

Supplementary materials

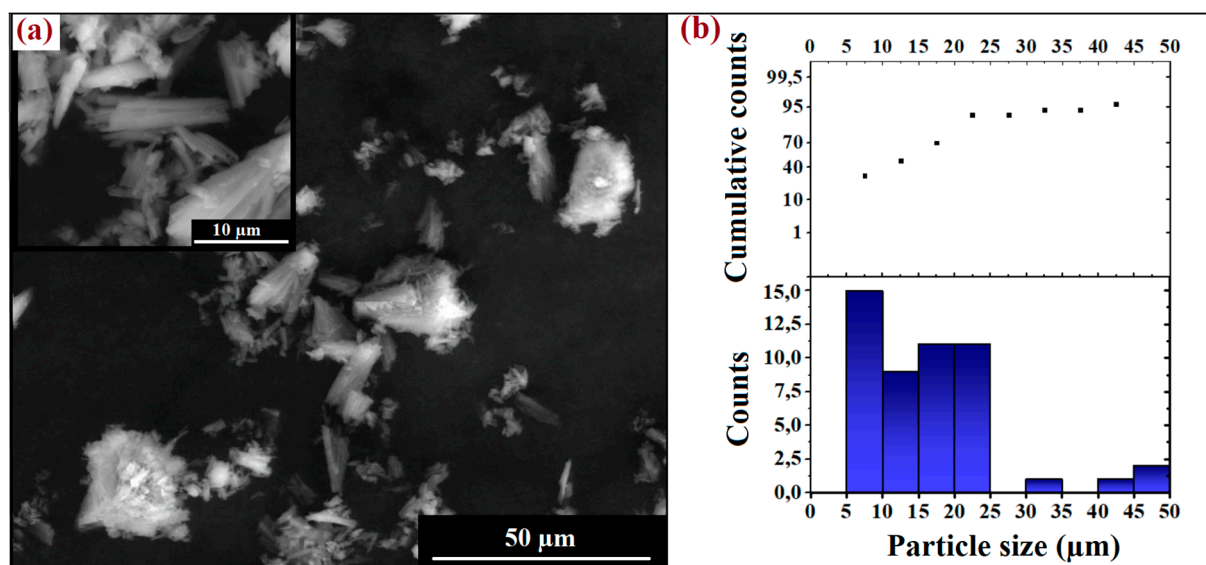


Figure S1. Nystatin powder: (a) Scanning electron microscopy images for nystatin powder and (b) particle size distribution histogram (particle sizes were determined using NIH Image J software).

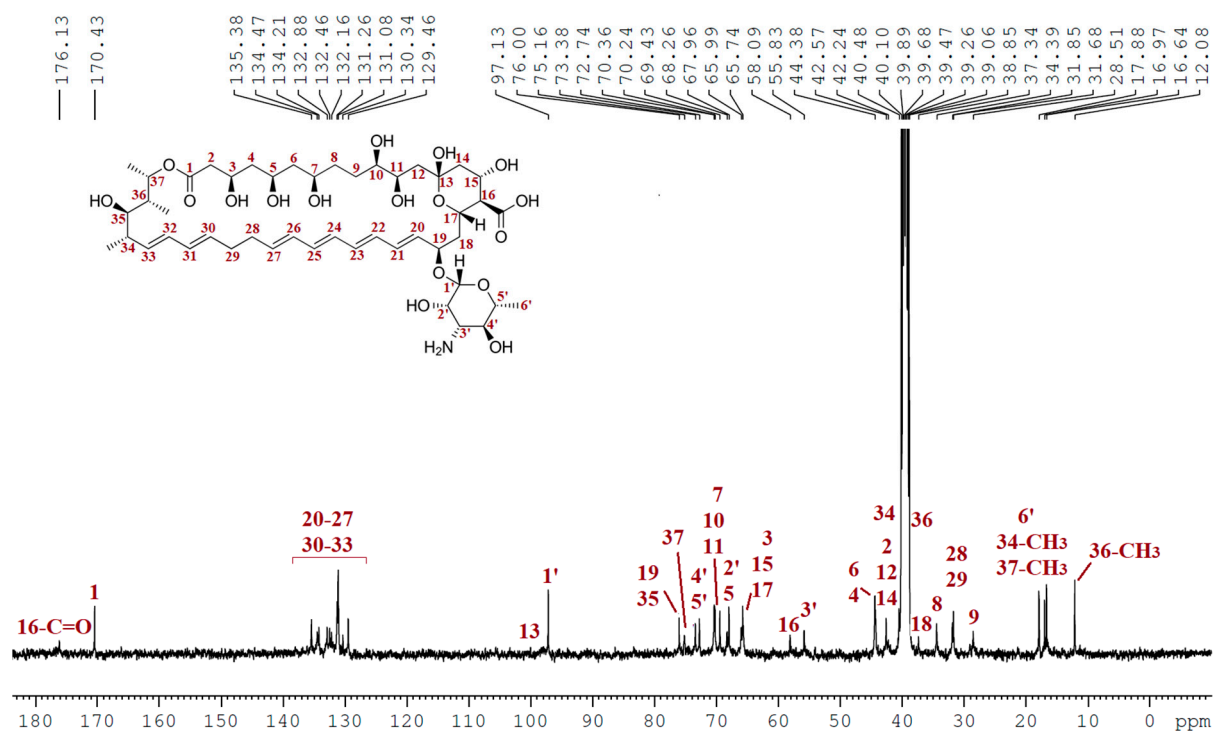


Figure S2. ^{13}C -RMN spectrum of nystatin; IUPAC Name: (1S,3R,4R,7R,9R,11R,15S,16R,17R,18S,19E, 21E,25E, 27E, 29E,31E,33R,35S,36R,37S)-33-[(2R,3S,4S,5S,6R)-4-amino-3,5-dihydroxy-6-methyloxan-2-yl]oxy-1,3,4,7,9,11,17,37-octahydroxy-15,16,18-trimethyl-13-oxo-14,39-dioxabicyclo[33.3.1]nonatriaconta-19,21,25,27,29,31-hexaene-36-carboxylic acid); signal attributions made according to ref. [44-46].

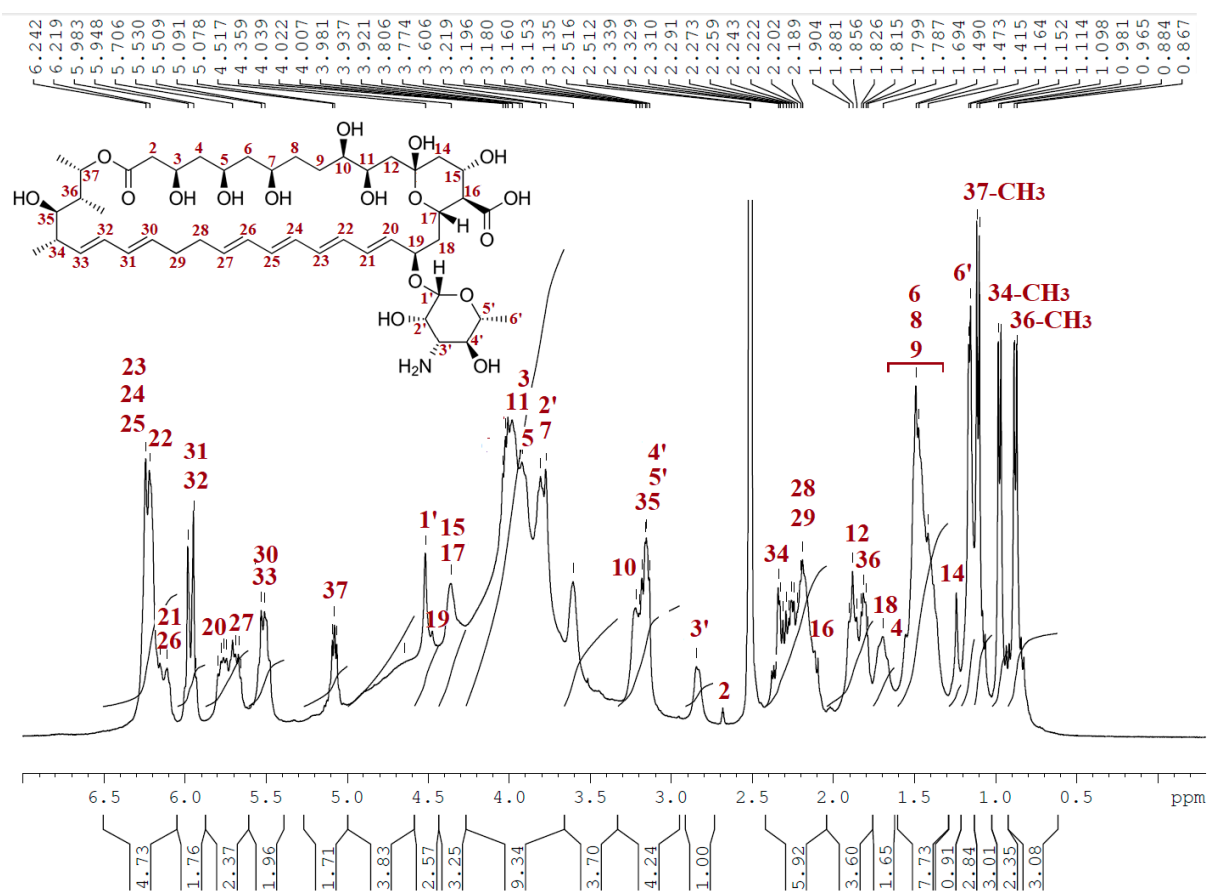


Figure S3. ¹H-RMN spectrum of nystatin. IUPAC Name: (1S,3R,4R,7R,9R,11R,15S,16R,17R,18S,19E, 21E,25E, 27E,29E,31E,33R,35S,36R,37S)-33-[(2R,3S,4S,5S,6R)-4-amino-3,5-dihydroxy-6-methyloxan-2-yl]oxy-1,3,4,7,9,11,17,37-octahydroxy-15,16,18-trimethyl-13-oxo-14,39-dioxabicyclo[33.3.1]nonatriaconta-19,21,25,27,29,31-hexaene-36-carboxylic acid); signal attributions made according to ref. [44-46].

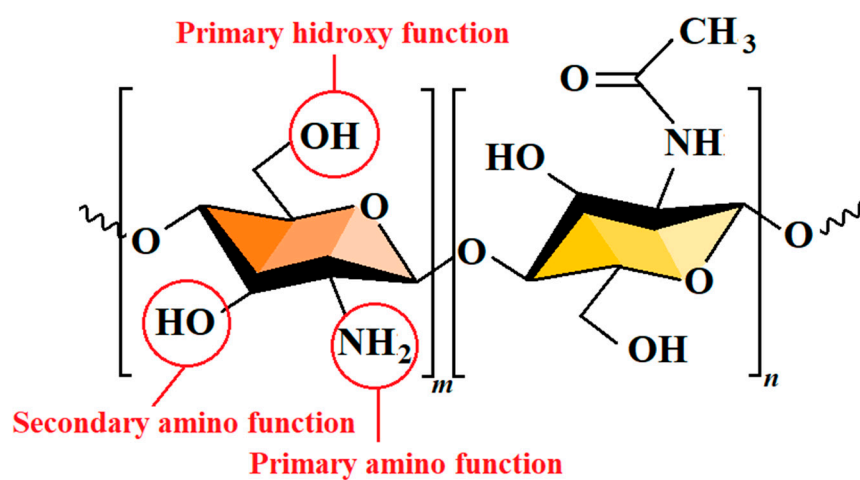


Figure S4. Chemical formula of chitosan with highlighting the functional groups: free primary amino groups (NH₂ to C2), primary hydroxyl group (OH to C6) and secondary hydroxyl groups (OH to C3); *m* and *n* represent the numbers of deacetylated (d-glucosamine) and N-acetyl-d-glucosamine repeating units, linked by (1-4)-β-glycosidic linkages, respectively.

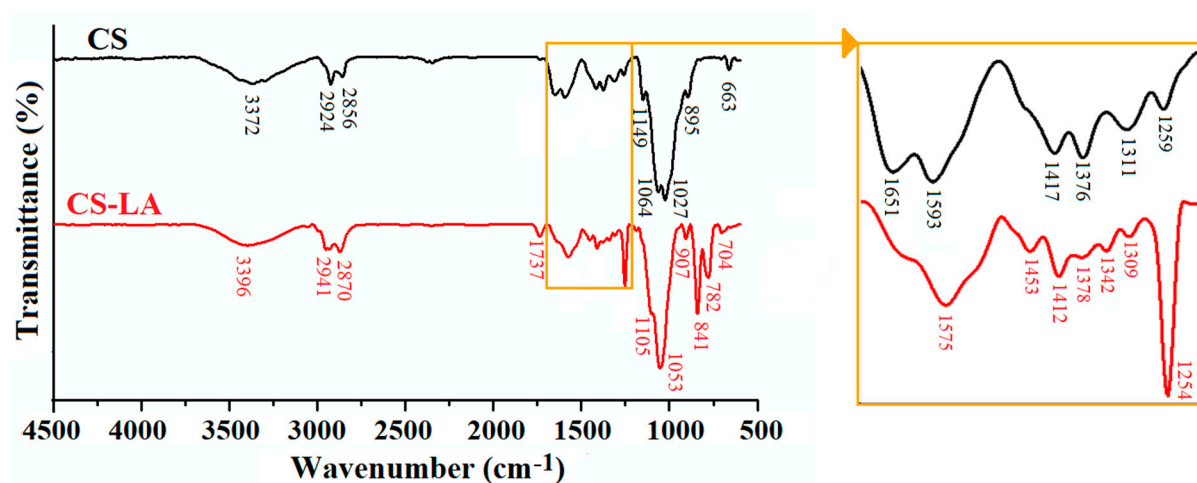


Figure S5. FTIR spectra of chitosan powder (CS) and chitosan gel (3%) after dispersion in 2% lactic acid solution (CS-LA).

The spectrum of chitosan shows polysaccharide characteristic absorption bands found in the range 4000–600 cm⁻¹: the broad band located between 3500–3100 cm⁻¹ results from the stretching vibrations of superimposed O–H and N–H bonds; the absorption bands corresponding to the asymmetric and symmetrical stretching vibrations of the C–H are observed at 2924 cm⁻¹, and 2856 cm⁻¹ respectively; the stretching vibration of the C=O bond of amide I (O=C–NHR) is distinguished at 1651 cm⁻¹; the band at 1593 cm⁻¹ is caused by the in-plane deformation vibration of the primary amine group NH₂ coupled with the stretching vibrations of amide II, and that at 1311 cm⁻¹ by the tensile vibrations of the C–N bond (amide III); the absorption bands given by the deformation vibrations of the CH₂ and CH₃ group appear at 1417 cm⁻¹, respectively 1376 cm⁻¹; the absorption band from 1149 cm⁻¹ is attributed to the asymmetric stretching vibrations of the C–O–C glycosidic bonds, and the bands from 1064 and 1027 cm⁻¹ are due to the C–O stretching vibrations, characteristic of the saccharide structure [19].

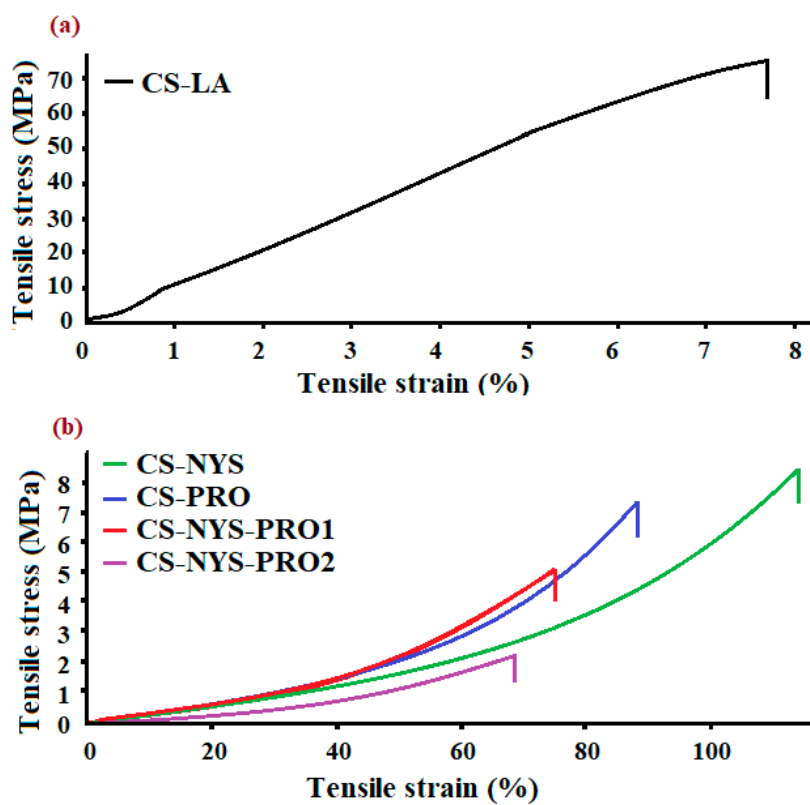


Figure S6. Characteristic stress-strain curves of the films: (a) without biological active compounds (CS-LA); (b) nystatin, propolis and nystatin/propolis loaded chitosan films (CS-NYS, CS-PRO, CS-NYS-PRO1, CS-NYS-PRO2).

Table S1. Parameters of pseudo-second order (PSO) and Korsmeyer-Peppas (K-P) kinetic models, where k_s is the constant of the swelling rate, S_e is the theoretical swelling capacity at equilibrium, k_p is a constant dependent on the polymeric network; n is the diffusion parameter of aqueous PBS in the formulation film and SD represents the standard deviation of $n=11$); swelling capacity in PBS of chitosan charged films after 5 and 24 h.

Film code	PSO model parameters	K-P model parameters	Swelling capacity after 5 h (%)	Swelling capacity after 24 h (%)
CS-NYS	$k_s = 1.86 \cdot 10^{-3}$	$k_p = 450.17$	618	751
	$S_e = 600.74$	$n = 5.9 \cdot 10^{-2}$		
	SD = 6.10	SD = 8.03		
CS-PRO	$k_s = 5.29 \cdot 10^2$	$k_p = 121.14$	122	122
	$S_e = 121.92$	$n = 4.87 \cdot 10^{-18}$		
	SD = 0.37	SD = 0.44		
CS-NYS-PRO1	$k_s = 1.21 \cdot 10^{-1}$	$k_p = 129.14$	129	129
	$S_e = 130.84$	$n = 1.40 \cdot 10^{-3}$		
	SD = 0.56	SD = 0.71		
CS-NYS-PRO2	$k_s = 6.54 \cdot 10^3$	$k_p = 117.93$	118	118
	$S_e = 119.35$	$n = 5.42 \cdot 10^{-18}$		
	SD = 0.56	SD = 0.71		

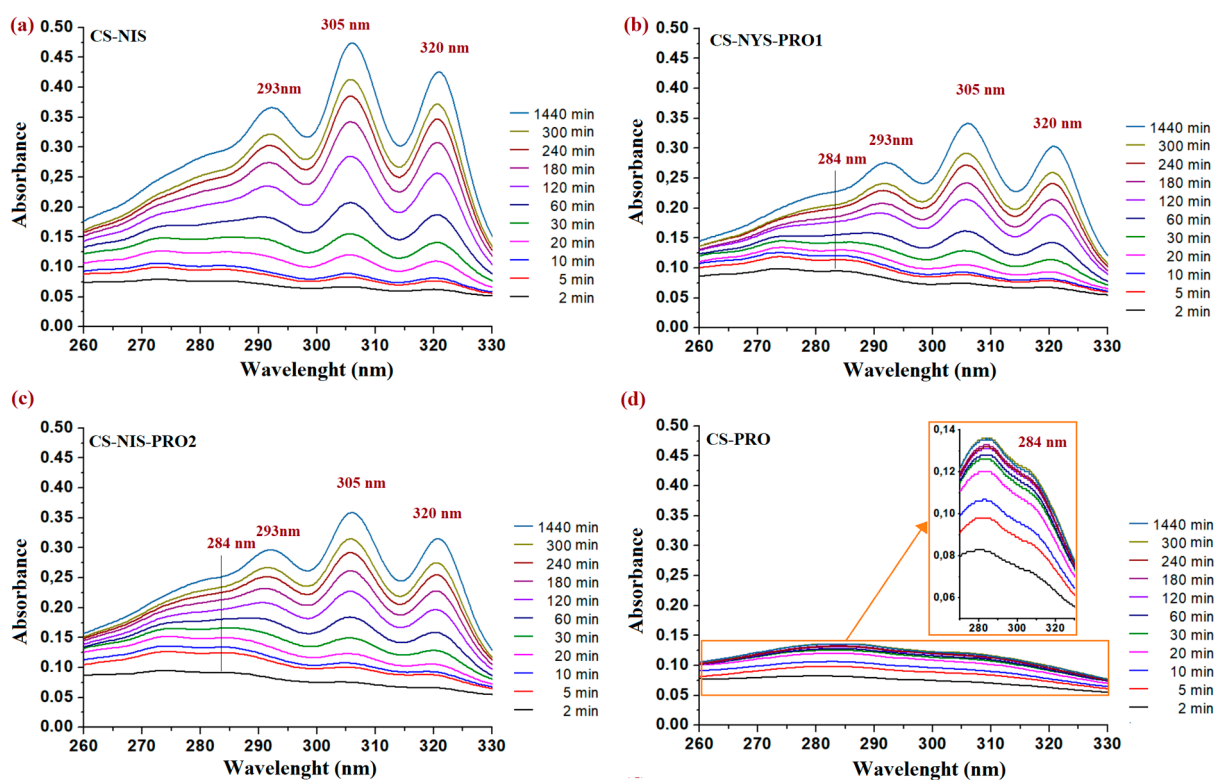


Figure S7. Evolution in time of UV-VIS spectra of: nystatin released from CS-NYS (a), CS-NYS-PRO1 (b) and CS-NYS-PRO2 (c); propolis released from CS-PRO (d).

Table S2. NYS and PRO release from chitosan films: Korsmeyer-Peppas kinetics parameters (k represents the transport constant, n is the diffusion exponent and SD is standard deviation of $n=11$), and release efficiency after 5 and 24 hours.

Film code	Drug released	Korsmeyer-Peppas model parameters	Drug release efficiency after 5 hours (%)	Drug release efficiency after 1 day (%)
CS-NYS	NYS	$n = 0.4189821$ $k = 0.045$ SD = 0.37	48.6	55.2
CS-NYS-PRO1	NYS	$n = 0.3209252$ $k = 0.072$ SD = 0.50	45.9	53.3
CS-NYS-PRO2	NYS	$n = 0.3098546$ $k = 0.071$ SD = 0.34	42.8	48.7
CS-PRO	PRO	$n = 0.0806324$ $k = 0.157$ SD = 0.35	24.2	24.2

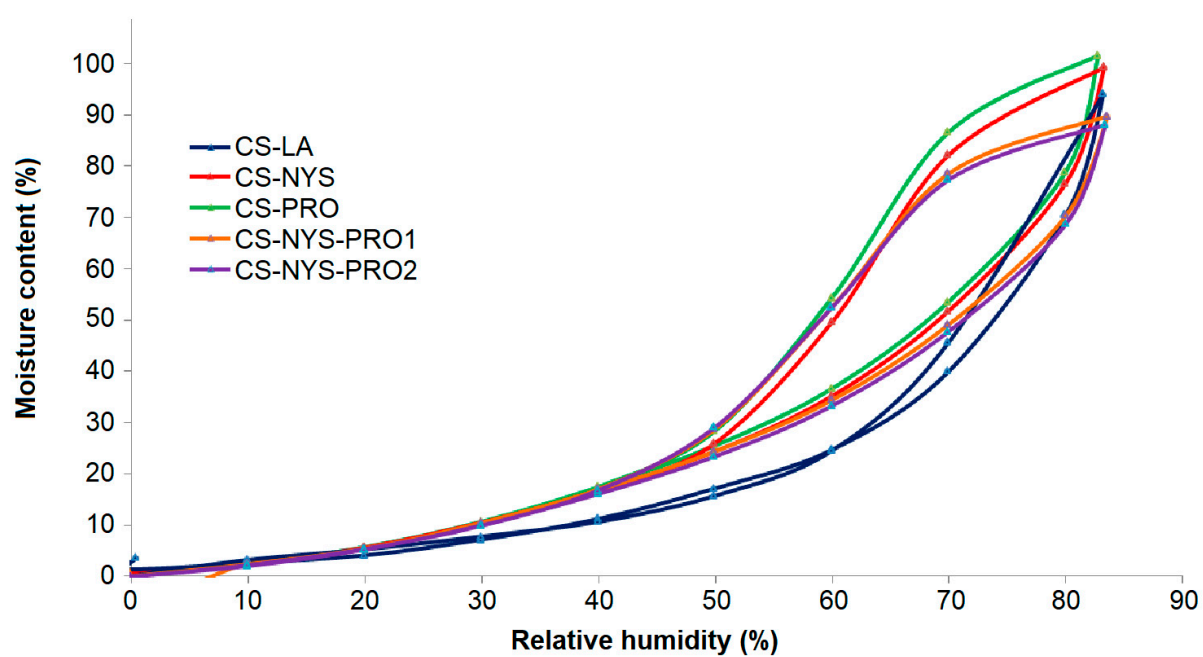


Figure S8. Sorption/desorption isotherms for the hydrogel formulations

Table S3. Surface parameters of the hydrogels evaluated based on adsorption/desorption isotherms: water vapor sorption capacity, final weight (W); average pore size (r_{pm}) and BET data (surface area and monolayer weight).

Hydrogel Surface	W (%)	r_{pm} (BJH model) (nm)	BET data *	
			Area (m ² /g)	Monolayer (mg/g)
CS-LA	94.1	6.8	275	788
CS-NYS	99.3	1.4	1379	399
CS-PRO	101.6	2.0	1035	294
CS-NYS-PRO1	89.8	1.2	1444	412
CS-NYS-PRO2	88.1	1.0	1727	492

Determined based on desorption branch of the isotherm (registered up to a relative humidity of 40 %)

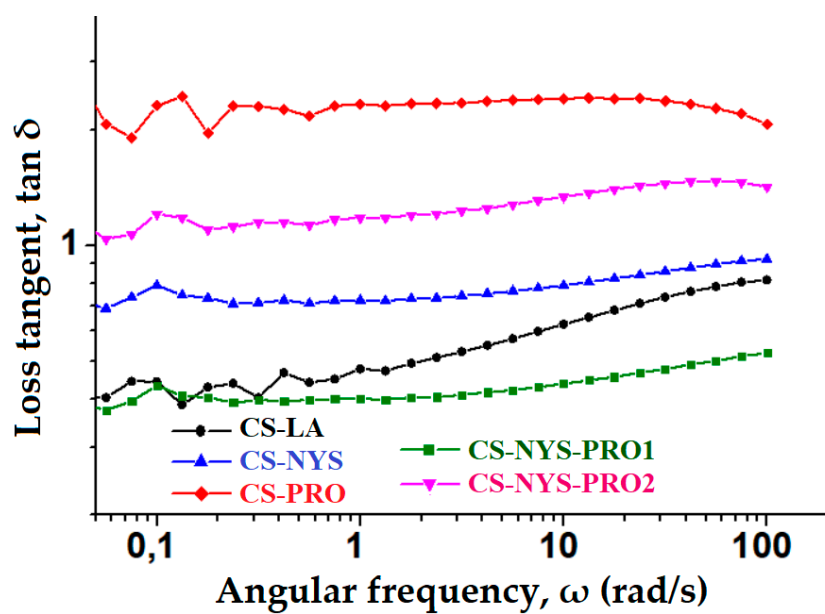


Figure S9. The loss tangent ($\tan \delta = G'' / G'$) as a function of angular frequency.

Table S4. Viscoelastic parameters (G' , G'' and $\tan \delta$) at a frequency of 1 Hz and viscous flow activation energy (E_η) calculated by fitting the temperature-dependence curve of the viscosity by Arrhenius equation.

Gel formulation	G' (Pa)	G'' (Pa)	$\tan \delta$	E_η (kcal/mol)
CS-LA	42.64	20.30	0.47	5.18
CS-NYS	20.70	14.70	0.73	5.51
CS-PRO	0.94	2.22	2.31	5.67
CS-NYS-PRO1	107.05	43.96	0.40	4.81
CS-NYS-PRO2	4.43	5.45	1.18	5.28

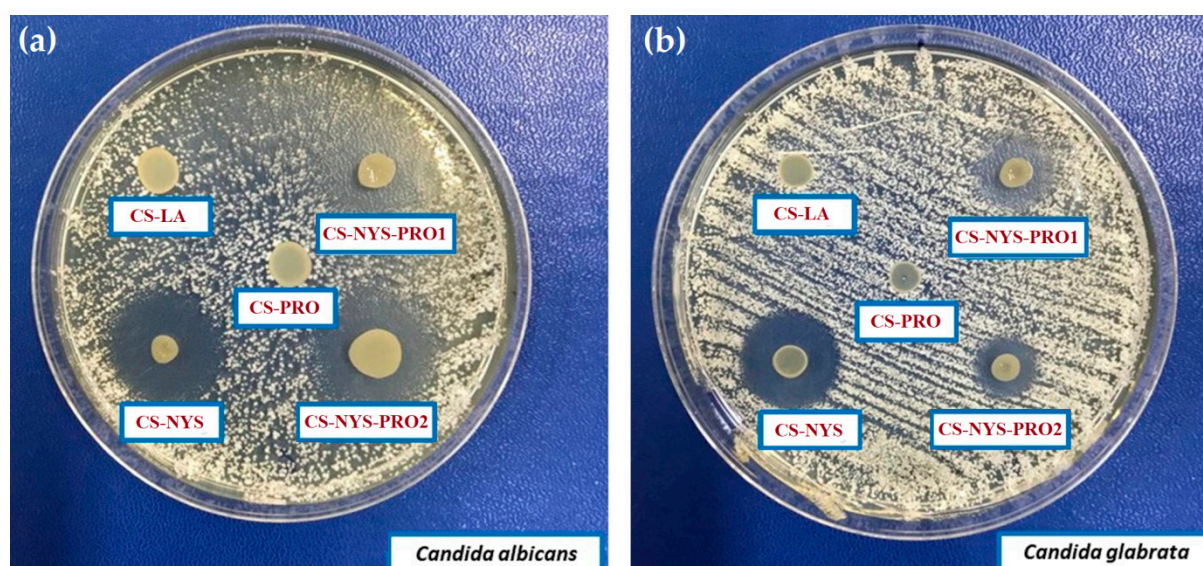


Figure S10. Antifungal activity of chitosan hydrogels against: (a) *C. albicans* and (b) *C. glabrata*

Table S5. Killing efficiency as a function of time for CS-NYS, CS-PRO and CS-NYS-PRO1 hydrogels, against *Candida albicans* in 24 h, as compared to the control test.

Time (hours)	<i>Candida albicans</i> (CFU/mL)			
	Control	CS-NYS	CS-PRO	CS-NYS-PRO1
0	1.24x10 ⁷	1.24x10 ⁷	1.24x10 ⁷	1.24x10 ⁷
6	1.64x10 ⁷	2.40x10 ²	7.60x10 ⁶	<10 ¹
12	1.96x10 ⁷	<10 ¹	3.00x10 ⁶	<10 ¹
24	4.32x10 ⁷	<10 ¹	2.28x10 ⁶	<10 ¹