

Article

Mineralogical and Elemental Composition of *Pectinatella magnifica* and Its Statoblasts

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Abstract: Several massive colonies of *Pectinatella magnifica* have been observed during the summer almost every year since 1974 in agricultural reservoir ponds and lakes with dirty freshwater environments in Ishikawa, Japan, which has posed serious environmental problems on the shores of Hokuriku District. We collected *Pectinatella magnifica* during the summer at Kahokugata Lake and Makiyama agricultural reservoir pond in June and July 2016. However, scientific data for the statoblasts of *Pectinatella magnifica* are limited. Our results for scanning electron microscopy equipped with energy-dispersive spectroscopy (SEM-EDS), inductively coupled plasma-mass spectrometry (ICP-MS), and X-ray powder diffraction (XRD) analyses of *Pectinatella magnifica* indicated immobilization of the chemical elements that were involved in the mass during the summer. We also reported the characterization of an invasive species of bryozoan (*Pectinatella magnifica*) in lakes and ponds in Ishikawa, Japan, based on field observations in 2016. We studied the microstructure, mineralogy, chemical composition, and radioactivity associated with these organisms, using a combination of micro-techniques, SEM-EDS, associated with ICP-MS, and XRD. This study aims to illustrate the capability of *Pectinatella magnifica* to produce minerals within statoblasts and gelatinous material. Obtained results may indicate forming quartz, palygorskite, dolomite, bischofite, pyrolusite, and pyrite, associated with native sulfur and copper in the statoblast. The mass of gelatinous material contains talc and vermiculite as well as non-crystalline phase. The mechanism of biomineral formation has important implications for water–mineral–organism or microorganism interactions both in lower drainage basin systems, such as Kahokugata Lake, and upper water areas, such as Makiyama agricultural reservoir pond. Many types with variety of sizes and shapes of bryozoan (*Pectinatella magnifica*) were found in lakes and ponds in Japan. The biomineralization systems will be made available for use not only in researching bryozoans (*Pectinatella magnifica*), but also for environmental change systems from upstream to downstream of the lake. To date, there have been no reports on related electron microscopy observations, including the real-life occurrence of “bioremediation”. These observations could lead to simple methods of removing statoblasts of the

invasive alien species *Pectinatella magnifica* from agricultural and reservoir environments, because there was limited microbial immobilization of the ions during the winter.

Keywords: agriculture reservoir; lake; bryozoa; Phylactolaemata; invasive alien species *Pectinatella magnifica*; statoblast; gelatinous material; SEM-EDS; ICP-MS; XRD; biomineralization; palygorskite; pyrite; dolomite

1. Introduction

The distribution of *Pectinatella magnifica* has been restricted to eastern North America and central Europe. *Pectinatella magnifica* is also abundant on the Korean peninsula, northeastern China, and Japan. Several gelatinous colonial masses of *P. magnifica* appeared in the water near the shore of Lake Kawaguchi, at the foot of Mt. Fuji, Yamanashi Prefecture, Japan, in the autumn of 1972 [1]. Shuzitu Oda found giant colonial masses occurred in Lake Shoji, Yamanashi Prefecture, Japan, in 1973 [2]. Several colonial masses of *Pectinatella magnifica* appeared in Lake Kiba near Lake Shibayama, Ishikawa Prefecture, Japan, in 1974 and 1975 [2,3].

Since 1973, several massive colonies of *Pectinatella magnifica* have appeared every year in Lake Shoji, one of the five lakes at the foot of Mt. Fuji, in central Japan. In connection with a study on the life cycle of *Pectinatella magnifica*, Dr. Oda was able to obtain a cluster of floating statoblasts from this lake in early summer, and he distinguished six species of freshwater bryozoans among these statoblasts [4]. He reported the biological significance of these statoblasts and the distribution of *Pectinatella magnifica* in Japan. Every year in spring, a large number of hibernating statoblasts of *Pectinatella magnifica* float in clusters on the surface of Lake Shoji, though they are frozen in winter. By chance, he collected a small cluster of hibernating statoblasts from this lake on 29 June 1976. At that time, the water temperature was 21.6 °C. The majority of the statoblasts of *Pectinatella magnifica*, which was the main species in this cluster, had germinated. However, the valves of these statoblasts had opened and the germinal materials were lost, and only a few polypides appeared between the valves. The statoblasts of other species had not yet germinated [5]. Other species, such as *Plumatella repens*, *Plumatella emarginata*, and *Stephanella hina*, are also common in Japan.

Bryozoans comprise a phylum of sessile, colonial animals that inhabit marine and freshwater environments. Only around 90 species from fresh habitats have been described, most belonging to the class Phylactolaemata. Phylactolaemata can propagate asexually by forming encapsulated dormant bodies known as statoblasts, which have characteristics that are highly useful in their taxonomy and identification. To date, 23 species of Phylactolaemata in 10 genera have been reported in Japan; however, no taxonomical reviews on these have been published. Furthermore, the current taxonomy of Phylactolaemata depends heavily on the observation of statoblast microstructures using scanning electron microscopy, which poses difficulties associated with simple identification [6,7]. *Pectinatella magnifica* has absolutely distinctive statoblast and colony morphology. Therefore, statoblasts are useful for identification of plumatellid bryozoans.

On the other hand, the minerals and their chemical composition in bryozoans are very important for understanding the conditions of the bio-mineralization process. Therefore, there are many previous works on biomineralization of bryozoans [8,9]. Taylor et al. [9] reviewed biomineralization in bryozoans. In particular, they noted that the Mg/Ca ratio in Mg-bearing calcite, calcite/aragonite ratio, and temperature are important for understanding the location of mineral formation. To investigate these characters, they used the X-ray diffraction method. However, they did not review other minerals in bryozoans.

Various types of minerals were present in the statoblasts and gelatinous material. The chemical data of *Pectinatella* “gelatinous parts” (not really agar) were described with no reference to the single previous analysis of this gelatinous material [10]. There have been no reports of the mineralogical and

elemental composition of *Pectinatella magnifica* and its statoblasts on the surface and inside gelatinous material of *Pectinatella magnifica* to date.

In this study, the mineralogy and chemistry of colonies and statoblasts of the freshwater species *Pectinatella magnifica* in Kanazawa, Ishikawa, Japan, are investigated by inductively coupled plasma-mass spectrometry (ICP-MS) and X-ray powder diffraction (XRD) techniques. Furthermore, the characteristics of colony and statoblast micro morphology and chemistry were also determined for *Pectinatella magnifica* using electron microscopy with an energy-dispersive microanalyzer (SEM-EDS).

2. Materials and Methods

2.1. Exploring *Pectinatella magnifica* Field and Investigated Specimens

The freshwater Magnificent Bryozoan, *Pectinatella magnifica*, a confirmed native invertebrate of North America, was found in Ishikawa prefecture, Japan. This was visually confirmed on the surface of the water at four spots in ponds and lakes in Ishikawa prefecture, and samples were obtained from four locations (Figure 1), Shibayama Gata Lagoon (Figure 2a,b), Kahokugata Lake (Figure 2c,d), Makiyama agricultural reservoir pond (Figure 2e–h), and Tawara Pond [11] in Ishikawa prefecture, Japan, in July 2015 and 2016; as a result, the distribution of *Pectinatella magnifica* was confirmed and attracted attention. However, until now, there have been no reports of its chemical, physical, biological, and mineralogical composition.

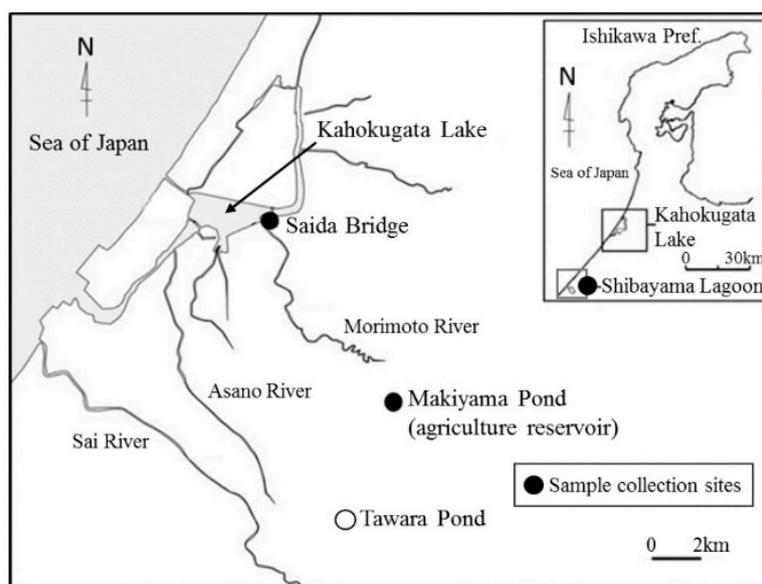


Figure 1. Location map of the study area where samples were collected in Ishikawa Prefecture, Japan.

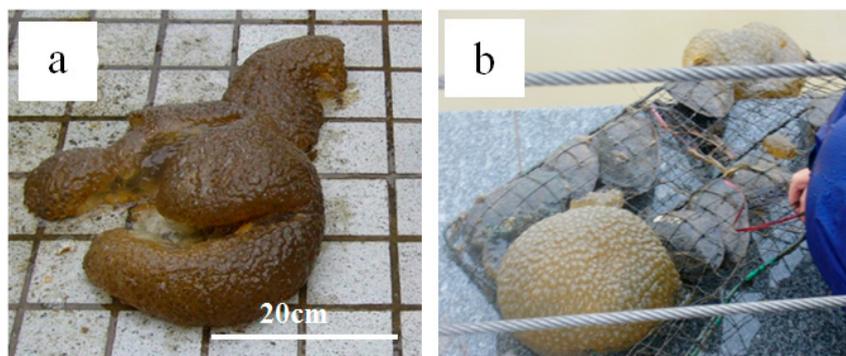


Figure 2. Cont.

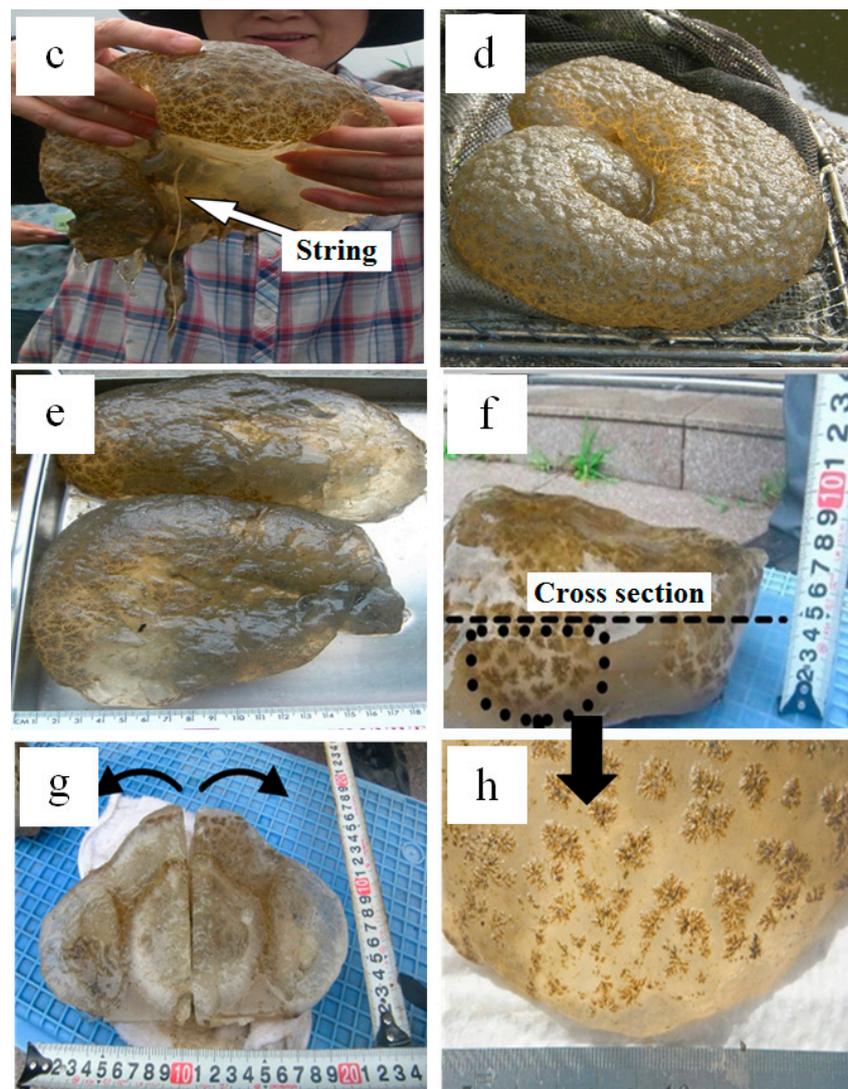


Figure 2. Colonies of *Pectinatella magnifica* appearing during the summer season in polluted freshwater environments at Shibayama Lagoon (a,b); Kahokugata Lake (c,d); and Makiyama agricultural reservoir pond (e) in Ishikawa Prefecture, Japan. The frozen *Pectinatella magnifica* collected from Makiyama agricultural reservoirs (f); a normal pattern of zooid clusters (g); and statoblast pattern on the surface (h).

Water characteristics such as pH, redox potential, electrical conductivity, dissolved oxygen concentration, and water temperature were measured at Makiyama agricultural reservoir pond and Kahokugata Lake during the summer of 2016 (Table 1). Furthermore, the radioactivity of water samples collected from Kahokugata Lake and Makiyama agricultural reservoir pond was recorded by measuring $\beta(\gamma)$ rays for dried statoblasts by an Aloka GM survey meter TGS-136 (Mitaka, Japan) with a Geiger–Müller (GM) tube (Table 1).

The samples were washed in distilled and deionized (DD) water and dried under the sunshine for a few days during the summer.

Table 1. Characteristics of water in Kahokugata Lake and Makiyama Pond in 2016. EC: electrical conductivity; DO: dissolved oxygen; BG: Back ground.

	Kahokugata Lake	Makiyama Agriculture Reservoirs Pond
Date	27 June, 14 July in 2016	18 June, 1 July in 2016
pH	6.8–7.7–9.0	6.8–7.5
Water Temp.	26–27 °C	33–36 °C
EC	12 µS/cm	12 µS/cm
DO	6.8–7.7 mg/L	3.0–4.0 mg/L
Water Color	light brown	light brown
Radio Activity	110–130 cpm (BG 80–100cpm)	110–120 cpm (BG 80–100 cpm)
Size of	30–40–60 cm	20–50 cm
<i>Pectinatella magnifica</i>	200 g, 800 g, 14 kg	500 g, 1 kg, 5 kg

The pH of the water was 6.8–7.5, the water temperature was 26–27 °C, the humidity was 48–56%, and the air temperature was 33–36 °C at noon on 18 June 2016 at Makiyama agricultural reservoir pond in Kanazawa. The dissolved oxygen (DO) was quite low at 3–4 mg/L compared with normal values (7–8 mg/L). We collected several *Pectinatella magnifica* samples from Makiyama agricultural pond (Figure 2e–h) and Kahokugata Lake (Figure 2c,d) to determine the major characteristics of the colonies and statoblasts. We observed their size and form and identified them as the species *Pectinatella magnifica*, characterizing their structure and properties based on the methods of Hirose [6,7,12] and Miyashita et al. [13]. Views of a cross section of frozen *Pectinatella magnifica* in Makiyama agricultural reservoir pond show two colonies 10 cm in diameter, with star patterns on their surfaces (Figure 2f–h with an arrow mark).

The frozen *Pectinatella magnifica* collected from Makiyama agricultural reservoir pond (f), a cross section of two colonies (g), and a statoblast pattern on the surface are shown (h) in Figure 2.

The gelatinous material varies widely in weight from 0.2 to 14 kg. Their radioactivity was 110–130 cpm (Back ground 80–100 cpm) in Kahokugata Lake during summer 2017. The water quality characteristics were pH 7.6–7.7, transparency 30–42 cm, water temperature 27 °C, electrical conductivity (EC) 12 µS/cm, and DO 6.8–7.7 mg/L for yellow brownish water on 7 and 14 July 2016.

We observed statoblast samples from Makiyama agricultural reservoir pond and Kahokugata Lake with an optical microscope (Nikon NTF2A, Tokyo, Japan) for the identification of *Pectinatella magnifica*.

2.2. ICP-MS Analyses

Semi-quantitative analyses of statoblast samples of *Pectinatella magnifica* collected from Kahokugata Lake were conducted using a Thermo Fisher Scientific (Bremen, Germany) inductively coupled argon plasma emission spectrometer (iCAP), ICP-MS Elementar Analysensysteme GmbH Vario MAX type instrument (ICP-MS) (Langenselbold, Germany).

2.3. Scanning Electron Microscopy (SEM) with Energy-Dispersive Spectroscopy (EDS)

A scanning electron microscope (S-3400N, Hitachi, Tokyo, Japan) equipped with an energy-dispersive X-ray analyzer (Horiba EMAX, Kyoto, Japan) was used to study the *Pectinatella magnifica* and *Asajirellar gelatinosa* samples collected from Makiyama agricultural reservoir pond and Kahokugata Lake. We attempted to analyze the appearance, distribution, tissue structure, and inorganic and organic constituents of the statoblasts, evaluating the microscale morphology, associated with elemental distribution maps. The operating conditions used were as follows: 15 kV accelerating voltage, 70–80 µA current, an analytical time of 1000 s, and an area of 10 mm × 10 mm on carbon double-sided tape with a Pt coating.

We also observed statoblast samples collected from Makiyama agricultural reservoir pond and performed observation and analysis at an acceleration voltage of 10 kV, in a low vacuum mode using a JEOL 6010 PLUS/LA (Tokyo, Japan) scanning electron microscope and an energy analysis of variance tools.

2.4. X-Ray Powder Diffraction Measurements

In this study, we focused on the mineralogical characterization of statoblasts and the gelatinous part of *Pectinatella magnifica*. Therefore, to identify minerals and analyze the structure of crystals and the crystallinity of the statoblasts and gelatinous material structure in statoblasts collected from Makiyama agricultural reservoir pond, we carried out X-ray powder diffraction measurements (XRD) under ambient conditions (Shimazu, Kyoto, Japan, XRD-6100; Cu-K α radiation, 40 kV, 40 mA, scan speed of 2°(2 θ)/min, scan range 2 θ = 5–60°). Three statoblast powder samples were prepared by placing them on a glass filter with a Zn plate and two samples of the gelatinous part of *Pectinatella magnifica* painted directly on the sample holder were used for measurements.

2.5. X-Ray Fluorescence Analyses (XRF) for Deposited Minerals on Makiyama Pond

For comparing XRD results, the chemical compositions of the sediment samples near the Makiyama agricultural reservoir pond were also analyzed by the X-ray fluorescence analysis (XRF) technique using a XRF analyzer (Rigaku Promus II, Tokyo, Japan), which operated at an acceleration voltage of 60 kV, current of 150 mA under vacuum conditions. The sediment samples were heated to 600 °C for 30 min using an electric furnace and were pressed for XRF measurements.

3. Results

3.1. Optical Microscopy Observation and Identification of Species

We observed and analyzed statoblast samples collected from Makiyama agricultural reservoir pond using an optical microscope and SEM (Figure 3) and identified *Pectinatella magnifica* according to the number of statoblasts in the collected cluster [11]. Most of the statoblasts collected from Makiyama agricultural reservoir pond could be identified as *Pectinatella magnifica* according to their statoblasts, which have 11–22 anchors around them. Some statoblasts have 11–15 anchors. On the other hand, a few oval statoblasts with a diameter of 0.5 mm can be seen. A few reddish *Cristatella mucedo* were also found in Makiyama agricultural reservoir pond.

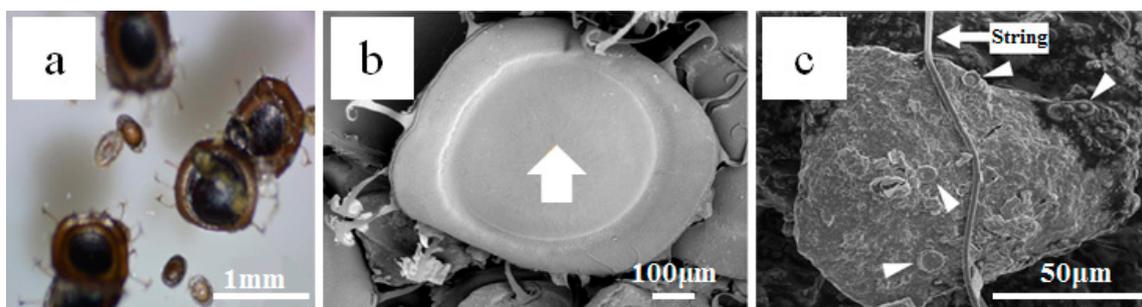


Figure 3. Optical micrographs of the statoblasts of *Pectinatella magnifica* (a) collected from Makiyama agricultural reservoir pond. Scanning electron microscope (SEM) micrograph of the statoblasts of *P. magnifica* (b) collected from Makiyama agricultural reservoir pond. SEM micrographs of massive colonies of *Pectinatella magnifica*, showing a long adherent string ((c), an arrow) and four spherical *Pectinatella magnifica* on the surface of gelatinous material ((c), four arrows).

3.2. ICP-MS Analyses of *Pectinatella magnifica*

Semi-quantitative ICP-MS results for dried colonies of *Pectinatella magnifica* collected from Kahokugata Lake are shown in Table 2. *Pectinatella magnifica* contained many kinds of elements; the main constituents were Fe (290 mg/L) and Al (204 mg/L), associated with S (53.7 mg/L), Mg (46.5 mg/L), Ca (45.6 mg/L), P (26.2 mg/L), K (22.4 mg/L), and Na (13.1 mg/L) with traces of Ti

(5.87 mg/L), Zn (1.58 mg/L), and Cu (0.15 mg/L) (Table 2). Traces of Sr (0.11 mg/L) can also be seen. Si cannot be measured by this method.

Table 2. ICP-MS semi-quantitative analytical results for dried colonies of *Pectinatella magnifica* collected from Kahokugata Lake. (The unit mg/L means the weight of the element in one liter of wet sample.)

Atomic Number	Elements	mg/L
5	B	0.046
11	Na	13.100
12	Mg	46.500
13	Al	204.000
14	Si	-
15	P	26.200
16	Si	53.700
17	Cl	1.400
19	K	22.400
20	Ca	45.600
22	Ti	5.870
23	V	0.630
24	Cr	0.320
25	Mn	3.980
26	Fe	290.000
27	Co	0.100
28	Ni	0.170
29	Cu	0.150
30	Zn	1.580
33	As	0.100
37	Rb	0.048
38	Sr	0.110
39	Y	0.084
40	Zr	ND
48	Cd	0.004
55	Ce	0.540
56	Ba	0.470
80	Hg	ND
82	Pb	0.150

-.: not measured; ND: not detected.

3.3. SEM Micrograph

SEM micrograph of a statoblast sample collected from Makiyama agricultural reservoir pond shows in Figure 3b. *Pectinatella magnifica* from Makiyama can be seen with a 1 mm diameter and more than 10 anchors around the rim.

3.4. SEM Observation and Energy-Dispersive Spectroscopic (EDS) Analyses

Figure 4 shows clusters and statoblasts from Makiyama agricultural reservoir pond. Clusters with various shapes, sizes (5–10–500 μm in diameter), and numbers of statoblasts were collected from Kahokugata Lake, whereas uniform sizes (900 μm in diameter) and simple round shapes with anchors were collected from Makiyama. The samples were observed by SEM-EDS. Various types of bryozoan statoblasts lived in Kahokugata Lake, whereas only one type of extraneous with anchor type of *Pectinatella magnifica* lived in Makiyama agricultural reservoir pond.

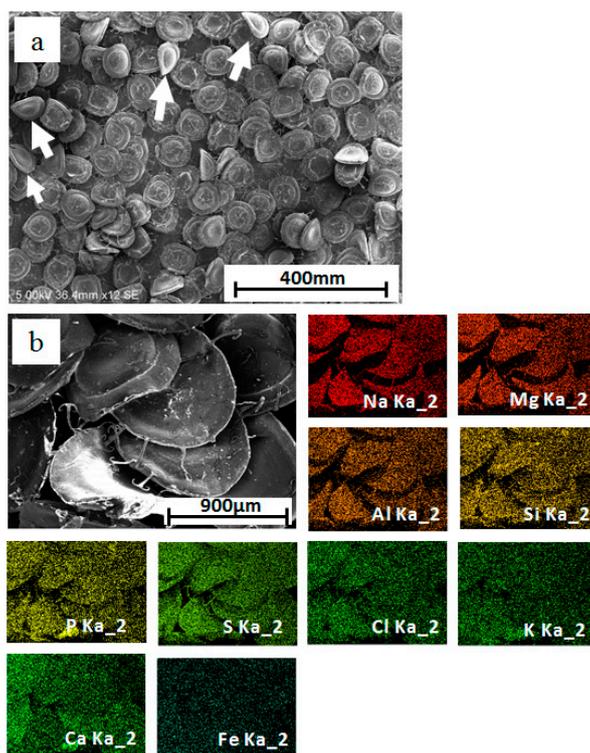


Figure 4. SEM micrographs of gathered statoblasts of *Pectinatella magnifica* (a) and the enlarged elemental distribution maps (b). The samples were collected from Makiyama agricultural reservoir Pond.

Semi-quantitative analysis data of whole gelatinous material or statoblasts collected from Kahokugata Lake and Makiyama agricultural reservoir pond are shown in Tables 3 and 4. Values are represented as mass %. The whole gelatinous material and statoblasts from Kahokugata Lake contained high Si (46–56%), associated with Al (7–15%) and Cu (7–14%) (Table 3). On the other hand, high concentrations of S (37–54%), Ca (12–15%), Si (6–10%), and traces of Sr (0.6%) can be seen in the Makiyama agricultural reservoir pond (Table 4). Statoblasts of *Pectinatella magnifica* from Makiyama agricultural reservoir pond showed richness in Si, S, and Ca, associated with traces of Sr, which indicates that only one anchor type of *Pectinatella magnifica* is present in Makiyama agricultural reservoir pond (Table 4 Points 1, 2, 3).

Table 3. Results of SEM-EDS semi-quantitative analyses of dried colonies of *Pectinatella magnifica* collected from Kahokugata Lake in July and September 2016. The unit is mass %.

Elements	1	2	3	4
Mg	1.60	1.22	2.09	3.45
Al	10.49	7.47	12.67	15.34
Si	46.05	56.48	48.67	47.03
P	3.52	3.18	3.54	3.39
S	7.98	7.92	7.06	3.16
Cl	0.43	0.31	0.68	0.18
K	1.14	0.26	1.24	1.31
Ca	1.70	1.51	2.04	1.30
Ti	0.24	ND	ND	ND
Mn	0.04	ND	ND	ND
Fe	6.50	4.86	4.73	10.90
Cu	13.69	11.09	13.71	6.55
Total	93.38	94.30	96.43	92.61

ND: not detected.

Table 4. Results of SEM-EDS semi-quantitative analyses of carbon-coated dried statoblasts and anchor-shaped thorns collected from Makiyama agricultural reservoir pond in November 2016, coated with carbon, indicating richness in Si, S, and Ca associated with traces of Sr. Values are represented as mass %.

Makiyama Pond: Carbon Coated Statoblasts (mass %, $\pm 3\delta$)			
Elements	1	2	3
Na	4.70	6.84	6.64
Mg	5.68	5.34	4.32
Al	2.26	1.97	1.20
Si	9.43	5.72	10.09
P	3.05	0.53	0.47
S	45.83	37.01	53.75
Cl	3.81	2.33	3.51
K	3.55	ND	3.02
Ca	14.78	12.78	12.43
Ti	0.42	ND	0.59
Mn	ND	ND	ND
Fe	4.01	ND	1.89
Ni	0.63	0.59	ND
Cu	ND	15.95	ND
Zn	ND	8.22	ND
Sr	ND	0.59	0.64
Total	98.15	97.87	98.55
	Statoblasts	Statoblasts	Anchor shape thorn

ND: not detected.

3.5. SEM Elemental Distribution Maps

SEM elemental distribution maps of statoblasts of *Asajirella gelatinosa* or *Cristatella mucedo* from Kahokugata Lake showed a constant distribution of P and S, whereas Al, Si, Fe, and Ca were partially concentrated at the rim or the central parts of statoblasts (Figure 5).

On the other hand, the elemental distribution maps of the statoblasts of *Pectinatella magnifica* from Makiyama agricultural reservoir pond showed an even distribution of almost all elements (Na, Mg, Al, Si, P, S, Cl, K, Ca, and Fe) over all the statoblasts (Figure 4b).

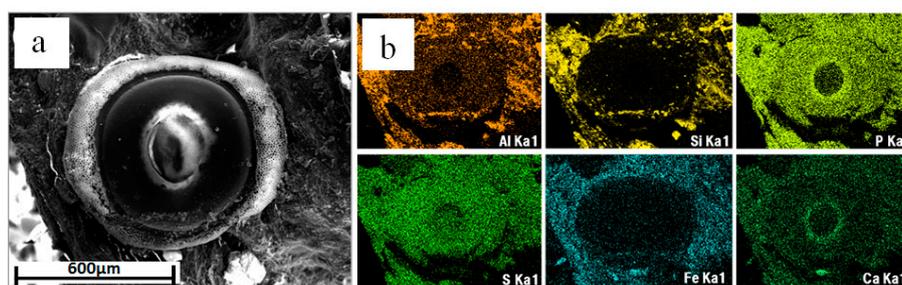


Figure 5. SEM-EDS elemental distribution maps of *Asajirella gelatinosa* collected from Kahokugata Lake. Elemental content maps indicated in this area associated with *Asajirella gelatinosa*. SEM image (a) Elemental content maps (b).

3.6. X-Ray Powder Diffraction (XRD) for Mineralogy of *Pectinatella magnifica* in Makiyama Agriculture Reservoir Pond

Some samples show almost equal XRD patterns. Therefore, we show typical profiles of each sample in Figure 6 and also show the diffraction peak position of minerals on Table 5. The mineralogy of the statoblasts of *Pectinatella magnifica* in Makiyama agricultural reservoir pond (Figure 6a), the gelatinous

Table 5. Peak positions and d values of XRD patterns.

(a) Statoblasts		
2θ (°)	d-Value (Å)	Relative Intensity
20.050	4.428	10
21.400	4.149	25
21.560	4.118	25
22.571	3.936	20
22.737	3.908	20
24.386	3.650	10
27.200	3.276	100
27.340	3.259	85
27.480	3.243	50
28.376	3.143	20
30.877	2.896	10
35.354	2.537	15
36.296	2.473	15
39.960	2.254	20
40.160	2.244	20
43.158	2.096	15
50.640	1.801	20
50.820	1.795	20
(b) Agar Part (Gelatinous Part)		
2θ (°)	d-Value (Å)	Relative Intensity
8.056	10.974	- *
12.237	7.233	- **
19.730	4.488	- *
35.939	2.497	40 ***
38.644	2.328	19 ***
42.881	2.107	100 ***
54.007	1.697	25 ***

* broad peak, ** weak peak, *** Zn plate of holder.

3.7. X-Ray Fluorescence Analyses (XRF) for Deposited Minerals in Makiyama Agricultural Reservoir Pond

The chemical compositions of sediment samples near the Makiyama agricultural reservoir pond obtained by XRF analyses are shown in Table 6.

Table 6. Results of XRF analyses of sediments in Makiyama agricultural reservoir pond, Kanazawa, Japan (mass %).

Components	1	2	3	4	5
SiO ₂	60.3000	58.1000	55.8000	56.8000	57.9000
Al ₂ O ₃	20.1000	19.5000	23.1000	14.9000	15.8000
CO ₂	6.5500	9.1400	9.3100	15.6000	13.0000
Fe ₂ O ₃	6.3000	4.5600	5.2000	4.6500	4.9900
K ₂ O	2.4100	2.2500	2.1700	2.3800	2.4100
MgO	1.9800	1.2200	1.1400	1.7500	1.8100
TiO ₂	0.8050	0.4590	0.4440	0.5730	0.6170
Na ₂ O	0.7310	1.6600	1.0500	1.2500	1.2500
CaO	0.4980	2.3200	1.1100	1.2500	1.1900
P ₂ O ₅	0.0947	0.4030	0.3540	0.4040	0.4620
SO ₃	0.0603	0.1540	0.1360	0.1850	0.1900
MnO	0.5290	0.0434	0.0588	0.1070	0.1060
Total	100.3590	99.8100	99.9280	99.8590	99.7250

4. Discussion

In this paper, mineralogical and elemental composition of *Pectinatella magnifica* and its statoblasts in freshwater are summarized (Figure 7). The distribution of elements and minerals inside of the bryozoan colony of *Pectinatella magnifica* indicates the role of bryozoans in mediating the transfer of metal solutes from hydrosphere to sediments. Abundant intracellular and cell wall Sr sorption has been identified in phytoplankton biomass and *Pectinatella magnifica* in Tawara Pond, Kanazawa, Ishikawa, Japan, at pH 7–8.5 and 20–30 °C, with C (42.9 wt %), H (5.8 wt %) and N (7.6 wt %) in summer 2015 [10]. Isolates of the phytoplankton community and *Pectinatella magnifica* were found to contain 0.65 mg/L of Sr by dry weight using the ICP-MS method. Concentration factors for Rb 1–1.55 mass % and Sr 5–7.5 mass % were found to be 5–7.5 mass % from point analyses, using the SEM-EDS method, confirming the role of microorganisms in mediating the transfer of metal solutes from the hydrosphere to sediments.

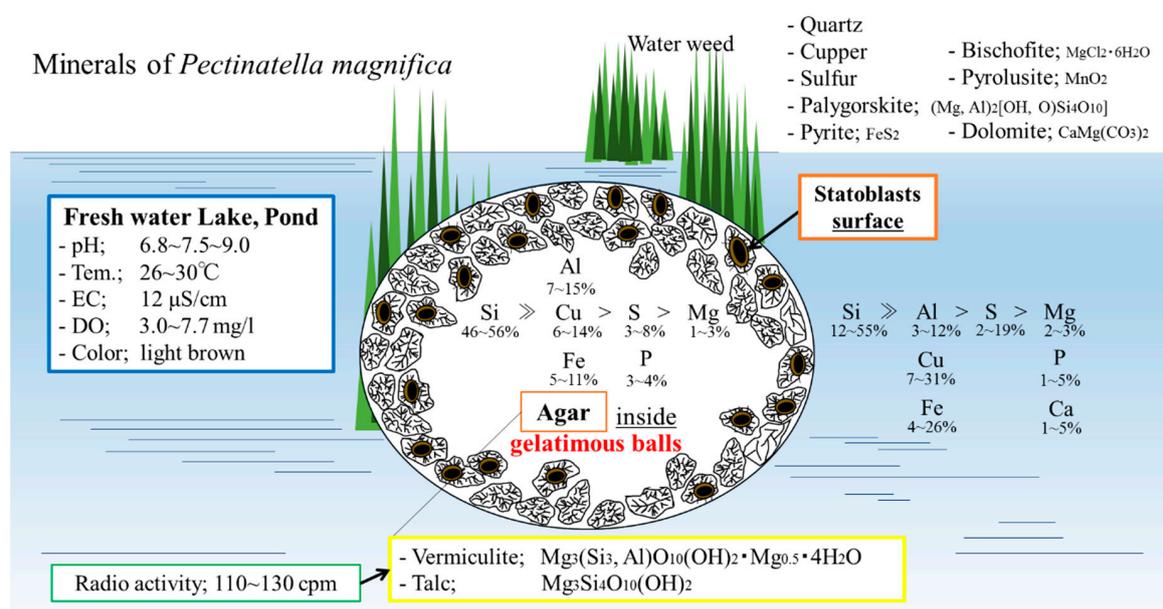


Figure 7. The distribution of elements and minerals inside the bryozoan colony, determined based on chemical data obtained from ICP-MS and SEM-EDS and mineralogical data from XRD. Characteristics of water, such as pH, temperature, EC, DO, and color are excerpted from Hashida et al. [14].

We think that the adsorption of part of some elements such as Al, Si, and Fe may be possible. On the other hand, the results of powder X-ray diffraction patterns (Figure 6) and rather high quantity of Si (Tables 3 and 4) show the existence of some silicate minerals such as vermiculite. Therefore, a high proportion of these elements exist as biominerals in *Pectinatella magnifica*.

4.1. Characteristics of Statoblasts

A key to species based on colony and statoblast gross micro morphology has already been provided [14] to aid in the identification of freshwater bryozoans in Makiyama agricultural reservoir pond and Kahokugata Lake using SEM-EDS, which provides useful characteristics related to the taxonomy of freshwater species described in this paper. Elemental distribution maps of statoblasts (Figure 4) have never previously been studied, and these showed high concentrations of P and S throughout the species, whereas Al, Si, and Fe were concentrated around the rim, showing the presence of minerals such as highly crystallized sulfur and palygorskite in statoblasts, whereas talc and vermiculite were present in the gelatinous material, as shown by XRD. Therefore, sulfur may be included in the statoblasts as native S and pyrite under redox conditions. On the other hand,

phosphorous may be included as non-crystalline matter. Because the XRD pattern shows a large background (halo) centered around $30^\circ(2\theta)$, analysis of non-crystalline matter is also important to understand *Pectinatella magnifica* totally. However, this kind of study requires more work. Therefore, we would like to deal with this material in a future work. On the other hand, the presence of Si is consistent with the quartz crystals identified in the XRD analysis. There is a possibility of quartz crystal due to the contamination of quartz crystal from sediments in Makiyama agricultural reservoir pond (Table 6). If the sediments contaminate the *Pectinatella magnifica* associated with gelatinous material, the XRD pattern of gelatinous materials (Figure 6b) may also show diffraction peaks of quartz. However, the XRD pattern of gelatinous materials did not show diffraction peaks of quartz. From these analyses, minerals in statoblasts detected by XRD measurements may be water-evaporated minerals and biominerals.

Otherwise, statoblasts of *Pectinatella magnifica* associated with gelatinous material will revive the following summer. The statoblasts of *Pectinatella magnifica* located upstream then drain downstream and are deposited in a lake, such as Kahokugata. It was particularly notable that quantities of *Pectinatella magnifica* have been found in Kahokugata Lake over a long period of time, since 1959. Furthermore, it is recognized that the factors affecting the size and quantity of *Pectinatella magnifica* are complex. Colonies accumulate in many lakes almost every summer in Japan [15–21].

4.2. Mineralogical and Elemental Composition of *Pectinatella magnifica* and Its Statoblasts

In this paper, we report not only the characterization of *Pectinatella magnifica* in freshwater systems, but also the formation of minerals inside gelatinous material and statoblasts on the surface of freshwater ponds and lakes in Kanazawa, Ishikawa, Japan, based on the data for chemicals and minerals in the field. Therefore, we report many data on the elements and minerals including in *Pectinatella magnifica* obtained using SEM-EDS, ICP-MS, and XRD techniques. These results, which are summarized in Figure 7, can provide insights into the identification of mineral species and characteristic structures. We analyzed the microchemistry of *Pectinatella magnifica* using a combination of micro-techniques. Previously, many mineralogical studies in bryozoans were limited to carbonate minerals such as calcite, Mg-rich calcite, and aragonite. However, in this work, we identified many other minerals as follows. The gelatinous material portions of *Pectinatella magnifica* may contain mainly of vermiculite and talc as well as organic non-crystalline materials, whereas highly crystalline quartz was associated with native sulfur, palygolskite, bischofite, pyrolusite, dolomite, pyrite, and copper in statoblasts of *Pectinatella magnifica* (Figure 6a). However, the presence of minerals such as palygolskite and copper is controversial. Therefore, more precise XRD analyses may be necessary for analyzing the mineralogy in detail.

The existence of clay minerals such as vermiculite and talc in gelatinous materials may be partially caused by the contamination of lake and pond sediments. It may be possible that materials including clay minerals flow into lakes and ponds based on the data on the chemical composition of the sediment. XRF data of the sediment for Makiyama agricultural reservoir pond (Table 6) are consistent with this interpretation. The existence of dolomite is consistent with similar quantities of Ca and Mg (Tables 2 and 3). Other minerals found in this study such as pyrite and native sulfur show reducing environment. Particular elements, such as abundant Si, Al, Cu, and Fe, were associated with S and Mg in the gelatinous material. Also, Si, Al, Cu, and Fe accumulated at the statoblast surfaces of *Pectinatella magnifica*, as shown in Figure 7, which illustrates *Pectinatella magnifica* based on chemical and mineralogical data. On the other hand, phosphorus may be included as a non-crystalline material, as mentioned above.

4.3. Distribution of Elements and Minerals inside the Bryozoan Colony

Elements often occur in close proximity in separate mineral phases, or substitute for one another as, for example, in silicate minerals, whose crystal chemical characteristics are now well known. Iron is also an essential element in the metabolic pathways of oxidation–reduction in all species from microbes

to man. From the simple aqueous chemistry of Si, Al, and Fe, it is clear that particular chemical species and clay minerals that occur in surficial environments are strongly dependent on environmental conditions [22–28].

This paper presents an overview of mineral species and essential elements, and the possible mechanisms involved in their precipitation. Depending on the specific environmental conditions, newly-formed Fe minerals are predominantly oxides, oxyhydroxides, sulfides, sulfates, carbonates, and phosphates. The main Fe mineral formation processes are discussed in many papers, e.g., by Schwertman and Fitzpatrick [29,30].

In *Actinopoda*, *Zoomastigina*, and *Mollusca*, different minerals are formed at different developmental stages. In the case of the oyster *Crassostrea virginica*, for example, the larval “shell gland” cells form aragonite, whereas in the adult, the mantle cells form calcite. In the holothurian species *Molpadiidae*, the juveniles initially form spicules of calcite. An example of a calcite spicule formed by a juvenile is reported in the literature [11,29–36].

It is also important to understand the possible mechanisms of metal uptake, whether biologically mediated, inorganically controlled, or working in concert, during the precipitation of minerals in Makiyama agricultural reservoir pond upstream and Kahokugata Lake downstream of the Morimoto River. Studying these mechanisms would add to our understanding of mineral formation and growth in surface water environments [37–40].

5. Conclusions

We studied the microstructure, mineralogy, chemical composition, and radioactivity associated with *Pectinatella magnifica* using a combination of micro techniques, scanning electron microscopy equipped with energy-dispersive spectroscopy (SEM-EDS), associated with inductively coupled plasma-mass spectrometry (ICP-MS), and X-ray powder diffraction analyses (XRD). This study aims to illustrate the capacity of *Pectinatella magnifica* to produce minerals within statoblasts, forming quartz, palygorskite, dolomite, bischofite, pyrolusite, pyrite, native sulfur, and native copper. The mass of gelatinous material may principally contain vermiculite and talc as well as non-crystalline organic materials. However, the presence of palygorskite, native copper, and talc is controversial. The mechanism of mineral formation has important implications for water–mineral–microorganism interactions in both lower drainage basin systems such as Kahokugata Lake, and upper water areas such as Makiyama agricultural reservoir pond. A SEM-EDS electron micrograph showing *Pectinatella magnifica* confirms the role of microorganisms in mediating the transfer of metal solutes (elemental distribution maps) from the hydrosphere to sediments. Many types of bryozoans (*Pectinatella magnifica*) were found in the lake and the pond in Japan. To date, there have been no reports describing electron microscopy observations, elementary content maps, and mineral formation. Therefore, this work is important for understanding the minerals include in bryozoans.

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References

1. Mawatari, S. New occurrence of *Pectinatella magnifica* (Leidy) in Japanese Lake. *Proc. Jpn. Soc. Syst. Zool.* **1973**, *9*, 41–43.
2. Oda, S. *Pectinatella magnifica* occurring in Lake Shoji, Japan. *Proc. Jpn. Soc. Syst. Zool.* **1974**, *10*, 31–39.
3. Oda, S. Notes on *Pectinatella magnifica* occurring in Japan. *Collect. Breed. Tokyo* **1975**, *37*, 40–53. (In Japanese)

4. Oda, S. A life of *Pectinatella magnifica* a freshwater bryozoan. *Ibid* **1976**, *38*, 434–443. (In Japanese)
5. Oda, S. A note on bryozoan in Lake Shoji, Japan. *Proc. Jpn. Soc. Syst. Zool.* **1978**, *15*, 19–23.
6. Hirose, M. Taxonomy as a basis and a goal of zoology, with some examples of bryozoology in museums. *TAXA Proc. Jpn. Soc. Syst. Zool.* **2013**, *34*, 25–31.
7. Hirose, M. Taxonomy of *Lophophorata*; Description of diversity of morphology and biology of bryozoans. *TAXA Proc. Jpn. Soc. Syst. Zool.* **2014**, *37*, 1–13.
8. Smith, A.M.; Key, M.M., Jr.; Gordon, D.P. Skeletal mineralogy of bryozoans: Taxonomic and temporal patterns. *Earth Sci. Rev.* **2010**, *78*, 271–277. [[CrossRef](#)]
9. Taylor, P.D.; Lombardi, C.; Cocito, S. Biomineralization in bryozoans: Present, past, future. *Biol. Rev.* **2015**, *90*, 1118–1150. [[CrossRef](#)] [[PubMed](#)]
10. Withrow, M. The chemical constitution of *Pectinatella*. *Science* **1930**, *71*, 265.
11. Tazaki, K.; Brandon, K.; Yamamoto, S.; Sato, M.; Hayashi, T.; Takehara, T. Strontium sorption in Phytoplankton biomass and *Pectinatella magnifica*; Implications for bioremediation. *Radioisotopes* **2016**, *65*, 383–392. [[CrossRef](#)]
12. Hirose, M. Taxonomy and identification of freshwater bryozoans in Japan. *TAXA Proc. Jpn. Soc. Syst. Zool.* **2012**, *33*, 17–32.
13. Miyashita, M.; Harada, M.; Noguch, M. *First Report of Magnificent Bryozoan Pectinatella magnifica (LEIDY) in Wakayama Prefecture and Its Distribution at Kinokawa and Iwada Cities*; Memoirs Institute of Advanced Technology, Kinki University: Osaka, Japan, 2014; Volume 19, pp. 1–12.
14. Hashida, Y.; Hashida, S.; Tazaki, K.; Nakamura, I.; Takehara, H.; Nakamura, K. Environmental activity and the ecotourism at Makiyama, Kanazawa, Ishikawa in Japan. *Kahokugata Lake Sci.* **2017**, *20*, 19–31.
15. Mukai, H. Germination of the statoblasts of freshwater bryozoan, *Pectinatella gelatinosa*. *J. Exp. Zool.* **1974**, *183*, 27–40. [[CrossRef](#)]
16. Oda, S. Freshwater bryozoan in Japan. *Biol. Jpn.* **1990**, *4*, 50–57.
17. Mawatari, S. USA originated *Pectinatella magnifica* (LEIDY) found in Japan. *Anim. Taxon.* **1973**, *9*, 41–44.
18. Kaneko, Y.; Iwatsuki, K.; Noda, Y. *Pectinatella magnifica* in Kagawa Prefecture. *J. Biol. Kagawa* **1994**, *21*, 63–66.
19. Hirose, M.; Ikezawa, H.; Kanek, N.; Mawatari, S. Distribution of freshwater bryozoans in Ibaragi Prefecture. *Ibaragi Nat. Mus. Rep.* **2006**, *9*, 31–46.
20. Iwasaki, K. Introduced sessile animals and invasive alien species act in Japan. *Sess. Org.* **2006**, *23*, 13–24.
21. Oda, S. Germination of the statoblasts in freshwater Bryozoa. *Sci. Rep. Tokyo Kyoiku Daigaku B* **1959**, *9*, 90–132.
22. Robin, N.; Bernard, S.; Miot, I.; Blanc-Valleron, M.M.; Charbonnier, S.; Petit, G. Calcification and diagenesis of bacterial colonies. *Minerals* **2015**, *5*, 488–506. [[CrossRef](#)]
23. Bindschedler, S.; Cailleau, G.; Verrecchia, E. Role of Fungi in the biomineralization of calcite. *Minerals* **2016**, *6*, 41. [[CrossRef](#)]
24. Simkiss, K.; Wilbur, K.M. *Biomineralization: Cell Biology and Mineral Deposition*; Academic Press Inc.: Waltham, MA, USA, 1989; p. 337.
25. Krumbein, W.E. *Microbial Geochemistry*; Blackwell Scientific Publications: Oxford, UK, 1983; p. 330.
26. Tazaki, K.; Okuno, M.; Furumoto, M.; Watanabe, H. Biomineralization of pisoliths in hot springs. *Mater. Sci. Eng. C* **2006**, *26*, 617–623. [[CrossRef](#)]
27. Tazaki, K.; Morii, I. Microbial immobilization of Si, Mn, Fe and Sr ions in the nacreous layer of *Sinohyloopsis schlegeli* and environmental factors. *Earth Sci. Front.* **2008**, *15*, 55–65. [[CrossRef](#)]
28. Tazaki, K. *Microbial Mats; Their Biomineralization and Bioremediation*; Takakuwa Art Printing Office: Ishikawa, Japan, 2009; 228p.
29. Lowenstam, H.A.; Weiner, S. *On Biomineralization*; Oxford University Press: Oxford, UK, 1989; pp. 45–46.
30. Mann, H.; Tazaki, K.; Fyfe, W.S.; Kerrich, R. *Microbial Accumulation of Iron and Manganese in Different Aquatic Environments: An Electron Optical Study*; CATENA SUPPLEMENT 21; CATENA: Cremlingen, Germany, 1992; pp. 115–131.
31. Tazaki, K.; Takehara, T.; Ishigaki, Y.; Nakagawa, H. The distribution of Sr, Cs, Np and Pu in the radioactive paddy rice field in Fukushima. *J. Foss. Res.* **2013**, *46*, 35–57.
32. Tazaki, K.; Simojima, Y.; Takehara, T.; Nakano, M. Formation of microbial mats and salt in radioactive paddy soils in Fukushima, Japan. *Minerals* **2015**, *5*, 849–862. [[CrossRef](#)]
33. Seckbach, J.; Oren, A. *Microbial Mats: Modern and Ancient Microorganisms in Stratified Systems*; Springer: New York, NY, USA, 2010; p. 606.

34. Morikawa, H.; Takahashi, M.; Kawamura, Y. New progress in environmental biotechnology using phytoremediation. *J. Environ. Biotechnol.* **2001**, *1*, 1–14.
35. Ehrlich, H.L.; Newman, D.K.; Kappler, A. *Ehrlich's Geomicrobiology*, 6th ed.; CRC Press, Taylor & Francis Group: Boca Raton, FL, USA, 2016; 635p.
36. Sekine, M.; Shimizu, F. Chemical Structure of Gel Material of *Pectinatella magnifica*. Abstract of Japanese Agricultural Chemical Society. 2013. Available online: <http://www.jsbba.or.jp/MeetingofJSBBA/2013/MeetingofJSBBA2013.pdf> (accessed on 24 September 2017).
37. Oono, K.; Shibasaki, T. The lagoon and bay sediments of the Hokuriku district. *Earth Sci.* **1967**, *21*, 25–34.
38. Tsukawaki, S.; Asano, I. *Sedimentation in Kahoku-gata Lagoon in Ishikawa Pref. Central Japan*; Bulletin of the Japan Sea Research Institute, Kanazawa University: Kanazawa, Japan, 1999; Volume 30, pp. 69–97.
39. Tazaki, K. Architecture of biomats reveals history of geo-, aqua-, and bio-systems. *Episodes* **1999**, *22*, 21–25.
40. Tazaki, K.; Tazaki, F.; Okuno, M.; Takehara, T.; Ishigaki, Y.; Nakagawa, H. Formation of pisolite at hot springs in Saturnia, Toscana, Italy. *J. Geol. Soc. Jpn.* **2016**, *122*, 45–60. [[CrossRef](#)]



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