

Review

OX-HDL: A Starring Role in Cardiorenal Syndrome and the Effects of Heme Oxygenase-1 Intervention

Stephen J. Peterson ^{1,2} , Abu Choudhary ², Amardeep K. Kalsi ², Shuyang Zhao ², Ragin Alex ³ and Nader G. Abraham ^{3,4,5,*} 

¹ Department of Medicine, Weill Cornell Medicine, New York, NY 10065, USA; Stp9039@nyp.org

² Department of Medicine, New York Presbyterian Brooklyn Methodist Hospital, Brooklyn, NY 11215, USA; Abc9031@nyp.org (A.C.); akk9013@nyp.org (A.K.K.); shz9027@nyp.org (S.Z.)

³ Department of Medicine, New York Medical College, Valhalla, NY 10595, USA; ralex@nymc.edu

⁴ Department of Pharmacology, New York Medical College, Valhalla, NY 10595, USA

⁵ Department of Medicine, Joan C. Edwards School of Medicine, Marshall University, Huntington, WV 25701, USA

* Correspondence: nader_abraham@nymc.edu; Tel.: +1-914-594-4153

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Abstract: In this review, we will evaluate how high-density lipoprotein (HDL) and the reverse cholesterol transport (RCT) pathway are critical for proper cardiovascular–renal physiology. We will begin by reviewing the basic concepts of HDL cholesterol synthesis and pathway regulation, followed by cardiorenal syndrome (CRS) pathophysiology. After explaining how the HDL and RCT pathways become dysfunctional through oxidative processes, we will elaborate on the potential role of HDL dysfunction in CRS. We will then present findings on how HDL function and the inducible antioxidant gene heme oxygenase-1 (HO-1) are interconnected and how induction of HO-1 is protective against HDL dysfunction and important for the proper functioning of the cardiovascular–renal system. This will substantiate the proposal of HO-1 as a novel therapeutic target to prevent HDL dysfunction and, consequently, cardiovascular disease, renal dysfunction, and the onset of CRS.

Keywords: heme oxygenase; cardiorenal syndrome; renal failure; congestive heart failure; oxidized HDL; HDL proteome

1. Introduction

Admission of heart failure (HF) patients in American hospitals exceeds 1 million people annually [1], with heart failure or coronary artery disease more prevalent in the more advanced stages of chronic kidney disease (CKD) [2]. Patients with heart failure are more predisposed to the development of acute kidney injury (AKI) [3,4]. This sets the stage for the relevance of the cardiovascular–renal system as an interrelationship between kidney function and cardiovascular health. Epidemiologic studies have investigated how renal dysfunction is a prominent risk factor for cardiovascular disease (CVD) [5] and how heart failure predisposes to kidney damage and/or the exacerbation of chronic kidney dysfunction. [6,7] However, *in vitro* and *in vivo* studies have yet to establish a definitive pathophysiological explanation for this phenomenon [8], otherwise known as cardiorenal syndrome (CRS) [9].

High-density lipoprotein–cholesterol (HDL–C) levels have become promising markers for the risk of CVD and even CKD [10,11]. Recent studies have cited that HDL function is more important than levels and that remodeling and dysfunction likely contribute to increased risk of CVD, CKD, and CRS [12–16]. HDL utilizes its protective effect through multiple mechanisms, including lowering tissue cholesterol

levels through reverse cholesterol transport, attenuation of low-density lipoprotein (LDL) oxidation, and decreasing inflammatory responses via association with paraoxonase 1 (PON1) [17–23].

Dysfunctional HDL can result from free radical attack or oxidation of “good” HDL, leading to Ox-HDL (“bad” HDL) [24–26]. Lipids and lipoproteins are the major culprits of free radical damage [27], which results in lipid peroxidation. Free-radical-mediated lipid peroxidation alters the biophysical properties of cell membranes, which may impair normal cellular function [28]. Furthermore, the generation of lipid peroxidation products, i.e., F2-isoprostane from arachidonic acid, may propagate the free radical damage via covalent modification of biomolecules [29–31]. It is pertinent to find an endogenous antioxidant that can prevent the remodeling of “good” HDL to proinflammatory and atherogenic “bad” HDL [32–35] (Figure 1). For CRS, it is crucial to find an antioxidant that can directly target the cardiovascular–renal system to protect from oxidative damage and dysfunction to the cardiovascular or renal systems [36].

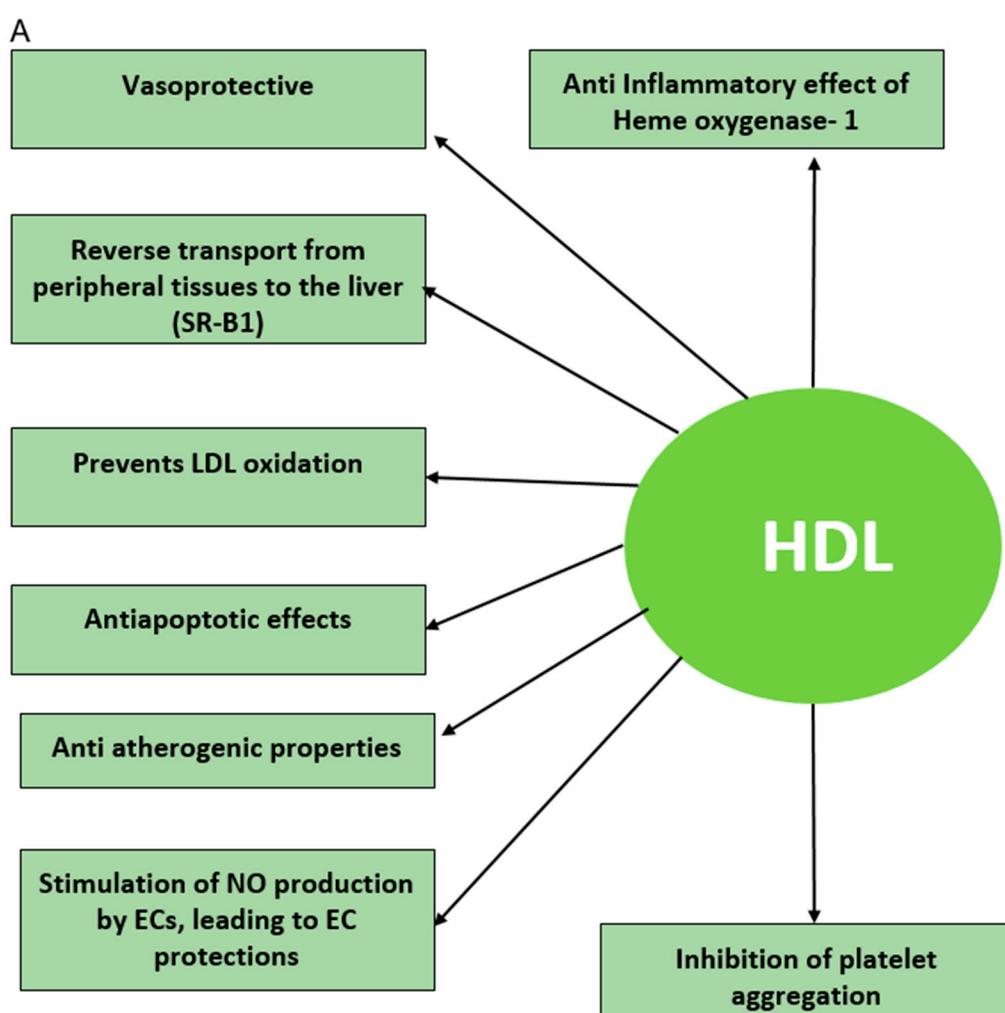


Figure 1. Cont.

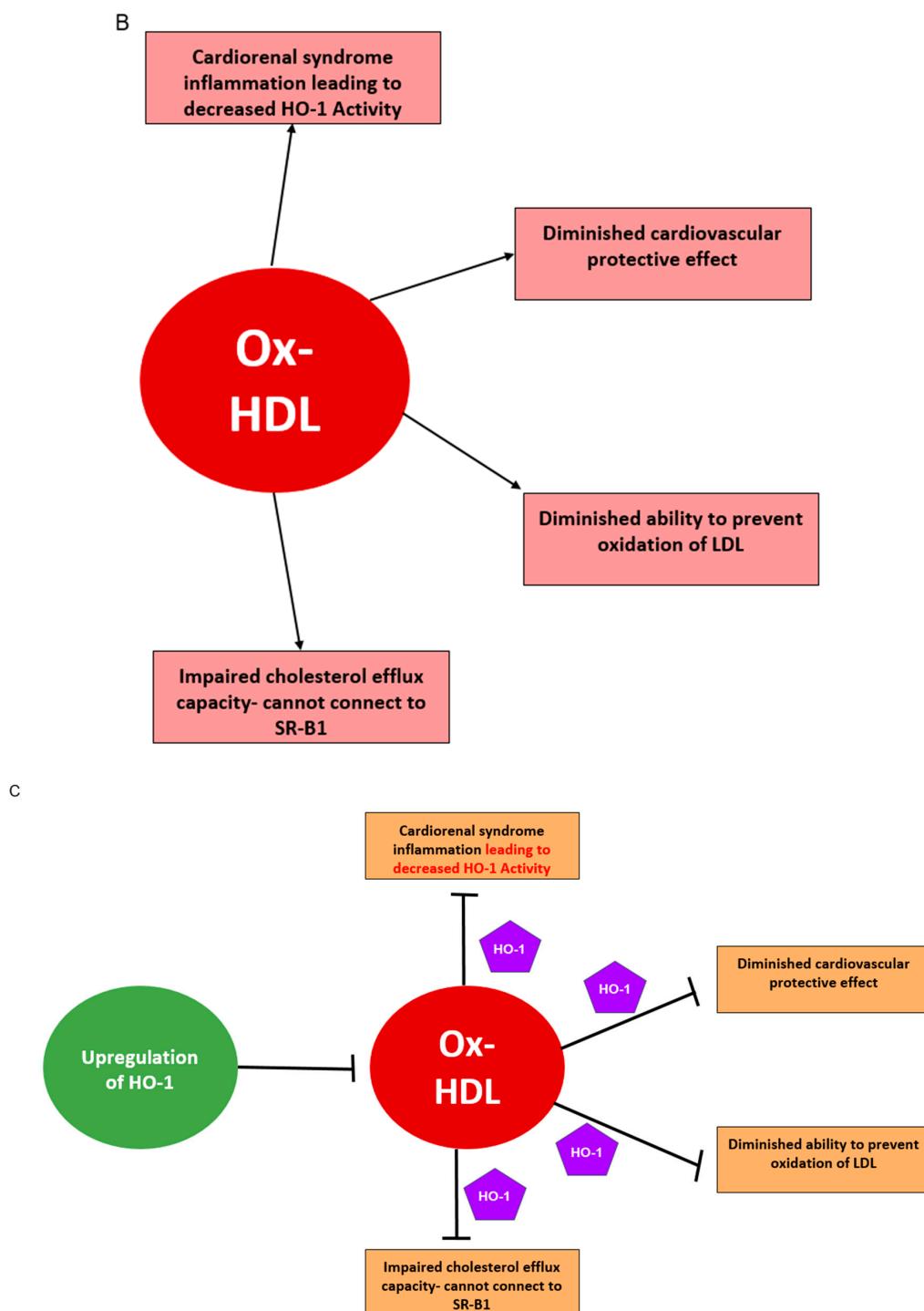


Figure 1. (A) The beneficial effects of high-density lipoprotein (HDL). HDL exhibits protective effects through multiple mechanisms, including lowering tissue cholesterol levels through reverse cholesterol transport, attenuation of LDL oxidation, and decreasing inflammatory responses. (B) The adverse effects of oxidized HDL (Ox-HDL). Oxidized or dysfunctional HDL is proinflammatory and atherogenic. (C) Role of HO-1. HO-1 has a protective effect against reactive oxygen species (ROS) and oxidative stress, and the upregulation of HO-1 decreases the detrimental effects of oxidized HDL. HO-1 = heme oxygenase 1; SR-B1 = scavenger receptor class B type 1; HDL=high-density lipoprotein cholesterol; LDL = low-density lipoprotein cholesterol; NO = nitric oxide; EC = endothelial cells.

2. The Heme Oxygenase System

Heme oxygenase-1 (HO-1) is an inducible enzyme within the body that is responsible for the catabolism of heme to equimolar parts carbon monoxide (CO) and biliverdin/bilirubin and the release of free iron [37]. HO-1 is known as a stress response protein [38], and we associate its induction with protection against reactive oxygen species (ROS) and, subsequently, oxidative stress [39]. We find heme oxygenase in the kidney in two isoforms: HO-1 (inducible form) and HO-2 [40]. Studies have shown HO-1 to be necessary and important for renal vascular and tubular function [41]. HO-1 has been shown to be important in vascular protection and function through the induction of adiponectin [42–44]. HO-1 is a potential and relevant therapeutic target for protection against and amelioration of CRS. HO-1 has been shown to decrease levels of angiotensin-II-mediated isoprostan production in endothelial cells [45]. Heme oxygenase has been proven to be cardioprotective, with associated induction of adiponectin expression [43,44,46,47], reduction in Ox-HDL with suppression of isoprostan production, and isoprostan binding to HDL. [48] Additionally, HO-1 may protect against heme-mediated damage in CRS, as hemoglobin has been shown to bind to “good” HDL and alter its conformation and functionality, causing HDL to become proinflammatory and atherogenic. [41,49].

Adiponectin is a cardioprotective protein hormone and has been shown to have increased plasma levels and improve vascular function after induction of HO-1 [37]. Adiponectin is well established as necessary for proper cardiovascular health, with clinical investigations proving adiponectin deficiency (hypoadiponectinemia) as an independent risk factor for CVD [50]. Researchers have also found hypoadiponectinemia to be strongly associated with renal dysfunction and CKD [51–53], and circulating adiponectin levels may be a predictor for CKD [54]. These studies show that adiponectin is crucial for vascular and renal function, with decreased levels leading to cardiovascular-renal dysfunction.

The heme oxygenase system may act as a potential therapeutic target for protection against and amelioration of cardiorenal syndrome through many pathways. Heme oxygenase is a potent endogenous antioxidant [55], and pharmacological induction of HO-1 improved type-1 cardiorenal syndrome in postischemic SCID mice [56]. HO-1 has been shown to decrease levels of angiotensin-II-mediated isoprostan-induced oxidative stress production in endothelial cells [45].

3. Structure of HDL and Reverse Cholesterol Transport Pathway

HDL is a small, dense lipoprotein particle with a high ratio of proteins to lipids [57], synthesized by liver hepatocytes [58]. HDL is a powerful anti-inflammatory agent that inhibits atherosclerosis [59,60]. The HDL proteome is very complex, with over 550 proteins reported in HDL [9]. HDL proteomics is a relatively novel approach to understanding the complex makeup and function of HDL in the setting of oxidative stress [61,62]. Sixteen HDL species with distinct proteomic signatures have been identified [63]. The HDL proteome is complex and separate from HDL cholesterol. Mass spectrometry has been used to understand complete HDL analysis and to identify biomarkers in order to better understand HDL function [61,64–67]. In fact, the HDL lipoprotein proteome has a high correlation with risk factors for cardiovascular disease and atherosclerotic burden and calcification on CT angiograms of coronary arteries [68]. The diversity of the HDL proteome is associated with clinical outcomes in patients with heart failure [69]. Alterations in the HDL proteome have been shown to result in dysfunctional HDL particles in type I diabetics [70,71].

HDL is primarily composed of several apolipoproteins, including Apo A-I, Apo A-II, and other proteins, including the enzyme paraoxonase. The external layer of HDL comprises free cholesterol, apolipoproteins (e.g., apoA-I, apoA-II, apoC, apoE), and phospholipids and is amphipathic [72]. The inner core of the HDL particle is highly concentrated with cholesterol esters, contains a minute amount of triglycerides, and is, therefore, hydrophobic [73].

HDL-C is considered the “good cholesterol” because of the physiologic function it performs in “reverse cholesterol transport” [9,74]. This is the process where the HDL particles move through the circulation and extract free cholesterol from less-dense particles and transport the free cholesterol to the liver to be processed and expelled, reducing the overall level of total cholesterol [75]. HDL is the

smallest of the lipoproteins but contains the highest apolipoprotein/lipid ratio [67]. The cholesterol delivered to the liver is excreted in bile and eventually converted into bile acids [76]. Delivery of HDL cholesterol to organs like adrenals, ovaries, and testes is critical for steroid hormone synthesis [34,77]. The efflux of cholesterol from HDL involves the following regulatory proteins.

ApoA-I has been shown to bind cholesterol in vitro and in vivo, showing its role in the uptake of extrahepatic cholesterol by HDL while traveling through the circulation [78–80]. Infusing patients with pro-apo-A-I was shown to increase RTC [81,82]. ApoA-I is synthesized in and interacts with the protein ATP-binding cassette transporter A 1 (ABCA1) in hepatocytes [83,84] and is then secreted into circulation as a lipid-poor particle. The lipid-poor apoA-1 particle removes cholesterol from the surfaces of macrophages in the arterial wall through interaction with ABCA1, forming nascent prebeta HDL or HDL₂ particles [85]. Phospholipid transfer protein (PLTP) is responsible for the transfer of phospholipids from triglyceride-rich lipoproteins to form nascent HDL particles and has been shown to interact with apoA-1 [86]. Apo A-1 oxidation has been shown to increase HDL oxidation and dysfunction [84,87].

The plasma enzyme [88] lecithin-cholesterol acyl transferase (LCAT) is responsible for the conversion of free cholesterol into cholesterol esters, which is a more hydrophobic form of cholesterol, making it easier to be sequestered into the core of the lipoprotein particle [89]. This eventually causes the newly synthesized HDL to assume a spherical shape [90]; the nascent HDL swells into a round, “mature” HDL particle. The mature HDL particle increases in size with the addition of more cholesterol and phospholipids from cells and other lipoproteins while circulating through the bloodstream [91].

A protein involved in the oxidation of HDL is myeloperoxidase (MPO), found in neutrophils and monocytes; it is released during acute inflammation. MPO generates Ox-HDLs, which cannot bind scavenger receptor class B type 1 (SR-B1) on the membrane of liver cells for RTC and have a proinflammatory function expressed through the upregulation of the protein (VCAM-1) on endothelial cells and the activation of NF- κ B and Ox-HDL, which lose their ability to activate eNOS and to inhibit caspase-3 and, therefore, lose their antiapoptotic activity [92] (Figure 1). HDL in patients with documented heart disease have elevated levels of nitrotyrosine and chlorotyrosine, both products of myeloperoxidase oxidative processes that facilitate the generation of dysfunctional HDL [93].

Alpha-HDL [72], containing free cholesterol and a cholesterol ester core, returns to the liver for selective uptake of cholesterol via contact with SR-B1. Cholesterol ester transfer protein (CETP) regulates the exchange of cholesterol esters from HDL to apo-B-containing lipoproteins for triglycerides [94,95]. Apo-containing lipoproteins then transfer cholesterol to the liver through interaction with hepatocyte LDL receptors [96].

The above processes are controlled by additional regulatory input from other proteins, including hormones and receptors. For example, the peroxisomal proliferator-activated receptor-alpha (PPAR α) has been shown to upregulate transcription of the ApoA-I gene [97]; transcription of the gene for ABCA1 is regulated by liver X receptors (LXRs) and retinoid X receptors (RXRs) [98]. Each of the HDL proteins, RCT pathway proteins, and regulatory factors highlighted above represent a potential point of intervention to raise the HDL-C number and/or promote reverse cholesterol transport.

4. Sexual Dimorphism and HDL

There are major differences in HDL levels between men and premenopausal women [99]. Obesity affects cardiometabolic function in both men and women but affects premenopausal women to a much lesser degree, even when matched to age and weight controls [100]. Sexual dimorphism is important to understand since there are major sex differences in fat distribution in visceral organs, skeletal muscle, and epicardial fat [101]. Adipose tissue distribution and adipose tissue health are responsible for differences in insulin sensitivity and consequent systemic inflammation. HDL levels are much higher in premenopausal women, but our group has shown a much higher Ox-HDL/HDL ratio in obese women, reducing the anti-inflammatory index of HDL and increasing their risk of endothelial cell dysfunction.

5. Cardiorenal Syndrome: A Definition

Cardiorenal syndrome is an umbrella term that encompasses the interaction between the heart and kidneys, such that injury to one organ causes dysfunction in the other [102]. There is well-established crosstalk in the pathophysiology of the heart and kidneys [103–106]. Patients on dialysis with end-stage renal disease (ESRD) have ten times the risk of death by a cardiovascular event than the general population [107]. Similarly, patients admitted with heart failure acquire renal dysfunction, and combined heart and kidney failure is associated with poor clinical outcomes [108]. The primary dysfunctional organ can be the heart or the kidney [109]. Cardiorenal syndrome is now defined more strictly than epidemiological outcomes in heart failure and dialysis patients. There are currently five types of cardiorenal syndrome, each with unique pathophysiology and progression of illness [110,111] (Table 1).

Table 1. Types of Cardiorenal Syndrome.

Type 1 Acute Cardiorenal Syndrome	Acute decompensated heart failure ⇔ Acute kidney injury
Type 2 Chronic Cardiorenal Syndrome	Chronic heart failure ⇔ Chronic kidney disease
Type 3 Acute Renocardiac syndrome	Acute kidney injury ⇔ Acute heart failure
Type 4 Chronic Renocardiac syndrome	Chronic kidney disease ⇔ Chronic heart failure
Type 5 Secondary Cardiorenal syndrome	Codevelopment of heart failure and chronic kidney disease due to acute or chronic systemic disorder

Type 1 cardiorenal syndrome is acute in nature, categorized as acute heart failure that leads to AKI, clinically presenting with inadequate renal perfusion due to an increase in venous pressure or a low cardiac output state, leading to kidney congestion [112] (Figure 2). Type 1 CRS is a common occurrence; AKI occurs in about 25% of hospitalized patients with HF, and declining renal function has been identified as an independent predictor of mortality. AKI activates the renin–angiotensin–aldosterone system (RAAS), salt and water imbalance, and vasoconstriction, all of which contribute to continued heart damage [113]. HF leads to decreased renal perfusion, along with monocyte and endothelial activation, causing cytokine secretion and further depressing renal function. This bidirectional pathophysiology is also exacerbated with common therapeutic agents such as ACE inhibitors and diuretics, leading to toxicity and vasoconstriction [114]. Production of epinephrine and angiotensin, along with a decreased sensitivity to vasodilators such as nitric oxide, causes excessive vasoconstriction, thereby worsening cardiorenal function. Fibrosis from inflammation is a common feature in HF and CKD [115] and may well be the unifying pathophysiology of the entire CRS continuum [116]. Evaluation of creatinine levels can be misleading, as the SOLVD trial showed that early initiation of RAAS inhibitors may reflect early changes in renal hemodynamics and may not reflect kidney injury [117,118]. Signs of CRS include an increase in the serum creatinine by 0.3 mg/dl in a 48-h period, an increase in the serum creatinine to 1.5 times baseline, or a urine volume less than 0.5 ml/kg/h over a six-hour period [119,120].

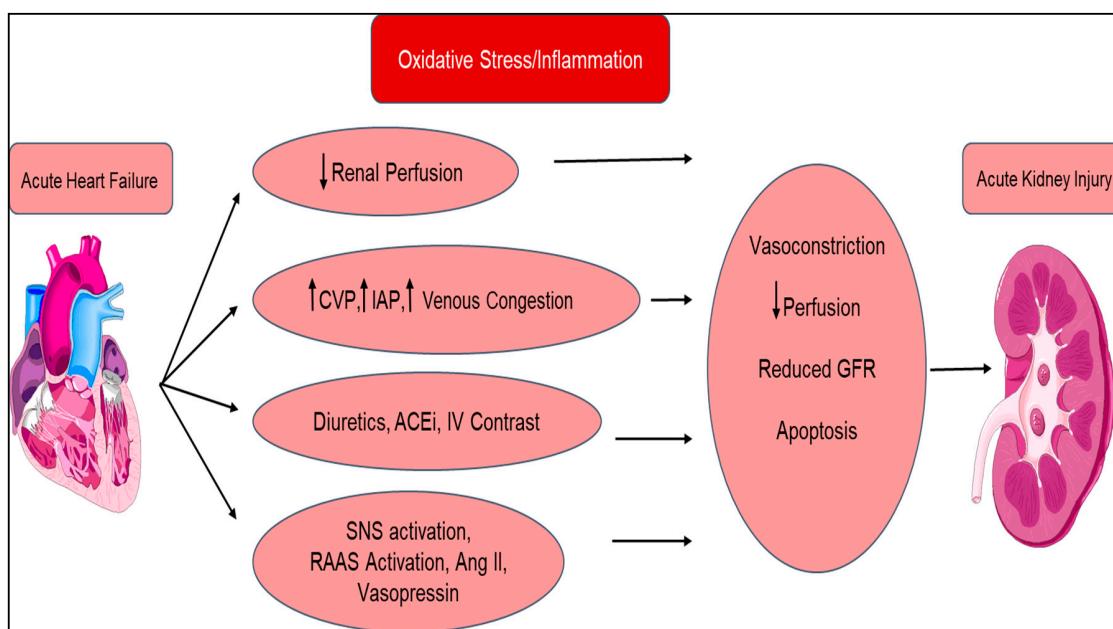


Figure 2. Pathophysiology of acute cardiorenal syndrome (type 1): mechanism of how acute heart failure leads to acute kidney injury (AKI) due to inadequate renal perfusion, endothelial activation, and cytokine production, which activates the RAAS, salt and water imbalance, and vasoconstriction, further exacerbating AKI. CVP = central venous pressure; IAP = intraabdominal pressure; ACEi = angiotensin converter enzyme inhibitor; RAAS = renin–angiotensin–aldosterone system; SNS = sympathetic nervous system; GFR = glomerular filtration rate.

Type 2 CRS is similar to type 1; however, where type 1 is acute, type 2 CRS is chronic (Figure 3). Type 2 CRS is characterized by chronic cardiac dysfunction (e.g., chronic congestive heart failure), which causes progressive CKD. The prevalence of renal dysfunction in chronic HF is significantly large; nearly 50% of patients with chronic HF appear to have decreased GFR. Even a minimal decrease in GFR is a strong independent predictor of mortality. Neurohormonal activation is also present in type 2 CRS, namely, increased production of vasoconstrictive mediators (angiotensin) and altered release of vasodilatory mediators (nitric oxide) [121]. Progression of CKD can be attributed to multiple factors including, but not limited to, low cardiac output, inflammation, endothelial dysfunction, accelerated atherosclerosis, chronic hypoperfusion, and increased renal vascular resistance. The progression of CKD further instigates cardiac dysfunction through RAAS activation, hypertension, and anemia.

Type 3 CRS is also called renocardiac syndrome and is categorized with abrupt kidney injury being the primary illness (e.g., ischemia, hypoperfusion, glomerulonephritis), leading to acute cardiac dysfunction (e.g., HF, arrhythmia, ischemia; Figure 4). AKI is a powerful predictor of hospital mortality; however, type 3 CRS is less common than type 1 CRS. AKI negatively impacts cardiac function through a variety of mechanisms. Renal ischemia has been shown to induce inflammation and apoptosis in cardiac cells. AKI contributes to acute heart dysfunction via familiar mechanisms from the abovementioned CRS types: RAAS activation, hypertension, decreased GFR, endothelial activation, and cytokine secretion [113]. Cytokines such as tumor necrosis factor (TNF), IL-1, and IL-6 play a diagnostic role and are also a pathogenic cause of myocardial cell damage and apoptosis during ischemic AKI. Furthermore, myeloperoxidase, a biomarker of oxidative stress and inflammation in acute coronary syndrome, may cause apoptosis and play a potential role in the pathogenesis of CRS.

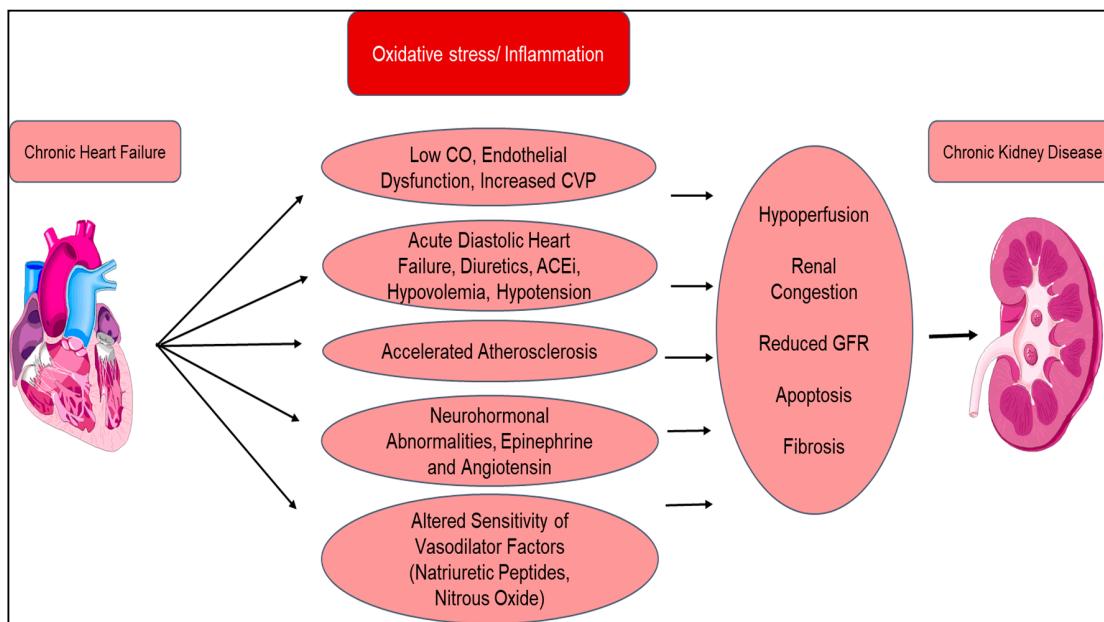


Figure 3. Pathophysiology of chronic cardiorenal syndrome (type 2): mechanism of how multiple effects of chronic heart failure lead to progressive and chronic kidney disease (CKD) because of neurohormonal upregulation, leading to altered vasoconstriction and vasodilation. CO = cardiac output; CVP = central venous pressure; ACEi = angiotensin-converting enzyme inhibitor.

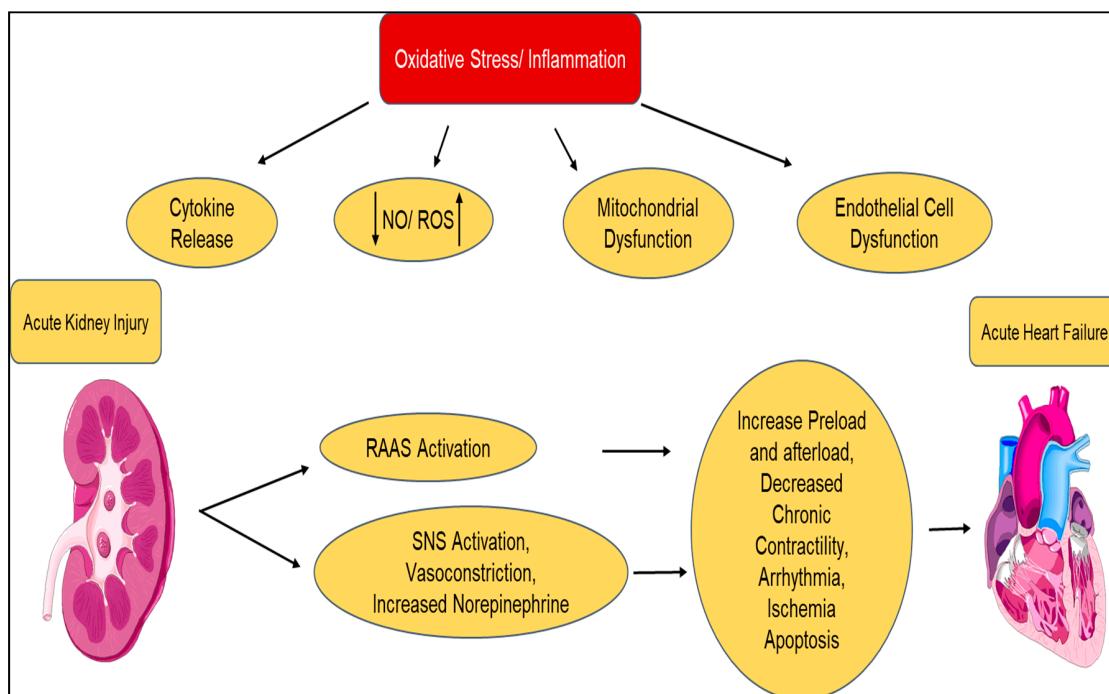


Figure 4. Pathophysiology of renocardiac syndrome (type 3): mechanism involves an acute insult to kidney function that results in the acute sympathetic nervous system activating a cascade of inflammatory responses, causing acute heart failure. NO = nitrous oxide; RAAS = renin–angiotensin–aldosterone system; ROS = reactive oxygen species; SNS = sympathetic nervous system.

Type 4 CRS is also known as chronic renocardiac syndrome, which involves primary CKD that contributes to worsening heart function and an increased risk for cardiovascular events (Figure 5). Current estimates for CKD in the general population exceed 10% of the US adult population. CKD is an

independent risk factor for cardiovascular-event-related mortality in individuals at all stages of CKD, especially for ESRD, which has an increased risk of cardiac death compared to patients without CKD. CKD is considered a more significant predictor of cardiovascular disease than diabetes mellitus [113]. Mechanisms for cardiac remodeling and decreased function from CKD are multifarious in nature. In Stages 1 and 2, risk factors resulting in CKD (e.g., obesity, hypertension, dyslipidemia, and chronic inflammation) contribute to decreased cardiac function. In Stages 3 and 4, anemia, uremia toxins, nutritional status, and BMI, along with chronic inflammation, lead to increased ischemic risk, coronary calcification, and neurohormonal abnormalities, as previously discussed. In late-stage CKD leading into dialysis, we see chronic inflammation (again), renal toxicity, endothelial dysfunction, oxidative stress, and accelerated atherosclerosis as contributing factors to the vicious cycle of cardiac dysfunction with renal failure.

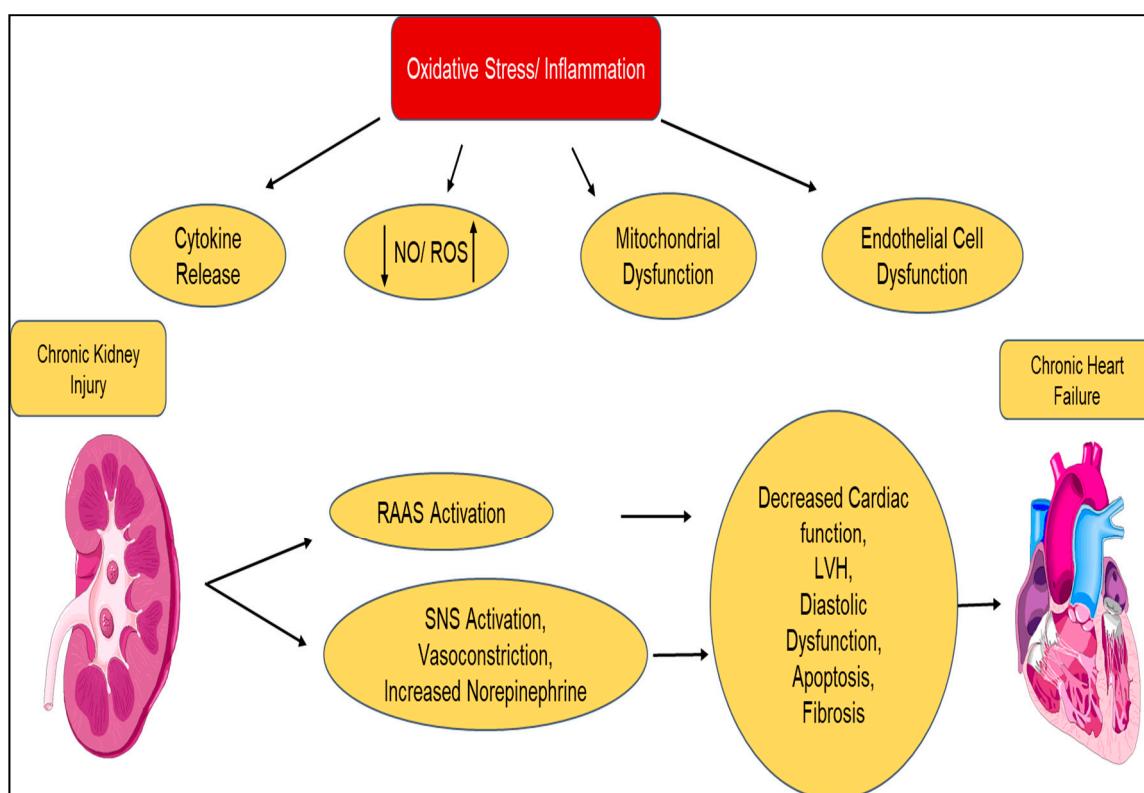


Figure 5. Pathophysiology of renocardiac syndrome (type 4): mechanism involving chronic kidney injury causing worsening heart function. The various causes of CKD, including diabetes and hypertension, accelerate cardiac remodeling, leading to poor cardiac function. This pathway involves RAAS activation and sympathetic nervous system activation.

Type 5 cardiorenal syndrome results in simultaneous renal and cardiac failure due to acute or chronic systemic disorders. Diabetes, sepsis, amyloidosis, and sarcoidosis are examples of such diseases affecting combined renal and cardiac function (Figure 6). Sepsis accounts for the most common condition that can acutely affect both organs, with the mechanisms being unclear but possibly involving TNF. Patients presenting with sepsis have multiorgan dysfunction over 50% of the time, especially involving cardiac and renal function, explaining the high mortality involved. [122] We observed this during the COVID-19 pandemic with systemic vascular inflammation [123–127].

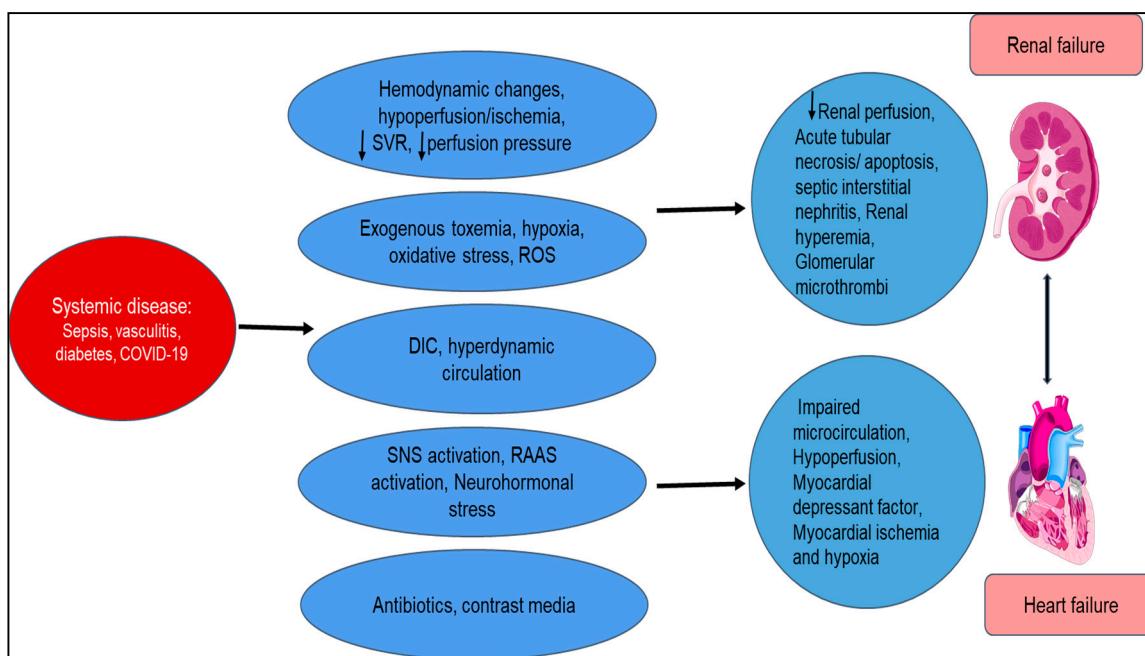


Figure 6. Summary of the pathophysiology of secondary cardiorenal syndrome (type 5): mechanism is characterized by combined cardiac and renal failure due to acute or chronic processes that cause hemodynamic instabilities, hypercoagulability, neurohormonal imbalances, and toxicity and hypoxia that cause poor renal perfusion, myocardial ischemia, and hypoxia. SVR = systemic vascular resistance; ROS = reactive oxygen species; DIC = disseminated intravascular coagulation; SNS = sympathetic nervous system; RAAS = renin–angiotensin–aldosterone system.

6. Cardiorenal Syndrome and HO-1

Physiological interaction between organs is necessary for the optimal equilibrium and functioning of the organism. Derangements in these interactions can initiate multiorgan dysfunction. In particular, heart and kidney functions are closely interrelated through a variety of dynamic and bidirectional mechanisms [128,129]; a pathological alteration in one organ can unfavorably affect function in another distant organ.

CRS involves complex interactions at the molecular level that induce vessel inflammation, atherosclerosis, cardiac fibrosis, and hypertrophy; in addition, structural and biochemical abnormalities can adversely affect cardiovascular or renal function [130].

Bright initially deliberated the causal association between chronic kidney disease (CKD) and cardiovascular risk in 1836. Patients with CKD are among the highest risk groups for adverse cardiovascular events and cardiovascular-related mortality and, therefore, require particular clinical attention. A recent study provided insight into the pathogenesis of CRS type 1, emphasizing the pivotal role of oxidative stress in CRS type 1 [128,131]. The study revealed that levels of oxidative stress markers (myeloperoxidase, nitric oxide, copper/zinc superoxide dismutase, and endogenous peroxidase activity) were significantly higher in CRS type 1 than in acute heart failure without CRS type 1 and in healthy controls [128,131]. In particular, CRS type 1 patients presented a significant increase in circulating ROS and RNS and an increased expression of the inflammatory cytokine IL-6 [132]. Monu et al. showed that ang-II-mediated recruitment of T-lymphocytes and increased oxidative stress is decreased by the upregulation of HO-1 in a model of postischemic heart failure [56]. The results showed that HO-1 induction decreased renal vasoconstriction and fibrosis and improved renal function in both immunocompetent and T-lymphocyte-suppressed mice [133]. Interestingly, treatment with SnMP, a known HO activity inhibitor, reversed the beneficial effects of HO-1 induction, suggesting that increased levels of HO activity play a central role in preventing MI-induced cardiac and renal damage in this CRS animal model. HO-1 induction reduces postischemic pathological

cardiac remodeling and, in mice with advanced heart failure and CRS, improves cardiac function and renal vasoconstriction. This renal vasoconstriction was demonstrated in a murine model of type 1 CRS, secondary to postischemic changes of LAD ligation [134].

Accumulating evidence suggests that hyperuricemia is one of the important factors that may significantly contribute to the development and progression of CRS. Elevated levels of uric acid have been associated with inflammation, oxidative stress, insulin resistance, dysglycemia, endothelial dysfunction, vascular, renal and cardiac stiffness, cardiac diastolic dysfunction, renal hyperfiltration, and proteinuria, all of which are components of CRS [135–137]. The significance of a westernized diet, which is high in fructose, and hyperuricemia in the development of CRS is underscored by the relationship between increased consumption of sugar-sweetened beverages, hyperuricemia, and all components of this syndrome [138]. Sodhi et al. showed that induction of HO-1 reduced expression of xanthine oxidase and NADPH oxidase, enzymatic systems that are important for ROS production, in adipocytes treated with fructose, a fuel source that increases uric acid levels [139]. Additionally, Khitan et al. showed that mice following a fructose diet presented an increase in isoprostanes that was associated with a decrease in HO-1 expression and an increase in heme levels. Isoprostanes and heme are regarded as valid markers of oxidative stress [140–142]. Upregulation of HO-1 presents cardio- and reno-protective functions mediated by its antioxidative, anti-inflammatory, and antiapoptotic properties. In animal models of myocardial ischemia (MI), both overexpression and pharmacological induction of HO-1 reduce infarct size and ventricular remodeling after ischemia-reperfusion damage by improving cardiac metabolism [143]. Increased HO-1 expression has a protective effect against ischemia-reperfusion injury in the kidney [144] and can correct blood pressure elevation following ang-II exposure [145].

Hyperglycemia-induced mitochondrial oxidative stress, a cause of metabolic CRS, is a contributory factor to increased risk of cardiovascular disease, which can induce cellular injury and cell dysfunction [146,147]. The molecular mechanism of mitochondrial dysfunction in CRS is driven via abnormalities involving the transcriptional coactivator peroxisome proliferator-activated receptor gamma coactivator 1 α (PGC-1 α), which controls the biogenesis of mitochondria and mitochondrial function in a variety of tissue and cell types. PGC-1 α is regulated by the endothelial NO synthase that plays an important role in mitochondrial biogenesis. Studies have shown that decreased expression of PGC-1 α -associated impairment of mitochondrial biogenesis may be responsible for various metabolic abnormalities in CRS [146] (Figure 7). Thus, the impairment of the complex steps in the regulation of mitochondrial biogenesis may contribute to CRS.

Activation of PGC-1 α reduced mitochondrial ROS, prevented adipogenesis in adipocytes, and protected diabetic hearts from hyperglycemia-mediated oxidative stress [148–150]. It has been shown that the protective effect of EETs in diabetic mice involves increased expression of PGC-1 α and SIRT1 [131]. In cardiomyocytes, pharmacological inhibition of SIRT1 was followed by decreased expression of PGC-1 α and increased ROS production [150,151]. At the same time, Singh et al. showed that PGC-1 α regulates HO-1 expression, confirming the beneficial effect of HO-1 in cardiovascular diseases and lipid metabolism [152,153]. Heme oxygenase-1 upregulation of PGC-1 α signaling in epicardial fat attenuated cardiovascular risk in both humans and mice [154]. Adipocyte-specific HO-1 gene therapy is very effective in reducing oxidative stress, improving both insulin resistance and vascular function in obese mice [155]. Finally, upregulation of HO-1 by icosapent ethyl, pomegranate seed oil, and black seed oil (thymoquine) have all reduced oxidative stress, improved insulin resistance, and improved mitochondrial function [156,157] (Figure 7).

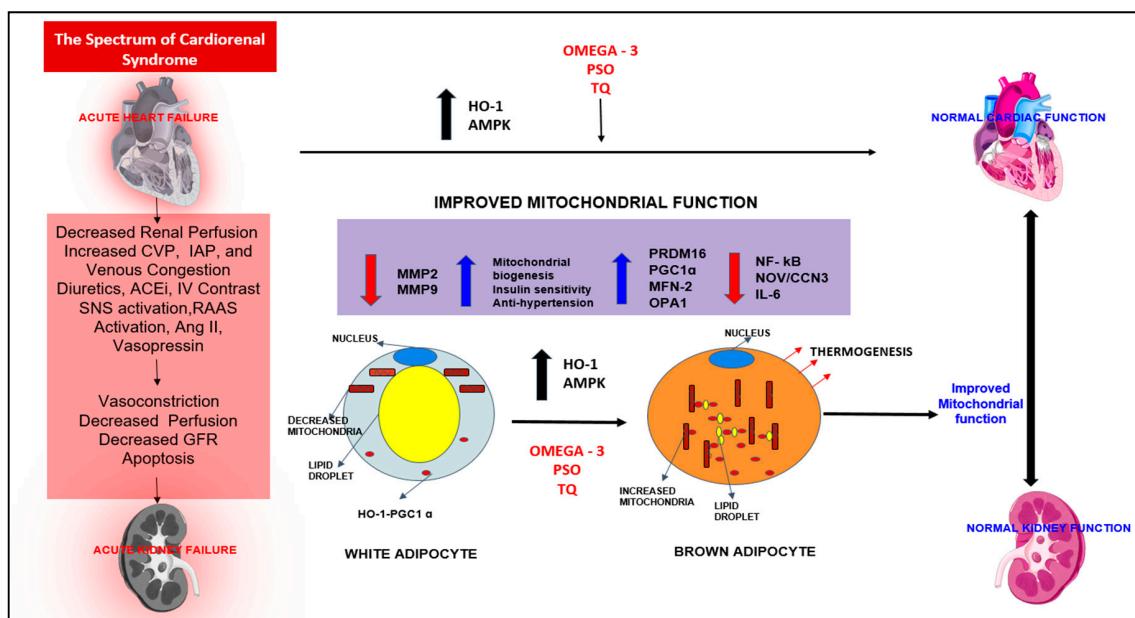


Figure 7. The spectrum of cardiorenal syndrome. This is a schematic representation of how HO-1 upregulation results in improved mitochondrial function and signaling pathways PGC1 α , PRDM 16, OPA1, and MFN-2. Upregulation also results in decreased inflammatory adipocytokines (NOV/CCN3, IL-6, and NF- κ B) and fibrotic markers (MMP2 and MMP9). The result was the “browning” of white adipocytes to “beige”, improving cardiac and renal functions. HO-1 = heme oxygenase 1; AMPK = AMP-activated protein kinase; PSO = pomegranate seed oil; TQ = thymoquinone; NF- κ B = nuclear factor kappa-light-chain-enhancer of activated B-cells; NOV/CCN3 = nephroblastoma overexpressed; IL-6 = interleukin 6; MMP2 = matrix metalloproteinase 2; MMP9 = matrix metalloproteinase 9; PRDM16 = PR domain containing 16; PGC-1 α = peroxisome proliferator-receptor gamma coactivator 1 α ; MFN-2 = mitofusin-2; OPA1 = mitochondrial dynamin-like GTPase.

More attention is being paid to the role of Ox-HDL in the chronic inflammatory state of obesity. Ox-HDL has been shown to independently upregulate the downstream activity of ang II and has been identified as part of a biomarker profile for early endothelial dysfunction in obese women without identified cardiovascular disease [158] (Figure 1). We have shown oxidized HDL to be part of a biomarker profile for cardiovascular risk in obese women [159]. PGC-1 α has been shown to improve organ function by upregulating mitochondrial enzymes, improving mitochondrial function in metabolic syndrome and nonalcoholic fatty liver disease (NAFLD), and reducing Ox-HDL [146]. Most importantly, this improvement in mitochondrial integrity and function will aid in the reprogramming of white fat to beige-like fat [160].

7. Lifestyle Interventions, Weight Loss Medications, and Nutraceuticals

Weight loss with diet and exercise has been successful in short-term supervised programs, with weight loss in excess of 5% body weight. None have approached the magic number of 10–12% of sustained body weight loss in BMIs above 35. The PREDIMED-PLUS study was a 12-month intervention that increased weight loss and improved cardiovascular risk factors [161]. There is confusing evidence as to the effect of aerobic exercise on HDL levels and cholesterol efflux [162]. Exercise has been shown to improve psychological health, particularly in the elderly [163]. Many nutraceuticals (like curcumin, CoQ 10, folic acid, berberine, alpha lipoic acid, astaxanthin, and policosanol) have tried to improve lipid and plasma glucose levels, with positive effects on lipid profiles [164]. Weight-loss drugs have been effective in supervised programs, especially those with close telemedicine follow-up [165–168]. There is clearly a role for diet, exercise, weight loss medications, and nutraceuticals as preventive measures in

obesity, metabolic syndrome, and type 2 diabetes. They are measures meant to prevent cardiorenal syndrome. We need more aggressive measures once we are confronted with cardiorenal syndrome.

8. Summary

Mitochondrial function is important for improving the function of the electron transport chain in both the heart and kidneys and converting white adipose tissue to beige, the “browning” of white adipose tissue. This results in an improvement of adiponectin levels and a severe reduction in the release of inflammatory adipocytokines. HO-1 upregulation is the key to improving mitochondrial function in both organs and reducing oxidative stress and Ox-HDL, which are important components of success in treating and preventing cardiorenal syndrome.

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Abbreviations

Ang II	angiotensin II
ARF	acute renal failure
AS	antisense
ASK-1	apoptotic signaling kinase-1
BP	blood pressure
BR	bilirubin-IX α
BV	biliverdin-IX α
BVR	biliverdin
Bw	body weight
CEC	circulating endothelial cell
CO	carbon monoxide
CoPP	cobalt protoporphyrin IX dichloride
CRP	C reactive protein
D-4F	apolipoprotein mimetic peptide
EC	endothelial cell
EC-SOD	extracellular superoxide dismutase
EETs	epoxyeicosatrienoic acids
eNOS	endothelial nitric oxide synthase
GFR	glomerular filtration rate
HCL	hypercholesterolemia
HETE	hydroxyeicosatetraenoic acid
HO	heme oxygenase
HO-1	heme oxygenase isozyme 1, inducible form
HO-2	heme oxygenase isozyme 2, constitutive form
ICAM-1	intracellular adhesion molecules-1
IL	interleukin
iNOS	inducible nitric oxide synthase
IRI	ischemia/reperfusion injury
LDL	low-density lipoprotein
MAPK	mitogen-activated protein kinase
MCP-1	monocyte chemoattractant protein-1

MSC	mesenchymal stem cells
NO	nitric oxide
NOS	nitric oxide synthase
CCN3/NOV	nephroblastoma overexpressed
O ₂ ⁻	superoxide anion radical
OONO ⁻	peroxynitrite
ROS	reactive oxygen species
SBP	systolic blood pressure
sEH	soluble epoxide hydrolase
SnMP	tin mesoporphyrin IX dichloride
SnPP	tin protoporphyrin IX dichloride
SOD	superoxide dismutase
T2DM	type 2 diabetes mellitus
TNF	tumor necrosis factor
TNF α	tumor necrosis factor-alpha
UVA	ultraviolet A (320–380 nm) radiation
VCAM-1	vascular adhesion molecules-1
VEGF	vascular endothelial growth factor
VEGFRI	vascular endothelial growth factor receptor I
VSMC	vascular smooth muscle cell
AMPK	AMP-activated protein kinase
AKT	serine/threonine-protein kinase
eNOS	endothelial NO synthase
FGF21	fibroblast growth factor 21
IL-10	interleukin 10
NF	kB nuclear factor-kappa B
NGF	nerve growth factor
PGC1 α	peroxisome proliferator-activated receptor gamma, coactivator 1
PPAR α	peroxisome proliferator-activated receptor
SIRT1	sirtuin 1
TNF- α	tumor necrosis factor- α
WAT	white adipose tissue
UCP1	Uncoupling protein 1
Ang II	Angiotensin II
aP2	adipocyte protein 2
CEBP α	CCAAT/enhancer-binding protein alpha
CoPP	cobalt protoporphyrin
eNOS	endothelial nitric oxide synthase
FAS	fatty acid synthase
H ₂ O ₂	hydrogen peroxide
MSC	mesenchymal stem cell
PPAR γ	peroxisome proliferator-activated receptor gamma

References

1. Haldeman, G.A.; Croft, J.B.; Giles, W.H.; Rashidee, A. Hospitalization of patients with heart failure: National Hospital Discharge Survey, 1985 to 1995. *Am. Heart J.* **1999**, *137*, 352–360. [[CrossRef](#)] [[PubMed](#)]
2. Foley, R.N. Clinical epidemiology of cardiovascular disease in chronic kidney disease. *J. Ren. Care* **2010**, *36* (Suppl. 1), 4–8. [[CrossRef](#)] [[PubMed](#)]
3. Adams, K.F., Jr.; Fonarow, G.C.; Emerman, C.L.; LeJemtel, T.H.; Costanzo, M.R.; Abraham, W.T.; Berkowitz, R.L.; Galvao, M.; Horton, D.P. Characteristics and outcomes of patients hospitalized for heart failure in the United States: Rationale, design, and preliminary observations from the first 100,000 cases in the Acute Decompensated Heart Failure National Registry (ADHERE). *Am. Heart J.* **2005**, *149*, 209–216. [[CrossRef](#)] [[PubMed](#)]

4. Fonarow, G.C.; Stough, W.G.; Abraham, W.T.; Albert, N.M.; Gheorghiade, M.; Greenberg, B.H.; O'Connor, C.M.; Sun, J.L.; Yancy, C.W.; Young, J.B. Characteristics, treatments, and outcomes of patients with preserved systolic function hospitalized for heart failure: A report from the OPTIMIZE-HF Registry. *J. Am. Coll. Cardiol.* **2007**, *50*, 768–777. [[CrossRef](#)] [[PubMed](#)]
5. Hui, X.; Matsushita, K.; Sang, Y.; Ballew, S.H.; Fulop, T.; Coresh, J. CKD and cardiovascular disease in the Atherosclerosis Risk in Communities (ARIC) study: Interactions with age, sex, and race. *Am. J. Kidney Dis.* **2013**, *62*, 691–702. [[CrossRef](#)] [[PubMed](#)]
6. Hillege, H.L.; Nitsch, D.; Pfeffer, M.A.; Swedberg, K.; McMurray, J.J.; Yusuf, S.; Granger, C.B.; Michelson, E.L.; Ostergren, J.; Cornel, J.H.; et al. Renal function as a predictor of outcome in a broad spectrum of patients with heart failure. *Circulation* **2006**, *113*, 671–678. [[CrossRef](#)]
7. Damman, K.; Masson, S.; Hillege, H.L.; Voors, A.A.; van Veldhuisen, D.J.; Rossignol, P.; Proietti, G.; Barbuzzi, S.; Nicolosi, G.L.; Tavazzi, L.; et al. Tubular damage and worsening renal function in chronic heart failure. *JACC Heart Fail.* **2013**, *1*, 417–424. [[CrossRef](#)]
8. Haase, M.; Muller, C.; Damman, K.; Murray, P.T.; Kellum, J.A.; Ronco, C.; McCullough, P.A. Pathogenesis of cardiorenal syndrome type 1 in acute decompensated heart failure: Workgroup statements from the eleventh consensus conference of the Acute Dialysis Quality Initiative (ADQI). *Contrib. Nephrol.* **2013**, *182*, 99–116.
9. Ronsein, G.E.; Vaisar, T. Deepening our understanding of HDL proteome. *Expert Rev. Proteom.* **2019**, *16*, 749–760. [[CrossRef](#)]
10. Navab, M.; Anantharamaiah, G.M.; Reddy, S.T.; Van Lenten, B.J.; Fogelman, A.M. HDL as a biomarker, potential therapeutic target, and therapy. *Diabetes* **2009**, *58*, 2711–2717. [[CrossRef](#)]
11. Vaziri, N.D.; Navab, M.; Fogelman, A.M. HDL metabolism and activity in chronic kidney disease. *Nat. Rev. Nephrol.* **2010**, *6*, 287–296. [[CrossRef](#)] [[PubMed](#)]
12. Fisher, E.A.; Feig, J.E.; Hewing, B.; Hazen, S.L.; Smith, J.D. High-density lipoprotein function, dysfunction, and reverse cholesterol transport. *Arterioscler. Thromb. Vasc. Biol.* **2012**, *32*, 2813–2820. [[CrossRef](#)] [[PubMed](#)]
13. Imaizumi, S.; Navab, M.; Morgantini, C.; Charles-Schoeman, C.; Su, F.; Gao, F.; Kwon, M.; Ganapathy, E.; Meriwether, D.; Farias-Eisner, R.; et al. Dysfunctional high-density lipoprotein and the potential of apolipoprotein A-1 mimetic peptides to normalize the composition and function of lipoproteins. *Circ. J.* **2011**, *75*, 1533–1538. [[CrossRef](#)] [[PubMed](#)]
14. Sherman, C.B.; Peterson, S.J.; Frishman, W.H. Apolipoprotein A-I mimetic peptides: A potential new therapy for the prevention of atherosclerosis. *Cardiol. Rev.* **2010**, *18*, 141–147. [[CrossRef](#)]
15. Navab, M.; Anantharamaiah, G.M.; Reddy, S.T.; Van Lenten, B.J.; Ansell, B.J.; Hama, S.; Hough, G.; Bachini, E.; Grijalva, V.R.; Wagner, A.C.; et al. The double jeopardy of HDL. *Ann. Med.* **2005**, *37*, 173–178. [[CrossRef](#)]
16. Kennedy, D.J.; Tang, W.H.; Fan, Y.; Wu, Y.; Mann, S.; Pepoy, M.; Hazen, S.L. Diminished antioxidant activity of high-density lipoprotein-associated proteins in chronic kidney disease. *J. Am. Heart Assoc.* **2013**, *2*, e000104. [[CrossRef](#)]
17. Boemi, M.; Leviev, I.; Sirolla, C.; Pieri, C.; Marra, M.; James, R.W. Serum paraoxonase is reduced in type 1 diabetic patients compared to non-diabetic, first degree relatives; influence on the ability of HDL to protect LDL from oxidation. *Atherosclerosis* **2001**, *155*, 229–235. [[CrossRef](#)]
18. Tall, A.R. Cholesterol efflux pathways and other potential mechanisms involved in the athero-protective effect of high density lipoproteins. *J. Intern. Med.* **2008**, *263*, 256–273. [[CrossRef](#)]
19. Odegaard, J.I.; Chawla, A. Old HDL learns a new (anti-inflammatory) trick. *Nat. Immunol.* **2014**, *15*, 138–139. [[CrossRef](#)]
20. Kotosai, M.; Shimada, S.; Kanda, M.; Matsuda, N.; Sekido, K.; Shimizu, Y.; Tokumura, A.; Nakamura, T.; Murota, K.; Kawai, Y.; et al. Plasma HDL reduces nonesterified fatty acid hydroperoxides originating from oxidized LDL: A mechanism for its antioxidant ability. *Lipids* **2013**, *48*, 569–578. [[CrossRef](#)]
21. Berrougui, H.; Momo, C.N.; Khalil, A. Health benefits of high-density lipoproteins in preventing cardiovascular diseases. *J. Clin. Lipidol.* **2012**, *6*, 524–533. [[CrossRef](#)] [[PubMed](#)]
22. Chang, C.T.; Lim, Y.P.; Lee, C.W.; Liao, H.Y.; Chen, F.Y.; Chang, C.M.; Tang, F.Y.; Yang, C.Y.; Chen, C.J. PON-1 carbamylation is enhanced in HDL of uremia patients. *J. Food Drug. Anal.* **2019**, *27*, 542–550. [[CrossRef](#)] [[PubMed](#)]

23. Vaisar, T.; Kanter, J.E.; Wimberger, J.; Irwin, A.D.; Gauthier, J.; Wolfson, E.; Bahnam, V.; Wu, I.H.; Shah, H.; Keenan, H.A.; et al. High Concentration of Medium-Sized HDL Particles and Enrichment in HDL Paraoxonase 1 Associate with Protection From Vascular Complications in People with Long-standing Type 1 Diabetes. *Diabetes Care* **2020**, *43*, 178–186. [CrossRef] [PubMed]
24. Pan, B.; Yu, B.; Ren, H.; Willard, B.; Pan, L.; Zu, L.; Shen, X.; Ma, Y.; Li, X.; Niu, C.; et al. High-density lipoprotein nitration and chlorination catalyzed by myeloperoxidase impair its effect of promoting endothelial repair. *Free Radic. Biol. Med.* **2013**, *60*, 272–281. [CrossRef] [PubMed]
25. Proudfoot, J.M.; Barden, A.E.; Loke, W.M.; Croft, K.D.; Puddey, I.B.; Mori, T.A. HDL is the major lipoprotein carrier of plasma F2-isoprostanes. *J. Lipid Res.* **2009**, *50*, 716–722. [CrossRef] [PubMed]
26. Nicholls, S.J.; Zheng, L.; Hazen, S.L. Formation of dysfunctional high-density lipoprotein by myeloperoxidase. *Trends Cardiovasc. Med.* **2005**, *15*, 212–219. [CrossRef] [PubMed]
27. Vazzana, N.; Ganci, A.; Cefalu, A.B.; Lattanzio, S.; Noto, D.; Santoro, N.; Saggini, R.; Puccetti, L.; Averna, M.; Davi, G. Enhanced lipid peroxidation and platelet activation as potential contributors to increased cardiovascular risk in the low-HDL phenotype. *J. Am. Heart Assoc.* **2013**, *2*, e000063. [CrossRef]
28. Cazzola, R.; Cassani, E.; Barichella, M.; Cestaro, B. Impaired fluidity and oxidizability of HDL hydrophobic core and amphipathic surface in dyslipidemic men. *Metabolism* **2013**, *62*, 986–991. [CrossRef]
29. Puchades, M.J.; Saez, G.; Munoz, M.C.; Gonzalez, M.; Torregrosa, I.; Juan, I.; Miguel, A. Study of oxidative stress in patients with advanced renal disease and undergoing either hemodialysis or peritoneal dialysis. *Clin. Nephrol.* **2013**, *80*, 177–186. [CrossRef]
30. Montuschi, P.; Barnes, P.J.; Roberts, L.J. Isoprostanes: Markers and mediators of oxidative stress. *FASEB J.* **2004**, *18*, 1791–1800. [CrossRef]
31. Montuschi, P.; Barnes, P.; Roberts, L.J. Insights into oxidative stress: The isoprostanes. *Curr. Med. Chem.* **2007**, *14*, 703–717. [CrossRef] [PubMed]
32. Navab, M.; Reddy, S.T.; Van Lenten, B.J.; Anantharamaiah, G.M.; Fogelman, A.M. The role of dysfunctional HDL in atherosclerosis. *J. Lipid Res.* **2009**, *50*, S145–S149. [CrossRef] [PubMed]
33. Ansell, B.J.; Fonarow, G.C.; Fogelman, A.M. The paradox of dysfunctional high-density lipoprotein. *Curr. Opin. Lipidol.* **2007**, *18*, 427–434. [CrossRef] [PubMed]
34. Rhainds, D.; Tardif, J.C. From HDL-cholesterol to HDL-function: Cholesterol efflux capacity determinants. *Curr. Opin. Lipidol.* **2019**, *30*, 101–107. [CrossRef]
35. He, D.; Zhao, M.; Wu, C.; Zhang, W.; Niu, C.; Yu, B.; Jin, J.; Ji, L.; Willard, B.; Mathew, A.V.; et al. Apolipoprotein A-1 mimetic peptide 4F promotes endothelial repairing and compromises reendothelialization impaired by oxidized HDL through SR-B1. *Redox Biol.* **2018**, *15*, 228–242. [CrossRef]
36. Wilkins, J.T.; Seckler, H.S. HDL modification: Recent developments and their relevance to atherosclerotic cardiovascular disease. *Curr. Opin. Lipidol.* **2019**, *30*, 24–29. [CrossRef]
37. Cao, J.; Sodhi, K.; Inoue, K.; Quilley, J.; Rezzani, R.; Rodella, L.; Vanella, L.; Germinario, L.; Stec, D.E.; Abraham, N.G.; et al. Lentiviral-human heme oxygenase targeting endothelium improved vascular function in angiotensin II animal model of hypertension. *Hum. Gene Ther.* **2011**, *22*, 271–282. [CrossRef]
38. Zager, R.A. Uremia induces proximal tubular cytoresistance and heme oxygenase-1 expression in the absence of acute kidney injury. *Am. J. Physiol. Renal Physiol.* **2009**, *296*, F362–F368. [CrossRef]
39. Ghattas, M.H.; Chuang, L.T.; Kappas, A.; Abraham, N.G. Protective effect of HO-1 against oxidative stress in human hepatoma cell line (HepG2) is independent of telomerase enzyme activity. *Int. J. Biochem. Cell Biol.* **2002**, *34*, 1619–1628. [CrossRef]
40. Goodman, A.I.; Quan, S.; Yang, L.; Synghal, A.; Abraham, N.G. Functional expression of human heme oxygenase-1 gene in renal structure of spontaneously hypertensive rats. *Exp. Biol. Med.* **2003**, *228*, 454–458. [CrossRef]
41. Abraham, N.G.; Kappas, A. Heme oxygenase and the cardiovascular-renal system. *Free Radic. Biol. Med.* **2005**, *39*, 1–25. [CrossRef]
42. Vanella, L.; Kim, D.H.; Sodhi, K.; Barbagallo, I.; Burgess, A.P.; Falck, J.R.; Schwartzman, M.L.; Abraham, N.G. Crosstalk between EET and HO-1 downregulates Bach1 and adipogenic marker expression in mesenchymal stem cell derived adipocytes. *Prostaglandins Other Lipid Mediat.* **2011**, *96*, 54–62. [CrossRef]

43. Kim, D.H.; Burgess, A.P.; Li, M.; Tsenovoy, P.L.; Addabbo, F.; McClung, J.A.; Puri, N.; Abraham, N.G. Heme oxygenase-mediated increases in adiponectin decrease fat content and inflammatory cytokines, tumor necrosis factor-alpha and interleukin-6 in Zucker rats and reduce adipogenesis in human mesenchymal stem cells. *J. Pharmacol. Exp. Ther.* **2008**, *325*, 833–840. [CrossRef]
44. Abraham, N.G.; Tsenovoy, P.L.; McClung, J.; Drummond, G.S. Heme oxygenase: A target gene for anti-diabetic and obesity. *Curr. Pharm. Des.* **2008**, *14*, 412–421. [CrossRef]
45. Abraham, N.G. Heme oxygenase attenuated angiotensin II-mediated increase in cyclooxygenase activity and decreased isoprostane F2alpha in endothelial cells. *Thromb. Res.* **2003**, *110*, 305–309. [CrossRef]
46. Peterson, S.J.; Drummond, G.; Kim, D.H.; Li, M.; Kruger, A.L.; Ikehara, S.; Abraham, N.G. L-4F treatment reduces adiposity, increases adiponectin levels, and improves insulin sensitivity in obese mice. *J. Lipid Res.* **2008**, *49*, 1658–1669. [CrossRef]
47. Vanella, L.; Li, V.G.; Guccione, S.; Rappazzo, G.; Salvo, E.; Pappalardo, M.; Schwartzman, M.L.; Abraham, N.G. Heme oxygenase-2/adiponectin protein-protein interaction in metabolic syndrome. *Biochem. Biophys. Res. Commun.* **2013**, *432*, 606–611. [CrossRef]
48. Khitan, Z.; Harsh, M.; Sodhi, K.; Shapiro, J.I.; Abraham, N.G. HO-1 Upregulation Attenuates Adipocyte Dysfunction, Obesity, and Isoprostanate Levels in Mice Fed High Fructose Diets. *J. Nutr. Metab.* **2014**, *2014*, 980547. [CrossRef]
49. Femlak, M.; Gluba-Brzozka, A.; Franczyk, B.; Rysz, J. Diabetes-induced Alterations in HDL Subfractions Distribution. *Curr. Pharm. Des.* **2020**, *26*, 3341–3348. [CrossRef]
50. Hui, X.; Lam, K.S.; Vanhoutte, P.M.; Xu, A. Adiponectin and cardiovascular health: An update. *Br. J. Pharmacol.* **2012**, *165*, 574–590. [CrossRef]
51. Fang, F.; Liu, G.C.; Kim, C.; Yassa, R.; Zhou, J.; Scholey, J.W. Adiponectin attenuates angiotensin II-induced oxidative stress in renal tubular cells through AMPK and cAMP-Epac signal transduction pathways. *Am. J. Physiol. Renal Physiol.* **2013**, *304*, F1366–F1374. [CrossRef]
52. Rutkowski, J.M.; Wang, Z.V.; Park, A.S.; Zhang, J.; Zhang, D.; Hu, M.C.; Moe, O.W.; Susztak, K.; Scherer, P.E. Adiponectin promotes functional recovery after podocyte ablation. *J. Am. Soc. Nephrol.* **2013**, *24*, 268–282. [CrossRef]
53. Cheng, C.F.; Lian, W.S.; Chen, S.H.; Lai, P.F.; Li, H.F.; Lan, Y.F.; Cheng, W.T.; Lin, H. Protective effects of adiponectin against renal ischemia-reperfusion injury via prostacyclin-PPARalpha-heme oxygenase-1 signaling pathway. *J. Cell. Physiol.* **2011**, *227*, 239–249. [CrossRef]
54. Lo, M.M.; Salisbury, S.; Scherer, P.E.; Furth, S.L.; Warady, B.A.; Mitsnefes, M.M. Serum adiponectin complexes and cardiovascular risk in children with chronic kidney disease. *Pediatr. Nephrol.* **2011**, *26*, 2009–2017. [CrossRef]
55. Abraham, N.G.; Asija, A.; Drummond, G.; Peterson, S. Heme oxygenase-1 gene therapy: Recent advances and therapeutic applications. *Curr. Gene Ther.* **2007**, *7*, 89–108. [CrossRef]
56. Monu, S.R.; Pesce, P.; Sodhi, K.; Boldrin, M.; Puri, N.; Fedorova, L.; Sacerdoti, D.; Peterson, S.J.; Abraham, N.G.; Kappas, A. HO-1 induction improves the type-1 cardiorenal syndrome in mice with impaired angiotensin II-induced lymphocyte activation. *Hypertension* **2013**, *62*, 310–316. [CrossRef]
57. Burstein, M.; Scholnick, H.R.; Morfin, R. Rapid method for the isolation of lipoproteins from human serum by precipitation with polyanions. *J. Lipid Res.* **1970**, *11*, 583–595.
58. Rader, D.J. Molecular regulation of HDL metabolism and function: Implications for novel therapies. *J. Clin. Investig.* **2006**, *116*, 3090–3100. [CrossRef]
59. Kajani, S.; Curley, S.; McGillicuddy, F.C. Unravelling HDL—Looking beyond the Cholesterol Surface to the Quality within. *Int. J. Mol. Sci.* **2018**, *19*, 1971. [CrossRef]
60. Schwertani, A.; Choi, H.Y.; Genest, J. HDLs and the pathogenesis of atherosclerosis. *Curr. Opin. Cardiol.* **2018**, *33*, 311–316. [CrossRef]
61. Shao, B.; Heinecke, J.W. Quantifying HDL proteins by mass spectrometry: How many proteins are there and what are their functions? *Expert Rev. Proteom.* **2018**, *15*, 31–40. [CrossRef] [PubMed]
62. Asztalos, B.F.; Niisuke, K.; Horvath, K.V. High-density lipoprotein: Our elusive friend. *Curr. Opin. Lipidol.* **2019**, *30*, 314–319. [CrossRef] [PubMed]
63. Furtado, J.D.; Yamamoto, R.; Melchior, J.T.; Andraski, A.B.; Gamez-Guerrero, M.; Mulcahy, P.; He, Z.; Cai, T.; Davidson, W.S.; Sacks, F.M. Distinct Proteomic Signatures in 16 HDL (High-Density Lipoprotein) Subspecies. *Arterioscler. Thromb. Vasc. Biol.* **2018**, *38*, 2827–2842. [CrossRef] [PubMed]

64. Chang, C.T.; Yang, C.Y.; Tsai, F.J.; Lin, S.Y.; Chen, C.J. Mass Spectrometry-Based Proteomic Study Makes High-Density Lipoprotein a Biomarker for Atherosclerotic Vascular Disease. *Biomed. Res. Int.* **2015**, *2015*, 164846. [CrossRef] [PubMed]
65. Ng, L.L.; Voors, A.A. Quantifying the HDL proteome by mass spectrometry: How many proteins truly associate with HDL? REPLY to Marsche. *Eur. J. Heart Fail.* **2018**, *20*, 1077–1078. [CrossRef] [PubMed]
66. Marsche, G. Quantifying the HDL proteome by mass spectrometry: How many proteins truly associate with HDL? *Eur. J. Heart Fail.* **2018**, *20*, 1077. [CrossRef] [PubMed]
67. Zhang, Y.; Gordon, S.M.; Xi, H.; Choi, S.; Paz, M.A.; Sun, R.; Yang, W.; Saredy, J.; Khan, M.; Remaley, A.T.; et al. HDL subclass proteomic analysis and functional implication of protein dynamic change during HDL maturation. *Redox Biol.* **2019**, *24*, 101222. [CrossRef] [PubMed]
68. Gordon, S.M.; Chung, J.H.; Playford, M.P.; Dey, A.K.; Sviridov, D.; Seifuddin, F.; Chen, Y.C.; Pirooznia, M.; Chen, M.Y.; Mehta, N.N.; et al. High density lipoprotein proteome is associated with cardiovascular risk factors and atherosclerosis burden as evaluated by coronary CT angiography. *Atherosclerosis* **2018**, *278*, 278–285. [CrossRef] [PubMed]
69. Emmens, J.E.; Jones, D.J.L.; Cao, T.H.; Chan, D.C.S.; Romaine, S.P.R.; Quinn, P.A.; Anker, S.D.; Cleland, J.G.; Dickstein, K.; Filippatos, G.; et al. Proteomic diversity of high-density lipoprotein explains its association with clinical outcome in patients with heart failure. *Eur. J. Heart Fail.* **2018**, *20*, 260–267. [CrossRef] [PubMed]
70. Gourgari, E.; Ma, J.; Playford, M.P.; Mehta, N.N.; Goldman, R.; Remaley, A.T.; Gordon, S.M. Proteomic alterations of HDL in youth with type 1 diabetes and their associations with glycemic control: A case-control study. *Cardiovasc. Diabetol.* **2019**, *18*, 43. [CrossRef] [PubMed]
71. Manjunatha, S.; Distelmaier, K.; Dasari, S.; Carter, R.E.; Kudva, Y.C.; Nair, K.S. Functional and proteomic alterations of plasma high density lipoproteins in type 1 diabetes mellitus. *Metabolism* **2016**, *65*, 1421–1431. [CrossRef] [PubMed]
72. Phillips, M.C. New insights into the determination of HDL structure by apolipoproteins: Thematic review series: High density lipoprotein structure, function, and metabolism. *J. Lipid Res.* **2013**, *54*, 2034–2048. [CrossRef] [PubMed]
73. Shah, P.K.; Kaul, S.; Nilsson, J.; Cersek, B. Exploiting the vascular protective effects of high-density lipoprotein and its apolipoproteins: An idea whose time for testing is coming, part I. *Circulation* **2001**, *104*, 2376–2383. [CrossRef] [PubMed]
74. Tall, A.R. An overview of reverse cholesterol transport. *Eur. Heart J.* **1998**, *19* (Suppl. A), A31–A35. [CrossRef]
75. Fielding, C.J.; Fielding, P.E. Molecular physiology of reverse cholesterol transport. *J. Lipid Res.* **1995**, *36*, 211–228.
76. Brufau, G.; Groen, A.K.; Kuipers, F. Reverse cholesterol transport revisited: Contribution of biliary versus intestinal cholesterol excretion. *Arterioscler. Thromb. Vasc. Biol.* **2011**, *31*, 1726–1733. [CrossRef]
77. Kanungo, S.; Soares, N.; He, M.; Steiner, R.D. Sterol metabolism disorders and neurodevelopment—an update. *Dev. Disabil. Res. Rev.* **2013**, *17*, 197–210. [CrossRef]
78. DiDonato, J.A.; Huang, Y.; Aulak, K.S.; Even-Or, O.; Gerstenecker, G.; Gogonea, V.; Wu, Y.; Fox, P.L.; Tang, W.H.; Plow, E.F.; et al. Function and distribution of apolipoprotein A1 in the artery wall are markedly distinct from those in plasma. *Circulation* **2013**, *128*, 1644–1655. [CrossRef]
79. Huang, Y.; DiDonato, J.A.; Levison, B.S.; Schmitt, D.; Li, L.; Wu, Y.; Buffa, J.; Kim, T.; Gerstenecker, G.S.; Gu, X.; et al. An abundant dysfunctional apolipoprotein A1 in human atheroma. *Nat. Med.* **2014**, *20*, 193–203. [CrossRef]
80. Sevugan, C.P.; Mayne, L.; Kan, Z.Y.; Lund-Katz, S.; Englander, S.W.; Phillips, M.C. Apolipoprotein A-I helical structure and stability in discoidal high-density lipoprotein (HDL) particles by hydrogen exchange and mass spectrometry. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 11687–11692. [CrossRef]
81. Brewer, H.B., Jr. High-density lipoproteins: A new potential therapeutic target for the prevention of cardiovascular disease. *Arterioscler. Thromb. Vasc. Biol.* **2004**, *24*, 387–391. [CrossRef] [PubMed]
82. Neufeld, E.B.; Sato, M.; Gordon, S.M.; Durbakhula, V.; Francone, N.; Aponte, A.; Yilmaz, G.; Sviridov, D.; Sampson, M.; Tang, J.; et al. ApoA-I-Mediated Lipoprotein Remodeling Monitored with a Fluorescent Phospholipid. *Biology* **2019**, *8*, 53. [CrossRef] [PubMed]
83. Kang, M.H.; Zhang, L.H.; Wijesekara, N.; de Haan, W.; Butland, S.; Bhattacharjee, A.; Hayden, M.R. Regulation of ABCA1 protein expression and function in hepatic and pancreatic islet cells by miR-145. *Arterioscler. Thromb. Vasc. Biol.* **2013**, *33*, 2724–2732. [CrossRef] [PubMed]

84. He, Y.; Ronsein, G.E.; Tang, C.; Jarvik, G.P.; Davidson, W.S.; Kothari, V.; Song, H.D.; Segrest, J.P.; Bornfeldt, K.E.; Heinecke, J.W. Diabetes Impairs Cellular Cholesterol Efflux From ABCA1 to Small HDL Particles. *Circ. Res.* **2020**, *127*, 1198–1210. [CrossRef] [PubMed]
85. Yokoyama, S. ABCA1 and biogenesis of HDL. *J. Atheroscler. Thromb.* **2006**, *13*, 1–15. [CrossRef]
86. Pussinen, P.J.; Jauhainen, M.; Metso, J.; Pyle, L.E.; Marcel, Y.L.; Fidge, N.H.; Ehnholm, C. Binding of phospholipid transfer protein (PLTP) to apolipoproteins A-I and A-II: Location of a PLTP binding domain in the amino terminal region of apoA-I. *J. Lipid Res.* **1998**, *39*, 152–161.
87. Oberbach, A.; Adams, V.; Schlichting, N.; Heinrich, M.; Kullnick, Y.; Lehmann, S.; Lehmann, S.; Feder, S.; Correia, J.C.; Mohr, F.W.; et al. Proteome profiles of HDL particles of patients with chronic heart failure are associated with immune response and also include bacteria proteins. *Clin. Chim. Acta* **2016**, *453*, 114–122. [CrossRef]
88. Rousset, X.; Shamburek, R.; Vaisman, B.; Amar, M.; Remaley, A.T. Lecithin cholesterol acyltransferase: An anti- or pro-atherogenic factor? *Curr. Atheroscler. Rep.* **2011**, *13*, 249–256. [CrossRef]
89. Martinez, L.R.; Santos, R.D.; Miname, M.H.; Deus, D.F.; Lima, E.S.; Maranhao, R.C. Transfer of lipids to high-density lipoprotein (HDL) is altered in patients with familial hypercholesterolemia. *Metabolism* **2013**, *62*, 1061–1064. [CrossRef]
90. Duka, A.; Fotakis, P.; Georgiadou, D.; Katefides, A.; Tzavlaiki, K.; von, E.L.; Stratikos, E.; Kardassis, D.; Zannis, V.I. ApoA-IV promotes the biogenesis of apoA-IV-containing HDL particles with the participation of ABCA1 and LCAT. *J. Lipid Res.* **2013**, *54*, 107–115. [CrossRef]
91. Kuivenhoven, J.A.; Pritchard, H.; Hill, J.; Frohlich, J.; Assmann, G.; Kastelein, J. The molecular pathology of lecithin:cholesterol acyltransferase (LCAT) deficiency syndromes. *J. Lipid Res.* **1997**, *38*, 191–205. [PubMed]
92. Undurti, A.; Huang, Y.; Lupica, J.A.; Smith, J.D.; DiDonato, J.A.; Hazen, S.L. Modification of high density lipoprotein by myeloperoxidase generates a pro-inflammatory particle. *J. Biol. Chem.* **2009**, *284*, 30825–30835. [CrossRef] [PubMed]
93. Peter, B.; Bibi, S.; Eisenwort, G.; Wingelhofer, B.; Berger, D.; Stefanl, G.; Blatt, K.; Herrmann, H.; Hadzijusufovic, E.; Hoermann, G.; et al. Drug-induced inhibition of phosphorylation of STAT5 overrides drug resistance in neoplastic mast cells. *Leukemia* **2018**, *32*, 1016–1022. [CrossRef] [PubMed]
94. Kappelle, P.J.; Gansevoort, R.T.; Hillege, H.J.; Wolffenbuttel, B.H.; Dullaart, R.P. Common variation in cholestryler ester transfer protein: Relationship of first major adverse cardiovascular events with the apolipoprotein B/apolipoprotein A-I ratio and the total cholesterol/high-density lipoprotein cholesterol ratio. *J. Clin. Lipidol.* **2013**, *7*, 56–64. [CrossRef] [PubMed]
95. Morton, R.E.; Liu, Y. The lipid transfer properties of CETP define the concentration and composition of plasma lipoproteins. *J. Lipid Res.* **2020**, *61*, 1168–1179. [CrossRef] [PubMed]
96. Leusink, M.; Onland-Moret, N.C.; Asselbergs, F.W.; Ding, B.; Kotti, S.; van Zuydam, N.R.; Papp, A.C.; Danchin, N.; Donnelly, L.; Morris, A.D.; et al. Cholestryler ester transfer protein polymorphisms, statin use, and their impact on cholesterol levels and cardiovascular events. *Clin. Pharmacol. Ther.* **2014**, *95*, 314–320. [CrossRef]
97. Do, H.Q.; Nazih, H.; Luc, G.; Arveiler, D.; Ferrieres, J.; Evans, A.; Amouyel, P.; Cambien, F.; Ducimetiere, P.; Bard, J.M. Influence of cholestryler ester transfer protein, peroxisome proliferator-activated receptor alpha, apolipoprotein E, and apolipoprotein A-I polymorphisms on high-density lipoprotein cholesterol, apolipoprotein A-I, lipoprotein A-I, and lipoprotein A-I:A-II concentrations: The Prospective Epidemiological Study of Myocardial Infarction study. *Metabolism* **2009**, *58*, 283–289.
98. Uehara, Y.; Miura, S.; von Eckardstein, A.; Abe, S.; Fujii, A.; Matsuo, Y.; Rust, S.; Lorkowski, S.; Assmann, G.; Yamada, T.; et al. Unsaturated fatty acids suppress the expression of the ATP-binding cassette transporter G1 (ABCG1) and ABCA1 genes via an LXR/RXR responsive element. *Atherosclerosis* **2007**, *191*, 11–21. [CrossRef]
99. Kannel, W.B.; Wilson, P.W. Risk factors that attenuate the female coronary disease advantage. *Arch. Intern. Med.* **1995**, *155*, 57–61. [CrossRef]
100. Jin, H.; Nicodemus-Johnson, J. Gender and Age Stratified Analyses of Nutrient and Dietary Pattern Associations with Circulating Lipid Levels Identify Novel Gender and Age-Specific Correlations. *Nutrients* **2018**, *10*, 1760. [CrossRef]
101. Goossens, G.H.; Jocken, J.W.E.; Blaak, E.E. Sexual dimorphism in cardiometabolic health: The role of adipose tissue, muscle and liver. *Nat. Rev. Endocrinol.* **2020**. [CrossRef] [PubMed]

102. Chan, E.J.; Dellasperger, K.C. Cardiorenal Syndrome: The Clinical Cardiologists' Perspective. *Cardiorenal. Med.* **2011**, *1*, 13–22. [CrossRef] [PubMed]
103. Virzi, G.M.; Day, S.; de, C.M.; Vescovo, G.; Ronco, C. Heart-kidney crosstalk and role of humoral signaling in critical illness. *Crit. Care* **2014**, *18*, 201. [CrossRef] [PubMed]
104. Schroten, N.F.; Gaillard, C.A.; van Veldhuisen, D.J.; Szymanski, M.K.; Hillege, H.L.; de Boer, R.A. New roles for renin and prorenin in heart failure and cardiorenal crosstalk. *Heart Fail. Rev.* **2012**, *17*, 191–201. [CrossRef] [PubMed]
105. Briet, M.; Schiffrin, E.L. Aldosterone: Effects on the kidney and cardiovascular system. *Nat. Rev. Nephrol.* **2010**, *6*, 261–273. [CrossRef] [PubMed]
106. Muhlberger, I.; Monks, K.; Fechete, R.; Mayer, G.; Oberbauer, R.; Mayer, B.; Perco, P. Molecular pathways and crosstalk characterizing the cardiorenal syndrome. *OMICS* **2012**, *16*, 105–112. [CrossRef] [PubMed]
107. Foley, R.N.; Parfrey, P.S.; Sarnak, M.J. Clinical epidemiology of cardiovascular disease in chronic renal disease. *Am. J. Kidney Dis.* **1998**, *32* (Suppl. 3), S112–S119. [CrossRef]
108. Cruz, D.N. Cardiorenal syndrome in critical care: The acute cardiorenal and renocardiac syndromes. *Adv. Chronic Kidney Dis.* **2013**, *20*, 56–66. [CrossRef]
109. Bongartz, L.G.; Cramer, M.J.; Doevedans, P.A.; Joles, J.A.; Braam, B. The severe cardiorenal syndrome: 'Guyton revisited'. *Eur. Heart J.* **2005**, *26*, 11–17. [CrossRef]
110. McCullough, P.A.; Kellum, J.A.; Haase, M.; Muller, C.; Damman, K.; Murray, P.T.; Cruz, D.; House, A.A.; Schmidt-Ott, K.M.; Vescovo, G.; et al. Pathophysiology of the cardiorenal syndromes: Executive summary from the eleventh consensus conference of the Acute Dialysis Quality Initiative (ADQI). *Contrib. Nephrol.* **2013**, *182*, 82–98.
111. House, A.A.; Haapio, M.; Lassus, J.; Bellomo, R.; Ronco, C. Therapeutic strategies for heart failure in cardiorenal syndromes. *Am. J. Kidney Dis.* **2010**, *56*, 759–773. [CrossRef] [PubMed]
112. Costanzo, M.R. The Cardiorenal Syndrome in Heart Failure. *Heart Fail. Clin.* **2020**, *16*, 81–97. [CrossRef] [PubMed]
113. Ronco, C.; Bellasi, A.; Di Lullo, L. Cardiorenal Syndrome: An Overview. *Adv. Chronic Kidney Dis.* **2018**, *25*, 382–390. [CrossRef] [PubMed]
114. Funahashi, Y.; Chowdhury, S.; Eiwaz, M.B.; Hutchens, M.P. Acute Cardiorenal Syndrome: Models and Heart-Kidney Connectors. *Nephron* **2020**, *1–5*. [CrossRef] [PubMed]
115. Zannad, F.; Rossignol, P. Cardiorenal Syndrome Revisited. *Circulation* **2018**, *138*, 929–944. [CrossRef]
116. Raina, R.; Nair, N.; Chakraborty, R.; Nemer, L.; Dasgupta, R.; Varian, K. An Update on the Pathophysiology and Treatment of Cardiorenal Syndrome. *Cardiol. Res.* **2020**, *11*, 76–88. [CrossRef]
117. Lam, P.H.; Dooley, D.J.; Fonarow, G.C.; Butler, J.; Bhatt, D.L.; Filippatos, G.S.; Deedwania, P.; Forman, D.E.; White, M.; Fletcher, R.D.; et al. Similar clinical benefits from below-target and target dose enalapril in patients with heart failure in the SOLVD Treatment trial. *Eur. J. Heart Fail.* **2018**, *20*, 359–369. [CrossRef]
118. Lam, P.H.; Packer, M.; Fonarow, G.C.; Faselis, C.; Allman, R.M.; Morgan, C.J.; Singh, S.N.; Pitt, B.; Ahmed, A. Early Effects of Starting Doses of Enalapril in Patients with Chronic Heart Failure in the SOLVD Treatment Trial. *Am. J. Med.* **2020**, *133*, e25–e31. [CrossRef]
119. de Boer, I.H.; Caramori, M.L.; Chan, J.C.N.; Heerspink, H.J.L.; Hurst, C.; Khunti, K.; Liew, A.; Michos, E.D.; Navaneethan, S.D.; Olowu, W.A.; et al. Executive summary of the 2020 KDIGO Diabetes Management in CKD Guideline: Evidence-based advances in monitoring and treatment. *Kidney Int.* **2020**, *98*, 839–848. [CrossRef]
120. Kidney Disease: Improving Global Outcomes (KDIGO) Diabetes Work Group. KDIGO 2020 Clinical Practice Guideline for Diabetes Management in Chronic Kidney Disease. *Kidney Int.* **2020**, *98*, S1–S115. [CrossRef]
121. Kumar, U.; Wettersten, N.; Garimella, P.S. Cardiorenal Syndrome: Pathophysiology. *Cardiol. Clin.* **2019**, *37*, 251–265. [CrossRef] [PubMed]
122. Kotecha, A.; Vallabhajosyula, S.; Coville, H.H.; Kashani, K. Cardiorenal syndrome in sepsis: A narrative review. *J. Crit. Care* **2018**, *43*, 122–127. [CrossRef] [PubMed]
123. Fried, J.A.; Ramasubbu, K.; Bhatt, R.; Topkara, V.K.; Clerkin, K.J.; Horn, E.; Rabbani, L.; Brodie, D.; Jain, S.S.; Kirtane, A.J.; et al. The Variety of Cardiovascular Presentations of COVID-19. *Circulation* **2020**, *141*, 1930–1936. [CrossRef] [PubMed]
124. Yan, Y.; Yang, Y.; Wang, F.; Ren, H.; Zhang, S.; Shi, X.; Yu, X.; Dong, K. Clinical characteristics and outcomes of patients with severe covid-19 with diabetes. *BMJ Open Diabetes Res. Care* **2020**, *8*. [CrossRef] [PubMed]

125. Pei, G.; Zhang, Z.; Peng, J.; Liu, L.; Zhang, C.; Yu, C.; Ma, Z.; Huang, Y.; Liu, W.; Yao, Y.; et al. Renal Involvement and Early Prognosis in Patients with COVID-19 Pneumonia. *J. Am. Soc. Nephrol.* **2020**, *31*, 1157–1165. [[CrossRef](#)]
126. Apetrii, M.; Enache, S.; Siriopol, D.; Burlacu, A.; Kanbay, A.; Kanbay, M.; Scripcariu, D.; Covic, A. A brand-new cardiorenal syndrome in the COVID-19 setting. *Clin. Kidney J.* **2020**, *13*, 291–296.
127. Sankritayayan, H.; Kale, A.; Sharma, N.; Anders, H.J.; Gaikwad, A.B. Evidence for Use or Disuse of Renin-Angiotensin System Modulators in Patients Having COVID-19 with an Underlying Cardiorenal Disorder. *J. Cardiovasc. Pharmacol. Ther.* **2020**, *25*, 299–306. [[CrossRef](#)]
128. Clementi, A.; Virzi, G.M.; Battaglia, G.G.; Ronco, C. Neurohormonal, Endocrine, and Immune Dysregulation and Inflammation in Cardiorenal Syndrome. *Cardiorenal. Med.* **2019**, *9*, 265–273. [[CrossRef](#)]
129. Savira, F.; Magaye, R.; Liew, D.; Reid, C.; Kelly, D.J.; Kompa, A.R.; Sangaralingham, S.J.; Burnett, J.C., Jr.; Kaye, D.; Wang, B.H. Cardiorenal syndrome: Multi-organ dysfunction involving the heart, kidney and vasculature. *Br. J. Pharmacol.* **2020**, *177*, 2906–2922. [[CrossRef](#)]
130. Kingma, J.G.; Simard, D.; Rouleau, J.R.; Drolet, B.; Simard, C. The Physiopathology of Cardiorenal Syndrome: A Review of the Potential Contributions of Inflammation. *J. Cardiovasc. Dev. Dis.* **2017**, *4*, 21. [[CrossRef](#)]
131. Hou, J.; Xie, X.; Tu, Q.; Li, J.; Ding, J.; Shao, G.; Jiang, Q.; Yuan, L.; Lai, X. SIRT1 gene polymorphisms are associated with nondiabetic type 1 cardiorenal syndrome. *Ann. Hum. Genet.* **2019**, *83*, 445–453. [[CrossRef](#)] [[PubMed](#)]
132. Virzi, G.M.; Clementi, A.; de Cal, M.; Brocca, A.; Day, S.; Pastori, S.; Bolin, C.; Vescovo, G.; Ronco, C. Oxidative stress: Dual pathway induction in cardiorenal syndrome type 1 pathogenesis. *Oxid. Med. Cell. Longev.* **2015**, *2015*, 391790. [[CrossRef](#)] [[PubMed](#)]
133. Elmarakby, A.A.; Imig, J.D. Obesity is the major contributor to vascular dysfunction and inflammation in high fat diet hypertensive rats. *Clin. Sci.* **2010**, *118*, 291–301. [[CrossRef](#)] [[PubMed](#)]
134. Pesce, P.; Cecchetto, L.; Brocco, S.; Bolognesi, M.; Sodhi, K.; Abraham, N.G.; Sacerdoti, D. Characterization of a murine model of cardiorenal syndrome type 1 by high-resolution Doppler sonography. *J. Ultrasound.* **2015**, *18*, 229–235. [[CrossRef](#)] [[PubMed](#)]
135. Duni, A.; Liakopoulos, V.; Roumeliotis, S.; Peschos, D.; Dounousi, E. Oxidative Stress in the Pathogenesis and Evolution of Chronic Kidney Disease: Untangling Ariadne’s Thread. *Int. J. Mol. Sci.* **2019**, *20*, 3711. [[CrossRef](#)]
136. Fan, P.C.; Chang, C.H.; Chen, Y.C. Biomarkers for acute cardiorenal syndrome. *Nephrology* **2018**, *23* (Suppl. 4), 68–71. [[CrossRef](#)] [[PubMed](#)]
137. Virzi, G.M.; Breglia, A.; Brocca, A.; de Cal, M.; Bolin, C.; Vescovo, G.; Ronco, C. Levels of Proinflammatory Cytokines, Oxidative Stress, and Tissue Damage Markers in Patients with Acute Heart Failure with and without Cardiorenal Syndrome Type 1. *Cardiorenal. Med.* **2018**, *8*, 321–331. [[CrossRef](#)]
138. Chaudhary, K.; Malhotra, K.; Sowers, J.; Aroor, A. Uric Acid—Key ingredient in the recipe for cardiorenal metabolic syndrome. *Cardiorenal. Med.* **2013**, *3*, 208–220. [[CrossRef](#)]
139. Sodhi, K.; Hilgefert, J.; Banks, G.; Gilliam, C.; Stevens, S.; Ansinelli, H.A.; Getty, M.; Abraham, N.G.; Shapiro, J.I.; Khitan, Z. Uric Acid-Induced Adipocyte Dysfunction Is Attenuated by HO-1 Upregulation: Potential Role of Antioxidant Therapy to Target Obesity. *Stem Cells Int.* **2016**, *2016*, 8197325. [[CrossRef](#)]
140. Khaw, K.T.; Barrett-Connor, E. Endogenous sex hormones, high density lipoprotein cholesterol, and other lipoprotein fractions in men. *Arterioscler. Thromb.* **1991**, *11*, 489–494. [[CrossRef](#)]
141. Khitan, Z.; Kim, D.H. Fructose: A key factor in the development of metabolic syndrome and hypertension. *J. Nutr. Metab.* **2013**, *2013*, 682673. [[CrossRef](#)]
142. van ‘t Erve, T.J.; Kadiiska, M.B.; London, S.J.; Mason, R.P. Classifying oxidative stress by F2-isoprostanate levels across human diseases: A meta-analysis. *Redox Biol.* **2017**, *12*, 582–599. [[CrossRef](#)]
143. Cao, J.; Tsenevov, P.L.; Thompson, E.A.; Falck, J.R.; Touchon, R.; Sodhi, K.; Rezzani, R.; Shapiro, J.I.; Abraham, N.G. Agonists of epoxyeicosatrienoic acids reduce infarct size and ameliorate cardiac dysfunction via activation of HO-1 and Wnt1 canonical pathway. *Prostaglandins Other Lipid Mediat.* **2015**, *116–117*, 76–86. [[CrossRef](#)]
144. Hinds, T.D., Jr.; Stec, D.E. Bilirubin Safeguards Cardiorenal and Metabolic Diseases: A Protective Role in Health. *Curr. Hypertens. Rep.* **2019**, *21*, 87. [[CrossRef](#)]

145. Bellner, L.; Lebovics, N.B.; Rubinstein, R.; Buchen, Y.D.; Sinatra, E.; Sinatra, G.; Abraham, N.G.; McClung, J.A.; Thompson, E.A. Heme Oxygenase-1 Upregulation: A Novel Approach in the Treatment of Cardiovascular Disease. *Antioxid. Redox Signal.* **2020**, *32*, 1045–1060. [[CrossRef](#)]
146. Aroor, A.R.; Mandavia, C.; Ren, J.; Sowers, J.R.; Pulakat, L. Mitochondria and Oxidative Stress in the Cardiorenal Metabolic Syndrome. *Cardiorenal. Med.* **2012**, *2*, 87–109. [[CrossRef](#)]
147. Sam, F.; Kerstetter, D.L.; Pimental, D.R.; Mulukutla, S.; Tabaei, A.; Bristow, M.R.; Colucci, W.S.; Sawyer, D.B. Increased reactive oxygen species production and functional alterations in antioxidant enzymes in human failing myocardium. *J. Card. Fail.* **2005**, *11*, 473–480. [[CrossRef](#)]
148. Wald, H.; Scherzer, P.; Rubinger, D.; Popovtzer, M.M. Effect of indomethacin in vivo and PGE2 in vitro on MTAL Na-K-ATPase of the rat kidney. *Pflugers Arch.* **1990**, *415*, 648–650. [[CrossRef](#)]
149. Waldman, M.; Bellner, L.; Vanella, L.; Schragenheim, J.; Sodhi, K.; Singh, S.P.; Lin, D.; Lakhkar, A.; Li, J.; Hochhauser, E.; et al. Epoxyeicosatrienoic Acids Regulate Adipocyte Differentiation of Mouse 3T3 Cells, Via PGC-1alpha Activation, Which Is Required for HO-1 Expression and Increased Mitochondrial Function. *Stem Cells Dev.* **2016**, *25*, 1084–1094. [[CrossRef](#)]
150. Waldman, M.; Nudelman, V.; Shainberg, A.; Abraham, N.G.; Kornwoski, R.; Aravot, D.; Arad, M.; Hochhauser, E. PARP-1 inhibition protects the diabetic heart through activation of SIRT1-PGC-1alpha axis. *Exp. Cell Res.* **2018**, *373*, 112–118. [[CrossRef](#)]
151. Waldman, M.; Cohen, K.; Yadin, D.; Nudelman, V.; Gorfil, D.; Laniado-Schwartzman, M.; Kornwoski, R.; Aravot, D.; Abraham, N.G.; Arad, M.; et al. Regulation of diabetic cardiomyopathy by caloric restriction is mediated by intracellular signaling pathways involving ‘SIRT1 and PGC-1alpha’. *Cardiovasc. Diabetol.* **2018**, *17*, 111. [[CrossRef](#)] [[PubMed](#)]
152. Radhakrishnan, N.; Yadav, S.P.; Sachdeva, A.; Pruthi, P.K.; Sawhney, S.; Piplani, T.; Wada, T.; Yachie, A. Human heme oxygenase-1 deficiency presenting with hemolysis, nephritis, and asplenia. *J. Pediatr. Hematol. Oncol.* **2011**, *33*, 74–78. [[CrossRef](#)] [[PubMed](#)]
153. Singh, S.P.; McClung, J.A.; Bellner, L.; Cao, J.; Waldman, M.; Schragenheim, J.; Arad, M.; Hochhauser, E.; Falck, J.R.; Weingarten, J.A.; et al. CYP-450 Epoxygenase Derived Epoxyeicosatrienoic Acid Contribute to Reversal of Heart Failure in Obesity-Induced Diabetic Cardiomyopathy via PGC-1 alpha Activation. *Cardiovasc. Pharmacol. (Open Access)* **2018**, *7*, 233.
154. Singh, S.; McClung, J.; Thompson, E.; Glick, Y.; Greenberg, M.; Acosta-Baez, G.; Edris, B.; Shapiro, J.; Abraham, N.G. Cardioprotective heme oxygenase-1-PGC-1 α signaling in epicardial fat attenuates cardiovascular risk in humans as in obese mice. *Obesity (Silver Spring)* **2019**, *27*, 1634–1643. [[CrossRef](#)] [[PubMed](#)]
155. Singh, S.P.; Greenberg, M.; Glick, Y.; Bellner, L.; Favero, G.; Rezzani, R.; Rodella, L.F.; Agostinucci, K.; Shapiro, J.I.; Abraham, N.G. Adipocyte Specific HO-1 Gene Therapy is Effective in Antioxidant Treatment of Insulin Resistance and Vascular Function in an Obese Mice Model. *Antioxidants* **2020**, *9*, 40. [[CrossRef](#)] [[PubMed](#)]
156. Raffaele, M.; Licari, M.; Amin, S.; Alex, R.; Shen, H.H.; Singh, S.P.; Vanella, L.; Rezzani, R.; Bonomini, F.; Peterson, S.J.; et al. Cold Press Pomegranate Seed Oil Attenuates Dietary-Obesity Induced Hepatic Steatosis and Fibrosis through Antioxidant and Mitochondrial Pathways in Obese Mice. *Int. J. Mol. Sci.* **2020**, *21*, 5469. [[CrossRef](#)]
157. Shen, H.H.; Peterson, S.J.; Bellner, L.; Choudhary, A.; Levy, L.; Gancz, L.; Sasson, A.; Trainer, J.; Rezzani, R.; Resnick, A.; et al. Cold-Pressed Nigella Sativa Oil Standardized to 3% Thymoquinone Potentiates Omega-3 Protection against Obesity-Induced Oxidative Stress, Inflammation, and Markers of Insulin Resistance Accompanied with Conversion of White to Beige Fat in Mice. *Antioxidants* **2020**, *9*, 489. [[CrossRef](#)]
158. Peterson, S.J.; Rubinstein, R.; Faroqui, M.; Raza, A.; Boumaza, I.; Zhang, Y.; Stec, D.; Abraham, N.G. Positive Effects of Heme Oxygenase Upregulation on Adiposity and Vascular Dysfunction: Gene Targeting vs. Pharmacologic Therapy. *Int. J. Mol. Sci.* **2019**, *20*, 2514. [[CrossRef](#)]
159. Peterson, S.J.; Shapiro, J.I.; Thompson, E.; Singh, S.; Liu, L.; Weingarten, J.A.; O’Hanlon, K.; Bialczak, A.; Bhesaria, S.R.; Abraham, N.G. Oxidized HDL, Adipokines, and Endothelial Dysfunction: A Potential Biomarker Profile for Cardiovascular Risk in Women with Obesity. *Obesity (Silver Spring)* **2019**, *27*, 87–93. [[CrossRef](#)]

160. Liu, L.; Puri, N.; Raffaele, M.; Schragenheim, J.; Singh, S.P.; Bradbury, J.A.; Bellner, L.; Vanella, L.; Zeldin, D.C.; Cao, J.; et al. Ablation of soluble epoxide hydrolase reprogram white fat to beige-like fat through an increase in mitochondrial integrity, HO-1-adiponectin in vitro and in vivo. *Prostaglandins Other Lipid Mediat.* **2018**, *138*, 1–8. [[CrossRef](#)]
161. Salas-Salvado, J.; Diaz-Lopez, A.; Ruiz-Canela, M.; Basora, J.; Fito, M.; Corella, D.; Serra-Majem, L.; Warnberg, J.; Romaguera, D.; Estruch, R.; et al. Effect of a Lifestyle Intervention Program with Energy-Restricted Mediterranean Diet and Exercise on Weight Loss and Cardiovascular Risk Factors: One-Year Results of the PREDIMED-Plus Trial. *Diabetes Care* **2019**, *42*, 777–788. [[CrossRef](#)] [[PubMed](#)]
162. Ruiz-Ramie, J.J.; Barber, J.L.; Sarzynski, M.A. Effects of exercise on HDL functionality. *Curr. Opin. Lipidol.* **2019**, *30*, 16–23. [[CrossRef](#)] [[PubMed](#)]
163. Cassiano, A.D.N.; Silva, T.S.D.; Nascimento, C.Q.D.; Wanderley, E.M.; Prado, E.S.; Santos, T.M.M.; Mello, C.S.; Barros-Neto, J.A. [Effects of physical exercise on cardiovascular risk and quality of life in hypertensive elderly people]. *Cien. Saude Colet.* **2020**, *25*, 2203–2212. [[CrossRef](#)] [[PubMed](#)]
164. Pirro, M.; Mannarino, M.R.; Bianconi, V.; Simental-Mendia, L.E.; Bagaglia, F.; Mannarino, E.; Sahebkar, A. The effects of a nutraceutical combination on plasma lipids and glucose: A systematic review and meta-analysis of randomized controlled trials. *Pharmacol. Res.* **2016**, *110*, 76–88. [[CrossRef](#)] [[PubMed](#)]
165. Marso, S.P.; Daniels, G.H.; Brown-Frandsen, K.; Kristensen, P.; Mann, J.F.; Nauck, M.A.; Nissen, S.E.; Pocock, S.; Poulter, N.R.; Ravn, L.S.; et al. Liraglutide and Cardiovascular Outcomes in Type 2 Diabetes. *N. Engl. J. Med.* **2016**, *375*, 311–322. [[CrossRef](#)]
166. Mann, J.F.E.; Orsted, D.D.; Brown-Frandsen, K.; Marso, S.P.; Poulter, N.R.; Rasmussen, S.; Torne, K.; Zinman, B.; Buse, J.B.; Committee, L.S. Investigators, Liraglutide and Renal Outcomes in Type 2 Diabetes. *N. Engl. J. Med.* **2017**, *377*, 839–848. [[CrossRef](#)]
167. Apovian, C.M.; Aronne, L.; Rubino, D.; Still, C.; Wyatt, H.; Burns, C.; Kim, D.; Dunayevich, E.; Group, C.-I.S. A randomized, phase 3 trial of naltrexone SR/bupropion SR on weight and obesity-related risk factors (COR-II). *Obesity (Silver Spring)* **2013**, *21*, 935–943. [[CrossRef](#)]
168. Apovian, C.M.; Aronne, L.J.; Bessesen, D.H.; McDonnell, M.E.; Murad, M.H.; Pagotto, U.; Ryan, D.H.; Still, C.D.; Endocrine, S. Pharmacological management of obesity: An endocrine Society clinical practice guideline. *J. Clin. Endocrinol. Metab.* **2015**, *100*, 342–362. [[CrossRef](#)]

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