



Leptospirosis and the Environment: A Review and Future Directions

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Abstract: Leptospirosis is a zoonotic disease of global importance with significant morbidity and mortality. However, the disease is frequently overlooked and underdiagnosed, leading to uncertainty of the true scale and severity of the disease. A neglected tropical disease, leptospirosis disproportionately impacts disadvantaged socioeconomic communities most vulnerable to outbreaks of zoonotic disease, due to contact with infectious animals and contaminated soils and waters. With growing evidence that *Leptospira* survives, persists, and reproduces in the environment, this paper reviews the current understanding of the pathogen in the environment and highlights the unknowns that are most important for future study. Through a systematic Boolean review of the literature, our study finds that detailed field-based study of *Leptospira* prevalence, survival, and transmission in natural waters and soils is lacking from the current literature. This review identified a strong need for assessment of physical characteristics and biogeochemical processes that support long-term viability of *Leptospira* in the environment followed by epidemiological assessment of the transmission and movement of the same strains of *Leptospira* in the present wildlife and livestock as the first steps in improving our understanding of the environmental stage of the leptospirosis transmission cycle.

Keywords: leptospirosis; Leptospira; environmental zoonoses; neglected tropical diseases; one health



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1. Introduction

1.1. Overview and Biology of Leptospirosis

Leptospirosis is a global zoonotic disease estimated to cause around 1 million cases and 60,000 deaths annually [1]. Caused by infection with bacteria from the genus *Leptospira*, leptospirosis presents a variety of symptomology ranging from asymptomatic to mild febrile illness to severe acute infection resulting in organ failure and death [1–5]. Additionally, as many as 30% of leptospirosis cases result in long-term health impacts [4,6]. There are many serovars and strains of *Leptospira*, with those that are pathogenic being of primary concern [2,3,5,7]. Leptospira reproduction usually occurs in the renal tubules of infected mammals [2,4,5,8]. Leptospira are excreted, via urine, into the environment where they may infect other organisms [1,8]. *Leptospira* infections usually occur through abrasions or wounds in skin or through contact with mucosa [5]. Leptospira can be transmitted directly to humans through the handling of infected animals, making leptospirosis an occupational hazard for people that regularly handle animals, such as livestock producers, abattoirs, veterinarians, hunters and game managers, animal control workers, and scientists [1–4,7,9–11]. As the local prevalence of leptospirosis and the frequency of exposure to Leptospira determines the risk of infection for an individual, individuals in regions with increased workplace procedures and protections such as surveillance, diagnostic testing and treatment of infected animals, and access to personal protective equipment-such as gloves, goggles, and boots—are less likely to contract leptospirosis [1,3,4,7,11–13]. More commonly, leptospirosis is contracted indirectly through contact with contaminated water or soil [2,4,5,7,8,11]. As a result, leptospirosis remains an occupational hazard for individuals that work closely with soil and water systems, such as sewage and waste workers, construction, military, aquaculture workers, and farmers [1,4,7–9,11,12,14–18]. Leptospirosis is also a growing environmental hazard for outdoor recreationalists—such as kayakers, swimmers, fishermen, etc. [7,12,14–16,19–33]. As outdoor recreation continues to grow, diagnosis of leptospirosis is expected to grow comparably.

In addition to being an occupational and environmental hazard, leptospirosis has been designated by the U.S. Centers for Disease Control (CDC) as a neglected tropical disease (NTD), characterized by its impacts on vulnerable populations and the diversity of epidemiological settings the disease is found in [3,11,34]. Due to the disproportionate impacts of the disease, epidemiological efforts to control leptospirosis are best focused on the most vulnerable populations, including socioeconomically disadvantaged populations in high-density urban environments and those in closest contact with infected animals, such as rural farmers [1,4,11,16,35]. Studies across the globe find significantly increased cases of leptospirosis and disproportionate transmission rates of *Leptospira* in communities of lower socioeconomic status due, in no small part, to inadequate sanitation infrastructure and/or lack of enforcement of safe drinking water standards [4,16,35–37]. Increased local prevalence of the disease and frequency of exposure determines the transmission rates, so these findings result from greater local prevalence of the disease-causing pathogen and greater frequency of exposure in socioeconomically disadvantaged communities. The frequency of exposure to the pathogen and the local prevalence of the disease is largely due to environmental conditions, which we will discuss in further detail later in the review. However, it is important to note that many of the environmental conditions associated with lower socioeconomic status and rates of leptospirosis result from social inequality. These conditions include contact with sewer water, trash accumulation, and reduced rates of governmental sanitation interventions, such as failure to control rodent populations, maintain municipal trash removal and sanitary sewers, and provide universal access to potable drinking water [4,38]. In addition to the conditions associated with leptospirosis that result from socioeconomic inequality, the burden of leptospirosis on socioeconomically disadvantaged communities further facilitates cycles of poverty due to the expense of medical treatment, lost wages while ill, and persistent long-term health effects of the disease. Thus, leptospirosis can serve as both the result of and the cause of poverty [3,34,39]. The burden of leptospirosis and its role in increasing poverty is only expected to grow through increasing urbanization, which often outpaces the establishment of governmental sanitation interventions in socioeconomically disadvantaged communities [40,41]. Rapidly increasing urbanization will likely increase the conditions associated with the disease, as will be further discussed later in this review.

1.2. Current Status of Leptospirosis

Currently, leptospirosis is a disease of significant concern. The World Health Organization (WHO) estimates that globally the endemic human leptospirosis rate is 5 cases per 100,000 people annually and the epidemic human leptospirosis rate is 14 cases per 100,000 people annually [42]. However, more recent estimates suggest that leptospirosis is "among the leading zoonotic causes of morbidity and mortality" and causes around 1,000,000 cases annually [1]. As shown by the conflicting estimates, the current global burden of leptospirosis is difficult to determine.

The current challenges surrounding obtaining a global estimate of leptospirosis morbidity and mortality largely stem from the compounding issues that arise when critical components of a disease's lifecycle are not understood. For example, in an effort to clarify the global issue of leptospirosis, the World Health Organization (WHO) leads an initiative seeking to improve global estimates of morbidity and mortality of the disease through efforts to address the lack of surveillance, adequate diagnostic tests, and prompt treatment as well as the rates of misdiagnosis and underdiagnosis of the disease [1,11,16,43–48]. The issue of misdiagnoses and underdiagnoses results from and are due to, in part, to a lack of understanding on the prevalence of leptospirosis in a given region, as this hinders physicians from knowing whether leptospirosis should be tested for [1,44,49]. Additionally, of concern is who (individuals, health insurance, or public health initiatives) is responsible for the costs of testing for and treatment of the disease. The ability to measure the prevalence of a disease usually depends on standard surveillance of livestock, wildlife, and human infection. In the United States, these are federally mandated and funded disease surveillance programs that include screening, sampling, testing, treatment, and testing components with required reporting. Outside of the United States, the WHO makes recommendations for testing and reporting within member nations. Depending on the level of urgency and committed funds, some testing and reporting of NTDs, such as leptospirosis, occurs at a member nation's laboratories or at regional WHO reference laboratories. Lower priority diseases may only be available through private laboratories. The major limitation of such surveillance approaches is that the priority ranking and allocation of funding is based on national or regional prevalence and transmission rates, which can be greatly masked or suppressed by inadequate testing. Additionally, the standard methods of disease surveillance are inadequate for leptospirosis as there is growing evidence supporting significant Leptospira transmission in the environment [1]. However, very little is known about the survival, persistence, and reproduction of *Leptospira* in the environment or about the routes of transmission of Leptospira within and from the environment. This makes efforts to improve our understanding of the environmental stage of the leptospirosis transmission cycle essential to understanding global morbidity and mortality rates. As our knowledge of where the disease is most prevalent, has the most severe impact, and the routes of transmission that facilitate the persistence and spread of the disease in such communities increases, our ability to respond to and reduce the disproportionate impacts of leptospirosis will improve.

While already imperative, addressing the disproportionate impacts of leptospirosis will become increasingly more crucial as climate change increases the frequency and severity of severe weather events and the rapidly changing climate increases the burdens on socioeconomically disadvantaged communities in tropical nations [4]. Closely associated with severe weather events, leptospirosis outbreaks are common across the world following floods, storms, and other mass precipitation events [8,50–53]. As such, it is crucial that efforts are made to better understand the environmental stage of the disease cycle so that better disease surveillance, diagnostics, and disease interventions can be implemented.

1.3. Objectives of this Review

In response to leptospirosis' status as a neglected tropical disease and the growing need to understand the environmental stage of the disease's transmission cycle, this review examines the literature, synthesizes the current understanding, and highlights future directions for further research. We will cover leptospirosis in the three major components of the environment: water, soil, and the organisms that facilitate the movement and transmission of the pathogen in the environment. In addition, we will also review how these components may differ along the urban–rural gradient. Due to the global extent of *Leptospira* and the impacts that leptospirosis has on human and animal health worldwide, our review will follow a broad, global discussion of the literature and findings.

2. Materials and Methods

To compile this review, we used numerous literature bases, including Web of Science, BioOne Complete, Elesvier, and several others (see list of databases searched at hhtps://www.lib.auburn.edu/find/bytitle.php, accessed from 1 September 2022). Within these literature bases, we conducted a Boolean search including the combination of the following search terms: [Leptospir* AND (transmission OR water OR soil OR water OR wildlife OR environment OR mortality OR ecology OR survivorship OR urban OR rural)]. No age range was selected. Potential publications were reviewed by the authors to determine those that were comparable in study subject, study focus, and methodological approach; this was performed to focus on leptospirosis in context of the environment and remove strictly papers unrelated to *Leptospira* in the environment. If the papers did not contain a focus on the environment or contain a focus on the given search terms, then they were excluded. Then the results of these studies were synthesized for this review, whether their findings contrasted or supported each other. A total of 1,463 publications were found using our literature bases for use in this review. Once these publications were collected, more than 500 duplicate publications, more than 300 out-of-scope publications, and more than 300 publications without full-text were removed from our list of reviewable manuscripts. Ultimately, ~220 publications were selected to be cited in this review. Any additional publications cited in this review were used because they covered information pertaining to topics covered in the review or they otherwise improved this review with their content. These papers were largely examples of environmental studies that have been performed on other zoonotic pathogens, such as *Escherichia coli*, but have yet to be conducted for *Leptospira*.

3. Leptospirosis in the Environment: Water

While leptospirosis is reliant on the continuous cycle of transmission of *Leptospira* from infected to uninfected animals, there is growing evidence that there is long-term survival and viability of the pathogen in the environmental stage of the transmission cycle [8,54–56]. This is important because the opportunity for transmission of the pathogen increases the longer that *Leptospira* survives and remains a persistent feature in the environment. With the recent laboratory-based discovery of *Leptospira* replication within waterlogged soil emphasizing the critical role that water serves for *Leptospira* outside of animal hosts, it is more apparent than ever that future research should be conducted to better understand the survival, persistence, and reproduction of *Leptospira* in waterways [54,57].

3.1. Current Understanding

Due to the frequent linkage of outbreaks with exposure to contaminated waters, leptospirosis has often been discussed as a water-borne disease [3,58]. Additionally, higher rates of leptospirosis are linked to regions with greater percentages of riparian habitat and surface waters [59]. While not strictly water-borne, outbreaks of leptospirosis following floods, storms, and other mass precipitation events have been documented across the world [8,50–53].

Outbreaks of leptospirosis following floods, storms, and other mass precipitation events originally linked the disease to environmental waters; however, many detection techniques have developed over the decades to facilitate active surveillance of the disease in environmental waters. The most promising techniques for the study of pathogenic Leptospira in waters in the environment are quantitative real-time polymerase chain reaction (PCR) tests that target only pathogenic *Leptospira* spp. [60,61]. There are key genetic features that appear to be conserved in pathogenic *Leptospira*, making detection techniques that target these retained gene patterns the preferential method as they allow for discussion of public health implications and comparison to animal and human studies in the area (provided that they used serological and molecular diagnostic techniques that target the same pathogenic gene patterns) [55,60,62]. Optimization in recent years of these methods for the quantification of pathogenic *Leptospira* in environmental waters has accelerated studies of environmental Leptospira and tracing of human leptospirosis to environmentally present Leptospira in regions across the world [63,64,64–66]. Additionally, these advances are accelerating the rate of further optimization of methods, which improves the testing in other conditions (such as soil) and the results available for modeling [67,68].

One of the most critical and currently perplexing issues in the study of leptospirosis and the survival of *Leptospira* in environmental waters is the range of conditions in which the pathogen is present and surviving [8]. Survivorship or hardiness of *Leptospira* in water covers a wide range of conditions. For example, *Leptospira*, both pathogenic and not, appear capable of surviving long periods under both low and high temperature ranges [8,69–71]. They also appear to survive in a range of pH levels, dependent on the strain and the presence of other microbiota, such as bacteria and fungi [8,69–74]. Additionally, pathogenic

Leptospira seem capable of surviving in waters with low nutrients, which indicates that they are likely present in streams and waterways across a far greater range of physiographic and hydrologic conditions than previously expected [8,69,70]. Not only do they appear capable of surviving in low nutrient conditions, but they also appear to retain virulence over long periods of starvation [75]. However, in our review of the literature, we found an extremely limited number of field-based environmental studies attempting to evaluate the survival of *Leptospira* in the environment [55,56,76]. Due to the lack of studies, we have yet to identify results about environmental conditions that facilitate the survival of *Leptospira* in surface waters [Table 1].

Table 1. Environmental studies of *Leptospira* survival in water. Only studies conducted in the field were included in the table.

Study Location	Leptospira Studied	Sampling Success	Survival	pН	Temperature	Citation
Tawain	Pathogenic and intermediate	NA ¹	7 days	7–8	0–30 °C	Ryu et al., 1966 [76]
Philippines and Japan	Multiple pathogenic and intermediate	31/73	0 days	NA ²	NA ²	Saito et al., 2013 [56]
New Caledonia	<i>L. interrogans</i> Pyrogenes	0/10	0 days	NA ²	~28 °C	Thibeaux et al., 2017 [55]

¹ Involved inoculated samples being placed into environment. ² Not measured.

3.2. Future Directions

As we have previously mentioned, there are many unknowns about *Leptospira* in environmental waters. Through our review, we have determined several key areas in need of targeted study. Primarily, there is a substantial need to focus on gathering information on the conditions of survival, persistence, and potential reproduction of pathogenic *Leptospira* in natural waters. It is essential that these studies be conducted in field conditions so that the variability of natural conditions informs the finding of the probable capabilities of *Leptospira*. Additionally, there is a substantial body of research in the literature exploring methods of detection and survivability of *Leptospira* in water sources, but there is still a substantial need to apply these methods in natural conditions to determine the survivorship of viable and infectious *Leptospira* [8,60]. Studies should use natural systems, target viable and infectious pathogenic *Leptospira*, and consider the complex processes and systems that drive the results of the study. With these specific considerations in place, we recommend the following additional future directions:

Firstly, lack of surveillance, which results from, and is at least partially attributable to, the costs and challenges with detection methods, limits our current understanding of the role of environmental waters on the transmission cycle of leptospirosis. One of the limitations for the measurement of *Leptospira* in the environment is that methods are often optimized for sampling under certain conditions over others and, thus, result in variability between conditions that hinder interpretability of results. For example, sampling of environmental water sources by methods that incorporate filtration can be complicated by the clogging of the sample filters as a result of physiochemical characteristics of the water such as increased concentrations of suspended solids, dissolved organic carbon, and other dissolved nutrients [63,67,77,78]. However, alterations to methods and procedures in a recent study provide evidence that variability in quantification of Leptospira collected from different water sources—such as ponds, streams, and sewage waters—can be reduced and accounted for [63]. Another limitation for the surveillance of *Leptospira* in the environment results from the complex and time-consuming nature of Leptospira isolation. However, methods for the identification of Leptospira from environmental sampling have been improved in recent years resulting in reduced laboratory time constraints [63,67,78,79]. One such method involves the enrichment culturing followed by PCR targeting the gene *lipL32*, which allows researchers to confirm the presence of pathogenic *Leptospira* [78]. Other studies target the 16s rRNA gene described by Schneider et al. 2018 [8,67,80]. The latest studies are beginning to evaluate the use of next-generation sampling—such as Oxford Nanopore Technologies—and mass spectrometry—such as matrix-assisted laser desorption ionization time-of-flight mass spectrometry using MALDI Biotyper Systems—for the sequencing of samples directly [78]. These improvements in methodology have resulted in studies reporting the identification of novel *Leptospira* from the environment, with one such study resulting in the identification of twelve novel species [79,81]. In addition to allowing for direct sequencing of environmental samples, the improvements to methods in recent years have created promising solutions to one of the most critical distinctions when studying *Leptospira* in the environment: whether the measured *Leptospira* are pathogenic or not. One of the final limitations to environmental surveillance of *Leptospira* is the difficulties with identifying whether measured *Leptospira* are virulent or not. As the point of conducting environmental studies of *Leptospira* transmission is to determine whether the environment is serving as a reservoir and contributing to the transmission of the disease to uninfected organisms, measures of the infectability and viability of environmental Leptospira are critical. However, recent applications of methods and techniques, such as culturing/optical densities or viability PCR, are creating promising solutions to this issue [55,74,82]. By utilizing approaches that focus on the viability of pathogenic strains of *Leptospira* from environmental samples, studies are beginning to identify potential factors that facilitate the survival of *Leptospira* in surface waters, such as calcium, iron, and pH [74]. As such, the advancements in methods and approaches in the past decade are rapidly addressing many of the current limitations of environmental surveillance of Leptospira and will serve a critical role in addressing the unknowns of its environmental persistence, survival, and transmission.

Secondly, understanding the driving mechanisms behind the persistence, survival, and transmission of *Leptospira* through water has proven elusive [54]. The characteristics of the waters in question, as well as the biogeochemical and hydrologic processes these characteristics result from, have significant influences on these driving mechanisms. As mentioned previously, pathogenic *Leptospira* survives for long periods in a range of different pH levels, nutrient levels, and temperatures. Such water quality metrics originate from biogeochemical and hydrologic processes associated with the stream environment. Despite this, few studies exist that consider such factors. As a result, there is a severe scarcity of information on the impacts of basic water quality metrics, such as nitrate levels, on the abundance and survivorship of Leptospira. Additionally, the standard water quality metrics that studies are including have the potential of being misinterpreted without additional biogeochemical and hydrologic processes being considered. For example, reports suggest that Leptospira concentrations increase with the turbidity of streams, a stream characteristic that often rises following large rainfall events and flooding events [8,83]. One theory behind this observation is the capacity of elevated turbidity levels to shield bacteria from UV irradiation through increased levels of suspended solids, which is a direct threat to *Leptospira* persistence and survival in surface waters in tropical regions [84]. The mechanism of suspended solids shielding pathogens from UV radiation has been demonstrated with fecal indicator species in natural conditions [85]. However, as a stream characteristic and water quality metric, turbidity reflects cloudiness in water; turbidity cannot be used directly as a measure of suspended sediments since cloudiness may stem from other sources including tannins. This is just one example of the potential for misinterpretation without attention being paid to specific processes and stream characteristics that may influence what is being observed. Additionally, this study did not identify a *Leptospira*-specific study of this mechanism and, thus, it remains unknown in the literature.

Another future direction is more attention to the intrinsic properties and behaviors of *Leptospira* and how that may facilitate survival, persistence, and reproduction in environmental waters. One such example is the ability of some strains, particularly pathogenic strains, to metabolize urea, which may improve the ability to survive for longer periods in smaller waterways particularly those that are nutrient-poor [86–88]. However, we

are unaware of studies that attempt to assess this in natural waters so future research is needed. Additionally, part of the survival of *Leptospira* in freshwater may be the result of interactions with environmental bacteria and the formation of cellular aggregations called biofilms [89–92]. These aggregations are held together through an extracellular matrix made of nutrients such as nucleic acids, proteins, lipids, and polysaccharides, which creates a more hospitable environment for the pathogen alongside other microbial species that may be present in the environment [89,93]. Future studies into the survival and viability of *Leptospira* in the environment should focus on biofilms including microbial relationships and conditions found in communities of *Leptospira* in streams. Existing methods found in water microcosms may be improved by the methods and frameworks used in the extraction and quantification of biofilms in medical contexts [94,95]. If these methods prove useful, then attempts to use them in field studies would be the next steps for assessing the impacts of these intrinsic properties of *Leptospira* and their interactions with other bacteria.

Finally, in addition to future studies considering the biogeochemical processes and hydrologic characteristics of the waters being studied, hydrologic and climatic modeling could be used to improve the risk maps created by socioeconomic and epidemiological modeling that help identify areas most suitable for long-term persistence and transmission of *Leptospira* [37,68,96–100]. These types of models are improved with the addition of new data, so long-term collaborative mapping between experts in different specialties would provide the maps most useful for targeting surveillance, management, and future research in regions where the use of such resources would be most effective. These maps would also facilitate discussion of region-specific trends of environmental persistence and transmission of *Leptospira*, which—due to limited studies to draw from—are currently infeasible to draw conclusions on.

4. Leptospirosis in the Environment: Soil

Leptospira, both pathogenic and not, have been found in soils globally [8,56,57,62,64,95,101–109]. Contact with *Leptospira* in the soil is one of the leading routes of infection with leptospirosis, resulting in outbreaks across the globe [2–4,7,9,12]. As mentioned previously, outbreaks of leptospirosis following floods, storms, and other mass precipitation events have been documented globally [8,50–53]. The leading hypothesis for this phenomenon is that outbreaks of leptospirosis result from the resuspension of the pathogen from the soil, where viable *Leptospira* may persist for extended periods [8,52,53]. However, just as with the current understanding of *Leptospira* in water, very little is currently understood about the growth, survival, and long-term persistence in soils. As new evidence supporting *Leptospira* multiplication in soil suggests, understanding the current knowledge and working to address existing gaps in the literature on *Leptospira* survival, persistence, and reproduction is essential [57].

4.1. Current Understanding

As discussed earlier, a general understanding of the risk of leptospirosis has emerged regarding the environmental conditions under which transmission to and subsequent infection of humans may be most susceptible to exposure. A high risk of transmission is associated with tropical settings, major precipitation events such as hurricanes, and contact with wet soils and surface waters that are contaminated by infected mammals and other animals [8,110]. These risk factors are sufficiently well-established to be used in alerts provided by public health agencies, such as the CDC [111]. Similarly, most agree that survival and persistence of *Leptospira* are higher in soils than in surface water, although this could reflect the difficulty of detecting the pathogen after major dilution occurs in surface waters [8,107]. Ultimately, we lack a detailed understanding of why there might be greater Leptospira numbers in soils or what potential mechanisms would be driving such a phenomenon [54].

We are unaware of studies assessing the texture and other physical properties of soils associated with Leptospira survival. However, the texture—portion of sand, silt, and clay-

sized particles that comprise the mineral fraction of the soil—and physical properties of soils—such as soil moisture content, pH, organic matter, soil organic carbon, total nitrogen, and the carbon to nitrogen ratio—can have significant impacts on the survival and persistence of zoonotic pathogens and microbial populations in soils [112–114]. Additionally, the texture and physical properties of soils in streams and waterways can directly influence Leptospira survival and persistence. Fine sediment material is readily suspended in the water column, while large soil particles, such as sand, usually drop out of the column. This can affect the deposition and resuspension of zoonotic pathogens in natural streams [115,116]. Following major rainfall events, clay sediments tend to be carried easily with sheet flow and add to turbidity as would organic particles. In addition, Leptospira bacteria may adhere to soil particles through adsorption, which is a trait that facilitates stream suspension [8,55]. Clayey soils and organic particles are more active in the adsorption of materials compared to sands since they have more surface area per unit of mass. Consequently, we suggest that soils providing the most protection and the highest likelihood of movement with floodwater would be finer textured clay soils with significant organic matter content. In addition to the capacity of clayey soils to facilitate the movement of water-borne pathogens through their resuspension rates and high runoff potential, clay soils have a greater field capacity than sandier soils and, thus, hold on to more water than other soil textures [117]. The greater capacity of clay soils to hold on to water likely facilitates *Leptospira* survival. Statistical support for this hypothesis is demonstrated within studies from America Samoa and the Netherlands, which both identified a significant positive association between cases of leptospirosis and clay soils [105,118]. Although more research into this potential association is needed, their findings support the widespread agreement that wetter soils, i.e., greater than 20% moisture, and waterlogged soils serve as environmental reservoirs for Leptospira [54,57]. However, it is unclear from some of these studies whether the soils used in the studies were truly anoxic or not. If these soils were truly under anoxic conditions, then not only would the transformations of nutrients—such as nitrogen—occur following anaerobic mechanisms but also by anaerobic bacteria species. An example of this is clearly demonstrated with the transformation of nitrogen in anaerobic soils conditions through anammox or through denitrification, both of which are processes led by species from different genera of prokaryotes [117]. Thus, depending on the conditions the soils being studied are exposed to, there could be significantly different microbial communities and biofilms, which may greatly influence the survival and persistence of Leptospira. Additionally, if Leptospira bacteria are found to be surviving and persisting in truly anoxic conditions, then it may be time to reclassify Leptospira from obligate aerobes to facultative anaerobes [119]. This is an important distinction, because if the bacterium is facultatively anaerobic, then it may be persistent across a wider range of soil conditions than previously thought.

Additionally, there is little information, much of which is conflicting, regarding the range of soil physiochemical properties that are most conducive to Leptospira persistence [54,104]. Some reports indicate that neutral to high pH conditions (i.e., 7-8) are most suitable [54,55]. However, this assertion conflicts with positive correlations between Leptospira persistence and concentrations of Fe, Mn, and Cu, elements that increase in concentration under acidic pH conditions, and negative correlations with Ca, which increases with higher pH conditions [104]. As in waters, the ability of Leptospira to survive and persist under such a range of often adverse soil conditions may be the result of biofilms, which create more suitable conditions for bacterial survival in soil, as has been seen with other zoonotic pathogens such as *Escherichia coli* [120]. Early study into this suggests that Leptospira are capable of producing these biofilms in soils and with several other species of bacteria found in the environment [93,94,121,122].

4.2. Future Directions

Much of the current literature gaps involving leptospirosis and soils exist due to a lack of field study and research on soil characteristics and dynamics. We know very little about what strains of *Leptospira* are present in the environment. As strains of *Leptospira*

from across the genus are present in soils, future studies should seek to assess the full diversity of *Leptospira* that persist, survive, and reproduce in soils. We expect great diversity of *Leptospira* in the soil as 12 new species were identified in a single study [79]. Assessment of *Leptospira* in soil should not be confined to searches for strains found in the traditionally 'pathogenic' subclade P1, as strains from both subclades are linked with human cases of leptospirosis and survival in soil. For example, both *L. interrogans* and *L. licerasiae* are linked to human cases of leptospirosis and survival in soil environments [95,123].

In addition to identifying the strains of *Leptospira* that are present in the environment, future study is needed to determine the soil conditions required for the persistence, survival, and reproduction of each strain. Our first recommendation is that care must be taken not to neglect describing and measuring the soils being used within future studies. As soil microbiologists can attest to, the biogeochemical processes of soils are complex and dynamic systems that are certain to greatly influence the strains of *Leptospira* that survive, persist, and reproduce in the soil. Future study is also needed to assess the conditions that facilitate the virality of *Leptospira* in the environment, as this is a key question of epidemiological concern. Knowing so little about the persistence, survivorship, and reproduction of individual Leptospira spp., we cannot begin to determine or speculate whether the required conditions for each are common among the genus or are specific to each strain. As a result, we highly recommend that future studies include reference to the soil survey for the study site, which contains the soil classification, general information about the geology, topography, and climate of the area, and list the types and volumes of soils in the area. Such surveys within the United States can be accessed online via the National Cooperative Soil Survey. Outside of the United States, local soil scientists should be contacted for assistance accessing local soil surveys. In addition to referencing the soil survey for the classification of the studied soil, an assessment of the physiochemical properties of the soils is essential. We recommend that such assessments include physical and chemical indicators of soil health, such as nutrient levels, bulk density, soil moisture, and texture.

While there is evidence that *Leptospira* can interact with other soil biota and create biofilms that may improve their persistence and survival in adverse soil conditions, there is little else known about the interaction of *Leptospira* with other soil microbiota and the role that these interactions (direct or otherwise) may have on the pathogen's survival [91,121,122]. With so little known about the topic, any future studies would be widely beneficial to beginning to form hypotheses on the long-term survival, persistence, and reproduction of *Leptospira*. In addition, as was emphasized in the discussion of *Leptospira* and environmental waters, it is imperative that future study be conducted in natural environments. Lab-based studies are unlikely to include the tremendous variability in biotic, soil, and climatic conditions that are known to influence soil microbiota. As a result, the findings of such lab-based studies are unlikely to represent the probable survival, persistence, or reproduction of *Leptospira* in natural conditions.

5. Leptospirosis Enzootic Persistence in the Environment: Organisms of Concern

Based on our current understanding of the lifecycle of *Leptospira*, the enzootic persistence of the pathogen in water and soil is reliant on the continual excretion of the pathogen by infected animals. Leptospirosis affects many animal species, both domestic and wild [1,8,124]. Despite most mammals serving as competent hosts and vectors of *Leptospira*, very little is known about the pathogen load excreted by different species or the role that specific species play in establishing and maintaining environmental sources of the disease [54]. Additionally, as impetus for a universal vaccine does not appear to be available soon, and with international trade of domestic and wild animals likely to increase and contribute to the global spread of leptospirosis, we have outlined some of the current knowledge surrounding the disease in native and introduced species, with particular attention paid to species with the most potential or known capacity to spread the disease [125,126].

5.1. Domestic

Domestic, non-native species are often the dominant reservoir of *Leptospira* in rural regions and urban regions [4,127]. Some domestic species are extremely widespread, transient in nature, and continue to shed *Leptospira* persistently in their urine long after their initial infection has subsided [127]. For this review, four such domestic animal groups (rats, dogs, pigs, and cattle) are used to illustrate the capacity that domestic species serve in the persistence, spread, and levels of *Leptospira* in the environment.

The black rat, *Rattus rattus*, and the brown rat, *Rattus norvegicus*, are largely considered the most important sources of leptospirosis [3,4,12,20,54,59,127–132]. Introduced globally, the black and brown rat continues to thrive alongside human populations in both urban and rural settings. Rats are asymptomatic carriers (hosts) for many strains of pathogenic Leptospira, are commensal hosts that are persistently infected, and shed the pathogen at levels that exceed many other competent hosts for the disease $(7 \times 10^7 \text{ per day})$ [132–134]. Linked historically to human outbreaks of leptospirosis across the globe, rats continue to play a significant role in the transmission of the disease in urban environments, particularly in low-income communities [128–131,134–140]. However, the role of rats in the disease cycle of leptospirosis is not limited to urban settings, as they can be quite prevalent in rural regions as well [134,141]. Perhaps the best studied of the animals associated with leptospirosis, there appear to be critical gaps in the literature remaining to study about the role that rats play in the persistence and load of Leptospira in the environment. Studies that seek to link strains of Leptospira in the environment with their animal host will encounter rats due to their global distribution, high population densities, and competency to spread the pathogen.

Domestic dogs are also highly competent hosts and participate in the spread of the pathogen globally [127,142–144]. Despite spending so much time in close proximity to humans, dogs do not seem to contribute to outbreaks in humans and instead seem to impact the disease cycle through their contribution to environmental contamination [145,146]. The majority of strains of canine leptospirosis cases are now associated with the environment, largely from contaminated waters [142]. Once infected, domestic dogs can spread the Leptospira (via urinary excretion) across large distances as a result of their highly mobile behavior (averaging 5 km per day and up to 330 km during human-mediated activity in one study) [147]. Not only are dogs able to spread *Leptospira* long distances, but they also excrete large numbers of the organism in their urine $(1.6 \times 10^5 \text{ per day})$ [132]. In addition to the long distances traveled by dogs and the large number of the pathogen excreted, shedding of Leptospira has been documented to last anywhere from four to six weeks—and in some cases for several years [148]. However, despite their capacity to spread *Leptospira* long distances and in great quantity over a substantial time period, little to no research has been conducted to evaluate whether leptospirosis outbreaks in wildlife are the result of canine-shed Leptospira. This is a key gap in the literature because vaccinations are available for canines and could be used to reduce the environmental load of Leptospira. As mass vaccination efforts are resource and effort-intensive, it is important that the capacity of dogs to contribute to wildlife leptospirosis outbreaks be assessed accurately and quantitatively prior to such efforts being made. Additionally, as indicated in the great spatial range facilitated by the movement of dogs by humans, dogs—particularly unowned, unconfined, or free-roaming dogs-may serve a critical role in the transport of infectious Leptospira along the urban–rural gradient.

Leptospirosis is a well-established disease of the ungulate species, *Sus scrofa*. Pigs, domestic or wild, can spread the pathogen to humans directly as frequent infections in hunters and pork producers illustrate [149–154]. Pigs may be significantly impacted by leptospirosis, causing reproductive failures including infertility, premature and stillbirth, and fetal death [154–156]. Additionally, infected pigs may have persistent renal infections and persistent shedding of *Leptospira*. Due to the unique foraging behavior of pigs called rooting, which involves the turning of soil with their snouts, they are exposed to *Leptospira* in the soil that has direct contact with the mucosa in their snouts. Another of their behaviors,

wallowing in the soil, may also play a role in the transmission of *Leptospira*, as other pigs and many other wildlife species drink from these wallows [157]. A study evaluating the prevalence of *Leptospira* in pig wallows and molecular tracing of the disease in wildlife from the area would reveal whether these wallows are playing a role in the transmission of leptospirosis in the environment. In addition, their natural social behavior encourages and provides close contact between potentially infected and non-infected individuals in wild environments as well as in domestic production [158]. As such, pigs are uniquely well-suited for contributing to both the introduction and persistence of *Leptospira* in the environment. However, very few studies have investigated the role that pigs have in the environmental transmission of Leptospira. Most existing studies have focused on the prevalence of leptospirosis in pigs and the capacity of wild pigs to serve as the reservoir for *Leptospira* to transmit to domestic pigs and livestock [159–162]. The potential for wild pigs to spread the disease is of significant concern throughout their range, both native and non-native [150,163–171]. This is due to a striking increase in feral pig populations across the globe and how common zoonotic pathogens are in the species, with 87% of swine pathogens listed by the World Organization for Animal Health being zoonotic, impacting humans, livestock, and wildlife species [163]. Their foraging behavior puts them in close contact with soils, and pigs spend significant time in and around waterways due to their inability to effectively thermoregulate. As a result, they contaminate waterways directly through their time in and around waters and indirectly through their waste [172]. Although we are unaware of studies directly evaluating soil and water contamination with *Leptospira* resulting from runoff and spills from swine sewage lagoons from swine-rearing facilities, studies indicate that swine pathogens can contaminate waters and soils surrounding the swine industry [173]. Therefore, future studies evaluating pig contamination of waters and soils are needed. Additionally, despite their tremendous potential for disease transmission, we were unable to find a study evaluating the levels of *Leptospira* excreted by pigs in their urine.

Due to the economic burden of the disease on livestock production, leptospirosis in cattle has long been studied [174,175]. Much of this economic impact results from reproductive failures including infertility, premature and stillbirth, and fetal death [159,176]. In addition to the economic impacts, leptospirosis threatens humans through direct transmission of *Leptospira* from cattle [4,16,174,177,178]. Due to management strategies such as allowing unrestricted access to natural water sources, cattle are infected by and contaminate surface waters with *Leptospira* [178,179]. The unrestricted access to surface water also puts cattle in close contact with potentially infected domestic species such as pigs or wildlife species. As discussed, one of the major concerns about wild pigs is their potential ability to spread leptospirosis to cattle. We are unaware of a study linking leptospirosis infection in cattle with strains of Leptospira found in nearby wildlife species, such as wild pigs. Future studies should evaluate this, as direct evidence may support modification of the management of livestock in regions with endemic or highly prevalent *Leptospira* in the environment. Other livestock management strategies such as large herd sizes, the introduction of new cattle to existing herds, and keeping pets on the farm all result in a greater risk of leptospirosis among the herd [180]. Additionally, cattle are highly capable of shedding large amounts of *Leptospira* (6.3×10^8 per day) [132]. Fortunately, considering the high rates of Leptospira shedding found in cattle, there are highly effective vaccinations available for cattle [181]. However, one consideration of leptospirosis vaccination in cattle is that it is generally only effective for about a year and livestock producers need to continuously vaccinate their herds to prevent the transmission of the disease in their herds. Additionally, evidence suggests that the available vaccines only protect against selected significantly pathogenic/commercially important strains of *Leptospira* [182].

5.2. Wildlife

Generally, large herbivores and small mammals are considered to be the most important wildlife species of concern for transmission and enzootic persistence of leptospirosis [8]. However, some evidence suggests that large carnivores such as lynx and wolf species are exposed to leptospirosis frequently—likely in the small rodents and other mammalian prey that they consume [183–185]. Wild rodents, including large rodents such as capybara (*Hydrochoerus hydrochaeris*), beavers (*Castor fiber* and *Castor canadensis*), and various species of smaller rodents such as squirrels and mice, are also common wildlife species that carry leptospirosis [186–201]. Leptospirosis can be extremely common among small mammal populations in an environment [185,186,202], as shown in one such study that reveals 62.4% of small mammals tested carried *Leptospira* [203]. However, as previously discussed, it is difficult to begin to speculate on their impacts on environmental *Leptospira* without further research assessing the shedding rate and urine loads of *Leptospira* in small mammals. It is also clear from the existing literature that future studies must consider the movement of leptospirosis among wildlife species in a broader context with specific attention to what species the disease routinely transmits to and from [204].

While primarily a pathogen associated with mammalian hosts, evidence exists of pathogenic and non-pathogenic Leptospira infection in many species of herpetofauna (amphibians and non-avian reptiles) [110,205–218]. Human leptospirosis is known to occur with the handling of infected herpetofauna species, particularly during interactions with captive species [211,212,219]. Many of the herpetofauna species that have tested positive for infection with *Leptospira*, particularly turtle species, spend significant time in and around waterways where they may be contributing to the transmission of zoonotic pathogenic *Leptospira* spp. such as *Leptospira interrogans* [220]. In tropical forests, ground-dwelling herpetofauna, such as some species of snakes, may contaminate the leaf litter and soil, potentially infecting small ground-dwelling mammals. As mammalian species are best known as hosts of leptospirosis, they may be more competent reservoirs and hosts of the pathogen. This could mean that herpetofauna transmits the pathogen to species that are more significant in the disease transmission cycle. Conversely, the opposite may be true. Unfortunately, very little is known about the disease cycle in herpetofauna. A critical gap is the lack of knowledge on the infectious load of *Leptospira* and rate of *Leptospira* excretion in infected species. Future leptospirosis studies in herpetofauna should address the routes of infection, the rate of excretion into the environment, and the role herpetofauna play in the persistence of *Leptospira* in the environment.

Another concern regarding the enzootic persistence of *Leptospira* in the environment is the potential impact on threatened and endangered species. It is well-established that the threat of disease increases as a species becomes more endangered [221]. Endangered wildlife species populations, such as that of the Iberian lynx (Lynx pardinus) and the island fox (Urocyon littoralis), may be threatened by leptospirosis as well as play a role in the environmental transmission cycle [222,223]. Leptospirosis may also be a threat to threatened marine mammals such as the Amazonian manatee (Trichechus inunguis) and the North American manatee (Trichechus manatus) [224-229]. Much is unknown about the disease in manatees, while other marine mammals suffer clinical disease and reproductive failure with *Leptospira* infection [230]. Therefore, further research is needed to understand the impacts of leptospirosis on threatened and endangered marine mammals. Additionally, Leptospira in the environment may threaten the implementation and success of efforts to re-introduce endangered species. Leptospirosis was a serious threat to the re-introduction of the Eurasian beaver (Caster fiber), which was formerly extirpated in much of its range and recently reintroduced in several European nations such as Scotland. Infection with Leptospira sometimes result in fatal clinical disease in Eurasian beavers [192,231]. Due to the potential for fatal clinical disease, careful monitoring and surveillance were taken with the re-introduction effort [232–236]. Another such study investigating the re-introduction of the endangered water vole, Arvicola amphibius, in the UK found that 42.9% of re-introduced voles were infected with Leptospira only four months post-release [237]. While there is currently no indication of impacts on this specific population from this infection (although this may be due to lack of study), leptospirosis is an exposure-dependent disease and other co-occurring species may be vulnerable to the elevated levels of *Leptospira* resulting from

the re-introduction of a new population of reservoir hosts. This is not an unlikely scenario as studies have shown that tropical regions have some of the greatest occurrences of highly endangered biodiversity and are frequently targeted for conservation efforts [238].

6. Leptospirosis in the Environment: Across the Urban-Rural Gradient

The prevalence, rate of transmission, and route of transmission of *Leptospira* is not uniform across the landscape. This is unsurprising as there are significant environmental differences across the urban-rural gradient, including to (but not limited to) differences in hydrology, soils, and animal life. In our review of the literature, discussion of environmental conditions associated with leptospirosis tended to revolve around discussion of urbanization; however, studies found equivalent rates of leptospirosis regardless of urbanization [1,54,132,239]. Ultimately, we found that leptospirosis is not a disease that can be adequately described through rate of urbanization. In fact, meta-analysis of decades of leptospirosis research shows that morbidity of the disease is not significantly associated with urbanization and is greater in rural and tropical regions [1]. In contrast, there is significant evidence of greater severity of leptospirosis and mortality associated with urbanization [1,61,239]. While we found no clear unifying trend that explains these patterns of leptospirosis morbidity or mortality across urban-rural gradients, clear trends emerged in many regions along the urban–rural gradient. As a result, we suspect that the trends along the urban–rural gradient in a given region are indicative of routes of transmission, contributors of environmental persistence, and the most likely risks of spillover from environmental sources. By this, we mean that leptospirosis is likely very regionally specific; thus, understanding the disease in a region will elucidate the dominant factors associated with the persistence and transmission of *Leptospira*. By studying such trends along the urban–rural gradient, opportunities for the management of the disease are likely to emerge.

6.1. Current Understanding

Leptospirosis is associated with urbanization in some regions. The association of urbanization and leptospirosis most likely results from environmental conditions that contribute to environmental persistence, increase the risk of spillover from environmental sources, and facilitate common routes of transmission. In one such study, the primary risk factors proved to be variation in rat populations and exposure to mud flows and flooding regardless of whether the rats or flooding occurred in an urban or rural environment [239]. In urban environments, high-density low-income housing is associated with the transmission of *Leptospira* as a result of the contamination of housing and soils by infected urine from rat infestations [3,128,130,131,240,241]. In regions with high populations of people and rats, there is a greater risk of leptospirosis infection among both animals and humans, as studies suggest a population-associated accumulation of disease load [68]. Essentially, leptospirosis rates should be greatest where there is a high population of infected individuals and a high population of susceptible individuals. This is one of the prevailing theories behind studies that suggest greater leptospirosis in urban regions [132].

Additionally, socioeconomically disadvantaged communities have disproportionate rates of leptospirosis [1,11,34,35,37,38]. In urban areas, socioeconomically disadvantaged communities often reside in high-density housing with conditions that facilitate the spread of leptospirosis by increasing the contact with environmental sources of the disease. These conditions include reduced rates of governmental sanitation interventions, trash accumulation, poor infrastructure, and contact with sewer water [8,35,38,52,239,240]. Low-income and socioeconomically disadvantaged communities receive reduced rates of governmental sanitation interventions such as trash pickup and pest control, which increases rodent populations that play a large role in urban leptospirosis [128–131,134–140]. The aging or impaired infrastructure commonly found in low-income urban communities results in increased contact with sewer water for residents. For example, many outdated urban stormwater drains connect with sewer systems. In urban areas, these outdated stormwater systems are most commonly found in historically marginalized and socioeconomically

disadvantaged communities [242]. High percentages of impervious surfaces in urban watersheds increase the urban-heat effect and the amount of storm runoff. This results in stormwater surges filling sewers and causing aboveground overflows of sewage and stormwater that contribute to the transmission of pathogens [243,244]. This situation promotes unsanitary, wet soils that are prone to frequent flooding. Consequently, many urban areas are characterized by high-risk conditions following large precipitation events. In addition, low-income housing is often constructed on wet soils in flood-prone regions along streams and other waterways [36]. Residing in flood-risk regions with wet soils has been significantly linked to risk of leptospirosis infection and outbreak during mass precipitation and flooding events [1,8,36,50,53,245,246]. Residents are also at greater risk outside of these mass precipitation and flooding events since they have greater contact with the wet soils that appear to be linked with long-term Leptospira persistence and survival. Additionally, as people are most likely to recreate in streams and waterways close to home, residents may be spending significant time in and around contaminated waterways. Residents may be unaware of their risk of Leptospira exposure and infection, as low-income and socioeconomically disadvantaged communities often do not receive equitable public health outreach [247]. In addition, low-income and socioeconomically disadvantaged communities have been shown to face disproportionate impacts from floods and other precipitation-based natural disasters due to reduced or delayed governmental assistance, lack of resources, and inequitable access to medical care [248–250].

The disproportionate impacts of leptospirosis on socioeconomically disadvantaged communities are not limited to urban contexts and extend to rural communities [1,11,34,35,37,38]. As we mentioned earlier in the review, there is significant evidence that leptospirosis cases occur most commonly in rural tropical regions [1]. Through our review, we found that the greater morbidity of leptospirosis in tropical rural communities seems to be associated with greater local prevalence of the pathogen and increased contact with potential environmental sources. Tropical rural regions likely have greater leptospirosis prevalence due to favorable environmental conditions that facilitate the greater environmental presence of other zoonotic pathogens. The warmer temperatures and increased precipitation rates found in the tropics are favorable for zoonotic pathogens [251]. These climatic conditions facilitate soil dynamics that are favorable for the survival of other enteric zoonotic pathogens such as E. coli [252]. The greater abundance and biodiversity of wildlife in tropical regions then facilitate greater environmental transmission of zoonotic pathogens [253]. Thus, there is greater potential for zoonotic pathogens such as Leptospira to complete their lifecycles through environmental transmission and wildlife in tropical regions, resulting in potentially greater prevalence in tropical rural regions. In addition, livestock and agricultural practices in tropical regions have increased human-wildlife-livestock interactions [254]. This may also facilitate greater rates of leptospirosis in tropical and rural regions. Additionally, agriculture and livestock production are generally located in rural communities. Thus, there are usually greater numbers of agricultural and livestock workers in rural communities and, thus, a greater risk of zoonotic disease [255]. This close contact with contaminated soils, waters, and animals likely facilitates greater local prevalence of leptospirosis in rural regions.

The leptospirosis risks associated with flooding and contaminated stormwaters are just as prevalent in rural communities as in urban [239]. In addition, inadequate sanitation interventions are also prevalent in socioeconomically disadvantaged rural communities and result in trash accumulation, contact with sewage, and other environmental exposures [239]. Similar to the impacts that outdated combined sewer overflows have on leptospirosis risk, inadequate and failing infrastructure increase the potential of leptospirosis exposure in rural communities, such as in the overflow of sewage lagoons and other livestock waste management solutions [256–260]. Similar to socioeconomically disadvantaged communities in urban settings, the impacts of leptospirosis on individuals have disproportionate impacts in rural areas. Lack of resources, quality medical care, and reduced or delayed governmental assistance can tremendously increase the burden and cycles of poverty [248–250,261].

6.2. Future Directions

As discussed, increased risk of leptospirosis along the urban–rural gradient often stems from a combination of factors associated with poverty, including lack of resources, poor infrastructure, and reduced governmental sanitation interventions. Additionally, increased risk of leptospirosis along the urban–rural gradient may also result from land management decisions that alter the hydrology and soil dynamics of waterways in tropical regions [262,263]. Alterations to hydrologic and biogeochemical conditions has been shown to determine virulence of disease, although this has yet to be studied in leptospirosis [264,265].

Since many of the increased risks associated with increased rates of leptospirosis are related to infrastructure and land management decisions, we suggest that by studying trends along the urban-rural gradient, opportunities for reducing the risk of leptospirosis outbreaks are likely to emerge alongside the increased understanding of where risk associated with environmental conditions is most severe. For example, prevalence within cattle herds increases along the urban-rural gradient as the proximity to the city increases [180]. The mechanism behind this trend is uncertain at this time, so further study is needed to understand the transmission cycles among domestic animals. There also appears to be a prevalence trend of leptospirosis among small mammal populations that occurs along the urban-rural gradient with different prevalence of infection between urban, suburban, and rural ecosystems [245]. The mechanisms behind this are also unknown. This highlights the lack of understanding on the cycles of Leptospira transmission in wildlife. Without a clear understanding of the routes of transmission, contributors of environmental persistence, and the most likely risks of spillover from environmental sources, efforts to control the disease are unlikely to be as effective as they could be. Due to this, it is imperative to study the dynamics of the disease across the urban-rural gradient alongside studies into the environmental persistence, survival, and reproduction of *Leptospira*. Targeted public health outreach to improve communities' ability to participate in reducing disproportionate exposure to leptospirosis requires that we discover more about the environmental persistence and survival of Leptospira [266].

7. Conclusions

In this review, we discussed the current understanding of leptospirosis in the environment and highlighted future directions of study that we suggest will greatly assist in disease surveillance and management. Although the need for expanded active surveillance, treatment, and epidemiological transmission studies is not new, increasing pressures on natural systems are accelerating the frequency and severity of zoonotic disease outbreaks [34]. As climate change increases the variability, frequency, and severity of precipitation and storm events, the conditions required for outbreaks of zoonotic diseases such as leptospirosis will occur more frequently and in greater severity [267]. Urbanization concentrates human populations in high-density environments where outbreaks of disease are more common and have greater transmission rates, while the spread of urbanization globally places more and more people in increased contact with wildlife and zoonotic pathogens [268]. As these pressures increase the occurrence of *Leptospira* transmission, the impacts of this disease will disproportionately impact people of lower socioeconomic status without access to timely, quality healthcare, and will further exacerbate systemic cycles of poverty in these communities [3,4,34]. In addition, water resources are under more and more demand for use for consumption and recreation; as this demand grows, the risk of leptospirosis infection and outbreak will increase as well [16,31]. As a result, it is imperative that further study into the prevalence and transmission of *Leptospira* in the environment is conducted, with specific attention being paid to objectives that improve disease surveillance and management efforts.

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References

- Costa, F.; Hagan, J.E.; Calcagno, J.; Kane, M.; Torgerson, P.; Martinez-Silveira, M.S.; Stein, C.; Abela-Ridder, B.; Ko, A.I. Global Morbidity and Mortality of Leptospirosis: A Systematic Review. *PLoS Negl. Trop. Dis.* 2015, *9*, e0003898. [CrossRef] [PubMed]
 Example to K.W. Calcagne, L. La transformation of the second statement of the s
- Evangelista, K.V.; Coburn, J. Leptospira as an Emerging Pathogen: A Review of Its Biology, Pathogenesis and Host Immune Responses. *Future Microbiol.* 2010, 5, 1413–1425. [CrossRef] [PubMed]
- 3. Karpagam, K.B.; Ganesh, B. Leptospirosis: A Neglected Tropical Zoonotic Infection of Public Health Importance—An Updated Review. *Eur. J. Clin. Microbiol. Infect. Dis.* **2020**, *39*, 835–846. [CrossRef] [PubMed]
- 4. Haake, D.A.; Levett, P.N. Leptospirosis in Humans. Curr. Top. Microbiol. Immunol. 2015, 387, 65–97. [CrossRef] [PubMed]
- Faisal, S.M.; McDonough, S.P.; Chang, Y.-F. Leptospira: Invasion, Pathogenesis and Persistence. In *The Pathogenic Spirochetes:* Strategies for Evasion of Host Immunity and Persistence; Embers, M.E., Ed.; Springer US: Boston, MA, USA, 2012; pp. 143–172, ISBN 978-1-4614-5404-5.
- Izurieta, R.; Galwankar, S.; Clem, A. Leptospirosis: The "Mysterious" Mimic. J. Emerg. Trauma Shock 2008, 1, 21–33. [CrossRef] [PubMed]
- Traxler, R.M.; Callinan, L.S.; Holman, R.C.; Steiner, C.; Guerra, M.A. Leptospirosis-Associated Hospitalizations, United States, 1998–2009. Emerg. Infect. Dis. 2014, 20, 1273–1279. [CrossRef]
- 8. Bierque, E.; Thibeaux, R.; Girault, D.; Soupé-Gilbert, M.-E.; Goarant, C. A Systematic Review of Leptospira in Water and Soil Environments. *PLoS ONE* **2020**, *15*, e0227055. [CrossRef]
- 9. López-Robles, G.; Córdova-Robles, F.N.; Sandoval-Petris, E.; Montalvo-Corral, M. Leptospirosis at Human-Animal-Environment Interfaces in Latin-America: Drivers, Prevention, and Control Measures. *Biotecnia* **2021**, *23*, 89–100.
- Rahman, M.H.A.A.; Hairon, S.M.; Hamat, R.A.; Jamaluddin, T.Z.M.T.; Shafei, M.N.; Idris, N.; Osman, M.; Sukeri, S.; Wahab, Z.A.; Mohammad, W.M.Z.W.; et al. Leptospirosis Health Intervention Module Effect on Knowledge, Attitude, Belief, and Practice among Wet Market Workers in Northeastern Malaysia: An Intervention Study. *Int. J. Environ. Res. Public Health* 2018, 15, 1396. [CrossRef]
- 11. Sykes, J.E.; Haake, D.A.; Gamage, C.D.; Mills, W.Z.; Nally, J.E. A Global One Health Perspective on Leptospirosis in Humans and Animals. J. Am. Vet. Med. Assoc. 2022, 1, 1589–1596. [CrossRef]
- Hartskeerl, R.A.; Collares-Pereira, M.; Ellis, W.A. Emergence, Control and Re-Emerging Leptospirosis: Dynamics of Infection in the Changing World. *Clin. Microbiol. Infect.* 2011, 17, 494–501. [CrossRef] [PubMed]
- 13. Biscornet, L.; de Comarmond, J.; Bibi, J.; Mavingui, P.; Dellagi, K.; Tortosa, P.; Pagès, F. An Observational Study of Human Leptospirosis in Seychelles. *Am. J. Trop. Med. Hyg.* **2020**, *103*, 999–1008. [CrossRef] [PubMed]
- 14. Narita, M.; Fujitani, S.; Haake, D.A.; Paterson, D.L. Leptospirosis After Recreational Exposure to Water in the Yaeyama Islands, Japan. *Am. J. Trop. Med. Hyg.* **2005**, *73*, 652–656. [CrossRef] [PubMed]
- 15. Mwachui, M.A.; Crump, L.; Hartskeerl, R.; Zinsstag, J.; Hattendorf, J. Environmental and Behavioural Determinants of Leptospirosis Transmission: A Systematic Review. *PLoS Negl. Trop. Dis.* **2015**, *9*, e0003843. [CrossRef]
- 16. Pappas, G.; Papadimitriou, P.; Siozopoulou, V.; Christou, L.; Akritidis, N. The Globalization of Leptospirosis: Worldwide Incidence Trends. *Int. J. Infect. Dis.* **2008**, *12*, 351–357. [CrossRef]
- Abdullah, N.M.; Mohammad, W.M.Z.W.; Shafei, M.N.; Sukeri, S.; Idris, Z.; Arifin, W.N.; Nozmi, N.; Saudi, S.N.S.; Samsudin, S.; Zainudin, A.-W.; et al. Leptospirosis and Its Prevention: Knowledge, Attitude and Practice of Urban Community in Selangor, Malaysia. *BMC Public Health* 2019, 19, 628. [CrossRef]
- 18. Gómez-Martín, M.; Lozano, C.; Luque, R.; Luque-Romero, L.; Rodríguez-Benjumeda, L.; Aznar-Martin, J. Red Swamp Crayfish Collecting: A Risk Activity for Leptospirosis. *Clin. Microbiol. Infect.* **2020**, *26*, 1103–1104. [CrossRef]
- 19. Waitkins, S.A. Leptospirosis in Man, British Isles: 1984. Br. Med. J. (Clin. Res. Ed.) 1986, 292, 1324. [CrossRef]

- 20. Wilkins, E.; Cope, A.; Waitkins, S. Rapids, Rafts, and Rats. Lancet 1988, 332, 283–284. [CrossRef]
- Guillois, Y.; Bourhy, P.; Ayral, F.; Pivette, M.; Decors, A.; Aranda Grau, J.H.; Champenois, B.; Malhère, C.; Combes, B.; Richomme, C.; et al. An Outbreak of Leptospirosis among Kayakers in Brittany, North-West France, 2016. *Euro Surveill.* 2018, 23, 1700848. [CrossRef]
- 22. Lau, C.; Smythe, L.; Weinstein, P. Leptospirosis: An Emerging Disease in Travellers. *Travel Med. Infect. Dis.* **2010**, *8*, 33–39. [CrossRef] [PubMed]
- 23. Shaw, R.D. Kayaking as a Risk Factor for Leptospirosis. Mo. Med. 1992, 89, 354–357. [PubMed]
- Agampodi, S.B.; Karunarathna, D.; Jayathilala, N.; Rathnayaka, H.; Agampodi, T.C.; Karunanayaka, L. Outbreak of Leptospirosis after White-Water Rafting: Sign of a Shift from Rural to Recreational Leptospirosis in Sri Lanka? *Epidemiol. Infect.* 2014, 142, 843–846. [CrossRef] [PubMed]
- Jevon, T.R.; Knudson, M.P.; Smith, P.A.; Whitecar, P.S.; Blake, R.L. A Point-Source Epidemic of Leptospirosis. *Postgrad. Med.* 1986, 80, 121–129. [CrossRef] [PubMed]
- 26. Anderson, D.C.; Folland, D.S.; Fox, M.D.; Patton, C.M.; Kaufmann, A.F. Leptospirosis: A Common-Source Outbreak Due to Leptospires of the Grippotyphosa Serogroup. *Am. J. Epidemiol.* **1978**, *107*, 538–544. [CrossRef]
- 27. Nelson, K.E.; Ager, E.A.; Galton, M.M.; Gillespie, R.W.H.; Sulzer, C.R. An Outbreak of Leptospirosis in Washington State. *Am. J. Epidemiol.* **1973**, *98*, 336–347. [CrossRef]
- Morgan, J.; Bornstein, S.L.; Karpati, A.M.; Bruce, M.; Bolin, C.A.; Austin, C.C.; Woods, C.W.; Lingappa, J.; Langkop, C.; Davis, B.; et al. Outbreak of Leptospirosis among Triathlon Participants and Community Residents in Springfield, Illinois, 1998. *Clin. Infect. Dis.* 2002, 34, 1593–1599. [CrossRef]
- Brockmann, S.; Piechotowski, I.; Bock-Hensley, O.; Winter, C.; Oehme, R.; Zimmermann, S.; Hartelt, K.; Luge, E.; Nöckler, K.; Schneider, T.; et al. Outbreak of Leptospirosis among Triathlon Participants in Germany, 2006. BMC Infect. Dis. 2010, 10, 91. [CrossRef]
- Sejvar, J.; Bancroft, E.; Winthrop, K.; Bettinger, J.; Bajani, M.; Bragg, S.; Shutt, K.; Kaiser, R.; Marano, N.; Popovic, T.; et al. Leptospirosis in "Eco-Challenge" Athletes, Malaysian Borneo, 2000. *Emerg. Infect. Dis.* 2003, *9*, 702–707. [CrossRef]
- 31. Walker, M.D. Leptospirosis: The Possible Risk to Those Participating in Water-Based Sports and Activities. *Br. J. Gen. Pract.* 2018, 68, 394–395. [CrossRef]
- 32. Haake, D.A.; Dundoo, M.; Cader, R.; Kubak, B.M.; Hartskeerl, R.A.; Sejvar, J.J.; Ashford, D.A. Leptospirosis, Water Sports, and Chemoprophylaxis. *Clin. Infect. Dis.* **2002**, *34*, e40–e43. [CrossRef] [PubMed]
- Pagès, F.; Larrieu, S.; Simoes, J.; Lenabat, P.; Kurtkowiak, B.; Guernier, V.; Minter, G.L.; Lagadec, E.; Gomard, Y.; Michault, A.; et al. Investigation of a Leptospirosis Outbreak in Triathlon Participants, Réunion Island, 2013. *Epidemiol. Infect.* 2016, 144, 661–669. [CrossRef]
- Hotez, P.J. Neglected Infections of Poverty in the United States of America. PLoS Negl. Trop. Dis. 2008, 2, e256. [CrossRef] [PubMed]
- Bacallao, J.; Schneider, M.C.; Najera, P.; Aldighieri, S.; Soto, A.; Marquiño, W.; Sáenz, C.; Jiménez, E.; Moreno, G.; Chávez, O.; et al. Socioeconomic Factors and Vulnerability to Outbreaks of Leptospirosis in Nicaragua. *Int. J. Environ. Res. Public Health* 2014, 11, 8301–8318. [CrossRef]
- Reis, R.B.; Ribeiro, G.S.; Felzemburgh, R.D.M.; Santana, F.S.; Mohr, S.; Melendez, A.X.T.O.; Queiroz, A.; Santos, A.C.; Ravines, R.R.; Tassinari, W.S.; et al. Impact of Environment and Social Gradient on Leptospira Infection in Urban Slums. *PLoS Negl. Trop. Dis.* 2008, 2, e228. [CrossRef]
- 37. Zhao, J.; Liao, J.; Huang, X.; Zhao, J.; Wang, Y.; Ren, J.; Wang, X.; Ding, F. Mapping Risk of Leptospirosis in China Using Environmental and Socioeconomic Data. *BMC Infect. Dis.* **2016**, *16*, 343. [CrossRef]
- Khalil, H.; Santana, R.; de Oliveira, D.; Palma, F.; Lustosa, R.; Eyre, M.T.; Carvalho-Pereira, T.; Reis, M.G.; Ko, A.I.; Diggle, P.J.; et al. Poverty, Sanitation, and Leptospira Transmission Pathways in Residents from Four Brazilian Slums. *PLoS Negl. Trop. Dis.* 2021, 15, e0009256. [CrossRef]
- 39. Sun, N.; Amon, J.J. Addressing Inequity. Health Hum. Rights 2018, 20, 11–25. [PubMed]
- 40. Hotez, P.J. Global Urbanization and the Neglected Tropical Diseases. PLoS Negl. Trop. Dis. 2017, 11, e0005308. [CrossRef]
- 41. UN Population Division. World Urbanization Prospects: The 2018 Revisions, 2nd ed.; Population Studies; United Nations: New York, NY, USA, 2019; ISBN 978-92-1-148319-2.
- 42. World Health Organization. Report of the Second Meeting of the Leptospirosis Burden Epidemiology Reference Group; World Health Organization: Geneva, Switzerland, 2011.
- Flannery, B.; Pereira, M.M.; Velloso, L.D.F.; Carvalho, C.D.C.; Codes, L.G.D.; Orrico, G.D.S.; Dourado, C.M.; Riley, L.W.; Reis, M.G.; Ko, A.I. Referral Pattern of Leptospirosis Cases during a Large Urban Epidemic of Dengue. *Am. J. Trop. Med. Hyg.* 2001, 65, 657–663. [CrossRef]
- Reller, M.E.; Wunder, E.A.; Miles, J.J.; Flom, J.E.; Mayorga, O.; Woods, C.W.; Ko, A.I.; Dumler, J.S.; Matute, A.J. Unsuspected Leptospirosis Is a Cause of Acute Febrile Illness in Nicaragua. *PLoS Negl. Trop. Dis.* 2014, *8*, e2941. [CrossRef]
- Ismail, T.F.; Wasfy, M.O.; Abdul-Rahman, B.; Murray, C.K.; Hospenthal, D.R.; Abdel-Fadeel, M.; Abdel-Maksoud, M.; Samir, A.; Hatem, M.E.; Klena, J.; et al. Retrospective Serosurvey of Leptospirosis among Patients with Acute Febrile Illness and Hepatitis in Egypt. Am. J. Trop. Med. Hyg. 2006, 75, 1085–1089. [CrossRef]

- 46. Turmel, J.-M.; Olive, C.; Bigeard, B.; Abel, S.; Banydeen, R.; Daoud, L.; Fayolle, P.-M.; Cabié, A. Case Report: Pulmonary Leptospirosis Misdiagnosed as COVID-19. *Am. J. Trop. Med. Hyg.* **2022**, *107*, 97–99. [CrossRef] [PubMed]
- 47. Musso, D.; La Scola, B. Laboratory Diagnosis of Leptospirosis: A Challenge. J. Microbiol. Immunol. Infect. 2013, 46, 245–252. [CrossRef] [PubMed]
- Abela-Ridder, B.; Sikkema, R.; Hartskeerl, R.A. Estimating the Burden of Human Leptospirosis. Int. J. Antimicrob. Agents 2010, 36 (Suppl. S1), S5–S7. [CrossRef] [PubMed]
- 49. Md-Lasim, A.; Mohd-Taib, F.S.; Abdul-Halim, M.; Mohd-Ngesom, A.M.; Nathan, S.; Md-Nor, S. Leptospirosis and Coinfection: Should We Be Concerned? *Int. J. Environ. Res. Public Health* **2021**, *18*, 9411. [CrossRef]
- Ko, A.I.; Reis, M.G.; Dourado, C.M.R.; Johnson, W.D., Jr.; Riley, L.W. Urban Epidemic of Severe Leptospirosis in Brazil. *Lancet* 1999, 354, 820–825. [CrossRef]
- Maskey, M.; Shastri, J.; Saraswathi, K.; Surpam, R.; Vaidya, N. Leptospirosis in Mumbai: Post-Deluge Outbreak 2005. *Indian J.* Med. Microbiol. 2006, 24, 337–338. [CrossRef]
- Hacker, K.P.; Sacramento, G.A.; Cruz, J.S.; de Oliveira, D.; Nery, N.; Lindow, J.C.; Carvalho, M.; Hagan, J.; Diggle, P.J.; Begon, M.; et al. Influence of Rainfall on Leptospira Infection and Disease in a Tropical Urban Setting, Brazil. *Emerg. Infect. Dis.* 2020, 26, 311. [CrossRef]
- Casanovas-Massana, A.; Costa, F.; Riediger, I.N.; Cunha, M.; de Oliveira, D.; Mota, D.C.; Sousa, E.; Querino, V.A.; Nery, N., Jr.; Reis, M.G.; et al. Spatial and Temporal Dynamics of Pathogenic Leptospira in Surface Waters from the Urban Slum Environment. Water Res. 2018, 130, 176–184. [CrossRef]
- 54. Barragan, V.; Olivas, S.; Keim, P.; Pearson, T. Critical Knowledge Gaps in Our Understanding of Environmental Cycling and Transmission of *Leptospira* spp. *Appl. Environ. Microbiol.* **2017**, *83*, e01190-17. [CrossRef] [PubMed]
- Thibeaux, R.; Geroult, S.; Benezech, C.; Chabaud, S.; Soupé-Gilbert, M.-E.; Girault, D.; Bierque, E.; Goarant, C. Seeking the Environmental Source of Leptospirosis Reveals Durable Bacterial Viability in River Soils. *PLoS Negl. Trop. Dis.* 2017, *11*, e0005414. [CrossRef] [PubMed]
- Saito, M.; Villanueva, S.Y.A.M.; Chakraborty, A.; Miyahara, S.; Segawa, T.; Asoh, T.; Ozuru, R.; Gloriani, N.G.; Yanagihara, Y.; Yoshida, S. Comparative Analysis of Leptospira Strains Isolated from Environmental Soil and Water in the Philippines and Japan. *Appl. Environ. Microbiol.* 2013, 79, 601–609. [CrossRef]
- Yanagihara, Y.; Villanueva, S.Y.A.M.; Nomura, N.; Ohno, M.; Sekiya, T.; Handabile, C.; Shingai, M.; Higashi, H.; Yoshida, S.; Masuzawa, T.; et al. Leptospira Is an Environmental Bacterium That Grows in Waterlogged Soil. *Microbiol. Spectr.* 2022, 10, e02157-21. [CrossRef] [PubMed]
- 58. Wynwood, S.J.; Graham, G.C.; Weier, S.L.; Collet, T.A.; McKay, D.B.; Craig, S.B. Leptospirosis from Water Sources. *Pathog. Glob. Health* **2014**, *108*, 334–338. [CrossRef] [PubMed]
- 59. Wasiński, B.; Dutkiewicz, J. Leptospirosis—Current Risk Factors Connected with Human Activity and the Environment. *Ann. Agric. Environ. Med.* **2013**, *20*, *6*.
- 60. Beigel, B.; Verma, A. Leptospira: Molecular Detection of Pathogenic Species in Natural Sources. *Curr. Protoc. Microbiol.* **2017**, 47, 12E.6.1–12E.6.8. [CrossRef]
- Ganoza, C.A.; Matthias, M.A.; Collins-Richards, D.; Brouwer, K.C.; Cunningham, C.B.; Segura, E.R.; Gilman, R.H.; Gotuzzo, E.; Vinetz, J.M. Determining Risk for Severe Leptospirosis by Molecular Analysis of Environmental Surface Waters for Pathogenic Leptospira. *PLoS Med.* 2006, *3*, e308. [CrossRef]
- Thibeaux, R.; Iraola, G.; Ferrés, I.; Bierque, E.; Girault, D.; Soupé-Gilbert, M.-E.; Picardeau, M.; Goarant, C. Deciphering the Unexplored Leptospira Diversity from Soils Uncovers Genomic Evolution to Virulence. *Microb. Genom.* 2018, 4, e000144. [CrossRef]
- 63. Riediger, I.N.; Hoffmaster, A.R.; Casanovas-Massana, A.; Biondo, A.W.; Ko, A.I.; Stoddard, R.A. An Optimized Method for Quantification of Pathogenic Leptospira in Environmental Water Samples. *PLoS ONE* **2016**, *11*, e0160523. [CrossRef]
- Casanovas-Massana, A.; Neves Souza, F.; Curry, M.; de Oliveira, D.; de Oliveira, A.S.; Eyre, M.T.; Santiago, D.; Aguiar Santos, M.; Serra, R.M.R.; Lopes, E.; et al. Effect of Sewerage on the Contamination of Soil with Pathogenic Leptospira in Urban Slums. *Environ. Sci. Technol.* 2021, 55, 15882–15890. [CrossRef] [PubMed]
- 65. Narkkul, U.; Thaipadungpanit, J.; Srisawat, N.; Rudge, J.W.; Thongdee, M.; Pawarana, R.; Pan-ngum, W. Human, Animal, Water Source Interactions and Leptospirosis in Thailand. *Sci. Rep.* **2021**, *11*, 3215. [CrossRef]
- Houéménou, H.; Gauthier, P.; Houéménou, G.; Mama, D.; Alassane, A.; Socohou, A.; Dossou, H.-J.; Badou, S.; Picardeau, M.; Tweed, S.; et al. Pathogenic Leptospira and Water Quality in African Cities: A Case Study of Cotonou, Benin. *Sci. Total Environ.* 2021, 774, 145541. [CrossRef] [PubMed]
- 67. Narkkul, U.; Thaipadungpanit, J.; Srilohasin, P.; Singkhaimuk, P.; Thongdee, M.; Chaiwattanarungruengpaisan, S.; Krairojananan, P.; Pan-ngum, W. Optimization of Culture Protocols to Isolate *Leptospira* spp. from Environmental Water, Field Investigation, and Identification of Factors Associated with the Presence of *Leptospira* spp. in the Environment. *Trop. Med. Infect. Dis.* 2020, *5*, 94. [CrossRef]
- Pedra, G.G. Predicting Environmental Risk of Transmission of Leptospirosis. Ph.D. Thesis, University of Liverpool, Liverpool, UK, 2019.
- 69. Andre-Fontaine, G.; Aviat, F.; Thorin, C. Waterborne Leptospirosis: Survival and Preservation of the Virulence of Pathogenic *Leptospira* spp. in Fresh Water. *Curr. Microbiol.* **2015**, *71*, 136–142. [CrossRef]

- 70. DIESCH, S.L.; McCulloch, W.F.; Braun, J.L.; Crawford, R.P., Jr. Environmental Studies on the Survival of Leptospires in a Farm Creek Following a Human Leptospirosis Outbreak in Iowa. *Bull. Wildl. Dis. Assoc.* **1969**, *5*, 166–173. [CrossRef] [PubMed]
- Parker, J.; Walker, M. Survival of a Pathogenic Leptospira Serovar in Response to Combined in Vitro PH and Temperature Stresses. *Vet. Microbiol.* 2011, 152, 146–150. [CrossRef]
- 72. Smith, C.E.; Turner, L.H. The Effect of PH on the Survival of Leptospires in Water. Bull. World Health Organ. 1961, 24, 35–43.
- 73. de Oliveira, D.; Airam Querino, V.; Sara Lee, Y.; Cunha, M.; Nery, N., Jr.; Wessels Perelo, L.; Rossi Alva, J.C.; Ko, A.I.; Reis, M.G.; Casanovas-Massana, A.; et al. Relationship between Physicochemical Characteristics and Pathogenic Leptospira in Urban Slum Waters. *Trop. Med. Infect. Dis.* 2020, *5*, 146. [CrossRef]
- Wongbutdee, J.; Jittimanee, J. The Viability of Leptospira Is Related to Physicochemical Properties of the Surface Water Surrounding an Agricultural Area and HemO and LipL32 Gene Expression in Response to Iron in Water. *Trans. R. Soc. Trop. Med. Hyg.* 2022, 116, 609–621. [CrossRef]
- 75. Bierque, E.; Soupé-Gilbert, M.-E.; Thibeaux, R.; Girault, D.; Guentas, L.; Goarant, C. Leptospira Interrogans Retains Direct Virulence After Long Starvation in Water. *Curr. Microbiol.* **2020**, *77*, 3035–3043. [CrossRef] [PubMed]
- 76. Ryu, E.; Liu, C.-K. The Viability of Leptospires in the Summer Paddy Water. Jpn. J. Microbiol. 1966, 10, 51–57. [CrossRef] [PubMed]
- Kaboosi, H.; Razavi, M.R. Efficiency of Filtration Technique for Isolation of Leptospires from Surface Waters: Role of Different Membranes with Different Pore Size and Materials. *Afr. J. Microbiol. Res.* 2010, *4*, 671–676.
- Gorman, M.; Xu, R.; Prakoso, D.; Salvador, L.C.M.; Rajeev, S. Leptospira Enrichment Culture Followed by ONT Metagenomic Sequencing Allows Better Detection of Leptospira Presence and Diversity in Water and Soil Samples. *PLoS Negl. Trop. Dis.* 2022, 16, e0010589. [CrossRef]
- Thibeaux, R.; Girault, D.; Bierque, E.; Soupé-Gilbert, M.-E.; Rettinger, A.; Douyère, A.; Meyer, M.; Iraola, G.; Picardeau, M.; Goarant, C. Biodiversity of Environmental Leptospira: Improving Identification and Revisiting the Diagnosis. *Front. Microbiol.* 2018, 9, 816. [CrossRef]
- Schneider, A.G.; Casanovas-Massana, A.; Hacker, K.P.; Wunder, E.A., Jr.; Begon, M.; Reis, M.G.; Childs, J.E.; Costa, F.; Lindow, J.C.; Ko, A.I. Quantification of Pathogenic Leptospira in the Soils of a Brazilian Urban Slum. *PLoS Negl. Trop. Dis.* 2018, 12, e0006415. [CrossRef]
- Casanovas-Massana, A.; Hamond, C.; Santos, L.A.; de Oliveira, D.; Hacker, K.P.; Balassiano, I.; Costa, F.; Medeiros, M.A.; Reis, M.G.; Ko, A.I.; et al. Leptospira Yasudae Sp. Nov. and Leptospira Stimsonii Sp. Nov., Two New Species of the Pathogenic Group Isolated from Environmental Sources. *Int. J. Syst. Evol. Microbiol.* 2020, 70, 1450–1456. [CrossRef]
- 82. Richard, E.; Bourhy, P.; Picardeau, M.; Moulin, L.; Wurtzer, S. Effect of Disinfection Agents and Quantification of Potentially Viable Leptospira in Fresh Water Samples Using a Highly Sensitive Integrity-QPCR Assay. *PLoS ONE* **2021**, *16*, e0251901. [CrossRef]
- 83. Viau, E.J.; Boehm, A.B. Quantitative PCR-Based Detection of Pathogenic Leptospira in Hawai'ian Coastal Streams. *J. Water Health* 2011, *9*, 637–646. [CrossRef]
- Cantwell, R.E.; Hofmann, R. Inactivation of Indigenous Coliform Bacteria in Unfiltered Surface Water by Ultraviolet Light. Water Res. 2008, 42, 2729–2735. [CrossRef]
- 85. Walters, E.; Graml, M.; Behle, C.; Müller, E.; Horn, H. Influence of Particle Association and Suspended Solids on UV Inactivation of Fecal Indicator Bacteria in an Urban River. *Water Air Soil Pollut.* **2013**, 225, 1822. [CrossRef]
- 86. Bejo, S.; Bahaman, A.; Saad, M.; Mutalib, A. The Survival of Leptospira Interrogans Serovar Hardjo in the Malaysian Environment. *J. Anim. Vet. Adv.* **2004**, *3*, 123–129.
- 87. Kadis, S.; Pugh, W.L. Urea Utilization by Leptospira. Infect. Immun. 1974, 10, 793–801. [CrossRef]
- Leonard, F.; Quinn, P.J.; Ellis, W.A. Possible Effect of PH on the Survival of Leptospires in Cattle Urine. Vet. Rec. 1992, 131, 53–54.
 [CrossRef]
- Trueba, G.; Zapata Mena, S.; Madrid, K.; Cullen, P.; Haake, D. Cell Aggregation: A Mechanism of Pathogenic Leptospira to Survive in Fresh Water. *Int. Microbiol. Off. J. Span. Soc. Microbiol.* 2004, 7, 35–40.
- Barragan, V.A.; Mejia, M.E.; Trávez, A.; Zapata, S.; Hartskeerl, R.A.; Haake, D.A.; Trueba, G.A. Interactions of Leptospira with Environmental Bacteria from Surface Water. *Curr. Microbiol.* 2011, 62, 1802–1806. [CrossRef]
- 91. Thibeaux, R.; Soupé-Gilbert, M.-E.; Kainiu, M.; Girault, D.; Bierque, E.; Fernandes, J.; Bähre, H.; Douyère, A.; Eskenazi, N.; Vinh, J.; et al. The Zoonotic Pathogen Leptospira Interrogans Mitigates Environmental Stress through Cyclic-Di-GMP-Controlled Biofilm Production. *NPJ Biofilms Microbiomes* **2020**, *6*, 24. [CrossRef]
- 92. Vinod Kumar, K.; Lall, C.; Vimal Raj, R.; Vedhagiri, K.; Vijayachari, P. Molecular Detection of Pathogenic Leptospiral Protein Encoding Gene (LipL32) in Environmental Aquatic Biofilms. *Lett. Appl. Microbiol.* **2016**, *62*, 311–315. [CrossRef]
- Meganathan, Y.; Vishwakarma, A.; Mohandass, R. Biofilm Formation and Social Interaction of Leptospira in Natural and Artificial Environments. *Res. Microbiol.* 2022, 173, 103981. [CrossRef]
- 94. Mandakhalikar, K.D.; Rahmat, J.N.; Chiong, E.; Neoh, K.G.; Shen, L.; Tambyah, P.A. Extraction and Quantification of Biofilm Bacteria: Method Optimized for Urinary Catheters. *Sci. Rep.* **2018**, *8*, 8069. [CrossRef]
- Casanovas-Massana, A.; Pedra, G.G.; Wunder, E.A.; Diggle, P.J.; Begon, M.; Ko, A.I. Quantification of Leptospira Interrogans Survival in Soil and Water Microcosms. *Appl. Environ. Microbiol.* 2018, 84, e00507-18. [CrossRef] [PubMed]
- 96. Vega-Corredor, M.C.; Opadeyi, J. Hydrology and Public Health: Linking Human Leptospirosis and Local Hydrological Dynamics in Trinidad, West Indies. *Earth Perspect.* **2014**, *1*, 3. [CrossRef]

- Ferreira, M.C.; Ferreira, M.F.M. Influence of Topographic and Hydrographic Factors on the Spatial Distribution of Leptospirosis Disease in São Paulo County, Brazil: An Approach Using Geospatial Techniques and GIS Analysis. *ISPRS-Int. Arch. Photogramm. Remote Sens. Spat. Inf. Sci.* 2016, 41B8, 197–201. [CrossRef]
- Convertino, M.; Reddy, A.; Liu, Y.; Munoz-Zanzi, C. Eco-Epidemiological Scaling of Leptospirosis: Vulnerability Mapping and Early Warning Forecasts. *Sci. Total Environ.* 2021, 799, 149102. [CrossRef] [PubMed]
- Ahangarcani, M.; Farnaghi, M.; Shirzadi, M.R.; Pilesjö, P.; Mansourian, A. Predictive Risk Mapping of Human Leptospirosis Using Support Vector Machine Classification and Multilayer Perceptron Neural Network. *Geospat. Health* 2019, 14. [CrossRef] [PubMed]
- Ercides Péres, W.; Russo, A.; Nunes, B. The Association between Hydro-Meteorological Events and Leptospirosis Hospitalizations in Santa Catarina, Brazil. Water 2019, 11, 1052. [CrossRef]
- 101. Flores, B.; Escobar, K.; Muzquiz, J.L.; Sheleby-Elías, J.; Mora, B.; Roque, E.; Torres, D.; Chávez, Á.; Jirón, W. Detection of Pathogenic Leptospires in Water and Soil in Areas Endemic to Leptospirosis in Nicaragua. *Trop. Med. Infect. Dis.* **2020**, *5*, 149. [CrossRef]
- 102. Jara, M.; Escobar, L.E.; Rodriges, R.O.; Frias-De-Diego, A.; Sanhueza, J.; Machado, G. Spatial Distribution and Spread Potential of Sixteen Leptospira Serovars in a Subtropical Region of Brazil. *Transbound. Emerg. Dis.* **2019**, *66*, 2482–2495. [CrossRef]
- Cucchi, K.; Liu, R.; Collender, P.A.; Cheng, Q.; Li, C.; Hoover, C.M.; Chang, H.H.; Liang, S.; Yang, C.; Remais, J.V. Hydroclimatic Drivers of Highly Seasonal Leptospirosis Incidence Suggest Prominent Soil Reservoir of Pathogenic *Leptospira* spp. in Rural Western China. *PLoS Negl. Trop. Dis.* 2019, 13, e0007968. [CrossRef]
- 104. Lall, C.; Vinod Kumar, K.; Raj, R.V.; Vedhagiri, K.; Sunish, I.P.; Vijayachari, P. Correlation Between Physicochemical Properties of Soil and Presence of Leptospira. *EcoHealth* **2018**, *15*, 670–675. [CrossRef]
- 105. Rood, E.J.J.; Goris, M.G.A.; Pijnacker, R.; Bakker, M.I.; Hartskeerl, R.A. Environmental Risk of Leptospirosis Infections in the Netherlands: Spatial Modelling of Environmental Risk Factors of Leptospirosis in the Netherlands. *PLoS ONE* 2017, 12, e0186987. [CrossRef] [PubMed]
- 106. Stone, N.E.; Hall, C.M.; Ortiz, M.; Hutton, S.M.; Santana-Propper, E.; Celona, K.R.; Williamson, C.H.D.; Bratsch, N.; Fernandes, L.G.V.; Busch, J.D.; et al. Diverse Lineages of Pathogenic Leptospira Species Are Widespread in the Environment in Puerto Rico, USA. PLoS Negl. Trop. Dis. 2022, 16, e0009959. [CrossRef] [PubMed]
- 107. Miller, E.; Barragan, V.; Chiriboga, J.; Weddell, C.; Luna, L.; Jiménez, D.J.; Aleman, J.; Mihaljevic, J.R.; Olivas, S.; Marks, J.; et al. Leptospira in River and Soil in a Highly Endemic Area of Ecuador. *BMC Microbiol.* **2021**, *21*, 17. [CrossRef] [PubMed]
- Masuzawa, T.; Sakakibara, K.; Saito, M.; Hidaka, Y.; Villanueva, S.Y.A.M.; Yanagihara, Y.; Yoshida, S. Characterization of Leptospira Species Isolated from Soil Collected in Japan. *Microbiol. Immunol.* 2018, 62, 55–59. [CrossRef] [PubMed]
- Wojcik-Fatla, A.; Zajac, V.; Wasinski, B.; Sroka, J.; Cisak, E.; Sawczyn, A.; Dutkiewicz, J. Occurrence of Leptospira DNA in Water and Soil Samples Collected in Eastern Poland. Ann. Agric. Environ. Med. 2014, 21, 730–732. [CrossRef]
- Verma, A.; Brandt, L.; Runser, S.; Gruszynski, K.; Gallatin, K.; Morgan, J.; Barnhart, H.; Duke, C.; Brovarney, S.; Geer, A.; et al. Detection of Pathogenic *Leptospira* spp. in Herpetofauna in Central Appalachia. *Zoonoses Public Health* 2022, 69, 325–332. [CrossRef]
- 111. Leptospirosis Risk in Outdoor Activities | Leptospirosis | CDC. Available online: https://www.cdc.gov/leptospirosis/features/ outdoor-activities.html (accessed on 20 September 2022).
- 112. Erickson, M.C.; Habteselassie, M.Y.; Liao, J.; Webb, C.C.; Mantripragada, V.; Davey, L.E.; Doyle, M.P. Examination of Factors for Use as Potential Predictors of Human Enteric Pathogen Survival in Soil. *J. Appl. Microbiol.* **2014**, *116*, 335–349. [CrossRef]
- 113. Mohammad, A. Assessing Changes in Soil Microbial Population with Some Soil Physical and Chemical Properties. *Int. J. Plant Anim. Environ. Sci.* 2015, *5*, 117–123.
- Mubiru, D.N.; Coyne, M.S.; Grove, J.H. Mortality of *Escherichia coli* O157:H7 in Two Soils with Different Physical and Chemical Properties. J. Environ. Qual. 2000, 29, 1821–1825. [CrossRef]
- 115. Jamieson, R.C.; Joy, D.M.; Lee, H.; Kostaschuk, R.; Gordon, R.J. Resuspension of Sediment-Associated *Escherichia coli* in a Natural Stream. *J. Environ. Qual.* 2005, 34, 581–589. [CrossRef]
- 116. Jamieson, R.C.; Gordon, R.J.; Sharples, K.E.; Madani, A.; Stratton, G.W. Movement and Persistence of Fecal Bacteria in Agricultural Soils and Subsurface Drainage Water: A Review. Can. Biosyst. Eng./Le Genie Des Biosyst. Au Can. 2002, 44, 1.1–1.9.
- 117. Weil, R.; Brady, N. The Nature and Properties of Soils, 15th ed.; Pearson: San Antonio, TX, USA, 2017; ISBN 978-0-13-325448-8.
- 118. Lau, C.L.; Clements, A.C.A.; Skelly, C.; Dobson, A.J.; Smythe, L.D.; Weinstein, P. Leptospirosis in American Samoa—Estimating and Mapping Risk Using Environmental Data. *PLoS Negl. Trop. Dis.* **2012**, *6*, e1669. [CrossRef] [PubMed]
- Hines, M.T. Chapter 32—Leptospirosis. In *Equine Infectious Diseases*, 2nd ed.; Sellon, D.C., Long, M.T., Eds.; W.B. Saunders: St. Louis, MO, USA, 2014; pp. 302–311.e5. ISBN 978-1-4557-0891-8.
- Vogeleer, P.; Tremblay, Y.D.N.; Mafu, A.A.; Jacques, M.; Harel, J. Life on the Outside: Role of Biofilms in Environmental Persistence of Shiga-Toxin Producing *Escherichia coli*. Front. Microbiol. 2014, 5, 317. [CrossRef] [PubMed]
- 121. Vinod Kumar, K.; Lall, C.; Raj, R.V.; Vedhagiri, K.; Vijayachari, P. Coexistence and Survival of Pathogenic Leptospires by Formation of Biofilm with Azospirillum. *FEMS Microbiol. Ecol.* **2015**, *91*, fiv051. [CrossRef]
- 122. Vinod Kumar, K.; Lall, C.; Raj, R.V.; Vijayachari, P. Coaggregation and Biofilm Formation of Leptospira with Staphylococcus Aureus. *Microbiol. Immunol.* 2019, 63, 147–150. [CrossRef]
- 123. Tsuboi, M.; Koizumi, N.; Hayakawa, K.; Kanagawa, S.; Ohmagari, N.; Kato, Y. Imported Leptospira Licerasiae Infection in Traveler Returning to Japan from Brazil. *Emerg. Infect. Dis.* **2017**, *23*, 548–549. [CrossRef]

- 124. Cilia, G.; Bertelloni, F.; Albini, S.; Fratini, F. Insight into the Epidemiology of Leptospirosis: A Review of Leptospira Isolations from "Unconventional" Hosts. *Animals* **2021**, *11*, 191. [CrossRef]
- 125. Grassmann, A.A.; Souza, J.D.; McBride, A.J.A. A Universal Vaccine against Leptospirosis: Are We Going in the Right Direction? *Front. Immunol.* 2017, *8*, 256. [CrossRef]
- 126. Garba, B.; Bahaman, A.R.; Bejo, S.K.; Zakaria, Z.; Mutalib, A.R.; Bande, F. Major Epidemiological Factors Associated with Leptospirosis in Malaysia. *Acta Trop.* **2018**, *178*, 242–247. [CrossRef]
- 127. Goarant, C. Leptospirosis: Risk Factors and Management Challenges in Developing Countries. *Res. Rep. Trop. Med.* **2016**, *7*, 49–62. [CrossRef]
- 128. Thiermann, A.B.; Frank, R.R. Human Leptospirosis in Detroit and the Role of Rats as Chronic Carriers. *Int. J. Zoonoses* **1980**, *7*, 62–72. [PubMed]
- 129. Demers, R.Y.; Frank, R.; Demers, P.; Clay, M. Leptospiral Exposure in Detroit Rodent Control Workers. *Am. J. Public Health* **1985**, 75, 1090–1091. [CrossRef] [PubMed]
- Costa, F.; Ribeiro, G.S.; Felzemburgh, R.D.M.; Santos, N.; Reis, R.B.; Santos, A.C.; Fraga, D.B.M.; Araujo, W.N.; Santana, C.; Childs, J.E.; et al. Influence of Household Rat Infestation on Leptospira Transmission in the Urban Slum Environment. *PLoS Negl. Trop. Dis.* 2014, 8, e3338. [CrossRef]
- 131. Santos, N.d.J.; Sousa, E.; Reis, M.G.; Ko, A.I.; Costa, F. Rat Infestation Associated with Environmental Deficiencies in an Urban Slum Community with High Risk of Leptospirosis Transmission. *Cad. De Saúde Pública* **2017**, *33*, 1463–1474. [CrossRef]
- 132. Barragan, V.; Nieto, N.; Keim, P.; Pearson, T. Meta-Analysis to Estimate the Load of Leptospira Excreted in Urine: Beyond Rats as Important Sources of Transmission in Low-Income Rural Communities. *BMC Res. Notes* **2017**, *10*, 71. [CrossRef]
- 133. Hathaway, S.C.; Blackmore, D.K. Ecological Aspects of the Epidemiology of Infection with Leptospires of the Ballum Serogroup in the Black Rat (*Rattus rattus*) and the Brown Rat (*Rattus norvegicus*) in New Zealand. *J. Hyg.* **1981**, *87*, 427–436. [CrossRef]
- 134. Boey, K.; Shiokawa, K.; Rajeev, S. Leptospira Infection in Rats: A Literature Review of Global Prevalence and Distribution. *PLoS Negl. Trop. Dis.* **2019**, *13*, e0007499. [CrossRef]
- Loan, H.K.; Van Cuong, N.; Takhampunya, R.; Kiet, B.T.; Campbell, J.; Them, L.N.; Bryant, J.E.; Tippayachai, B.; Van Hoang, N.; Morand, S.; et al. How Important Are Rats As Vectors of Leptospirosis in the Mekong Delta of Vietnam? *Vector Borne Zoonotic Dis.* 2015, 15, 56–64. [CrossRef]
- 136. Koizumi, N.; Muto, M.; Tanikawa, T.; Mizutani, H.; Sohmura, Y.; Hayashi, E.; Akao, N.; Hoshino, M.; Kawabata, H.; Watanabe, H. 2009 Human Leptospirosis Cases and the Prevalence of Rats Harbouring Leptospira Interrogans in Urban Areas of Tokyo, Japan. J. Med. Microbiol. 2009, 58, 1227–1230. [CrossRef]
- 137. Sarkar, U.; Nascimento, S.F.; Barbosa, R.; Martins, R.; Nuevo, H.; Kalafanos, I.; Grunstein, I.; Flannery, B.; Dias, J.P.; Riley, L.W.; et al. Population-Based Case-Control Investigation of Risk Factors for Leptospirosis during an Urban Epidemic. Am. J. Trop. Med. Hyg. 2002, 66, 605–610. [CrossRef]
- Masali, K.A.; Pulare, M.V.; Kachare, V.K.; Patil, M.B.; Reddi, S. Control and Prevention of Rat Fever (Leptospirosis) Outbreak in Six Villages of Raichur District, Karnataka. J. Indian Med. Assoc. 2007, 105, 632, 634–636.
- 139. Socolovschi, C.; Angelakis, E.; Renvoisé, A.; Fournier, P.-E.; Marié, J.L.; Davoust, B.; Stein, A.; Raoult, D. Strikes, Flooding, Rats, and Leptospirosis in Marseille, France. *Int. J. Infect. Dis.* 2011, *15*, e710–e715. [CrossRef] [PubMed]
- Strand, T.M.; Lohmus, M.; Vinnersten, T.P.; Råsbäck, T.; Sundström, K.; Bergström, T.; Lundkvist, Å. Highly Pathogenic Leptospira Found in Urban Brown Rats (*Rattus norvegicus*) in the Largest Cities of Sweden. *Vector-Borne Zoonotic Dis.* 2015, 15, 779–781. [CrossRef] [PubMed]
- 141. Webster, J.P.; Ellis, W.A.; Macdonald, D.W. Prevalence of Leptospira and Other Zoonoses in Wild Brown Rats on UK Farms. *Mammalia* **1995**, *59*, 615–622. [CrossRef]
- Ghneim, G.S.; Viers, J.H.; Chomel, B.B.; Kass, P.H.; Descollonges, D.A.; Johnson, M.L. Use of a Case-Control Study and Geographic Information Systems to Determine Environmental and Demographic Risk Factors for Canine Leptospirosis. *Vet. Res.* 2007, 38, 37–50. [CrossRef] [PubMed]
- Major, A.; Schweighauser, A.; Francey, T. Increasing Incidence of Canine Leptospirosis in Switzerland. Int. J. Environ. Res. Public Health 2014, 11, 7242–7260. [CrossRef] [PubMed]
- Weekes, C.C.; Everard, C.O.; Levett, P.N. Seroepidemiology of Canine Leptospirosis on the Island of Barbados. *Vet. Microbiol.* 1997, 57, 215–222. [CrossRef] [PubMed]
- 145. Guagliardo, S.A.J.; Iverson, S.A.; Reynolds, L.; Yaglom, H.; Venkat, H.; Galloway, R.; Levy, C.; Reindel, A.; Sylvester, T.; Kretschmer, M.; et al. Despite High-Risk Exposures, No Evidence of Zoonotic Transmission during a Canine Outbreak of Leptospirosis. *Zoonoses Public Health* 2019, 66, 223–231. [CrossRef]
- Venkataraman, K.S.; Nedunchelliyan, S. Epidemiology of an Outbreak of Leptospirosis in Man and Dog. Comp. Immunol. Microbiol. Infect. Dis. 1992, 15, 243–247. [CrossRef]
- 147. Sparkes, J.; Körtner, G.; Ballard, G.; Fleming, P.J.S. Spatial and Temporal Activity Patterns of Owned, Free-Roaming Dogs in Coastal Eastern Australia. *Prev. Vet. Med.* 2022, 204, 105641. [CrossRef]
- 148. Altheimer, K.; Jongwattanapisan, P.; Luengyosluechakul, S.; Pusoonthornthum, R.; Prapasarakul, N.; Kurilung, A.; Broens, E.M.; Wagenaar, J.A.; Goris, M.G.A.; Ahmed, A.A.; et al. Leptospira Infection and Shedding in Dogs in Thailand. *BMC Vet. Res.* 2020, 16, 89. [CrossRef] [PubMed]

- 149. Orr, B.; Westman, M.E.; Malik, R.; Purdie, A.; Craig, S.B.; Norris, J.M. Leptospirosis Is an Emerging Infectious Disease of Pig-Hunting Dogs and Humans in North Queensland. *PLoS Negl. Trop. Dis.* **2022**, *16*, e0010100. [CrossRef] [PubMed]
- Brown, V.R.; Bowen, R.A.; Bosco-Lauth, A.M. Zoonotic Pathogens from Feral Swine That Pose a Significant Threat to Public Health. *Transbound. Emerg. Dis.* 2018, 65, 649–659. [CrossRef] [PubMed]
- 151. Baker, T.F.; McEwen, S.A.; Prescott, J.F.; Meek, A.H. The Prevalence of Leptospirosis and Its Association with Multifocal Interstitial Nephritis in Swine at Slaughter. *Can. J. Vet. Res.* **1989**, *53*, 290–294.
- 152. Dreyfus, A.; Benschop, J.; Collins-Emerson, J.; Wilson, P.; Baker, M.G.; Heuer, C. Sero-Prevalence and Risk Factors for Leptospirosis in Abattoir Workers in New Zealand. *Int. J. Environ. Res. Public Health* **2014**, *11*, 1756–1775. [CrossRef]
- 153. Stephens, D.P.; Thomas, J.H. Leptospirosis: An Unusual Presentation. Crit. Care Resusc. 2006, 8, 215–218.
- 154. Macaluso, G.; Torina, A.; Blanda, V.; Guercio, A.; Lastra, A.; Giacchino, I.; D'Agostino, R.; Sciacca, C.; D'Incau, M.; Bertasio, C.; et al. Leptospira in Slaughtered Fattening Pigs in Southern Italy: Serological Survey and Molecular Typing. *Animals* 2022, 12, 585. [CrossRef]
- 155. Bolin, C.A.; Cassells, J.A.; Hill, H.T.; Frantz, J.C.; Nielsen, J.N. Reproductive Failure Associated with Leptospira Interrogans Serovar Bratislava Infection of Swine. J. Vet. Diagn. Investig. **1991**, *3*, 152–154. [CrossRef]
- 156. Kazami, A.; Watanabe, H.; Hayashi, T.; Kobayashi, K.; Ogawa, Y.; Yamamoto, K.; Adachi, Y. Serological Survey of Leptospirosis in Sows with Premature Birth and Stillbirth in Chiba and Gunma Prefectures of Japan. J. Vet. Med. Sci. 2002, 64, 735–737. [CrossRef]
- Eckert, K.D.; Keiter, D.A.; Beasley, J.C. Animal Visitation to Wild Pig (Sus scrofa) Wallows and Implications for Disease Transmission. J. Wildl. Dis. 2019, 55, 488–493. [CrossRef]
- 158. Gray, S.M.; Roloff, G.J.; Montgomery, R.A.; Beasley, J.C.; Pepin, K.M. Wild Pig Spatial Ecology and Behavior. In *Invasive Wild Pigs in North America: Ecology, Impacts, and Management*; VerCauteren, K.C., Ditchkoff, S.S., Beasley, J.C., Mayer, J.J., Roloff, G.J., Strickland, B.K., Eds.; CRC Press: Boca Raton, FL, USA, 2019; pp. 33–56. ISBN 978-1-315-23305-5.
- 159. Mori, M.; Bakinahe, R.; Vannoorenberghe, P.; Maris, J.; de Jong, E.; Tignon, M.; Marin, M.; Desqueper, D.; Fretin, D.; Behaeghel, I. Reproductive Disorders and Leptospirosis: A Case Study in a Mixed-Species Farm (Cattle and Swine). *Vet. Sci.* 2017, 4, 64. [CrossRef]
- 160. Poudel, A.; Hoque, M.M.; Madere, S.; Bolds, S.; Price, S.; Barua, S.; Adekanmbi, F.; Kalalah, A.; Kitchens, S.; Brown, V.; et al. Molecular and Serological Prevalence of *Leptospira* spp. in Feral Pigs (*Sus scrofa*) and Their Habitats in Alabama, USA. *Pathogens* 2020, 9, 857. [CrossRef] [PubMed]
- 161. Pearson, H.E.; Toribio, J.-A.L.M.L.; Lapidge, S.J.; Hernández-Jover, M. Evaluating the Risk of Pathogen Transmission from Wild Animals to Domestic Pigs in Australia. *Prev. Vet. Med.* **2016**, *123*, 39–51. [CrossRef] [PubMed]
- 162. Petri, F.A.M.; Sonalio, K.; de Souza Almeida, H.M.; Mechler-Dreibi, M.L.; Galdeano, J.V.B.; Mathias, L.A.; de Oliveira, L.G. Cross-Sectional Study of *Leptospira* spp. in Commercial Pig Farms in the State of Goiás, Brazil. *Trop. Anim. Health Prod.* 2020, 53, 13. [CrossRef]
- Miller, R.S.; Sweeney, S.J.; Slootmaker, C.; Grear, D.A.; Di Salvo, P.A.; Kiser, D.; Shwiff, S.A. Cross-Species Transmission Potential between Wild Pigs, Livestock, Poultry, Wildlife, and Humans: Implications for Disease Risk Management in North America. *Sci. Rep.* 2017, 7, 7821. [CrossRef] [PubMed]
- 164. Krawczyk, M. Serological studies on leptospirosis in wild boar. Med. Weter. 2000, 56, 440-443.
- Cleveland, C.A.; DeNicola, A.; Dubey, J.P.; Hill, D.E.; Berghaus, R.D.; Yabsley, M.J. Survey for Selected Pathogens in Wild Pigs (*Sus scrofa*) from Guam, Marianna Islands, USA. *Vet. Microbiol.* 2017, 205, 22–25. [CrossRef]
- Bengsen, A.J.; Gentle, M.N.; Mitchell, J.L.; Pearson, H.E.; Saunders, G.R. Impacts and Management of Wild Pigs Sus Scrofa in Australia. *Mammal Rev.* 2014, 44, 135–147. [CrossRef]
- 167. Corn, J.L.; Yabsley, M.J. Diseases and Parasites That Impact Wild Pigs and Species They Contact. In *Invasive Wild Pigs in North America: Ecology, Impacts and Management*; VerCauteren, K.C., Beasley, J.C., Ditchkoff, S.S., Mayer, J.J., Roloff, G.J., Strickland, B.K., Eds.; CRC Press: Boca Raton, FL, USA, 2019; pp. 83–126. ISBN 978-1-315-23305-5.
- 168. Hutton, T.; DeLiberto, T.; Owen, S.; Morrison, B. Disease Risks Associated with Increasing Feral Swine Numbers and Distribution in the United States. *Mich. Bov. Tuberc. Bibliogr. Database* **2006**.
- 169. Lewis, J.S.; Corn, J.L.; Mayer, J.J.; Jordan, T.R.; Farnsworth, M.L.; Burdett, C.L.; VerCauteren, K.C.; Sweeney, S.J.; Miller, R.S. Historical, Current, and Potential Population Size Estimates of Invasive Wild Pigs (*Sus scrofa*) in the United States. *Biol. Invasions* 2019, 21, 2373–2384. [CrossRef]
- Snow, N.P.; Miller, R.S.; Beasley, J.C.; Pepin, K.M. Wild Pig Population Dynamics. In *Invasive Wild Pigs in North America: Ecology, Impacts, and Management*; VerCauteren, K.C., Beasley, J.C., Ditchkoff, S.S., Mayer, J.J., Roloff, G.J., Strickland, B.K., Eds.; CRC Press: Boca Raton, FL, USA, 2019; pp. 57–82, ISBN 978-1-315-23305-5.
- 171. Barrios-Garcia, M.N.; Ballari, S.A. Impact of Wild Boar (*Sus scrofa*) in Its Introduced and Native Range: A Review. *Biol. Invasions* **2012**, 14, 2283–2300. [CrossRef]
- 172. Mînzat, R.M.; Tomescu, V. Detection of pathogenic leptospira in the waste water and sewage sludge of large pig breeding sites. *Arch. Exp. Veterinarmed.* **1975**, *29*, 557–562. [PubMed]
- 173. Ziemer, C.J.; Bonner, J.M.; Cole, D.; Vinjé, J.; Constantini, V.; Goyal, S.; Gramer, M.; Mackie, R.; Meng, X.J.; Myers, G.; et al. Fate and Transport of Zoonotic, Bacterial, Viral, and Parasitic Pathogens during Swine Manure Treatment, Storage, and Land Application1. *J. Anim. Sci.* **2010**, *88*, E84–E94. [CrossRef] [PubMed]
- 174. Baker, J.A.; Little, R.B. Leptospirosis in Cattle. J. Exp. Med. 1948, 88, 295–308. [CrossRef]

- 175. Lilenbaum, W.; Martins, G. Leptospirosis in Cattle: A Challenging Scenario for the Understanding of the Epidemiology. *Transbound. Emerg. Dis.* **2014**, *61*, 63–68. [CrossRef]
- 176. Ellis, W.A. Leptospirosis as a Cause of Reproductive Failure. Vet. Clin. N. Am. Food Anim. Pract. 1994, 10, 463–478. [CrossRef]
- 177. Ratnam, S.; Sundararaj, T.; Subramanian, S. Serological Evidence of Leptospirosis in a Human Population Following an Outbreak of the Disease in Cattle. *Trans. R. Soc. Trop. Med. Hyg.* **1983**, 77, 94–98. [CrossRef]
- 178. Zarantonelli, L.; Suanes, A.; Meny, P.; Buroni, F.; Nieves, C.; Salaberry, X.; Briano, C.; Ashfield, N.; Silveira, C.D.S.; Dutra, F.; et al. Isolation of Pathogenic Leptospira Strains from Naturally Infected Cattle in Uruguay Reveals High Serovar Diversity, and Uncovers a Relevant Risk for Human Leptospirosis. *PLoS Negl. Trop. Dis.* **2018**, *12*, e0006694. [CrossRef]
- 179. Zamir, L.; Baum, M.; Bardenstein, S.; Blum, S.E.; Moran-Gilad, J.; Perry Markovich, M.; King, R.; Lapid, R.; Hamad, F.; Even-Tov, B.; et al. The Association between Natural Drinking Water Sources and the Emergence of Zoonotic Leptospirosis among Grazing Beef Cattle Herds during a Human Outbreak. *One Health* **2022**, *14*, 100372. [CrossRef]
- Yatbantoong, N.; Chaiyarat, R. Factors Associated with Leptospirosis in Domestic Cattle in Salakphra Wildlife Sanctuary, Thailand. Int. J. Environ. Res. Public Health 2019, 16, 1042. [CrossRef]
- Martins, G.; Lilenbaum, W. Control of Bovine Leptospirosis: Aspects for Consideration in a Tropical Environment. *Res. Vet. Sci.* 2017, 112, 156–160. [CrossRef] [PubMed]
- 182. Sonada, R.B.; de Azevedo, S.S.; Soto, F.R.M.; da Costa, D.F.; de Morais, Z.M.; de Souza, G.O.; Gonçales, A.P.; Miraglia, F.; Vasconcellos, S.A. Efficacy of Leptospiral Commercial Vaccines on the Protection against an Autochtonous Strain Recovered in Brazil. *Braz. J. Microbiol.* 2018, 49, 347–350. [CrossRef] [PubMed]
- Žele-Vengušt, D.; Lindtner-Knific, R.; Mlakar-Hrženjak, N.; Jerina, K.; Vengušt, G. Exposure of Free-Ranging Wild Animals to Zoonotic Leptospira Interrogans Sensu Stricto in Slovenia. *Animals* 2021, 11, 2722. [CrossRef] [PubMed]
- 184. Millán, J.; Candela, M.G.; López-Bao, J.V.; Pereira, M.; Jiménez, M.Á.; León-Vizcaíno, L. Leptospirosis in Wild and Domestic Carnivores in Natural Areas in Andalusia, Spain. *Vector-Borne Zoonotic Dis.* 2009, 9, 549–554. [CrossRef] [PubMed]
- 185. Pedersen, K.; Anderson, T.D.; Maison, R.M.; Wiscomb, G.W.; Pipas, M.J.; Sinnett, D.R.; Baroch, J.A.; Gidlewski, T. Leptospira Antibodies Detected in Wildlife in the Usa and the Us Virgin Islands. *J. Wildl. Dis.* **2018**, *54*, 450–459. [CrossRef] [PubMed]
- 186. Anderson, A.N.; Long, A.M.; LaCour, J.M.; Bresnan, A.M.; Bushaw, J.D.; Gerrits, A.P.; Hunt, J.D.; Moscicki, D.J.; Raginski, N.M.; Stafford, B.; et al. Seroprevalence of Leptospira Among Selected Mammals on a Wildlife Management Area in Louisiana, USA. J. Wildl. Dis. 2021, 58, 183–187. [CrossRef]
- 187. de Albuquerque, N.F.; Martins, G.; Medeiros, L.; Lilenbaum, W.; Ribeiro, V.M.F. The Role of Capybaras as Carriers of Leptospires in Periurban and Rural Areas in the Western Amazon. *Acta Trop.* **2017**, *169*, 57–61. [CrossRef]
- 188. Obiegala, A.; Woll, D.; Karnath, C.; Silaghi, C.; Schex, S.; Eßbauer, S.; Pfeffer, M. Prevalence and Genotype Allocation of Pathogenic Leptospira Species in Small Mammals from Various Habitat Types in Germany. *PLoS Negl. Trop. Dis.* **2016**, *10*, e0004501. [CrossRef]
- Shearer, K.E.; Harte, M.J.; Ojkic, D.; DeLay, J.; Campbell, D. Detection of *Leptospira* spp. in Wildlife Reservoir Hosts in Ontario through Comparison of Immunohistochemical and Polymerase Chain Reaction Genotyping Methods. *Can. Vet. J.* 2014, 55, 240–248.
- 190. Fiechter, G.I. Leptospirosis in Switzerland: An Emerging Disease or Emerging Awareness? Ph.D. Thesis, University of Geneva, Geneva, Switzerland, 2020.
- 191. Maas, M.; Glorie, J.; Dam-Deisz, C.; de Vries, A.; Franssen, F.F.J.; Jaarsma, R.I.; Hengeveld, P.D.; Dierikx, C.M.; van der Giessen, J.W.B.; Opsteegh, M. Zoonotic Pathogens in Eurasian Beavers (*Castor fiber*) in the Netherlands. J. Wildl. Dis. 2022, 58, 404–408. [CrossRef]
- 192. Nolet, B.A.; Broekhuizen, S.; Dorrestein, G.M.; Rienks, K.M. Infectious Diseases as Main Causes of Mortality to Beavers Castor Fiber after Translocation to the Netherlands. *J. Zool.* **1997**, 241, 35–42. [CrossRef]
- López-Pérez, A.M.; Carreón-Arroyo, G.; Atilano, D.; Vigueras-Galván, A.L.; Valdez, C.; Toyos, D.; Mendizabal, D.; López-Islas, J.; Suzán, G. Presence of Antibodies to *Leptospira* spp. in Black-Tailed Prairie Dogs (*Cynomys ludovicianus*) and Beavers (*Castor canadensis*) in Northwestern Mexico. J. Wildl. Dis. 2017, 53, 880–884. [CrossRef] [PubMed]
- 194. Diesch, S.L.; Crawford, R.P.; McCulloch, W.F.; Top, F.H. Human Leptospirosis Acquired from Squirrels. *N. Engl. J. Med.* **1967**, 276, 838–842. [CrossRef] [PubMed]
- 195. Shotts, E.J.; Andrews, C.L.; Harvey, T.W. Leptospirosis in Selected Wild Mammals of the Florida Panhandle and Southwestern Georgia. J. Am. Vet. Med. Assoc. 1975, 167, 587–589.
- 196. Nawtaisong, P.; Robinson, M.T.; Khammavong, K.; Milavong, P.; Rachlin, A.; Dittrich, S.; Dubot-Pérès, A.; Vongsouvath, M.; Horwood, P.F.; Dussart, P.; et al. Zoonotic Pathogens in Wildlife Traded in Markets for Human Consumption, Laos. *Emerg. Infect. Dis.* 2022, *28*, 860–864. [CrossRef]
- 197. Omonona, A.O.; Jubril, A.J.; Durosinmi, O.M.; Adetuga, A.T. Squirrels and Transmission of Leptospirosis: Awareness and Perception of the University of Ibadan Main Campus Residents, Nigeria. *Afr. J. Health Saf. Environ.* **2022**, *3*, 01–12. [CrossRef]
- 198. Dirsmith, K.; VanDalen, K.; Fry, T.; Charles, B.; VerCauteren, K.; Duncan, C. Leptospirosis in Fox Squirrels (*Sciurus niger*) of Larimer County, Colorado, USA. J. Wildl. Dis. 2013, 49, 641–645. [CrossRef]
- 199. Daud, A.; Fuzi, N.M.H.M.; Arshad, M.M.; Kamarudin, S.; Mohammad, W.M.Z.W.; Amran, F.; Ismail, N. Leptospirosis Seropositivity and Its Serovars among Cattle in Northeastern Malaysia. *Vet. World* **2018**, *11*, 840–844. [CrossRef]

- Anan'ina, I.V.; Korenberg, E.I.; Tserennorov, D.; Savel'eva, O.V.; Batjav, D.; Otgonbaatar, D.; Enkhbold, N.; Tsend, E.; Erdenechimeg, B. Detection of leptospirosis infection in certain wild and domestic animals in Mongolia. *Zh. Mikrobiol. Epidemiol. Immunobiol.* 2011, 36–39.
- Thayaparan, S.; Robertson, I.D.; Abdullah, M.T. Serological and Molecular Detection of *Leptospira* spp. from Small Wild Mammals Captured in Sarawak, Malaysia. *Malays. J. Microbiol.* 2015, 11, 93–101. [CrossRef]
- Hathaway, S.C.; Blackmore, D.K.; Marshall, R.B. Leptospirosis in Free-Living Species in New Zealand. JWDI 1981, 17, 489–496.
 [CrossRef]
- 203. Verma, A.; Beigel, B.; Smola, C.C.; Kitts-Morgan, S.; Kish, D.; Nader, P.; Morgan, J.; Roberson, J.; Christmann, U.; Gruszynski, K.; et al. Evidence of Leptospiral Presence in the Cumberland Gap Region. *PLoS Negl. Trop. Dis.* 2019, 13, e0007990. [CrossRef] [PubMed]
- 204. Vieira, A.S.; Pinto, P.S.; Lilenbaum, W. A Systematic Review of Leptospirosis on Wild Animals in Latin America. Trop. Anim. Health Prod. 2018, 50, 229–238. [CrossRef] [PubMed]
- 205. Dezzutto, D.; Barbero, R.; Canale, G.; Acutis, P.L.; Biolatti, C.; Dogliero, A.; Mitzy, M.D.; Francone, P.; Colzani, A.; Bergagna, S.; et al. Detection of *Leptospira* spp. in Water Turtle (*Trachemys scripta*) Living in Ponds of Urban Parks. *Vet. Sci.* 2017, 4, 51. [CrossRef] [PubMed]
- Oliveira, J.P.; Kawanami, A.E.; Silva, A.S.L.; Chung, D.G.; Werther, K. Detection of *Leptospira* spp. in Wild Phrynops Geoffroanus (Geoffroy's Side-Necked Turtle) in Urban Environment. *Acta Trop.* 2016, 164, 165–168. [CrossRef]
- 207. Miranda, J.M.S.; Rocha, K.d.S.; Monteiro, L.H.; Baia, I.W.M.; Monteiro, T.R.M.; Brito, J.d.S.; Mesquita, E.Y.E.; de Moraes, C.C.G. Presence of Anti-*Leptospira* spp. Antibodies in Captive Yellow-Spotted River Turtles (*Podocnemis unifilis*) in the Eastern Amazon. *Cienc. Rural* 2020, 50. [CrossRef]
- 208. Lindtner-Knific, R.; Vergles-Rataj, A.; Vlahović, K.; Zrimšek, P.; Dovč, A. Prevalence of Antibodies against Leptospira Sp. in Snakes, Lizards and Turtles in Slovenia. *Acta Vet. Scand.* 2013, *55*, 65. [CrossRef]
- Rodamilans, G.M.; Fonseca, M.S.; Paz, L.N.; Fernandez, C.C.; Biondi, I.; Lira-da-Silva, R.M.; Meyer, R.; Pinna, M.H.; Portela, R.D. Leptospira Interrogans in Wild Boa Constrictor Snakes from Northeast Brazil Peri-urban Rainforest Fragments. *Acta Trop.* 2020, 209, 105572. [CrossRef]
- Rockwell, K.E.; Thompson, D.; Maddox, C.; Mitchell, M.A. Blanding's Turtles (*Emydoidea blandingii*) as a Reservoir for *Leptospira* spp. *PLoS ONE* 2019, 14, e0210688. [CrossRef]
- 211. Rocha, K.d.S.; Baia, I.W.M.; Monteiro, L.H.; Miranda, J.M.S.; Monteiro, T.R.M.; da Silva, A.F.; dos Reis, T.A.; Ferreira, M.F.S.; Mesquita, E.Y.E.; de Moraes, C.C.G. Identification of Antibodies to *Leptospira* spp. in the Spot-Legged Turtle (*Rhinoclemmys punctularia*) Maintained in Captivity. *Semin. Ciências Agrárias* 2019, 40, 3763–3768. [CrossRef]
- Pérez-Flores, J.; Charruau, P.; Cedeño-Vázquez, R.; Atilano, D. Evidence for Wild Crocodiles as a Risk for Human Leptospirosis, Mexico. *EcoHealth* 2017, 14, 58–68. [CrossRef] [PubMed]
- 213. Rossetti, C.A.; Uhart, M.; Romero, G.N.; Prado, W. Detection of Leptospiral Antibodies in Caimans from the Argentinian Chaco. *Vet. Rec.* 2003, 153, 632–633. [CrossRef] [PubMed]
- Silva, É.F.; Seyffert, N.; Cerqueira, G.M.; Leihs, K.P.; Athanazio, D.A.; Valente, A.L.S.; Dellagostin, O.A.; Brod, C.S. Serum Antileptospiral Agglutinins in Freshwater Turtles from Southern Brazil. *Braz. J. Microbiol.* 2009, 40, 227–230. [CrossRef] [PubMed]
- Van Der Hoeden, J.; Szenberg, E.; Evenchik, Z. Leptospira-Agglutmating Factors in Turtle Sera. Nature 1961, 190, 95–96. [CrossRef] [PubMed]
- 216. Biscola, N.P.; Fornazari, F.; Saad, E.; Richini-Pereira, V.B.; Campagner, M.V.; Langoni, H.; Barraviera, B.; Ferreira Junior, R.S. Serological Investigation and PCR in Detection of Pathogenic Leptospires in Snakes. *Pesq. Vet. Bras.* 2011, *31*, 806–811. [CrossRef]
- Everard, C.O.; Carrington, D.G.; Korver, H.; Burke, R.; Everard, J.D.; Gravekamp, C. Leptospires in the Whistling Frog (*Eleutherodactylus johnstonei*) on Barbados. J. Trop. Med. Hyg. 1990, 93, 140–145.
- 218. Gravekamp, C.; Korver, H.; Montgomery, J.; Everard, C.O.R.; Carrington, D.; Ellis, W.A.; Terpstra, W.J. Leptospires Isolated from Toads and Frogs on the Island of Barbados. *Zentralblatt Für Bakteriol.* **1991**, 275, 403–411. [CrossRef]
- Rodrigues, T.C.S.; Santos, A.L.Q.; Lima, A.M.C.; Gomes, D.O.; Brites, V.L.C. Anti-*Leptospira* spp. Antibodies in Crotalus Durissus Collilineatus Kept in Captivity and Its Zoonotic Relevance. *Acta Trop.* 2016, 158, 39–42. [CrossRef]
- 220. Glosser, J.W.; Sulzer, C.R.; Eberhardt, M.; Winkler, W.G. Cultural and Serologic Evidence of *Leptospira interrogans* Serotype Tarassovi Infection in Turtles. *J. Wildl. Dis.* **1974**, *10*, 429–435. [CrossRef]
- 221. Heard, M.J.; Smith, K.F.; Ripp, K.J.; Berger, M.; Chen, J.; Dittmeier, J.; Goter, M.; Mcgarvey, S.T.; Ryan, E. The Threat of Disease Increases as Species Move Toward Extinction. *Conserv. Biol.* **2013**, *27*, 1378–1388. [CrossRef]
- 222. Millán, J.; Candela, M.G.; Palomares, F.; Cubero, M.J.; Rodríguez, A.; Barral, M.; de la Fuente, J.; Almería, S.; León-Vizcaíno, L. Disease Threats to the Endangered Iberian Lynx (*Lynx pardinus*). Vet. J. 2009, 182, 114–124. [CrossRef] [PubMed]
- Clifford, D.L.; Mazet, J.A.K.; Dubovi, E.J.; Garcelon, D.K.; Coonan, T.J.; Conrad, P.A.; Munson, L. Pathogen Exposure in Endangered Island Fox (*Urocyon littoralis*) Populations: Implications for Conservation Management. *Biol. Conserv.* 2006, 131, 230–243. [CrossRef] [PubMed]
- 224. Aragón-Martínez, A.; Olivera-Gómez, L.D.; Jiménez-Domínguez, D. Seasonal Prevalence of Antibodies to *Leptospira interrogans* in Antillean Manatees from a Landlocked Lake in Tabasco, Mexico. J. Wildl. Dis. 2014, 50, 505–511. [CrossRef] [PubMed]
- 225. Delgado, P.M.; Perea, N.S.; Garcia, C.B.; Davila, C.R.G. Detection of Infection with *Leptospira* spp. in Manatees (*Trichechus inunguis*) of the Peruvian Amazon. *Lat. Am. J. Aquat. Mamm.* **2015**, *10*, 58–61. [CrossRef]

- 226. Mathews, P.D.; da Silva, V.M.F.; Rosas, F.C.W.; d'Affonseca Neto, J.A.; Lazzarini, S.M.; Ribeiro, D.C.; Dubey, J.P.; Vasconcellos, S.A.; Gennari, S.M. Occurrence of Antibodies to Toxoplasma Gondii and *Lepstospira* spp. in Manatees (*Trichechus inunguis*) of the Brazilian Amazon. *J. Zoo Wildl. Med.* 2012, 43, 85–88. [CrossRef]
- 227. Sulzner, K.; Kreuder Johnson, C.; Bonde, R.K.; Auil Gomez, N.; Powell, J.; Nielsen, K.; Luttrell, M.P.; Osterhaus, A.D.M.E.; Aguirre, A.A. Health Assessment and Seroepidemiologic Survey of Potential Pathogens in Wild Antillean Manatees (*Trichechus manatus manatus*). PLoS ONE 2012, 7, e44517. [CrossRef]
- 228. Rodrigues, T.C.S.; Santos, A.L.Q.; Pinheiro, E.S.; Piatti, R.M.; Castro, V.; Buiatte, A.B.G.; Lima, A.M.C.; Marmontel, M. Survey for Leptospira and Brucella in Amazonian Manatees, Amazon River Dolphins, and a Tucuxi in the Brazilian Amazon. *Dis. Aquat. Org.* 2022, 150, 17–29. [CrossRef]
- 229. Sánchez-Sarmiento, A.M.; Carvalho, V.L.; Meirelles, A.C.O.; Gravena, W.; Marigo, J.; Sacristán, C.; Costa-Silva, S.; Groch, K.R.; Silva, N.d.S.; Neto, J.S.F.; et al. Survey of *Brucella* spp. and *Leptospira* spp. Antibodies in Cetaceans and Manatees of the Amazon Basin and Atlantic Ocean, Brazil. *Dis. Aquat. Org.* 2018, 132, 1–11. [CrossRef]
- 230. Smith, A.W.; Brown, R.J.; Skilling, D.E.; DeLong, R.L. Leptospira Pomona and Reproductive Failure in California Sea Lions. J. Am. Vet. Med. Assoc. 1974, 165, 996–998.
- Marreros, N.; Zürcher-Giovannini, S.; Origgi, F.C.; Djelouadji, Z.; Wimmershoff, J.; Pewsner, M.; Akdesir, E.; Batista Linhares, M.; Kodjo, A.; Ryser-Degiorgis, M.-P. Fatal Leptospirosis in Free-Ranging Eurasian Beavers (*Castor fiber L.*), Switzerland. *Transbound*. *Emerg. Dis.* 2018, 65, 1297–1306. [CrossRef]
- Girling, S.J.; Naylor, A.; Fraser, M.; Campbell-Palmer, R. Reintroducing Beavers Castor Fiber to Britain: A Disease Risk Analysis. Mammal Rev. 2019, 49, 300–323. [CrossRef]
- 233. Goodman, G.; Girling, S.; Pizzi, R.; Meredith, A.; Rosell, F.; Campbell-Palmer, R. Establishment of a Health Surveillance Program for Reintroduction of the Eurasian Beaver (*Castor fiber*) into Scotland. *J. Wildl. Dis.* **2012**, *48*, 971–978. [CrossRef] [PubMed]
- Girling, S.J.; Goodman, G.; Burr, P.; Pizzi, R.; Naylor, A.; Cole, G.; Brown, D.; Fraser, M.; Rosell, F.N.; Schwab, G.; et al. Evidence of Leptospira Species and Their Significance during Reintroduction of Eurasian Beavers (*Castor fiber*) to Great Britain. *Vet. Rec.* 2019, 185, 482. [CrossRef] [PubMed]
- 235. Campbell-Palmer, R.; Rosell, F.; Naylor, A.; Cole, G.; Mota, S.; Brown, D.; Fraser, M.; Pizzi, R.; Elliott, M.; Wilson, K.; et al. Eurasian Beaver (*Castor fiber*) Health Surveillance in Britain: Assessing a Disjunctive Reintroduced Population. *Vet. Rec.* 2021, 188, e84. [CrossRef]
- 236. Goodman, G.; Meredith, A.; Girling, S.; Rosell, F.; Campbell-Palmer, R. Outcomes of a 'One Health' Monitoring Approach to a Five-Year Beaver (*Castor fiber*) Reintroduction Trial in Scotland. *EcoHealth* **2017**, *14*, 139–143. [CrossRef]
- 237. Gelling, M.; Zochowski, W.; Macdonald, D.W.; Johnson, A.; Palmer, M.; Mathews, F. Leptospirosis Acquisition Following the Reintroduction of Wildlife. *Vet. Rec.* 2015, 177, 440. [CrossRef]
- Hrdina, A.; Romportl, D. Evaluating Global Biodiversity Hotspots—Very Rich and Even More Endangered. J. Landsc. Ecol. 2017, 10, 108–115. [CrossRef]
- Galan, D.I.; Roess, A.A.; Pereira, S.V.C.; Schneider, M.C. Epidemiology of Human Leptospirosis in Urban and Rural Areas of Brazil, 2000–2015. PLoS ONE 2021, 16, e0247763. [CrossRef]
- Hagan, J.E.; Moraga, P.; Costa, F.; Capian, N.; Ribeiro, G.S.; Wunder, E.A., Jr.; Felzemburgh, R.D.M.; Reis, R.B.; Nery, N.; Santana, F.S.; et al. Spatiotemporal Determinants of Urban Leptospirosis Transmission: Four-Year Prospective Cohort Study of Slum Residents in Brazil. *PLoS Negl. Trop. Dis.* 2016, 10, e0004275. [CrossRef]
- 241. Vinetz, J.M.; Glass, G.E.; Flexner, C.E.; Mueller, P.; Kaslow, D.C. Sporadic Urban Leptospirosis. *Ann. Intern. Med.* **1996**, 125, 794–798. [CrossRef]
- 242. Miller, A.G.; Ebelt, S.; Levy, K. Combined Sewer Overflows and Gastrointestinal Illness in Atlanta, 2002–2013: Evaluating the Impact of Infrastructure Improvements. *Environ. Health Perspect.* **2022**, *130*, 57009. [CrossRef] [PubMed]
- 243. Pathogens in Urban Stormwater Systems; Urban Water Resources Research Council: Reston, VA, USA, 2014.
- 244. Clary, J.; Ervin, J.; Steets, B.; Olson, C. Pathogens in Urban Stormwater Systems: Where Are We Now? J. Sustain. Water Built Environ. 2022, 8, 02521004. [CrossRef]
- 245. Yusof, M.A.; Mohd-Taib, F.S.; Ishak, S.N.; Md-Nor, S.; Md-Sah, S.A.; Mohamed, N.Z.; Azhari, N.N.; Neela, V.; Sekawi, Z. Microhabitat Factors Influenced the Prevalence of Pathogenic *Leptospira* spp. in Small Mammal Host. *EcoHealth* 2019, 16, 260–274. [CrossRef]
- 246. Benacer, D.; Woh, P.Y.; Mohd Zain, S.N.; Amran, F.; Thong, K.L. Pathogenic and Saprophytic Leptospira Species in Water and Soils from Selected Urban Sites in Peninsular Malaysia. *Microbes Environ.* **2013**, *28*, 135–140. [CrossRef] [PubMed]
- 247. Jackson, C.S.; Gracia, J.N. Addressing Health and Health-Care Disparities: The Role of a Diverse Workforce and the Social Determinants of Health. *Public Health Rep.* 2014, 129, 57–61. [CrossRef] [PubMed]
- 248. Deria, A.; Ghannad, P.; Lee, Y.-C. Evaluating Implications of Flood Vulnerability Factors with Respect to Income Levels for Building Long-Term Disaster Resilience of Low-Income Communities. *Int. J. Disaster Risk Reduct.* 2020, 48, 101608. [CrossRef]
- Koks, E.E.; Jongman, B.; Husby, T.G.; Botzen, W.J.W. Combining Hazard, Exposure and Social Vulnerability to Provide Lessons for Flood Risk Management. *Environ. Sci. Policy* 2015, 47, 42–52. [CrossRef]
- Rufat, S.; Tate, E.; Burton, C.G.; Maroof, A.S. Social Vulnerability to Floods: Review of Case Studies and Implications for Measurement. Int. J. Disaster Risk Reduct. 2015, 14, 470–486. [CrossRef]

- Tidman, R.; Abela-Ridder, B.; de Castañeda, R.R. The Impact of Climate Change on Neglected Tropical Diseases: A Systematic Review. *Trans. R. Soc. Trop. Med. Hyg.* 2021, 115, 147–168. [CrossRef]
- Brennan, F.P.; O'Flaherty, V.; Kramers, G.; Grant, J.; Richards, K.G. Long-Term Persistence and Leaching of *Escherichia coli* in Temperate Maritime Soils. *Appl. Environ. Microbiol.* 2010, 76, 1449–1455. [CrossRef]
- 253. Keesing, F.; Belden, L.K.; Daszak, P.; Dobson, A.; Harvell, C.D.; Holt, R.D.; Hudson, P.; Jolles, A.; Jones, K.E.; Mitchell, C.E.; et al. Impacts of Biodiversity on the Emergence and Transmission of Infectious Diseases. *Nature* 2010, 468, 647–652. [CrossRef]
- Wegner, G.I.; Murray, K.A.; Springmann, M.; Muller, A.; Sokolow, S.H.; Saylors, K.; Morens, D.M. Averting Wildlife-Borne Infectious Disease Epidemics Requires a Focus on Socio-Ecological Drivers and a Redesign of the Global Food System. *eClinicalMedicine* 2022, 47, 101386. [CrossRef] [PubMed]
- 255. Jones, B.A.; Grace, D.; Kock, R.; Alonso, S.; Rushton, J.; Said, M.Y.; McKeever, D.; Mutua, F.; Young, J.; McDermott, J.; et al. Zoonosis Emergence Linked to Agricultural Intensification and Environmental Change. *Proc. Natl. Acad. Sci. USA* 2013, 110, 8399–8404. [CrossRef]
- Ryu, S.; Lau, C.L.; Chun, B.C. The Impact of Livestock Manure Control Policy on Human Leptospirosis in Republic of Korea Using Interrupted Time Series Analysis. *Epidemiol. Infect.* 2017, 145, 1320–1325. [CrossRef] [PubMed]
- Faine, S.; World Health Organization. Guidelines for the Control of Leptospirosis; World Health Organization: Geneva, Switzerland, 1982; ISBN 978-92-4-170067-2.
- Crim, J.F.; Schoonover, J.E.; Lockaby, B.G. Assessment of Fecal Coliform and *Escherichia coli* Across a Land Cover Gradient in West Georgia Streams. Water Qual. Expo. Health 2012, 4, 143–158. [CrossRef]
- Nagy, R.C.; Lockaby, B.G. Urbanization in the Southeastern United States: Socioeconomic Forces and Ecological Responses along an Urban-Rural Gradient. Urban Ecosyst. 2011, 14, 71–86. [CrossRef]
- Paul, M.J.; Meyer, J.L. Streams in the Urban Landscape. In Urban Ecology: An International Perspective on the Interaction Between Humans and Nature; Marzluff, J.M., Shulenberger, E., Endlicher, W., Alberti, M., Bradley, G., Ryan, C., Simon, U., ZumBrunnen, C., Eds.; Springer: Boston, MA, USA, 2008; pp. 207–231, ISBN 978-0-387-73412-5.
- 261. Umeh, C.A.; Feeley, F.G. Inequitable Access to Health Care by the Poor in Community-Based Health Insurance Programs: A Review of Studies From Low- and Middle-Income Countries. *Glob. Health Sci. Pract.* **2017**, *5*, 299–314. [CrossRef]
- Nagy, R.C.; Lockaby, B.G.; Helms, B.; Kalin, L.; Stoeckel, D. Water Resources and Land Use and Cover in a Humid Region: The Southeastern United States. J. Environ. Qual. 2011, 40, 867–878. [CrossRef]
- Nagy, R.C.; Lockaby, B.G.; Kalin, L.; Anderson, C. Effects of Urbanization on Stream Hydrology and Water Quality: The Florida Gulf Coast. *Hydrol. Process.* 2012, 26, 2019–2030. [CrossRef]
- Glas, M.S.; Sato, Y.; Ulstrup, K.E.; Bourne, D.G. Biogeochemical Conditions Determine Virulence of Black Band Disease in Corals. ISME J. 2012, 6, 1526–1534. [CrossRef]
- Liu, S.; Jiang, Z.; Deng, Y.; Wu, Y.; Zhang, J.; Zhao, C.; Huang, D.; Huang, X.; Trevathan-Tackett, S.M. Effects of Nutrient Loading on Sediment Bacterial and Pathogen Communities within Seagrass Meadows. *MicrobiologyOpen* 2018, 7, e00600. [CrossRef] [PubMed]
- Freudenberg, N.; Pastor, M.; Israel, B. Strengthening Community Capacity to Participate in Making Decisions to Reduce Disproportionate Environmental Exposures. *Am. J. Public Health* 2011, 101, S123–S130. [CrossRef] [PubMed]
- Joyce, L.A.; Coulson, D. Climate Scenarios and Projections: A Technical Document Supporting the USDA Forest Service 2020 RPA Assessment; General Technical Report RMRS-GTR-413; U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station: Fort Collins, CO, USA, 2020; 85p. [CrossRef]
- Hassell, J.M.; Begon, M.; Ward, M.J.; Fèvre, E.M. Urbanization and Disease Emergence: Dynamics at the Wildlife–Livestock– Human Interface. *Trends Ecol. Evol.* 2017, 32, 55–67. [CrossRef] [PubMed]

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