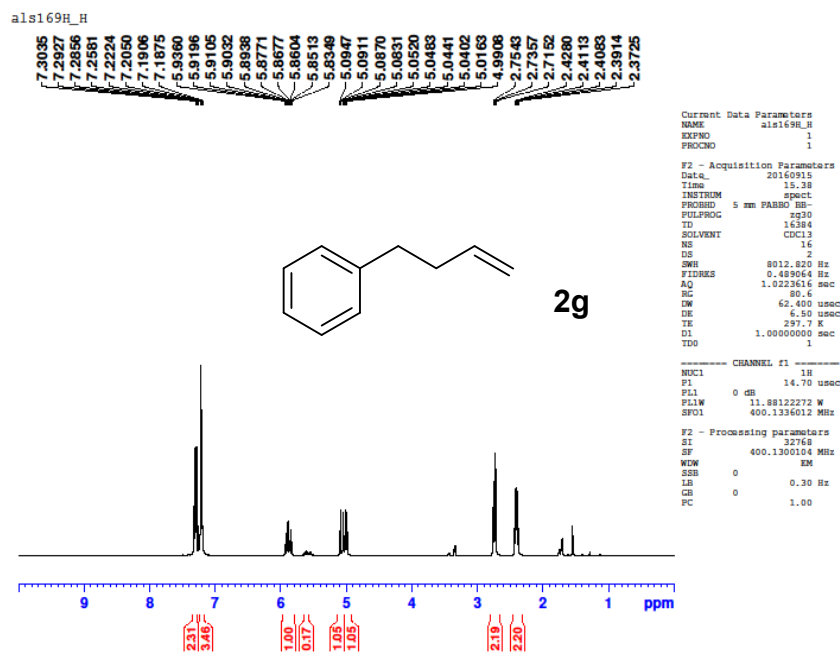
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Instrument : 5977
Sample Name: als184et2
Misc Info :
Vial Number: 2



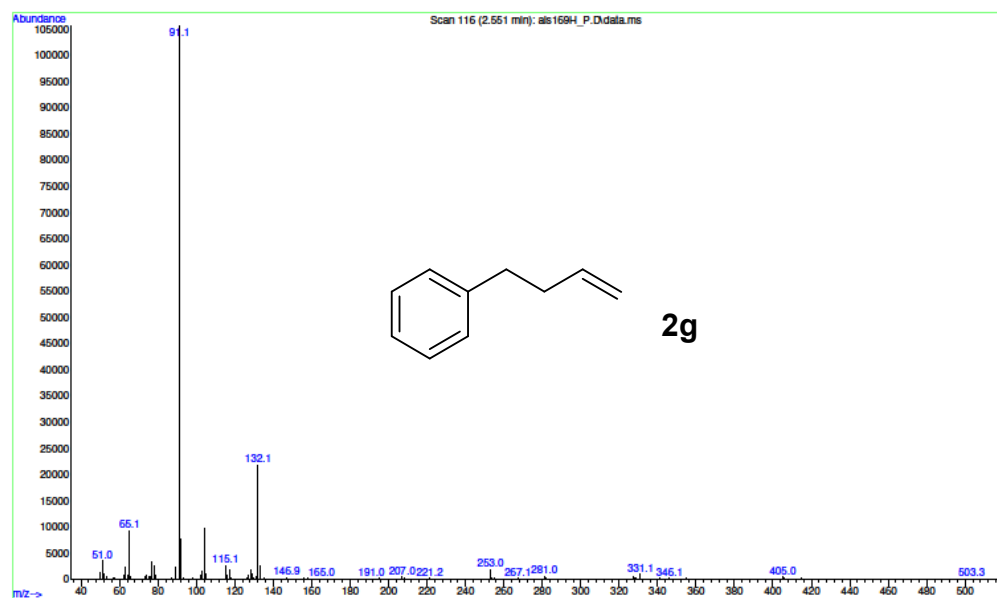


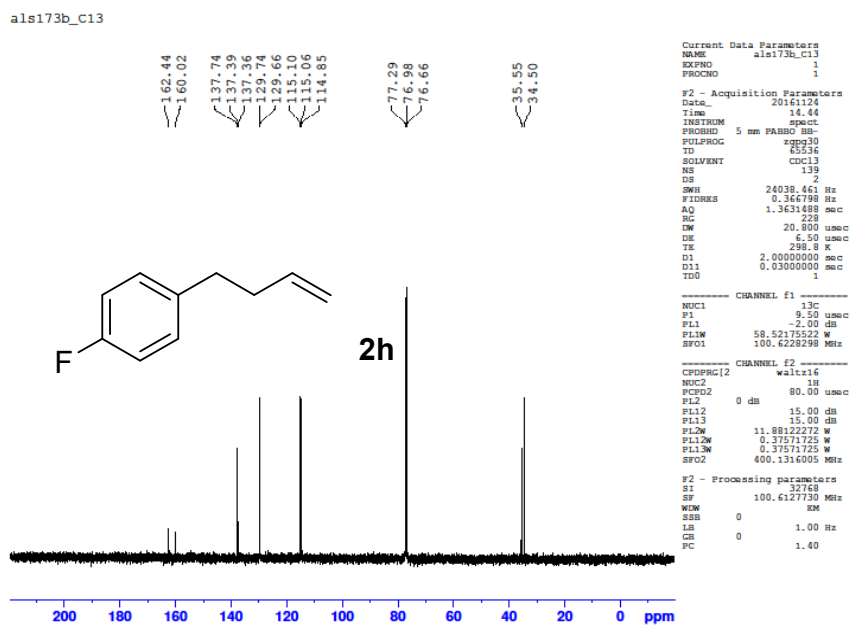
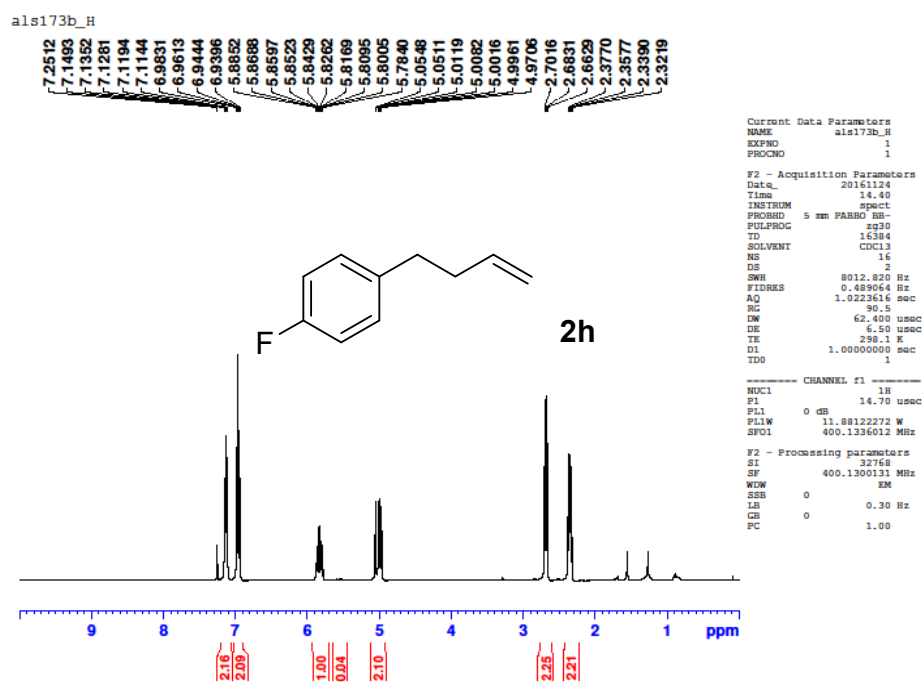
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Vial Number: 4



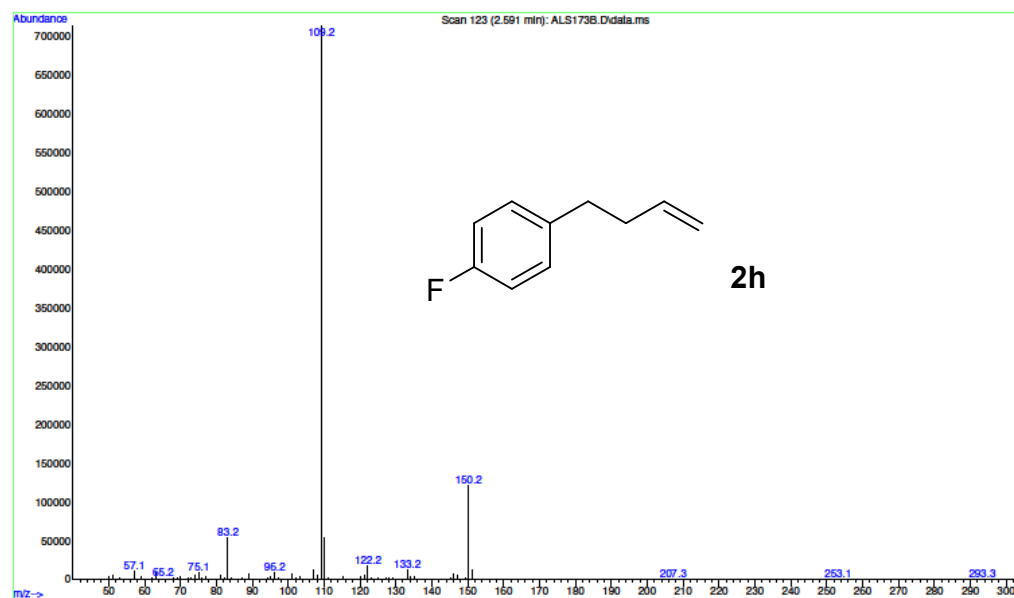


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Misc Info :
Vial Number: 20

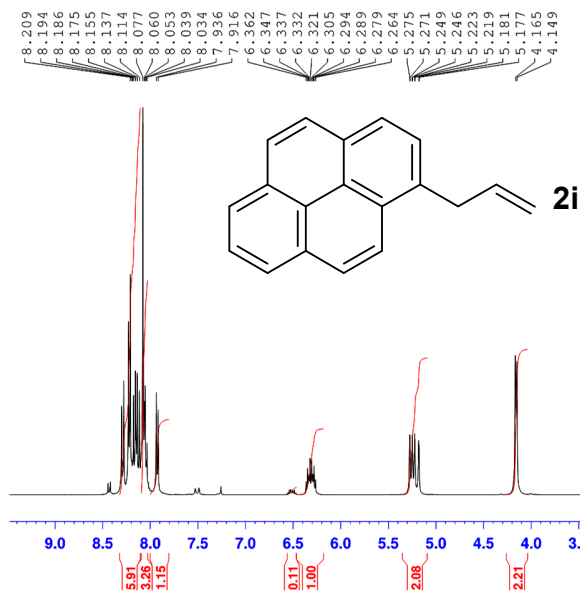




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Misc Info :
Vial Number: 2



als175e_H



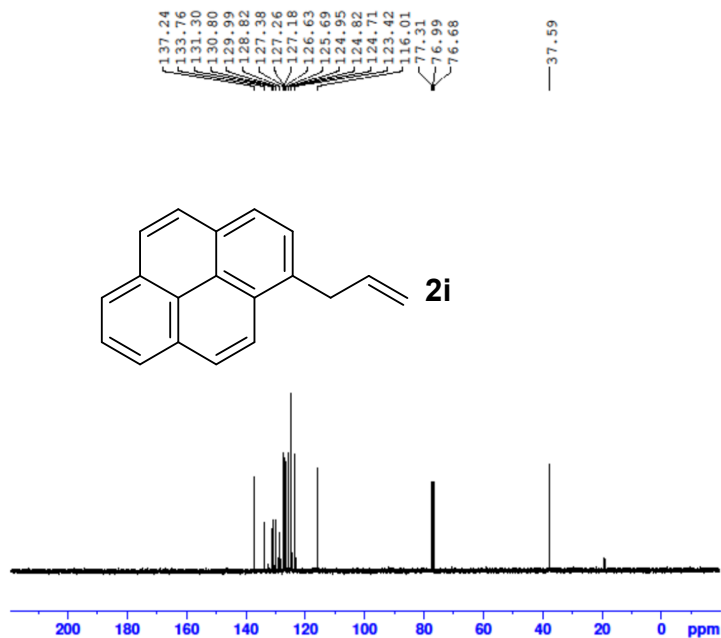
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 PROCNO 1

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 DE 6.50 usec
 TE 297.9 K
 D1 1.00000000 sec
 TDO 1

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 SFO1 400.1336012 MHz

F2 - Processing parameters
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 WDW EM
 SSB 0
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 PC 1.00

als175e_C13



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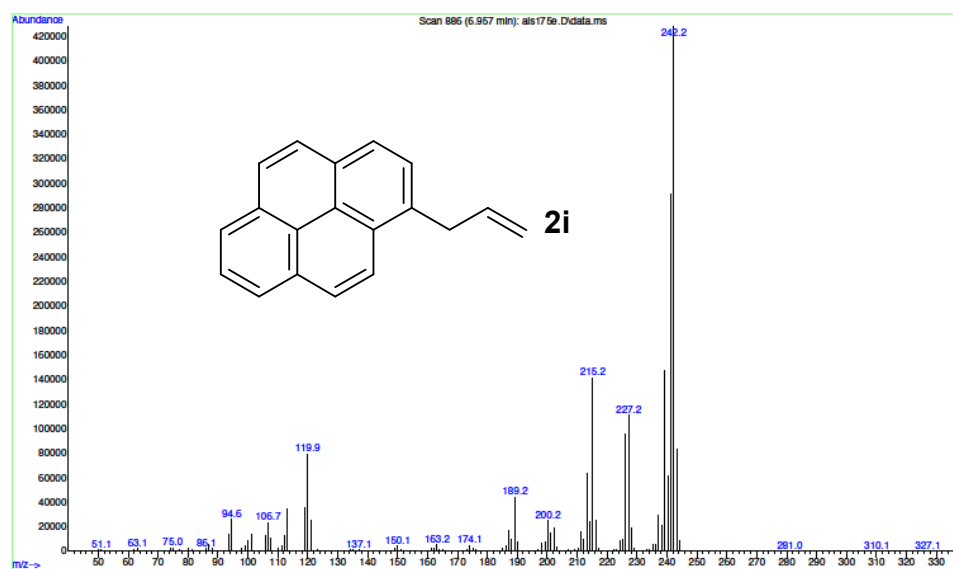
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 SFO1 100.6228298 MHz

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 PL13 15.00 dB
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Instrument : 5977
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8. X-ray crystalline structure for Pd-complexes

Figure S6: Crystalline structure of $[\text{PdCl}_2(\text{C}_{28}\text{H}_{21}\text{P})]_2$, Pd-CP-1 complex

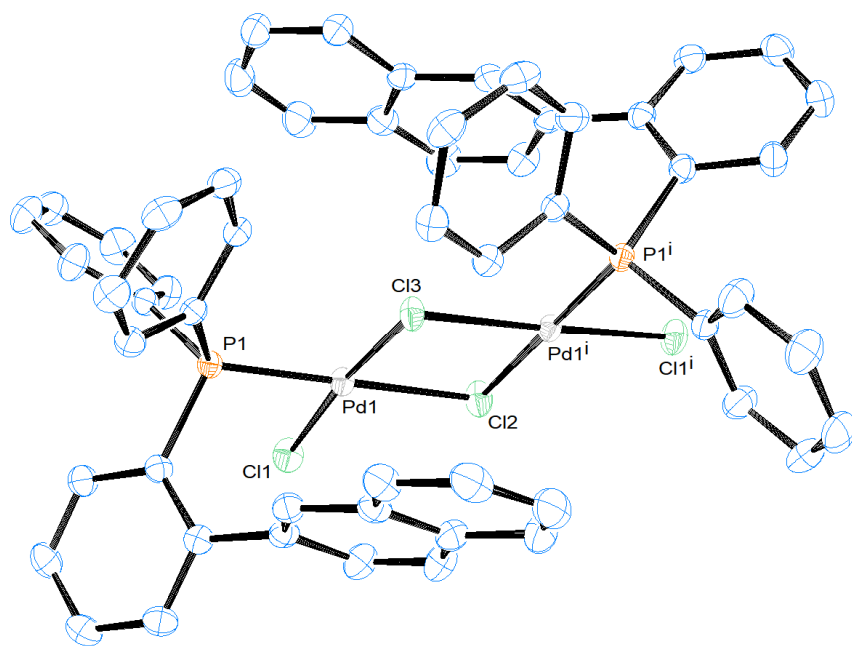


Table S2. Crystal data and structure refinement for [PdCl₂(C₂₈H₂₁P)]₂, Pd-CP-1.

Identification code	ALS1702
Empirical formula	C ₅₈ H ₄₆ Cl ₈ P ₂ Pd ₂
Formula weight	1301.29
Temperature	298(2) K
Wavelength	0.71073 Å
Crystal system, space group	Monoclinic, C 2/c
Unit cell dimensions	a = 26.9421(86) Å alpha = 90 deg. b = 11.1527(33) Å beta = 114.7781(86) deg. c = 19.7203(59) Å gamma = 90 deg.
Volume	5380(3) Å ³
Z, Calculated density	4, 1.607 Mg/m ³
Absorption coefficient	1.164 mm ⁻¹
F(000)	2608
Crystal size	0.16 x 0.10 x 0.08 mm
Theta range for data collection	2.76 to 27.51 deg.
Limiting indices	-34 ≤ h ≤ 34, -14 ≤ k ≤ 14, -25 ≤ l ≤ 25
Reflections collected / unique	60625 / 6168 [R(int) = 0.0426]
Completeness to theta = 27.51	99.5 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.9126 and 0.8356
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	6168 / 0 / 317

Goodness-of-fit on F^2 1.029

Final R indices [$I > 2\sigma(I)$] $R_1 = 0.0342$, $wR_2 = 0.0774$

R indices (all data) $R_1 = 0.0515$, $wR_2 = 0.0850$

Largest diff. peak and hole 0.675 and -0.910 e. \AA^{-3}

Table S3. Bond lengths [Å] and angles [deg] for [PdCl₂(C₂₈H₂₁P)]₂, Pd-CP-1.

Pd(1)-P(1)	2.2563(8)
Pd(1)-Cl(1)	2.2937(9)
Pd(1)-Cl(3)	2.3181(8)
Pd(1)-Cl(2)	2.4293(9)
Cl(2)-Pd(1)#1	2.4293(9)
Cl(3)-Pd(1)#1	2.3181(8)
P(1)-C(1)	1.820(3)
P(1)-C(13)	1.824(3)
P(1)-C(7)	1.830(3)
C(1)-C(6)	1.389(4)
C(1)-C(2)	1.392(4)
C(2)-C(3)	1.378(4)
C(2)-H(2A)	0.9300
C(3)-C(4)	1.375(5)
C(3)-H(3A)	0.9300
C(4)-C(5)	1.374(5)
C(4)-H(4A)	0.9300
C(5)-C(6)	1.381(4)
C(5)-H(5A)	0.9300
C(6)-H(6A)	0.9300
C(7)-C(8)	1.383(4)
C(7)-C(12)	1.386(4)
C(8)-C(9)	1.395(5)
C(8)-H(8A)	0.9300
C(9)-C(10)	1.364(6)
C(9)-H(9A)	0.9300
C(10)-C(11)	1.363(5)
C(10)-H(10A)	0.9300
C(11)-C(12)	1.387(5)
C(11)-H(11A)	0.9300
C(12)-H(12A)	0.9300

C(13)-C(14)	1.396(4)
C(13)-C(18)	1.399(4)
C(14)-C(15)	1.384(4)
C(14)-H(14A)	0.9300
C(15)-C(16)	1.366(5)
C(15)-H(15A)	0.9300
C(16)-C(17)	1.385(4)
C(16)-H(16A)	0.9300
C(17)-C(18)	1.390(4)
C(17)-H(17A)	0.9300
C(18)-C(19)	1.504(4)
C(19)-C(20)	1.364(4)
C(19)-C(28)	1.414(4)
C(20)-C(21)	1.431(4)
C(20)-H(20A)	0.9300
C(21)-C(26)	1.407(4)
C(21)-C(22)	1.410(4)
C(22)-C(23)	1.370(5)
C(22)-H(22A)	0.9300
C(23)-C(24)	1.398(6)
C(23)-H(23A)	0.9300
C(24)-C(25)	1.342(6)
C(24)-H(24A)	0.9300
C(25)-C(26)	1.439(5)
C(25)-H(25A)	0.9300
C(26)-C(27)	1.396(5)
C(27)-C(28)	1.372(5)
C(27)-H(27A)	0.9300
C(28)-H(28A)	0.9300
C(29)-Cl(4)	1.698(5)
C(29)-Cl(5)	1.712(5)
C(29)-H(29A)	0.9700
C(29)-H(29B)	0.9700

P(1)-Pd(1)-Cl(1)	87.83(3)
P(1)-Pd(1)-Cl(3)	96.18(3)
Cl(1)-Pd(1)-Cl(3)	173.89(3)
P(1)-Pd(1)-Cl(2)	179.65(2)
Cl(1)-Pd(1)-Cl(2)	92.09(4)
Cl(3)-Pd(1)-Cl(2)	83.93(3)
Pd(1)#1-Cl(2)-Pd(1)	93.09(4)
Pd(1)-Cl(3)-Pd(1)#1	99.06(4)
C(1)-P(1)-C(13)	106.77(13)
C(1)-P(1)-C(7)	101.29(13)

C(13)-P(1)-C(7)	106.90(13)
C(1)-P(1)-Pd(1)	117.12(9)
C(13)-P(1)-Pd(1)	108.83(9)
C(7)-P(1)-Pd(1)	115.14(10)
C(6)-C(1)-C(2)	118.7(3)
C(6)-C(1)-P(1)	121.5(2)
C(2)-C(1)-P(1)	119.6(2)
C(3)-C(2)-C(1)	120.2(3)
C(3)-C(2)-H(2A)	119.9
C(1)-C(2)-H(2A)	119.9
C(4)-C(3)-C(2)	120.5(3)
C(4)-C(3)-H(3A)	119.7
C(2)-C(3)-H(3A)	119.7
C(5)-C(4)-C(3)	119.7(3)
C(5)-C(4)-H(4A)	120.1
C(3)-C(4)-H(4A)	120.1
C(4)-C(5)-C(6)	120.3(3)
C(4)-C(5)-H(5A)	119.9
C(6)-C(5)-H(5A)	119.9
C(5)-C(6)-C(1)	120.4(3)
C(5)-C(6)-H(6A)	119.8
C(1)-C(6)-H(6A)	119.8
C(8)-C(7)-C(12)	118.6(3)
C(8)-C(7)-P(1)	122.0(2)
C(12)-C(7)-P(1)	119.3(2)
C(7)-C(8)-C(9)	119.7(3)
C(7)-C(8)-H(8A)	120.2
C(9)-C(8)-H(8A)	120.2
C(10)-C(9)-C(8)	120.8(4)
C(10)-C(9)-H(9A)	119.6
C(8)-C(9)-H(9A)	119.6
C(11)-C(10)-C(9)	120.2(3)
C(11)-C(10)-H(10A)	119.9
C(9)-C(10)-H(10A)	119.9
C(10)-C(11)-C(12)	119.8(3)
C(10)-C(11)-H(11A)	120.1
C(12)-C(11)-H(11A)	120.1
C(7)-C(12)-C(11)	120.9(3)
C(7)-C(12)-H(12A)	119.5
C(11)-C(12)-H(12A)	119.5
C(14)-C(13)-C(18)	119.4(3)
C(14)-C(13)-P(1)	119.5(2)
C(18)-C(13)-P(1)	121.0(2)
C(15)-C(14)-C(13)	120.8(3)

C(15)-C(14)-H(14A)	119.6
C(13)-C(14)-H(14A)	119.6
C(16)-C(15)-C(14)	119.9(3)
C(16)-C(15)-H(15A)	120.1
C(14)-C(15)-H(15A)	120.1
C(15)-C(16)-C(17)	120.1(3)
C(15)-C(16)-H(16A)	119.9
C(17)-C(16)-H(16A)	119.9
C(16)-C(17)-C(18)	121.2(3)
C(16)-C(17)-H(17A)	119.4
C(18)-C(17)-H(17A)	119.4
C(17)-C(18)-C(13)	118.7(3)
C(17)-C(18)-C(19)	117.7(3)
C(13)-C(18)-C(19)	123.6(2)
C(20)-C(19)-C(28)	119.0(3)
C(20)-C(19)-C(18)	121.1(3)
C(28)-C(19)-C(18)	119.7(3)
C(19)-C(20)-C(21)	121.2(3)
C(19)-C(20)-H(20A)	119.4
C(21)-C(20)-H(20A)	119.4
C(26)-C(21)-C(22)	120.1(3)
C(26)-C(21)-C(20)	118.7(3)
C(22)-C(21)-C(20)	121.2(3)
C(23)-C(22)-C(21)	120.0(3)
C(23)-C(22)-H(22A)	120.0
C(21)-C(22)-H(22A)	120.0
C(22)-C(23)-C(24)	120.3(4)
C(22)-C(23)-H(23A)	119.8
C(24)-C(23)-H(23A)	119.8
C(25)-C(24)-C(23)	121.1(3)
C(25)-C(24)-H(24A)	119.5
C(23)-C(24)-H(24A)	119.5
C(24)-C(25)-C(26)	120.8(4)
C(24)-C(25)-H(25A)	119.6
C(26)-C(25)-H(25A)	119.6
C(27)-C(26)-C(21)	119.4(3)
C(27)-C(26)-C(25)	123.0(3)
C(21)-C(26)-C(25)	117.6(3)
C(28)-C(27)-C(26)	120.8(3)
C(28)-C(27)-H(27A)	119.6
C(26)-C(27)-H(27A)	119.6
C(27)-C(28)-C(19)	120.9(3)
C(27)-C(28)-H(28A)	119.6
C(19)-C(28)-H(28A)	119.6

Cl(4)-C(29)-Cl(5)	115.8(3)
Cl(4)-C(29)-H(29A)	108.3
Cl(5)-C(29)-H(29A)	108.3
Cl(4)-C(29)-H(29B)	108.3
Cl(5)-C(29)-H(29B)	108.3
H(29A)-C(29)-H(29B)	107.4

Symmetry transformations used to generate equivalent atoms:

#1 -x+1,y,-z+1/2

Table S4. Torsion angles [deg] for ALS1702.

P(1)-Pd(1)-Cl(2)-Pd(1)#1	107(5)
Cl(1)-Pd(1)-Cl(2)-Pd(1)#1	-175.36(2)
Cl(3)-Pd(1)-Cl(2)-Pd(1)#1	0.0
P(1)-Pd(1)-Cl(3)-Pd(1)#1	-179.66(2)
Cl(1)-Pd(1)-Cl(3)-Pd(1)#1	49.5(3)
Cl(2)-Pd(1)-Cl(3)-Pd(1)#1	0.0
Cl(1)-Pd(1)-P(1)-C(1)	173.04(10)
Cl(3)-Pd(1)-P(1)-C(1)	-2.34(10)
Cl(2)-Pd(1)-P(1)-C(1)	-109(5)
Cl(1)-Pd(1)-P(1)-C(13)	51.90(10)
Cl(3)-Pd(1)-P(1)-C(13)	-123.48(9)
Cl(2)-Pd(1)-P(1)-C(13)	130(5)
Cl(1)-Pd(1)-P(1)-C(7)	-68.06(11)
Cl(3)-Pd(1)-P(1)-C(7)	116.56(10)
Cl(2)-Pd(1)-P(1)-C(7)	10(5)
C(13)-P(1)-C(1)-C(6)	-14.4(3)
C(7)-P(1)-C(1)-C(6)	97.3(2)
Pd(1)-P(1)-C(1)-C(6)	-136.6(2)
C(13)-P(1)-C(1)-C(2)	171.2(2)
C(7)-P(1)-C(1)-C(2)	-77.1(2)
Pd(1)-P(1)-C(1)-C(2)	49.0(2)
C(6)-C(1)-C(2)-C(3)	4.0(4)
P(1)-C(1)-C(2)-C(3)	178.5(2)
C(1)-C(2)-C(3)-C(4)	-2.0(4)
C(2)-C(3)-C(4)-C(5)	-2.0(5)
C(3)-C(4)-C(5)-C(6)	3.9(5)
C(4)-C(5)-C(6)-C(1)	-1.9(5)
C(2)-C(1)-C(6)-C(5)	-2.1(4)
P(1)-C(1)-C(6)-C(5)	-176.5(2)
C(1)-P(1)-C(7)-C(8)	149.4(3)
C(13)-P(1)-C(7)-C(8)	-99.0(3)
Pd(1)-P(1)-C(7)-C(8)	22.0(3)
C(1)-P(1)-C(7)-C(12)	-30.1(3)
C(13)-P(1)-C(7)-C(12)	81.5(3)
Pd(1)-P(1)-C(7)-C(12)	-157.4(2)
C(12)-C(7)-C(8)-C(9)	-0.1(5)
P(1)-C(7)-C(8)-C(9)	-179.5(3)
C(7)-C(8)-C(9)-C(10)	0.3(6)
C(8)-C(9)-C(10)-C(11)	-0.2(6)
C(9)-C(10)-C(11)-C(12)	0.0(6)
C(8)-C(7)-C(12)-C(11)	-0.2(5)
P(1)-C(7)-C(12)-C(11)	179.3(3)

C(10)-C(11)-C(12)-C(7)	0.2(6)
C(1)-P(1)-C(13)-C(14)	105.3(2)
C(7)-P(1)-C(13)-C(14)	-2.5(3)
Pd(1)-P(1)-C(13)-C(14)	-127.5(2)
C(1)-P(1)-C(13)-C(18)	-79.5(2)
C(7)-P(1)-C(13)-C(18)	172.8(2)
Pd(1)-P(1)-C(13)-C(18)	47.8(2)
C(18)-C(13)-C(14)-C(15)	-0.9(5)
P(1)-C(13)-C(14)-C(15)	174.4(3)
C(13)-C(14)-C(15)-C(16)	-0.2(5)
C(14)-C(15)-C(16)-C(17)	0.5(5)
C(15)-C(16)-C(17)-C(18)	0.4(5)
C(16)-C(17)-C(18)-C(13)	-1.5(5)
C(16)-C(17)-C(18)-C(19)	179.1(3)
C(14)-C(13)-C(18)-C(17)	1.7(4)
P(1)-C(13)-C(18)-C(17)	-173.5(2)
C(14)-C(13)-C(18)-C(19)	-178.9(3)
P(1)-C(13)-C(18)-C(19)	5.8(4)
C(17)-C(18)-C(19)-C(20)	-99.0(3)
C(13)-C(18)-C(19)-C(20)	81.6(4)
C(17)-C(18)-C(19)-C(28)	75.8(4)
C(13)-C(18)-C(19)-C(28)	-103.6(3)
C(28)-C(19)-C(20)-C(21)	0.4(4)
C(18)-C(19)-C(20)-C(21)	175.3(3)
C(19)-C(20)-C(21)-C(26)	1.4(4)
C(19)-C(20)-C(21)-C(22)	-177.6(3)
C(26)-C(21)-C(22)-C(23)	1.5(5)
C(20)-C(21)-C(22)-C(23)	-179.5(3)
C(21)-C(22)-C(23)-C(24)	0.4(5)
C(22)-C(23)-C(24)-C(25)	-1.3(6)
C(23)-C(24)-C(25)-C(26)	0.2(6)
C(22)-C(21)-C(26)-C(27)	177.0(3)
C(20)-C(21)-C(26)-C(27)	-2.0(4)
C(22)-C(21)-C(26)-C(25)	-2.5(4)
C(20)-C(21)-C(26)-C(25)	178.5(3)
C(24)-C(25)-C(26)-C(27)	-177.8(4)
C(24)-C(25)-C(26)-C(21)	1.7(5)
C(21)-C(26)-C(27)-C(28)	0.8(5)
C(25)-C(26)-C(27)-C(28)	-179.7(3)
C(26)-C(27)-C(28)-C(19)	1.0(6)
C(20)-C(19)-C(28)-C(27)	-1.7(5)
C(18)-C(19)-C(28)-C(27)	-176.6(3)

Symmetry transformations used to generate equivalent atoms:

#1 $-x+1, y, -z+1/2$

Figure S7: Crystalline structure of $\text{Pd}(\text{C}_{27}\text{H}_{20}\text{NP})(\text{CH}_3\text{CO}_2)_2$, Pd-NP-1 complex

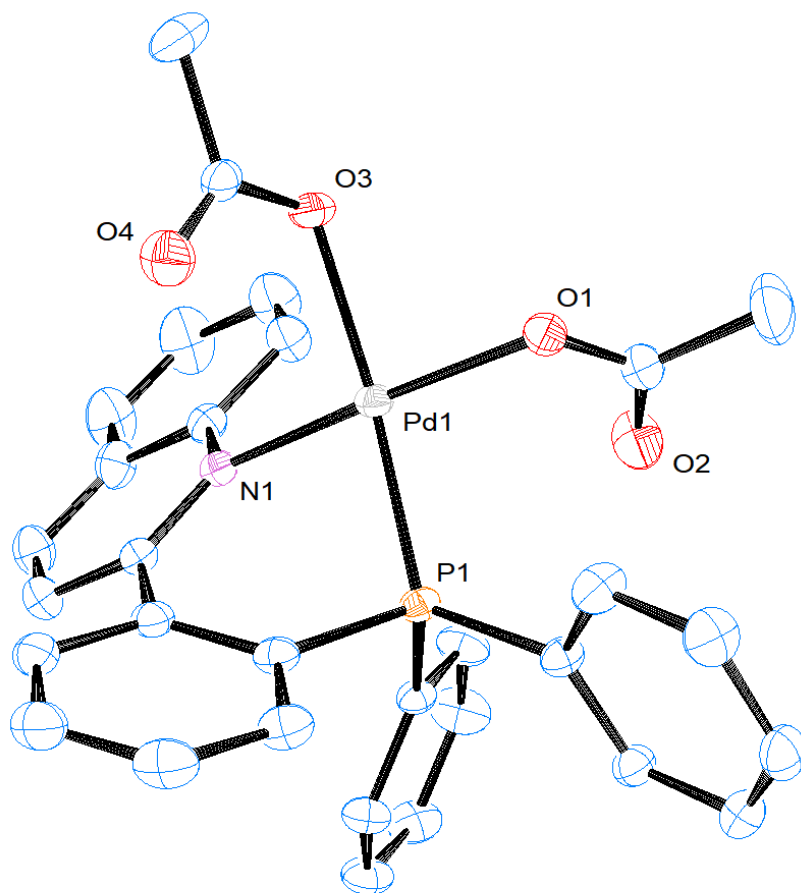


Table S3. Crystal data and structure refinement for**Pd(C₂₇H₂₀NP)(CH₃CO₂)₂.****Pd-NP-1 complex.**

Identification code	yoy6	
Empirical formula	Pd(C ₂₇ H ₂₀ NP)(CH ₃ CO ₂) ₂ .CH ₂ Cl ₂	
Formula weight	698.82	
Temperature	297(2) K	
Wavelength	0.71073 Å	
Crystal system	Triclinic	
Space group	P-1	
Unit cell dimensions	a = 10.0468(5) Å	∠ = 81.570(2)°.
	b = 12.3263(7) Å	∠ = 75.746(2)°.
	c = 13.6449(8) Å	∠ = 69.269(2)°.
Volume	1528.42(15) Å ³	
Z	2	
Density (calculated)	1.518 Mg/m ³	
Absorption coefficient	0.872 mm ⁻¹	
F(000)	708	
Crystal size	0.12 x 0.06 x 0.04 mm ³	
Theta range for data collection	2.22 to 30.61°.	
Index ranges	-14 ≤ h ≤ 14, -17 ≤ k ≤ 17, -19 ≤ l ≤ 19	
Reflections collected	135040	
Independent reflections	9404 [R(int) = 0.0355]	
Completeness to theta = 30.61°	99.7 %	
Absorption correction	None	
Max. and min. transmission	0.7461 and 0.7084	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	9404 / 0 / 386	
Goodness-of-fit on F ²	1.004	
Final R indices [I > 2σ(I)]	R1 = 0.0377, wR2 = 0.1031	
R indices (all data)	R1 = 0.0498, wR2 = 0.1175	
Largest diff. peak and hole	1.080 and -0.961 e.Å ⁻³	

Table 2. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for YOY6. $U(\text{eq})$ is defined as one third of the trace of the orthogonalized U^{ij} tensor.

	x	y	z	$U(\text{eq})$
Pd(1)	4058(1)	2545(1)	2262(1)	32(1)
P(1)	2214(1)	3634(1)	1591(1)	31(1)
O(1)	4557(1)	3896(1)	2554(1)	47(1)
O(2)	2383(2)	4511(2)	3583(1)	67(1)
O(3)	5983(1)	1448(1)	2684(1)	45(1)
O(4)	6770(2)	1053(2)	1068(1)	61(1)
N(1)	3368(2)	1194(1)	2137(1)	34(1)
C(1)	3044(2)	996(2)	1306(1)	37(1)
C(2)	2518(2)	78(2)	1296(2)	50(1)
C(3)	2310(3)	-611(2)	2143(2)	57(1)
C(4)	2604(2)	-407(2)	3040(2)	51(1)
C(5)	2384(4)	-1084(2)	3961(2)	77(1)
C(6)	2693(4)	-858(3)	4808(2)	89(1)
C(7)	3209(3)	53(2)	4788(2)	69(1)
C(8)	3434(3)	732(2)	3919(2)	51(1)
C(9)	3142(2)	510(2)	3021(1)	39(1)
C(10)	3187(2)	1766(2)	369(1)	38(1)
C(11)	2768(2)	2979(2)	382(1)	37(1)
C(12)	2862(2)	3644(2)	-533(2)	47(1)
C(13)	3357(2)	3122(2)	-1450(2)	58(1)
C(14)	3757(3)	1935(2)	-1462(2)	60(1)
C(15)	3659(2)	1257(2)	-558(2)	50(1)
C(16)	465(2)	3482(1)	2177(1)	32(1)
C(17)	-496(2)	3468(2)	1604(2)	44(1)
C(18)	-1829(2)	3355(2)	2079(2)	52(1)
C(19)	-2208(2)	3255(2)	3109(2)	57(1)
C(20)	-1261(3)	3259(3)	3682(2)	64(1)
C(21)	80(2)	3371(2)	3223(2)	50(1)
C(22)	1930(2)	5163(2)	1272(1)	35(1)
C(23)	544(2)	5971(2)	1362(2)	40(1)

Supporting information

C(24)	362(2)	7123(2)	1030(2)	50(1)
C(25)	1546(3)	7470(2)	618(2)	52(1)
C(26)	2929(3)	6684(2)	538(2)	56(1)
C(27)	3136(2)	5527(2)	869(2)	50(1)
C(28)	3591(2)	4572(2)	3207(2)	50(1)
C(29)	4032(3)	5536(2)	3453(3)	93(1)
C(30)	6949(2)	957(2)	1935(2)	40(1)
C(31)	8394(3)	218(3)	2192(3)	77(1)
C(32)	-1263(5)	-3615(4)	4140(3)	111(1)
Cl(1)	-3158(2)	-2997(2)	4085(1)	194(1)
Cl(2)	-478(2)	-2605(1)	3974(1)	143(1)

Table 3. Bond lengths [\AA] and angles [$^\circ$] for YOY6.

Pd(1)-O(1)	2.0164(15)
Pd(1)-N(1)	2.0601(16)
Pd(1)-O(3)	2.0901(13)
Pd(1)-P(1)	2.1885(4)
P(1)-C(16)	1.8052(16)
P(1)-C(22)	1.8065(18)
P(1)-C(11)	1.8166(19)
O(1)-C(28)	1.284(2)
O(2)-C(28)	1.220(3)
O(3)-C(30)	1.276(2)
O(4)-C(30)	1.223(3)
N(1)-C(1)	1.331(2)
N(1)-C(9)	1.385(2)
C(1)-C(2)	1.411(3)
C(1)-C(10)	1.485(3)
C(2)-C(3)	1.349(3)
C(2)-H(2A)	0.94(3)
C(3)-C(4)	1.404(4)
C(3)-H(3A)	0.9300
C(4)-C(9)	1.409(3)
C(4)-C(5)	1.419(3)
C(5)-C(6)	1.357(5)
C(5)-H(5A)	0.9300
C(6)-C(7)	1.387(5)
C(6)-H(6A)	0.9300
C(7)-C(8)	1.367(3)
C(7)-H(7A)	0.9300
C(8)-C(9)	1.413(3)
C(8)-H(8A)	0.9300
C(10)-C(15)	1.394(3)
C(10)-C(11)	1.404(3)
C(11)-C(12)	1.391(3)
C(12)-C(13)	1.389(3)

C(12)-H(12A)	0.9300
C(13)-C(14)	1.375(4)
C(13)-H(13A)	0.9300
C(14)-C(15)	1.388(3)
C(14)-H(14A)	0.9300
C(15)-H(15A)	0.9300
C(16)-C(21)	1.383(3)
C(16)-C(17)	1.390(3)
C(17)-C(18)	1.383(3)
C(17)-H(17A)	0.95(2)
C(18)-C(19)	1.361(3)
C(18)-H(18A)	0.9300
C(19)-C(20)	1.374(4)
C(19)-H(19A)	0.9300
C(20)-C(21)	1.384(3)
C(20)-H(20A)	0.9300
C(21)-H(21A)	0.9300
C(22)-C(23)	1.385(2)
C(22)-C(27)	1.399(3)
C(23)-C(24)	1.388(3)
C(23)-H(23A)	0.9300
C(24)-C(25)	1.369(3)
C(24)-H(24A)	1.03(3)
C(25)-C(26)	1.374(3)
C(25)-H(25A)	0.93(2)
C(26)-C(27)	1.388(3)
C(26)-H(26A)	0.9300
C(27)-H(27A)	0.9300
C(28)-C(29)	1.512(4)
C(29)-H(29A)	0.9600
C(29)-H(29B)	0.9600
C(29)-H(29C)	0.9600
C(30)-C(31)	1.511(3)
C(31)-H(31A)	0.9600
C(31)-H(31B)	0.9600

C(31)-H(31C)	0.9600
C(32)-Cl(2)	1.658(5)
C(32)-Cl(1)	1.799(5)
C(32)-H(32A)	0.9700
C(32)-H(32B)	0.9700
O(1)-Pd(1)-N(1)	172.84(6)
O(1)-Pd(1)-O(3)	87.72(6)
N(1)-Pd(1)-O(3)	93.76(6)
O(1)-Pd(1)-P(1)	94.60(4)
N(1)-Pd(1)-P(1)	84.95(4)
O(3)-Pd(1)-P(1)	171.50(4)
C(16)-P(1)-C(22)	107.64(8)
C(16)-P(1)-C(11)	106.12(9)
C(22)-P(1)-C(11)	105.15(9)
C(16)-P(1)-Pd(1)	116.67(6)
C(22)-P(1)-Pd(1)	120.49(6)
C(11)-P(1)-Pd(1)	98.70(5)
C(28)-O(1)-Pd(1)	115.41(14)
C(30)-O(3)-Pd(1)	112.74(12)
C(1)-N(1)-C(9)	119.49(17)
C(1)-N(1)-Pd(1)	124.92(12)
C(9)-N(1)-Pd(1)	115.43(13)
N(1)-C(1)-C(2)	121.37(18)
N(1)-C(1)-C(10)	120.37(17)
C(2)-C(1)-C(10)	118.23(18)
C(3)-C(2)-C(1)	120.2(2)
C(3)-C(2)-H(2A)	122.4(18)
C(1)-C(2)-H(2A)	117.4(18)
C(2)-C(3)-C(4)	119.9(2)
C(2)-C(3)-H(3A)	120.0
C(4)-C(3)-H(3A)	120.0
C(3)-C(4)-C(9)	118.4(2)
C(3)-C(4)-C(5)	122.8(2)
C(9)-C(4)-C(5)	118.9(2)

C(6)-C(5)-C(4)	120.6(3)
C(6)-C(5)-H(5A)	119.7
C(4)-C(5)-H(5A)	119.7
C(5)-C(6)-C(7)	120.4(3)
C(5)-C(6)-H(6A)	119.8
C(7)-C(6)-H(6A)	119.8
C(8)-C(7)-C(6)	121.2(3)
C(8)-C(7)-H(7A)	119.4
C(6)-C(7)-H(7A)	119.4
C(7)-C(8)-C(9)	120.0(2)
C(7)-C(8)-H(8A)	120.0
C(9)-C(8)-H(8A)	120.0
N(1)-C(9)-C(4)	120.62(19)
N(1)-C(9)-C(8)	120.33(19)
C(4)-C(9)-C(8)	119.05(19)
C(15)-C(10)-C(11)	119.43(18)
C(15)-C(10)-C(1)	117.74(18)
C(11)-C(10)-C(1)	122.66(16)
C(12)-C(11)-C(10)	119.04(18)
C(12)-C(11)-P(1)	121.65(16)
C(10)-C(11)-P(1)	119.23(13)
C(13)-C(12)-C(11)	120.8(2)
C(13)-C(12)-H(12A)	119.6
C(11)-C(12)-H(12A)	119.6
C(14)-C(13)-C(12)	120.1(2)
C(14)-C(13)-H(13A)	120.0
C(12)-C(13)-H(13A)	120.0
C(13)-C(14)-C(15)	120.0(2)
C(13)-C(14)-H(14A)	120.0
C(15)-C(14)-H(14A)	120.0
C(14)-C(15)-C(10)	120.6(2)
C(14)-C(15)-H(15A)	119.7
C(10)-C(15)-H(15A)	119.7
C(21)-C(16)-C(17)	119.40(17)
C(21)-C(16)-P(1)	118.99(15)

C(17)-C(16)-P(1)	121.61(14)
C(18)-C(17)-C(16)	120.06(19)
C(18)-C(17)-H(17A)	120.5(16)
C(16)-C(17)-H(17A)	119.2(16)
C(19)-C(18)-C(17)	120.3(2)
C(19)-C(18)-H(18A)	119.8
C(17)-C(18)-H(18A)	119.8
C(18)-C(19)-C(20)	120.1(2)
C(18)-C(19)-H(19A)	120.0
C(20)-C(19)-H(19A)	120.0
C(19)-C(20)-C(21)	120.6(2)
C(19)-C(20)-H(20A)	119.7
C(21)-C(20)-H(20A)	119.7
C(16)-C(21)-C(20)	119.5(2)
C(16)-C(21)-H(21A)	120.2
C(20)-C(21)-H(21A)	120.2
C(23)-C(22)-C(27)	119.36(17)
C(23)-C(22)-P(1)	121.35(14)
C(27)-C(22)-P(1)	119.17(13)
C(22)-C(23)-C(24)	119.89(19)
C(22)-C(23)-H(23A)	120.1
C(24)-C(23)-H(23A)	120.1
C(25)-C(24)-C(23)	120.48(19)
C(25)-C(24)-H(24A)	117.4(17)
C(23)-C(24)-H(24A)	122.1(17)
C(24)-C(25)-C(26)	120.4(2)
C(24)-C(25)-H(25A)	115.9(14)
C(26)-C(25)-H(25A)	123.8(14)
C(25)-C(26)-C(27)	120.1(2)
C(25)-C(26)-H(26A)	119.9
C(27)-C(26)-H(26A)	119.9
C(26)-C(27)-C(22)	119.77(18)
C(26)-C(27)-H(27A)	120.1
C(22)-C(27)-H(27A)	120.1
O(2)-C(28)-O(1)	125.3(2)

O(2)-C(28)-C(29)	120.5(2)
O(1)-C(28)-C(29)	114.1(2)
C(28)-C(29)-H(29A)	109.5
C(28)-C(29)-H(29B)	109.5
H(29A)-C(29)-H(29B)	109.5
C(28)-C(29)-H(29C)	109.5
H(29A)-C(29)-H(29C)	109.5
H(29B)-C(29)-H(29C)	109.5
O(4)-C(30)-O(3)	124.87(18)
O(4)-C(30)-C(31)	120.6(2)
O(3)-C(30)-C(31)	114.6(2)
C(30)-C(31)-H(31A)	109.5
C(30)-C(31)-H(31B)	109.5
H(31A)-C(31)-H(31B)	109.5
C(30)-C(31)-H(31C)	109.5
H(31A)-C(31)-H(31C)	109.5
H(31B)-C(31)-H(31C)	109.5
Cl(2)-C(32)-Cl(1)	111.9(3)
Cl(2)-C(32)-H(32A)	109.2
Cl(1)-C(32)-H(32A)	109.2
Cl(2)-C(32)-H(32B)	109.2
Cl(1)-C(32)-H(32B)	109.2
H(32A)-C(32)-H(32B)	107.9

Symmetry transformations used to generate equivalent atoms:

Table 4. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for YOY6. The anisotropic displacement factor exponent takes the form: $-2\pi^2 [h^2 a^{*2} U^{11} + \dots + 2 h k a^* b^* U^{12}]$

	U^{11}	U^{22}	U^{33}	U^{23}	U^{13}	U^{12}
Pd(1)	25(1)	32(1)	39(1)	-2(1)	-10(1)	-8(1)
P(1)	24(1)	32(1)	35(1)	1(1)	-6(1)	-8(1)
O(1)	33(1)	43(1)	69(1)	-9(1)	-11(1)	-16(1)
O(2)	59(1)	76(1)	68(1)	-22(1)	6(1)	-32(1)
O(3)	33(1)	49(1)	49(1)	-6(1)	-16(1)	-4(1)
O(4)	61(1)	70(1)	50(1)	-9(1)	-7(1)	-20(1)
N(1)	31(1)	32(1)	39(1)	-3(1)	-10(1)	-8(1)
C(1)	31(1)	35(1)	43(1)	-7(1)	-10(1)	-5(1)
C(2)	51(1)	46(1)	59(1)	-12(1)	-19(1)	-17(1)
C(3)	66(1)	44(1)	73(1)	-7(1)	-17(1)	-28(1)
C(4)	59(1)	37(1)	58(1)	-1(1)	-9(1)	-20(1)
C(5)	116(2)	56(1)	71(2)	12(1)	-14(2)	-51(1)
C(6)	145(2)	70(1)	58(2)	21(1)	-18(2)	-56(2)
C(7)	99(2)	69(1)	44(1)	10(1)	-19(1)	-38(1)
C(8)	65(1)	50(1)	41(1)	3(1)	-14(1)	-25(1)
C(9)	39(1)	35(1)	42(1)	0(1)	-8(1)	-10(1)
C(10)	29(1)	45(1)	37(1)	-6(1)	-9(1)	-6(1)
C(11)	27(1)	45(1)	36(1)	-1(1)	-7(1)	-9(1)
C(12)	40(1)	55(1)	41(1)	6(1)	-7(1)	-13(1)
C(13)	48(1)	85(2)	34(1)	6(1)	-6(1)	-18(1)
C(14)	47(1)	85(2)	39(1)	-17(1)	-7(1)	-6(1)
C(15)	41(1)	58(1)	45(1)	-15(1)	-12(1)	-2(1)
C(16)	25(1)	33(1)	38(1)	2(1)	-6(1)	-10(1)
C(17)	38(1)	54(1)	45(1)	4(1)	-15(1)	-18(1)
C(18)	40(1)	61(1)	66(1)	7(1)	-22(1)	-25(1)
C(19)	35(1)	66(1)	72(1)	13(1)	-9(1)	-27(1)
C(20)	52(1)	100(2)	45(1)	8(1)	-2(1)	-39(1)
C(21)	40(1)	76(1)	39(1)	1(1)	-8(1)	-28(1)
C(22)	33(1)	34(1)	37(1)	4(1)	-7(1)	-11(1)
C(23)	35(1)	38(1)	45(1)	-3(1)	-9(1)	-9(1)

Supporting information

C(24)	51(1)	37(1)	60(1)	-2(1)	-16(1)	-8(1)
C(25)	70(1)	36(1)	53(1)	4(1)	-17(1)	-21(1)
C(26)	58(1)	52(1)	60(1)	9(1)	-7(1)	-32(1)
C(27)	38(1)	45(1)	61(1)	7(1)	-3(1)	-16(1)
C(28)	50(1)	43(1)	61(1)	-6(1)	-16(1)	-17(1)
C(29)	83(2)	71(1)	139(3)	-48(2)	-6(2)	-39(1)
C(30)	37(1)	30(1)	54(1)	-2(1)	-12(1)	-10(1)
C(31)	45(1)	69(2)	97(2)	-14(2)	-21(1)	11(1)
C(32)	119(3)	111(3)	83(2)	-22(2)	-15(2)	-13(2)
Cl(1)	217(2)	275(2)	144(1)	20(1)	-109(1)	-110(1)
Cl(2)	159(1)	109(1)	137(1)	-2(1)	-26(1)	-20(1)

Table 5. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^{-3}$) for YOY6.

	x	y	z	U(eq)
H(2A)	2310(30)	-20(20)	690(20)	68(8)
H(3A)	1972	-1220	2132	69
H(5A)	2025	-1689	3985	93
H(6A)	2558	-1315	5405	107
H(7A)	3404	204	5376	83
H(8A)	3780	1339	3920	61
H(12A)	2589	4447	-531	57
H(13A)	3419	3576	-2056	70
H(14A)	4092	1586	-2076	72
H(15A)	3910	456	-570	60
H(17A)	-190(30)	3460(20)	889(19)	52(7)
H(18A)	-2471	3348	1694	63
H(19A)	-3108	3185	3425	68
H(20A)	-1523	3186	4385	77
H(21A)	717	3371	3615	60
H(23A)	-265	5741	1644	48
H(24A)	-650(30)	7750(20)	1070(20)	70(8)
H(25A)	1340(20)	8248(19)	394(17)	41(6)
H(26A)	3729	6926	262	67
H(27A)	4073	4996	823	59
H(29A)	4623	5245	3952	190(20)
H(29B)	3177	6163	3714	230
H(29C)	4577	5813	2850	230
H(31A)	9034	-165	1604	142(17)
H(31B)	8248	-353	2728	170
H(31C)	8820	704	2407	170
H(32A)	-1172	-4035	4793	133
H(32B)	-764	-4166	3620	133

Table 6. Torsion angles [°] for YOY6.

O(1)-Pd(1)-P(1)-C(16)	-121.62(8)
N(1)-Pd(1)-P(1)-C(16)	51.21(8)
O(3)-Pd(1)-P(1)-C(16)	132.8(3)
O(1)-Pd(1)-P(1)-C(22)	11.96(8)
N(1)-Pd(1)-P(1)-C(22)	-175.21(8)
O(3)-Pd(1)-P(1)-C(22)	-93.6(3)
O(1)-Pd(1)-P(1)-C(11)	125.35(8)
N(1)-Pd(1)-P(1)-C(11)	-61.82(8)
O(3)-Pd(1)-P(1)-C(11)	19.8(3)
N(1)-Pd(1)-O(1)-C(28)	-21.0(5)
O(3)-Pd(1)-O(1)-C(28)	-123.09(16)
P(1)-Pd(1)-O(1)-C(28)	65.11(15)
O(1)-Pd(1)-O(3)-C(30)	-110.24(14)
N(1)-Pd(1)-O(3)-C(30)	76.78(14)
P(1)-Pd(1)-O(3)-C(30)	-4.2(4)
O(1)-Pd(1)-N(1)-C(1)	135.3(4)
O(3)-Pd(1)-N(1)-C(1)	-123.03(13)
P(1)-Pd(1)-N(1)-C(1)	48.54(12)
O(1)-Pd(1)-N(1)-C(9)	-40.1(5)
O(3)-Pd(1)-N(1)-C(9)	61.63(12)
P(1)-Pd(1)-N(1)-C(9)	-126.80(11)
C(9)-N(1)-C(1)-C(2)	-2.1(2)
Pd(1)-N(1)-C(1)-C(2)	-177.26(13)
C(9)-N(1)-C(1)-C(10)	176.07(14)
Pd(1)-N(1)-C(1)-C(10)	0.9(2)
N(1)-C(1)-C(2)-C(3)	1.1(3)
C(10)-C(1)-C(2)-C(3)	-177.09(18)
C(1)-C(2)-C(3)-C(4)	0.6(3)
C(2)-C(3)-C(4)-C(9)	-1.2(3)
C(2)-C(3)-C(4)-C(5)	178.8(2)
C(3)-C(4)-C(5)-C(6)	179.6(3)
C(9)-C(4)-C(5)-C(6)	-0.4(4)
C(4)-C(5)-C(6)-C(7)	1.0(5)

C(5)-C(6)-C(7)-C(8)	-0.8(5)
C(6)-C(7)-C(8)-C(9)	0.0(4)
C(1)-N(1)-C(9)-C(4)	1.4(2)
Pd(1)-N(1)-C(9)-C(4)	177.02(14)
C(1)-N(1)-C(9)-C(8)	-177.90(17)
Pd(1)-N(1)-C(9)-C(8)	-2.3(2)
C(3)-C(4)-C(9)-N(1)	0.3(3)
C(5)-C(4)-C(9)-N(1)	-179.8(2)
C(3)-C(4)-C(9)-C(8)	179.58(19)
C(5)-C(4)-C(9)-C(8)	-0.5(3)
C(7)-C(8)-C(9)-N(1)	180.0(2)
C(7)-C(8)-C(9)-C(4)	0.7(3)
N(1)-C(1)-C(10)-C(15)	143.99(18)
C(2)-C(1)-C(10)-C(15)	-37.8(2)
N(1)-C(1)-C(10)-C(11)	-40.8(2)
C(2)-C(1)-C(10)-C(11)	137.42(19)
C(15)-C(10)-C(11)-C(12)	-1.2(3)
C(1)-C(10)-C(11)-C(12)	-176.38(18)
C(15)-C(10)-C(11)-P(1)	-177.94(15)
C(1)-C(10)-C(11)-P(1)	6.9(2)
C(16)-P(1)-C(11)-C(12)	112.26(16)
C(22)-P(1)-C(11)-C(12)	-1.64(18)
Pd(1)-P(1)-C(11)-C(12)	-126.61(15)
C(16)-P(1)-C(11)-C(10)	-71.13(16)
C(22)-P(1)-C(11)-C(10)	174.97(14)
Pd(1)-P(1)-C(11)-C(10)	49.99(15)
C(10)-C(11)-C(12)-C(13)	0.1(3)
P(1)-C(11)-C(12)-C(13)	176.76(17)
C(11)-C(12)-C(13)-C(14)	0.3(4)
C(12)-C(13)-C(14)-C(15)	0.3(4)
C(13)-C(14)-C(15)-C(10)	-1.4(4)
C(11)-C(10)-C(15)-C(14)	1.9(3)
C(1)-C(10)-C(15)-C(14)	177.2(2)
C(22)-P(1)-C(16)-C(21)	-98.30(17)
C(11)-P(1)-C(16)-C(21)	149.53(16)

Pd(1)-P(1)-C(16)-C(21)	40.78(17)
C(22)-P(1)-C(16)-C(17)	82.06(17)
C(11)-P(1)-C(16)-C(17)	-30.11(17)
Pd(1)-P(1)-C(16)-C(17)	-138.86(14)
C(21)-C(16)-C(17)-C(18)	0.7(3)
P(1)-C(16)-C(17)-C(18)	-179.71(16)
C(16)-C(17)-C(18)-C(19)	-0.1(3)
C(17)-C(18)-C(19)-C(20)	-0.4(4)
C(18)-C(19)-C(20)-C(21)	0.4(4)
C(17)-C(16)-C(21)-C(20)	-0.6(3)
P(1)-C(16)-C(21)-C(20)	179.70(19)
C(19)-C(20)-C(21)-C(16)	0.1(4)
C(16)-P(1)-C(22)-C(23)	-8.39(19)
C(11)-P(1)-C(22)-C(23)	104.44(17)
Pd(1)-P(1)-C(22)-C(23)	-145.60(14)
C(16)-P(1)-C(22)-C(27)	175.58(17)
C(11)-P(1)-C(22)-C(27)	-71.59(19)
Pd(1)-P(1)-C(22)-C(27)	38.37(19)
C(27)-C(22)-C(23)-C(24)	1.5(3)
P(1)-C(22)-C(23)-C(24)	-174.49(17)
C(22)-C(23)-C(24)-C(25)	-0.3(3)
C(23)-C(24)-C(25)-C(26)	-0.7(4)
C(24)-C(25)-C(26)-C(27)	0.5(4)
C(25)-C(26)-C(27)-C(22)	0.8(4)
C(23)-C(22)-C(27)-C(26)	-1.8(3)
P(1)-C(22)-C(27)-C(26)	174.34(19)
Pd(1)-O(1)-C(28)-O(2)	-6.5(3)
Pd(1)-O(1)-C(28)-C(29)	176.7(2)
Pd(1)-O(3)-C(30)-O(4)	-3.6(3)
Pd(1)-O(3)-C(30)-C(31)	176.06(17)

Symmetry transformations used to generate equivalent atoms:

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