

-Article

Dual independent roles of the p24 complex in selectivity of secretory cargo export from the endoplasmic reticulum

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Supplementary data

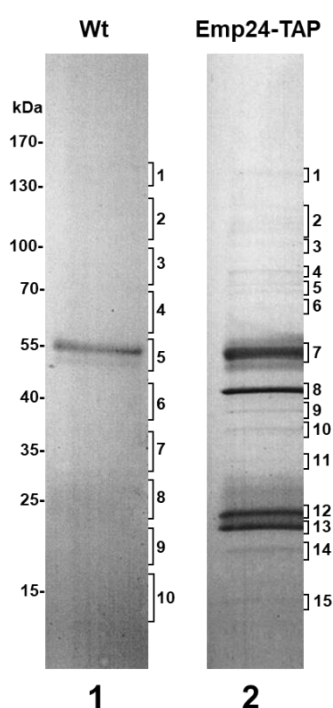


Figure S1. Mass spectrometry analysis of Emp24-TAP interactors. Enriched ER membrane fractions of wild-type yeast cells or cells expressing Emp24-TAP were isolated and solubilized with 1% digitonin. The extract was incubated with IgG-coupled magnetic beads and co-precipitated proteins were analyzed by SDS-PAGE and Coomassie Blue staining. Visualized bands (lane 2) and regions from the control reaction (lane 1) were numbered, cut out, subjected to in-gel digestion with trypsin and their protein content determined by LC-MS/MS techniques (Supplementary material Table S1, S2 and S3)

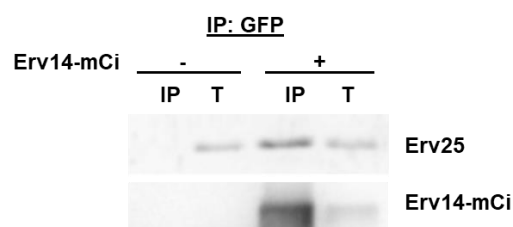


Figure S2. The p24 complex member Erv25 associates with the cargo receptor Erv14. Co-immunoprecipitation assay between Erv14-mCi and Erv25. Enriched ER membrane fractions of wild-type cells with or without expressing Erv14-mCi were solubilized, and immunoprecipitated (IP) with anti-GFP antibody, followed by immunoblotting with anti-Erv25 and anti-GFP antibodies. T represents a 0.5% of the solubilized input material.

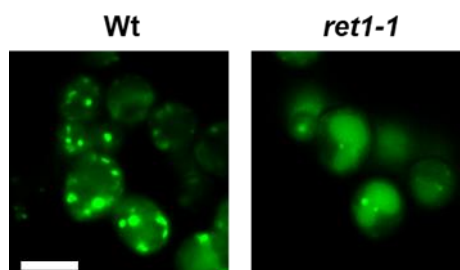


Figure S3. The thermosensitive *ret1-1* mutation impairs the Golgi-ER retrograde transport at restrictive temperature. Live images of wild-type and *ret1-1* cells expressing the recycling receptor Rer1-GFP at 37°C. In wild-type, Rer1-GFP showed the typical dotted Golgi pattern, whereas in *ret1-1*, it was mainly localized at the vacuole, indicating a defective retrograde transport. Scale bar, 5 μ m.

Table S1. Description of proteins identified by LCQ Deca XP Plus in Emp24-TAP samples by 2 or more peptides,

Table S2. Peptides identified in emp24-TAP and control samples analyzed by LCQ XP Plus analysis.