

Contribution of Manure-Spreading Operations to Bioaerosols and Antibiotic Resistance Genes' Emission

Mahsa Baghdadi ^{1,2}, Patrick Brassard ³, Stéphane Godbout ³, Valérie Létourneau ^{1,2}, Nathalie Turgeon ^{1,2}, Florent Rossi ^{1,2}, Émie Lachance ^{1,2}, Marc Veillette ^{1,2}, Marie-Lou Gaucher ⁴ and Caroline Duchaine ^{1,2,5,*}

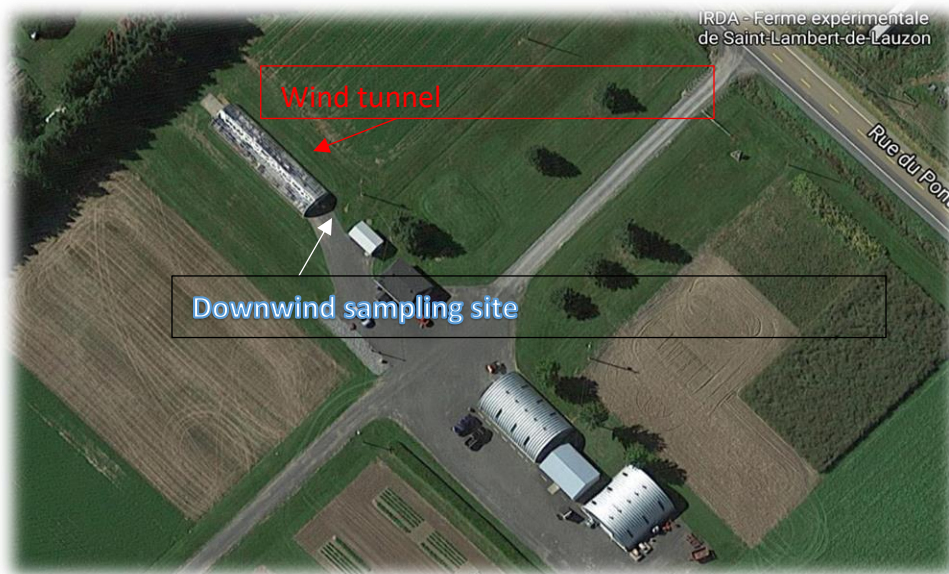
Supplementary Material

Section S1. Wind tunnel characteristics

A field-scale wind tunnel was built at IRDA (*Institut de recherche et de développement en agroenvironnement, Québec, Canada*) to spread manure in a controlled environment and measure gas, odor and bioaerosol emissions. The wind tunnel was scaled to accommodate agricultural machinery used for small-scale farming operations (8 m width x 30 m length x 4 m height). Air was drawn into the wind tunnel by ten, speed tunable, 24-inch diameter fans who allow wind speed of 0.2 to 1.2 m/s (5 to 30 m³/s). The environmental conditions (wind speed, temperature, and humidity) are closely monitored in the tunnel with an INTERCAP® humidity and temperature probe (VAISALA - HMP60). The speed of the fans was always 0.32 m/s which is equal to 8 m³/s in this experiment [20]. Inside the wind tunnel, an (8 m width x 22 m length x 0.6 height) strip of soil harvested from the nearby farming sites was disposed. The wind tunnel is accessible by two doors (front and back). Spreader-vehicles' entrance was located at the back of the tunnel (upwind) and spreading was always performed from the back to the front of the tunnel (downwind to upwind) by operating two successive back and forth on the strip of soil for solid manure, and only one pass for pig slurry. All doors were closed during spreading. Because the strip of soil tends dry quickly inside the wind tunnel, the soil was irrigated before each spreading on the entire surface to prevent artificial particle emissions caused by the passage of the spreader-vehicle.



A.



B.

Figure S1. A: Positioning of the exhaust fans of the wind tunnel and **B:** a view of wind tunnel and the downwind sampling site



Figure S2. Pictures of the different spreaders used to apply different types of manure inside the wind tunnel. Splash plate (A, D) and dribble bar (E) were connected through a hose to a storage tank (B) and used to spread pig slurry. Horizontal beater spreader (C) was used to apply solid (poultry and cow) manure.

Table S1. Physical characteristics of manure

	Number of samplings	Dry Matter content (%)
Cow Manure	Field trip 6	36.5
	Field trip 7	34.8
	Field trip 8	29.3
	Field trip 9	35.4
	Field trip 10	39.4
	Field trip 11	34.5
Pig slurry - Splash plate	Field trip 12	8.89
	Field trip 13	7.90
	Field trip 14	7.91
	Field trip 15	9.04
	Field trip 16	8.47
	Field trip 17	8.55
Poultry Manure	Field trip 18	47.9
	Field trip 19	48.8
	Field trip 20	59.6
	Field trip 21	54.3
	Field trip 22	50.5
	Field trip 23	53.7
Pig slurry - Dribble Bar	Field trip 24	6.67
	Field trip 25	6.47
	Field trip 26	6.16
	Field trip 27	6.44
	Field trip 28	11.2
	Field trip 29	11.2

Table S2. qPCR thermoprotocols

qPCR Protocols	Total bacteria (EUB)	40 cycles
Thermo Protocol	95 °C	3 min
	95 °C	20s
	62 °C	1 min
	Go to step 2	39x
	<i>Aerococcus</i> Phage	40 cycles
Thermo Protocol	95 °C	3 min
	95 °C	15s
	60°C	60s
	Go to step 2	39x
	<i>Archaea- rRNA 16S</i>	40 cycles
Thermo Protocol	95 °C	3 min
	95 °C	10s
	55.5 °C	20s
	72 °C	25s
	Go to step 2	39X
	72 °C	10 min
Melting Curve	50 °C	31s
	50 °C	0.02 + 0.5/s
	Go to step 9	90X
	<i>Entero- rRNA 16S</i>	40 cycles
Thermo Protocol	95 °C	3 min
	94 °C	15s
	57 °C	30s
	72 °C	30s
	Go to step 2	39X
	72 °C	10 min
Melting Curve	50 °C	31s
	50 °C	0.02 + 0.5/s
	Go to step 9	90X
	<i>E.coli- rRNA 16S</i>	40 cycles
Thermo Protocol	95 °C	3 min
	95 °C	15s
	55 °C	30s
	72 °C	30s

	Go to step 2	39X
	72 °C	10 min
Melting Curve	50 °C	31s
	50 °C	0.02 + 0.5/s
	Go to step 9	90X

Table S1. Sequences of primers and probes used for quantification by PCR of total bacteria [44,45], *E. coli*, *Enterococcus* [46], *Archaea* [47,48], the phage *vB_AviM_AVP* of *Aerococcus viridans*, ARGs and MGEs [22].

Targeted microorganisms or antibiotic resistances	Targeted genes	Sequence of primers and probes
Total bacteria	16S rRNA	Forward:GACARCCATGCASCACCTG Reverse:GGTAGTCYAYGCMSTAAACG Probe: FAM/TKC GCG TTG/ZEN/CDT CGA ATT AAW CCA C/3IABkFQ
<i>Enterococcus</i>	16S rRNA	Forward : CCCTTATTGTTAGTTGCCATCATT Reverse : ACTCGTTGTACTTCCCATTGT
<i>E. coli</i>	16S rRNA	Forward : GTTAATACCTTTGCTCATTGA Reverse : ACCAGGTATCTAATCTCTGTT
<i>Archaea</i>	16S rRNA	Forward : CCGACGTGAGRGRYGAA Reverse : YCCGGCGTTGAMTCCAAT T
Phage <i>vB_AviM_AVP</i> of <i>Aerococcus viridans</i>	DNA polymerase	Forward : CTACACAGACATGGGYGGATATG Reverse : CTACACAGACATGGGYGGATATG Probe: TGATGCCTTAGAGGACTACAAGAAGA
Aminoglycoside resistance	aac(6')-II (8)	Forward : CGACCCGACTCCGAACAA Reverse: GCACGAATCCTGCCTTCTCA
Aminoglycoside resistance	aac(6')-Ib (95)	Forward: CGTCGCCGAGCAACTTG Reverse: CGGTACCTTGCCTCTCAAACC
Aminoglycoside resistance	aac(3)-iid_iii_iif_iiia_iiie (410)	Forward: CGATGGTCGCGGTTGGTC Reverse: TCGGCGTAGTGCAATGCG
Beta-Lactamase resistance	blaCMY2 (108)	Forward: AAAGCCTCATGGGTGCATAAA Reverse: ATAGCTTTTGTTCAGCATCA
Beta-Lactamase resistance	blaGES (120)	Forward: GCAATGTGCTCAACGTTCAAG Reverse: GTGCCTGAGTCAATCTTTCAAAG
Beta-Lactamase resistance	blaVEB (38)	Forward: CCCGATGCAAAGCGTTATG Reverse: GAAAGATTCCTTTATCTATCTCAGACAA
Beta-Lactamase resistance	blaTEM (164)	Forward: AGCATCTTACGGATGGCATGA Reverse: TCCTCCGATCGTTGTGAGAAGT
Beta-Lactamase resistance	blaVIM (147)	Forward: GCACTTCTCGCGGAGATTG Reverse : CGACGGTGATGCGTACGTT
Beta-Lactamase resistance	blaIMP (324)	Forward: GGAATAGAGTGGCTTAATTC Reverse: GGTTAACAAAAACAACCACC
Beta-Lactamase resistance	blaMOX (34)	Forward: CTATGTCAATGTGCCGAAGCA Reverse: GGCTTGTCTCTTTTCAATAGC
Beta-Lactamase resistance	blaSHVII (1110)	Forward: TTGACCGCTGGGAAACGG Reverse: TCCGGTCTTATCGGCGATAAAC

Beta-Lactamaase resistance	blaOXA (1506)	Forward:CGCAATTATCGGCCTA GAAACT Reverse: TTGGCTTTCCGTCCCATT
Beta-Lactamaase resistance	blaCTX-M-1	Forward: CGG GCR ATG GCG CAR AC Reverse : TGC RCC GGT SGT ATT GCC Probe: FAM/CCA RCG GGC/ZEN/GCA GYT GGT GAC/3IABkFQ
Erythromycine resistance	ermB (804)	Forward: GAACACTAGGGTTGTTCTTGCA Reverse: CTGGAACATCTGTGGTATGGC
Erythromycine resistance	ermF (23)	Forward: CAGCTTTGGTTGAACATTTACGAA Reverse: AAATTCCTAAAATCACAACCGACAA
Erythromycine resistance	ermT (137)	Forward:GTTCACTAGCACTATTTTAATGACA GAAGT Reverse: GAAGGGTGTCTTTTAATACAATTAACGA
Erythromycine resistance	ermX (209)	Forward:GCTCAGTGGTCCC CATGGT Reverse: ATCCCCCGTCAACGTT
Erythromycine resistance	erm35 (815)	Forward: CCTTCAGTCAGAACCGGCAA Reverse: GCTGATTGACAGTTGGTGGTG
Tetracycline resistance	tet32 (54)	Forward: CCATTACTTCGGACAACGGTAGA Reverse: CAATCTCTGTGAGGCATTTAACA
Tetracycline resistance	tetA (180)	Forward: CTCACCAGCTGACCTCGAT Reverse: CACGTTGTTATAGAAGCCGCATAG
Tetracycline resistance	tetL (195)	Forward:ATGGTTGTAGTTGCGCG CTATAT Reverse: ATCGCTGGACCGACTCCTT
Tetracycline resistance	tetO (192)	Forward :CAACATTAACGGAAAGTTTATTG TATACCA Reverse: TTGACGCTCCAAATTCATTGTATC
Tetracycline resistance	tetQ (185)	Forward:CGCCTCAGAAGTAAGTTCATAC ACTAAG Reverse: TCGTTTCATGCGGATATTATCAGAAT
Tetracycline resistance	tetS (200)	Forward:TTAAGGACAAACTTCTGAC GACATC Reverse: TGTCTCCCATTTGTTCTGGTTCA
Tetracycline resistance	tetW (191)	Forward: ATGAACATTCCCACCGTTATCTTT Reverse: ATATCGGCGGAGAGCTTATCC
Tetracycline resistance	tetX (196)	Forward: AAATTTGTTACCGACCGGAAGTT Reverse: CATAGCTGAAAAATCCAGGACAGTT
Tetracycline resistance	tetM (1513)	Forward: GGAGCGATTACAGAATTAGGAAG C Reverse: TCCATATGTCCTGGCGTGTG
Vancomycin resistance	vanA (1514)	Forward: GGGCTGTGAGGTCGGTTG Reverse: TTCAGTACAATGCGGCCGTTA
Vancomycin resistance	vanB (211)	Forward: TTGTCGGCGAAGTGGATCA Reverse: AGCCTTTTCCGGCTCGTT
Vancomycin resistance	vanRA (216)	Forward: CCCTTACTCCCACCGAGTTTT Reverse: TTCGTCGCCCCATATCTCAT
Vancomycin resistance	vanSA (218)	Forward: CGCGTCATGCTTTCAAAATTC Reverse: TCCGCAGAAAGCTCAATTGT
Sulfonamide resistance	sul1 (363)	Forward: GCCGATGAGATCAGACGTATT G Reverse: CGCATAGCGCTGGGTTTC

Sulfonamide resistance	sul2 (133)	Forward: TCATCTGCCAAACTCGTCGTTA Reverse: GTCAAAGAACGCCGCAATGT
Quinolone resistance	qnrB (1202)	Forward: TCACCACCCGCACCTG Reverse: GGATATCTAAATCGCCCAGTTCC
Colistine resistance	mcr-1	Forward: ATGGCAGGTCTATGATA Reverse: CGGATAATCCACCTTAACA Probe: FAM/CTA CAG ACC/ZEN/GAC CAA GCC GA/3IABkFQ/-3'
Mobile Genetic Element resistance	is26 (1546)	Forward: ATGGATGAAACCTACGTGAAGGTC Reverse: CGGTACTTAATCTGTCCGGTGTCA
Mobile Genetic Element resistance	tnpA (207)	Forward: AATTGATCGGCACGGCTTAA Reverse: TCACCAAACCTGTTATGGAGTCGTT
Mobile Genetic Element resistance	int1-A (336)	Forward: CGAAGTCGAGGCATTTCTGTG Reverse: GCCTTCCAGAAAACCGAGGA

Table S4. Insert sequences used to construct standard curves that was used for qPCR detection of targeted ARGs and MGEs

Genes	Insert sequences
aac(6')-II	ATCTTCTCCGCACGAATCCTGCCTTCTCATAGCAGCGTATGGCTCGATGGTTGTTCGGAGTCGGGTCGGTCTGAATCT
aac(6')-Ib	TTAGGCATCACTGCGTGTTCGCTCGAATGCCTGGCGTGTGTTGAACCATGTACACGGCTGGACCATATGGGGTGGTTA
aac(3)-iid_iii_iif_iaa_iee	CTAACCTGAAGGCTCGCAAGAGCGCTCGACGGCTCTGTCGGAGGCACGATCGGAGTGGTTCGAAATGCTTCTCAA GATAGGTGACGCCGACGTCACGATGTCCTGCGCGTCGAACAGGTAGCACTGAGCAAGCCACGACACCTTCTCGAT GGCGACCGAGCTTACGTAAGCATTGCTATAGTTTCAACCGCATCCGGCTTTCCTTCGATAGCAAGCAATCGAGAATG CCGTTTGAATCGTAATCCGATGCCGTTTCCAGGCGACTTACCCTCTCTTCCAAGCATCGGCATCTACATCGTCAACCCAC CGTTTGTGGGGATATCGGCAACCGCC
blaCMY2	CCCCGAGTGAAAGCCTCATGGGTGCATAAAAACGGGCTCCACTGGTGGATTGGCAGTACGTAGCCTTCGTTCCAGAAAA AACCTTGGCATCGTGATGCTGGCAACAAAGCTATCCTAACCTG
blaGES	GCAGCGTTTTGCAATGTGCTCAACGTTCAAGTTTCCGCTAGCCGCGCTGGTCTTTGAAAGAATTGACTCAGGCACCGAGCGGGG
blaVEB	ACTTCCATTTCCCGATGCAAAAGCGTTATGAAATTTCGATTGCTTTAGCCGTTTGTCTGAGATAGATAAAGGGAATCTTCTTTTGAACAA
blaTEM	GTCACAGAAAAGCATCTTACGGATGGCATGACAGTAAGAGAATTATGCACTGCTGCCATAACCATGAGTGATAACACTGCTGCCAATTAC TTCTGACAACGATCGGAGGACCGAAGGAGC
blaVIM	ATGTTCAAACCTTTTGAAGTAAGTTATTGGTCTATTGACCGCTCTATCATGGCTATTGGCAGTCCGCTCGCTTTTCCGTAAGATTCTAGCGGTG AGTATCCGACAGTCAGCGAAATTCGGTTCGGGGAGGTCCGGCTTACCAGATTGCCGATGGTGTGTTGGTTCGATATCGCAACGCACTGCTTT GATGGCGCAGTCTACCCGTCGAATGGTCTCATGTGCGGTGATGGTGATGAGTTGCTTTTGAATTGATACAGCGTGGGTGCGAAAAACACA GCG
blaIMP	ATGAGCAAGTTATCTGTATTCTTTATATTTTGTGTTTGTAGCATTGCTACCGCAGCAGAGCCTTGGCAGATTTAAAAATTGAAAACTTGAT GAAGGGGTTTATGTTCTACTACTGTTTGAAGAAGTTAACGGGTGGGGCGTTTTCTTAACATGGTTTGGTTGTTCTGTAGATGCTGAAGCTTA TCT AATTGACACTCCATTTACGGCTAAAGATACTGAAAGTTAGTCACTTGGTTTGTGGAACGTGGCTATAAAATAAAGGCAGTATTTCTCTC ATTT TCATAGTGACAGCACGGGC
blaMOX	ATGCAACAACGCAATCCATCCTGTGGGGCGCTCTGGCCACCCTGATGTGGCCGGTCTGGCCCATGCAGGTGAGACTTACCGGTGATCCC CTGCGCCCCGTGGTGGATGCCAGCATCCGGCCGTGCTCAAGGAGCACAGGATCCCGGGCATGGCGGTGGCCGTGCTCAAGGATGGCAAGGC CCACTATTCAACTACGGTGTGGCCGATCGGGAGCGCGCAGTCGGTGTGACGAGCAGACCTGTTTCGAGATAGGCTCCGTGAGCAAGCCCT GACCGCGACCTAGGAGCCTATGCGGTGGTCAAGGGAGCGATGCAACTGGATGACAAGGCGAGCCGGCAGCCCCCTGGCTCAAGGGATCCG CCTTTGACAGCATCACCATGGGGGAGTGGCTACCTACAGCGCGGGCGGCTTGCCTGCAATTCCCGAGGAGGTGGATTGCTCGAGAAGA TGCAGGCTACTACCGCAGTGGACCCAGCCTACTCGCGGGTTCCATCGCCAGTACTTAACCCAGCATAGGGCTTTCGGCCAC CTGG CGCGGAGCAGCATGAAGCAGCCGTTTGGCCAGTGTAGTGAGCAGACGCTCTGCGGGGCTTGGCTGCACCCAC
blaSHVII	TTAGCGTTGCCAGTGTGATCAGCGCCGCGCGATCCCGCGATTGCTGATTTCGCTCGGCCATGCTCGCCGGCGTATCCCGCAGA TAAATCACCACAATGCGCTCTGCTTTGTTATTTCGGGCCAAGCAGGGCGACAATCCCGCGCGCACCCCGCTTGCTAGC-TCCGGTCTTATC GGGATATAAC-CAGCCCGCGGACGACGAGCGGATCAACGGTCCGGCGACCCGATCGTCCACCATCCACTGCAGCAGTCCGCTTG CGAACGGGCGCTCAGACGCTGGCTGGTACGACGCTTGGCAGGGTCCGCGCCATGCTGGCCGGGGTAGTGGTGTGCGGGCGCTCGC CGGGAAGCGCTCATTCAGTTCCGTTTCCAGCGGTCAAGGCGGGTGACGTTGTGCCGATCTGGCGCAAAAGGCAGTCAATCCTGGC GGGCGGCCGAGCGTGGCCAGCAGCAGATTGGCGGCGCTGTTATCGCTCATGGTAATGGCGGGCGCGCAGAGTTCCGCGACCGTCAATGC CGTCGGCAAGGTGTTTTTCGCTGACCGGGAGTAGTCCACAGATCCTGCTGGCGATAGTGATCTTTTCGCTCAGCTGTTCGTCAACCG CATCCACCCGCGCAGCACTGCGCGCAGAGCACTACTTAAAGGTGCTCATCATGGGAAAGCGTTCATCGGCGCGCAGGCGGTCAAGC GTGCGGCGCTGGCCAGATCCATTCTATCATGCTACCGCGCCGACAGCTGGCTTTCGCTTTGTTTAAATTGCTCAAGCGGCTGCGGG CTGGCGTGTACCGCCAGCGCGAGGTGGCTAACAGGGAGATAATACAGCGCAATATAACGCAT
blaOXA	ATGAACATTAAAGCACTTACTTATAACAAGCGCTATTTTATTTACGCTGCTCACCTTATATAGTGACTGCTAATCCAAATCACAGCGC TTCAAATCTGATGTAAAGCAGAGAAAAATTAAATAATTTTAAACGAAGCACACACTACGGGTGTTTTAGTTATCCAAACAGGCCAAATC AACAAGCATGGTAATGATCTTGTCTGTGCT

blaCTX-M-1	TGCACCGGTGGTATTGCCTTTCATCCATGTCAACAGCTGCGCCCGTTGGCTGTCGCCAATGCTTTACCCAGCGTCAGATTCCGCAGAGT TTGCCCAATTGCCCG
ermB	ATGAACAAAAATATAAAATATTCTCAAAACCTTTTAAACGAGTGAAAAAGTACTCAACCAAATAATAAAACAATTGAATTTAAAGAAACCGATA
	CCGTTTACGAAATTTGGAACAGGTAAGGGGCATTAAACGACGAAACTGGCTAAAATAAGTAAACAGGTAACGTTCTATTGAATTAGACAGTCAT CTATTCAACTTATCGTCAGAAAAATTAACACTGAATACTCGTGTCACCTTAAATTCACCAAGATATTCTACAGTTTCAATTCCCTAACAAACAGA GGTATAAAATTGTTGGGAATATTCTTACCATTAAAGCACACAAATTATTAATAAAGTGGTTTTGAAAGCCATGCGCTGACATCTATCTGA TTGTTGAAGAAGGATTCTACAAGCGTACCTTGGATATTACC
ermF	TGAAAACGACACAGCTTTGGTTGAACATTTCAGAAAATATTCTCTGATGCCGAAATGTTCAAGTTGTCGGTTGTGATTTTAGGAATTTTGC AGTTCCG
ermT	AATACAAATCGTTCACTAGCACTATTTTAAATGACAGAAGTTGATATATCCATATTAAGTAAATCCCTAGAGAATACTTTTATCCAAAACTTA G AGTTAATAGCTCGTTAATTGTATTAAAAAGACACCCCTTCAAAAATATCA
ermX	GATGATGACGGCTCAGTGGTCCCATGGTTTCACATTTTACCTGGGTTCTCGGGTACCAAGGTCTGCTTTCCGGCCACAGCCAAACGTTGA CG GGGGGATCTTAGTGATC
erm35	GAATTCAGTATTCACTGTAAGAAGTAAATAATGACAAAAAGAAATGGCCGTCGTTTACGGGTCAGCACTTTACTATTGACAAAGTGCTT ATTAAAGATGCAATAAAAGAATCAAAATATAAATCAACACGATACAGTTTAGATATTGGAGCTGGTAAGGGTTTTCTAACTGTTCATCTCTAAA A AATGTCGATAAAGTTATTGCCATTGAAACGATGTTGCATTAAAGTCAACATTTGCGCAAAAAATTCATTACGCTCAAAAACGTTCAAGTGGTTAG TTGTGATTATAGAAATTTTGGTTCCGAAAGTTCCATTAAAGTAGTTTCAAAATATTCCTTTTGGTATTACATCTGATATTTTTAGTAGTCTG ATG TTTGAATAATGTCGAATATTTTCTATGCGGTTCATTAAT
tet32	TAATCCCATGCCATTACTTCGGACAACCGTAGAGCCGCAAAAGCCGGAGCAAAAGGGAAGCCCTGTTAAATGCCCTCACAGAGATTGCTGA TACAGA
tetA	ACTGGCGCGCTCACAGCCTGACCTCGATCGTCGGACCCCTCCTCTTCACGGCGATCTATGCGGCTTCTATAACAACGTGGAACGGGT GG
tetL	AGCACTCGTAATGGTTGTAGTTGCGCGCTATATCCAAAGGAAAAATAGGGTAAAGCATTGGTCTTATTGGATCGATAGATAGCCATGGGA GAA GGAGTCGGTCCAGCGATTGGTGGAAAG
tetO	GCAGGAAAGACAACATTAACGGGAAAGTTTATTGTATACCAAGTGGTGAATTCGAGAAGTACAGGAGCGTAGATGAAGGCACAACAAGGACA GAT ACAATGAATTTGGAGCGTCAAAGGGGAATCA
tetQ	GTAAGACTACGCTCAGAAAGTAAAGTTACATACTAAGGGCTTAGGGCTTTTATGGTCAAGCCATGCGGGTATCAAAATAACAAAAGGCGA TTA TTCTGATAATATCCGATGAACGAAAAAGATAAA
tetS	TATCAAGATATTAAGGACAACCTTTCTGACGACATCATAAATTAAGCAGACTGTGAATCTAAATTTGAAACCTTATGTAAATAGATTACTGAA CCA GAACAATGGGAGACAGTAATTGTGG
tetW	CCCTGCGGAAAAATGAACATTTCCACCGTTATCTTTATCAACAAGATCGACCAAGTTGGCGTTGATTGTCAGGGCGGTATCAGTCTGTTCG GG ATAAGCTCTCCGCGATATTATCATCAAG
tetX	GTTTCATTGTCATAGCTGAAAAAATCCAGGACAGTTTATCTCTGTTGATGAATATCGGCTTGATATITGAAAGTACCTGTTTCTTCAACTTC CG TGTCGGTAACAAATTTTCTTACCTTG
tetM	ATGGAGGAAAAATCACATGAAAATTATTAATATTGGAGTTTAGCTCATGTTGATGCGGGAAAAACTACCTTAACAGAAAGCTTATTATATAAC AGT
vanA	TCACCCCTTTAACGCTAATACGATCAAGCGGTCAATCAGTTTCGGGAAGTGCAATACCTGCAGCGCCATCATACGGGGATAACGACTGTA TGA CGTGAAACCGGGCAGAGTATTGACTTCG
vanB	GATGATTGTGATGTCGGCGAAGTGGATCAAATCCGGTTAGGCCACGGTATCTTCCGATCCATCAGGAAAACGAGCCGAAAAAGGCTCA GAGAATGC
vanRA	TGCTGAAATATTCTGTCGCCCATATCTCATGAAATAGCAGCTCGGAGCTAACCACATTCCTTGTTCACAGAGGATTGCGAGTATTGAA AAC TCGGTGGGAGTAAGGGATAACTGCT
vanSA	TATTCTATGTCGCGTATGCTTTCAAAATTCGCAAAATACTTTGACGAGATAAATACCGGATGATGTACTTATTACAGAACGAAGATAAACA AA TTGAGCTTTCTGCGGAAATGGATGTT
sul1	ATGGTGACGGTGTTCGGCATTCTGAATCTCACCGAGGACTCTTCTTCATGATGAGAGCCGGCGGCTAGACCCCGCCGGCTGTCAACCGC G CGCATCGAAATGCTGCGAGTCGGATCAGACGTCGTGGATGTCGGACCGCGCCGACGCAATCCGGACGCGAGGCTGTATCGCCG
sul2	ATGAATAAATCGCTCATCATTTTCGGCATCGTCAACATAACCTCGGACAGTTTCTCCGATGGAGGCCGATCTGGCGCCAGACGACGCC ATTGCGCAGGCGCTAAGCTGATGGCCGAGGGGCGAGATGTGATCGACCTCGGTCCGGCATCCAGCAATCCCGACGCCGCGCTGTTT CGTCCGACACAGAAATCGCGCTATCGCGCCGGTGTGGACGCGCTCAAGGCAGATGGCATTCCTGCTCGCTCGACAGTTATCAACCC GCGACCAAGCCTATGCTTGTGCGGTGGTGTGGCCTATCTCAATGATATTCGCGGTTTTCAGACGCTGCGTTCTATCCGCAATTGGCG AAA
qnrB	TTAACCCATGACAGCGATACCAAGACGTTCCAGGAGCAACGATGCCTGGTAGCTGTCCAGTTTGACGCTTGCAAAATCAACCCCGC
mcr-1	ATGGCACGGTCTATGATACGACCATGCTCCTCAAAATGCCCTACAGACCGACCAAGCCGAGACCAAGGATCTATTAACGCGAGCGTTTATCA TGCATATCATTTGGTTTGGGTGTGCTACCAAGTTTGGCTTTTGTAAAGGTGGATTATCCG
is26	TTACATTTTAAAAACTCTGCTTACCAGGCGCATTTTCGCCAGGGGATCACCATAATAAAATGCTGAGGCTTGGCTTTGCGTAGTG CACGCATCACTCAATACCTTTGATGGTGGCGTAAGCCGCTCTCATGGAATTAATCCAGCGTGGCGCGGATTATCCGTTTCAGT TTGCCATGATCGCATTCATCAACGTTGTTT
tnpA	TTTATCTGCAATTTGATGCGGACGGCTTAACCTTAGATATCTGGTTACGAAAGAAATGGGATACGCAAGCAGCCTATGCTTTCTTAA AACGACTCCATAAACAGTTTGGTGAGCGGAAAGCA
int1-A	ATGAAACCCGCACTGGCCGTTACCACCGCTGCGTTCCGTTCAAGGTTCTGGACCAAGTTGCGTGAGCGCATACGCTACTTGCATT ACAGCTTACGAACCGAACAGGCTTATGTCCACTGGGTTCTGCTCTTACCGTTTCCACGGTGTGCGTACCCGGCAACCTTGGGCAGCA G

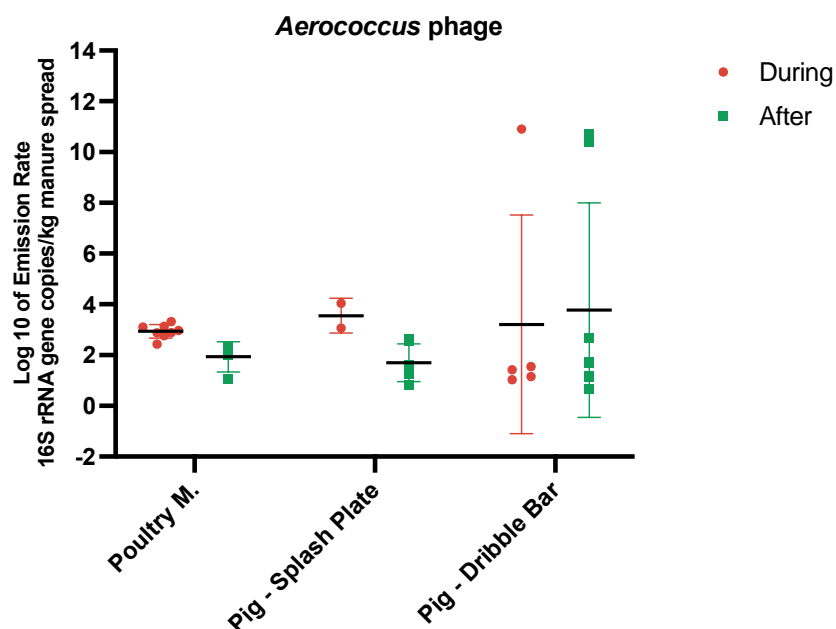


Figure S3. Emission rate of the phage *vB_AviM_AVP* of *Aerococcus viridans* for tested manure types; No difference was observed among the paired groups regarding emission rates of the phage.

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