



Article Genetic Factors Contributing to the Pathogenesis of Essential Hypertension in Two African Populations

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Abstract: The African continent has the highest prevalence of hypertension globally, with South Africa reporting the highest prevalence in Southern Africa. While the influence of genetic variability in the pathogenesis of hypertension is well described internationally, limited reports are available for African populations. This study aimed to assess the association of genetic variants and essential hypertension in a cohort of two ethnic South African population groups. Two hundred and seventy-seven hypertensive and one hundred and seventy-six normotensive individuals were genotyped for 78 variants. Genotyping was performed using the Illumina GoldenGate Assay and allele-specific polymerase chain reaction. The association of variants was assessed using the Fisher Exact test under the additive and allelic genetic models, while multivariate logistic regression was used to predict the development of hypertension. Five variants (*CYP11B2* rs179998, *AGT* rs5051 and rs699, *AGTR1* rs5186, and *ACE* rs4646994) were significantly associated with essential hypertension in the cohort under study. Furthermore, *AGTR1* rs5186 and *AGT* rs699 were identified as risk factors for the development of hypertension in both ethnic groups. In two ethnic South African populations, an association was observed between renin–angiotensin–aldosterone system (RAAS)-related genes and the development of hypertension.

Keywords: essential hypertension; South Africa; mixed ancestry; Xhosa; renin–angiotensin–aldosterone system; *AGTR1*; *AGT*; *ACE*; *CYP11B2*

1. Introduction

Essential hypertension (EH), defined as a systolic blood pressure (BP) of \geq 140 mm Hg and/or diastolic BP of \geq 90 mm Hg, is a major modifiable risk factor for cardiovascular disease and premature death worldwide [1–3]. Globally, the highest prevalence of EH is reported in the African region, with South Africa reporting the highest prevalence in Southern Africa [3,4]. As of 2016, the prevalence of EH in South Africa was 48.2%, an increase from 38.4%, as reported in 2012 [5,6]. A comparative analysis of two recent South African surveys (South African National Health and Nutrition Examination Survey (SANHANES) in 2012 and the 2016 South African Demographic and Health Survey (DHS)) reported a higher prevalence of EH in males, those of mixed ancestry, and those residing in urban areas [3].

The increasing prevalence of EH in South Africa has resulted in a rise in the number of patients being treated for EH. However, the proportion of patients achieving BP control has remained suboptimal, with an estimated 22.1% of treated patients achieving BP control in 2017 [7].

Several lifestyle factors have been attributed to the increasing prevalence and inadequate control of hypertension in African populations [3,4,7,8]. While these lifestyle factors may explain a portion of patients with inadequate control of BP, racial disparities in the



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Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). clinical presentation and control of EH in African populations have been described [9–12]. A recent study concluded that African ancestry patients were more likely to develop EH at an earlier age and have a higher prevalence of EH, but less likely to have their BP controlled when compared to their Caucasian counterparts [10,12,13].

Severe and resistant hypertension has also been observed at greater levels in patients of African descent [10,11]. Often, these patients have biological differences due to a genetic predisposition to salt and water retention, supressed plasma renin activity, and differential response to anti-hypertensive drugs [11]. This genetic predisposition is hypothesised to be the result of historical environmental pressures and has been referred to as the Sodium Retention Hypothesis [14]. The Sodium Retention Hypothesis suggests that, evolutionarily, the capacity to retain salt provided a biological advantage and increased the fitness of salt retainers in tropical hunter-gatherer societies. However, with urbanisation and dietary changes in the modern era, the environmental pressure to retain salt was removed. As a result, the genetic predisposition to salt retention became disadvantageous and subsequently led to a rise in BP [11,14,15].

Despite the clinical differences observed in hypertensive patients of African descent, a limited number of studies have investigated genetic factors contributing to the pathogenesis of EH in these populations [16–26]. Genetic variants showing an association with EH in African populations are reported in Table 1.

Gene	Single Nucleotide Polymorphism (SNP)	Association	Reference
ACE	rs1799752 (also referred to as rs4646994)	DD genotype is involved in susceptibility to hypertension in Burkinabe and Ethiopian populations. D allele is associated with EH in Sub-Saharan African and Ethiopian populations.	[19,21,27,28]
AGTR1	rs5186	A allele is associated with EH in an Egyptian population.	[29]
ATP2B1	rs17249754	GG genotype had a higher risk of developing hypertension than AA+AG in Burkinabe.	[20]
CYP11B2	rs179998 (-344C/T)	T allele is associated with EH in Egyptian patients.	[30]
GSTM1 and GSTT1 (null)		<i>GSTM1</i> -null and <i>GSTT1</i> -null genotypes are potential factors to predict the development of EH in Egyptian patients.	[31]
GSTT1	(null)	<i>GSTT1</i> -null genotype is associated with EH in Burkinabe.	[32]
MTHFR	rs1801133 (C677T)	TT genotype is associated with the risk of hypertension in a Moroccan population. T allele associated with a predisposition to hypertension in a South-West Cameroonian population.	[22,23]
NOS3	rs2070744 -786T/C	CC genotype was associated with EH in a Sudanese population. C allele is associated with an increased risk of hypertension in an Algerian population, a Tunisian population, and a Sudanese population.	[24,25,33]
NOS3	rs1799983 G894T	TT genotype is associated with EH in a Moroccan population.	[34]

Table 1. Genetic variants associated with EH in African populations.

To date, no genetic association with EH has been described in South African cohorts [18, 35,36], though genetic associations in distinct South African ethnic groups have been described with blood pressure traits [37,38] and uncontrolled hypertension [16,39,40].

South Africa, with its rich ethnic diversity [41–43], presents a unique opportunity to unearth ethnic-specific associations, shedding light on the complex interplay between genetics and EH. With a focus on two South African ethnic groups, this study aims to investigate the association between well-described genetic variants, and the development of EH in Cape Town, South Africa.

2. Materials and Methods

2.1. Ethical Approval and Study Cohort

The University of Cape Town (UCT) Human Research Ethics Committee (HREC) approved this study (UCT HREC 328/2010). All individuals participating in the study provided informed consent. Individuals were recruited from Groote Schuur Hospital (GSH), an academic facility affiliated with the UCT in Cape Town, South Africa. Additional clinical information including the individuals' age, sex, ethnicity, comorbidities (diabetes, a history of transient ischaemic attack, or cerebrovascular accident), and lifestyle factors (smoking status and alcohol consumption) was obtained.

Registered nurses drew two 5 mL Ethylenediaminetetraacetic acid (EDTA) tubes of blood by means of venesection from all individuals recruited into the study. Three consecutive BP measurements using a Dinamap (Soma Tech International, Bloomfield, NJ, USA) were taken and an average of the three readings were used as the final measurement of BP. Individuals were classified as hypertensive either based on the BP reading (the average systolic blood pressure reading was \geq 140 mmHg and/or the average diastolic blood pressure was \geq 90 mmHg), or because the individual recruited was a known hypertensive patient on treatment at the Hypertension Clinic at GSH. Normotensive individuals were classified as such when BP < 140/90 mmHg and the individual was not on any antihypertensive treatment at the time of study. As an exclusion criterion, no related patients were included in the cohort under study.

2.2. Identification of Genetic Variants under Study

Ninety-three (93) genetic single nucleotide polymorphisms (SNPs) postulated to influence the development of EH were identified via the literature and an analysis of the BP regulatory pathways. The complete list of variants identified may be found in the supplementary material (Supplementary Table S1).

2.3. DNA Isolation and Genotyping

A salting-out DNA isolation method [44] was used to extract genomic DNA from whole blood. The extracted DNA was quantified using the NanoDrop[®] ND-1000 (Thermo Scientific, Wilmington, NC, USA). Two genotyping methods were utilized: the Illumina GoldenGate Assay (Illumina Inc, San Diego, CA, USA) and an allele-specific polymerase chain reaction.

2.3.1. Illumina GoldenGate Assay

A total of 92 SNPs were genotyped using the Illumina GoldenGate Assay, a medium throughput genotyping method, as per the manufacturer's protocol [45–47]. The analysis of the run was performed using the Illumina GenomeStudio Genotyping Module v2.0 software (Illumina Inc., San Diego, CA, USA), which uses a clustering algorithm for automated genotype clustering and calling [47,48]. Prior to genotype analysis, quality metrics were assessed for each SNP and each sample processed. Samples with low GenCall scores, indicative of a sample with poor performance on the assay, were excluded from the analysis. Furthermore, SNPS with low Cluster Separation Scores, low intensity for genotypes to be reliably called, and overlapping clusters were excluded from the analysis.

2.3.2. Allele-Specific PCR

Intron 16 of the Angiotensin converting enzyme (*ACE*) gene harbours a 287 base pair polymorphism which could not be resolved using the Illumina Golden Gate Assay. To

identify the presence or absence of this *ACE* sequence in this study, two allele-specific PCRs were sequentially performed, using a previously published protocol [49–51]. Further details on the protocol utilized are available in Supplementary Material S2.

2.4. Statistical Analysis

All statistical analysis was performed using R version 4.2.2 [52]. The Fisher exact method was used to test the association between the genetic variants under study and the hypertensive phenotype. For the association study, *p*-values of less than 0.00054 (post Bonferroni correction with 93 variants under study (p = 0.05/93)) were considered statistically significant.

Two genetic models were assessed; an additive genetic model (which assumes that there is an increased risk in disease per genotype) and an allelic genetic model (which assumes that one allele has a greater effect than the alternate allele) [53].

Multiple Logistic Regression

The generalized linear model (glm) function in R was used to perform multiple logistic regression to test the prediction of EH using the additive genetic model. Initially, all potential risk factors (clinical and lifestyle data, as described in Section 2.1) were included in the analysis. Backward elimination was used to remove each least significant predictor from the model. To assess model fit, both the predictive power of the model and the goodness of fit were used as indicators. The McFadden (pseudo) R2 was used to measure the predictive power of the model, while the goodness-of-fit indictors included the c-statistic and the Hosmer–Lemeshow statistic.

3. Results

3.1. Study Population

Four hundred and fifty-seven participants recruited into the study were genotyped. Four samples failed to meet the data quality metrics of the Illumina GoldenGate Assay and were thus excluded from the study. The resulting cohort under study included a total of 453 participants and classified as follows: based on the phenotypic criteria described in Section 2.1, 277 participants classified as hypertensive while 176 were classified as normotensive. The cohort could be further sub-classified based on self-reported ethnicity (Table 2). The sex distribution of the cohort is available in Supplementary Table S3a. The clinical and demographic data of the cohort are available in Supplementary Table S3b.

Table 2. Stratification of the study population.

Study Population	Mixed Ancestry	Xhosa	Total
Hypertensive Individuals	197	80	277
Normotensive Individuals	116	60	176
Total	313	140	453

3.2. Genotyping Results

Seventy-nine variants were successfully genotyped. Nine variants failed to meet the recommended data quality metrics and therefore could not be confidently used in the analysis. An additional five variants could not be successfully genotyped for all patients in the cohort and were also excluded from the analysis.

Patients known with the *SCNN1B* R563Q (rs149868979 (ENaC)) variant were not included in this study. The cohort was assessed for this variant and all patients under study were genotyped as homozygous wild type. This mutation, previously identified by Rayner in a South African cohort [54], is associated with low-renin-low-aldosterone hypertension and pre-eclampsia in black African and mixed-ancestry individuals. Accordingly, 78 variants were used for statistical analysis.

3.3. Statistical Analysis: Association Study

The Fisher exact method was used to test associations between the 78 variants and EH under the additive genetic model. The results of these associations can be found in Tables 3 and 4. With no stratification of the cohort, five variants (*CYP11B2* (rs1799998), *AGT* (rs5051), *AGTR1* (rs5186), *AGT* (rs699), and *ACE* (rs4646994)) were significant post Bonferroni correction (Table 3).

Gene	Reference SNP Identification Number	Genotype	Hypertensive (N = 277)	Hypertensive %	Normotensive (N = 176)	Normotensive %	<i>p-</i> Value	
		CC	145	52%	26	15%		
CYP11B2	rs1799998	СТ	107	39%	60	34%	$<2.2 \times 10^{-16}$	
	-344C>1	TT	25	9%	90	51%		
		GG	24	9%	86	49%		
AGT	rs5051 -30-3273G>T	GT	134	48%	56	32%	0.0001635	
		TT	119	43%	34	19%		
		AA	78	28%	124	70%		
AGTR1	rs5186 A1166C	AC	56	20%	44	25%	$<2.2 \times 10^{-16}$	
	-	CC	143	52%	8	5%		
		TT	9	3%	96	55%		
AGT	rs699	TC	75	27%	56	32%	$3.841 imes 10^{-5}$	
	1770C	CC	193	70%	24	14%		
		II	50	18%	38	22%		
ACE	rs4646994	ID	121	44%	120	68%	$4.323 imes 10^{-11}$	
	INDEL -	DD	106	38%	18	10%		

Table 3. Variants significantly associated with EH in the study population.

Fifty-two percent of the hypertensive cohort under study harboured the CC genotype in the *CYP11B2* rs1799998 variant, while fifty one percent of the normotensive cohort harboured the TT variant. The *CYP11B2* rs1799998 CC genotype was significantly associated with EH ($p < 2.2 \times 10^{-16}$) in this study (Table 3). This association remained when stratified by ethnicity (Table 4: Mixed Ancestry $p = 1.045 \times 10^{-14}$; Xhosa $p = 1.042 \times 10^{-13}$) and by sex (Supplementary Table S4a: Female $p = 1.19 \times 10^{-15}$; Supplementary Table S4b: Male $p = 4.40 \times 10^{-9}$).

Two variants within the *AGT* gene, rs5051 and rs699, were significantly associated with EH in the cohort (Table 3: p = 0.0001635 and $p = 3.841 \times 10^{-5}$, respectively). Both variants are known to be in linkage; however, the linkage disequilibrium blocks are reported to differ between Caucasian and Black populations [31]. Only the *AGT* rs699 variant demonstrated an association with EH in both the Mixed Ancestry (Table 4: $p = 4.556 \times 10^{-5}$) and Xhosa (Table 4: $p = 4.775 \times 10^{-6}$) ethnic populations. When the cohort was stratified by sex, the *AGT* rs699 variant was only associated with EH in females (Supplementary Table S4a: $p = 5.79 \times 10^{-6}$).

The *AGTR1* rs5186 variant and the *ACE* insertion/deletion polymorphism were significantly associated with EH in the unstratified cohort (Table 3: *AGTR1* rs5186 $p < 2.2 \times 10^{-16}$; *ACE* rs4646994 $p = 4.323 \times 10^{-11}$). The association was upheld when the cohort was stratified by ethnicity (Table 4: *AGTR1* rs5186 $p < 2.2 \times 10^{-16}$ in Mixed Ancestry and $p = 1.831 \times 10^{-11}$ in Xhosa; *ACE* rs4646994 Mixed Ancestry $p = 9.446 \times 10^{-17}$; Xhosa $p = 1.395 \times 10^{-5}$) and sex (Supplementary Table S4a for females: *AGTR1* rs5186 $p < 2.2 \times 10^{-16}$ *ACE* rs4646994 $p = 1.00 \times 10^{-16}$; Supplementary Table S4b for males: *AGTR1* rs5186 $p < 2.2 \times 10^{-16}$ and *ACE* rs4646994 $p = 1.39 \times 10^{-15}$, respectively).

					Mixed Ancestry					Xhosa		
Gene	SNP ID	Genotype	Hypertensive (N = 189)	Hypertensive (%)	Normotensive (N = 116)	Normotensive (%)	<i>p</i> -Value	Hypertensive (N = 88)	Hypertensive (%)	Normotensive (N = 60)	Normotensive (%)	<i>p</i> -Value
CYP11B2	rs1799998 -344C>T	CC CT TT	89 79 21	47% 42% 11%	16 44 56	14% 38% 48%	1.045×10^{-14}	56 28 4	64% 32% 5%	10 16 34	17% 27% 57%	1.042×10^{-13}
AGTR1	rs5186 A1166C	AA AC CC	42 50 97	22% 26% 51%	82 28 6	71% 24% 5%	$<2.2 \times 10^{-16}$	36 6 46	41% 7% 52%	42 16 2	70% 27% 3%	1.831×10^{-11}
AGT	rs699 T776C	TT TC CC	8 71 110	4% 38% 58%	24 36 56	21% 31% 48%	4.556×10^{-5}	1 4 83	1% 5% 94%	0 20 40	0% 33% 67%	4.775×10^{-6}
ACE	rs4646994 INDEL	II ID DD	37 90 62	20% 48% 33%	22 84 10	19% 72% 9%	$9.446 imes 10^{-7}$	13 31 44	15% 35% 50%	16 36 8	27% 60% 13%	1.395×10^{-5}

Table 4. Variants significantly associated with EH in the Mixed Ancestry and Xhosa populations under study.

3.3.1. Allelic Genetic Model

The Fisher exact method was used to test associations of the major and minor alleles of each variant under study and EH. As was observed in the additive genetic model (Table 3), four variants were significant post Bonferroni correction: *CYP11B2* (rs1799998), *AGTR1* (rs5186), *AGT* (rs699), and *ACE* (rs4646994) (Table 5).

In this study, the C allele (rs1799998) of the *CYP11B2* gene was more prevalent in hypertensives than their normotensive counterparts, and conferred a 5.40 increased risk for the development of EH when compared to the T allele (Table 5: $p < 2.2 \times 10^{-16}$; 95% CI 4.010–7.324; OR 5.40). This effect was also significant when the cohort was stratified by ethnicity (Table 6: Mixed Ancestry $p < 2.2 \times 10^{-16}$; 95% CI 3.030–6.280; OR 4.35; and Table 6: Xhosa $p < 2.2 \times 10^{-16}$; 95% CI 5.140–16.071; OR 8.99) and sex (Supplementary Table S5a: Females $p = 2.49 \times 10^{-6}$; 95% CI 1.620–3.412; OR 2.35; Supplementary Table S5b Males $p = 5.67 \times 10^{-16}$; 95% CI 4.104–11.80; OR 6.89).

The *AGTR1* rs5186 A allele was associated with a decreased odds ratio for EH (Table 6: $p < 2.2 \times 10^{-16}$; 95% CI 0.090–0.178; OR 0.13) in the study cohort. This association remained when the cohort was stratified by ethnicity (Table 6: Mixed Ancestry $p < 2.2 \times 10^{-16}$; 95% CI 0.0747–0.173; OR 0.114; and Table 6: Xhosa $p = 6.41 \times 10^{-12}$; 95% CI 0.0859–0.288; OR 0.16).

The insertion allele in the *ACE* gene (rs4646994) was also found to confer a decreased risk for the development of EH (Table 5: $p = 4.4 \times 10^{-6}$; 95% CI 0.399–0.698; OR 0.529). On cohort stratification, this association was only significant in the Xhosa cohort (Table 6: $p = 4.306 \times 10^{-5}$; 95% CI 0.220–0.608; OR 0.367) and males (Supplementary Table S5b: p = 0.0001247; 95% CI 0.266–0.6683; OR 0.418).

3.3.2. Multiple Logistic Regression

The model of best fit for the prediction of EH included four SNPs (*CYP11B2* (rs1799998), *AGT* (rs5051 and rs699), *AGTR1* (rs5186), and *ACE* rs4646994) (Table 7).

As per the fitted model, the *CYP11B2* r1799998 CT and rs1799998 TT genotype and *ACE* rs4646994 ID genotype resulted in a decreased risk of developing hypertension, with odds ratios of 0.2017, 0.0538, and 0.4329, respectively. These decreased risks were also observed in the allelic model (*CYP11B2* rs1799998 T allele and *ACE* rs4646994 I allele) (Table 5).

Conversely, the *AGT* rs5051 GT, *AGTR1* rs5186 AC and CC, and *AGT* rs699 CC genotypes resulted in an increased risk of developing hypertension, with odds ratios of 2.7688, 2.9494, 63.3178, and 10.6507, respectively. It is important, however, to caution the effects of the *AGTR1* rs5186 CC genotype and the *AGT* rs699 CC genotype due to large confidence intervals (AGTR1 rs5186 CC 95%CI: 23.7907–244.2167; and AGT rs699 95 CI 1.9382–72.7814). The model showed good discrimination (c-statistic: 0.91) and good fit (Hoslem–Lemeshow statistic: *p*-value = 0).

Gene	SNP ID	Allele	Hypertensive (N = 554)	Hypertensive (%)	Normotensive (N = 352)	Normotensive (%)	p-Value	95% CI	OR
CYP11B2	rs1799998 -344C>T	C T	397 157	72% 28%	112 240	32% 68%	$<2.2 \times 10^{-16}$	4.010-7.324	5.40
AGTR1	rs5186 A1166C	A C	212 342	38% 62%	292 60	83% 17%	$<2.2 \times 10^{-16}$	0.090-0.178	0.13
AGT	rs699 T776C	T C	93 461	17% 83%	104 248	30% 70%	$7.6 imes 10^{-6}$	0.345–0.670	0.48
ACE	rs4646994 INDEL	I D	221 333	40% 60%	196 156	56% 44%	$4.4 imes 10^{-6}$	0.399–0.698	0.529

Table 5. Alleles significantly associated with EH in the study population.

Table 6. a. Alleles significantly associated with EH in the Mixed Ancestry population. b. Alleles significantly associated with EH in the Xhosa population.

а											
				Mixed Ancestry							
Gene	SNP ID	Allele	Hypertensive (N = 378)	Hypertensive (%)	Normotensive (N =232)	Normotensive (%)	<i>p</i> -Value	95% CI	OR		
CYP11B2	rs1799998 -344C>T	C T	257 121	68% 32%	76 156	33% 67%	$<2.2 \times 10^{-16}$	3.030-6.280	4.35		
AGTR1	rs5186 A1166C	A C	134 244	35% 65%	192 40	83% 17%	$<2.2 \times 10^{-16}$	0.0747-0.173	0.114		

b

			Xhosa						
Gene	SNP ID	Allele	Hypertensive (N = 176)	Hypertensive (%)	Normotensive (N = 120)	Normotensive (%)	<i>p</i> -Value	95% CI	OR
CYP11B2	rs1799998 -344C>T	C T	140 36	80% 20%	36 84	30.00% 70.00%	$<2.2 \times 10^{-16}$	5.140-16.071	8.99
AGTR1	rs5186 A1166C	A C	78 98	44% 56%	100 20	83% 17%	6.41×10^{-12}	0.0859-0.288	0.16
ACE	rs4646994 INDEL	I D	57 119	32% 68%	68 52	57% 43%	4.306×10^{-5}	0.220 0 0.608	0.367

Gene	Coefficients	Estimate Std.	Error	z Value	Pr(>\z\)		OR	95% CI
	(Intercept)	-0.9927	0.9831	-1.010	0.31265		0.3706	0.04867-2.3612
	Gender Male	0.3958	0.3094	1.279	0.20081		1.4856	0.8133-2.7447
CYP11B2	rs1799998 CT rs1799998 TT	-1.6009 -2.9223	0.3730 0.4229	$-4.292 \\ -6.910$	$\begin{array}{c} 1.77 \times 10^{-5} \\ 4.83 \times 10^{-12} \end{array}$	*** ***	0.2017 0.0538	0.0950-0.4119 0.0226-0.1195
AGT	rs5051 GT rs5051 TT	$1.0184 \\ -1.0739$	0.4276 0.7834	2.382 -1.371	0.01722 0.17044	*	2.7688 0.3417	1.2181–6.5618 0.0741–1.6306
AGTR1	rs5186 AC rs5186 CC	1.0816 4.2242	0.3541 0.5868	3.055 7.198	$\begin{array}{c} 0.00225 \\ 6.11 \times 10^{-13} \end{array}$	** ***	2.9494 68.3178	1.4899–5.9991 23.7907–244.2167
AGT	rs699 TC rs699 CC	0.7752 2.3656	0.8298 0.9166	0.934 2.581	0.35015 0.00985	**	2.1711 10.6507	0.4545–12.1796 1.9382–72.7814
ACE	rs4646994 ID rs4646994 DD	-0.8372 0.4453	$0.3554 \\ 0.4546$	-2.355 0.980	0.01850 0.32723	*	0.4329 1.5610	0.2131–0.8627 0.6440–3.8519

Table 7. Multinomial logistic regression results. * denotes *p*-values less than 0.05, ** denotes *p*-values less than 0.01, *** denotes *p*-values less than 0.001.

4. Discussion

This ground-breaking study delves into the intricate genetic landscape of EH within the diverse ethnic tapestry of South Africa. Recognizing the distinctive genetic makeup of the country's population, this investigation focussed on two specific South African population groups: the Xhosa population group and the Mixed Ancestry population, an admixed population colloquially known as the Coloured population. This study was conducted in the Western Cape province, the third most populated province in South Africa. This region comprises 38.8% of Black South Africans (which includes the Xhosa ethnic group) and 41.2% Mixed Ancestry South Africans [55].

Our investigation identified significant associations between EH and five variants in genes related to the renin–angiotensin–aldosterone system (RAAS): *CYP11B2* rs179998, *AGT* rs5051, *AGT* rs699, *AGTR1* rs5186, and *ACE* rs4646994. These associations were evident under both additive and allelic genetic models, highlighting the robust genetic influence on EH within the South African context. Stratifying the cohort by ethnicity further unveiled nuanced associations. Notably, the *CYP11B2* rs179998, *AGT* rs699, *AGTR1* rs5186, and *ACE* rs4646994 variants demonstrated consistent associations with EH in both the Mixed Ancestry and Xhosa ethnic populations. Multinomial logistic regression pinpointed specific risk factors, emphasizing the intricate interplay between genetic variants and the development of EH in these distinct populations.

The RAAS system is a critical pathway in the regulation of BP and has been the focus of several studies investigating the genetic basis of EH [17,37,56,57]. This system is also the target of currently available hypertensive treatment [58]. The observed associations between RAAS variants and EH in our study support the Sodium Retention Hypothesis [59] and highlight the need for pharmacogenetic research of anti-hypertensive treatment in these population groups.

Delving into individual variants, the *CYP11B2* rs1799998 variant, located in the promoter region of the *CYP11B2* gene, exhibited associations with EH, thus corroborating studies linking this variant to higher plasma aldosterone-to-renin ratios and increased BP [60,61]. As an added layer to the complexity to the genetic landscape of EH, the prevalence of the C allele of the *CYP11B2* rs1799998 variant has demonstrated ethnic variation globally [30,62–64].

The *AGT* gene variants, rs5051 and rs699, showcased associations with EH, correlating to previously described studies. The rs5051 T allele is reported to correlate to higher plasma angiotensinogen levels in African populations, while the rs699 variant has been associated with elevated plasma angiotensin levels, contributing to increased BP [65,66]. The *AGTR1* rs5186 SNP in the angiotensin II receptor type 1 gene has also demonstrated ethnic-specific

associations, aligning with the broader disparities observed in previous studies across Caucasian [67], Chinese [62], and African populations [17,29].

The ACE gene's rs4646994 variant, an insertion/deletion polymorphism, unveiled opposing associations with EH, echoing the complex nature of genetic influences on BP regulation. This study highlighted disparities in associations across diverse ethnic cohorts previously reported [21,27,28,57,63,68–73], reinforcing the need for ethnicity-based considerations in genetic studies. The rs1799983 SNP in the *NOS3* gene presented sex-specific associations, emphasizing the importance of considering sex-specific genetic influences on EH described in a 2021 study on Brazilian women of African descent [74].

This study's robust findings deviate from the existing literature on African populations [16–18], emphasizing the need for context-specific research. The distinct genetic associations uncovered herein could be attributed to the meticulous characterization of hypertensive patients attending a specialized clinic, shedding light on potential genetic predispositions within this subset. Additionally, this study marks the first exploration of genetic variants and hypertension in the Mixed Ancestry population, a significant contribution to the genetic understanding of EH in South Africa.

Acknowledging the study's limitations, including a limited sample size for the Xhosaspeaking population group, we emphasize the necessity of expanding research to encompass a more extensive array of ethnic populations within South Africa. The identification of novel microRNAs in an African hypertensive population [66] underscores the importance of incorporating omics approaches in future studies to unveil previously undiscovered genetic variants contributing to EH.

In conclusion, our study reveals compelling associations between five genetic variants and EH in the Mixed Ancestry and Xhosa ethnic groups of South Africa. These findings underscore the importance of ethnicity in understanding the genetic underpinnings of hypertension. As we navigate the complexities of genetic influences on BP regulation, future research endeavours must adopt large-scale omics approaches in indigenous African populations, fostering a deeper understanding of the intricate genetic architecture governing EH in this unique demographic.

Supplementary Materials: The following supporting information can be downloaded at: https: //www.mdpi.com/article/10.3390/jpm14030323/s1, Supplementary material S1: Genetic variants under study; Supplementary material S2: ACE insertion-deletion methodology; Supplementary material S3: Sex stratification and baseline characteristics of the study cohort; Supplementary material S4: Variants significantly associated with EH when stratified by sex; Supplementary material S5: Alleles significantly associated with EH when stratified by sex.

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References

- Mills, K.T.; Stefanescu, A.; He, J. The Global Epidemiology of Hypertension. *Nat. Rev. Nephrol.* 2020, 16, 223–237. [CrossRef] [PubMed]
- 2. Zhou, B.; Perel, P.; Mensah, G.A.; Ezzati, M. Global Epidemiology, Health Burden and Effective Interventions for Elevated Blood Pressure and Hypertension. *Nat. Rev. Cardiol.* **2021**, *18*, 785–802. [CrossRef] [PubMed]
- 3. Kandala, N.-B.; Nnanatu, C.C.; Dukhi, N.; Sewpaul, R.; Davids, A.; Reddy, S.P. Mapping the Burden of Hypertension in South Africa: A Comparative Analysis of the National 2012 SANHANES and the 2016 Demographic and Health Survey. *Int. J. Environ. Res. Public Health* **2021**, *18*, 5445. [CrossRef]
- Reddy, S.P.; Mbewu, A.D.; Williams, D.R.; Harriman, N.W.; Sewpaul, R.; Morgan, J.W.; Sifunda, S.; Manyaapelo, T.; Mabaso, M. Race, Geographical Location and Other Risk Factors for Hypertension: South African National Health and Nutrition Examination Survey 2011/12. SSM—Popul. Health 2021, 16, 100986. [CrossRef]
- 5. Shisana, O.; Labadarios, D.; Rehle, T.; Simbayi, L.; Zuma, K.; Dhansay, A.; Reddy, P.; Parker, W.; Hoosain, E.; Naidoo, P.; et al. South African National Health and Nutrition Examination Survey (SANHANES-1); HSRC Press: Cape Town, South Africa, 2013.
- 6. National Department of Health (NDOH); Statistics South Africa (Stats SA). *South Africa Demographic and Health Survey* 2016; National Department of Health (NDOH): Pretoria, South Africa, 2019.
- Benade, M.; Mchiza, Z.; Raquib, R.V.; Prasad, S.K.; Yan, L.D.; Brennan, A.T.; Davies, J.; Sudharsanan, N.; Manne-Goehler, J.; Fox, M.P.; et al. Health Systems Performance for Hypertension Control Using a Cascade of Care Approach in South Africa, 2011-2017. PLoS Glob. Public Health 2023, 3, e0002055. [CrossRef]
- 8. Adeloye, D.; Basquill, C. Estimating the Prevalence and Awareness Rates of Hypertension in Africa: A Systematic Analysis. *PLoS ONE* **2014**, *9*, e104300. [CrossRef]
- 9. Abrahamowicz, A.A.; Ebinger, J.; Whelton, S.P.; Commodore-Mensah, Y.; Yang, E. Racial and Ethnic Disparities in Hypertension: Barriers and Opportunities to Improve Blood Pressure Control. *Curr. Cardiol. Rep.* **2023**, *25*, 17–27. [CrossRef]
- Howard, G.; Prineas, R.; Moy, C.; Cushman, M.; Kellum, M.; Temple, E.; Graham, A.; Howard, V. Racial and Geographic Differences in Awareness, Treatment, and Control of Hypertension: The REasons for Geographic And Racial Differences in Stroke Study. *Stroke* 2006, 37, 1171–1178. [CrossRef]
- 11. Spence, J.D.; Rayner, B.L. Hypertension in Blacks: Individualized Therapy Based on Renin/Aldosterone Phenotyping. *Hypertension* 2018, 72, 263–269. [CrossRef]
- Aggarwal, R.; Chiu, N.; Wadhera, R.K.; Moran, A.E.; Raber, I.; Shen, C.; Yeh, R.W.; Kazi, D.S. Racial/Ethnic Disparities in Hypertension Prevalence, Awareness, Treatment, and Control in the United States, 2013 to 2018. *Hypertension* 2021, 78, 1719–1726. [CrossRef]
- 13. Lackland, D.T. Racial Differences in Hypertension: Implications for High Blood Pressure Management. *Am. J. Med. Sci.* 2014, 348, 135–138. [CrossRef]
- 14. Batuman, V. Salt and Hypertension: Why Is There Still a Debate? Kidney Int. Suppl. 2013, 3, 316–320. [CrossRef] [PubMed]
- 15. Malinowska, J.K.; Żuradzki, T. Towards the Multileveled and Processual Conceptualisation of Racialised Individuals in Biomedical Research. *Synthese* **2023**, 201, 11. [CrossRef] [PubMed]
- 16. Mabhida, S.E.; Mashatola, L.; Kaur, M.; Sharma, J.R.; Apalata, T.; Muhamed, B.; Benjeddou, M.; Johnson, R. Hypertension in African Populations: Review and Computational Insights. *Genes* **2021**, *12*, 532. [CrossRef]
- Yako, Y.Y.; Balti, E.V.; Matsha, T.E.; Dzudie, A.; Kruger, D.; Sobngwi, E.; Agyemang, C.; Kengne, A.P. Genetic Factors Contributing to Hypertension in African-Based Populations: A Systematic Review and Meta-Analysis. *J. Clin. Hypertens.* 2018, 20, 485–495. [CrossRef] [PubMed]
- Ranjith, N.; Pegoraro, R.J.; Rom, L.; Lanning, P.A.; Naidoo, D.P. Renin-Angiotensin System and Associated Gene Polymorphisms in Myocardial Infarction in Young South African Indians. *Cardiovasc. J. S. Afr. Off. J. S. Afr. Card. Soc. S. Afr. Soc. Card. Pract.* 2004, 15, 22–26.
- Tchelougou, D.; Kologo, J.K.; Karou, S.D.; Yaméogo, V.N.; Bisseye, C.; Djigma, F.W.; Ouermi, D.; Compaoré, T.R.; Assih, M.; Pietra, V.; et al. Renin-Angiotensin System Genes Polymorphisms and Essential Hypertension in Burkina Faso, West Africa. *Int. J. Hypertens.* 2015, 2015, 979631. [CrossRef]
- Sombié, H.K.; Kologo, J.K.; Tchelougou, D.; Ouédraogo, S.Y.; Ouattara, A.K.; Compaoré, T.R.; Nagalo, B.M.; Sorgho, A.P.; Nagabila, I.; Soubeïga, S.T.; et al. Positive Association between ATP2B1 Rs17249754 and Essential Hypertension: A Case-Control Study in Burkina Faso, West Africa. *BMC Cardiovasc. Disord.* 2019, 19, 155. [CrossRef]
- 21. Abouelfath, R.; Habbal, R.; Laaraj, A.; Khay, K.; Harraka, M.; Nadifi, S. ACE Insertion/Deletion Polymorphism Is Positively Associated with Resistant Hypertension in Morocco. *Gene* **2018**, *658*, 178–183. [CrossRef]
- 22. Nassereddine, S.; Kassogue, Y.; Korchi, F.; Habbal, R.; Nadifi, S. Association of Methylenetetrahydrofolate Reductase Gene (C677T) with the Risk of Hypertension in Morocco. *BMC Res. Notes* **2015**, *8*, 775. [CrossRef]
- 23. Ghogomu, S.M.; Ngolle, N.E.; Mouliom, R.N.; Asa, B.F. Association between the MTHFR C677T Gene Polymorphism and Essential Hypertension in South West Cameroon. *Genet. Mol. Res. GMR* **2016**, *15*. [CrossRef]
- 24. Gamil, S.; Erdmann, J.; Abdalrahman, I.B.; Mohamed, A.O. Association of NOS3 Gene Polymorphisms with Essential Hypertension in Sudanese Patients: A Case Control Study. *BMC Med. Genet.* 2017, *18*, 128. [CrossRef] [PubMed]

- Jemaa, R.; Kallel, A.; Sediri, Y.; Omar, S.; Feki, M.; Elasmi, M.; Haj-Taieb, S.; Sanhaji, H.; Kaabachi, N. Association between -786TC Polymorphism in the Endothelial Nitric Oxide Synthase Gene and Hypertension in the Tunisian Population. *Exp. Mol. Pathol.* 2011, 90, 210–214. [CrossRef]
- Kabadou, I.A.; Soualmia, H.; Jemaa, R.; Feki, M.; Kallel, A.; Souheil, O.; Taieb, S.H.; Sanhaji, H.; Kaabachi, N. G Protein Beta3 Subunit Gene C825T and Angiotensin Converting Enzyme Gene Insertion/Deletion Polymorphisms in Hypertensive Tunisian Population. *Clin. Lab.* 2013, *59*, 85–92. [CrossRef]
- 27. Birhan, T.A.; Molla, M.D.; Abdulkadir, M.; Tesfa, K.H. Association of Angiotensin-Converting Enzyme Gene Insertion/Deletion Polymorphisms with Risk of Hypertension among the Ethiopian Population. *PLoS ONE* **2022**, *17*, e0276021. [CrossRef]
- 28. Mengesha, H.G.; Petrucka, P.; Spence, C.; Tafesse, T.B. Effects of Angiotensin Converting Enzyme Gene Polymorphism on Hypertension in Africa: A Meta-Analysis and Systematic Review. *PLoS ONE* **2019**, *14*, e0211054. [CrossRef]
- 29. Farrag, W.; Eid, M.; El-Shazly, S.; Abdallah, M. Angiotensin II Type 1 Receptor Gene Polymorphism and Telomere Shortening in Essential Hypertension. *Mol. Cell. Biochem.* 2011, 351, 13–18. [CrossRef]
- Abdel Ghafar, M.T. Association of Aldosterone Synthase CYP11B2 (-344C/T) Gene Polymorphism with Essential Hypertension and Left Ventricular Hypertrophy in the Egyptian Population. *Clin. Exp. Hypertens.* 2019, 41, 779–786. [CrossRef]
- 31. Bessa, S.S.; Ali, E.M.M.; Hamdy, S.M. The Role of Glutathione S- Transferase M1 and T1 Gene Polymorphisms and Oxidative Stress-Related Parameters in Egyptian Patients with Essential Hypertension. *Eur. J. Intern. Med.* **2009**, *20*, 625–630. [CrossRef]
- Sombié, H.K.; Sorgho, A.P.; Kologo, J.K.; Ouattara, A.K.; Yaméogo, S.; Yonli, A.T.; Djigma, F.W.; Tchelougou, D.; Somda, D.; Kiendrébéogo, I.T.; et al. Glutathione S-Transferase M1 and T1 Genes Deletion Polymorphisms and Risk of Developing Essential Hypertension: A Case-Control Study in Burkina Faso Population (West Africa). *BMC Med. Genet.* 2020, 21, 55. [CrossRef] [PubMed]
- Amrani-Midoun, A.; Kiando, S.R.; Treard, C.; Jeunemaitre, X.; Bouatia-Naji, N. Genetic Association Study between T-786C NOS3 Polymorphism and Essential Hypertension in an Algerian Population of the Oran City. *Diabetes Metab. Syndr.* 2019, 13, 1317–1320. [CrossRef]
- Nassereddine, S.; Hassani Idrissi, H.; Habbal, R.; Abouelfath, R.; Korch, F.; Haraka, M.; Karkar, A.; Nadifi, S. The Polymorphism G894 T of Endothelial Nitric Oxide Synthase (eNOS) Gene Is Associated with Susceptibility to Essential Hypertension (EH) in Morocco. *BMC Med. Genet.* 2018, 19, 127. [CrossRef]
- 35. Mabhida, S.E.; Sharma, J.R.; Apalata, T.; Masilela, C.; Nomatshila, S.; Mabasa, L.; Fokkens, H.; Benjeddou, M.; Muhamed, B.; Shabalala, S.; et al. The Association of MTHFR (Rs1801133) with Hypertension in an Indigenous South African Population. *Front. Genet.* **2022**, *13*, 937639. [CrossRef]
- Nkeh, B.; Samani, N.J.; Badenhorst, D.; Libhaber, E.; Sareli, P.; Norton, G.R.; Woodiwiss, A.J. T594M Variant of the Epithelial Sodium Channel Beta-Subunit Gene and Hypertension in Individuals of African Ancestry in South Africa. *Am. J. Hypertens.* 2003, 16, 847–852. [CrossRef]
- Tiago, A.D.; Badenhorst, D.; Nkeh, B.; Candy, G.P.; Brooksbank, R.; Sareli, P.; Libhaber, E.; Samani, N.J.; Woodiwiss, A.J.; Norton, G.R. Impact of Renin-Angiotensin-Aldosterone System Gene Variants on the Severity of Hypertension in Patients with Newly Diagnosed Hypertension. *Am. J. Hypertens.* 2003, *16*, 1006–1010. [CrossRef]
- Hendry, L.M.; Sahibdeen, V.; Choudhury, A.; Norris, S.A.; Ramsay, M.; Lombard, Z.; of the AWI-Gen Study and as Members of the H3Africa Consortium. Insights into the Genetics of Blood Pressure in Black South African Individuals: The Birth to Twenty Cohort. BMC Med. Genomics 2018, 11, 2. [CrossRef] [PubMed]
- Masilela, C.; Pearce, B.; Ongole, J.J.; Adeniyi, O.V.; Benjeddou, M. Genomic Association of Single Nucleotide Polymorphisms with Blood Pressure Response to Hydrochlorothiazide among South African Adults with Hypertension. *J. Pers. Med.* 2020, 10, 267. [CrossRef] [PubMed]
- 40. Masilela, C.; Adeniyi, O.V.; Benjeddou, M. Single Nucleotide Polymorphisms in Amlodipine-Associated Genes and Their Correlation with Blood Pressure Control among South African Adults with Hypertension. *Genes* **2022**, *13*, 1394. [CrossRef]
- 41. Choudhury, A.; Sengupta, D.; Ramsay, M.; Schlebusch, C. Bantu-Speaker Migration and Admixture in Southern Africa. *Hum. Mol. Genet.* **2021**, *30*, R56–R63. [CrossRef]
- 42. Choudhury, A.; Aron, S.; Botigué, L.R.; Sengupta, D.; Botha, G.; Bensellak, T.; Wells, G.; Kumuthini, J.; Shriner, D.; Fakim, Y.J.; et al. High-Depth African Genomes Inform Human Migration and Health. *Nature* **2020**, *586*, 741–748. [CrossRef]
- Sengupta, D.; Choudhury, A.; Fortes-Lima, C.; Aron, S.; Whitelaw, G.; Bostoen, K.; Gunnink, H.; Chousou-Polydouri, N.; Delius, P.; Tollman, S.; et al. Genetic Substructure and Complex Demographic History of South African Bantu Speakers. *Nat. Commun.* 2021, 12, 2080. [CrossRef]
- 44. Miller, S.A.; Dykes, D.D.; Polesky, H.F. A Simple Salting out Procedure for Extracting DNA from Human Nucleated Cells. *Nucleic Acids Res.* **1988**, *16*, 1215. [CrossRef]
- 45. Illumina Technical Note: Designing Custom GoldenGate Genotyping Assay. Available online: https://www.illumina.com/ Documents/products/technote_goldengate_design.pdf (accessed on 16 March 2024).
- Illumina Technical Note: GoldenGate Assay Workflow. Available online: https://www.illumina.com/documents/products/ workflows/workflow_goldengate_assay.pdf (accessed on 16 March 2024).
- 47. González-Neira, A. The GoldenGate Genotyping Assay: Custom Design, Processing, and Data Analysis. *Methods Mol. Biol.* 2013, 1015, 147–153. [CrossRef] [PubMed]

- Software Guide: GenomeStudio Genotyping Module v2.0. Available online: https://support.illumina.com/content/dam/ illumina-support/documents/documentation/software_documentation/genomestudio/genomestudio-2-0/genomestudiogenotyping-module-v2-user-guide-11319113-01.pdf (accessed on 16 March 2024).
- 49. Rigat, B.; Hubert, C.; Corvol, P.; Soubrier, F. PCR Detection of the Insertion/Deletion Polymorphism of the Human Angiotensin Converting Enzyme Gene (DCP1) (Dipeptidyl Carboxypeptidase 1). *Nucleic Acids Res.* **1992**, *20*, 1433. [CrossRef]
- 50. Odawara, M.; Matsunuma, A.; Yamashita, K. Mistyping Frequency of the Angiotensin-Converting Enzyme Gene Polymorphism and an Improved Method for Its Avoidance. *Hum. Genet.* **1997**, *100*, 163–166. [CrossRef]
- Lindpaintner, K.; Pfeffer, M.A.; Kreutz, R.; Stampfer, M.J.; Grodstein, F.; LaMotte, F.; Buring, J.; Hennekens, C.H. A Prospective Evaluation of an Angiotensin-Converting-Enzyme Gene Polymorphism and the Risk of Ischemic Heart Disease. *N. Engl. J. Med.* 1995, 332, 706–711. [CrossRef]
- 52. R Core Team. *R: A Language and Environment for Statistical Computing;* R Foundation for Statistical Computing: Vienna, Austria, 2021.
- Horita, N.; Kaneko, T. Genetic Model Selection for a Case-Control Study and a Meta-Analysis. *Meta Gene* 2015, 5, 1–8. [CrossRef] [PubMed]
- Rayner, B.L.; Owen, E.P.; King, J.A.; Soule, S.G.; Vreede, H.; Opie, L.H.; Marais, D.; Davidson, J.S. A New Mutation, R563Q, of the Beta Subunit of the Epithelial Sodium Channel Associated with Low-Renin, Low-Aldosterone Hypertension. *J. Hypertens.* 2003, 21, 921–926. [CrossRef]
- 55. Statistics South Africa (Stats SA). Post-Enumeration Survey (PES) 2022; Stats SA: Pretoria, South Africa, 2022.
- Fountain, J.H.; Kaur, J.; Lappin, S.L. Physiology, Renin Angiotensin System. Available online: https://pubmed.ncbi.nlm.nih.gov/ 29261862/ (accessed on 13 December 2023).
- 57. Mondry, A.; Loh, M.; Liu, P.; Zhu, A.-L.; Nagel, M. Polymorphisms of the Insertion/Deletion ACE and M235T AGT Genes and Hypertension: Surprising New Findings and Meta-Analysis of Data. *BMC Nephrol.* **2005**, *6*, 1. [CrossRef]
- Al-Makki, A.; DiPette, D.; Whelton, P.K.; Murad, M.H.; Mustafa, R.A.; Acharya, S.; Beheiry, H.M.; Champagne, B.; Connell, K.; Cooney, M.T.; et al. Hypertension Pharmacological Treatment in Adults: A World Health Organization Guideline Executive Summary. *Hypertension* 2022, *79*, 293–301. [CrossRef] [PubMed]
- 59. Strazzullo, P.; Galletti, F. Genetics of Salt-Sensitive Hypertension. Curr. Hypertens. Rep. 2007, 9, 25–32. [CrossRef]
- Connell, J.M.C.; Fraser, R.; MacKenzie, S.M.; Friel, E.C.; Ingram, M.C.; Holloway, C.D.; Davies, E. The Impact of Polymorphisms in the Gene Encoding Aldosterone Synthase (CYP11B2) on Steroid Synthesis and Blood Pressure Regulation. *Mol. Cell. Endocrinol.* 2004, 217, 243–247. [CrossRef] [PubMed]
- 61. Takeuchi, F.; Yamamoto, K.; Katsuya, T.; Sugiyama, T.; Nabika, T.; Ohnaka, K.; Yamaguchi, S.; Takayanagi, R.; Ogihara, T.; Kato, N. Reevaluation of the Association of Seven Candidate Genes with Blood Pressure and Hypertension: A Replication Study and Meta-Analysis with a Larger Sample Size. *Hypertens. Res. Off. J. Jpn. Soc. Hypertens.* **2012**, *35*, 825–831. [CrossRef] [PubMed]
- Wang, L.; Zhang, B.; Li, M.; Li, C.; Liu, J.; Liu, Y.; Wang, Z.; Zhou, J.; Wen, S. Association between Single-Nucleotide Polymorphisms in Six Hypertensive Candidate Genes and Hypertension among Northern Han Chinese Individuals. *Hypertens. Res. Off. J. Jpn. Soc. Hypertens.* 2014, *37*, 1068–1074. [CrossRef] [PubMed]
- 63. Gouissem, I.; Midani, F.; Soualmia, H.; Bouchemi, M.; Ouali, S.; Kallele, A.; Romdhane, N.B.; Mourali, M.S.; Feki, M. Contribution of the ACE (Rs1799752) and CYP11B2 (Rs1799998) Gene Polymorphisms to Atrial Fibrillation in the Tunisian Population. *Biol. Res. Nurs.* **2022**, *24*, 31–39. [CrossRef] [PubMed]
- 64. Shah, W.A.; Jan, A.; Khan, M.A.; Saeed, M.; Rahman, N.; Zakiullah; Afridi, M.S.; Khuda, F.; Akbar, R. Association between Aldosterone Synthase (CYP11B2) Gene Polymorphism and Hypertension in Pashtun Ethnic Population of Khyber Pakhtunkwha, Pakistan. *Genes* 2023, *14*, 1184. [CrossRef] [PubMed]
- 65. Nakajima, T.; Wooding, S.; Sakagami, T.; Emi, M.; Tokunaga, K.; Tamiya, G.; Ishigami, T.; Umemura, S.; Munkhbat, B.; Jin, F.; et al. Natural Selection and Population History in the Human Angiotensinogen Gene (AGT): 736 Complete AGT Sequences in Chromosomes from around the World. *Am. J. Hum. Genet.* **2004**, *74*, 898–916. [CrossRef]
- 66. Powell, N.R.; Shugg, T.; Leighty, J.; Martin, M.; Kreutz, R.P.; Eadon, M.T.; Lai, D.; Lu, T.; Skaar, T.C. Analysis of the Combined Effect of Rs699 and Rs5051 on Angiotensinogen Expression and Hypertension. *Chronic Dis. Transl. Med.* **2023**, 1–16. [CrossRef]
- Semianiv, M.M.; Sydorchuk, L.P.; Dzhuryak, V.S.; Gerush, O.V.; Vasylovich Gerush, O.; Palamar, A.O.; Muzyka, N.Y.; Korovenkova, O.M.; Blazhiievska, O.M.; Sydor, V.V.; et al. Association of AGTR1 (Rs5186), VDR (Rs2228570) Genes Polymorphism with Blood Pressure Elevation in Patients with Essential Arterial Hypertension. J. Med. Life 2021, 14, 782–789. [CrossRef]
- Charoen, P.; Eu-Ahsunthornwattana, J.; Thongmung, N.; Jose, P.A.; Sritara, P.; Vathesatogkit, P.; Kitiyakara, C. Contribution of Four Polymorphisms in Renin-Angiotensin-Aldosterone-Related Genes to Hypertension in a Thai Population. *Int. J. Hypertens.* 2019, 2019, 4861081. [CrossRef]
- 69. Gupta, S.; Agrawal, B.K.; Goel, R.K.; Sehajpal, P.K. Angiotensin-Converting Enzyme Gene Polymorphism in Hypertensive Rural Population of Haryana, India. *J. Emerg. Trauma Shock* **2009**, *2*, 150–154. [CrossRef]
- Alsafar, H.; Hassoun, A.; Almazrouei, S.; Kamal, W.; Almaini, M.; Odama, U.; Rais, N. Association of Angiotensin Converting Enzyme Insertion-Deletion Polymorphism with Hypertension in Emiratis with Type 2 Diabetes Mellitus and Its Interaction with Obesity Status. *Dis. Mark.* 2015, 2015, 536041. [CrossRef]
- 71. Kooffreh, M.E.; Anumudu, C.I.; Kumar, P.L. Insertion/Deletion Polymorphism of the Angiotensin-Converting Enzyme Gene and the Risk of Hypertension among Residents of Two Cities, South-South Nigeria. *Adv. Biomed. Res.* **2014**, *3*, 118. [CrossRef]

- 72. Krishnan, R.; Sekar, D.; Karunanithy, S.; Subramanium, S. Association of Angiotensin Converting Enzyme Gene Insertion/Deletion Polymorphism with Essential Hypertension in South Indian Population. *Genes Dis.* **2016**, *3*, 159–163. [CrossRef] [PubMed]
- 73. Morshed, M.; Khan, H.; Akhteruzzaman, S. Association between Angiotensin I-Converting Enzyme Gene Polymorphism and Hypertension in Selected Individuals of the Bangladeshi Population. J. Biochem. Mol. Biol. 2002, 35, 251–254. [CrossRef] [PubMed]
- 74. Neto, A.B.L.; Vasconcelos, N.B.R.; Dos Santos, T.R.; Duarte, L.E.C.; Assunção, M.L.; de Sales-Marques, C.; Ferreira, H.D.S. Prevalence of IGFBP3, NOS3 and TCF7L2 Polymorphisms and Their Association with Hypertension: A Population-Based Study with Brazilian Women of African Descent. *BMC Res. Notes* **2021**, *14*, 186. [CrossRef] [PubMed]

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