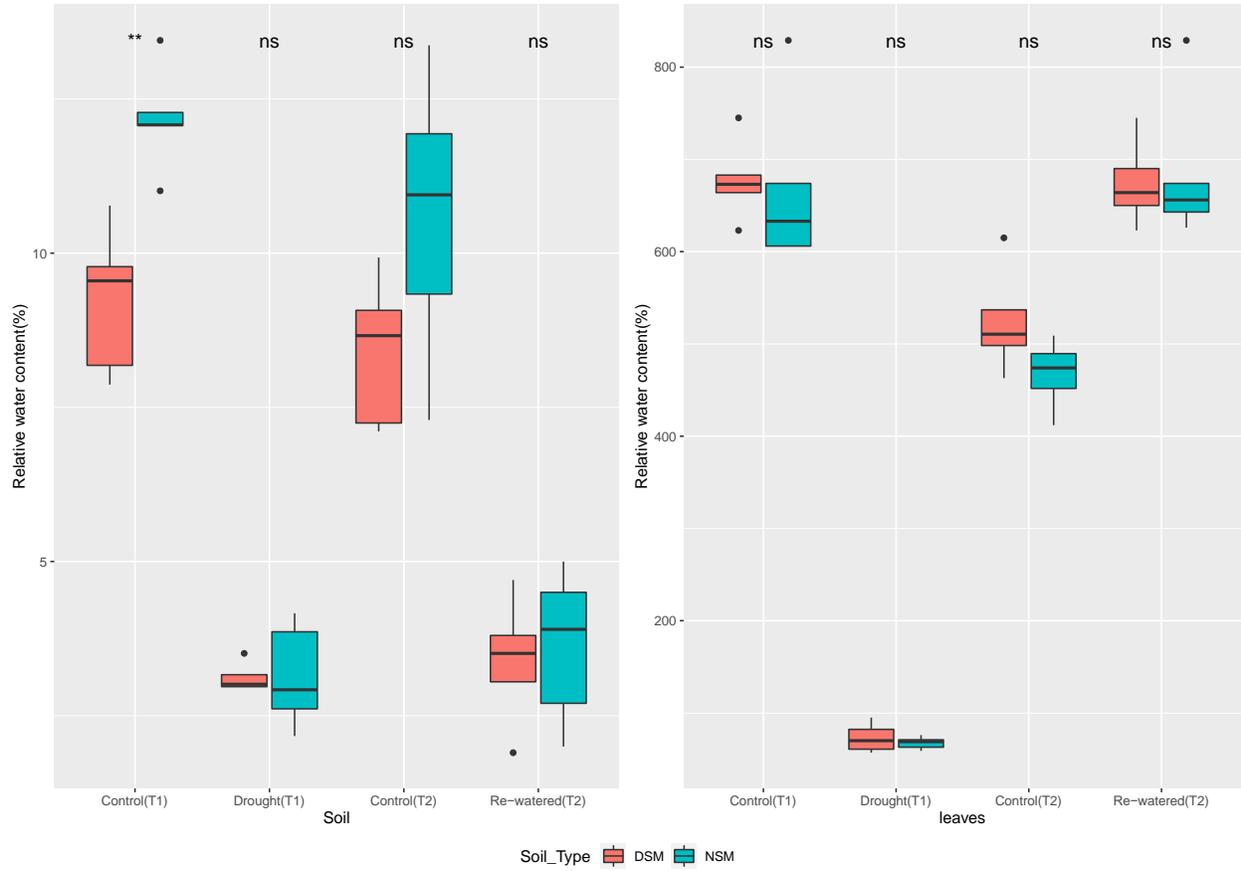
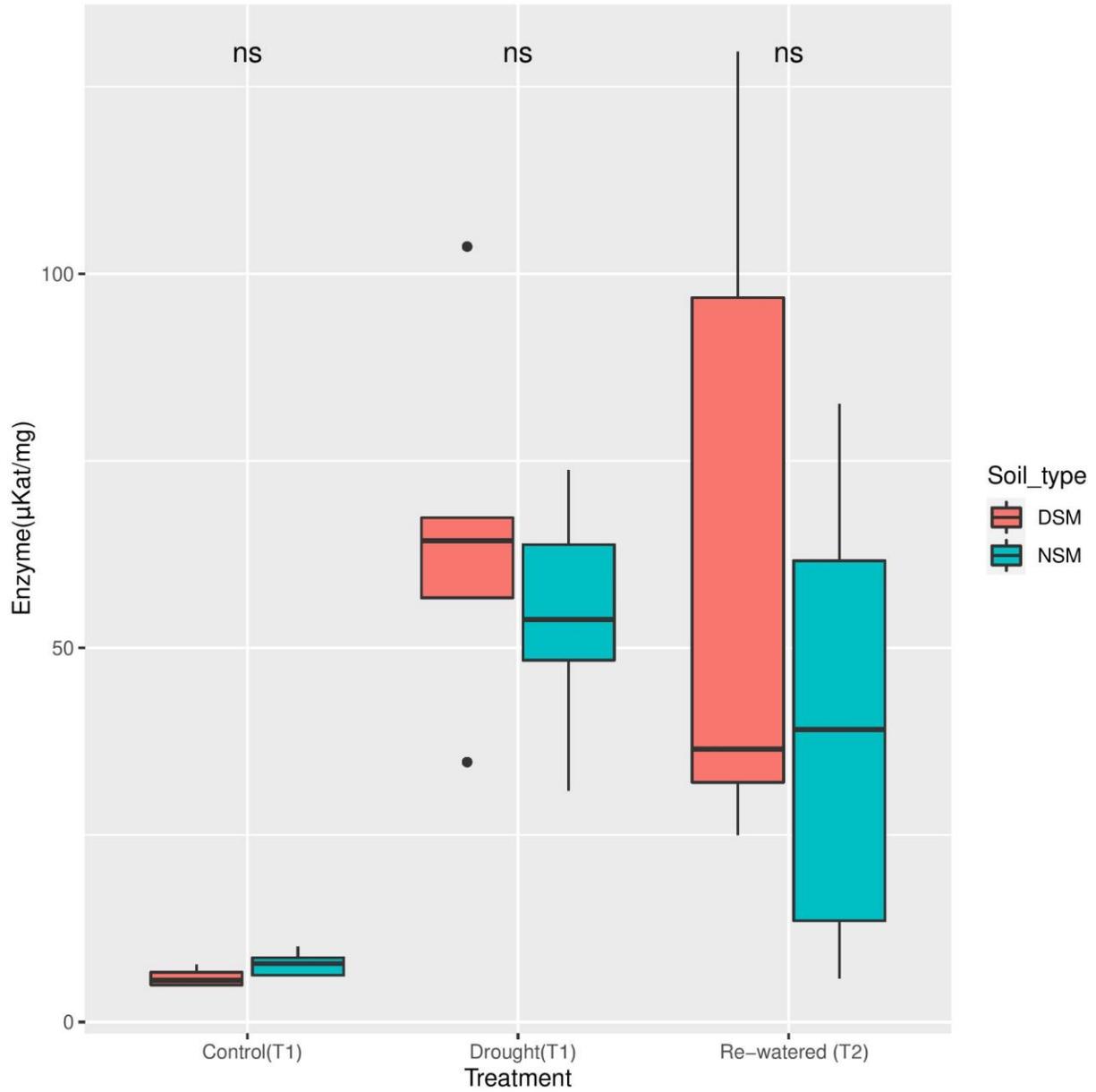


Supplementary table 1: 16S rRNA and ITS primers and PCR conditions

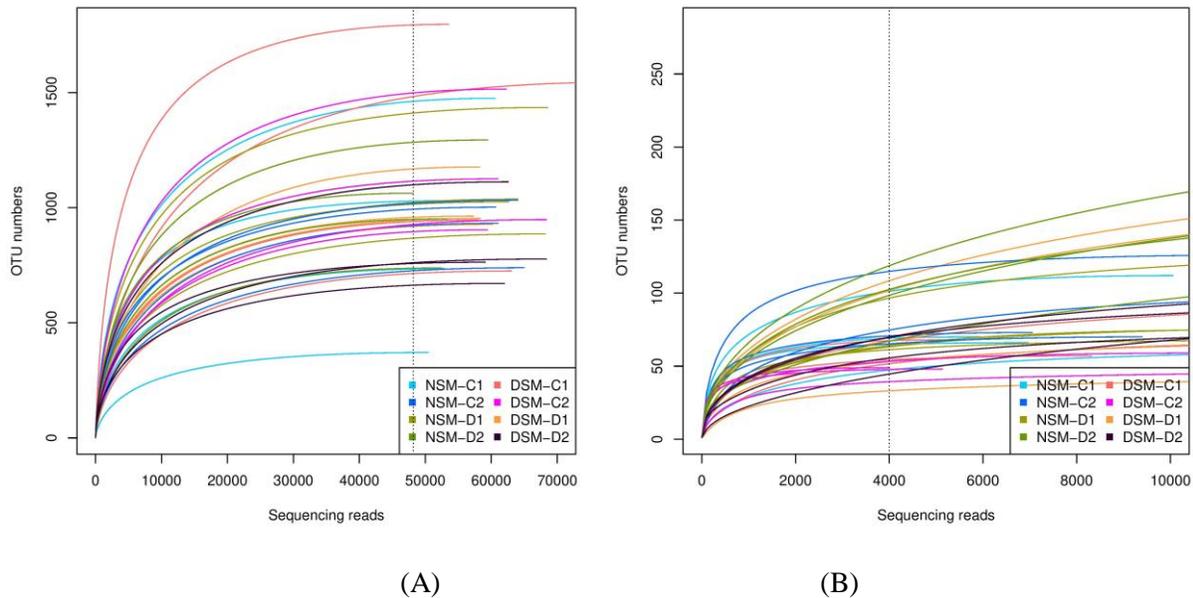
| rRNA primer set | seq 5'->3' | | |
|--|---|---------------------|----------|
| 335f | CADACTCCTACGGGAGGC | | |
| 769r | ATCCTGTTGMTMCCCVRC | | |
| Reference | | | |
| Dorn et al 2015 | | | |
| Reagent | Stock concentration | Final concentration | µl /tube |
| H2O | | | |
| NEB Next High Fidelity Master Mix | 2x | 1x | 12.5 |
| 335f | 10 pmol.µl-1 | 5 pmol | 0.5 |
| 769r | 10 pmol.µl-1 | 5 pmol | 0.5 |
| BSA | 15 µg.µl-1 | | 2.5 |
| sample | | | |
| Set | temperature (°C) | time | |
| hotstart | | 98 2 min | |
| denaturation | | 98 10 s | |
| annealing | | 60 30 s | |
| elongation | | 72 30 S | x 30 |
| | | 72 5 min | |
| ITS primers | seq 5'->3' (forward-reverse combinations) | | |
| ITS3-mix* | | | |
| ITS3-Mix1 (Fungi) | TCGTCGGCAGCGTCAGATGTGTATAAGAGACAG-CATCGATGAAGAACGCAG | | |
| ITS3-Mix2 (Chytridiomycota) | TCGTCGGCAGCGTCAGATGTGTATAAGAGACAG-CAACGATGAAGAACGCAG | | |
| ITS3-Mix3 (Sebacinales) | TCGTCGGCAGCGTCAGATGTGTATAAGAGACAG-CACCGATGAAGAACGCAG | | |
| ITS3-Mix4 (Glomeromycota) | TCGTCGGCAGCGTCAGATGTGTATAAGAGACAG-CATCGATGAAGAACGTAG | | |
| ITS3-Mix5 (Sordariales) | TCGTCGGCAGCGTCAGATGTGTATAAGAGACAG-CATCGATGAAGAACGTGG | | |
| ITS4-mix* | | | |
| ITS4-Mix1 (Fungi) | GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAG-TCCTCCGCTTATTGATATGC | | |
| ITS4-Mix2 (Chaetothyriales) | GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAG-TCCTGCGCTTATTGATATGC | | |
| ITS4-Mix3 (Archaeorhizomycetes) | GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAG-TCCTCGCCTTATTGATATGC | | |
| ITS4-Mix4 (Tulasnellaceae) | GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAG-TCCTCCGCTGAWTAATATGC | | |
| * All primers were diluted to 10 pmol and mixed in equimolar concentration | | | |
| Reference | | | |
| Tedesoo et al 2014 /2015 | | | |
| Reagent | Stock concentration | Final concentration | µl /tube |
| H2O | | | |
| NEB Next High Fidelity Master Mix | 2x | 1x | 12.5 |
| ITS3-mix | 10 pmol.µl-1 | 5 pmol | 0.5 |
| ITS4-mix | 10 pmol.µl-1 | 5 pmol | 0.5 |
| Tetramethylammonium | 15 µg.µl-1 | | 2.5 |
| sample | | | |
| Set | temperature (°C) | time | |
| hotstart | | 98 2 min | |
| denaturation | | 98 10 s | |
| annealing | | 61 30 s | |
| elongation | | 72 30 S | x 30 |
| | | 72 5 min | |



Supplementary Fig. S1 Relative water content of (A) soil and (B) leaves in DSM and NSM under different treatments at sampling time T1 and T2 (n=5). (ns: not significant, **: $p < 0.01$)

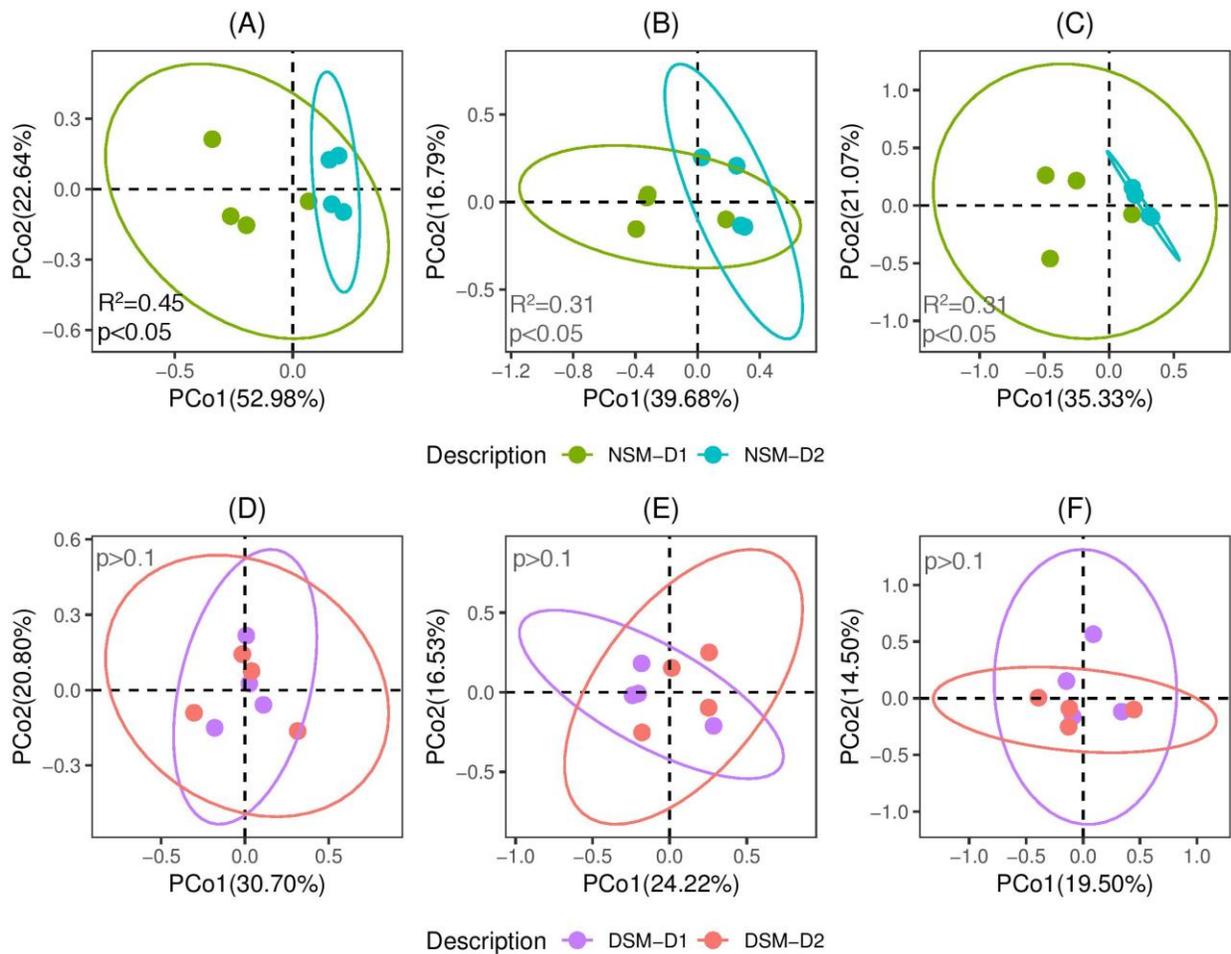


Supplementary Fig. S2 Peroxidase activity in leaves of barley plants grown in in DSM (soil with disturbed microbiome) and NSM (soil with natural microbiome) under control, drought and Re-watered conditions (n=5)

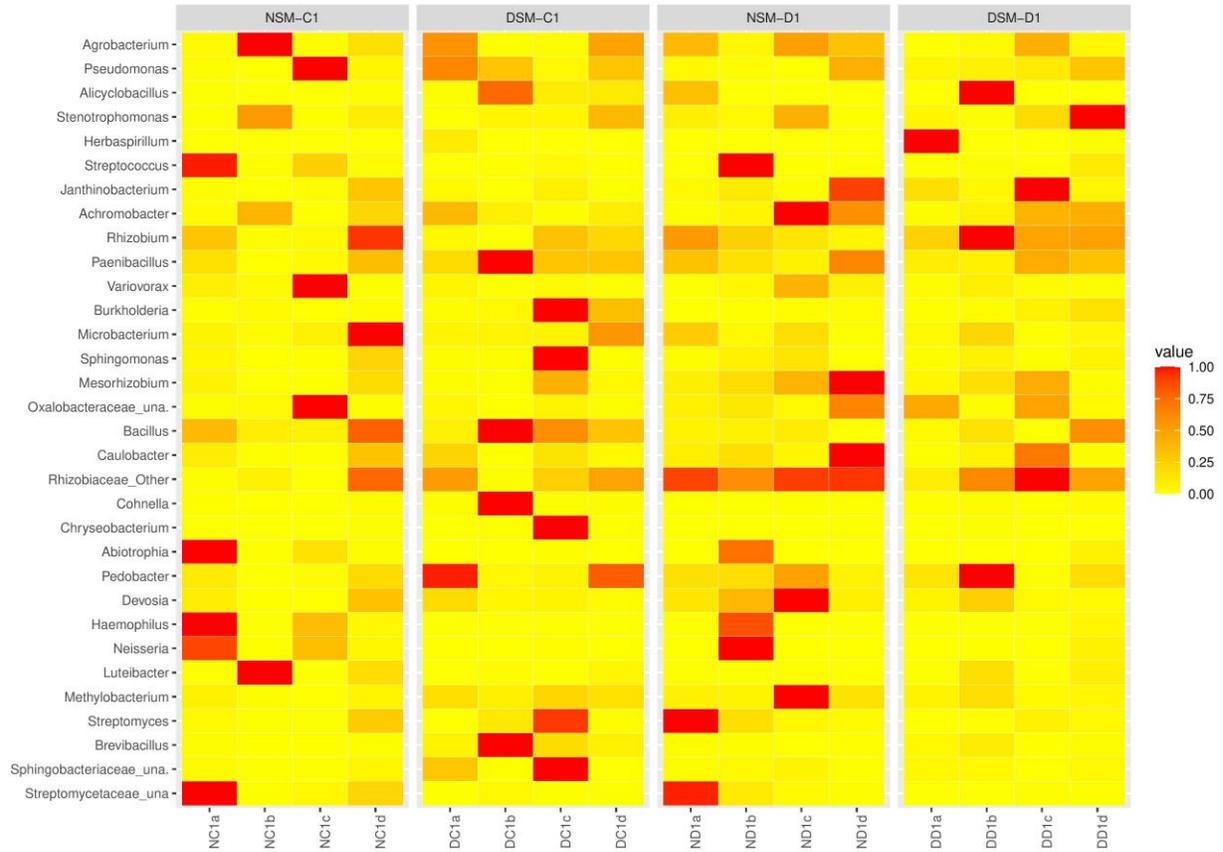


Supplementary Fig. S3 Rarefaction curves of root endophytic (A) bacteria and (B) fungi (n=4). The bacteria were rarefied at the sequencing depth of 48166 while the fungi were rarefied at the depth of 4002.

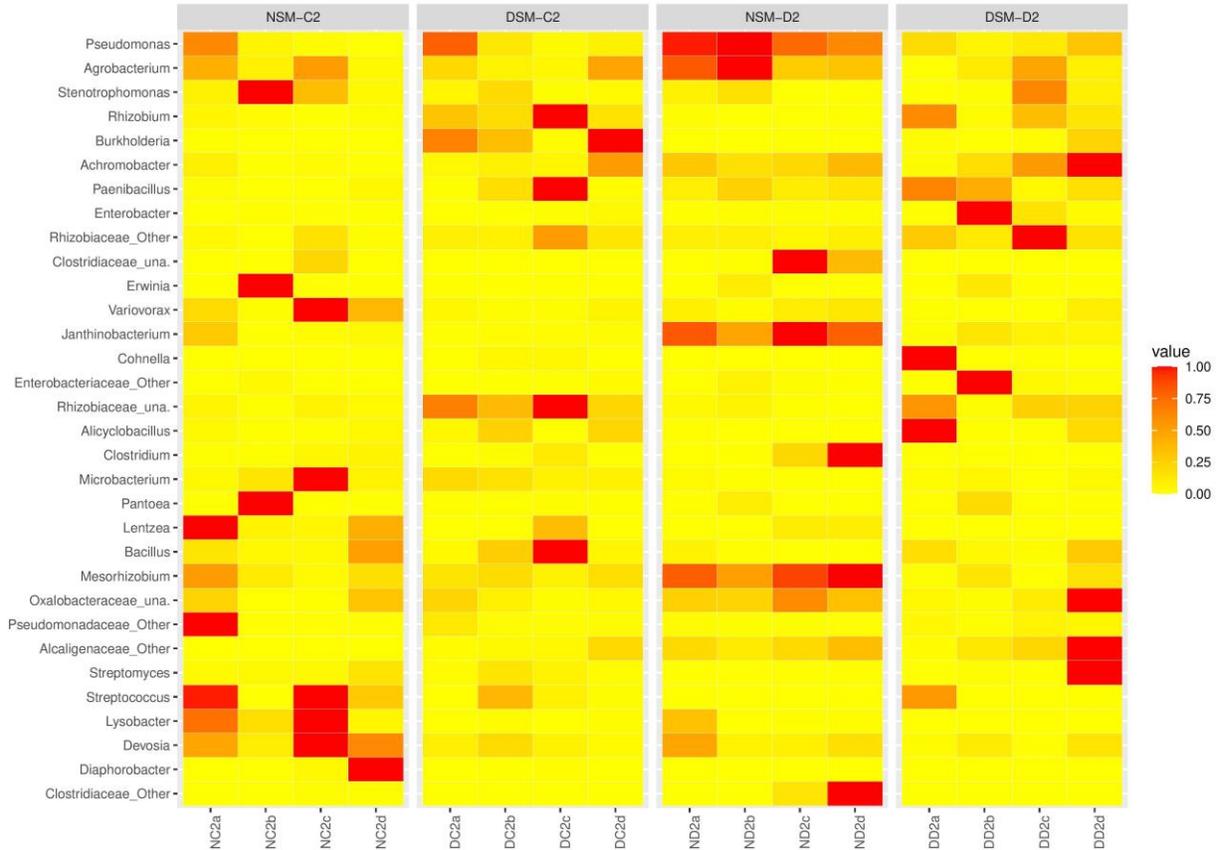
(NSM-C1:soil with natural microbiome under regular watering at first sampling;
 NSM-C2: soil with natural microbiome under regular watering at second sampling;
 NSM-D1: soil with natural microbiome after drought stress;
 NSM-D2: soil with natural microbiome drought stressed and re-watered;
 DSM-C1: soil with disturbed microbiome under regular watering at first sampling;
 DSM-C2: soil with disturbed microbiome under regular watering at second sampling;
 DSM-D1: soil with disturbed microbiome after drought stress;
 DSM-D2: soil with disturbed microbiome drought stressed and re-watered)



Supplementary Fig. S4 PCoA plots of root endophytic bacteria in NSM using (A) weighted (B) unweighted Unifrac distances and in DSM using (D) weighted and (E) unweighted Unifrac distances under drought and re-watered condition. PCoA plots of root endophytic fungi using Bray-Curtis dissimilarity in (C) NSM and (F) DSM under drought and re-watered condition ($n=4$). The ellipses represent 95% confidence interval of corresponding samples. (DSM-D1: soil with disturbed microbiome under drought stress; DSM-D2: soil with disturbed microbiome drought stressed and re-watered; NSM-D1: soil with natural microbiome under drought stress; NSM-D2: soil with natural microbiome drought stressed and re-watered)



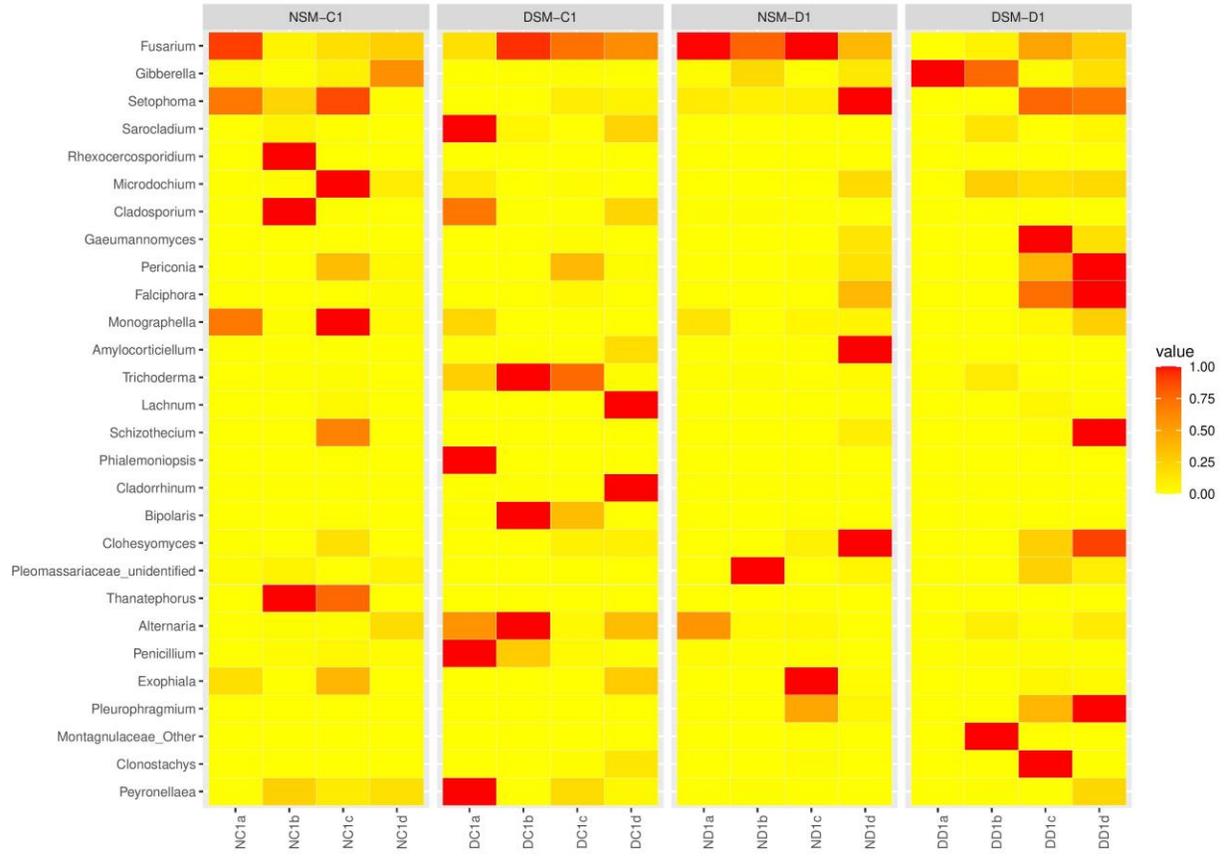
(A)



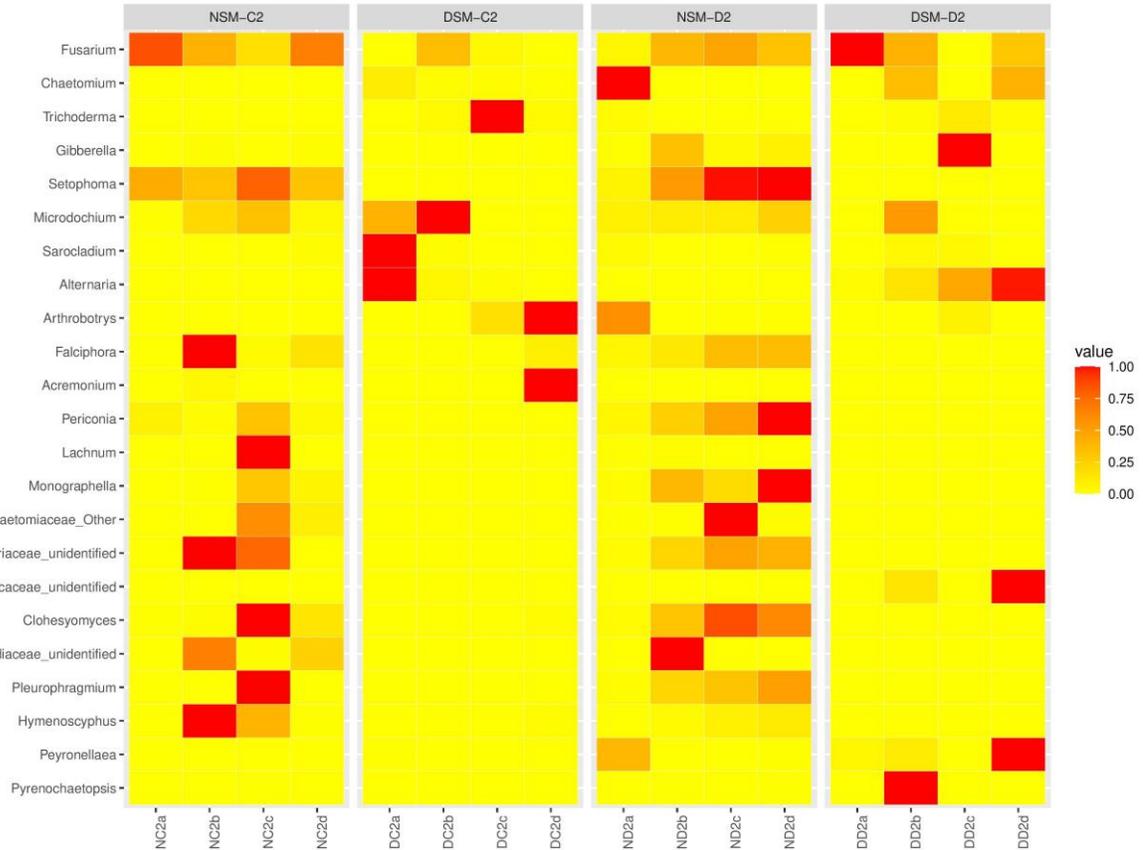
(B)

Supplementary Fig. S5 Major genera of bacterial endophytes in roots grown in NSM and DSM at (A) T1 and (B) T2 (n=4)

(NSM-C1: soil with natural microbiome under control conditions at the first sampling; NSM-D1: soil with natural microbiome under drought stress at the first sampling; DSM-C1: soil with disturbed microbiome under control conditions at the first sampling; DSM-D1: soil with disturbed microbiome under drought stress at the first sampling; NSM-C2: soil with natural microbiome under regular watering at second sampling; NSM-D2: soil with natural microbiome drought stressed and re-watered; DSM-C2: soil with disturbed microbiome under regular watering at second sampling; DSM-D2: soil with disturbed microbiome drought stressed and re-watered)



(A)



(B)

Supplementary Fig. S6 Major genera of fungal endophytes in roots grown in NSM and DSM at (A) T1 and (B) T2 (n=4)

(NSM-C1: soil with natural microbiome under control conditions at the first sampling; NSM-D1: soil with natural microbiome under drought stress at the first sampling; DSM-C1: soil with disturbed microbiome under control conditions at the first sampling; DSM-D1: soil with disturbed microbiome under drought stress at the first sampling; NSM-C2: soil with natural microbiome under regular watering at second sampling; NSM-D2: soil with natural microbiome drought stressed and re-watered; DSM-C2: soil with disturbed microbiome under regular watering at second sampling; DSM-D2: soil with disturbed microbiome drought stressed and re-watered)