Table S1. Fluorochrome panel of monoclonal antibodies used to immunophenotype moDCs.

Laser	488 nm				640 nm	
Emission filters	525/40	585/42	525/40	585/42	665/20	660/20
Fluorochrome	FITC	PE	FITC	PE	APC	Alexa Fluor® 647
Relative Brightness	3	5	3	5	4	4
Biomarker	CD14	CD83	MHC II	HLA ABC	CD11c	CD1a
Monoclonal antibody	Mouse anti-Human CD14	Mouse anti-Human CD83	Rat anti-Dog MHC Class II	Mouse anti-Human HLA ABC	Mouse anti-Human CD11c	Mouse anti-Human CD1a
Clone	TuK4	HB15e	YKIX334.2	W6/32	BU15	NA1/34-HLK
Company	ThermoFisher Scientific	Invitrogen	Bio-Rad	Bio-Rad	ThermoFisher Scientific	Bio-Rad

Table S2. List of forward (FW) and reverse (RV) primers, base-pair number (bp) of amplified fragments, and primer annealing temperature (TAN) for each gene studied. * indicates the primers designed with Primer3 software.

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	Primer	Reference	Fragment size (bp)	TA (ºC)
β-Actin	FW-5'ACGGAGCGTGGCTACAGC3' RW-5'TCCTTGATGTCACGCACGA3'	Sauter et al., 2005	61	60.5
TLR2	FW-5'AATCCCCCGTTCAAGTTGTG3' RW-5'ATGGTTTTGCGGCTCTTCTC 3'	Ishii et al.,2006	101	61
TLR4	FW-5'AGAGGATTTCCCCATTGGAC3' RW-5'ACGCAGGTAGCTTGAAGGAA3'	*	86	56
TLR9	FW-5'ACCACATCATCACCCTGGCACCT3' RW-5' CGGCGACAGTTCCCACCCAC 3'	Rodrigues et al., 2017	82	64
NOD1	FW-5' CCTTGGCTGTCGGAGATTGGCT 3' RW-5' ACCTGCTTACTGGGTCGGTGT 3'	Rodrigues et al., 2017	82	61
NOD2	FW-5' TGGCGTGGGAGCAGGGTTTC 3' RW-5' CGCTGGGAGGATGTGAAGATGG 3'	Rodrigues et al., 2017	76	66
NLRP10	FW-5'CCATGAGTGTGCGTGGATAC3' RW-5'TGTGCAAGGGTGTGTTTCAT3'	*	73	57
CD80	FW-5'GCAGCAGAAGCCATGGATTAC3' RW-5'CACCAAGAGCTGAGAGACCTTGA3'	Yasunaga et al., 2003	84	60
CD86	FW-5'CGAAACCCACCCCTGATG3' RV-5'CACAAAATGACCAACATTACAAGCA3'	Yasunaga et al., 2003	70	60
TGF-β	FW-5'CAGAATGGCTGTCCTTTGATGTC3' RW-5'AGGCGAAAGCCCTCGACTT3'	Huang, 2008	79	60
IL-10	FW-5'CAAGCCCTGTCGGAGATGAT3' RW-5'CTTGATGTCTGGGTCGTGGTT3'	Yu et al., 2010	78	54
IL-12p35	FW-5'ATGACGGTCCTGTGCCTTAG3' RW-5'CTGCCTCTTGGGATCCATTA3'	*	102	57
IL-12p40	FW-5'CAGCAGAGAGGGTCAGAGTGG3' RW-5'ACGACCTCGATGGGTAGGC3'	Peters et al., 2005	109	58
IL-1α	FW-5'GTTGTTCATTGCCACACAGG3' RW-5'CAAGGCTGGGTTTCCAGTAA3'	*	93	56
IL-1β	Fw-5'CCATGAGTGTGCGTGGATAC3' Rv-5'TGTGCAAGGGTGTGTTTCAT3'	*	80	60.5
IL-8	FW-5'CACTCCACACCTTTCCATCC3' RW-5'GTCCAGGCACACCTCATTTC3'	Harman et al., 2014	120	60
IL-18	FW-5'CTCTCCTGTAAGAACAAAACTATTTCCTT3' RW-5'GAACACTTCTCTGAAAGAATATGATGTCA3'	Argyle et al., 1999	100	56

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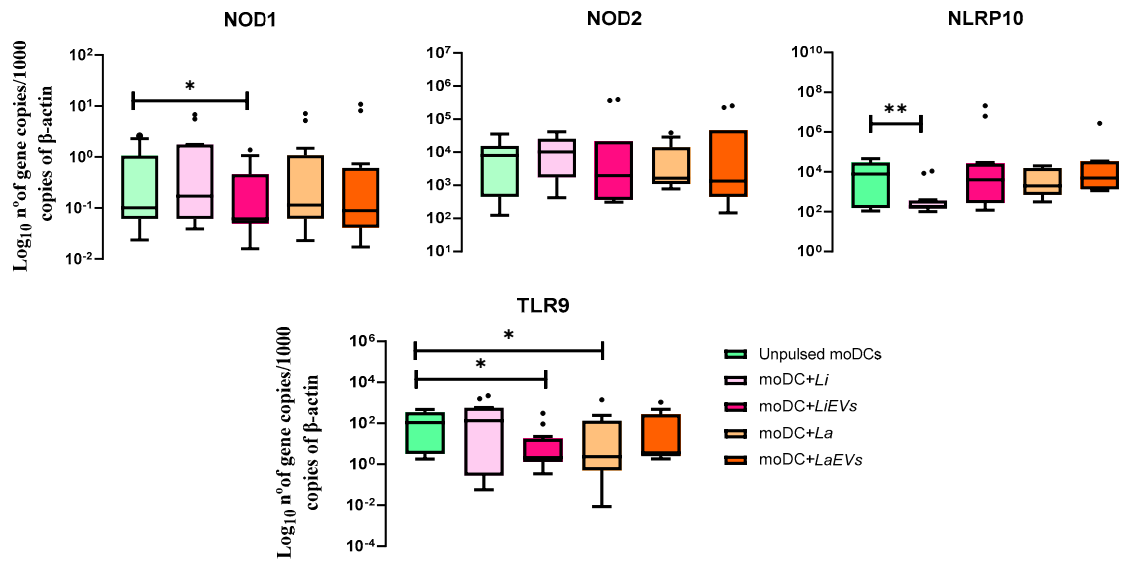
Figure S1. Gene expression of NOD1, NOD2, NLRP10, and TLR9 by *L. infantum* and *L. amazonensis* infected moDCs. RNA extracted from infected moDCs (moDC+*Li* and moDC+*La*) and moDCs exposed to parasite EVs (moDCs+LiEVs and moDCs+LaEVs) for 24 hours were used to evaluate NOD1, NOD2, NLRP10, and TLR9 gene expression. Results of at least eight samples are represented by box plots, including the median, interquartile ranges, and minimum and maximum values. Black dots are indicative of outliers. Nonparametric Wilcoxon's test was used for statistical comparisons. * ($p<0.05$) indicates statistically significant differences.
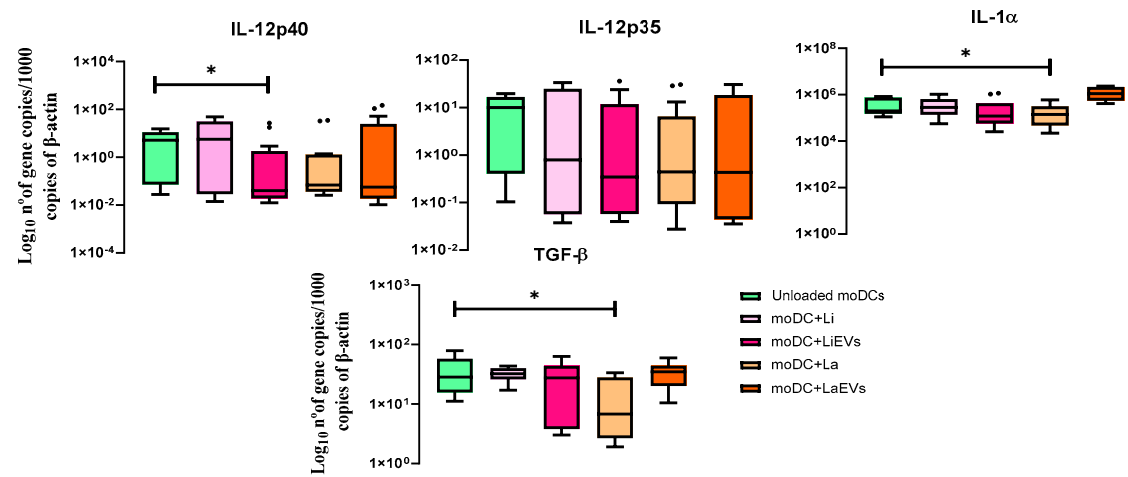
Figure S2. Gene expression of cytokines by *L. infantum* and *L. amazonensis* infected moDCs. RNA extracted from infected moDCs (moDC+Li and moDC+La) and moDCs exposed to (moDCs+LiEVs and moDCs+LaEVs) for 24 hours were used to evaluate *IL-12p40, IL-12p35, IL-1a,* and *TGF-b* gene expression. In parallel, cytokines of unloaded moDCs were also assessed. The results of at least 10 samples are represented by box plots, including the median, interquartile ranges, and minimum and maximum values. Black dots are indicative of outliers. Nonparametric Wilcoxon's test was used for statistical comparisons. *(*p*<0.05) indicates significant differences.

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