Supplementary Materials

Organization of Bone Mineral: The Role of Mineral– Water Interactions

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Figure S1. Individual ³¹P signals of the crystalline nanoparticle core and the amorphous surface layer within bone-like hydroxyapatite particles, together with their evolution from dry to wet conditions. Shown are the normalized one-dimensional (1D) individual ³¹P NMR signals of the crystalline nanoparticle core (left) and the amorphous surface layer (right) of the synthetic Carbonated HydroxyApatite (CHA) particles in dry conditions (bottom) and soaked in water (top). These individual ³¹P NMR signals were extracted from the two-dimensional (2D) {¹H}³¹P Heteronuclear Correlation (HetCor) NMR spectra that are shown here. The sums of the F2 slices taken at the OH ions position (from δ^1 H = -2 to 2 ppm) in F1 have been used to generate the individual ³¹P signals of

the crystalline nanoparticle core both in dry and wet conditions. The sums of the F2 slices taken at the bound water molecules position (from δ^1 H = 3 to 7 ppm) or the residual structural water molecules together with HPO₄²⁻ ions position (from δ^1 H = 3 to 17 ppm) in F1 have been used to generate the individual ³¹P signals of the amorphous surface layer, respectively in wet and dry conditions.



Figure S2. Observations of bone-like platelet-shaped hydroxyapatite particles seen edge-on. Transmission electron micrograph (TEM) of the synthetic Carbonated HydroxyApatite (CHA) particles. Some of them are here standing vertically such that their smallest side faces are exposed while their largest side faces are parallel to the electron beam.



Figure S3. Spatial proximities between phosphorus and hydrogen species within the needle/rodshaped carbonated hydroxyapatite particles. (**A**) Two-dimensional (2D) {¹H}³¹P Heteronuclear Correlation (HetCor) NMR spectrum of the synthetic needle/rod-shaped Carbonated HydroxyApatite (CHA-needle/rod) sample. Signal intensity increases from blue to red. (**B**) Onedimensional (1D) ¹H projection of the vertical F1 dimension extracted from the 2D {¹H}³¹P HetCor NMR spectrum displayed in (A).



Figure S4. Behavior of the needle/rod-shaped carbonated hydroxyapatite particles in water. Shown is a representative cryogenic transmission electron micrograph (cryo-TEM) of the synthetic needle/rod-shaped Carbonated HydroxyApatite (CHA-needle/rod) particles dispersed in water.



Figure S5. Spatial proximities between phosphorus and hydrogen species within monetite and brushite. Two-dimensional (2D) {¹H}³¹P Heteronuclear Correlation (HetCor) NMR spectra of synthetic monetite (**A**) and brushite (**C**) samples soaked in water (pH 4). Signal intensity increases from blue to red. Normalized one-dimensional (1D) ¹H projections of the vertical F1 dimensions extracted from the 2D {¹H}³¹P HetCor NMR spectra of monetite (**B**) and brushite (**D**). For monetite, the spatial proximities among the various ¹H and ³¹P nuclei in the HPO₄²⁻ ions are detected as two correlation peaks observable at $\delta^{31}P = -0.2$ and -1.4 ppm in the F2 (³¹P) dimension, which are both associated with two correlation peaks observable at $\delta^{1}H = 13.3$ and 16.1 ppm in the F1 (¹H) dimension. For brushite, the spatial proximities among the various ¹H and ³¹P nuclei in the HPO₄²⁻ ions are detected as a single correlation peak observable at $\delta^{31}P = 1.4$ ppm in the F2 (³¹P) dimension and in the range of $\delta^{1}H = 8-15$ ppm in the F1 (¹H) dimension. In addition, the structural water molecules present in brushite are detected as a broad correlation peak observable at $\delta^{31}P = 1.4$ ppm in the F2 (³¹P) dimension and in the range of $\delta^{1}H = 8-15$ ppm in the F1 (¹H) dimension. In addition, the structural water molecules present in brushite are detected as a broad correlation peak observable at $\delta^{31}P = 1.4$ ppm in the F2 (³¹P) dimension and centered at around $\delta^{1}H = 5$ ppm in the F1 (¹H) dimension. Last, the red arrows indicate the presence of tiny signals of bound water which are not observable is dry conditions.