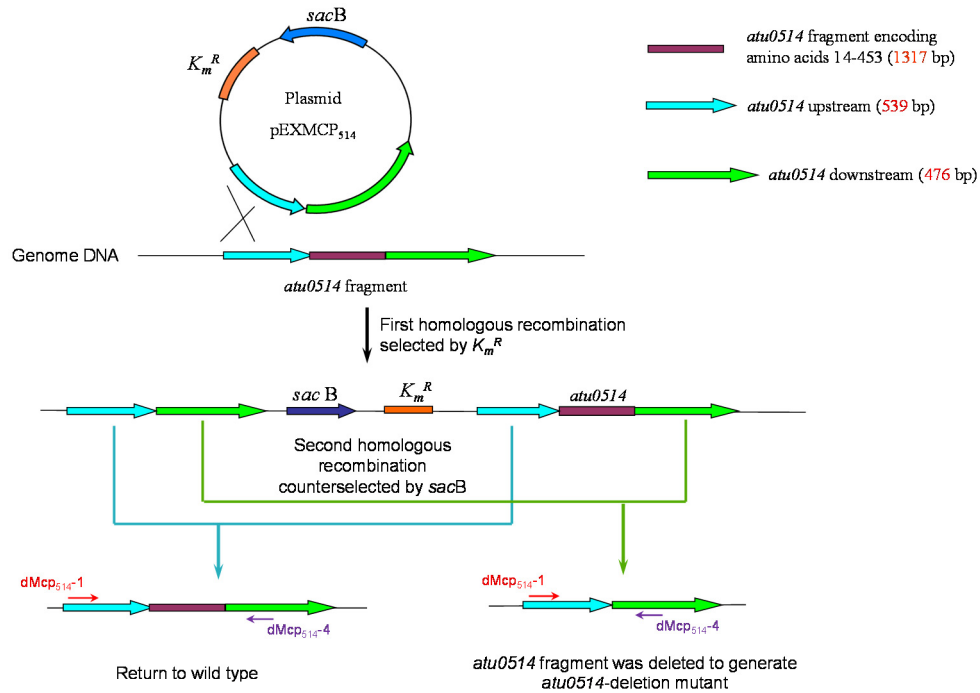
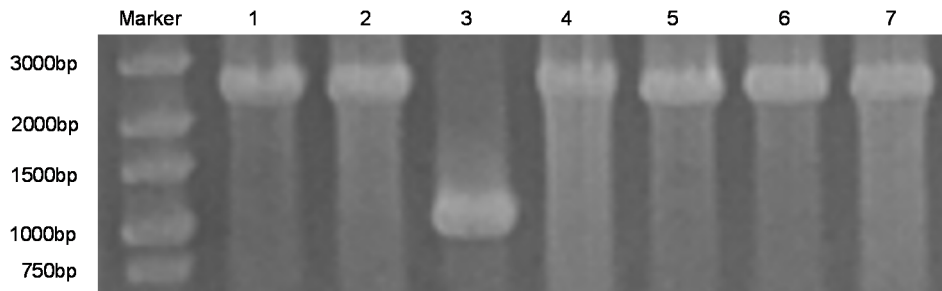


### A. Schematic diagram of gene replacement strategy (First cross occurs in upstream)



### B. Screening of *atu0514*-deletion mutant.



### C. Sequencing result of PCR fragment amplified from the mutant.

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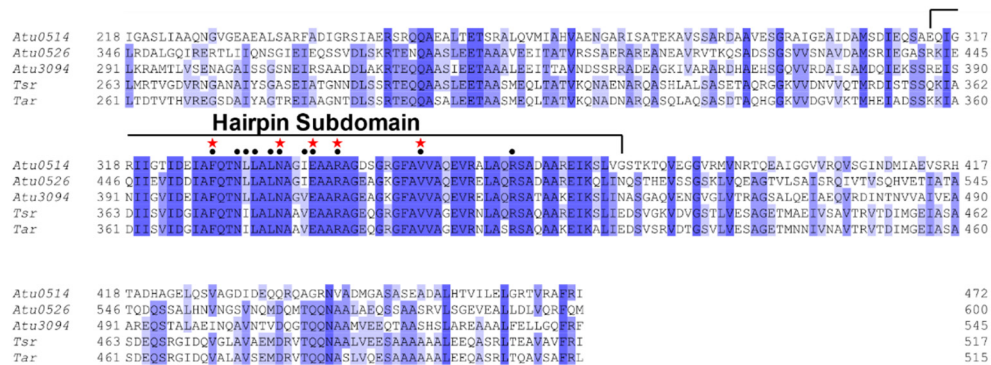
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C58 2201 CAGGTCTCATGGCCGCGCGAAATCCTGGGAAGAGGATGGCAAGCCCTTCGTTTCGCAAGGGTTCCGATGCATTGCACAAGACCATGAAACTGATAG 2300

A514 981 GCGTCGATATCGACCATTTGCTCCCTAAGGAGATG 1015
C58 2301 GCGTCGATATCGACCATTTGCTCCCTAAGGAGATG 2335

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**Figure S1.** Construction procedure, screening and verification of *atu0514*-deficient mutants. **(A)** Schematic diagram of gene replacement strategy. The suicide plasmid pEXMCP<sub>514</sub> carrying the upstream (light blue arrow) and downstream (green arrow) sequences of *atu0514* was introduced to the recipient cell. Because this plasmid can not be replicated in *A. fabrum*, only the transformants that the plasmid was integrated into the genome DNA at the locus of the target gene by homologous recombination can grow under the selection pressure of kanamycin. The K<sub>m</sub><sup>R</sup> (orange rectangles) represents kanamycin-resistant gene. The *sacB* (blue arrows) is a counterselectable marker gene (sucrose suicide gene) that confers sucrose sensitivity. A fragment of *atu0514* was shown in dark red rectangles. Colonies having undergone single cross-over homologous recombination (×) were screened as kanamycin resistance and sucrose sensitivity. Second cross-over homologous recombination event was selected by plating the single cross-over colonies on media containing 5% sucrose. The *atu0514*-deletion (or replacement) mutants were isolated from the kanamycin-sensitive and sucrose-resistant colonies. **(B)** Screening of *atu0514*-deletion mutants. The *atu0514*-deletion mutants were screened by PCR. Two oligonucleotides, dMcp<sub>514</sub>-1 (annealed to upstream of *atu0514*) and dMcp<sub>514</sub>-4 (annealed to downstream of *atu0514*), were used as the primers. PCR fragment amplified from wild type template is longer than that from mutant template. After agarose gel electrophoresis, only the PCR product in lane 3 was amplified from the *atu0514*-deletion mutant template. **(C)** Sequencing result of the PCR fragment amplified from the mutant. The red border rectangles indicate the rest fragment of *atu0514* and the black dot indicate the nucleotide shared by *atu0514* and *atu0515*.



**Figure S2.** Sequences of three *A. fabrum* MCPs and two *E. coli* MCPs were aligned by Jalview. The numbers in two sides indicate the position of amino acid. The black dots represent the key residues of trimer contact in *E. coli* MCPs[52]. The red stars indicate the residues of the single residue substitution for MCP<sub>514</sub>.

**Table S1.** Bacterial strains and plasmids used in this study

Bacterial strains	Relevant feature(s)	Reference/source
<i>Escherichia coli</i>		
E. coli DH5 $\alpha$	F <sup>-</sup> , $\phi$ 80dlacZ $\Delta$ M15, $\Delta$ (lacZYAargF)U169,deoR,recA1,endA1,hsdR17(rK <sup>-</sup> ,mK <sup>+</sup> ), phoA, supE44, $\lambda$ <sup>-</sup> , thi-1, gyrA96, relA1, for DNA cloning	Bethesda Research Laboratories
XL1 Blue	Report strain for Bacterial Two-hybrid System	Stratagene
<i>Agrobacterium fabrum</i>		
C58	Nopaline type strain; pTiC58, pAtC58	[63]
C58 $\Delta$ 514	C58 derivative deletional mutant in locus <i>atu0514</i>	This study
C58 $\Delta$ 514-C	C58 $\Delta$ mcp514 in which <i>cheW1</i> expression was restored by pCBMCP <sub>514</sub>	This study
C58 $\Delta$ w	C58 derivative deletional mutant in locus both <i>cheW1</i> and <i>cheW2</i>	[40]
C58 $\Delta$ a	C58 derivative deletional mutant in locus <i>chea</i>	[40]
$\Delta$ 514 $\Delta$ w	C58 derivative triple-deletion mutant in locus <i>cheW1</i> , <i>cheW2</i> , and <i>atu0514</i>	This study
<b>Plasmid</b>		
pEX18Km	Derivative of pEX18Tc in which Tc <sup>r</sup> was replaced by <i>nptIII</i> from pCB301; Km <sup>R</sup> , Sur <sup>S</sup>	[38]
pEXMCP <sub>514</sub>	pEX18Km, carrying a 1015bp fragment, which consisted of 539bp upstream fragment (from -500 to +39) and 476bp downstream fragment (from +1359 to +1835) of <i>atu0514</i>	This study
pUCA-19	pUC19 carrying an agrobacterial replicon; Ap <sup>R</sup> , Cr <sup>R</sup>	[38]
pUCA-GMCP <sub>514</sub>	pUCA-19, carrying an eGFP-MCP <sub>514</sub> expression cassette at HindIII and BamHI; Ap <sup>R</sup> , Cr <sup>R</sup>	This study
pUCA-GMCP <sub>514</sub> <sup>F328A</sup>	pUCA-19, carrying an eGFP-MCP <sub>514</sub> expression cassette with single residue-substitution F382A at HindIII and BamHI; Ap <sup>R</sup> , Cr <sup>R</sup>	This study
pUCA-GMCP <sub>514</sub> <sup>F328P</sup>	pUCA-19, carrying an eGFP-MCP <sub>514</sub> expression cassette with single residue-substitution F382P at HindIII and BamHI; Ap <sup>R</sup> , Cr <sup>R</sup>	This study
pUCA-GMCP <sub>514</sub> <sup>F328W</sup>	pUCA-19, carrying an eGFP-MCP <sub>514</sub> expression cassette with single residue-substitution F382W at HindIII and BamHI; Ap <sup>R</sup> , Cr <sup>R</sup>	This study
pCB301	A minim binary vector plasmid, Km <sup>R</sup>	[38]
pCBMCP <sub>514</sub>	pCB301, carrying a 2207bp MCP <sub>514</sub> expression cassette at BamHI and EcoRI, which consisted of 662bp upstream and 1545bp orf of <i>atu0514</i> ; Km <sup>R</sup>	This study
pCBMCP <sub>514</sub> <sup>F328A</sup>	pCB301, carrying a 2207bp MCP <sub>514</sub> expression cassette with single residue-substitution F382A; Km <sup>R</sup>	This study
pCBMCP <sub>514</sub> <sup>N336A</sup>	pCB301, carrying a 2207bp MCP <sub>514</sub> expression cassette with single residue-substitution N336A; Km <sup>R</sup>	This study
pCBMCP <sub>514</sub> <sup>E340A</sup>	pCB301, carrying a 2207bp MCP <sub>514</sub> expression cassette with single residue-substitution E340A; Km <sup>R</sup>	This study
pCBMCP <sub>514</sub> <sup>R343A</sup>	pCB301, carrying a 2207bp MCP <sub>514</sub> expression cassette with single residue-substitution R343A; Km <sup>R</sup>	This study
pCBMCP <sub>514</sub> <sup>V353A</sup>	pCB301, carrying a 2207bp MCP <sub>514</sub> expression cassette with single residue-substitution V353A; Km <sup>R</sup>	This study
pBT	Bait plasmid, Cm <sup>R</sup>	Stratagene
pBT-MCP <sub>514</sub> MA	pBT, carrying <i>atu0514</i> orf at BamHI and XhoI, Cm <sup>R</sup>	This study
pTRG	Target plasmid, Tc <sup>R</sup>	Stratagene
pTRG-W <sub>1</sub>	pTRG, carrying <i>cheW1</i> orf at BamHI and XhoI, Tc <sup>R</sup>	[40]
pTRG-W <sub>2</sub>	pTRG, carrying <i>cheW2</i> orf at BamHI and XhoI, Tc <sup>R</sup>	[40]

References

63. Thomashow, M.; Nutter, R.; Montoya, A.; Gordon, M.; Nester, E. Integration and organization of Ti plasmid sequences in crown gall tumors. *Cell* **1980**, *19*, 729-739, doi:10.1016/S0092-8674(80)80049-3.

**Table S2.** Primers used in this study

Primers	Sequences	Purpose
dMCP <sub>514</sub> -1	GACTCTAGAGGATCCCGTTGATTTGTTTTAGGGGAAC	To amplify the upstream sequence of <i>atu0514</i> to construct pEXMCP <sub>514</sub>
dMCP <sub>514</sub> -2	GACGGTGTGCAGCGCATCGAAATCGGATTGCATCTCATCAAG	
dMCP <sub>514</sub> -3	GAGATGCAATCCGATTTCGATGCGCTGCACACCGTCAT	To amplify the downstream sequence of <i>atu0514</i> to construct pEXMCP <sub>514</sub>
dMCP <sub>514</sub> -4	TGCTGCCAACTCGAGCATCTCCTTAGGGAGCAAATGGT	
egfp-MCP <sub>514</sub> -1	GATTACGCCAAGCTTGATGGTGAGCAAGGGCGAGG	To amplify the sequence of <i>egfp</i>
egfp-MCP <sub>514</sub> -2	GCTGCCACCTCCACCCCTGTACAGCTCGTCCATGCC	
egfp-MCP <sub>514</sub> -3	GGTGGAGGTGGCAGCATGGGTATGTTTCGTTCAAAAATC	To amplify the sequence of <i>atu0514</i> fusing to <i>egfp</i>
egfp-MCP <sub>514</sub> -4	GGTACCCGGGGATCCTTACATGACTCCCTGACGTCTG	
pCBMCP <sub>514</sub> -1	AGAAGTAGTGGATCCCGAGGGTACCTGAAAGCCGAT	To amplify the complement sequence of pCBMCP <sub>514</sub>
pCBMCP <sub>514</sub> -2	CTTGATATCGAATTCCTTACATGACTCCCTGACGTCTG	
F328A-1	GAGATCGCCGCCAGACCAACCTTCTCGCCCT	To amplify the sequence of MCP <sub>514</sub> with the single residue-substitution F328A
F328A-2	GGTTGGTCTGGGCGGCGATCTCGTCGATGGTGC	
F328P-1	GAGATCGCCCCCAGACCAACCTTCTCGCCCT	To amplify the sequence of MCP <sub>514</sub> with the single residue-substitution F328P
F328P-2	GGTTGGTCTGGGGGGCGATCTCGTCGATGGTGC	
F328W-1	GAGATCGCCTGGCAGACCAACCTTCTCGCCCT	To amplify the sequence of MCP <sub>514</sub> with the single residue-substitution F328W
F328W-2	GGTTGGTCTGCCAGGCGATCTCGTCGATGGTGC	
N336A-1	TCGCCCTCGCCGCCGGCATCGAGGCG	To amplify the sequence of MCP <sub>514</sub> with the single residue-substitution N336A
N336A-2	TGCCGGCGGCGAGGGCGAGAAGGTTGGTCT	
E340A-1	CCGGCATCGCCGCGGCACGGGCCC	To amplify the sequence of MCP <sub>514</sub> with the single residue-substitution E340A
E340A-2	GTGCCGCGGCGATGCCGGCATTGAGGGCG	
R343A-1	GGCGGCAGCCGCCGGCGACAGCGGA	To amplify the sequence of MCP <sub>514</sub> with the single residue-substitution R343A
R343A-2	CGCCGGCGGCTGCCGCCTCGATGCCG	
V353A-1	GGTTTCGCCGCCGTTGCGCAGGAAGTCCGGG	To amplify the sequence of MCP <sub>514</sub> with the single residue-substitution V353A
V353A-2	TGCGCAACGGCGGCGAAACCGCGTCCG	
MCP <sub>514</sub> MA-1	GGCCGCATCGAATTCTCGCTGATAGCCGCCAG	To amplify the sequence of MCP <sub>514</sub> cytoplasmic signal domain
MCP <sub>514</sub> MA-2	TTAACTCGAGGATCCTTAGCGGGCAATACGGAACG	