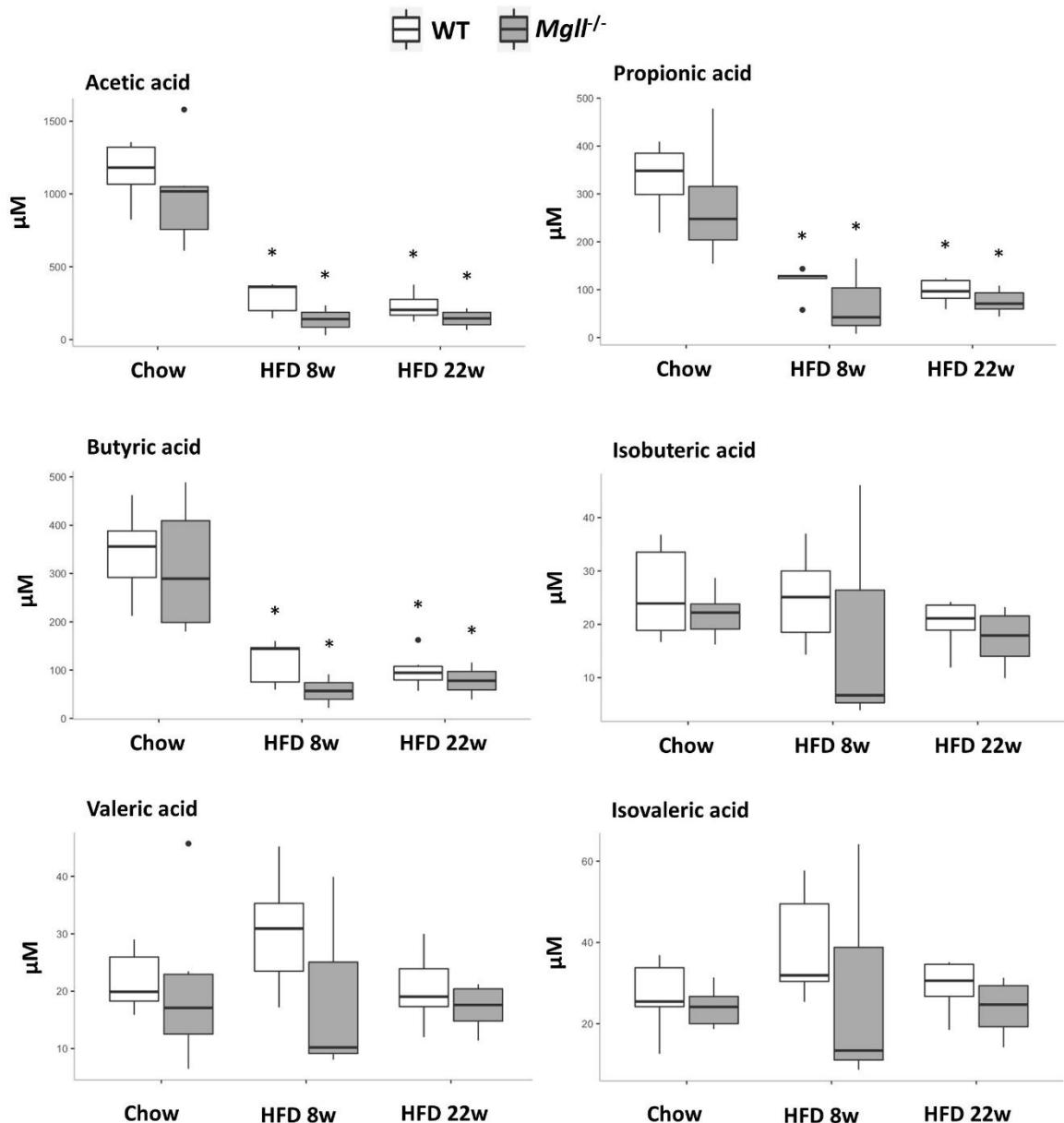


# Supplemental Materials



**Supplemental Figure S1.** Faecal SCFA levels are not different between WT and *Mgll*<sup>-/-</sup> mice. SCFA levels were measured in faecal samples from WT and *Mgll*<sup>-/-</sup> mice on chow diets and after 8 and 22 weeks of HFD feeding. \*  $p < 0.05$  vs. relevant chow control .

**Supplemental Table S1.** Culture conditions used in the targeted culturomics.

Culture Conditions	Media Composition
1. Preincubation in BACTEC Lytic Anaerobic medium with 5% sheep blood and 5% of rumen fluid, anaerobic condition at 37 °C	BACTEC Lytic Anaerobic medium (BD, Franklin Lakes, New Jersey, USA); Sheep blood (Fisher scientific, Hampton, New Hampshire, USA); Rumen fluid (Centre de recherche en sciences animales de Deschambault)
2. Preincubation in BACTEC Lytic Anaerobic medium with 5% of rumen fluid, anaerobic condition at 37 °C	BACTEC Lytic Anaerobic medium (BD, Franklin Lakes, New Jersey, USA); Rumen fluid (Centre de recherche en sciences animales de Deschambault, Quebec, Canada)

3. Preincubation in BACTEC Lytic Anaerobic medium with 5% sheep blood, anaerobic condition at 37 °C	BACTEC Lytic Anaerobic medium (BD, Franklin Lakes, New Jersey, USA); Sheep blood (Fisher scientific, Hampton, New Hampshire, USA)
4. Preincubation in BACTEC Lytic Anaerobic medium after thermic shock at 80°C/20mn, anaerobic condition at 37°C	BACTEC Lytic Anaerobic medium (BD, Franklin Lakes, New Jersey, USA); Sheep blood (Fisher scientific, Hampton, New Hampshire, USA); Rumen fluid (Centre de recherche en sciences animales de Deschambault, Quebec, Canada)
5. Preincubation in BACTEC Lytic Anaerobic medium with stool filtered at 5µm, anaerobic condition at 37°C	BACTEC Lytic Anaerobic medium (BD, Franklin Lakes, New Jersey, USA); Sheep blood
6. Preincubation in Chopped Meat Medium Carbohydrate (CMC), anaerobic condition at 37°C	Chopped Meat Medium Carbohydrate (Morgan Hill, California, USA)
7. Preincubation in Chopped Meat Medium (CM), anaerobic condition at 37°C	Chopped Meat Medium (Morgan Hill, California, USA)
8. Preincubation in Yeast Casitone Fatty Acids broth with Carbohydrates -YCFAC Broth, anaerobic condition at 37°C	Yeast Casitone Fatty Acids broth with Carbohydrates - YCFAC (Morgan Hill, California, USA)
9. Preincubation in MTGE Broth anaerobic condition at 37°C	MTGE Broth (Morgan Hill, California, USA)
10. Preincubation in PYG Broth after thermic shock at 80°C/20mn, anaerobic condition at 37°C	Peptone Yeast Extract Broth with Glucose-PYG broth
11. Preincubation in Brain Hart Infusion (BHI) Broth with stool filtered at 5µml, anaerobic condition at 37°C	Brain Hart Infusion (Anaerobe systems, Morgan Hill, California, USA)
12. Preincubation in Thioglycolate Broth, anaerobic condition at 37°C	Thioglycolate Broth (Anaerobe systems, Morgan Hill, California, USA)
13. Preincubation in BACTEC Lytic Anaerobic medium with 5% sheep blood and 5% of rumen fluid, aerobic condition at 37°C	BACTEC Lytic Anaerobic medium (BD, Franklin Lakes, New Jersey, USA); Sheep blood (Fisher scientific, Hampton, New Hampshire, USA); Rumen fluid (Centre de recherche en sciences animales de Deschambault)
14. Preincubation in BACTEC Lytic Anaerobic medium with rumen fluid, aerobic condition at 37°C	BACTEC Lytic Anaerobic medium (BD, Franklin Lakes, New Jersey, USA); Rumen fluid (Centre de recherche en sciences animales de Deschambault)
15. Preincubation in BACTEC Lytic Anaerobic medium with 5% sheep blood, aerobic condition at 37°C	BACTEC Lytic Anaerobic medium (BD, Franklin Lakes, New Jersey, USA); Sheep blood (Fisher scientific, Hampton, New Hampshire, USA)
16. Preincubation in Columbia broth, anaerobic condition at 37°C	Columbia broth (Sheep blood (Fisher scientific, Hampton, New Hampshire, USA))
17. Preincubation in BACTEC Lytic Anaerobic medium with stool filtered at 5µm, aerobic condition 37°C	BACTEC Lytic Anaerobic medium (BD, Franklin Lakes, New Jersey, USA)
18. Preincubation in Brain Hart Infusion (BHI) Broth, aerobic condition at 37°C	Brain Hart Infusion (Anaerobe systems, Morgan Hill, California, USA)

**Supplemental Table S2.** Changes in abundances of bacterial taxa according to Diet, Genotype and Genotype\*Diet over the course of the protocol.

Bacteria family	p-value		
	Diet	Genotype	Geno*Diet

			<b>p-value</b>	
		<b>Diet</b>	<b>Genotype</b>	<b>Geno*Diet</b>
<i>Anaeroplasmataceae</i>		2.6e-05	0.071	ns
<i>Bacteroidaceae</i>		ns	ns	ns
<i>Bdellovibrionaceae</i>		0.00017	ns	ns
<i>Christensenellaceae</i>		0.058	ns	ns
<i>Clostridiaceae_1</i>		7.3e-09	ns	ns
<i>Clostridiales_Incertae_Sedis_XIII</i>		0.011	ns	ns
<i>Coriobacteriaceae</i>		ns	0.0029	ns
<i>Desulfovibrionaceae</i>		6.6e-09	0.099	0.091
<i>Enterobacteriaceae</i>		ns	ns	ns
<i>Erysipelotrichaceae</i>		ns	0.011	ns
<i>Eubacteriaceae</i>		ns	0.0095	ns
<i>Lachnospiraceae</i>		0.0082	ns	ns
<i>Lactobacillaceae</i>		0.036	4.2e-06	0.015
<i>Mycoplasmataceae</i>		0.031	ns	ns
<i>Peptococcaceae_1</i>		5.1e-07	ns	ns
<i>Peptostreptococcaceae</i>		0.0029	ns	ns
<i>Porphyromonadaceae</i>		6.3e-06	ns	ns
<i>Prevotellaceae</i>		0.0032	0.049	ns
<i>Rhodospirillaceae</i>		ns	ns	ns
<i>Rikenellaceae</i>		0.023	ns	ns
<i>Ruminococcaceae</i>		0.0017	0.084	0.047
<i>Sutterellaceae</i>		8.2e-12	ns	ns
No match		6.1e-05	ns	ns
<b>Bacterial genera</b>				
<i>Acetanaerobacterium</i>		0.031	ns	ns
<i>Acetatifactor</i>		1.8e-12	ns	0.068
<i>Acetitomaculum</i>		0.046	ns	ns
<i>Aestuariispira</i>		ns	ns	ns
<i>Alistipes</i>		0.0014	ns	ns
<i>Allobaculum</i>		ns	0.021	ns
<i>Alloprevotella</i>		ns	0.038	ns
<i>Anaeroplasma</i>		2.4e-05	0.1	ns
<i>Anaerostipes</i>		0.0071	ns	ns
<i>Anaerotruncus</i>		0.001	ns	ns
<i>Anaerovorax</i>		0.0069	0.095	ns
<i>Bacteroides</i>		ns	0.06	ns
<i>Barnesiella</i>		1.5e-14	ns	ns
<i>Butyricicoccus</i>		9.8e-06	ns	ns
<i>Christensenella</i>		ns	ns	ns
<i>Clostridium_III</i>		0.0026	ns	ns
<i>Clostridium_IV</i>		2.8e-05	ns	ns
<i>Clostridium_XIVa</i>		0.00044	0.079	0.019
<i>Clostridium_XIVb</i>		ns	ns	ns
<i>Coprobacillus</i>		0.1	0.002	ns
<i>Coprobacter</i>		0.0051	ns	ns
<i>Coprococcus</i>		ns	0.012	ns
<i>Desulfovibrio</i>		ns	ns	ns
<i>Dorea</i>		1.8e-07	4.9e-06	0.0086
<i>Eisenbergiella</i>		1.1e-11	ns	0.023
<i>Enterorhabdus</i>		0.04	0.039	0.0016

<i>Escherichia</i>	ns	ns	ns
<i>Eubacterium</i>	ns	0.02	ns
<i>Falsiporphyromonas</i>	0.001	0.0055	0.056
<i>Flavonifractor</i>	1.3e-06	ns	ns
<i>Gemmiger</i>	1e-04	ns	ns
<i>Hydrogenoanaerobacterium</i>	5.3e-11	ns	0.009
<i>Intestinimonas</i>	0.013	ns	ns
<i>Lactobacillus</i>	0.012	6.1e-07	0.0033
<i>Marvinbryantia</i>	0.072	ns	ns
<i>Odoribacter</i>	0.0016	ns	ns
<i>Oscillibacter</i>	7.4e-12	ns	ns
<i>Parabacteroides</i>	ns	ns	ns
<i>Parasutterella</i>	1.1e-11	ns	ns
<i>Peptococcus</i>	3.2e-08	ns	ns
<i>Prevotella</i>	7.1e-11	ns	ns
<i>Pseudoflavonifractor</i>	4.6e-05	ns	ns
<i>Robinsoniella</i>	0.0077	ns	ns
<i>Romboutsia</i>	0.0024	ns	ns
<i>Roseburia</i>	1.9e-05	0.022	0.008
<i>Ruminococcus</i>	5.9e-15	0.018	0.097
<i>Ruminococcus2</i>	0.0012	ns	0.078
<i>Stomatobaculum</i>	0.073	5.1e-07	ns
<i>Ureaplasma</i>	0.058	ns	ns
<i>Vampirovibrio</i>	0.00011	ns	ns
No match	8.3e-05	ns	0.071