

PROJECT PE117519 *Herramientas para la enseñanza de la  
Geomática con programas de código abierto*

SUPPLEMENTARY MATERIAL

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# Analysis of High Temporal Resolution Land Use/Land Cover Trajectories

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## 1 Introduction

In this document, a R script to carry out an analysis of land use / cover sequences similar to the one reported in the article *Analysis of High Temporal Resolution Land Use/Land Cover Trajectories* is presented. Methods, originally developed to assess life course trajectories and implemented in the R package TraMineR (<http://traminer.unige.ch/>), are explored in order to evaluate land change through the analysis of sequences of land use / cover.

We will process a very small portion of the maps from the MapBiomass project ([mapbiomas.org/](http://mapbiomas.org/)) used in the paper along with covariates maps (distance from roads, elevation and slope). The maps show the distribution of land use/cover in a small region of Northeastern Brazil for eleven dates from circa 1986 to 2016. The sequences analysis is mainly conducted using the R package **TraMineR** (Gabadinho et al., 2011).

## 2 Materials and preprocessing

### 2.1 Importing cartographic data into R

The maps (raster tif files) are:

1. Time serie of land use / cover maps (serie.tif),
2. Distance from roads (dist\_roads.tif),
3. Elevation (dem.tif),

#### 4. Slope (slope.tif).

It is also necessary to install the packages **TraMineR**, **raster** and **fastcluster**. First, the tif files are imported as raster objects into R.

```
# Working directory and libraries
setwd("/home/jf/Dropbox/sequences/SupplementaryMaterials")
library(TraMineR)
library(raster)
library(fastcluster)

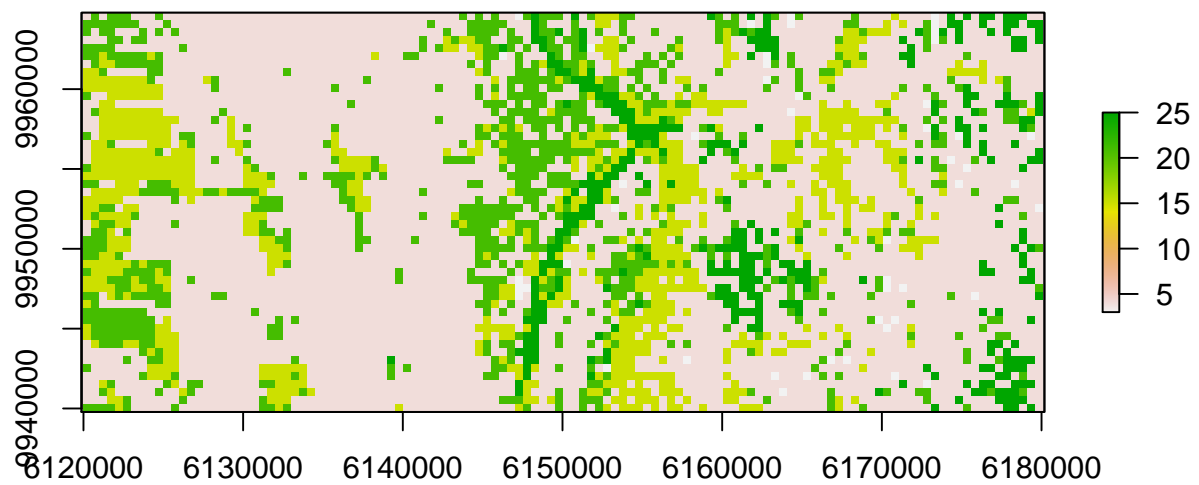
## Read rasters of time serie of land cover and covariates

## LUC maps for eleven dates
serie <- stack("serie.tif")
summary(serie)

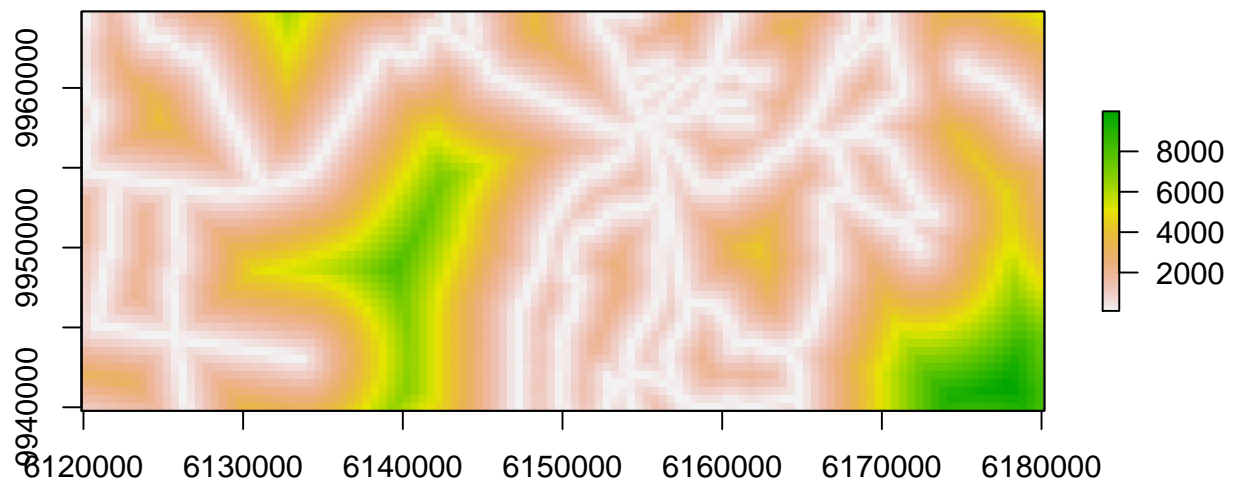
##          serie.1 serie.2 serie.3 serie.4 serie.5 serie.6 serie.7 serie.8
## Min.           3      3      3      3      3      3      3      3
## 1st Qu.         4      4      4      4      4      4      4      4
## Median          4      4      4      4      4      4      4      4
## 3rd Qu.        15     15     15     15     15     15     15     15
## Max.           25     25     25     25     25     25     25     25
## NA's            0      0      0      0      0      0      0      0
##          serie.9 serie.10 serie.11
## Min.           3      3      3
## 1st Qu.         4      4      4
## Median          4      4      4
## 3rd Qu.        15     15     15
## Max.           25     25     25
## NA's            0      0      0

# Distance from roads
dist <- raster("dist_roads.tif")
# Elevation
dem <- raster("dem.tif")
# Slope
slope <- raster("slope.tif")

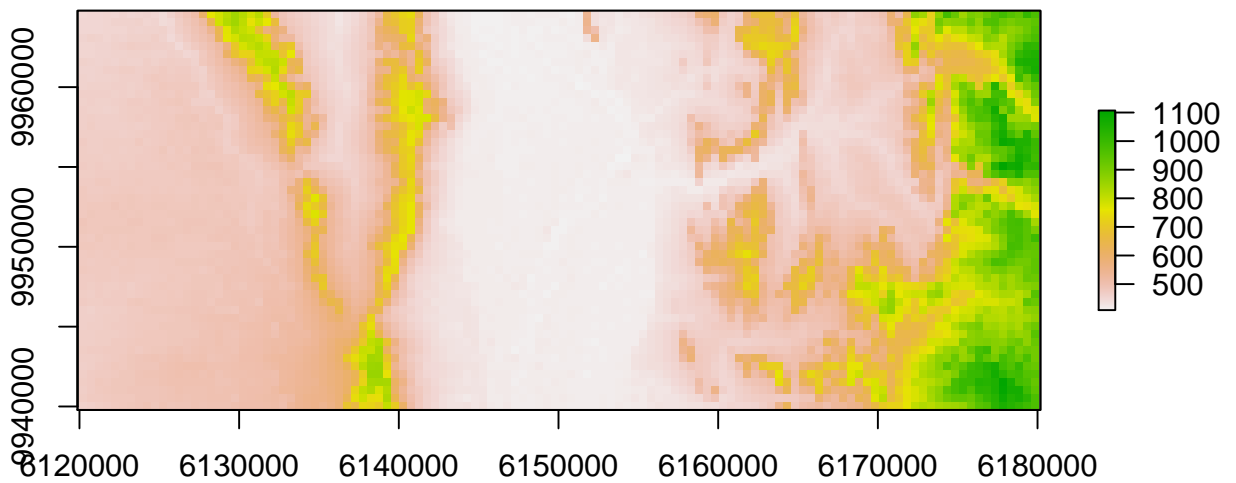
## Plot the maps
plot(serie[[1]])
```



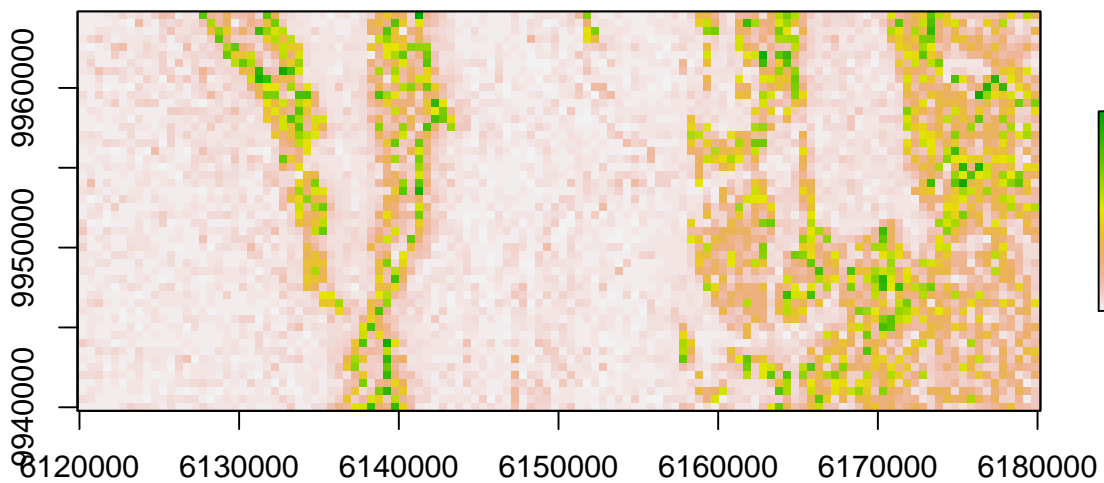
```
plot(dist)
```



```
plot(dem)
```



```
plot(slope)
```



## 2.2 Creation of tabular data

For further analysis, information from the raster data should be presented as tabular data. In this table, each row represents a cell, and each column its value in the maps (category number and

value of the covariates). Additional columns are created to hold the information of the covariates as categorical data. A column with the entire concatenated sequence is also created. The function **rasterToPoints** enables us to obtain the coordinates of each cell, this information will permit the spatial visualization of the results.

```
## Elaboration of a table which describes for each pixel the land category
# at each time step + value of covariates

## using rasterToPoints(r) to get coordinates
tab <- as.data.frame(rasterToPoints(stack(serie,dem,slope,dist)))
names(tab) <- c("x","y","c1986","c1989","c1992","c1995","c1998","c2001","c2004",
               "c2007","c2010","c2013","c2016","dem","slope","dist")
# A look at the first rows of the table:
head(tab)
```

##		x	y	c1986	c1989	c1992	c1995	c1998	c2001	c2004	c2007	c2010
## 1		6120280	9964535	4	21	21	21	15	15	15	15	15
## 2		6120780	9964535	21	21	21	21	15	21	15	15	15
## 3		6121280	9964535	4	21	21	21	21	21	15	15	15
## 4		6121780	9964535	21	21	21	15	15	15	15	15	15
## 5		6122280	9964535	21	21	15	15	15	15	15	15	15
## 6		6122780	9964535	21	21	15	15	15	15	15	15	15

```
##      c2013 c2016 dem slope dist
## 1      15     15 451     1  100
## 2      21     15 452     1  600
## 3      15     15 453     1 1100
## 4      15     15 457     2 1600
## 5      21     21 454     2 1264
## 6      21     15 458     5  806

# and a summary:
summary(tab)
```

##		x	y	c1986	c1989
##	Min.	:6120280	Min. :9940035	Min. : 3.000	Min. : 3.000
##	1st Qu.:	:6135155	1st Qu.:9946035	1st Qu.: 4.000	1st Qu.: 4.000
##	Median :	:6150030	Median :9952285	Median : 4.000	Median : 4.000
##	Mean :	:6150030	Mean : 9952285	Mean : 9.148	Mean : 9.778
##	3rd Qu.:	:6164905	3rd Qu.:9958535	3rd Qu.:15.000	3rd Qu.:15.000
##	Max. :	:6179780	Max. :9964535	Max. :25.000	Max. :25.000

##		c1992	c1995	c1998	c2001
##	Min.	: 3.000	Min. : 3.000	Min. : 3.000	Min. : 3.000
##	1st Qu.:	: 4.000	1st Qu.: 4.000	1st Qu.: 4.000	1st Qu.: 4.000
##	Median :	: 4.000	Median : 4.000	Median : 4.000	Median : 4.000
##	Mean :	: 9.591	Mean : 9.357	Mean : 9.075	Mean : 9.399
##	3rd Qu.:	:15.000	3rd Qu.:15.000	3rd Qu.:15.000	3rd Qu.:15.000
##	Max. :	:25.000	Max. :25.000	Max. :25.000	Max. :25.000

```
##      c2004      c2007      c2010      c2013
## Min.    : 3.000  Min.    : 3.000  Min.    : 3.000  Min.    : 3.000
## 1st Qu.: 4.000  1st Qu.: 4.000  1st Qu.: 4.000  1st Qu.: 4.000
## Median : 4.000  Median : 4.000  Median : 4.000  Median : 4.000
## Mean   : 9.373  Mean   : 9.063  Mean   : 9.112  Mean   : 8.493
## 3rd Qu.:15.000  3rd Qu.:15.000  3rd Qu.:15.000  3rd Qu.:15.000
## Max.   :25.000  Max.   :25.000  Max.   :25.000  Max.   :25.000
##      c2016      dem      slope      dist
## Min.    : 3.000  Min.    : 409.0  Min.    : 0.00  Min.    : 100
## 1st Qu.: 4.000  1st Qu.: 442.0  1st Qu.: 2.00  1st Qu.: 608
## Median : 4.000  Median : 482.0  Median : 4.00  Median :1486
## Mean   : 8.673  Mean   : 539.5  Mean   :10.08  Mean   :2136
## 3rd Qu.:15.000  3rd Qu.: 578.0  3rd Qu.:13.00  3rd Qu.:3041
## Max.   :25.000  Max.   :1107.0  Max.   :76.00  Max.   :9985

## Create categorical data from covariates
tab$demK<-cut(tab$dem, c(400,700,1000,1200))
tab$slopeK<-cut(tab$slope, c(-1,2,6,102))
tab$distK<-cut(tab$dist, c(0,500,1000,2000,5000,10000))
## Two categories for distance and slope (binary variable)
tab$distK2<-cut(tab$dist, c(0,1000,10000))
tab$slopeK2<-cut(tab$slope, c(-1,6,102))

# Put an extra column with the concatenated sequence
tab$sec <- as.vector(paste(tab[,3],tab[,4],tab[,5],tab[,6],tab[,7],
  tab[,8],tab[,9],tab[,10],tab[,11],tab[,12],tab[,13],sep="-"))
```

## 2.3 Preliminar sequence analysis

As a first step, the categories which appear in the entire time serie are determined. There are six land cover categories (3, 4, 15, 18, 21, 25) which are 1) Forest, 2) Savanna, 3) Grasslands, 4) Mosaic of agriculture and pastures and 5) others. Then, existing sequences, and their respective frequency, are computed.

```
# Verify all the categories for all the time steps
classes <- c()
for (year in 3:13){
  classes <- unique(c(classes,tab[,year]))
}
print(sort(classes))

## [1]  3  4 15 18 21 25

# [1]  3  4 15 18 21 25

### Determine the number of different sequences
```



```

unique_traj <- sort(unique(tab$sec))
length(unique_traj) # 1651 different trajectories

## [1] 1651

# Determine the frequency of these trajectories
tab_freq <- as.data.frame(table(tab$sec))

# Sort the sequences from more to less frequent
tab_freq2 <- tab_freq[order(tab_freq$Freq,decreasing = TRUE),]
head(tab_freq2)

##                               Var1 Freq
## 1651          4-4-4-4-4-4-4-4-4-4 2515
## 1      15-15-15-15-15-15-15-15-15 271
## 867    25-25-25-25-25-25-25-25-25 142
## 551    21-21-21-21-21-21-21-21-21  49
## 695          21-21-4-4-4-4-4-4-4  44
## 795          21-4-4-4-4-4-4-4-4  36

```

## 2.4 Creation of a state sequence object

To carry out the analysis with **TraMineR**, a state sequence object should be created. It comprises information as the names and the colors assigned to each one of the land cover category ("states"). In sequence analysis, the alphabet is the finite set of possible states.

```

# Creation of a state sequence object to which names are assigned
# short state names (for printed output) and long state labels
# (for the graphics' legend).
# Also define the alphabet (be sure that the order of the names and labels
# is the same of the alphabet and a palette of colors for display.

## State sequence object
alphabet <- c(3,4,15,18,21,25)
labels <- c("Forest","Savanna","Pasture","Agriculture","Mosaic Agri/Past","Others")
short_labels <- c("F","Sav","Past","Agri","Mosaic","Others")
palette <- c("darkolivegreen","green","yellow","red","orange","grey")
tab.seq <- seqdef(tab, 3:13, alphabet = alphabet, states = short_labels,
                  cpal = palette, labels = labels)

## [>] state coding:
##      [alphabet] [label] [long label]
##      1          3F      Forest
##      2          4Sav     Savanna
##      3          15Past   Pasture
##      4          18Agri   Agriculture

```

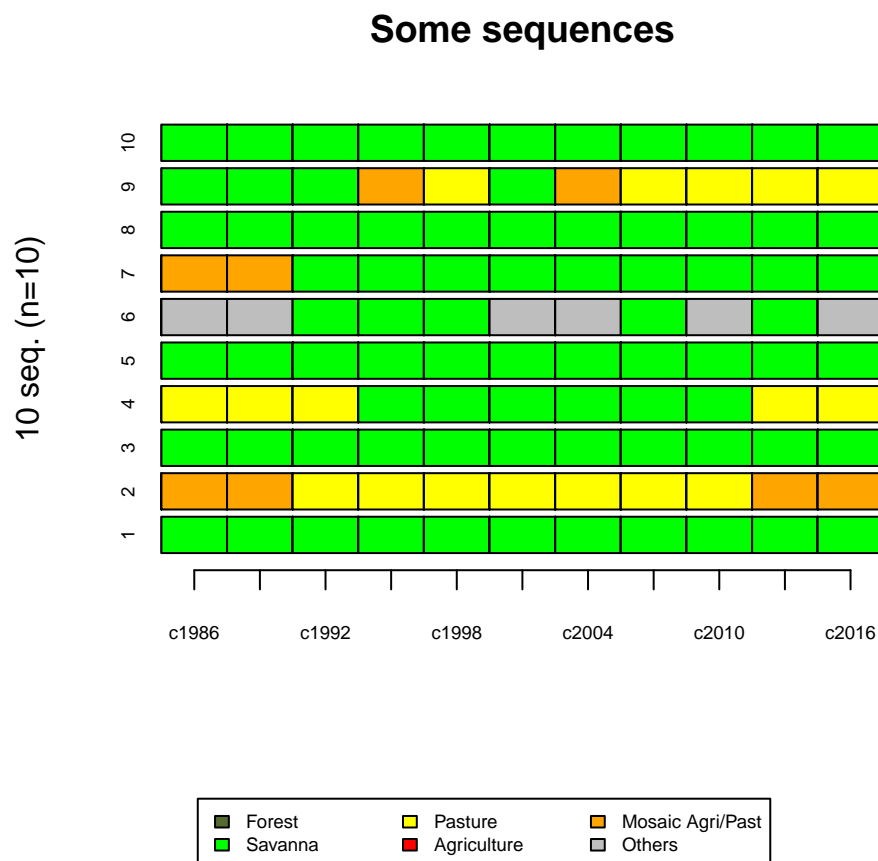
```
##      5      21Mosaic Mosaic Agri/Past
##      6      250others Others
##  [>] 6000 sequences in the data set
##  [>] min/max sequence length: 11/11
```

### 3 Visualization of the sequences

Figures which describe the observed sequences are elaborated: graph of some representative or random sequences, graph of all the observed sequences, most common sequences, land cover (states) proportion over time, landscape entropy over time, most frequent (dominant) category, average time of permanence of each land cover category.

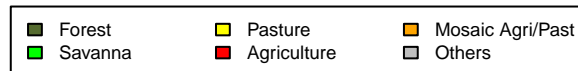
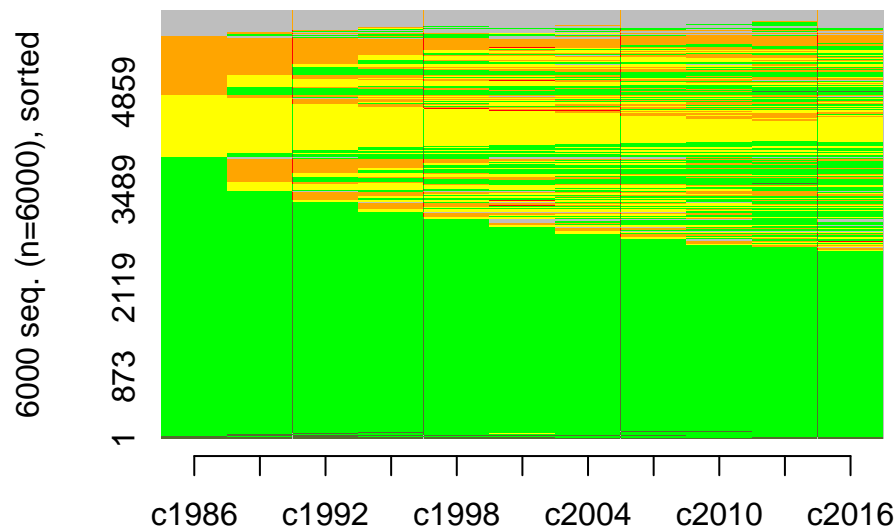
```
## Visualize the sequence data set

# Plot 10 sequences in the tab.seq sequence object (chosen to show diversity)
some_seq <- tab.seq[c(19,5,973,976,34,84,930,3893,993,995),]
seqiplot(some_seq, with.legend = T, cex.legend = 0.6, border = T,
          cex.axis=0.6, main = "Some sequences")
```



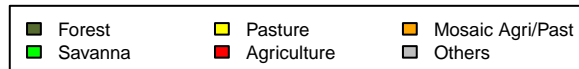
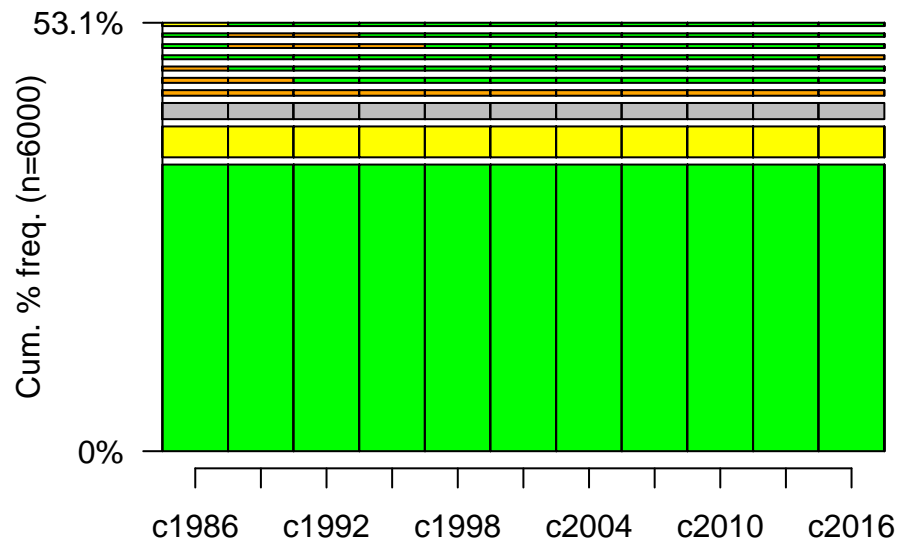
```
# Plot all the sequences in the data set, sorted by states from start.
seqIplot(tab.seq, sortv = "from.start", with.legend = T,
         cex.legend = 0.6, main = "Sequences 1985-2017")
```

## Sequences 1985-2017



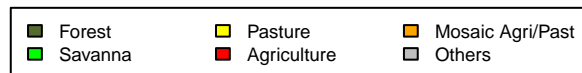
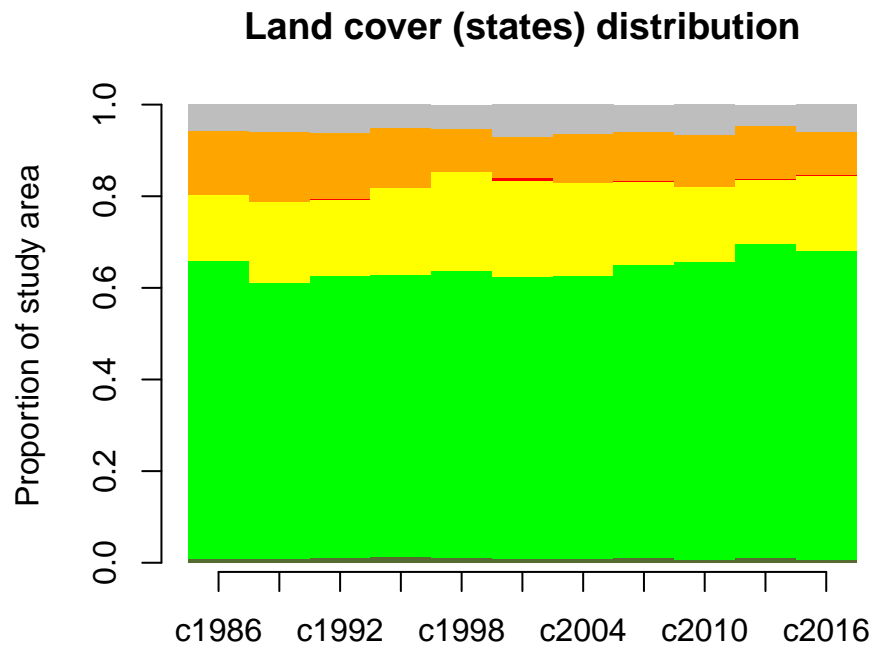
```
# Plot the 10 most frequent sequences.
seqfplot(tab.seq, with.legend = T, cex.legend = 0.6,
         main="Most common sequences")
```

## Most common sequences

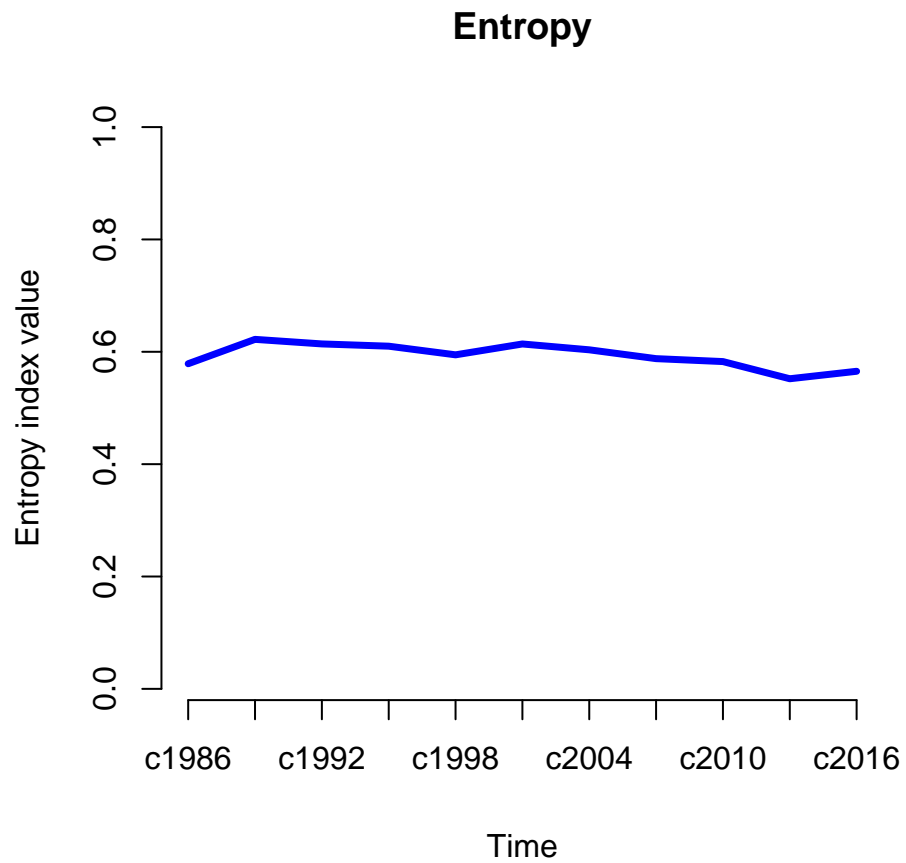


```
## Explore the sequence data set by computing and visualizing
# descriptive statistics

# Compute and plot the state distributions by time step.
# With border = NA, borders surrounding the bars are removed.
seqdplot(tab.seq, with.legend = T, border = NA,, cex.legend = 0.6,
main="Land cover (states) distribution", ylab="Proportion of study area")
```

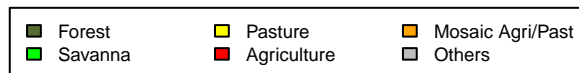
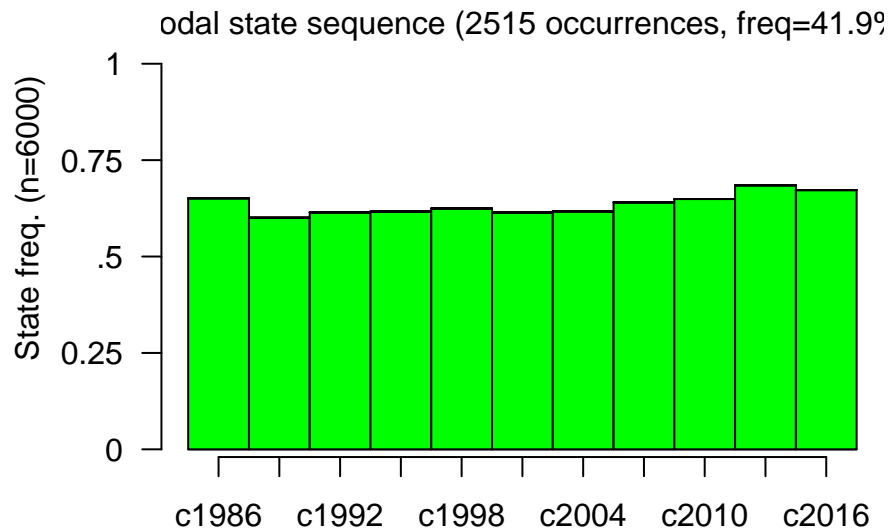


```
# Compute and plot the transversal entropy index (Landscape entropy over time)
seqHtpplot(tab.seq, main = "Entropy", ylab="Entropy index value", xlab=("Time"))
```

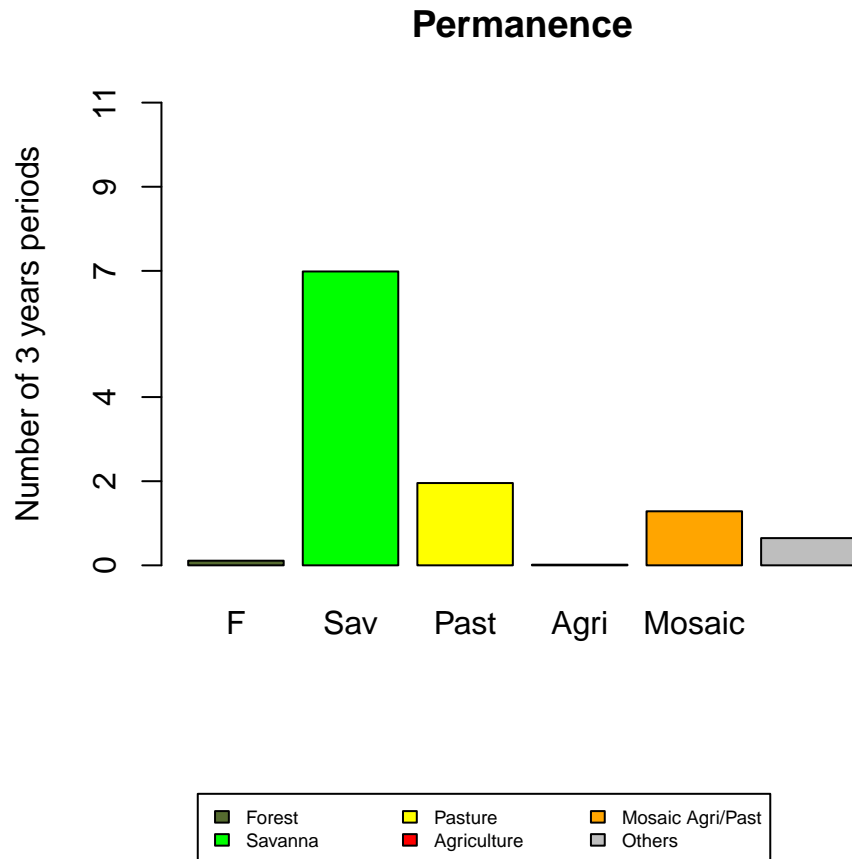


```
#Plot the sequence of modal states (dominant land cover)  
# of the transversal state distributions.  
seqmsplot(tab.seq, with.legend = T, main = "Most frequent land cover",  
           cex.legend = 0.6)
```

## Most frequent land cover



```
# Plot the mean time spent in each land cover category.
seqmtplot(tab.seq, with.legend = T, main = "Permanence", cex.legend = 0.6,
  ylab="Number of 3 years periods")
```



Spatial complexity is evaluated by computing and mapping the longitudinal entropy index (also known as Shannon index) and the turbulence index (Elzinga, 2007).

```
### Longitudinal turbulence and entropy indices
# Computed for each pixel over the time
tab$Turb <- seqST(tab.seq, norm=FALSE)

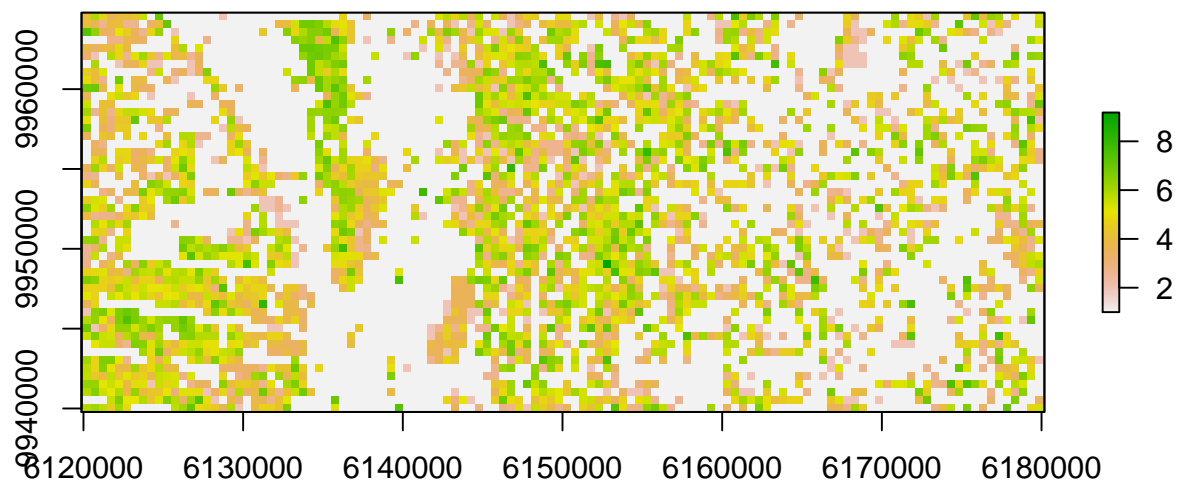
## [>] extracting symbols and durations ...
## [>] computing turbulence for 6000 sequence(s) ...

tab$Entrop <- sequent(tab.seq, norm=TRUE, base=exp(1))

## [>] computing entropy for 6000 sequences ...
## [>] computing state distribution for 6000 sequences ...

# Generate rasters which represent these indices
xyt <- as.data.frame(cbind(tab$x,tab$y,tab$Turb))
names(xyt) <- c("x","y","t")
coordinates(xyt) <- ~ x + y
gridded(xyt) <- TRUE
raster_turb <- raster(xyt)
plot(raster_turb)
```





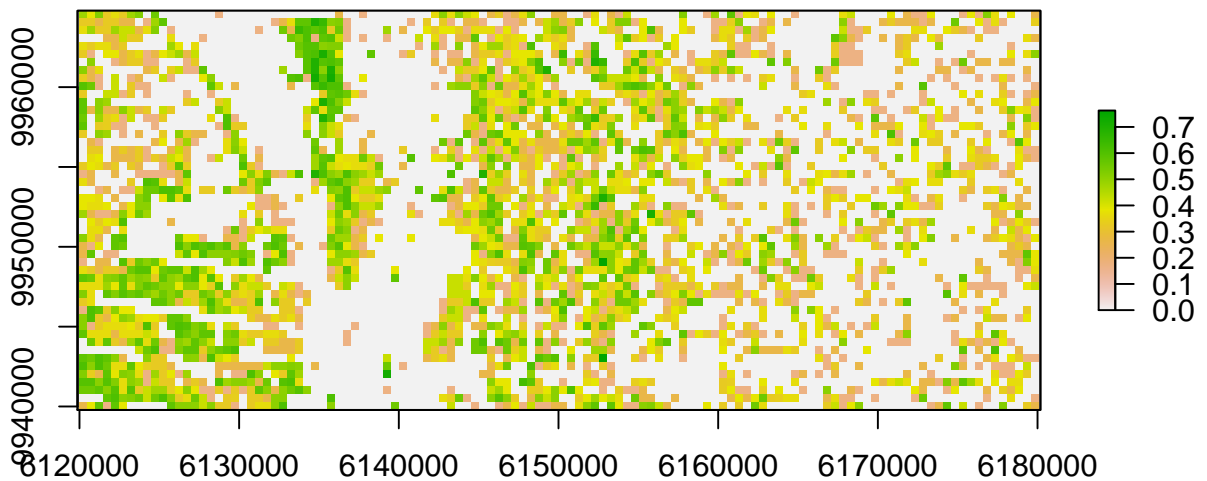
```

xye <- as.data.frame(cbind(tab$x, tab$y, tab$Entrop))
names(xye) <- c("x", "y", "e")
head(xye)

##           x           y           e
## 1 6120280 9964535 0.4799569
## 2 6120780 9964535 0.3845434
## 3 6121280 9964535 0.5217050
## 4 6121780 9964535 0.3270264
## 5 6122280 9964535 0.3658313
## 6 6122780 9964535 0.3270264

coordinates(xye) <- ~ x + y
gridded(xye) <- TRUE
raster_entrop <- raster(xye)
plot(raster_entrop)

```



```
# Calculate the correlation between both indices
cor(tab$Turb,tab$Entrop) # 0.9394874
```

```
## Entropy
## Turbulence 0.9404563
```

Finally, the transition rates between land cover categories can be easily computed.

```
## Computes the transition rates
tr_rates <- seqtrate(tab.seq)
```

```
## [>] computing transition probabilities for states F/Sav/Past/Agri/Mosaic/Others
...
```

```
print(tr_rates)
```

```
##           [-> F]    [-> Sav]    [-> Past]    [-> Agri] [-> Mosaic]
## [F ->]      7.594108e-01 0.22913257 0.001636661 0.0000000000 0.003273322
## [Sav ->]    3.669192e-03 0.93392815 0.016155004 0.0002375736 0.033973022
## [Past ->]   9.294544e-05 0.06022865 0.832326424 0.0033460359 0.099823404
## [Agri ->]   0.000000e+00 0.01449275 0.492753623 0.2898550725 0.202898551
## [Mosaic ->] 5.592059e-04 0.19795890 0.172095624 0.0039144415 0.600587166
## [Others ->] 5.673759e-04 0.12056738 0.011347518 0.0000000000 0.057872340
##           [-> Others]
## [F ->]      0.006546645
## [Sav ->]    0.012037061
## [Past ->]   0.004182545
```

```
## [Agri ->] 0.000000000
## [Mosaic ->] 0.024884664
## [Others ->] 0.809645390
```

## 4 Dissimilarity analysis

### 4.1 Computing of dissimilarities

Dissimilarity between sequences can be calculated using different methods. In this document, we used the longest common subsequence (LCS) distance, the longest common prefix (LCP) and the optimal matching (OM) based on transition rates (OM1) and on features (OM2) (Elzinga, 2007; Gabadinho et al., 2011; Studer and Ritschard, 2016). Then, a cluster hierarchical analysis is applied to identify similar sequences.

```
# Compute distances between sequences using different dissimilarity indices

## OM with substitution costs based on transition
## probabilities and indel set as half the maximum
## substitution cost
costs.tr <- seqcost(tab.seq, method = "TRATE", with.missing = FALSE)

## [>] creating substitution-cost matrix using transition rates ...
## [>] computing transition probabilities for states F/Sav/Past/Agri/Mosaic/Others
...
print(costs.tr)

## $indel
## [1] 1
##
## $sm
##           F->   Sav->   Past->   Agri-> Mosaic-> Others->
## F->         0.000000 1.767198 1.998270 2.000000 1.996167 1.992886
## Sav->       1.767198 0.000000 1.923616 1.985270 1.768068 1.867396
## Past->      1.998270 1.923616 0.000000 1.503900 1.728081 1.984470
## Agri->      2.000000 1.985270 1.503900 0.000000 1.793187 2.000000
## Mosaic->    1.996167 1.768068 1.728081 1.793187 0.000000 1.917243
## Others->    1.992886 1.867396 1.984470 2.000000 1.917243 0.000000

dist.om1 <- seqdist(tab.seq, method = "OM", indel = costs.tr$indel,
                    sm = costs.tr$sm, with.missing = F)

## [>] 6000 sequences with 6 distinct states
## [>] checking 'sm' (one value for each state, triangle inequality)
## [>] 1651 distinct sequences
## [>] min/max sequence length: 11/11
## [>] computing distances using the OM metric
## [>] elapsed time: 1.376 secs
```

```

dim(dist.om1)

## [1] 6000 6000

### OM based on features
tab_state_features <- data.frame(state=c(10,5,3,3,3,1))
costs.gower <- seqcost(tab.seq, method = "FEATURES",with.missing = FALSE,
                        state.features = tab_state_features)
print(costs.gower)

## $indel
## [1] 0.5
##
## $sm
##           F->      Sav->      Past->      Agri->      Mosaic->      Others->
## F->          0.0000000 0.5555556 0.7777778 0.7777778 0.7777778 1.0000000
## Sav->        0.5555556 0.0000000 0.2222222 0.2222222 0.2222222 0.4444444
## Past->       0.7777778 0.2222222 0.0000000 0.0000000 0.0000000 0.2222222
## Agri->       0.7777778 0.2222222 0.0000000 0.0000000 0.0000000 0.2222222
## Mosaic->    0.7777778 0.2222222 0.0000000 0.0000000 0.0000000 0.2222222
## Others->    1.0000000 0.4444444 0.2222222 0.2222222 0.2222222 0.0000000

dist.om2 <- seqdist(tab.seq, method = "OM",indel = costs.gower$indel,
                    sm = costs.gower$sm,with.missing = F)

## [>] 6000 sequences with 6 distinct states
## [>] checking 'sm' (one value for each state, triangle inequality)
## [>] 1651 distinct sequences
## [>] min/max sequence length: 11/11
## [>] computing distances using the OM metric
## [>] elapsed time: 1.076 secs

dim(dist.om2)

## [1] 6000 6000

## LCS
dist.lcs <- seqdist(tab.seq, method = "LCS")

## [>] 6000 sequences with 6 distinct states
## [>] creating a 'sm' with a substitution cost of 2
## [>] creating 6x6 substitution-cost matrix using 2 as constant value
## [>] 1651 distinct sequences
## [>] min/max sequence length: 11/11
## [>] computing distances using the LCS metric
## [>] elapsed time: 1.052 secs

## LCP
dist.lcp <- seqdist(tab.seq, method = "LCP")

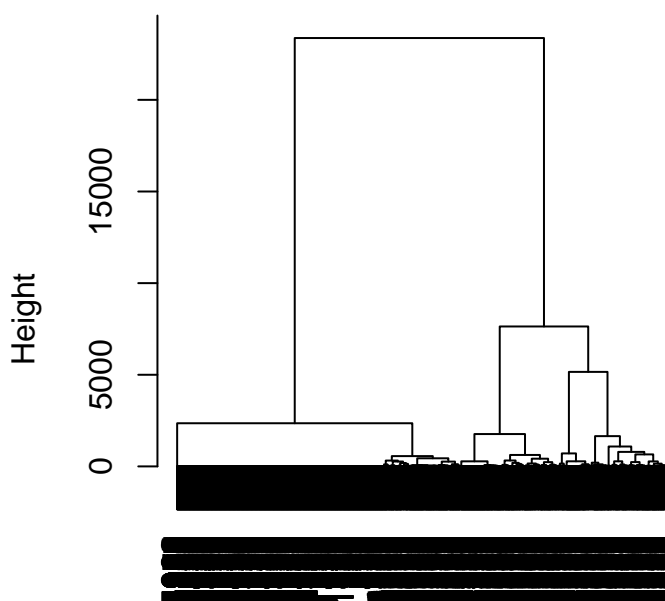
```

```
## [>] 6000 sequences with 6 distinct states
## [>] 1651 distinct sequences
## [>] min/max sequence length: 11/11
## [>] computing distances using the LCP metric
## [>] elapsed time: 0.664 secs

# Elaboration of a typology of the trajectories by hierarchical clustering
# of the sequences from the different distances and retrieve for each cell
# sequence the cluster membership of the 4 class solution.

## Cluster based on OM transition rates
clusterward_om1 <- hclust(as.dist(dist.om1),method="ward.D")
plot(clusterward_om1)
```

## Cluster Dendrogram



```
as.dist(dist.om1)
hclust (*, "ward.D")
```

```
cl_om1 <- cutree(clusterward_om1, k = 4)
tab$clusterom1 <- cl_om1
head(tab)
```

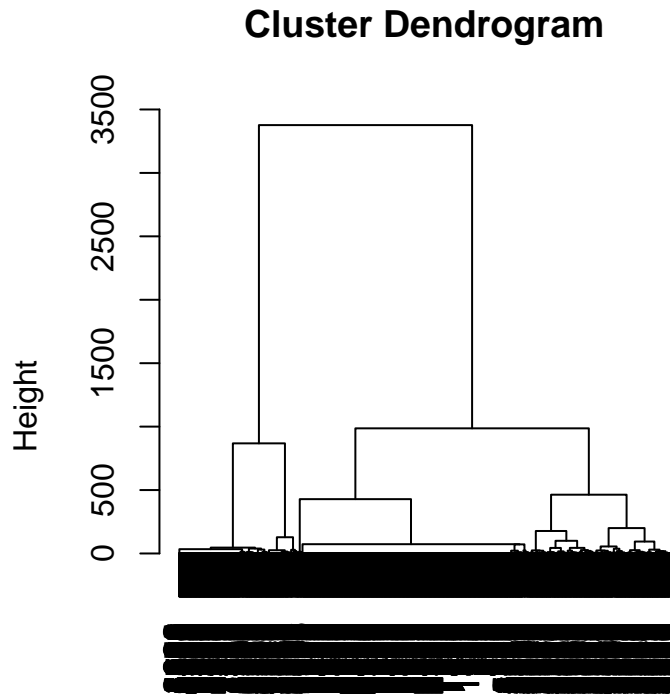
##	x	y	c1986	c1989	c1992	c1995	c1998	c2001	c2004	c2007	c2010
## 1	6120280	9964535	4	21	21	21	15	15	15	15	15
## 2	6120780	9964535	21	21	21	21	15	21	15	15	15

```

## 3 6121280 9964535      4      21      21      21      21      21      15      15      15
## 4 6121780 9964535      21      21      21      15      15      15      15      15      15
## 5 6122280 9964535      21      21      15      15      15      15      15      15      15
## 6 6122780 9964535      21      21      15      15      15      15      15      15      15
##      c2013 c2016 dem slope dist      demK slopeK      distK      distK2
## 1      15      15 451      1  100 (400,700] (-1,2]      (0,500]      (0,1e+03]
## 2      21      15 452      1  600 (400,700] (-1,2]      (500,1e+03]      (0,1e+03]
## 3      15      15 453      1 1100 (400,700] (-1,2]      (1e+03,2e+03]      (1e+03,1e+04]
## 4      15      15 457      2 1600 (400,700] (-1,2]      (1e+03,2e+03]      (1e+03,1e+04]
## 5      21      21 454      2 1264 (400,700] (-1,2]      (1e+03,2e+03]      (1e+03,1e+04]
## 6      21      15 458      5  806 (400,700] (2,6]      (500,1e+03]      (0,1e+03]
##      slopeK2      sec Turbulence      Entropy clusterom1
## 1 (-1,6] 4-21-21-21-15-15-15-15-15-15-15 4.075664 0.4799569      1
## 2 (-1,6] 21-21-21-21-15-21-15-15-15-21-15 5.899578 0.3845434      1
## 3 (-1,6] 4-21-21-21-21-21-15-15-15-15-15 4.740480 0.5217050      1
## 4 (-1,6] 21-21-21-15-15-15-15-15-15-15-15 3.551410 0.3270264      1
## 5 (-1,6] 21-21-15-15-15-15-15-15-15-21-21 4.022744 0.3658313      1
## 6 (-1,6] 21-21-15-15-15-15-15-15-15-21-15 4.088201 0.3270264      1

## Cluster based on OM features
clusterward_om2 <- hclust(as.dist(dist.om2),method="ward.D")
plot(clusterward_om2)

```



```
as.dist(dist.om2)
hclust (*, "ward.D")
```

```
cl_om2 <- cutree(clusterward_om2, k = 4)
tab$clusterom2 <- cl_om2
head(tab)
```

##	x	y	c1986	c1989	c1992	c1995	c1998	c2001	c2004	c2007	c2010
## 1	6120280	9964535	4	21	21	21	15	15	15	15	15
## 2	6120780	9964535	21	21	21	21	15	21	15	15	15
## 3	6121280	9964535	4	21	21	21	21	21	15	15	15
## 4	6121780	9964535	21	21	21	15	15	15	15	15	15
## 5	6122280	9964535	21	21	15	15	15	15	15	15	15
## 6	6122780	9964535	21	21	15	15	15	15	15	15	15

##	c2013	c2016	dem	slope	dist	demK	slopeK	distK	distK2
## 1	15	15	451	1	100 (400,700]	(-1,2]	(0,500]	(0,1e+03]	
## 2	21	15	452	1	600 (400,700]	(-1,2]	(500,1e+03]	(0,1e+03]	
## 3	15	15	453	1	1100 (400,700]	(-1,2]	(1e+03,2e+03]	(1e+03,1e+04]	
## 4	15	15	457	2	1600 (400,700]	(-1,2]	(1e+03,2e+03]	(1e+03,1e+04]	
## 5	21	21	454	2	1264 (400,700]	(-1,2]	(1e+03,2e+03]	(1e+03,1e+04]	
## 6	21	15	458	5	806 (400,700]	(2,6]	(500,1e+03]	(0,1e+03]	

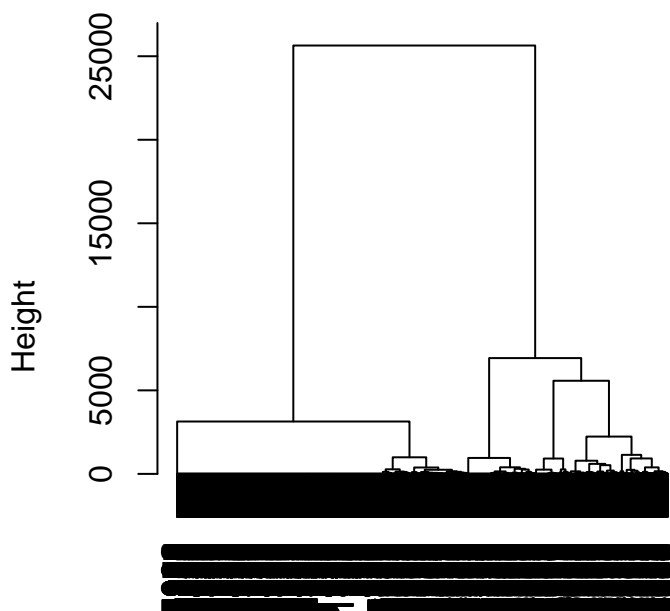
  

##	slopeK2	sec	Turbulence	Entropy	clusterom1
## 1	(-1,6]	4-21-21-21-15-15-15-15-15-15-15	4.075664	0.4799569	1
## 2	(-1,6]	21-21-21-21-15-21-15-15-15-21-15	5.899578	0.3845434	1

```
## 3  (-1,6]  4-21-21-21-21-21-15-15-15-15-15  4.740480 0.5217050      1
## 4  (-1,6]  21-21-21-15-15-15-15-15-15-15-15  3.551410 0.3270264      1
## 5  (-1,6]  21-21-15-15-15-15-15-15-15-15-21  4.022744 0.3658313      1
## 6  (-1,6]  21-21-15-15-15-15-15-15-15-15-21  4.088201 0.3270264      1
##  clusterom2
## 1          1
## 2          1
## 3          1
## 4          1
## 5          1
## 6          1

## Cluster based on LCS
clusterward_lcs <- hclust(as.dist(dist.lcs),method="ward.D")
plot(clusterward_lcs)
```

## Cluster Dendrogram



```
as.dist(dist.lcs)
hclust (*, "ward.D")
```

```
cl_lcs <- cutree(clusterward_lcs, k = 4)
tab$clusterlcs <- cl_lcs
head(tab)

##          x          y c1986 c1989 c1992 c1995 c1998 c2001 c2004 c2007 c2010
```



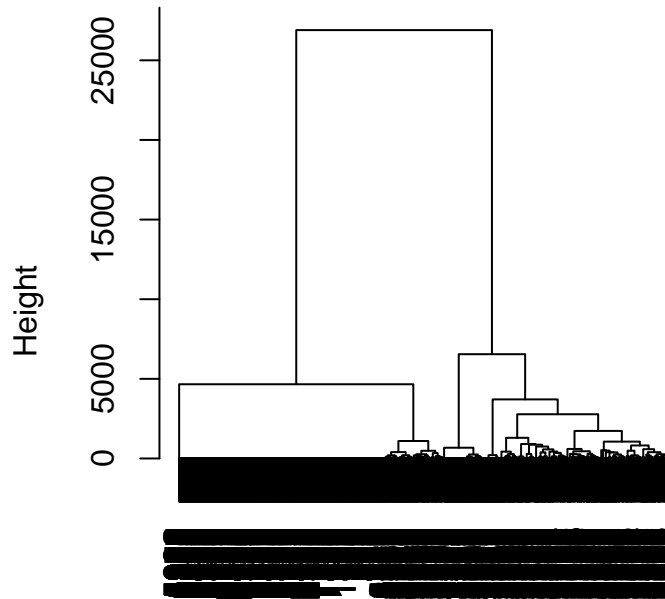
```

## 1 6120280 9964535      4      21      21      21      15      15      15      15      15
## 2 6120780 9964535     21      21      21      21      15      21      15      15      15
## 3 6121280 9964535      4      21      21      21      21      21      15      15      15
## 4 6121780 9964535     21      21      21      15      15      15      15      15      15
## 5 6122280 9964535     21      21      15      15      15      15      15      15      15
## 6 6122780 9964535     21      21      15      15      15      15      15      15      15
##      c2013 c2016 dem slope dist      demK slopeK      distK      distK2
## 1      15      15 451      1  100 (400,700] (-1,2]      (0,500]      (0,1e+03]
## 2      21      15 452      1   600 (400,700] (-1,2]      (500,1e+03]      (0,1e+03]
## 3      15      15 453      1 1100 (400,700] (-1,2]      (1e+03,2e+03] (1e+03,1e+04]
## 4      15      15 457      2 1600 (400,700] (-1,2]      (1e+03,2e+03] (1e+03,1e+04]
## 5      21      21 454      2 1264 (400,700] (-1,2]      (1e+03,2e+03] (1e+03,1e+04]
## 6      21      15 458      5   806 (400,700] (2,6]      (500,1e+03]      (0,1e+03]
##      slopeK2      sec Turbulence      Entropy clusterom1
## 1      (-1,6]      4-21-21-21-15-15-15-15-15-15-15      4.075664 0.4799569      1
## 2      (-1,6]      21-21-21-21-15-21-15-15-15-21-15      5.899578 0.3845434      1
## 3      (-1,6]      4-21-21-21-21-21-15-15-15-15-15      4.740480 0.5217050      1
## 4      (-1,6]      21-21-21-15-15-15-15-15-15-15-15      3.551410 0.3270264      1
## 5      (-1,6]      21-21-15-15-15-15-15-15-15-21-21      4.022744 0.3658313      1
## 6      (-1,6]      21-21-15-15-15-15-15-15-15-21-15      4.088201 0.3270264      1
##      clusterom2 clusterlcs
## 1              1              1
## 2              1              2
## 3              1              2
## 4              1              1
## 5              1              1
## 6              1              1

## Cluster based on LCP
clusterward_lcp <- hclust(as.dist(dist.lcp),method="ward.D")
plot(clusterward_lcp)

```

## Cluster Dendrogram



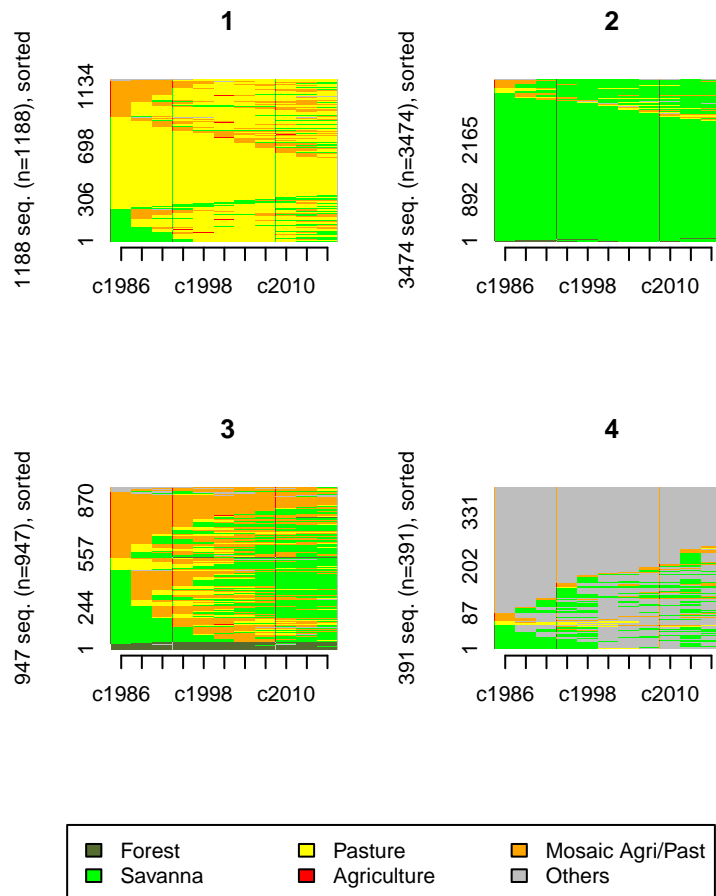
```
as.dist(dist.lcp)
hclust (*, "ward.D")
```

```
cl_lcp <- cutree(clusterward_lcp, k = 4)
tab$clusterlcp <- cl_lcp
head(tab)
```

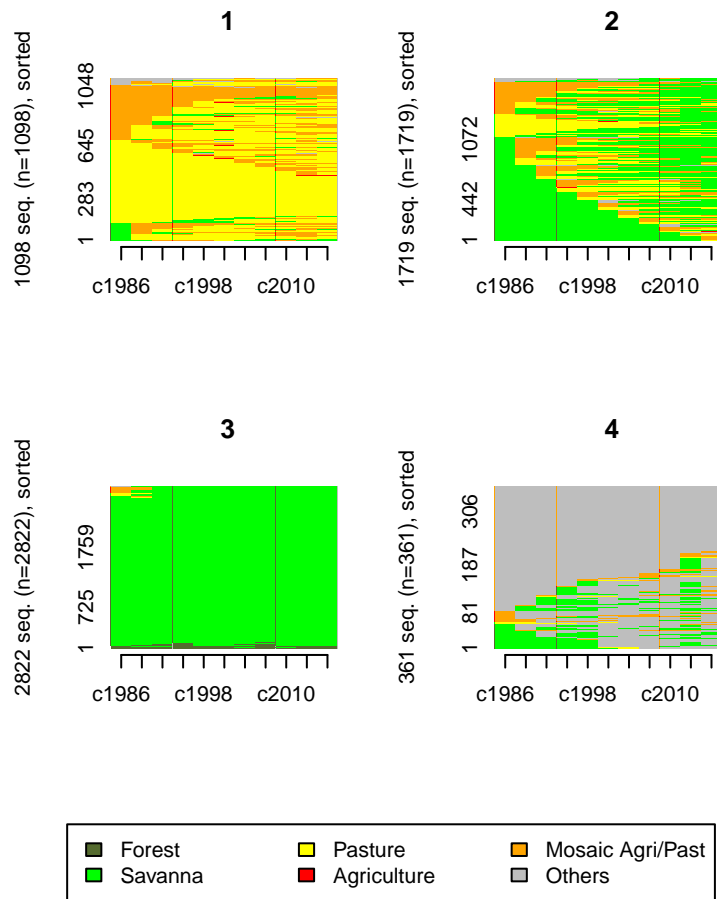
##	x	y	c1986	c1989	c1992	c1995	c1998	c2001	c2004	c2007	c2010
## 1	6120280	9964535	4	21	21	21	15	15	15	15	15
## 2	6120780	9964535	21	21	21	21	15	21	15	15	15
## 3	6121280	9964535	4	21	21	21	21	21	15	15	15
## 4	6121780	9964535	21	21	21	15	15	15	15	15	15
## 5	6122280	9964535	21	21	15	15	15	15	15	15	15
## 6	6122780	9964535	21	21	15	15	15	15	15	15	15
##	c2013	c2016	dem	slope	dist	demK	slopeK	distK	distK2		
## 1	15	15	451	1	100 (400,700]	(-1,2]	(0,500]	(0,1e+03]			
## 2	21	15	452	1	600 (400,700]	(-1,2]	(500,1e+03]	(0,1e+03]			
## 3	15	15	453	1	1100 (400,700]	(-1,2]	(1e+03,2e+03]	(1e+03,1e+04]			
## 4	15	15	457	2	1600 (400,700]	(-1,2]	(1e+03,2e+03]	(1e+03,1e+04]			
## 5	21	21	454	2	1264 (400,700]	(-1,2]	(1e+03,2e+03]	(1e+03,1e+04]			
## 6	21	15	458	5	806 (400,700]	(2,6]	(500,1e+03]	(0,1e+03]			
##	slopeK2	sec	Turbulence	Entropy	clusterom1						
## 1	(-1,6]	4-21-21-21-15-15-15-15-15-15-15	4.075664	0.4799569	1						
## 2	(-1,6]	21-21-21-21-15-21-15-15-15-21-15	5.899578	0.3845434	1						

```
## 3 (-1,6] 4-21-21-21-21-21-15-15-15-15-15 4.740480 0.5217050 1
## 4 (-1,6] 21-21-21-15-15-15-15-15-15-15-15 3.551410 0.3270264 1
## 5 (-1,6] 21-21-15-15-15-15-15-15-15-21-21 4.022744 0.3658313 1
## 6 (-1,6] 21-21-15-15-15-15-15-15-15-21-15 4.088201 0.3270264 1
## clusterom2 clusterlcs clusterlcp
## 1 1 1 1
## 2 1 2 1
## 3 1 2 1
## 4 1 1 1
## 5 1 1 1
## 6 1 1 1

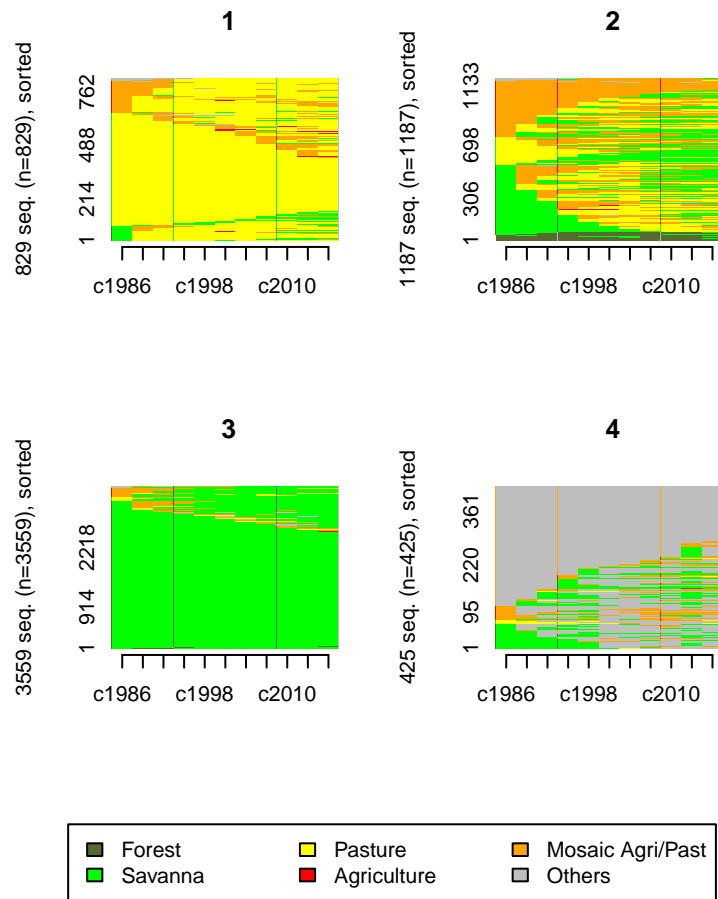
# Plot all the sequences within each cluster para los 4 métodos
# OM1
seqIplot(tab.seq, group = tab$clusterom1, sortv = "from.start")
```



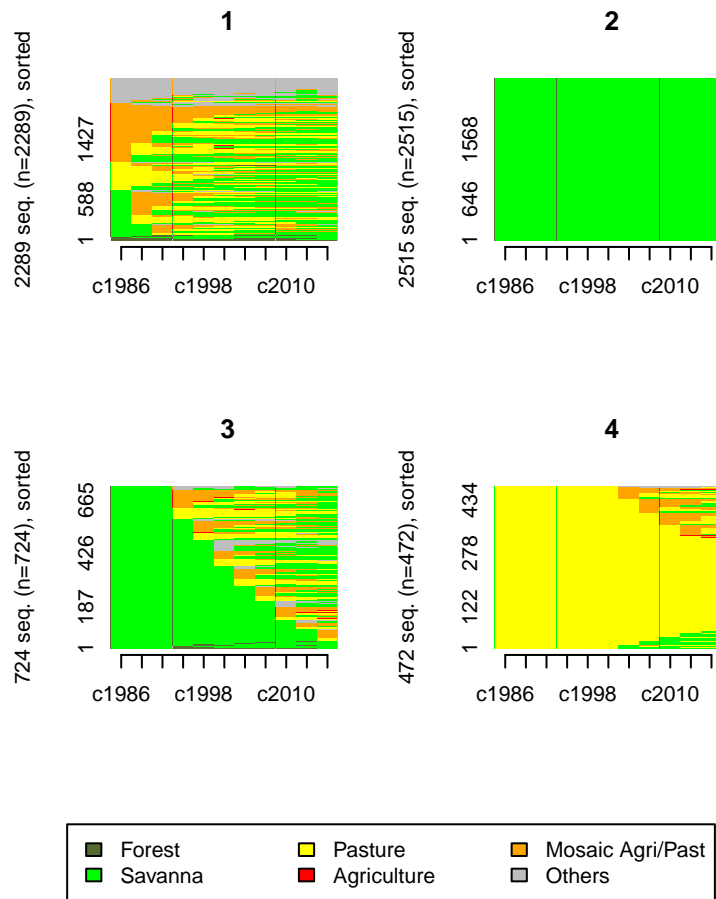
```
# OM2
seqIplot(tab.seq, group = tab$clusterom2, sortv = "from.start")
```



```
# LCS
seqIplot(tab.seq, group = tab$clusterlcs, sortv = "from.start")
```



```
# LCP
seqIplot(tab.seq, group = tab$clusterlcp, sortv = "from.start")
```

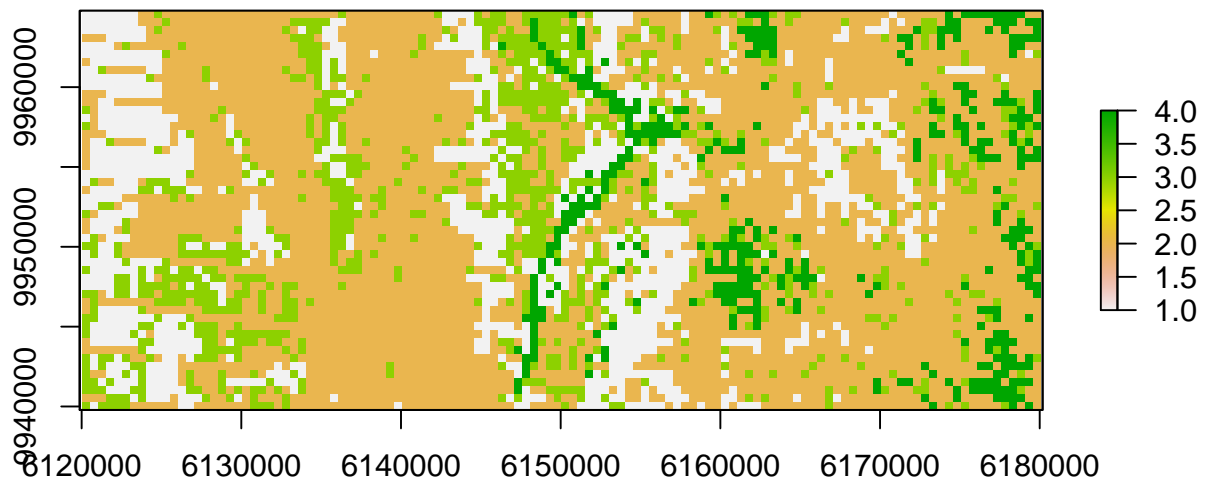


## 4.2 Plotting the clusters

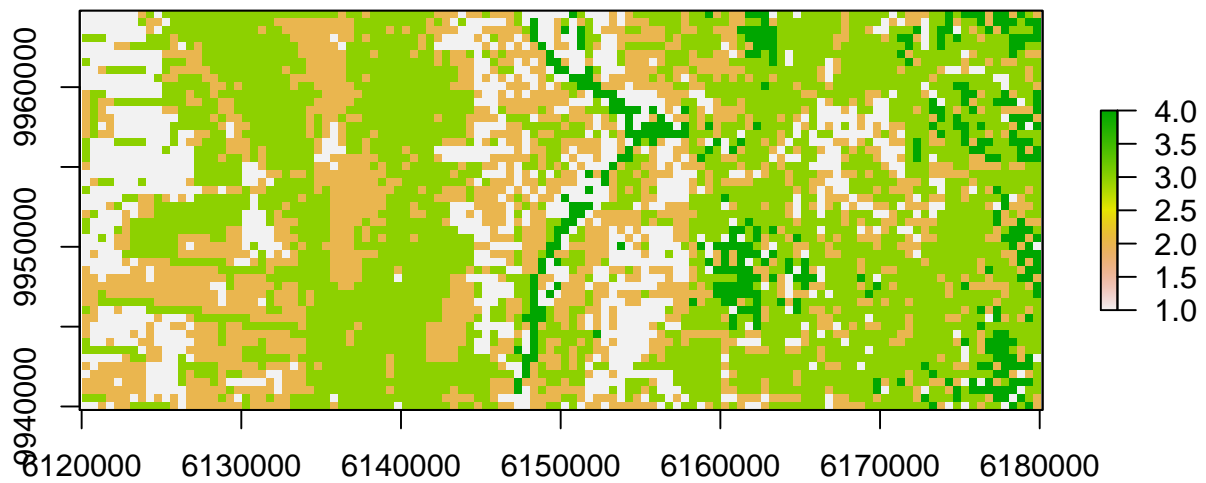
Rasters are created from the tabular data using the coordinates and the cluster number in order to assess the spatial distribution of the clusters.

```
##### Plot clusters' spatial distribution

# Elaborate raster OM1
xyz <- as.data.frame(cbind(tab$x, tab$y, tab$clusterom1))
names(xyz) <- c("x", "y", "z")
xyz <- xyz[complete.cases(xyz), ]
coordinates(xyz) <- ~ x + y
gridded(xyz) <- TRUE
raster_com1 <- raster(xyz)
plot(raster_com1)
```

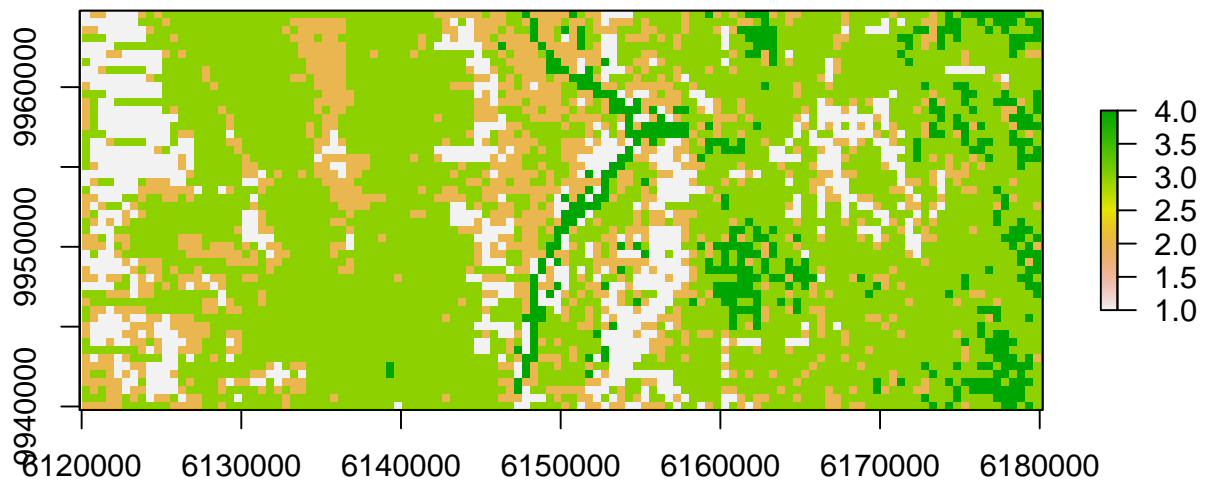


```
# Elaborate raster OM2
xyz <- as.data.frame(cbind(tab$x,tab$y,tab$clusterom2))
names(xyz) <- c("x","y","z")
xyz <- xyz[complete.cases(xyz), ]
coordinates(xyz) <- ~ x + y
gridded(xyz) <- TRUE
raster_com2 <- raster(xyz)
plot(raster_com2)
```

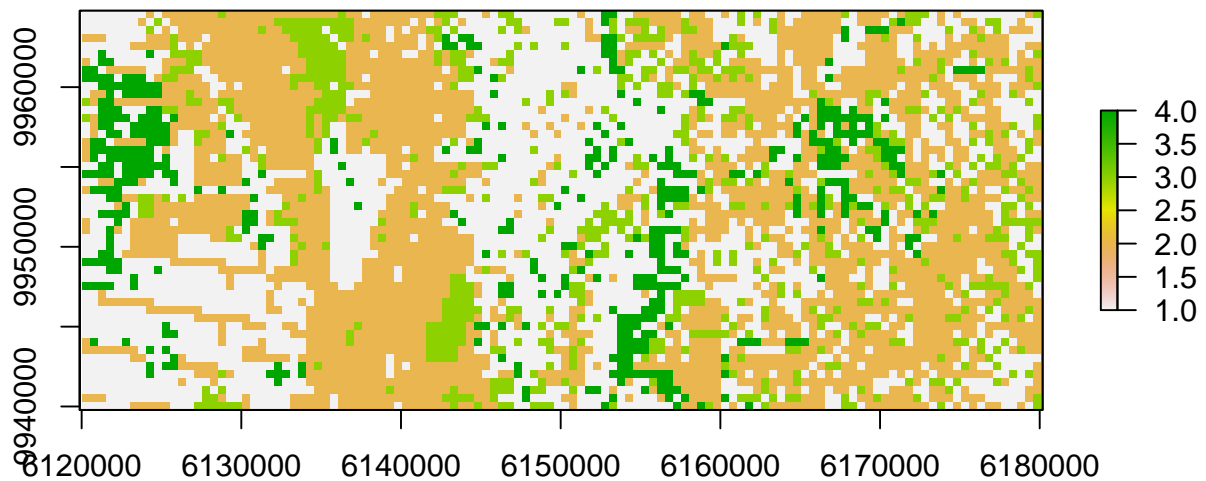


```
# Elaborate raster LCS
xyz <- as.data.frame(cbind(tab$x,tab$y,tab$clusterlcs))
names(xyz) <- c("x","y","z")
xyz <- xyz[complete.cases(xyz), ]
coordinates(xyz) <- ~ x + y
gridded(xyz) <- TRUE
raster_clcs <- raster(xyz)
plot(raster_clcs)
```





```
# Elaborate raster LCP
xyz <- as.data.frame(cbind(tab$x,tab$y,tab$clusterlcp))
names(xyz) <- c("x","y","z")
xyz <- xyz[complete.cases(xyz), ]
coordinates(xyz) <- ~ x + y
gridded(xyz) <- TRUE
raster_clcp <- raster(xyz)
plot(raster_clcp)
```



## 5 Assessing the effect of covariates

Understanding the role of drivers is an important topic in land science. Multi-factor analysis of variance (ANOVA) based on the dissimilarity matrix allows to evaluate the part of discrepancy explained by covariates. The chi-square test enables identifying subsequences that discriminate significantly groups based on covariates. These analysis can be focussed on a subset of sequences which contain specific subsequences.

```
## Run discrepancy analyses to study how sequences are related to covariates

# Compute and test the share of discrepancy explained by different categories
# on covariates
da1 <- dissassoc(dist.om1, group = tab$slopeK, R = 50)
print(da1$stat)

##                t0 p.value
## Pseudo F      290.97565167  0.02
## Pseudo Fbf    292.87606297  0.02
## Pseudo R2      0.08845655  0.02
## Bartlett      162.58157351  0.02
## Levene         255.53957691  0.02

da2 <- dissassoc(dist.om1, group = tab$distK, R = 50)
print(da2$stat)

##                t0 p.value
```

```

## Pseudo F      65.8847395    0.02
## Pseudo Fbf    68.3539356    0.02
## Pseudo R2     0.0421087    0.02
## Bartlett      78.7893707    0.02
## Levene        61.0976289    0.02

da3 <- dissassoc(dist.om1, group = tab$distK2, R = 50)
print(da3$stat)

##                t0 p.value
## Pseudo F      182.78595649    0.02
## Pseudo Fbf    172.82498629    0.02
## Pseudo R2     0.02957325    0.02
## Bartlett      35.10210286    0.02
## Levene        130.17397698    0.02

# Selecting event subsequences:
# The analysis was restricted to sequences that exhibit the state Mosaic

tabe.seq <- seqcreate(tab.seq, use.labels = FALSE)
mosaic <- seqcontain(tabe.seq, event.list = c("Mosaic"))
mosaic_tab <- tab[mosaic,]
mosaic.seq <- tab.seq <- seqdef(mosaic_tab, 3:13, alphabet = alphabet,
                                states = short_labels, cpal = palette, labels = labels)

## [>] state coding:
##      [alphabet] [label] [long label]
##      1          3F      Forest
##      2          4Sav     Savanna
##      3          15Past   Pasture
##      4          18Agri   Agriculture
##      5          21Mosaic Mosaic Agri/Past
##      6          25Others Others
## [>] 834 sequences in the data set
## [>] min/max sequence length: 11/11

mosaic.seq <- seqcreate(mosaic.seq, use.labels = FALSE)

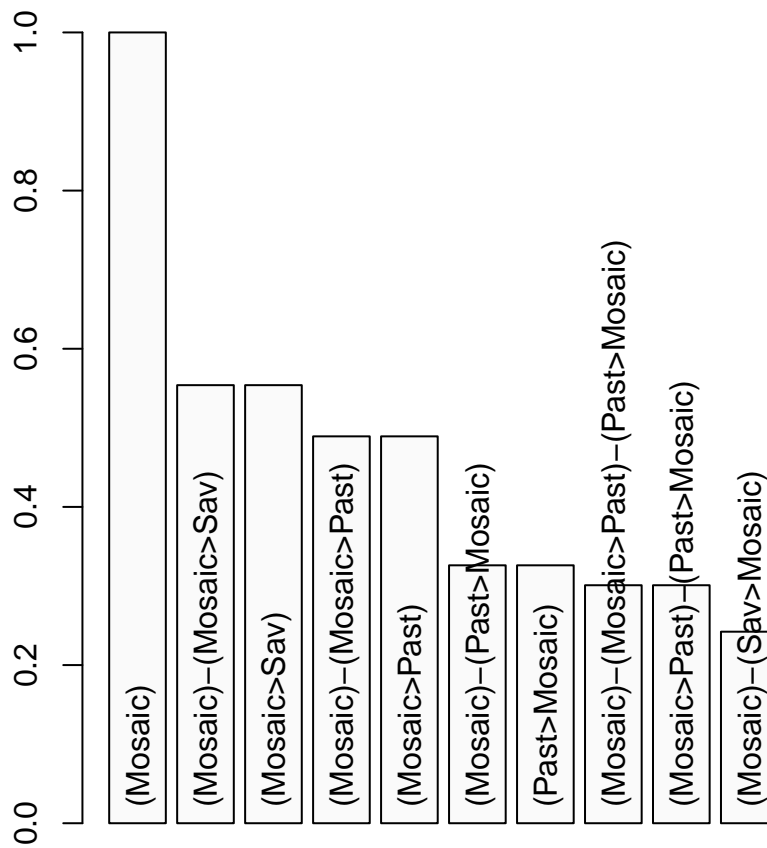
# Look for frequent event subsequences and plot the 10 most frequent ones.
fsubseq <- seqefsub(mosaic.seq, pmin.support = 0.05)
head(fsubseq)

##      Subsequence      Support Count
## 1      (Mosaic) 1.0000000    834
## 2 (Mosaic)-(Mosaic>Sav) 0.5539568    462
## 3      (Mosaic>Sav) 0.5539568    462
## 4 (Mosaic)-(Mosaic>Past) 0.4892086    408
## 5      (Mosaic>Past) 0.4892086    408

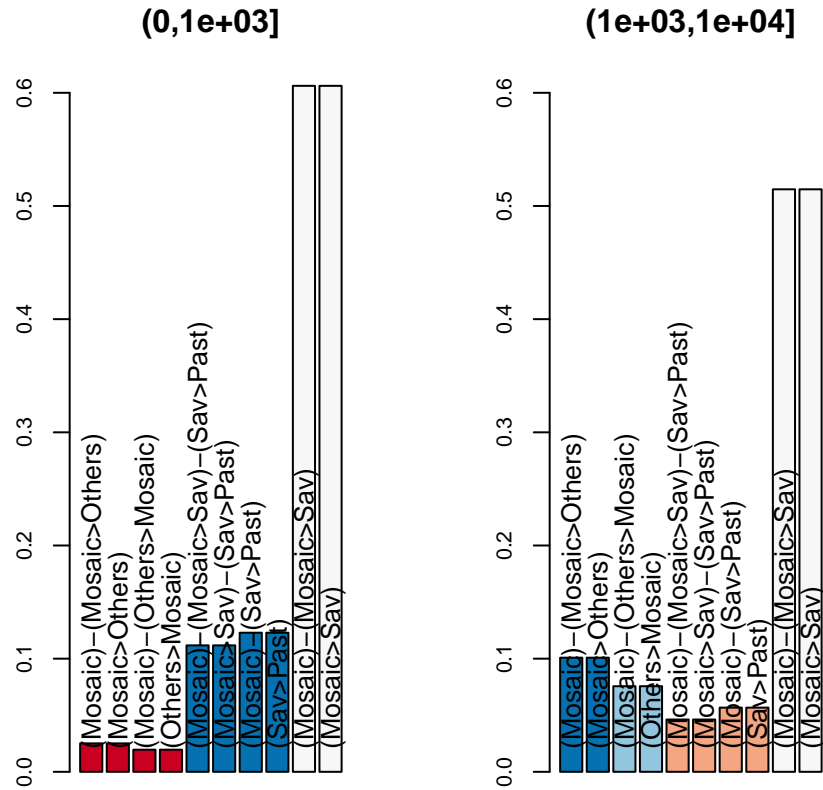
```

```
##
## Computed on 834 event sequences
##   Constraint Value
##   count.method  COBJ

# 10 Most common subsequences
plot(fsubseq[1:10], col = "grey98")
```



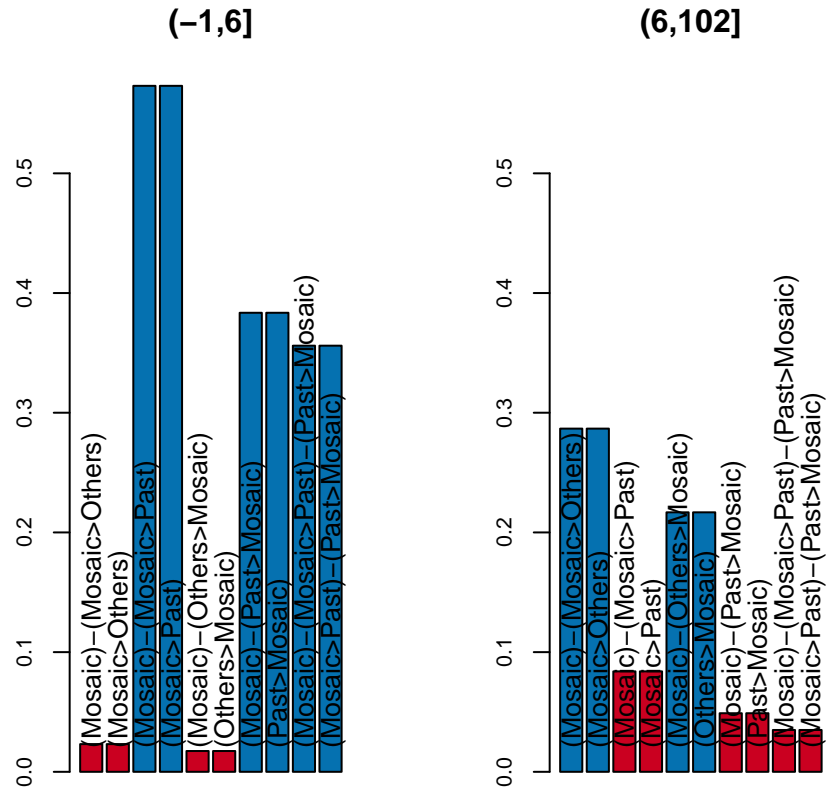
```
# Determine the subsequences of transitions which best discriminate the groups as
# areas close and faraway from roads
discr1 <- seqecmpgroup(fsubseq, group = mosaic_tab$distK2)
plot(discr1[1:10], cex=1, cex.legend=0.8, legend.title="Distance", cex.lab=0.8,
     cex.axis = 0.8)
```



**Distance**

■ Negative 0.01 ■ Negative 0.05 □ neutral ■ Positive 0.05 ■ Positive 0.01

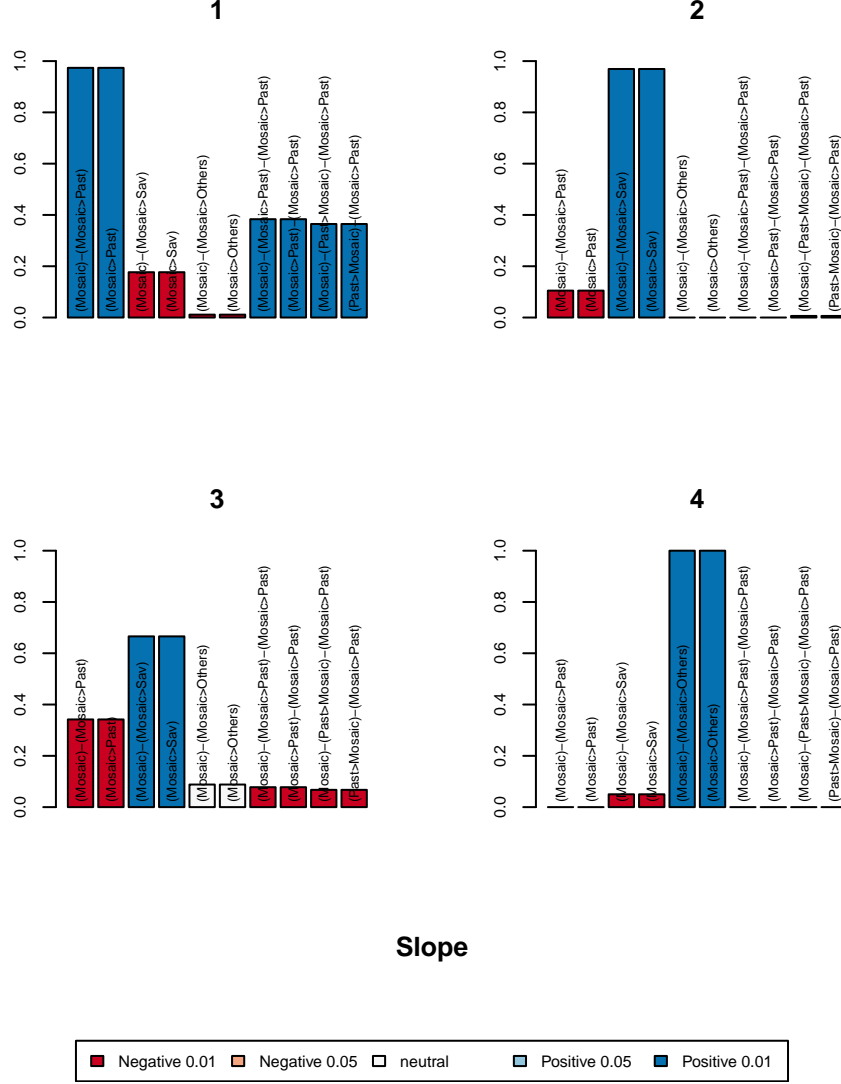
```
# areas with moderate vs steep slope
discr2 <- sequecmpgroup(fsubseq, group = mosaic_tab$slopeK2)
plot(discr2[1:10], cex=1, cex.legend=0.8, legend.title="Slope", cex.lab=0.8,
     cex.axis = 0.8)
```



## Slope

■ Negative 0.01 
 ■ Negative 0.05 
 □ neutral 
 ■ Positive 0.05 
 ■ Positive 0.01

```
# Clusters
discr3 <- seqecmpgroup(fsubseq, group = mosaic_tab$clusterom1)
plot(discr3[1:10], cex=0.6, cex.legend=0.8, legend.title="Slope", cex.lab=0.8,
     cex.axis = 0.8)
```



In this last example, the chi-square test identifies subsequences that discriminate significantly the sequences of each group obtained from the cluster analysis.

## 6 Acknowledgments

This material was elaborated with the support of the *Programa de Apoyo a Proyectos para la Innovación y Mejoramiento de la Enseñanza* - PAPIME-DGAPA at the *Universidad Nacional Autónoma de México* (Project PE117519 *Herramientas para la enseñanza de la Geomática con programas de código abierto*). I thank the authors of the package **TraMineR** for developing and sharing this excellent and useful program.

# Bibliography

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- Gabadinho, A., Ritschard, G., Müller, N., and Studer, M. (2011). Analyzing and Visualizing State Sequences in R with TraMineR. *Journal of Statistical Software, Articles*, 40(4):1–37.
- Studer, M. and Ritschard, G. (2016). What matters in differences between life trajectories: A comparative review of sequence dissimilarity measures. *Journal of the Royal Statistical Society: Series A (Statistics in Society)*, 179:481–511.