

# Supplementary Materials: Bioelectrochemical Changes during the Early Stages of Chalcopyrite Interaction with *Acidithiobacillus Thiooxidans* and *Leptospirillum* sp.

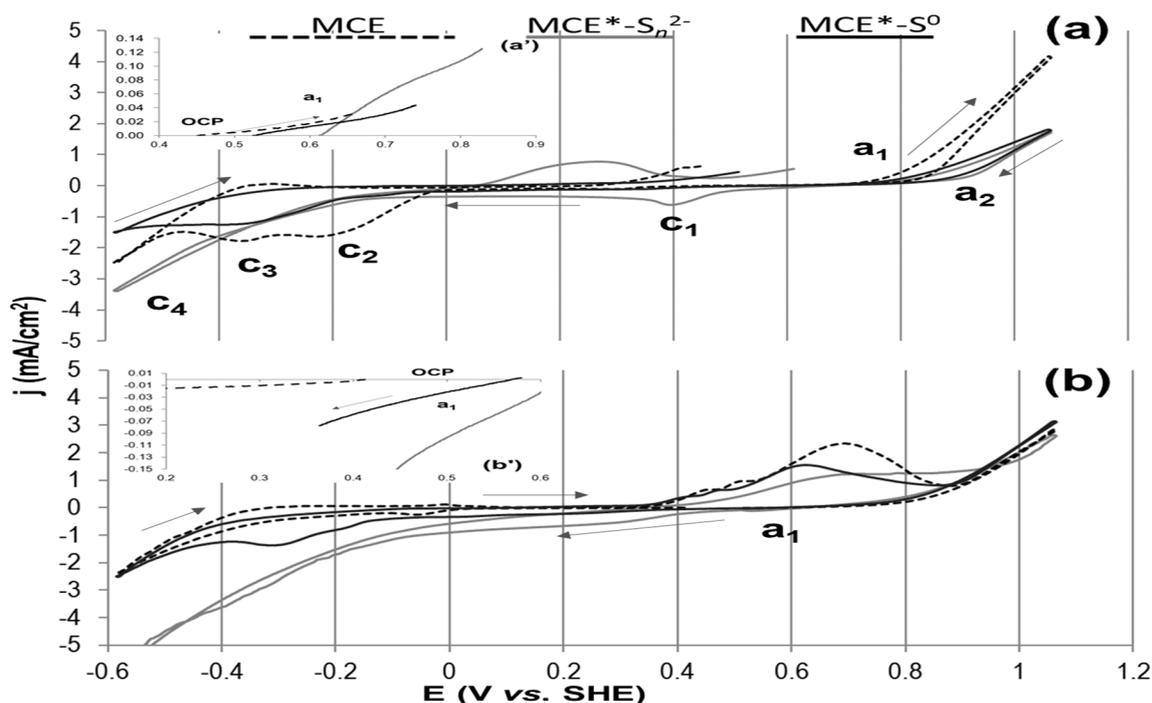
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## 1. Potentiostatic modification of massive chalcopyrite electrodes (MCE)

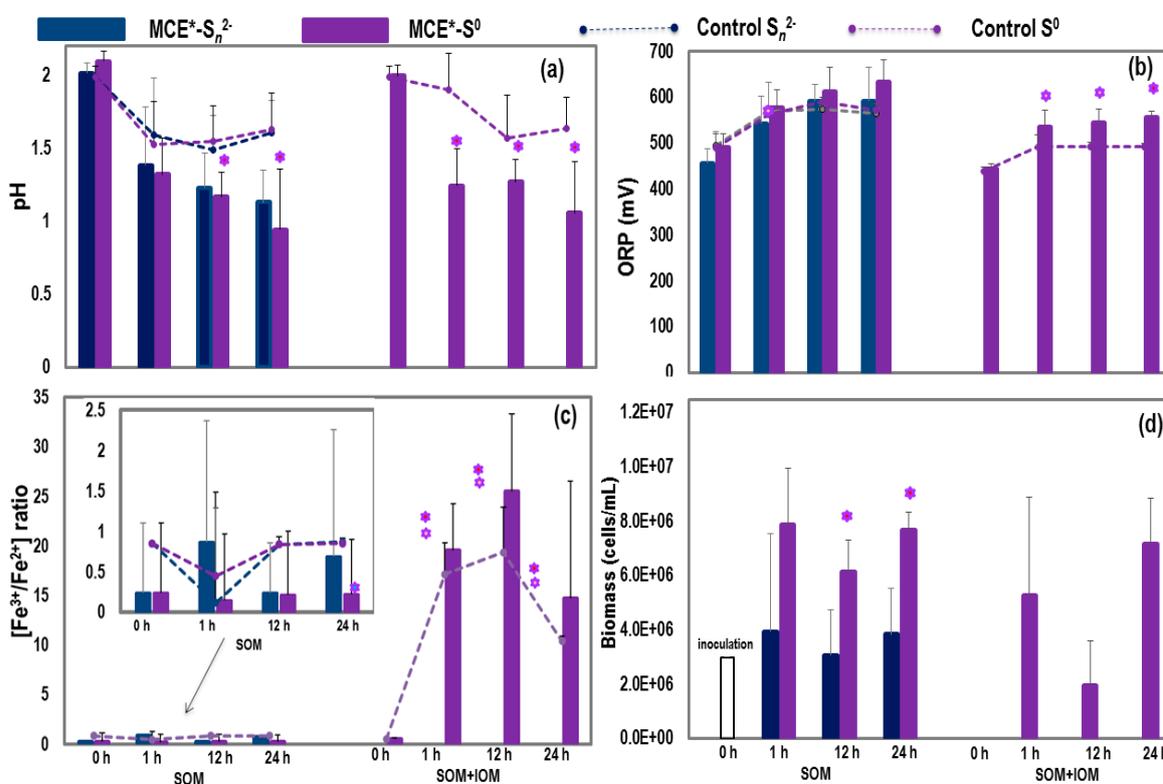
Mineral coupons of 1.0 cm<sup>2</sup> were coated with Cu via electrolytic deposit using a CuSO<sub>4</sub> solution, to improve the current distribution; a Cu wire was added with a silver solder to enhance the electrical contact of the MCE. Finally, the electrode was imbibed in epoxy resin and the exposed MCE surface was polished with a water sandpaper until it reached a mirror-like surface condition. The MCE were maintained in desiccators under anaerobic conditions until their use.

The potentiostatic modification was conducted with an EPSILON BASi 2.10.73 potentiostat (Indiana, USA), at 25°C, in a typical electrochemical, three-electrodes device; the MCE was used as working electrode, the reference electrode was a saturated sulfate electrode, Hg/Hg<sub>2</sub>SO<sub>4</sub> (0.615 V vs. SHE, the standard hydrogen electrode), and a graphite rod (Alfa Aesar, Massachusetts, USA, 99.9995% purity) was used as counter-electrode. These MCE\* were then achieved by application of anodic pulse (E<sub>an</sub>, 3600 s); ATCC-125 culture at pH 2.0 was the electrolyte, hence emerging S<sup>0</sup> and S<sub>n</sub><sup>2-</sup> compounds (reduced sulfur species, RSS), as a function of the applied E<sub>an</sub>, from 0.36 to 1.015 V vs. SHE. The Raman spectra were recorded with a triple subtractive monochromator (T64000 Jobin Yvon spectrometer, Kyoto, Japan) equipped with a confocal microscope, Olympus BH2-UMA (λ= 514 nm). At least 10 spectra were recorded for each MCE\* surface. Calibration was done using a Si wafer, which showed a single peak at 521 cm<sup>-1</sup>. The noise/signal ratio was better than 100.

After the CV and Raman analysis of the MCE, anodic E<sub>an</sub> of 695 mV and 915 mV were chosen to electrogenerated two different RSS, since at these E<sub>an</sub> was observed minor electrooxidation and low activation current (anodic peak a<sub>1</sub> at the open circuit potential, OCP; Figure. S1a and b). Raman peaks for these MCE\* indicate the predominance of S<sub>n</sub><sup>2-</sup> and heptagonal sulfur S<sub>7</sub> for E<sub>an</sub> < 695 mV; such electrode was referred as MCE\*-S<sub>n</sub><sup>2-</sup>. Octagonal sulfur S<sub>8</sub> was detected at E<sub>an</sub> > 915 mV, for MCE\*-S<sup>0</sup>. The reactivity j<sub>act</sub> and of the MCE\*-S<sub>n</sub><sup>2-</sup> was significantly lower than of the MCE\*-S<sup>0</sup> (Fig. S1 c and d); thus, more energy is necessary to (bio)oxidize the MCE\*-S<sup>0</sup> than for MCE\*-S<sub>n</sub><sup>2-</sup> surface.



**Figure S1.** Voltammograms in positive (a) and negative (b) potential sweep of pristine MCE (dotted line), and electrooxidized EMC at 695 mV, MCE\*-S<sub>n</sub><sup>2-</sup> (gray line), and at 915 mV, MCE\*-S<sup>0</sup> (black line), in ATCC-125 medium pH 2. Scan rate: 20 mV/s with stirring; the potential scan was initiated at the OCP.



**Figure S2.** Certain characteristics of the media as electrolyte (after 1, 12 and 24 h of the exposure of MCE\*-S<sub>n</sub><sup>2-</sup> (blue) and MCE\*-S<sup>0</sup> (purple) to SOM or SOM + IOM media (abiotic controls; dotted lines) and cultures (biotic; columns): pH (a); ORP (b), soluble Fe<sup>3+</sup>/Fe<sup>2+</sup> (c), and biomass of non-attached bacteria (d). Data: Average values (n = 3) and standard deviation (error bars). \*: Values significantly different from both, controls and surficial SRS (–S<sub>n</sub><sup>2-</sup> or –S<sup>0</sup>).