

Supplementary Methods

Establishing an Amplicon Targeted Sequencing Approach

To overcome sequencing errors and PCR-mediated recombination that might introduce biases (1-4), we adjusted a library preparation strategy established by others (5). This strategy offers the inclusion of a stretch of degenerate nucleotides included into the cDNA synthesis primer that enables “tagging” of the input cDNA template before proceeding on to PCR amplification steps. This stretch of nucleotides served as an identifier sequence and was previously referred to as “Primer ID”(5).

Since the focus of this study was to identify intra-host viral genetic diversity and we specifically aimed at sequencing single cells carrying a plethora of viral genomes, it was important to be able to identify low frequency genomes and to have as low sequencing bias as possible. The inclusion of the Primer ID sequence allowed each original template copy to be tagged with an identifier sequence. Finding multiple identical Primer ID sequences during the analysis process of the sequencing products indicated that genomes tagged with the same Primer ID originated from a single original input template. These were subsequently collapsed to create a consensus sequence for each original template.

We adapted the Primer ID approach to the Illumina MiSeq platform and amplified a 335 bp long fragment within the WNV genome that included the barcoded region within the NS4b segment (Supplementary Table, table of Illumina barcodes used for this study, excel sheet table-Illumina barcodes used for sample multiplexing (6). Library preparation is further detailed in the Experimental procedures section. Using the Primer ID method we were able to generate libraries from an input of as little as 50 viral genomes.

Supplementary Table S1. Table of Illumina barcodes.

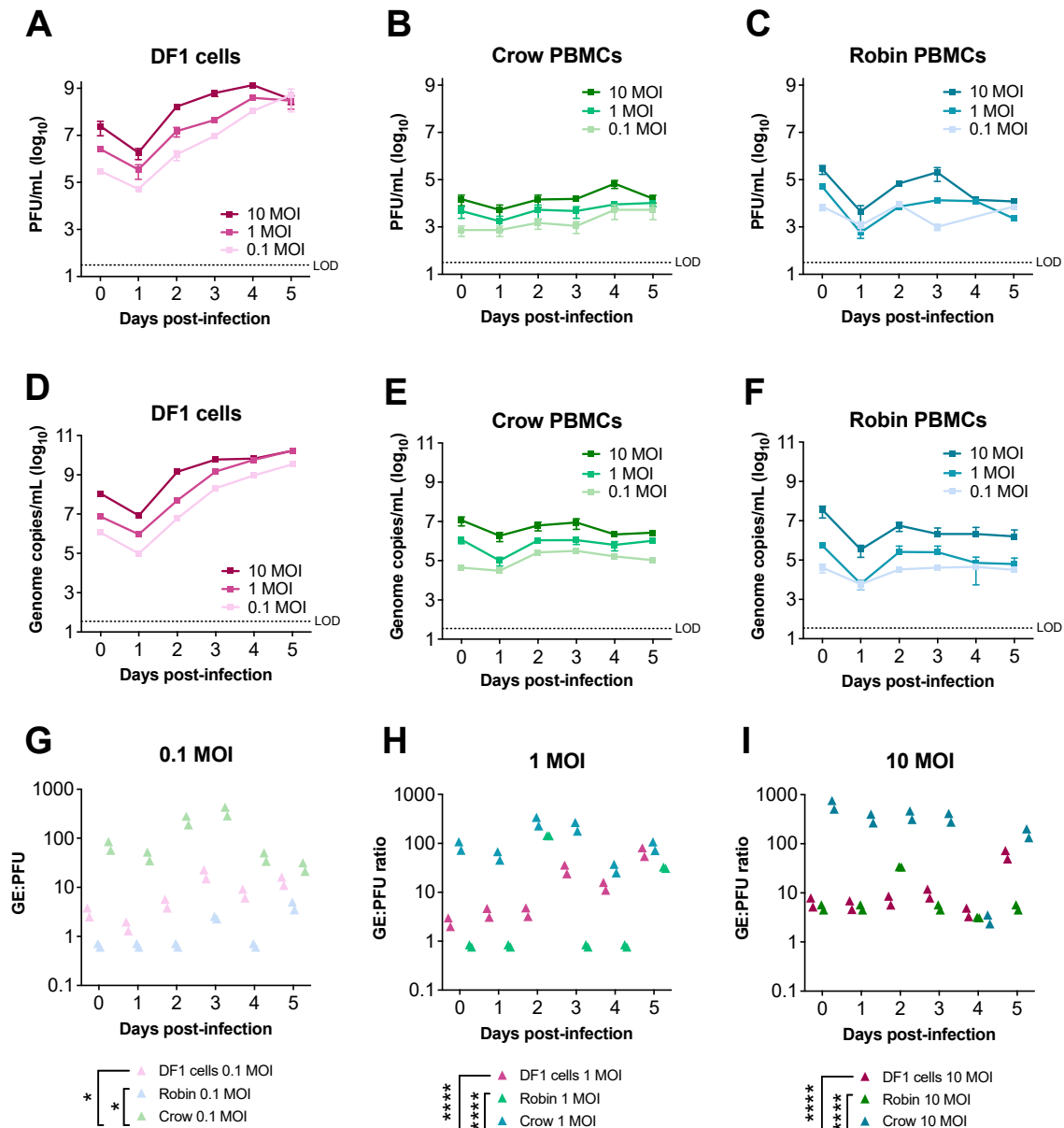
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i5003	gtaaggagagct	AATGATACGGCGACCACCGAGATCTACACgtaaggagagctTCGTCCGCAGCGTC
i5004	actgcataagct	AATGATACGGCGACCACCGAGATCTACAActgcataagctTCGTCCGCAGCGTC
i5005	aaggagtaagct	AATGATACGGCGACCACCGAGATCTACAAaaggagtaagctTCGTCCGCAGCGTC
i5006	ctaagcctagct	AATGATACGGCGACCACCGAGATCTACACctaagcctagctTCGTCCGCAGCGTC
i5007	cgtctaataagct	AATGATACGGCGACCACCGAGATCTACACcgtctaataagctTCGTCCGCAGCGTC
i5008	tctctccgagct	AATGATACGGCGACCACCGAGATCTACAtctctccgagctTCGTCCGCAGCGTC

i5009	tcgactagacgt	AATGATACGGCGACCACCGAGATCTACACtcgactagacgtTCGTCCGCAGCGTC
i5010	ttctagctacgt	AATGATACGGCGACCACCGAGATCTACACttctagctacgtTCGTCCGCAGCGTC
i5011	cctagagtacgt	AATGATACGGCGACCACCGAGATCTACACcctagagtacgtTCGTCCGCAGCGTC
i5012	gcgtaagaacgt	AATGATACGGCGACCACCGAGATCTACACgcgtaagaacgtTCGTCCGCAGCGTC
i5013	ctattaagacgt	AATGATACGGCGACCACCGAGATCTACACctattaagacgtTCGTCCGCAGCGTC
i5014	aaggctatacgt	AATGATACGGCGACCACCGAGATCTACACaaggctatacgtTCGTCCGCAGCGTC
i5015	gagccttaacgt	AATGATACGGCGACCACCGAGATCTACACgagccttaacgtTCGTCCGCAGCGTC
i5016	ttatgcgaacgt	AATGATACGGCGACCACCGAGATCTACACttatgcgaacgtTCGTCCGCAGCGTC
i5017	ctctctattgca	AATGATACGGCGACCACCGAGATCTACACctctctattgcaTCGTCCGCAGCGTC
i5018	tatctcttgcga	AATGATACGGCGACCACCGAGATCTACACtatctcttgcgaTCGTCCGCAGCGTC
i5019	gtaaggagtgcga	AATGATACGGCGACCACCGAGATCTACACgtaaggagtgcgaTCGTCCGCAGCGTC
i5020	actgcataatgca	AATGATACGGCGACCACCGAGATCTACACactgcataatgcaTCGTCCGCAGCGTC
i5021	aaggagtatgca	AATGATACGGCGACCACCGAGATCTACACaaggagtatgcaTCGTCCGCAGCGTC
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i5031	gagccttatgcga	AATGATACGGCGACCACCGAGATCTACACgagccttatgcgaTCGTCCGCAGCGTC
i5032	ttatgcgatgcga	AATGATACGGCGACCACCGAGATCTACACttatgcgatgcgaTCGTCCGCAGCGTC
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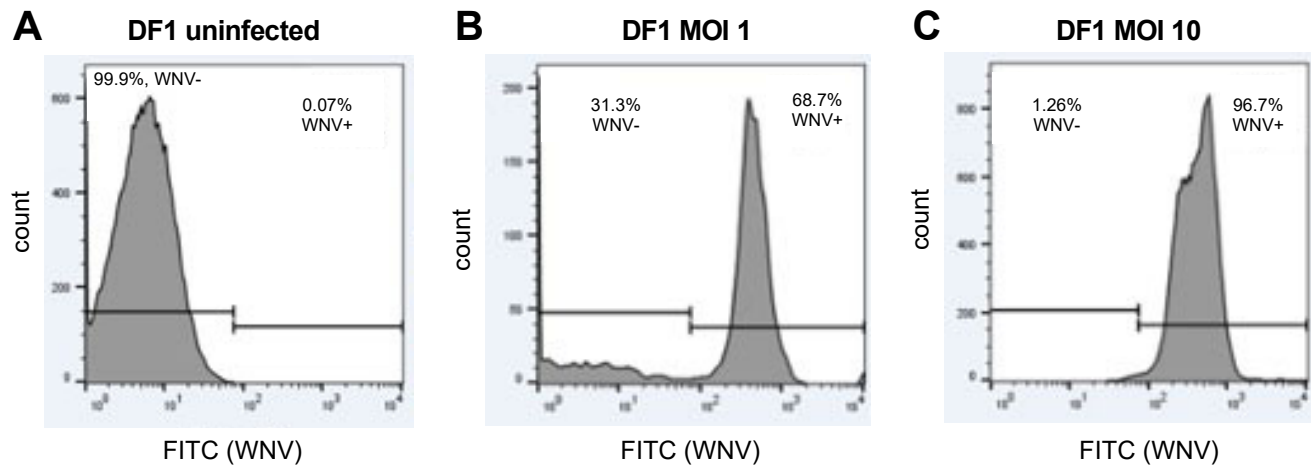
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i701	taaggcga	CAAGCAGAAGACGGCATAACGAGATtaaggcgaGTCTCGTGGGCTCGG
i702	cgtactag	CAAGCAGAAGACGGCATAACGAGATcgtactagGTCTCGTGGGCTCGG
i703	aggcagaa	CAAGCAGAAGACGGCATAACGAGATaggcagaaGTCTCGTGGGCTCGG
i704	tcctgagc	CAAGCAGAAGACGGCATAACGAGATtcctgagcGTCTCGTGGGCTCGG
i705	ggactcct	CAAGCAGAAGACGGCATAACGAGATggactcctGTCTCGTGGGCTCGG
i706	taggcatg	CAAGCAGAAGACGGCATAACGAGATtaggcatgGTCTCGTGGGCTCGG
i707	ctctctac	CAAGCAGAAGACGGCATAACGAGATctctctacGTCTCGTGGGCTCGG
i708	cagagagg	CAAGCAGAAGACGGCATAACGAGATcagagaggGTCTCGTGGGCTCGG
i709	gctacgct	CAAGCAGAAGACGGCATAACGAGATgctacgctGTCTCGTGGGCTCGG
i710	cgaggctg	CAAGCAGAAGACGGCATAACGAGATcgaggctgGTCTCGTGGGCTCGG
i711	aagaggca	CAAGCAGAAGACGGCATAACGAGATAagaggcaGTCTCGTGGGCTCGG
i712	gtagagga	CAAGCAGAAGACGGCATAACGAGATgtagaggaGTCTCGTGGGCTCGG

Supplementary Table S2. Table of primers for Illumina sample multiplexing and primer ID.

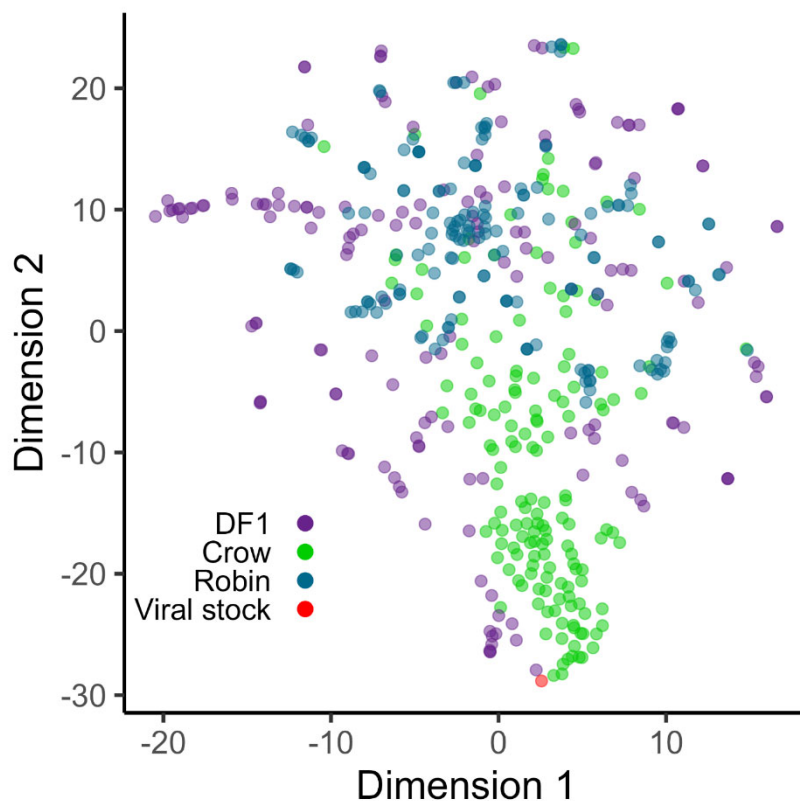
Name	Sequence
ID_cDNAWNV_7374_Rev	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGNNNNNNNNNCAGTGCCATCCACTACAGCGTTCT
R1_5'_WNV_for	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGNNNNNTCCCCTTCGTCGATGTTGG
5'_ID_Primer_Rev	GTCTCGTGGGCTCGGAGATGTGTAT
Illumina index i5 (N5XX)	AATGATACGGCGACCACCGAGATCTACAC[i5]TCGTCGGCAGCGTC
Illumina index i7 (N7XXX)	CAAGCAGAAGACGGCATAACGAGAT[i7]GTCTCGTGGGCTCGG



Supplementary Figure S1. WNV replicates in DF1 cells and *ex vivo* crow and robin PBMCs. Titers determined by plaque assay of DF1 cells (A), crow PBMCs (B) and robin PBMCs (C) infected with field strain WNV at MOI of 0.1, 1 and 10 (n=2 wells per cell type per MOI). Genome copies determined by qRT-PCR of DF1 cells (D), crow PBMCs (E) and robin PBMCs (F) (n=2 wells per cell type per MOI). WNV genome equivalents to plaque forming units (GE:PFU) at MOI of 0.1 (G) (*, DF1 vs crow PBMC, $P = 0.0215$; *, robin vs crow PBMC, $P = 0.0185$; 2-way ANOVA, Tukey's Multiple Comparison), 1 (H) (***, DF1 vs crow PBMC, $P < 0.0001$; ***, robin vs crow PBMC, $P < 0.0001$; 2-way ANOVA, Tukey's Multiple Comparison) and 10 (I) (***, DF1 vs crow PBMC, $P < 0.0001$; ***, robin vs crow PBMC, $P < 0.0001$; 2-way ANOVA, Tukey's Multiple Comparison) of DF1 cells, robin PBMCs and crow PBMCs.



Supplementary Figure S2. A) uninfected, B) MOI 1 and C) MOI 10 WNV-infected DF1 cells were stained using a FITC labeled-anti-West Nile virus monoclonal antibody. Cells were analyzed by flow cytometry and gated as positive or negative based on FITC (WNV) signal.



Supplementary Figure S3. T-Distributed Stochastic Neighbor Embedding (t-SNE) analysis to plot WNV barcode composition within individual cells in two-dimensional space.

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