

Review



Electrospun Nanofibrous Scaffolds: Review of Current Progress in the Properties and Manufacturing Process, and Possible Applications for COVID-19

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Abstract: Over the last twenty years, researchers have focused on the potential applications of electrospinning, especially its scalability and versatility. Specifically, electrospun nanofiber scaffolds are considered an emergent technology and a promising approach that can be applied to biosensing, drug delivery, soft and hard tissue repair and regeneration, and wound healing. Several parameters control the functional scaffolds, such as fiber geometrical characteristics and alignment, architecture, etc. As it is based on nanotechnology, the concept of this approach has shown a strong evolution in terms of the forms of the materials used (aerogels, microspheres, etc.), the incorporated microorganisms used to treat diseases (cells, proteins, nuclei acids, etc.), and the manufacturing process in relation to the control of adhesion, proliferation, and differentiation of the mimetic nanofibers. However, several difficulties are still considered as huge challenges for scientists to overcome in relation to scaffolds design and properties (hydrophilicity, biodegradability, and biocompatibility) but also in relation to transferring biological nanofibers products into practical industrial use by way of a highly efficient bio-solution. In this article, the authors review current progress in the materials and processes used by the electrospinning technique to develop novel fibrous scaffolds with suitable design and that more closely mimic structure. A specific interest will be given to the use of this approach as an emergent technology for the treatment of bacteria and viruses such as COVID-19.

Keywords: electrospinning technique; nanofiber scaffolds; bio-solution; innovative process; new biotechnology applications; COVID-19

1. Overview of the Electrospinning Technique

1.1. State of the Art

Electrospinning is widely attractive to industry and researchers for its scalability, versatility, and potential applications in many fields [1,2]. It is considered one of the most suitable techniques for fabricating nanofibrous scaffolds, which are known for their high physical porosity and huge potential to mimic defects, such as bone defects [3–5]. In terms of geometry, the diameter of each electrospun fiber depends on the polymer specificity and electrospinning processing parameters [3]. The electrospinning set-up consists of a high voltage source, an infusion syringe pump, and a collector; the collector might be stationary or portable metal or a coagulating bath [4–6]. Electrospinning technique produces more thinner, smoother, and folded scaffolds and achieves more uniform drug distribution with less residual liquid than the solvent casting [7]. Therefore, electrospinning induces well-controlled drug release profiles. In the next paragraph, the potential applications of nanofiber scaffolds will be further highlighted.



Citation: Kchaou, M.; Alquraish, M.; Abuhasel, K.; Abdullah, A.; Ali, A.A. Electrospun Nanofibrous Scaffolds: Review of Current Progress in the Properties and Manufacturing Process, and Possible Applications for COVID-19. *Polymers* **2021**, *13*, 916. https://doi.org/10.3390/ polym13060916

Academic Editors: Gianluca Cicala and Jianxun Ding

Received: 18 February 2021 Accepted: 5 March 2021 Published: 16 March 2021

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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Nowadays electrospinning is a well-known technology and it has been under a thorough investigations. One of the earliest studies of the electrified jetting phenomenon has been published by Zeleny [8]; in this paper the role of the surface instability in electrical discharges from charged droplets has been studied. A series of patents from 1934 to 1944 has been published by Formhals [9–12]. These patents are mainly describing an experimental setup utilizing electrostatic forces to produce fine dried polymer filaments. Another apparatus for the production of patterned, ultrathin, low weight, non-woven fabrics by using electrical spinning has been studied by Taylor [14–17], Taylor described a conical stable geometry at the end of the meniscus which is now known as a Taylor cone. In 1971, Baumgarten [18] used electrospinning apparatus to produce ultra-fine acrylic fibers with diameters below 1.1 μ m down to 500 nm. Although the electrospinning process has been extensively studied by many researchers since then for several decades in the literature, many parameters are still under investigation and not yet completely understood [19].

A research team in Akron University, USA lead by Professor Reneker [20] reintroduced the electrospinning technique to make submicron fibers from different types of synthetic and natural polymers. Yaril's team reported the production of hollow nanotubes by co-electrospining two polymer solutions (PCL and PMMA nanofibers) [21,22]. Researchers particularly highlighted the desorption limited mechanism of release from polymer nanofibers [23] and discussed electrospinning jets in the form of a polymer fiber with a diameter that can often be conveniently stated in nanometers [24].

Polyacrylonitrile (PAN) and copolymers of PAN are considered a very important candidate for electrospinning as they have commercial/ technological applications. Among various precursors of carbon nanofibers (CNFs), PAN is considered the best candidate, mainly due to its high carbon yield (up to 56%), flexibility and the ease of its heat treatment stabilization stage to form a stable Zigzag structure during nitrile polymerization [25–33]. Also, PAN has excellent characteristics, such as spinnability, environmentally friendly nature and varieties of commercial applications.

Inagaki et al. [34] describes the chemistry and applications of CNFs. Barhate and Ramakrishna [35] published a review on nanofibers as a filtering media for tiny materials. Li and Xia [36] discussed about the trends in nanofibers with emphasis on electrospinning techniques to produce nanofibers. PAN nanofibers and carbon nanotube (CNT) reinforced PAN nanofibers were successfully electrospun [37]. In our research team, Ali and et al. [38–49] published a series of publications studying the characteristics of the electrospun PAN/N, N-dimethylformamide (DMF) polymer solution using different types of collectors. Hot pressing technique has introduced as well to electrospun PAN nanofibers with and without nano reinforcements to produce carbon nano fibers. In their work optimization of the process for PAN nanofibers has been introduced by response surface methodology (RSM).

RSM has been used as a successful tool for optimizing the process in both polymer electrospinning and polymer hydrogels [50–52]. Process optimization of nanofibers has been investigated by RSM in order to predict the electrospinning parameters affecting the producing nanofiber diameter in order to achieve minimum fiber diameter in nano level. A quantitative relationship between electrospinning parameters and the responses (mean diameter and standard deviation) was established and then the final multi-layers structure of nanofibers and nanoparticles has been achieved for a controlled and robust process [53–55].

In the last decade, nanofibers have been generated by electrospinning under pressurized CO_2 . In fact, an evolution of the traditional technique was clarified by several researchers by adding CO_2 in the liquid polymeric solution [56–58]. The presence of CO_2 in the solution reduced the surface tension of the liquid to be processed and its viscosity. Wahyudiono et al. [59] demonstrated that CO_2 dissolved in the starting liquid polymeric solution introduced greater flexibility in the process with respect to the classical technique by the production of micro- and nanofibers of poly(vinylpyrrolidone (PVP) through the adoption of a more advanced configuration of the process. Baldino et al. [60] reported a more developed technique called supercritical assisted electrospraying (SA-ESPR), in which the addition of supercritical CO_2 (sc- CO_2) to a starting polymeric liquid solution produced a controlled size of micro- or nanoparticles. Attempts to produce nano-composite (polymer + drug) using this electrohydrodynamic process have shown encouraging results in the last years [61].

1.2. Electrospinning Parameters

Electrospinning parameters are essential to understand not only the nature of electrospinning but also the conversion of polymer solutions into nanofibers through Electrospinning. These parameters that affecting electrospinning process and electrospun fiber diameters can be classified under three main categories named as: polymer solution, processing, and environmental.

These parameters can affect morphological characteristic of electrospun fibers as well as its size. A summary discussion of these parameters and their effect on fiber characteristics will be presented:

1.2.1. Solution Parameters

Concentration and/or Berry's Number

The concentration or Berry's number of a polymer solution or melt play a crucial role in fiber formation during the electrospinning process. Four critical concentrations from low to high should be noted:

- 1. When the concentration is very low, micro to nano beads will be obtained. Here, electrospraying occurs instead of electrospinning, owing to the low viscosity and high surface tensions of the solution [62].
- As the concentration increases, a mixture of fine beads and microfibers will be obtained [63–65].
- 3. At a specific Berry's number and when the concentration is adequate, smooth nanofibers can be obtained.
- 4. At high concentrations, no nanofibers are obtained; helix-shaped, coil structures, and microribbons will be observed [66].

In general, as the concentration or Berry's number of solution or melt respectively increases the fiber diameter increases within the spin-able range.

Molecular Weight

Molecular weight of the polymer reflects the degree of polymer chain entanglement in solutions accordingly indicating solution viscosity. In case of keeping the concentration fixed, as molecular weight of the polymer decreases beads formation rather than smooth fiber is the probability. Enhancing molecular weight value results in smooth fiber. Increasing in molecular weight results in micro-ribbon formation [67].

It is also important to note that by increasing molecular weight even with low concentration micro-ribbon could be formed [68,69].

Intrinsic Viscosity

Viscosity of polymer solution is considered one of the most important parameters affecting electrospun fiber diameter and morphology. There is a suitable viscosity for the electrospinning to form fiber [70,71]. Group of publications on the correlation between polymer solution viscosity and formation of electrospun fiber have been published [67,72,73]. Concentration, viscosity and polymer molecular weight all are correlated to each other and one parameter has been used to describe them all named Berry's number it measures the degree of chain entanglement inside the solvent and can be calculated by the product of concentration by intrinsic viscosity. Ali and et al. [38–48] correlates such parameter with the optimization of electrospun PAN fibers.

Surface Tension

Surface tension is important factor in electrospinning. In 2004, Yang and Wang investigated the influence of surface tensions on electrospun PVP fibers with three different types of solvent namely ethanol, DMF, and MC [66].

As the surface tension of the solution decreases, beaded fibers can be converted into fine fibers [74–80]. Also, in this study, the surface tension and solution Viscosity can been adjusted by changing the mass ratio of solvents mix and fiber morphologies.

If all other conditions are constant, surface tension identifies the upper and lower boundaries of the electrospinning window [77,79].

Conductivity/Surface Charge Density

Solution conductivity is specified by polymer bond type, solvent solubility parameter and any other additives such as salt due to its ionic bond type. Usually, natural polymers are polyelectrolytic in nature, in which the ions increase the charge carrying ability of the polymer jet, subjecting to higher tension under the electric field, resulting in the poor fiber formation on the other hand the synthetic polymers are tending to form good fibers [80]. With the aid of ionic salts, nanofibers with small diameter can be obtained [81].

Sometimes high solution conductivity can be also achieved by using organic acid instead of regular solvent. Hou et al [81] used formic acid as the solvent to dissolve nylon 6,6 and obtained nano fibers of 3 nm diameter with beads. In their study, small amount of pyridine has been added into the solution to enhance capacity of solution to carry charge aiming to eliminate the beads by increasing the conductivity of the solution. In general, increase in the solution conductivity the more possibility of formation of thinner fibers especially in synthetic polymers.

1.2.2. Processing Parameters

Voltage

One of the important parameter in the electrospinning process is the applied voltage value. Only if the applied voltage overcomes the surface tension of the polymer meniscus then an ejected jet comes out from Taylor Cone. However, increasing in the value of applied voltage does not in the favor of the characteristics of the electrospun fiber diameter or/and morphology. Reneker and Chun [82] have proved that there is not much effect of electric field on the diameter of electrospun polyethylene oxide (PEO) nanofibers. Several groups suggested that higher voltages than required to form ejected charged jet implies to form large fiber diameter. Zhang et al [79] theoretically studied the effect of voltage on morphologies and fiber diameters distribution with poly (vinyl alcohol) (PVA)/water solution in a theoretical modeling approach.

Another research groups showed that higher applied voltages may be increase the electrostatic repulsive force on the charged jet and accordingly smaller fiber diameter can be formed. Yuan et al [83] investigated the effect of applied voltage on morphologies and fiber alignment with polysulfone (PSF)/DMAC/acetone as model. In addition to those phenomena, some groups also demonstrated that higher voltage offers the greater probability of beads formation [62,84,85].

Definitely and at the end of the previous discussion voltage does influence fiber diameters. However, the significance level differs according to polymer type, solution concentration and finally the distance between the tip of the spinneret and the nearest point on the collector [86].

Flow Rate

Another important processing parameter is the polymer solution flow rate. Generally, lower flow rate is more recommended as the polymer solution will get enough time for polarization. If the flow rate is very high, bead fibers with thick diameter will form rather than the fine fiber. That can be easily explained due to the short drying time period that the travelling jet has taken until it reaches the collector. Yuan et al [83] studied the effect of the

flow rate on the morphologies of the PSF fibers from 20 % PSF/DMAC solution at 10 kV. In their study, bead fibers with thicker diameters obtained as the flow rate is 0.66 mL/h.

Collectors

Metal collector is acting as the conductive substrate (ground) to collect the charged fibers. Aluminum foil is used as a cover sheet attached to the stationary metal grounded screen but it is difficult to transfer the collected nanofibers to other substrates for various applications. With the need of fibers transferring, diverse collectors have been developed including cone [87], pin [88], metal grids [89], parallel or gridded bar [46], rotating rods, cylinder or wheel [90], wet coagulating bath [91].

Distance or Spinning Height (H) between the Collector and the Tip of the Spinneret

Traveling distance between the tip of the charged spinneret and the nearest point of the grounded collector certainly affect the fiber diameter and its morphologies [73]. Conceptually, as the travelling bath increases more time is consumed by the travelling jet and more chance and possibility of getting rid of the solvent and consequently drying of the jet into fine fiber on the collector surface is expected [83].

1.2.3. Environmental Parameters

Temperature and Humidity are two important environmental parameters that can also affect the fiber diameters and its morphologies. Mituppatham et al. [92] proved that as temperature increases smaller and thinner fiber diameter is collected.

Low humidity tends to dry the solvent totally and increase the velocity of the solvent evaporation. On the contrary, high humidity will lead to thicker fiber diameters. Casper et al. [93] demonstrated that the variety of humidity can also affect the surface morphologies of electrospun polystyrene (PS) fibers.

2. Potential Applications of Electrospun Nanofiber Scaffolds (ENS)

2.1. Tissue Engineering Applications

Tissue engineering integrates science of biology and medicine to design artificial organs for regeneration of tissue function [94,95]. In order to mimic the extracellular matrix and provide tissue with oxygen and nutrient circulation, functional tissues were fabricated from several materials and specific structures, particularly nanofiber scaffolds [96,97]. In fact, nanofiber scaffolds are widely used in tissue repair, whether soft or hard regeneration [98]. Faced with therapeutic problems, damaged ligament, fracted bone cartilage, and blood vessels were restored, taking advantage of ENS to repair orthopedic tissues and/or develop organs with high similarity in term of characteristics, properties, and design [99–101]. Exploring their characteristics, ENS is used for wound healing. In fact, it is considered a solution for the loss of skin integrity caused by injury or illness. It provides additional biological stimuli to support cell and tissue function. As consequence, substantial psychological balance is noticed in the tissue-engineered skin patients [102–104].

2.2. Drug or Protein Delivery

Exploring the ability to control biomaterial properties (geometry, fiber alignment fiber diameter, composition, etc.), ENS are used to incorporate drugs/proteins into scaffold [104]. In fact, known for its large surface-to-volume ratio, it was demonstrated by several researchers that ENS could be a perfect vehicle for drug delivery, either by dissolving the drug into the electrospinning solution or by mixing the drug with this solution when the drug is not soluble [105–107]. As consequence, the drug contained inside in the fiber will be released. Several solutions are used to master the adsorption of a drug, particularly when the solubility of the drug is limited. Pillay et al. [108] proposed an immersion step of nanofibers into a drug solution to let the drug molecules chemically and physically bond or a drug after electrospinning by controlling physical adsorption (both when we use simple

physical adsorption or nanoparticle assembly on the surface or even layer by layer-by-layer multilayer assembly). Chemical adsorption methods can be explored by surface activation (such as plasma or ultraviolet treatments) or bioactive molecule immobilization [110,111].

The reliability of the drug delivery application is conditioned by the drug–polymer compatibility, as well as the foreign body's interaction with natural organ or tissue [112,113]. Varied results were noticed using drugs with different bioactivities, such as antimicrobial [114,115], anticancer [116,117], anti-inflammatory [118,119], cardiovascular [120,121], or antihistamine [122,123] drugs. Another factor that can influence the drug-release kinetic is the type of the polymer used as a matrix for the nanofiber's elaboration and its architecture (sandwiched with microparticle, sandwiched with microfibers, etc.) [124].

For protein delivery, surface immobilization of bioactive molecules has been used to load proteins into ENS fibers to protect the molecules from the effects of high voltage [125]. The method consists of the fixation of the protein scaffolds surface using suitable chemical conjugation with corresponding functional groups, such as carboxylate [126]. It was reported by Tigli et al. [127] that cell fate powered by peptides is conditioned by the control of the drug feeding ratio.

Cancer has become a leading disease, causing human death worldwide [128]. Cancer is responsible for 15% of human deaths worldwide, with 1.5 million new cases expected annually. Chemotherapy, hormonal therapy, radiation therapy, immunotherapy, and surgery are the current standard methods of treatment. Chemotherapeutic drugs in clinical use such as 5-fluorouracil (5-FU), cisplatin, carboplatin, paclitaxel, gemcitabine (Gemzar), etc. are characterized by either poor bioavailability, poor selectivity and specificity, or by liver accumulation and fast renal clearance [129]. Nanotechnology's evolution offers promise to tackle these problems. For instance, anticancer drug-loaded polymeric nanoparticles (NPs) are advantages to traditional anticancer drug formulations in terms of diminishing the adverse effects of drugs and enhancing their therapeutic efficacy. Generally, through the enhanced permeability and retention effect, some NPs are accumulated preferentially in tumors. Additionally, electrospun nanofibers (NFs) formulation became a promising techniquein the delivery of drug [130] and wound dressing [131]. Due to their high porosity and important surface area to molar ratio that mimics the extracellular matrix, they gained their importance in biomedical engineering [132]. Examples of these formulations include encapsulation of anticancer drugs into biodegradable polymeric nanofibers; 5-FU and salinomycin on poly(lactic-co-glycolic acid) (PLGA) [133]; Paclitaxel on PCL [134]; Doxorubicine HCL-block-poly(ethylene glycol)-block-poly(L-lactide) (HCl on PEG–PLLA) [135], hydroxycamptothecinon (Poly(lactic acid)-Poly(ethylene oxide)) PLA-PEG [136].

Pawłowska et al. [137] highlighted the development of smart drug delivery systems using the stimuli-responsive electrospun nanofibers. Their developed nanoctruture pillows have the specificity of fast photothermal responsiveness for near-infrared (NIR) light-controlled on-demand drug delivery. The innovative platform consists of electrospun PLLA loaded with a rhodamine B drug model that encapsulates platonic hydrogel P(NIPAAm-co-NIPMAAm)/AuNR [138]. The researchers demonstrated that this emergent nanotechnology is considered an excellent candidate for achieving on-demand drug release in synergy with photothermal treatment.

2.3. Agriculture, Food Industry, and Environment

As reported in many publications, technical reports, and communications, ENS are used in the biosensing [137,138], agriculture protection [139,140], the fermentation food industry [141,142], and biocatalytic remediation of the environment and energy [143,144]. In fact, to monitor food and agriculture using simpler, faster, and less expensive sensitive detection methods of foodborne illness, researchers and developers have exploited the potential given by the electrospinning technique to innovate 1D micro- and nanobiosensors [145]. Taking into account the surface properties, the high level of porosity, and the capability to interact with green elements, nanofibers have been functionalized with several types of nanomaterials such as graphene and carbone nanotubes to acquire the

singularity of multifunctional hybrid electrospun nanofibers. These specificities enhance the reactivity of the materials, improve adsorption, and increase the sites' numbers of catalysts loading and interacting. Zhang et al. [146] proved that the quality of such biochemical sensors is attributed to the strong stretching forces associated with electrospinning that induce polymer orientation along the longer axis of the fiber chains. Therefore, high charge-carrier mobility or polarized photoluminescence can be created [147]. Recent laboratories have incorporated new nanomaterials to create what is called ESN-based chemical and hybrid biosensors, which have been applied in the agri-food sector to address food quality and safety [148–150]. The conjugated use of polymer, ceramic, and inorganic other materials are reviewed by Mercante et al. [151]. In another branch of this very fertile sector, researchers used biocompatible ESN to entrap bioactive food ingredients, both hydrophilic and hydrophobic types [152]. Besides, they synthesized a food encapsulation method based on ESN to safeguard food from oxidation and damage [153,154]. Other researchers have produced ultra-fine poly(acrylamide) PAM fibers for use as a water super-absorber and soil erosion resistant agent in irrigation systems [155]. By electrospinning a 290 nm fiber diameter, the optimization of Berry's number, spinning height, and spinning angle on PAM fiber was investigated based on empirical and experimental approaches.

Th application of ESN in environment topics and issues is varied, from energy harvesting/conversion/storage to filtration membranes and catalytic supports [156–158]. In fact, specific devices made in ENS were developed and incorporated as solar cells, rechargeable batteries, and fuel cells [159]. Exploring the possibility to electrospin metal oxides and/or carbon nanofibers, they can be used as electrode materials thanks to their properties such as high surface-to-volume ratio, short diffusion distance, and large specific surface area [160]. As a consequence, they are able to rapidly transfer electrons and ions, with a large electrode/electrolyte contact area [161].

As reported by [162–164], the mass transport of reactant is feasible using a nonwoven mat of nanofibers. In fact, thanks to their high porosity level and their good interconnection, extensive contact between reactants and active sites on electrocatalysis are provided to fabric fuel cells, particularly from polymer, ceramic, and metal electrospun nanofibers. Their durability and efficiency were confirmed by several researchers and industries [165,166].

3. Innovation in Biomimetic Design, Materials Properties, and Structure Architecture

Nowadays, great efforts are focused on innovation in the design of biomimetic electrospun scaffolds for biotechnological applications. Natural and synthetic polymers as well as ceramic and metallic materials have been tested to improve properties and structure architecture.

3.1. Biomimetic Design

Biomimicry is a technologically oriented approach focused on creating innovative solutions that are inspired by nature's wealth [167,168]. In relation to our topic, researchers have concentrated on a few key sources of inspiration, in terms of shape, function, materials, or ecosystem [169,170]. They benefit from the easily tunable compositions and structures of electrospun fibers to successfully biomimic via electrospinning.

Wei et al. [171] reported the design of nacre-inspired porous scaffolds for bone repair. Their work was the result of product design strategy that included electrospinning, phase separation, and 3D printing to elaborate layer-by-layer a composite film with nacre-like structure from nano-platelets and polyamide. Nacre has also inspired newly developed coatings and implants (from simple to complex geometries) that functionalize the biomaterials surface in order to induce desirable biological responses [172–174].

Wang et al. developed engineered biomimetic superhydrophobic surfaces of electrospun nanomaterials inspired by the lotus leaf [175]. The same property was explored by investigated silver ragwort leaf and hillock bush leaf [101,176]. Other researchers have focused on the biomimetic of structure and functions of honeycombs, polar bear fur, and spider webs to inspire tissue structure and organ architecture (membrane, bone marrow, etc.) [177,178].

3.2. Materials Properties

To electrospin submicrometric fibers, a panel of materials can be used, from a natural source such as gelatin or collagen to a material from a synthetic category such as poly-caprolactone (PCL) and polylactide (PLA), as well as to hybrid types (e.g., PCL blended with collagen) [179–183].

Cell adhesion, migration, spreading, and differentiation are the essential characteristics related to the nanofiber surface's stiffness. In fact, after implementation, a successful integrated scaffold should provide good mechanical properties in terms of rigidity and flexibility, with an optimal porosity size and calibration [184]. Pennel et al. [185] made a direct relation between the infiltration and vascularization ability of the new material and its continual stability. Liu et al. [186] demonstrated that polylactide co-trim-ethylene carbonate nanofibers showed enhanced efficiencies as scaffold materials for tissue regeneration. Bao et al. [187] developed multifunctional fibrous scaffolds with shape memory specification. They were especially useful for tissue repair. Many other polymers such as chitosan, alginate, and silk fibroin have been explored for the production of healing mechanisms due to their biodegradability, drug release ability, acceptable hydrophilicity, and non-toxicity [188–193]. From the animal resource, a polymer solution is developed. Membranes with or without active agents are obtained by electrospining. The cultivation and separation process are operated to create new cells with antibacterial properties.

3.3. Architecture

An appropriate cellular environment is required to assume available scaffold architecture. This architecture is the result of various special arrangements (aligned, random, or cross aligned) [194]. These arrangements should ensure neo-tissue elaboration and proliferation, vascularization, and integration without risk to the original tissue [195]. Lutzweiler et al. [196] considered that the most sensitive property of a scaffold's structure is its ability to diffuse nutrients and metabolites thanks to an optimal pore size and stable architecture. It needs a suitable design for cell migration into scaffold and necessary ligand density on the scaffold surface [197]. According to the principle of regeneration, it should be gradually replaced with extracellular matrix, taking into account the biodegradability of the developed tissue, without any risk of toxication or surrounding organ disturbance [198,199].

Other progress related to the architectural point of view consists of the development of gradient structures to tailor cell orientation and their extracellular matrix deposition [200]. The evolution is related to the fabrication of scaffolding with random and aligned fibers on one section. As a consequence, a graded mechanical property throughout the tissue constructs is possible [201].

4. Current Progress on Elaboration Process, Implementation, and Manufacturing

The electrospun nanofibers' properties such as morphology and diameter are influenced by intrinsic (solution properties) and extrinsic (process and environment) factors [202–204]. Dorati et al. [205] reported that solvent concentration, viscosity, electrical conductivity, and elasticity are the most influential parameters on the geometry and morphology of electrospun nanofibers. In fact, to run electrospinning components, a researcher needs a small amount of solution with a suitable concentration to achieve a smooth and uniform nanofiber. Both low or high concentrations could induce a morphological problem or diameter variations according to the interaction of the solvent concentration with the viscosity and the surface tensions effects [206,207]. As consequence, non-uniform shape and non-mastered diameters of nanofibers have a great probability of being detected. Datta and Dhara [208] combined microfabrication and rolling process to design 3D bone grafts based on 2D ENS of synthetic polymer. This approach needed a graphical design of macro-pore that could be created by a laser-engraving machine on EN sheets to facilitate cellular infiltration into the 3D scaffold. Finally, multi-scalar porosity was rolled up to obtain a 3D scaffold, which was associated to the microfabricated nanofiber sheets for a final bio-engineered organ.

As more sophisticated method, a robocasting 3D printing technique was used to develop an electrospun organ with a complex structure, paving a new way to developing unprecedented scaffold microstructure [209,210]. A considerable amount of software is needed to control geometry, architectural structure, and manufacturing parameters to result in more closely mimicking material and highly efficient biomedical scaffolds [211].

Table 1. summarizes the most important applications of the electrospun nanofibrous scaffolds.

Application	Polymer and Solvent Used	Product Characteristics	Ref.
Cosmetic mask	A siliceous sponge spicules (SSS) and polylactic acid (PLA).	A nanofiber composite (PLA/SSS) of 50–450-nm with enhanced thermal and mechanical properties; a slight enhancement in human foreskin fibroblast cell proliferation; a decent cytocompatibility; and antibacterial	[212]
	A gelatin solution prepared in ethanol extracted from Crude Carissa Carandas fruits (CCE) and incorporating acetic acid.	A smooth and continuous gelatin fibers mats (GFM) with an average diameter of 235.69 ± 10.45 nm. could be obtained with the optimal conditions of 30% (w/v) gelatin solution, $25% (v/v)$ ethanol solution, $30\% (v/v)$ acetic acid, a fixed electrostatic field strength of 20 kV and a 15 cm distance between spinneret tip and collector. When $15\% (w/w)$ CCE is used, the CCE-GFM shows high DPPH radical scavenging and tyrosinase inhibitory activity.	[213]
	Anionic surfactants added to a natural biopolymer of galacturonic acid (PGuA) to enable its electrospinning to nanofibers.	Small spindled fibers of 2 to 10 μm length and 287 to 997 nm diameter. Large continuous fibers could be produced when an amount of 10 to 30% of high molecular weight PVA is used.	[214]
Drug Delivery	A poly(vinylpyrrolidone)/PVP electrospun to encapsulate β-carotene dissolved in ethanol.	PVP/ β -carotene composite nanofibers of 176 to 306 nm average diameter were able to protect the β -carotene properties	[215]
	A blend of poly (ε-caprolactone) and poly (ethylene oxide) (PCL/PEO) incorporating a nanosized hydroxyapatite (n-HA) to carry curcumin.	A nanofiber material with slow release rate of curcumin and with a high cytotoxicity against breast cancer cell line	[216]
	Chitosan/pullulan carried by a shell of polylactic acid (PLA)	A nanofiber with improved thermal properties and rapid dissolving capability in water.	[217]
Tissue engineering	A platelet-derived growth factor (PDGF-BB) contained within a shell of polylactic acid (PLA) and encapsulated within nanofibers	A 3D scaffolding nanofibers with microporous structure, acceptable mechanical properties and high cell compatibility.	[218]
	Crystalline cellulose (NCC) in a matrix of cellulose acetate (CA) polymer.	a bio-tissues of nanofibers with uniform diameter, moderate thermal properties and improved mechanical properties of 30 MPa tensile strength and 1.597 MPa module of elasticity.	[219]
	A blend of poly(ε -caprolactone), poly(ethylene glycol), poly(ε -caprolactone) (PCEC) along with polylactide (PLA).	A biodegradable polylactide (PLA)/PCEC fibrous membranes compatible with bone tissues.	[220]

Application	Polymer and Solvent Used	Product Characteristics	Ref.
Cancer therapy	A poly(ε -caprolactone) (PCL)	A scaffolding system of long nanofibers to carry breast cancer therapy	[221,222]
Wound dressing	A zein/Graphene oxide (GO) blend. The GO is loaded by Tetracycline hydrochloride (TCH).	A nanofiber composite with enhanced mechanical properties and improved release profile.	[223]
Gene delivery	A biopolymer incorporating nano-hydroxyapatite (nHAp) modified with linear polyethylenimine (LPEI), and poly(ε-caprolactone) (PCL).	A homogeneous and cohesive composite with structural characteristics, swelling and degradation behavior dependent on the size and amount of the included inorganic particles.	[224]
Filter media.	A poly(ε-caprolactone) (PCL)	Nanofibers (NF) of average diameters of 180 and 234 nm with improved bioprotective activity and filtration efficiency	[225,226]

Table 1. Cont.

5. Electrospun Nanofiber Applications for Medical Care in the Coronavirus COVID-19 Pandemic Crisis

COVID-19 is a disease caused by a new kind of coronavirus called severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) manifested in Wuhan, Hubei, China at the end of 2019 [227]. This novel coronavirus can cause fever, respiratory failure, septic shock, and even death. Researchers believe that SARS-cov2 has zoonotic origin as bat coronaviruses, pangolin coronaviruses, and previously discovered SARS-CoV [228]. To date, COVID-19 is expected to greatly impact society and the economy and to widely change our daily lifestyle [229]. Despite the number of research studies, the high kinetics of publication, and global strategies in relation to this pandemic [230–232], development of protection methods and solutions and the search for an efficient, well-controlled, and accurate means of drug dose delivery, especially with regard to the respiratory system of newly infected COVID-19 patients by using biodegradable electrospun polymer, are still considered potential issues until an effective vaccine is developed and made widely available [233].

As soon as the pandemic appeared, Tebyetekerwa et al. [234] innovated unique electrospun nanofibers for filtration membranes in face masks. They proposed the use of durable and yet reliable electrospun nonwoven filters with 10 nm fiber diameter, as the filtration efficiency of polymeric nanofibers mats is excellent and can adsorb submicronic and nanosized organisms [235].

In this context, Zussman's research group developed a sticker to upgrade surgical masks called "Maya" [236]. This innovative product showed successful results in the protection of respirators, trapping nanometric particles and, thanks to new functionalities given by the electropunk nanofibers, efficiently neutralizing the virus from droplets that might reach the mask surface.

Bin Ding's team developed other types of masks using nanofibrous membrane-based desiccants for energy efficient humidity control and atmospheric water harvesting [237]. The new technology of a wood-inspired moisture pump was adapted based on electrospun nanofibrous membrane for solar-driven continuous indoor dehumidification.

Other smart masks were developed by DooKim's group, which was exploring electrospun nanostructure performance [238]. The researchers designed a membrane to use as an additional filter for tissue-based face masks. Thanks to its nanostructured arrangement with 100–500 nm dimeter fibers, this electrospun membrane showed an excellent filtering efficiency even after being hand washed more than 20 times.

Another new generation of masks using electrospun nanostructure was proposed by Sio et al. [236]. They developed smart self-disinfecting face masks based on a multilayer

electrospun membrane. They introduced nanoclusters and plasmonic nanoparticles with a hierarchical arrangement to realize chemically driven and on-demand anti-pathogen activities.

Suitable disinfection methods and protocols should be produced so filters can be reused without compromising filtration efficiency. Khanzada et al. [239] developed aloe vera and polyvinyl alcohol electrospun nanofibers for protective clothes. The efficiency of the innovative product was confirmed by using antimicrobial activity tests to check against Gram-positive and Gram-negative bacteria. An optimum composition for high-antimicrobial activity against *S. aureus*, compared with *E. coli* bacteria was patented. For possible use as a fast absorbent carrier of anti-COVID-19 drug delivery, some researchers are trying to prepare scaffolding that would be able to carry such drugs while controlling the degree of a drug's absorption by the human body—something that would be indispensable [240,241]. However, this new applied nanotechnology is not yet mastered for use with newly discovered drugs.

6. Challenge of the Electrospun Nanofiber Scaffolds

Based on our review study, it is clear that electrospun nanofiber scaffolds are still facing many challenges with relation to the choice of materials (properties, performance, etc.), production policy (complexity, process, rate, etc.), and manipulation (scaffold storage, cost, etc.). In some applications, such as drug delivery, the improvement of drug–polymer compatibility is a primary focus for biologists, with the need to ameliorate the rate of matrix hydration and drug diffusion of the fiber-comprising polymer. Besides, ENS encounter some other practical limitations, such as scare cell infiltration and inadequate mechanical strength for load-bearing application, for example. Therefore, researchers are still focused on innovation in design, material, and architecture, as well as innovation in the manufacturing process.

Author Contributions: Conceptualization, M.A., M.K. and A.A.A.; methodology, M.K.; validation, M.K., M.A. and A.A.; formal analysis, M.K. and M.A.; investigation, M.K., M.A., A.A.A., and K.A.; resources, M.A.; data curation, A.A.; writing—original draft preparation, M.K.; writing—review and editing, M.K. and A.A.A.; visualization, A.A. and K.A.; supervision, K.A. and A.A.; project administration, M.A.; funding acquisition, M.A. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by Deanship of Science Research at University of Bisha, Saudi Arabia, grant number (UB-COVID-09-1441), and the APC was funded by the project indicated above.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data sharing not applicable.

Acknowledgments: The authors extend their appreciation to the Deanship of Science Research at University of Bisha, Saudi Arabia, for funding this work through the COVID-19 Initiative Project under Grant number (UB-COVID-09-1441).

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Mirjalili, M.; Zohoori, S. Review for Application of Electrospinning and Electrospun Nanofibers Technology in Textile Industry. J. Nanostruct Chem. 2016, 6, 207–213. [CrossRef]
- 2. Nandana, B.; Subhas, C.K. Electrospinning: A fascinating fiber fabrication technique. *Biotechnol. Adv.* 2010, 28, 325–347.
- Ramakrishna, S.; Fujihara, K.; Teo, W.-E.; Lim, T.-C.; Ma, Z. An Introduction to Electrospinning and Nanofibers; World Scientific Publishing: Singapore, 2005.
- 4. Ghosal, K.; Chandra, A.; Praveen, G.; Snigdha, S.; Roy, S.; Agatemor, C.; Thomas, S.; Provaznik, I. Electrospinning Over Solvent Casting: Tuning of Mechanical Properties of Membranes. *Sci. Rep.* **2018**, *8*, 5058. [CrossRef] [PubMed]
- Wang, Y.; Li, W.; Xia, Y.; Jiao, X.; Chen, D. Electrospun Flexible Self-Standing γ-Alumina Fibrous Membranes and Their Potential as High-Efficiency Fine Particulate Filtration Media. J. Mater. Chem. 2014, 2, 15124–15131. [CrossRef]

- 6. Persano, L.; Camposeo, A.; Tekmen, C.; Pisignano, D. Industrial Upscaling of Electrospinning and Applications of Polymer Nanofibers: A Review. *Macromol. Mater. Eng.* 2013, 298, 504–520. [CrossRef]
- Bhattarai, R.S.; Das, A.; Alzhrani, R.M.; Kang, D.; Bhaduri, S.B.; Boddu, S.H.S. Comparison of Electrospun and Solvent Cast Polylactic Acid (PLA)/Poly (vinyl alcohol) (PVA) Inserts as Potential Ocular Drug Delivery Vehicles. *Mater. Sci. Eng.* 2017, 77, 895–903. [CrossRef] [PubMed]
- 8. Zeleny, J. Instability of Electrified Liquid Surfaces. *Phys. Rev.* 1917, 10, 1–6. [CrossRef]
- 9. Formhals, A. Processing and Apparatus for Preparing Artificial Threads. U.S. Patent 2077373, 10 February 1934.
- 10. Formhals, A. Method of Producing Artificial Fiber. U.S. Patent 2158415, 16 May 1939.
- 11. Formhals, A. Production of Artificial Fibers Forming Liquids. U.S. Patent 2323025, 29 June 1943.
- 12. Formhals, A. Method and Apparatus for Spinning. U.S. Patent 2349950, 30 May 1944.
- 13. Simmons, H.L. Process and apparatus for producing patterned non-woven fabrics. U.S. Patent 3280229, 18 October 1966.
- 14. Taylor, G.I. Disintegration of Water Drops in an Electric Field. In Proceedings of the Royal Society, Series A, Mathematical, Physical & Engineering Sciences, London, UK, 28 July 1964; Volume 280, No. 1382. pp. 383–397.
- 15. Taylor, G.I.; McEwan, A.D. Stability of a Horizontal Fluid Interface in a Vertical Electric Field. *J. Fluid Mech.* **1965**, 22, 1–15. [CrossRef]
- 16. Melcher, J.R.; Taylor, G.I. Electrohydrodynamics: A Review of the Role of Interfacial Shear Stresses. *Annu. Rev. Fluid Mech.* **1969**, *1*, 111–146. [CrossRef]
- Taylor, G.I. Electrically Driven Jets. In Proceedings of the Royal Society of London, Series A, Mathematical. Physical & Engineering Sciences, London, UK, 2 December 1969; Volume 313, pp. 453–475.
- 18. Baumgarten, P.K. Electrostatic spinning of acrylic microfibers. J. Colloid Interface Sci. 1971, 36, 7. [CrossRef]
- 19. Haung, Z.M.; Zhang, Y.Z.; Kotaki, M.; Ramakrishna, S. Composites Science and Technology. *Health* **2003**, *63*, 2223.
- Doshi, J.; Reneker, D.H. Electrospinning Process and Applications of Electrospun Fibers. *J. Electrost.* 1995, *3*, 151–160. [CrossRef]
 Zussman, E.; Yarin, A.L.; Bazilevsky, A.V.; Avrahami, R.; Feldman, M. Electrospun Polyaniline/Poly(Methyl Methacrylate)-
- Zussman, E.; Yarin, A.L.; Bazilevsky, A.V.; Avrahami, R.; Feldman, M. Electrospun Polyaniline/Poly(Methyl Methacrylate)-Derived Turbostratic Carbon Micro. *Nanotubes* 2006, *18*, 348–353. [CrossRef]
- 22. Darrell, H.R.; Alexander, L.Y. Electrospinning Jets and Polymer Nanofibers. Polymer 2008, 49, 2387–2425.
- Srikar, R.; Yarin, A.L.; Megaridis, C.M.; Bazilevsky, A.V.; Kelley, E. Desorption-Limited Mechanism of Release from Polymer Nanofibers. *Langmuir* 2008, 24, 965–974. [CrossRef] [PubMed]
- 24. Fong, H.; Reneker, D.H. Electrospinning and the formation of nanofibers. In *Structure Formation in Polymeric Fibers*; Salem, D.R., Ed.; Carl Hanser Verlag: Munich, Germany, 2001; pp. 225–246.
- Liang, Y.; Ji, L.; Guo, B.; Lin, Z.; Yao, Y.F.; Li, Y.; Alcoutlabi, M.; Qiu, Y.; Zhang, X. Preparation and Electrochemical Characterization of Ionicconducting Lithium Lanthanum Titanate Oxide/Polyacrylonitrile Submicron Composite Fiber-Based Lithium-Ion Battery Separators. J. Power Sources 2011, 196, 436–441. [CrossRef]
- Schildknecht, C.E. Vinyl and Related Polymers. In *Their Preparations, Properties and Applications in Rubbers, Plastics and in Medical and Industrial Arts*; Wiley-Interscience: New York, NY, USA, 1952; p. 3878.
- Perepelkin, K.E.; Klyuchnikova, N.V.; Kulikova, N.A. Experimental Evaluation of Man-Made Fibre Brittleness. *Fibre Chem.* 1989, 21, 145–148. [CrossRef]
- Rahaman, M.S.A.; Ismail, A.F.; Mustafa, A. A Review of Heat Treatment on Polyacrylonitrile Fiber. *Polym. Degrad. Stab.* 2007, 92, 1421–1432. [CrossRef]
- 29. Litmanovich, A.D.; Plate, N.A. Alkaline Hydrolysis of Polyacrylonitrile; on the Reaction Mechanism. *Macromol. Chem. Phys.* 2000, 201, 2176–2180. [CrossRef]
- 30. Nataraja, S.K.; Yangb, T.M. Aminabhavib, Polyacrylonitrile-Based Nanofibers-A Atate-of-the-Art Review. Prog. Polym. Sci. 2011.
- Ko, F.K.; Khan, S.; Ali, A.A.; Gogotsi, Y.; Naguib, N.; Yang, G.L.; Li, C. Structure and Properties of Carbon Nanotube Reinforced Nano-Composites. In Proceedings of the AIAA/ASME/ASCE/AHS/ASC Structures, Structural Dynamics and Materials Conference, Denver, CO, USA, 22–25 April 2002.
- Ali, A.A.; Geshury, A.J.; Ko, F. Ultra-Fine Carbon Fibers and Fibrous Structures From Electro-Spun PAN Polymer Solution. In Proceedings of the Fiber Society Annual Technical Conference, Natick, MA, USA, 16–18 October 2002.
- 33. Ko, F.; Gogotsi, Y.; Ali, A.A.; Naguib, N.; Ye, H.; Yang, G.; Li, C.; Willis, P. Electrospinning of Continuous Carbon Nanotube-Filled Nanofiber Yarns. *Adv. Mater.* **2003**, *15*, 1161–1165. [CrossRef]
- 34. Inagaki, M.; Kaneko, K.; Nishizawa, T. Nanocarbons Recent Research in Japan. Carbon 2020, 42, 1401–1417. [CrossRef]
- 35. Barhate, R.S.; Ramakrishna, S. Review Nanofibrous Filtering Media: Filtration Problems and Solutions from Tiny Materials. J. Membr. Sci. 2007, 296, 1–8. [CrossRef]
- 36. Li, D.; Xia, Y. Electrospinning of Nanofibers: Reinventing the Wheel? Adv. Mater. 2004, 16, 1151–1170. [CrossRef]
- Mohammad, K.P.; Pirjo, H.; Ali, H. Preparation of Carbon Nanotube Embedded in PAN Nanofiber Composites by Electrospinning Process. AUTEX Res. J. 2012, 12, 1–6.
- Ali, A.A. Wet Electrospun Nanofibers. In Proceedings of the Al-Azhar Engineering Eights International Conference, Cairo, Egypt, 24–27 December 2004.
- 39. Ali, A.A.; El-Hamid, M. Electrospinning Optimization for Precursor Carbon Nanofiber. Compos. A 2006, 37, 1681–1687. [CrossRef]
- 40. Ali, A.A. Self-Assembled Ultra-Fine Carbon Coils by with Electrospinning. Mater. Lett. 2006, 60, 2858–2862. [CrossRef]

- 41. Ali, A.A. Presented at Synthesis, Characterization and Industrial Applications of Nanoparticles and Nanostructure Materials Work Shop hosted by MuCSAT and NSF, New Borg El-Arab, Alexandria, Egypt, 11–15 November 2005.
- 42. Ali, A.A.; Rutledge, G.C. Hot-Pressed Electrospun PAN Nanofibers: An Idea for Flexible Carbon Mat. J. Mater. Process. Technol. 2009, 209, 4617–4620. [CrossRef]
- 43. Ali, A.A.; Al-Asmari, A.K. Wet-Electrospun CuNP/Carbon Nano Fiber Composites: Potential Application for Surface Mounted Component. *Appl. Nanosci.* 2012, 2, 55–61. [CrossRef]
- Ali, A.A.; Mansour, A.; Gewefel, E.; Agwa, M. Electrospun EGNPs/PS Ultra-Thin Fibril Composites: An Idea for Luminescence and Thermal Controllable Strong Fabrics. In Proceedings of the International Conference in Nanotechnology, Biotechnology and Spectroscopy (ICNBS), iSeS, Delta Pyramids Hotel, Cairo, Egypt, 29 November–1 December 2013.
- 45. Ali, A.A. A Novel 3-D Graphite Structure from Thermally Stabilized Electrospun MWCNTs/PAN Nanofibril Composite Fabrics. *Int. J. Adv. Manuf. Technol.* 2014, 70, 1731–1738. [CrossRef]
- Ali, A.A.; Eldesouky, R.A.; Zoalfakar, H.S. Hot-Pressed Electrospun MWCNTs/Carbon Nano Fibril Composites: Potential Applications for Breaking Pads and Journal Bearings. In Proceedings of the 16th International AMME Conference, Cairo, Egypt, 27–29 May 2014.
- 47. Ali, A.A.; Eldesouky, R.A.; Zoalfakar, H.S. Mechanical and Tribological Properties of Hot-Pressed Electrospun MWCNTs/Carbon Nanofibril Composite Fabrics. *Int. J. Adv. Manuf. Technol.* **2014**, *74*, 983–993. [CrossRef]
- 48. Ali, A.A.; Eltabey, M.M.; Farouk, W.M.; Zoalfakar, H.S. Electrospun Precursor Carbon Nanofiber Optimization by Using Response Surface Methodology, I WIN EG 29–30 March 2013. *J. Electrost.* **2014**, *72*, 462–469. [CrossRef]
- 49. Megahed, A.A.; Zoalfakar, H.S.; Hassan, A.E.A.; Ali, A.A. A Novel Polystyrene/Epoxy Ultra-Fine Fabric by Electrospinning. *Polym. Adv. Technol.* **2018**, 29, 517–527. [CrossRef]
- 50. Sukigara, S.; Gandhi, M.; Ayutsede, J.; Micklus, M.; Ko, F. Regeneration of Bombyx Mori Silk by Electrospinning, Part 2. Process Optimization and Empirical Modeling Using Response Surface Methodology. *Polymer* **2004**, *45*, 3701. [CrossRef]
- 51. Wächter, R.; Cordery, A. Response Surface Methodology Modeling of Diamond like Carbon Film Deposition. *Carbon* **1999**, *37*, 1529. [CrossRef]
- 52. Hung, C.C.; Lin, H.C.; Shih, H.C. Response Surface Methodology Applied to Silicon Trench Etching in Cl2/HBr/O2 Using Transformer Coupled Plasma Technique. *Solid State Electron.* **2002**, *46*, 791. [CrossRef]
- 53. Rosoa, M.; Lorenzettia, A.; Bescoa, S.; Montib, M.; Berti, G.; Michele, B. Modestia, Application of Empirical Modeling in Multi-layers Membrane Manufacturing. *Comput. Chem. Eng.* **2011**, *35*, 2248–2256. [CrossRef]
- 54. Montgomery, D.C. Design and Analysis of Experiments; Wiley: New York, NY, USA, 2001.
- Ghabrial, S.R.; Ebeid, S.J.; Serag, S.M.; Ayad, M.M. A Mathematical Modeling for Electrochemical Turning. In Proceedings of the PEDAC 5th International Conference, Bangalore, India, 26–29 October 1992; pp. 45–49.
- 56. García-Mateos, F.; Ruiz–Rosas, R.; Rosas, J.M.; Rodríguez-Mirasol, J.; Cordero, T. Controlling the Composition, Morphology, Porosity and Surface Chemistry of Lignin-Based Electrospun Carbon Materials. *Front Mater.* **2019**, *6*, 114. [CrossRef]
- 57. Jeong, D.; Jie, W.; Adelodun, A.; Kim, S.; Jo, Y. Electrospun Melamine-Blended Activated Carbon Nanofibers for Enhanced Control of Indoor CO₂. J. Appl. Polym. Sci. 2019, 136, 47747. [CrossRef]
- 58. Yadav, D.; Amini, F.; Ehrmann, A. Recent Advances in Carbon Nanofibers and Their Applications A Review. *Eur. Polym. J.* **2020**, 138, 109963. [CrossRef]
- Wahyudiono, M.; Machmudah, S.; Kanda, H.; Okubayashi, S.; Goto, M. Formation of PVP Hollow Fibers by Electrospinning in One-Step Process at sub and Supercritical CO₂. *Chem. Eng. Process. Process. Intensif.* 2014, 77, 1–6. [CrossRef]
- 60. Baldino, L.; Cardea, S.; Reverchon, E. Supercritical Assisted Electrospray: An Improved Micronization Process Lucia Baldino, Stefano Cardea and Ernesto Reverchon. *Polymers* **2019**, *11*, 244. [CrossRef] [PubMed]
- 61. Baldino, L.; Cardea, S.; Reverchon, E. A Supercritical CO₂ Assisted Electrohydrodynamic Process Used to Produce Microparticles and Microfibers of a Model Polymer. *J. CO2 Util.* **2019**, *33*, 532–540. [CrossRef]
- 62. Deitzel, J.; Kleinmeyer, J.; Harris, D.; Tan, N.B. The effect of Processing Variables on the Morphology of Electrospun Nanofibers and Textiles. *Polymer* **2001**, *42*, 261–272. [CrossRef]
- 63. Eda, G.; Shivkumar, S. Bead-to-fiber Transition in Electrospun Polystyrene. J. Appl. Polym. Sci. 2007, 106, 475–487. [CrossRef]
- 64. Fong, H.; Chun, I.; Reneker, D. Beaded Nanofibers Formed During Electrospinning. Polymer 1999, 40, 4585–4592. [CrossRef]
- 65. Lee, K.; Kim, H.; Bang, H.; Jung, Y.; Lee, S. The Change of Bead Morphology Formed on Electrospun Polystyrene Fibers. *Polymer* **2003**, *44*, 4029–4034. [CrossRef]
- 66. Yang, Q.; Li, Z.; Hong, Y.; Zhao, Y.; Qiu, S.; Wang, C.; Wei, Y. Influence of Solvents on the Formation of Ultrathin Uniform poly (Vinyl Pyrrolidone) Nanofibers with Electrospinning. J. Polym. Sci. Part B Polym. Phys. 2004, 42, 3721–3726. [CrossRef]
- Koski, A.; Yim, K.; Shivkumar, S. Effect of Molecular Weight on Fibrous PVA Produced by Electrospinning. *Mater. Lett.* 2004, 58, 493–497. [CrossRef]
- Zhao, Y.Y.; Yang, Q.B.; Lu, X.F.; Wang, C.; Wei, Y. Study on Correlation of Morphology of Electrospun Products of Polyacrylamide with Ultrahigh Molecular Weight. J. Polym. Sci. Part B Polym. Phys. 2005, 43, 2190–2195. [CrossRef]
- 69. McKee, M.G.; Layman, J.M.; Cashion, M.P.; Long, T.E.; Enache, D.I.; Edwards, J.K.; Landon, P.; Solsona-Espriu, B.; Carley, A.F.; Herzing, A.A.; et al. Phospholipid Nonwoven Electrospun Membranes. *Sciences* **2006**, *311*, 353–355. [CrossRef] [PubMed]
- Larrondo, L.; St. Manley, J.R. Electrostatic Fiber Spinning from Polymer Melts, Experimental Observations on Fiber Formation and Properties. J. Polym. Sci. Polym. Phys. Ed. 1981, 19, 906–920. [CrossRef]

- 71. Sukigara, S.; Gandhi, M.; Ayutsede, J.; Micklus, M.; Ko, F. Regeneration of Bombyx Mori Silk by Electrospinning—part 1: Processing Parameters and Geometric Properties. *Polymer* **2003**, *44*, 5721–5727. [CrossRef]
- Ding, B.; Kim, H.-Y.; Lee, S.-C.; Shao, C.-L.; Lee, D.-R.; Park, S.-J.; Kwag, G.-B.; Choi, K.-J. Preparation and Characterization of a Nanoscale Poly (Vinyl Alcohol) Fiber Aggregate Produced by an Electrospinning Method. *J. Polym. Sci. Part B Polym. Phys.* 2002, 40, 1261–1268. [CrossRef]
- 73. Ki, C.S.; Baek, D.H.; Gang, K.D.; Lee, K.H.; Um, I.C.; Park, Y.H. Characterization of Gelatin Nanofiber Prepared from Gelatin– Formic Acid Solution. *Polymer* **2005**, *46*, 5094–5102. [CrossRef]
- 74. Kim, K.H.; Jeong, L.; Park, H.N.; Shin, S.Y.; Park, W.H.; Lee, S.C.; Kim, T.I.; Park, Y.J.; Seol, Y.J.; Lee, Y.M.; et al. Biological Efficacy of Silk Fibroin Nanofiber Membranes for Guided Bone Regeneration. J. Biotechnol. 2005, 120, 327–339. [CrossRef] [PubMed]
- Lee, J.S.; Choi, K.H.; Ghim, H.D.; Kim, S.S.; Chun, D.H.; Kim, H.Y.; Lyoo, W.S. Role of Molecular Weight of Atactic Poly (Vinyl Alcohol) (PVA) in the Structure and Properties of PVA Nanofabric Prepared by Electrospinning. *J. Appl. Polym. Sci.* 2004, *93*, 1638–1646. [CrossRef]
- 76. Zhang, Y.; Ouyang, H.; Lim, C.T.; Ramakrishna, S.; Huang, Z.-M. Electrospinning of Gelatin Fibers and Gelatin/PCL Composite Fibrous Scaffolds. *J. Biomed. Mater. Res.* **2004**, *72*, 156–165. [CrossRef]
- 77. Haghi, A.K.; Akbari, M. Trends in electrospinning of natural nanofibers. Phys. Status solidi 2007, 204, 1830–1834. [CrossRef]
- Pham, Q.P.; Sharma, U.; Mikos, A.G. Electrospun Poly(e-caprolactone) Microfiber and Multilayer Nanofiber/Microfiber Scaffolds: Characterization of Scaffolds and Measurement of Cellular Infiltration. *Biomacromolecules* 2006, 7, 2796–2805. [CrossRef] [PubMed]
- 79. Zhang, C.; Yuan, X.; Wu, L.; Han, Y.; Sheng, J. Study on Morphology of Electrospun Poly(Vinyl Alcohol) mats. *Eur. Polym. J.* 2005, 41, 423–432. [CrossRef]
- 80. Zong, X.; Kim, K.; Fang, D.; Ran, S.; Hsiao, B.S.; Chu, B. Structure and Process Relationship of Electrospun Bioabsorbable Nanofiber Membranes. *Polymer* **2002**, *43*, 4403–4412. [CrossRef]
- 81. Huang, C.; Chen, S.; Lai, C.; Reneker, D.H.; Qiu, H.; Ye, Y.; Hou, H. Electrospun Polymer Nanofibres with Small Diameters. *Nanomaterials* **2006**, *17*, 1558–1563. [CrossRef]
- 82. Reneker, D.H.; Chun, I. Nanometre Diameter Fibres of Polymer, Produced by Electrospinning. *Nanotechnology* **1996**, *7*, 216–223. [CrossRef]
- Yuan, X.; Zhang, Y.; Dong, C.; Sheng, J. Morphology of Ultrafine Polysulfone Fibers Prepared by Electrospinning. *Polym. Int.* 2004, 53, 1704–1710. [CrossRef]
- 84. Buchko, C.J.; Chen, L.C.; Shen, Y.; Martin, D.C. Processing and Microstructural Characterization of Porous Biocompatible Protein Polymer thin Films. *Polymer* **1999**, *40*, 7397–7407. [CrossRef]
- 85. Demir, M.; Yilgor, I.; Yilgor, E.; Erman, B. Electrospinning of polyurethane fibers. Polymer 2002, 43, 3303–3309. [CrossRef]
- Yördem, O.S.; Papila, M.; Menceloğlu, Y.Z. Effects of Electrospinning Parameters on Polyacrylonitrile Nanofiber Diameter: An Investigation by Response Surface Methodology. *Mater. Des.* 2008, 29, 34–44. [CrossRef]
- 87. Wang, X.; Um, I.C.; Fang, D.; Okamoto, A.; Hsiao, B.S.; Chu, B. Formation of Water-Resistant Hyaluronic Acid Nanofibers by Blowing-Assisted Electro-Spinning and Non-Toxic Post Treatments. *Polymer* **2005**, *46*, 4853–4867. [CrossRef]
- 88. Sundaray, B.; Subramanian, V.; Natarajan, T.S.; Xiang, R.Z.; Chang, C.C.; Fann, W.S. Electrospinning of Continuous Aligned Polymer Fibers. *Appl. Phys. Lett.* **2004**, *84*, 1222–1224. [CrossRef]
- Li, D.; Wang, Y.; Xia, Y. Electrospinning Nanofibers as Uniaxially Aligned Arrays and Layer-by-Layer Stacked Films. *Adv. Mater.* 2004, 16, 361–366. [CrossRef]
- 90. Xu, C.; Inai, R.; Kotaki, M.; Ramakrishna, S. Aligned Biodegradable Nanofibrous Structure: A Potential Scaffold for Blood Vessel Engineering. *Biomaterials* 2004, 25, 877–886. [CrossRef]
- 91. Ki, C.S.; Kim, J.W.; Hyun, J.H.; Lee, K.H.; Hattori, M.; Rah, D.K.; Park, Y.H. Electrospun three-dimensional silk fibroin nanofibrous scaffold. *J. Appl. Polym. Sci.* 2007, 106, 3922–3928. [CrossRef]
- 92. Mit-Uppatham, C.; Nithitanakul, M.; Supaphol, P. Ultrafine Electrospun Polyamide-6 Fibers: Effect of Solution Conditions on Morphology and Average Fiber Diameter. *Macroml Chem. Phys.* 2004, 205, 2327–2338. [CrossRef]
- 93. Casper, C.L.; Stephens, J.S.; Tassi, N.G.; Chase, D.B.; Rabolt, J.F. Controlling Surface Morphology of Electrospun Polystyrene Fibers: Effect of Humidity and Molecular Weight in the Electrospinning Process. *Macromolecules* **2004**, *37*, 573–578. [CrossRef]
- 94. Abdelhady, S.S.; Zoalfakar, S.H.; Agwa, M.A.; Ali, A.A. Mechanical and Thermal Characteristics of Optimized Electrospun Nylon 6,6 Nanofibers by Using Taguchi Method. *Nanofiber* **2019**, *14*, 1950139. [CrossRef]
- 95. Khang, G. Handbook of Intelligent Scaffolds for Tissue Engineering and Regenerative Medicine; Imprint Jenny Stanford Publishing: New York, NY, USA, 2017.
- Dzobo, K.; Thomford, N.E.; Senthebane, D.A.; Shipanga, H.; Rowe, A.; Dandara, C.; Pillay, M.; Motaung, K.S.C.M. Advances in regenerative medicine and tissue engineering: Innovation and transformation of medicine. *Stems Cells Int.* 2018, 2018, 1–24. [CrossRef] [PubMed]
- 97. Niemczyk-Soczynska, B.; Gradys, A.; Sajkiewicz, P. Hydrophilic Surface Functionalization of Electrospun Nanofibrous Scaffolds in Tissue Engineering. *Polymers* **2020**, *12*, 2636. [CrossRef]
- 98. Li, D.; Tao, L.; Shen, Y.; Sun, B.; Xie, X.; Ke, Q.; Mo, X.; Deng, B. Fabrication of Multilayered Nanofiber Scaffolds with a Highly Aligned Nanofiber Yarn for Anisotropic Tissue Regeneration. *ACS Omega* **2020**, *5*, 24340–24350. [CrossRef]
- 99. Rim, N.G.; Shin, C.S.; Shin, H. Current Approaches to Electrospun Nanofibers for Tissue Engineering. *Biomed. Mater.* **2013**, *8*, 014102. [CrossRef]

- 100. Ma, B.; Xie, J.; Jiang, J.; Shuler, F.D.; Bartlett, D.E. Rational Design of Nanofiber Scaffolds for Orthopedic Tissue Repair and Regeneration. *Nanomedicine* **2013**, *8*, 1459–1481. [CrossRef]
- Xianfeng, W.; Bin, D.; Jianyong, Y.; Moran, W. Engineering biomimetic superhydrophobic surfaces of electrospun nanomaterials. *Nano Today* 2011, 6, 510–530.
- Barajaa, M.A.; Nair, L.S.; Laurencin, C.T. Bioinspired Scaffold Designs for Regenerating Musculoskeletal Tissue Interfaces. *Regen. Eng. Transl. Med.* 2020, 6, 451–483. [CrossRef]
- 103. Vig, K.; Chaudhari, A.; Tripathi, S.; Dixit, S.; Sahu, R.; Pillai, S.; Dennis, V.; Singh, S. Advances in Skin Regeneration Using Tissue Engineering. *Int. J. Mol. Sci.* 2017, *18*, 789. [CrossRef]
- 104. Bacakova, L.; Pajorova, J.; Zikmundova, M.; Filova, E.; Mikes, P.; Jencova, V.; Kuzelova Kostakova, E.; Sinica, A. Nanofibrous Scaffolds for Skin Tissue Engineering and Wound Healing Based on Nature-Derived Polymers. In *Current and Future Aspects of Nanomedicine*; Khalil, I.A.H., Ed.; IntechOpen: London, UK, 2019.
- 105. Travis, J.; Sill, H.; von Recum, A. Electrospinning: Applications in Drug Delivery and Tissue Engineering. *Biomaterials* **2008**, 29, 1989–2006.
- 106. Romána, Z.; Dimitrios, A.L.; István, S. Recent Development of Electrospinning for Drug Delivery. Pharmaceutics 2020, 12, 5.
- Liu, M.; Zhang, Y.; Sun, S.; Abdur, R.K.; Ji, J.; Mingshi, Y.; Guangxi, Z. Recent Advances in Electrospun for Drug Delivery Purpose. J. Drug Targets 2019, 27, 270–282. [CrossRef] [PubMed]
- Pillay, V.; Dott, C.; Choonara, Y.E.; Tyagi, C.; Tomar, L.; Kumar, P.; du Toit Lisa, C.; Ndesendo, V.M.K. A Review of the Effect of Processing Variables on the Fabrication of Electrospun Nanofibers for Drug Delivery Applications. *J. Nanomater.* 2013, 789289, 1–22. [CrossRef]
- 109. Hyuk, S.Y.; Taek, G.K.; Tae, G.P. Surface-Functionalized Electrospun Nanofibers for Tissue Engineering and Drug Delivery. *Adv. Drug Deliv. Rev.* **2009**, *61*, 1033–1042.
- 110. Bishweshwar, P.; Park, M.; Park, S.J. Drug Delivery Applications of Core-Sheath Nanofibers Prepared by Coaxial Electrospinning: A Review. *Pharmaceutics* **2019**, *11*, 305.
- 111. Guarino, V.; Altobelli, R.; Cirillo, V.; Cummaro, A.; Ambrosio, L. Additive Electrospraying: A route to process Electrospun Scaffolds for Controlled Molecular Release. *Polym Adv. Technol.* **2015**, *26*, 1359–1369. [CrossRef]
- 112. Richa, S.; Meenakshi, B. Transdermal Drug Delivery System: A Review. Int. J. Res. Dev. Pharm. Life Sci. 2013, 3, 773–790.
- 113. Rafaela, Z.C.; Meira, I.B.F.; Biscaia, C.; Nogueira, F.S.; Murakami, L.; Bernardi, S.; Paulo, R. Oliveira Solid-State Characterization and Compatibility Studies of Penciclovir, Lysine Hydrochloride, and Pharmaceutical Excipients. *Materials* **2019**, *12*, 3154.
- 114. Karthikeyan, K.; Guhathakarta, S.; Rajaram, R.; Korrapati, P.S. Electrospun Zein/Eudragit Nanofibers Based Dual Drug Delivery System for the Simultaneous Delivery of Aceclofenac and Aantoprazole. *Int. J. Pharm.* **2012**, *438*, 1–2. [CrossRef]
- Modgill, V.; Garg, T.; Goyal, A.K.; Rath, G. Permeability Study of Ciprofloxacin from Ultra-thin Nanofibrous Film through Various Mucosal Membranes. *Artif. Cells Nanomed. Biotechnol* 2016, 44, 1–6. [CrossRef]
- Zhang, Y.; Liu, S.; Wang, X.; Zhang, Z.; Jing, X.; Zhang, P.; Xie, Z.G. Prevention of Local Liver Cancer Recurrence After Surgery Using Multilayered Cisplatin-Loaded Polylactide Electrospun Nanofibers. *Chinese J. Polym. Sci.* 2014, 32, 1111–1118. [CrossRef]
- 117. Singh, B.; Garg, T.; Goyal, A.K.; Rath, G. Development, Optimization, and Characterization of Polymeric Electrospun Nanofiber: A New Attempt in Sublingual Delivery of Nicorandil for the Management of Angina Pectoris. *Artif Cells Nanomed Biotechnol.* 2016, 44, 1498–1507. [CrossRef]
- 118. Canbolat, M.F.; Celebioglu, A.; Uyar, T. Drug Delivery System Based on Cyclodextrin-Naproxen Inclusion Complex Incorporated in Electrospun Polycaprolactone Nanofibers. *Colloids Surf. B Biointerfaces* **2014**, *115*, 15–21. [CrossRef] [PubMed]
- 119. Yu, D.-G.; Shen, X.-X.; Branford-White, C.; White, K.; Zhu, L.-M.; Annie Bligh, S.W. Oral Fast-Dissolving Drug Delivery Mem-branes Prepared from Electrospun Polyvinylpyrrolidone Ultrafine Fibers. *Nanotechnology* **2009**, *20*, 9. [CrossRef]
- Conn, L.; Hastings, E.; Roche, T.; Ruiz-Hernandez, E.; Schenke-Layland, K.; Walsh, C.J.; Duffy, G.P. Drug and Cell Delivery for Cardiac Regeneration. *Adv. Drug Deliv. Rev.* 2015, 84, 85–106.
- 121. Kenyatta, S.W.; Chris, A.B. Delivery of Antioxidant and Anti-inflammatory Agents for Tissue Engineered Vascular Grafts. *Front. Pharm.* **2017**, *8*, 659.
- 122. Shahriar, S.M.S.; Jagannath, M.; Mohammad, N.H.; Vishnu, R.; Lee, D.Y.; Lee, Y.-K. Electrospinning Nanofibers for Therapeutics Delivery. *Nanomaterials* 2019, *9*, 532. [CrossRef] [PubMed]
- Roberts, T.C.; Langer, R.; Wood, M.J.A. Advances in Oligonucleotide Drug Delivery. *Nat. Rev. Drug Discov.* 2020, 19, 673–694.
 [CrossRef] [PubMed]
- 124. Wong, B.S.; Teoh, S.H.; Kang, L. Polycaprolactone Scaffold as Targeted Drug Delivery System and Cell Attachment Scaffold for Postsurgical Care of Limb Salvage. *Drug Deliv. Transl. Res.* 2012, *2*, 272–283. [CrossRef]
- 125. Ji, W.; Sun, Y.; Yang, F.; van den Beucken, J.J.J.P.; Fan, M.; Chen, Z.; Jansen, J.A. Bioactive Electrospun Scaffolds Delivering Growth Factors and Genes for Tissue Engineering Applications. *Pharm. Res.* **2011**, *28*, 1259–1272. [CrossRef]
- 126. Suk, C.J.; Sang, Y.H. Electrospun Nanofibers Surface-modified with Fluorescent Proteins. J. Bioact. Compat. Polym. 2007, 22, 508–524.
- 127. Tıglı, R.S.; Kazaroğlu, N.M.; Mavış, B.; Gümüşderelioğlu, M. Cellular Behavior on Epidermal Growth Factor (EGF)-Immobilized PCL/Gelatin Nanofibrous Scaffolds. *J. Biomater. Sci. Polym. Ed.* **2011**, *22*, 207–223. [CrossRef]
- 128. Tao, W.; Zhu, X.; Yu, X.; Zeng, X.; Xiao, Q.; Zhang, X.; Ji, X.; Wang, X.; Shi, J.; Zhang, H.; et al. Black Phosphorus Nanosheets as a Robust Delivery Platform for Cancer Theragnostic. *Adv. Mater.* **2017**, *29*, 1603276. [CrossRef]

- 129. Gao, H. Progress and Perspectives on Targeting Nanoparticles for Brain Drug Delivery. *Acta Pharm. Sin. B* 2016, *6*, 268–286. [CrossRef] [PubMed]
- Zeng, J.; Xu, X.; Chen, X.; Liang, Q.; Bian, X.; Yang, L.; Jing, X. Biodegradable electrospun fibers for drug delivery. J. Control. Release 2003, 92, 227–231. [CrossRef]
- 131. Zahedi, P.; Rezaeian, I.; Ranaei-Siadat, S.O.; Jafari, S.H.; Supaphol, P. A Review on Wound Dressings with an Emphasis on Electrospun Nanofibrous Polymeric Bandages. *Polym. Adv. Technol.* **2010**, *21*, 77–95. [CrossRef]
- 132. Mohammadian, F.; Eatemadi, A. Drug Loading and Delivery Using Nanofibers Scaffolds. Artificial Cells. *Nanomed. Biotechnol.* **2017**, *45*, 881–888.
- Hu, J.; Wei, J.; Liu, W.; Chen, Y. Preparation and Characterization of Electrospun PLGA/Gelatin Nanofibers as a Drug Delivery System by Emulsion Electrospinning. J. Biomater. Sci. Polym. Ed. 2013, 24, 972–985. [CrossRef] [PubMed]
- 134. Norouzi, M.; Abdali, Z.; Liu, S.; Miller, D.W. Salinomycin-Loaded Nanofibers for Glioblastoma Therapy. *Sci. Rep.* **2018**, *8*, 9377. [CrossRef] [PubMed]
- Iqbal, S.; Rashid, M.H.; Arbab, A.S.; Khan, M. Encapsulation of Anticancer Drugs (5-Fluorouracil and Paclitaxel) into Polycaprolactone (PCL) Nanofibers and in Vitro Testing for Sustained and Targeted Therapy. J. Biomed. Nanotechnol. 2017, 13, 355–366.
 [CrossRef] [PubMed]
- Xu, X.; Yang, L.; Xu, X.; Wang, X.; Chen, X.; Liang, Q.; Zeng, J.; Jing, X. Ultrafine Medicated Fibers Electrospun from W/O Emulsions. J. Control. Release 2005, 108, 33–42. [CrossRef] [PubMed]
- Hassan, A.E.A.; Abou-Elkhair, R.A.I.; Parker, W.B.; Allan, P.W.; Secrist, J.A., III. 6-Methylpurine Derived Sugar Modified Nucleosides: Synthesis and Evaluation of Their Substrate Activity with Purine Nucleoside Phosphorylases. *Bioorg. Chem.* 2016, 65, 9–16. [CrossRef]
- 138. Pawłowska, S.; Rinoldi, C.; Nakielski, P.; Ziai, Y.; Urbanek, O.; Li, X.; Aleksander Kowalewski, T.; Ding, B.; Pierini, P. Ultraviolet Light-Assisted Electrospinning of Core–Shell Fully Cross-Linked P(NIPAAm-co-NIPMAAm) Hydrogel-Based Nanofibers for Hermally Induced Drug Delivery Self-Regulation. *Adv. Mater. Interfaces* 2020, 7, 1–13.
- Nakielski, P.; Pawłowska, S.; Rinoldi, C.; Ziai, Y.; De Sio, L.; Urbanek, O.; Pierini, F. Multifunctional Platform Based on Electrospun Nanofibers and Plasmonic Hydrogel: A Smart Nanostructured Pillow for Near-Infrared Light-Driven Biomedical Applications. ACS Appl. Mater. Interfaces 2020, 12, 54328–54342. [CrossRef]
- 140. Thenmozhi, S.; Dharmaraj, N.; Kadirvelu, K.; Kim, H.Y. Electrospun Nanofibers: New Generation Materials for Advanced Applications. *Mater. Sci. Eng. B* 2017, 217, 36–48. [CrossRef]
- 141. Zhang, P.; Zhao, X.; Zhang, X.; Lai, Y.; Wang, X.; Li, J.; Wei, G.; Su, Z. Electrospun Doping of Carbon Nanotubes and Platinum Nanoparticles into the β-Phase Polyvinylidene Difluoride Nanofibrous Membrane for Biosensor and Catalysis Applications. ACS Appl. Mater. Interfaces 2014, 6, 7563–7571. [CrossRef] [PubMed]
- 142. Masumeh, N. Electrospun Nanofibres in Agriculture and the Food Industry: A Review. J. Sci. Food Agric. 2016, 96, 1–16.
- Castano, L.M.; Flatau, A.B. Smart Fabric Sensors and e-Textile Technologies: A Review. Smart Mater. Struct. 2014, 23, 053001. [CrossRef]
- 144. Mohammadi, A.M.; Hosseini, S.M.; Mohammad, Y. Application of Electrospinning Technique in Development of Intelligent Food Packaging: A short Review of Recent Trends. *Food Sci. Nutr.* **2020**, *8*, 1–10. [CrossRef]
- 145. Krumreich, F.D.; Prietsch, L.P.; Antunes, M.D.; Jansen-Alves, C.; Mendonça, C.R.B.; Borges, C.D.; Zavareze, E.d.R.; Zambiazi, R.C. Avocado Oil Incorporated in Ultrafine Zein Fibers by Electrospinning. *Food Biophys.* **2019**, *14*, 383–392. [CrossRef]
- 146. Shuiliang, C.; Guanghua, H.; Alessandro, A.C.M.; Seema, A.; Greiner, A.; Haoqing, H.; Uwe, S. Electrospun Carbon Fiber Mat with Layered Architecture for Anode in Microbial Fuel Cells. *Electrochem. Commun.* **2011**, *13*, 1026–1029.
- 147. Veluru, J.B.; Manippady, K.K.; Rajendiren, M.; Mya Mya, K.; Rayavarapu, P.R.; Appukuttan, S.N.; Seeram, R. Photocatalytic Hydrogen Generation by Splitting of Water from Electrospun Hybrid Nanostructures. *Int. J. Hydrog. Energy* 2013, *38*, 4324–4333. [CrossRef]
- 148. Zhang, M.; Zhao, X.; Zhang, G.; Wei, G.; Su, Z. Electrospinning Design of Functional Nanostructures for Biosensor Applications. J. Mater. Chem. B 2017, 5, 1699–1711.
- 149. Zhang, P.; Zhao, X.; Ji, Y.; Ouyang, Z.; Wen, X.; Li, J.; Su, Z.; Wei, G. Electrospinning Graphene Quantum Dots into a Nanofibrous Membrane for Dual-Purpose Fluorescent and Electrochemical Biosensors. *J. Mater. Chem. B* 2015, *3*, 2487–2496.
- 150. Subhash, S. Nanofiber Electrodes for Biosensors. In *Handbook of Nanofibers*; Barhoum, A., Bechelany, M., Makhlouf, A., Eds.; Springer: Cham, Switzerland, 2018.
- 151. Shao, P.; Yan, Z.; Chen, H.; Xiao, J. Electrospun Poly (Vinyl Alcohol)/Permutite Fibrous Film Loaded with Cinnamaldehyde for Active Food Packaging. *J. Appl. Polym. Sci.* 2017, 40, 46117. [CrossRef]
- 152. Wang, K.N.; Burugapalli, W.S.J.; Halls, F.; Moussy, A.R.; Zheng, Y. Electrospun Fibro-Porou Spolyurethan Ecoating Sforimplantableglucosebiosensors. *Biomaterials* **2013**, *34*, 888–901. [CrossRef] [PubMed]
- 153. Neethirajan, S.; Ragavan, V.; Weng, X.; Chand, R. Biosensors for Sustainable Food Engineering: Challenges and Perspectives. *Biosensors* 2018, *8*, 23. [CrossRef]
- 154. Mercante, L.A.; Scagion, V.P.; Migliorini, F.L.; Mattoso, L.H.C.; Correa, D.S. Electrospinning-Based (bio)Sensors for Food and Agricultural Applications: A review. *TrAC Trends Anal. Chem.* **2017**, *91*, 91–103. [CrossRef]
- 155. Zhang, D.; Wu, X.; Chen, J.; Lin, K. The Development of Collagen Based Composite Scaffolds for Bone Regeneration. *Bioact. Mater.* **2018**, *3*, 129–138. [CrossRef]

- 156. Grgić, J.; Šelo, G.; Planinić, M.; Tišma, M.; Bucić-Kojić, A. Role of the Encapsulation in Bioavailability of Phenolic Compounds. *Antioxidants* **2020**, *9*, 923. [CrossRef]
- 157. Wen, P.; Wen, Y.; Zong, M.; Linhardt, R.J.; Wu, H. Encapsulation of Bioactive Compound in Electrospun Fibers and its Potential Application. J. Agric. Food Chem. 2017, 65, 9161–9179. [CrossRef]
- Ashraf, A.A. New Generation of Super Absorber Nano-Fibroses Hybrid Fabric by Electro-Spinning. J. Mater. Process. Technol. 2008, 199, 193–198.
- 159. Mohammad, M.; Hasani-Sadrabadi, I.S.; Masoud, S.; Homayoun, M. Novel Nanofiber-Based Triple-Layer Proton Exchange Membranes for Fuel Cell Applications. *J. Power Sources* **2011**, *196*, 4599–4603.
- Wu, J.; Wang, N.; Zhao, Y.; Jiang, L. Electrospinning of Multilevel Structured Functional Micro-/Nanofibers and Their Applications. J. Mater. Chem. A 2013, 1, 7290. [CrossRef]
- Afeesh, R.; Nasser, A.M.; Barakat, S.; Al-Deyab, S.; Yousef, A.; Kim, H.Y. Nematic Shaped Cadmium Sulfide Doped Electrospun Nanofiber Mat: Highly Efficient, Reusable. *Sol. Light Photocatal.* 2012, 409, 21–29.
- 162. Nguyen, T.D.; Kadri, O.E.; Sikavitsas, V.I.; Voronov, R.S. Scaffolds with a High Surface Area-to-Volume Ratio and Cultured Under Fast Flow Perfusion Result in Optimal O2 Delivery to the Cells in Artificial Bone Tissues. *Appl. Sci.* **2019**, *9*, 2381. [CrossRef]
- 163. Darbar, D.; Reddy, M.V.; Sundarrajan, S.; Pattabiraman, R.; Ramakrishna, S.; Chowdari, B.V.R. Anodic Electrochemical Performances of MgCo₂O₄ Synthesized by Oxalate Decomposition Method and Electrospinning Technique for Li-ion Battery Application. *Mater. Res. Bull.* **2016**, *73*, 369–376. [CrossRef]
- 164. Mannarino, M.M. Characterization and Modification of Electrospun Fiber Mats for Use in Composite Proton Exchange Membranes. Ph.D. Thesis, Massachusetts Institute of Technology, Cambridge, MA, USA, June 2013.
- 165. Mousavi, S.; Shahraki, F.; Aliabadi, M.; Haji, A.; Deuber, F.; Adlhart, C. Surface Enriched Nanofiber Mats for Efficient Adsorption of Cr(VI) Inspired by Mature. *J. Environ. Chem. Eng.* **2019**, *7*, 102817. [CrossRef]
- 166. Jian, F.; Tao, N.; Tong, L.; Gai, W. Applications of Electrospun Nanofibers. Chin. Sci. Bull. 2008, 53, 2265–2286.
- 167. Zhao, R.; Lu, X.; Wang, C. Electrospinning Based all-Nano Composite Materials: Recent Achievements and Perspectives. *Compos. Commun.* **2018**, *10*, 140–150. [CrossRef]
- Alvi, M.A.; Akhtar, M.S. An Effective and Low Cost Pd–Ce Bimetallic Decorated Carbon Nanofibers as Electro-Catalyst for Direct Methanol Fuel Cells Applications. J. Alloys Compd. 2016, 684, 524–529. [CrossRef]
- 169. Lurie-Luke, E. Product and Technology Innovation: What Can Biomimicry Inspire? *Biotechnol. Adv.* 2014, 32, 1494–1505. [CrossRef] [PubMed]
- 170. Emily, B.K.; Thomas, A.M. Biomimicry: Streamlining the Front End of Innovation for Environmentally Sustainable Products. *Res. Manag.* **2016**, *59*, 40–48.
- 171. Hayato, I.; Masato, S.; Seiji, A.; Tsutomu, K. Realistic Imitation of Mosquito's Proboscis: Electrochemically Etched Sharp and Jagged Needles and Their Cooperative Inserting Motion. *Sens. Actuators A Phys.* **2011**, *165*, 115–123.
- 172. Wang, X.; Ding, B.; Li, B. Biomimetic Electrospun Nanofibrous Structures for Tissue Engineering. *Mater. Today* **2013**, *16*, 229–241. [CrossRef]
- 173. Ito, Y.; Chen, X.; Kang, I.-K. Advances in Bioinspired and Biomedical Materials. ACS Symp. Ser. 2017, 1253, 153–167. [CrossRef]
- 174. Du, M.; Gu, J.; Wang, J.; Xue, Y.; Ma, Y.; Mo, X.; Xue, S. Silk Fibroin/Poly(L-lactic Acid-co-ε-Caprolactone) Electrospun NanoFibrous Scaffolds Exert a Protective Effect Following Myocardial Infarction. *Exp. Ther. Med.* 2019, 17, 3989–3998. [CrossRef] [PubMed]
- 175. Bhattarai, D.P.; Aguilar, L.E.; Park, C.H.; Kim, C.S. A Review on Properties of Natural and Synthetic Based Electrospun Fibrous Materials for Bone Tissue Engineering. *Membranes* **2018**, *8*, 62. [CrossRef] [PubMed]
- 176. Mousavi, S.M.; Zarei, M.; Hashemi, S.A.; Ramakrishna, S.; Chiang, W.-H.; Lai, C.W.; Gholami, A.; Omidifar, N.; Shokripour, M. Asymmetric Membranes: A Potential Scaffold for Wound Healing Applications. *Symmetry* **2020**, *12*, 1100. [CrossRef]
- 177. Roach, P.; Shirtcliffe, N.; Newton, J.; Michael, I. Progess in Superhydrophobic Surface Development. *Soft Matter* **2008**, *4*, 224. [CrossRef] [PubMed]
- 178. Lin, J.; Wang, X.; Ding, B.; Yu, J.; Sun, G.; Wang, M. Biomimicry via Electrospinning. *Crit. Rev. Solid State Mater. Sci.* 2012, 37, 94–114. [CrossRef]
- 179. Yan, G.; Yu, J.; Qiu, Y.; Yi, X.; Lu, J.; Zhou, X.; Bai, X. Self-Assembly of Electrospun Polymer Nanofibers: A General Phenomenon Generating Honeycomb-Patterned Nanofibrous Structures. *Langmuir* 2011, 27, 4285–4289. [CrossRef] [PubMed]
- 180. Ding, B.; Wang, X.; Yu, J.; Wang, M. Polyamide 6 Composite Nano-Fiber/net Functionalized by Polyethyleneimine on Quartz Crystal Microbalance for Highly Sensitive Formaldehyde Sensors. J. Mater. Chem. 2011, 21, 12784. [CrossRef]
- 181. Andreas, H. Electrospun Polycaprolactone Nanofiber Scaffolds for Tissue Engineering. Ph.D. Thesis, University of Arkansas, Fayetteville, NC, USA, 2012.
- 182. Nagam, H.S.; Rao, S. Multi-Functional Electrospun Nanofibers from Polymer Blends for Scaffold Tissue Engineering. *Fibers* **2019**, *7*, 66.
- Nemati, S.; Kim, S.; Shin, Y.M.; Shin, H. Current Progress in Application of Polymeric Nanofibers to Tissue Engineering. *Nano* Converg. 2019, 6, 36. [CrossRef] [PubMed]
- Yang, Y.; Wang, K.; Gu, X.; Leong, K.W. Biophysical Regulation of Cell Behavior—Cross Talk between Substrate Stiffness and Nanotopography. *Engineering* 2017, 3, 36–54. [CrossRef] [PubMed]

- 185. Pennel, T.; Fercana, G.; Bezuidenhout, D.; Simionescu, A.; Chuang, T.-H.; Zilla, P.; Simionescu, D. The Performance of Cross-linked Acellular Arterial Scaffolds as Vascular Grafts; Pre-Clinical Testing in Direct and Isolation Loop Circulatory Models. *Biomaterials* 2014, 35, 6311–6322. [CrossRef] [PubMed]
- Liu, S.; Qin, S.; He, M.; Zhou, D.; Qin, Q.; Wang, H. Current Applications of Poly(Lactic Acid) Composites in Tissue Engineering and Drug Delivery. *Compos. Part B Eng.* 2020, 199, 108238. [CrossRef]
- Bao, M.; Lou, X.; Zhou, Q.; Dong, W.; Yuan, H.; Zhang, Y. Electrospun Biomimetic Fibrous Scaffold from Shape Memory Polymer of PDLLA-co-TMC for Bone Tissue Engineering. ACS Appl. Mater. Interfaces 2014, 6, 2611–2621. [CrossRef]
- 188. Aragón, J.; Costa, C.; Coelhoso, I.; Mendoza, G.; Aguiar-Ricardo, A.; Irusta, S. Electrospun asymmetric membranes for wound dressing applications. *Mater. Sci. Eng. C* 2019, *103*, 109822. [CrossRef] [PubMed]
- Hajiabbas, M.; Alemzadeh, I.; Vossoughi, M. A Porous Hydrogel-Electrospun Composite Scaffold Made of Oxidized Alginate/Gelatin/Silk Fibroin for Tissue Engineering Application. *Carbohydr. Polym.* 2020, 245, 116465. [CrossRef]
- 190. Hajiabbas, I.; Alemzadeh, M.; Vossoughi, M.; Shamloo, A. In-Situ Crosslinking of Electrospun Gelatin-Carbodiimide Nanofibers: Fabrication, Characterization, and Modeling of Solution Parameter. *Chem. Eng. Commun.* **2020**, *10*, 1–17. [CrossRef]
- 191. Yousefzade, O.; Katsarava, R.; PuiggalÃ, J. Biomimetic Hybrid Systems for Tissue Engineering. Biomimetics 2020, 5, 49. [CrossRef]
- Dalton, P.D.; Woodfield, T.B.F.; Mironov, V.; Groll, J. Advances in Hybrid Fabrication toward Hierarchical Tissue Constructs. *Adv. Sci.* 2020, 7, 1902953. [CrossRef]
- 193. Ng, J.Y.; Obuobi, S.; Chua, M.L.; Zhang, C.; Hong, S.; Kumar, Y.; Gokhale, R.; Ee, P.L.R. Biomimicry of Microbial Polysaccharide Hydrogels for Tissue Engineering and Regenerative Medicine: A Review. *Carbohydr. Polym.* 2020, 241, 116345. [CrossRef] [PubMed]
- 194. An, J.; Teoh, J.; Ee, M.; Suntornnond, R.; Chua, C.K. Design and 3D Printing of Scaffolds and Tissues. *Engineering* **2015**, *1*, 261–268. [CrossRef]
- 195. Chen, F.-M.; Liu, X. Advancing Biomaterials of Human Origin for Tissue Engineering. *Prog. Polym. Sci.* 2016, 53, 86–168. [CrossRef] [PubMed]
- 196. Lutzweiler, G.; Ndreu, H.A.; Engin, V.N. The Overview of Porous, Bioactive Scaffolds as Instructive Biomaterials for Tissue Regeneration and Their Clinical Translation. *Pharmaceutics* **2020**, *7*, 602. [CrossRef]
- 197. Koichi, N. Chapter 1 in vitro biofabrication of Tissues and Organs, Biofabrication Micro- and Nano-Fabrication, Printing. *Patterning Assem.* **2013**, 1–21.
- 198. Nikolova, M.P.; Chavali, M.S. Recent Advances in Biomaterials for 3D Scaffolds: A review. *Bioact. Mater.* 2019, *4*, 271–292. [CrossRef]
- 199. Jan, H.; Dietmar, W.H. Design and Fabrication of Scaffold-Based Tissue Engineering. Bio Nano Mater. 2020, 14. [CrossRef]
- 200. Ameer, P.; Kasoju, R. Strategies to Tune Electrospun Scaffold Porosity for Effective Cell Response in Tissue Engineering. *J. Funct. Biomater.* **2019**, *10*, 30. [CrossRef]
- 201. Abbasi, N.; Ivanovski, S.; Gulati, K.; Love, R.M.; Hamlet, S. Role of Offset and Gradient Architectures of 3-D Melt Electrowritten Scaffold on Differentiation and Mineralization of Osteoblasts. *Biomater. Res.* 2020, 24, 2. [CrossRef]
- Zhang, W.; He, Z.; Han, Y.; Jiang, Q.; Zhan, C.; Zhang, K.; Li, Z.; Zhang, R. Structural Design and Environmental Applications of Electrospun Nanofibers. *Compos. Part A Appl. Sci. Manuf.* 2020, 137, 106009. [CrossRef]
- Liu, H.; Gough, C.R.; Deng, Q.; Gu, Z.; Wang, F.; Hu, X. Recent Advances in Electrospun Sustainable Composites for Biomedical, Environmental, Energy, and Packaging Applications. *Int. J. Mol. Sci.* 2020, 21, 4019. [CrossRef]
- 204. Soroush, S.; Khanian, N.; Choong, T.S.Y.; Rashid, U. Recent Progress in the Design and Synthesis of Nanofibers with Diverse Synthetic Methodologies: Characterization and Potential Applications. *New J. Chem.* **2020**, *44*, 9581.
- 205. Dorati, R.; Chiesa, E.; Pisani, S.; Genta, I.; Modena, T.; Bruni, G.; Brambilla, C.R.M.; Benazzo, M.; Conti, B. The Effect of Process Parameters on Alignment of Tubular Electrospun Nanofibers for Tissue Regeneration Purposes. J. Drug Deliv. Sci. Technol. 2020, 58, 101781. [CrossRef]
- 206. Xue, J.; Wu, T.; Dai, Y.; Xia, Y. Electrospinning and Electrospun Nanofibers: Methods, Materials, and Applications. *Chem. Rev.* 2019, 119, 5298–5415. [CrossRef] [PubMed]
- 207. Haider, A.; Haider, S.; Kang, I.-K. Comprehensive Review Summarizing Effect of Electrospinning Parameters and Potential Applications of Nanofibers in Biomedical and Biotechnology. *Arab. J. Chem.* **2015**, *11*, 1165–1188. [CrossRef]
- Datta, P.; Dhara, S. Engineering Porosity in Electrospun Nanofiber Sheets by Laser Engraving: A Strategy to Fabricate 3D Scaffolds for Bone Graft Applications. J. Indian. Inst. Sci. 2019, 99, 329–337. [CrossRef]
- Nestor, W.S.P.; Anton, S.; Nikita, P.; Anton, S.; Pavel, P. Direct Ink Writing Technology (3D Printing) of Graphene-Based Ceramic Nanocomposites: A Review. Nanomaterials 2020, 10, 1300.
- Sooriyaarachchi, D.; Minière, H.J.; Maharubin, S.; Tan, G.Z. Hybrid Additive Microfabrication Scaffold Incorporated with Highly Aligned Nanofibers for Musculoskeletal Tissues. *Tissue Eng. Regen. Med.* 2019, 16, 29–38. [CrossRef]
- 211. Rabionet, M.; Polonio, E.; Guerra, A.; Martin, J.; Puig, T.; Ciurana, J. Design of a Scaffold Parameter Selection System with Additive Manufacturing for a Biomedical Cell Culture. *Materials* **2018**, *11*, 1427. [CrossRef]
- 212. Wu, C.S.; Wu, D.Y.; Wang, S.S. Bio-Based Polymer Nanofiber with Siliceous Sponge Spicules Prepared by Electrospinning: Preparation, Characterisation, and Functionalisation. *Mater. Sci. Eng. C* 2020, *108*, 110506. [CrossRef]
- 213. Rongthong, W.; Niamnont, N.; Srisuwannaket, C.; Paradee, N.; Mingvanish, W. Electrospun Gelatin Fiber Mats Mixed With C.carandas Extract and its Enhanced Stability and Bioactivity. *J. Pharm. Sci.* **2021**, in press.

- Gupta, D.; Jassal, M.; Agrawal, A.K. Solution Properties and Electrospinning of Poly(Galacturonic Acid) Nanofibers. *Carbohydr. Polym.* 2019, 212, 102–111. [CrossRef]
- Reksamunandar, R.P.; Edikresnha, D.; Munir, M.M.; Damayanti, S. Encapsulation of β-Carotene in Poly(Vinylpyrrolidone) (PVP) by Electrospinning Technique. *Procedia Eng.* 2017, 170, 19–23. [CrossRef]
- 216. Eskitoros, T.Ş.M.; Bulbul, Y.E.; Dilsiz, N. Combination of Nano-Hydroxyapatite and Curcumin in a Biopolymer Blend Matrix: Characteristics and Drug Release Performance of Fibrous Composite Material Systems. *Int. J. Pharm.* 2020, 590, 119933. [CrossRef] [PubMed]
- 217. Qin, Z.; Jia, X.W.; Liu, Q.; Kong, B.; Wang, H. Fast Dissolving Oral Films for Drug Delivery Prepared from Chitosan/Pullulan Electrospinning Nanofibers. *Int. J. Biol. Macromol.* **2019**, *137*, 224–231. [CrossRef]
- 218. Baek, J.; Lee, E.; Lotz, M.K.; D'Lima, D.D. Bioactive Proteins Delivery through Core-Shell Nanofibers for Meniscal Tissue Regeneration. *Nanomed. Nanotechnol. Biol. Med.* 2020, 23, 102090. [CrossRef] [PubMed]
- Oliveira, P.E.; Petit-Breuilh, X.; Díaz, P.E.; Gacitúa, W. Manufacture of a Bio-Tissue Based on Nanocrystalline Cellulose from Chilean Bamboo Chusquea Quila and a Polymer Matrix Using Electrospinning. *Nano Struct. Nano Objects* 2020, 23, 100525. [CrossRef]
- 220. Singhal, P. Preparation and Characterization of Poly (E-CAPROLACTONE) Nano Fibers by Electrospinning Technique for Tissue Engineering Applications. *Mater. Today Proc.* 2021, *37*, 2997–3001. [CrossRef]
- 221. Rabionet, M.; Puig, T.; Ciurana, J. Manufacture of PCL Scaffolds through Electrospinning Technology to Accommodate Triple Negative Breast Cancer Cells Culture. *Procedia CIRP* 2020, *89*, 98–103. [CrossRef]
- 222. Sedghi, R.; Shaabani, A.; Mohammadi, Z.; Samadi, F.Y.; Isaei, E. Biocompatible Electrospinning Chitosan Nanofibers: A Novel Delivery System with Superior Local Cancer Therapy. *Carbohydr. Polym.* **2017**, *159*, 1–10. [CrossRef] [PubMed]
- 223. Asadi, H.; Ghaee, A.; Nourmohammadi, J.; Mashak, A. Electrospun Zein/Graphene Oxide Nanosheet Composite Nanofibers with Controlled Drug Release as Antibacterial Wound Dressing. Int. J. Polym. Mater. Polym. Biomater. 2020, 69, 173–185. [CrossRef]
- 224. Simionescu, B.C.; Drobota, M.; Timpu, D.; Vasiliu, T.; Constantinescu, C.A.; Rebleanu, D.; Calin, M.; David, G. Biopolymers/poly(ε-caprolactone)/Polyethylenimine Functionalized Nano-Hydroxyapatite Hybrid Cryogel: Synthesis, Characterization and Application in gene Delivery. *Mater. Sci. Eng. C* 2017, *81*, 167–176. [CrossRef] [PubMed]
- Zhang, S.; Liu, H.; Tang, N.; Zhou, S.; Yu, J.; Ding, B. Spider-Web-Inspired PM0.3 Filters Based on Self-Sustained Electrostatic Nanostructured Networks. Adv. Mater. 2020, 32, 2002361. [CrossRef]
- 226. Omori, Y.; Gu, T.; Bao, L.; Otani, Y.; Seto, T. Performance of Nanofiber/Microfiber Hybrid Air Filter Prepared by Wet Paper Processing. *Aerosol. Sci. Technol.* 2019, *53*, 1149–1157. [CrossRef]
- 227. Tabibzadeh, A.; Esghaei, M.; Soltani, S.; Yousefi, P.; Taherizadeh, M.; Safarnezhad, T.F.; Golahdooz, M.; Panahi, M.; Ajdarkosh, H.; Zamani, F.; et al. Evolutionary Study of COVID-19, Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) as an Emerging Coronavirus: Phylogenetic Analysis and Literature Review. *Vet. Med. Sci.* 2020, 00, 1–13. [CrossRef] [PubMed]
- 228. Chen, J.; Huang, C.; Zhang, Y.; Zhang, S.; Jin, M. Severe Acute Respiratory Syndrome Coronavirus 2-Specific Antibodies in Pets in Wuhan, China. J. Infect. 2020, 81, 68–69. [CrossRef]
- 229. Karunathilake, K. Positive and Negative Impacts of COVID-19, an Analysis with Special Reference to Challenges on the Supply Chain in South Asian countries. *J. Soc. Econ. Dev.* **2020**. [CrossRef]
- Sivaraman, D.; Pradeep, P.S.; Manoharan, S.S.; Bhat, C.R.; Leela, K.V.; Venugopal, V. Current Strategies and Approaches in Combating SARS-CoV-2 Virus that Causes COVID-19. *Lett. Drug Des. Discov.* 2020, 17, 670–672. [CrossRef]
- 231. Kchaou, M.; Abuhasel, K.; Khadr, M.; Hosni, F.; Alquraish, M. Surface Disinfection to Protect against Microorganisms: Overview of Traditional Methods and Issues of Emergent Nanotechnologies. *Appl. Sci.* 2020, *10*, 6040. [CrossRef]
- Global Research on Coronavirus Disease (COVID-19). Available online: https://www.who.int/emergencies/diseases/novelcoronavirus-2019/global-research-on-novel-coronavirus-2019-ncov (accessed on 16 December 2020).
- 233. Outlook on the Worldwide Market for Nanofibers to 2030—Initiatives Using Nanofibers to Aid the Response to COVID-19. Available online: https://www.globenewswire.com/news-release/2020/04/06/2011882/0/en/Outlook-on-the-Worldwide-Market-for-Nanofibers-to-2030-Initiatives-Using-Nanofibers-to-Aid-the-Response-to-COVID-19.html (accessed on 17 December 2020).
- 234. Tebyetekerwa, M.; Xu, Z.; Yang, S.; Ramakrishna, S. Electrospun Nanofibers-Based Face Masks. *Adv. Fiber Mater.* **2020**, *2*, 161–166. [CrossRef]
- 235. Liu, Z.; Ramakrishna, S.; Liu, X. Electrospinning and Emerging Healthcare and Medicine Possibilities. *APL Bioeng.* **2020**, *4*, 030901. [CrossRef]
- 236. De Sio, L.; Ding, B.; Focsan, M.; Kogermann, K.; Pascoal-Faria, P.; Petronella, F.; Pierini, F. Personalized Reusable Face Masks with Smart Nano-Assisted Destruction of Pathogens for COVID-19: A Visionary Road. *Chem. Eur. J.* **2021**, *27*, 1–20.
- 237. Zhang, Y.; Wu, L.; Wang, X.; Yu, J.; Ding, B. Super Hygroscopic Nanofibrous Membrane-Based Moisture Pump for Solar-Driven Indoor Dehumidification. *Nat. Commun.* **2020**, *11*, 3302. [CrossRef]
- 238. Recyclable Nano-Fiber Filtered Face Masks a Boon for Supply Fiasco. Available online: https://news.kaist.ac.kr/newsen/ html/news/?mode=V&mng_no=6530&skey=category&sval=research&list_s_date=&list_e_date=&GotoPage=1 (accessed on 3 January 2021).
- Khanzada, H.; Salam, A.; Qadir, M.B.; Phan, D.-N.; Hassan, T.; Munir, M.U.; Pasha, K.; Hassan, N.; Khan, M.Q.; Kim, I.S. Fabrication of Promising Antimicrobial Aloe Vera/PVA Electrospun Nanofibers for Protective Clothing. *Materials* 2020, 13, 3884. [CrossRef] [PubMed]

- 240. Castillo-Henríquez, L.; Brenes-Acuña, M.; Castro-Rojas, A.; Cordero-Salmerón, R.; Lopretti-Correa, M.; Vega-Baudrit, J.R. Biosensors for the Detection of Bacterial and Viral Clinical Pathogens. *Sensors* **2020**, *20*, 6926. [CrossRef] [PubMed]
- 241. Ding, Y.; Dou, C.; Chang, S.; Xie, Z.; Yu, D.G.; Liu, Y.; Shao, J. Core-Shell Eudragit S100 Nanofibers Prepared via Triaxial Electrospinning to Provide a Colon-Targeted Extended Drug Release. *Polymers* **2020**, *12*, 2034. [CrossRef] [PubMed]