





Review

Determining the Effects of Eugenol on the Bond Strength of Resin-Based Restorative Materials to Dentin: A Meta-Analysis of the Literature

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Abstract: The aim of this study was to determine whether the residual presence of eugenol in coronal dentin may compromise the bond strength of resin-based restorative materials. A search was performed on MEDLINE/Pubmed, Scopus, and by hand search for relevant papers. No restriction was applied for language and publication date. The studies selected for analysis tested specimens with reduced size (micro-shear bond strength (μ SBS) and micro-tensile bond strength (μ TBS)) of adhesive systems and resin-based restorative materials applied to coronary dentin “contaminated” with eugenol-based materials. The search provided 335 articles, but only 10 studies met the inclusion criteria. The pooled global analysis showed a significant influence of eugenol, as it negatively influenced the bond strength of resin-based restorations (5.79 (3.31–8.28) MPa, $p < 0.00001$). The subgroup analyses for conventional etch-and-rinse ($p = 0.003$) and self-etch ($p < 0.0004$) adhesive systems, as well as for μ SBS ($p = 0.01$) and μ TBS ($p < 0.0001$), showed a negative influence of eugenol on the bond strength. Data were statistically heterogeneous. However, it was possible to observe that eugenol could negatively affect the bonding of resin-based restorative materials to dentin. Further evidence is necessary in order to acquire more accurate information about this issue and confirm that the residual presence of eugenol in dentin compromises the bond strength of resin-based materials.

Keywords: eugenol; dentin; adhesive; zinc oxide–eugenol cement; dental cements; dental bonding

1. Introduction

Eugenol, in combination with zinc oxide (ZOE), is one of the most common materials used as a root canal sealer in endodontics [1–5], as well as a pulp sedative in cases of pulpotomies [6–9] and an impression material for edentulous patients [10–13]. In restorative dentistry, ZOE cements are frequently applied as temporary materials during indirect restorations [14]. It is also known that ZOE cements can perform as anodynes for pulpal pain [15], especially in those patients presenting dentin hypersensitivity after tooth preparation and temporary restorations [16].

ZOE cements used in temporary luting procedures are usually characterized by reasonable retention and compression strength and they can be removed quite easily [17]. Several techniques to eliminate residual provisional cements from dental preparations have been advocated. These include grating the tooth with a hand instrument (usually scalers or spoon excavators), cleaning the tooth preparation with a prophylaxis cup in combination with a water–pumice paste slurry, and/or using an intraoral air abrasion system commonly in combination with alumina or sodium bicarbonate. Although such temporary cements may be considered user-friendly materials in the clinic, there is evidence that the residual eugenol remaining on the dentin surface after setting can jeopardize the bonding performance of resin-based materials due to polymerization inhibition [1,18–20]. The current literature shows conflicting results and a distinct lack of robust evidence about this issue.

The remaining presence of eugenol-containing cements on dentin can act as a physical barrier [21,22], impairing the degree of monomer conversion of resin-based materials [23] due to the existence of radical-scavenging molecules in such phenolic compound [24]. Indeed, the reactivity of monomers is reduced due to the hydroxyl groups within the eugenol molecule, which can protonate free radicals during the polymerization reaction [25]. Hence, an optimal removal of provisional eugenol-containing cements must be achieved in order to accomplish a suitable adhesion of permanent resin-based restorations to dentin [1,26].

Dentin hybridization through the use of adhesive systems is an essential mechanism for tooth/material adhesion [27]. Adhesive systems are made of photocurable resin monomers, which are responsible for micromechanical interlocking with the dental tissues. Such adhesive systems can be classified according to their mode of interaction with the bonding substrate as (i) etch-and-rinse (ER) or (ii) self-etch (SE) systems [28]. Studies confirmed that eugenol can jeopardize their bonding performance, although it is still unclear which type of adhesive (ER or SE) is more at risk of debonding in case of contact with eugenol. A recent meta-analysis has shown that sealers with eugenol may be able to reduce the push-out strength of fiber posts applied in the root canal in combination with resin-based cements [29]. Although many studies tried to elucidate the effect of eugenol on the bond strength of resin adhesives to coronal dentin, there is still no consensus about this issue [18–20,30–35].

Therefore, this study aimed to analyze whether the residual presence of eugenol in the coronal dentin may compromise the bond strength of resin-based restorative materials. The hypothesis tested in this study was that the presence of residual eugenol on dentin surface would potentially interfere with SE and/or ER adhesive systems.

2. Material and Methods

This review was carried out following the guidelines of the PRISMA Statement (Preferred Reporting Items for Systematic Review and Meta-Analysis) [36,37]. A systematic electronic search was performed in MEDLINE/Pubmed and Scopus databases using the keywords “(eugenol) AND (bond strength)” (no restriction was applied for language and publication dates). Eligible studies were those that assessed immediate bond strength of adhesive systems to coronal dentin after contact with eugenol-containing materials by testing specimens with a reduced size (micro-shear bond strength (μ SBS) or micro-tensile bond strength (μ TBS)). Inclusion criteria comprised the presence of a comparison of bond strength values with and without (control group) eugenol, with mean and standard deviation values presented in megapascal (MPa).

The searched titles and abstracts were checked carefully, and noneligible articles were excluded. Subsequently, full texts of the selected studies were obtained, and data were extracted to gather information about (1) method, (2) teeth type, (3) adhesive system, (4) eugenol-based material, (5) eugenol contact time, (6) storage time of samples prior the bond strength test, (7) sample size, (8) mean bond strength, and (9) standard deviation.

To assess the individual risk of bias of each study, six methodological items were analyzed: (1) screening for caries and cracks on teeth, (2) blindness of the researcher, (3) teeth randomization, (4) use of materials according to manufacturers' instructions, (5) time of storage before debonding, and (6) presence of pretest failure that was included in statistical analyses. Such studies were classified as having high, moderate, or low risk of bias: studies with high risk failed to report four items or more, those with moderate risk failed to report 3 or 2 items, and those with low risk failed to report one item or less. The bias risk of the articles was evaluated by plotting the effect by the inverse of its standard error. The symmetry of the resulting funnel plot was visually assessed.

Meta-analysis was conducted using the "Review Manager Software (RMS)" (version 5.3) from the Cochrane Collaboration (Copenhagen, Denmark), and the mean difference with 95% confidence interval (CI) was calculated for mean bond strength of all groups of the studies included using the inverse variance method. The p -value for significance was set at $p < 0.05$. Statistical homogeneity (I^2) of the treatment effect was assessed by the modified chi-square test (Cochran's Q). The assessment indicated that there was statistical homogeneity when $p > 0.10$. Global and subgroup analyses were carried out (subgroups were arranged in ER or SE adhesive system and μ SBS or μ TBS).

3. Results

The latest search was performed on 9 November 2019. Figure 1 displays a flowchart representing the selection process of the studies involved in our work. Initially, 335 manuscripts were obtained (156 from PubMed and 179 from Scopus). Of those 335 studies, 147 were excluded as they were present in both databases. The title and abstract of each article were inspected, and 224 manuscripts were excluded because they were not relevant to the topic of this study or they did not meet the selection criteria (i.e., studies not related to dentistry or to dental restorations or based on macro-mechanical bonding tests; the latter were 29 studies). One study was excluded because the access to the manuscript was not possible, even after several attempts to contact the corresponding author [38]. A hand search was performed by checking the references in relevant papers, although no additional studies were found. Ten studies were included in the analysis, and their full texts were carefully examined.

Among the studies selected, nine were published in English and one in Portuguese. Eight studies assessed μ TBS, and two assessed μ SBS. Only two studies were performed using bovine teeth, while the other eight studies were performed using human teeth (one used the primary molar, and seven used the third molar). Five studies used only a eugenol-based material as a control group, whereas the other five also had a non-eugenol-based material group as a control. The adhesive systems tested in the studies were Adper Single Bond 2 (3M ESPE, Saint Paul, MN, USA) and Optibond FL (Kerr, Orange, CA, USA); the SE adhesive systems were Clearfil SE Bond (Kuraray, Okayama, Japan), iBond (Heraeus Kulzer, Hanau, Germany), One-Up Bond F (Tokuyama, Tokyo, Japan), Adper SE Plus (3M ESPE, St Paul, MN, USA), and Adper Prompt (3M ESPE, Saint Paul, MN, USA). The descriptive data of the 10 included studies are shown in Table 1.

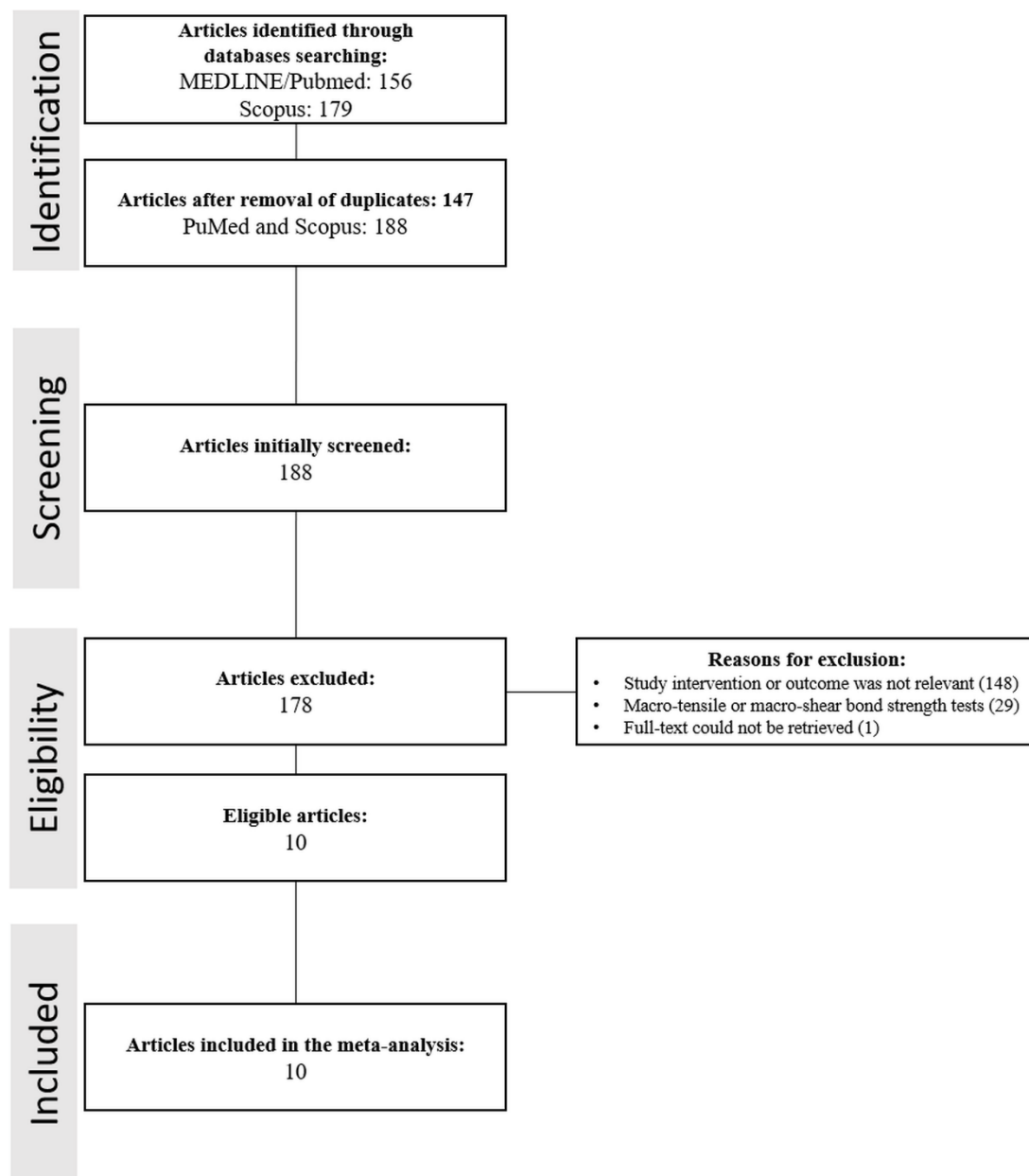


Figure 1. Systematic review flow chart of studies comparing the bond strength of adhesive systems/composite resin to coronal dentin after contact with eugenol-based materials and without contact.

Table 1. Identification and characteristics of the included studies.

| Author | Methodology | Substrate | Adhesive | Eugenol Material | Eugenol Contact Time | Storage Time | Group Sample Size | Bond Strength (MPa) | SD |
|--------------------------------|---|---------------------------|------------------|------------------|----------------------|--------------|-------------------|---|-------|
| Fonseca et al. 2005 [39] | μ TBS (sticks of 1 mm ²) | Bovine Teeth | Single Bond | Dycal | 7 days | - | 5 | 27.21 (Al ₂ O ₃ sandblasting) | 3.71 |
| | | | | | | - | 5 | 20.36 (Pumice-water slurry) | 3.99 |
| | | | | | | - | 5 | 15.98 (Hand scaler) | 3.22 |
| | | | | Provy | 7 days | - | 5 | 28.31 (Al ₂ O ₃ sandblasting) | 1.77 |
| | | | | | | - | 5 | 26.10 (Pumice-water slurry) | 5.63 |
| | | | | | | - | 5 | 20.73 (Hand scaler) | 5.29 |
| | | | | TempBond NE | 7 days | - | 5 | 28.28 (Al ₂ O ₃ sandblasting) | 3.29 |
| | | | | | | - | 5 | 25.39 (Pumice-water slurry) | 3.10 |
| | | | | | | - | 5 | 24.25 (Hand scaler) | 4.06 |
| Carvalho et al., 2007 [18] | μ SBS (Cylinders - H:0.5 mm \times D:0.75 mm) | Human Teeth (third molar) | Clearfil SE Bond | IRM | 24 h | 24 h | 6 | 23.7 | 1.7 |
| | | | | None | N/A | | 6 | 30.5 | 2.0 |
| | | | iBond | IRM | 24 h | | 6 | 19.7 | 8.5 |
| | | | | None | N/A | | 6 | 25.3 | 5.7 |
| | | | Single Bond | IRM | 24 h | | 6 | 28.3 | 3.8 |
| | | | | None | N/A | | 6 | 31.3 | 2.7 |
| Schwartzter, et al., 2007 [17] | μ TBS (sticks of 0.5 mm ² Aprox. 0.7 mm \times 0.7 mm) | Bovine Teeth | One-UP Bond F | TempCem | 7 days | 24 h | 5 | 39.3 | 15.72 |
| | | | | None | N/A | | 5 | 44.67 | 13.31 |
| | | | | TempCem NE | 7 days | | 5 | 41.35 | 13.42 |

Table 1. Cont.

| Author | Methodology | Substrate | Adhesive | Eugenol Material | Eugenol Contact Time | Storage Time | Group Sample Size | Bond Strength (MPa) | SD |
|----------------------------|---|----------------------------|---------------------|------------------|----------------------|--------------|-------------------|---------------------|------|
| Sanabe; Hebling, 2009 [40] | μ TBS (sticks of 0.81 mm ²) | Human Teeth (third molar) | Adper Single Bond | Cavit | 7 days | 24 h | 4 | 37.2 | 12.8 |
| | | | | IRM | 7 days | | 4 | 41.7 | 15.1 |
| | | | | None | N/A | | 4 | 45.5 | 15.1 |
| | | | Clearfil SE Bond | Cavit | 7 days | | 4 | 42.1 | 11.0 |
| | | | | IRM | 7 days | | 4 | 30.1 | 13.8 |
| | | | | None | N/A | | 4 | 38.9 | 13.5 |
| Ribeiro, et al., 2011 [19] | μ TBS (rectangular beams of 0.9 mm ²) | Human Teeth (third molars) | Single Bond | TempBond | 7 days | 24 h | 5 | 39.4 | 15.6 |
| | | | | None | N/A | | 5 | 44.9 | 15.6 |
| | | | | Freegenol | 7 days | | 5 | 47.4 | 18.8 |
| | | | Adper Prompt | TempBond | 7 days | | 5 | 27.4 | 12.3 |
| | | | | None | N/A | | 5 | 32.4 | 10.8 |
| | | | | Freegenol | 7 days | | 5 | 31.1 | 12.8 |
| Silva et al., 2011 [20] | μ SBS (Cylinders -H:2mmxD:1mm) | Human Teeth (third molars) | Adper SE Plus | IRM | 24 h | Immediate | 10 | 13.9 | 3.4 |
| | | | | | 7 days | | 10 | 26.0 | 3.8 |
| | | | | | 14 days | | 10 | 24.1 | 4.2 |
| | | | | None | N/A | | 10 | 24.3 | 8.4 |
| Koch, et al., 2013 [1] | μ TBS (sticks of 1.07 mm ²) | Human Teeth (molars) | Optibond FL | IRM | 1 day | 7 days | 21 | 12.5 | 5.3 |
| | | | | | 7 days | | 21 | 17.2 | 9.8 |
| | | | | | 28 days | | 21 | 17.0 | 8.0 |
| | | | | None | N/A | | 21 | 26.3 | 7.1 |
| Pinto et al., 2014 [41] | μ TBS (sticks of 0.8 mm ²) | Human Teeth (third molars) | Adper Single Bond 2 | IRM | 24 h | Immediate | 5 | 46.8 | 3.4 |
| | | | | | 7 days | | 5 | 63.0 | 3.2 |
| | | | | | 45 days | | 5 | 59.3 | 2.3 |
| | | | | None | N/A | | 5 | 60.4 | 5.2 |
| | | | Clearfil S3 Bond | IRM | 24 h | | 5 | 20.4 | 2.2 |
| | | | | | 7 days | | 5 | 18.1 | 2.1 |
| | | | | | 45 days | | 5 | 35.2 | 3.9 |
| | | | | None | N/A | | 5 | 39.1 | 4.2 |

Table 1. Cont.

| Author | Methodology | Substrate | Adhesive | Eugenol Material | Eugenol Contact Time | Storage Time | Group Sample Size | Bond Strength (MPa) | SD |
|--------------------------------|--|-----------------------------|---------------------|---|----------------------|--------------|-------------------|---------------------|------|
| Pires et al., 2018 [6] | μ TBS (sticks of 1 mm ²) | Human Teeth (primary molar) | Adper Single Bond 2 | Zinc oxide and eugenol | 15 min | 24 h | 8 | 6.6 | 1.5 |
| | | | | Iodoform-based Guedes-Pinto paste | 15 min | | 8 | 10.2 | 2.5 |
| | | | | Calcium hydroxide paste thickened with zinc oxide | 15 min | | 8 | 9.5 | 1.5 |
| | | | | None | N/A | | 8 | 10.2 | 2.3 |
| Wongsorachai et al., 2018 [42] | μ TBS (sticks of 1 mm ²) | Human Teeth (third molar) | Optibond FL | IRM | 24 h | 24 h | 10 | 34.39 | 5.84 |
| | | | | None | N/A | | 10 | 52.52 | 3.41 |
| | | | Clearfil SE Bond | IRM | 24 h | | 10 | 20.14 | 4.16 |
| | | | | None | N/a | | 10 | 46.03 | 5.21 |

The provisional eugenol-based materials used were Provy (Herpo/Dentsply, São Paulo, Brazil), IRM (Dentsply, Milford, USA), TempCem (Vigodent, Rio de Janeiro, Brazil), TempBond (Kerr, Orange, CA, USA), and ZOE and eugenol (Biodinâmica, Ibiporá, Brazil and Maquira, Maringá, Brazil). The provisional non-eugenol-based materials were Dycal (Dentsply/Caulk, Milford, DE, USA), TempCem NE (Vigodent, Rio de Janeiro, Brazil), Freegenol (GC, Kasugai, Japan), iodoform-based Guedes-Pinto paste (Biodinâmica, Ibiporá, Brazil; Maquira, Maringá, Brazil; Fórmula&Ação, compounding pharmacy, São Paulo, Brazil), and calcium hydroxide paste thickened with ZOE (S. S. White Artigos Dentários Ltd.a., Rio de Janeiro, RJ, Brazil and Biodinâmica, Ibiporá, Brazil). In several studies, the effect of different eugenol contact times was tested, being the bond strength normally tested 24 h after the bonding step. In all studies, the primary outcome was bond strength expressed in MPa. The lowest mean for bond strength was 6.6 MPa, and the highest was 63 MPa (Table 1). During the qualitative analysis (Table 2), three studies were classified as having a high individual risk of bias, six as having a moderate individual risk of bias, and one as having a low individual risk of bias. The funnel plot (Figure 2) indicates the asymmetry among the studies.

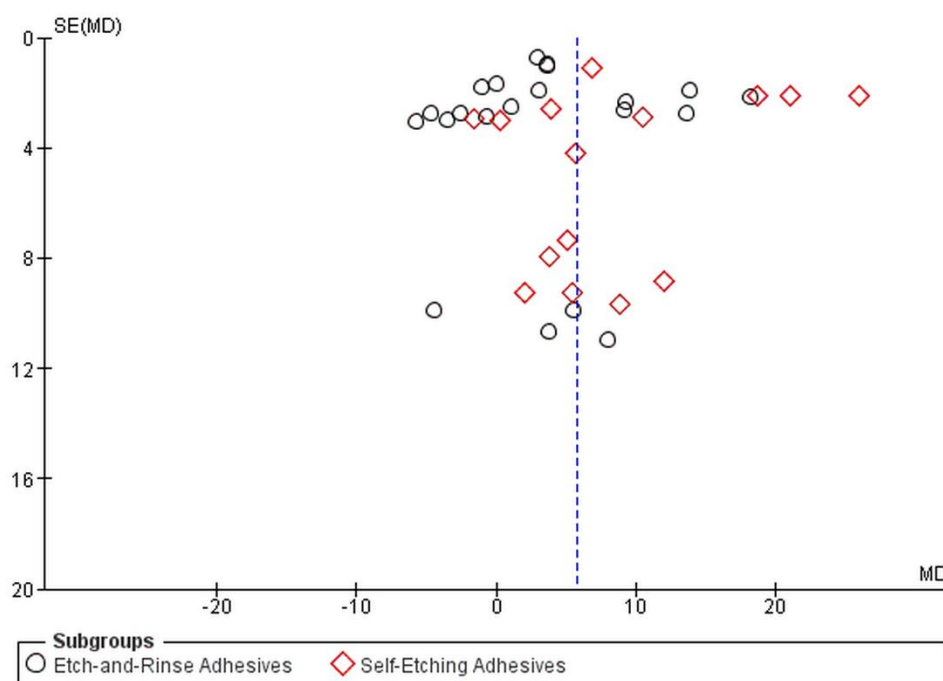


Figure 2. Funnel plot of the included studies.

The meta-analysis conducted on 10 studies with subgroups in ER and SE adhesive systems resulted in 36 comparisons (Figure 3): 21 in the ER adhesive system and 15 in the SE adhesive system. The random effects model was used, and the pooled global analysis showed a significant influence of eugenol ($p < 0.00001$). Eugenol had a negative effect on the bond strength of resin-based restorations (5.79 (3.31–8.28) MPa). Within the subgroup analyses, a previous contact with eugenol-based materials also negatively affected the bond strength (ER: 3.63 MPa (1.22–6.05), $p = 0.003$; SE: 9.16 (4.08–14.23) MPa, $p = 0.0004$). The heterogeneity was calculated via τ^2 , Cochrane's Q value, and I^2 . For the ER subgroup, τ^2 was 22.08 (Cochrane's Q value $p = 0.00001$), and I^2 was 86%. For the SE subgroup, τ^2 was 75.34 (Cochrane's Q value $p < 0.00001$), and I^2 was 90%. For the global analysis, τ^2 was 42.18 (Cochrane's Q value $p < 0.00001$), and I^2 was 90%. Therefore, significant heterogeneity was detected within comparisons and in the global analysis. The test for subgroup difference showed $I^2 = 73.1\%$.

Table 2. Individual risk of bias of the included studies.

| Study | Teeth Free of Caries | Blinded Researcher | Teeth Randomization | Manufacturer's Instructions | Storage Time | Pre-Test Failure | Classification |
|--------------------------------|----------------------|--------------------|---------------------|-----------------------------|--------------|------------------|----------------|
| Fonseca et al., 2005 [39] | + | - | - | + | ? | - | High |
| Carvalho et al., 2007 [18] | + | - | - | + | + | - | Moderate |
| Schwartz et al., 2007 [17] | + | - | - | + | + | - | Moderate |
| Sanabe; Hebling, 2009 [40] | + | - | - | - | + | - | High |
| Ribeiro et al., 2011 [19] | + | - | + | + | + | ? | Moderate |
| Silva et al., 2011 [20] | + | - | + | + | - | - | Moderate |
| Koch et al., 2013 [1] | - | - | + | - | + | - | High |
| Pinto et al., 2014 [41] | - | - | - | + | + | - | High |
| Pires et al., 2018 [6] | + | + | + | + | + | - | Low |
| Wongsorachai et al., 2018 [42] | + | - | + | + | + | - | Moderate |

+ reported; - not reported; ? unclear.

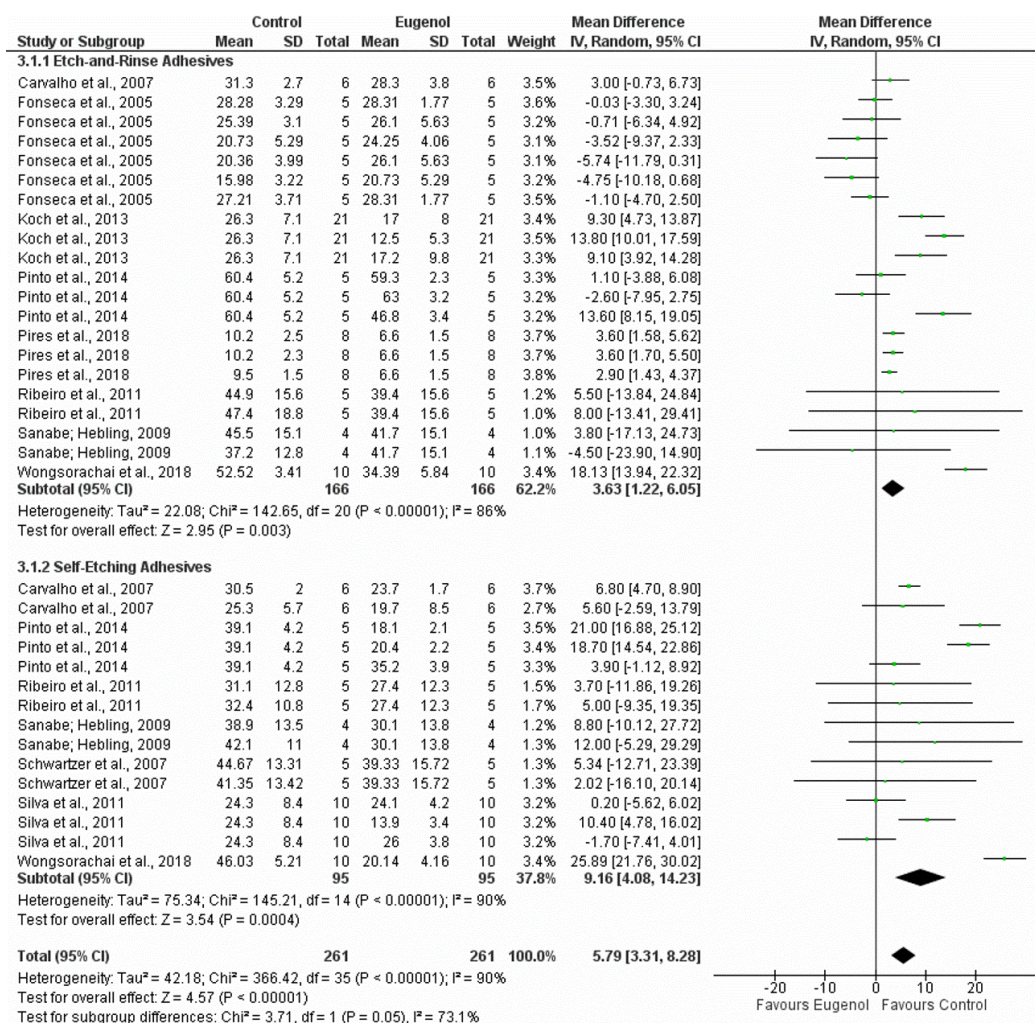


Figure 3. Forest plot of global and subgroups (etch-and-rinse (ER) or self-etch (SE) adhesive system) meta-analyses. The bond strengths of adhesive systems/resin-based restorative materials to coronal dentin after contact with eugenol-based materials and without contact were compared.

Besides the meta-analysis comparing ER and SE adhesive systems, the differences between subgroups for μ SBS and μ TBS were compared (Figure 4). Starting from 36 comparisons, only 6 were used for μ SBS and 30 for μ TBS. For both subgroups, the presence of eugenol before bonding negatively affected the bond strength of the tested adhesives (μ SBS: 4.21 (0.91–7.51) MPa; μ TBS: 6.39 (3.39–9.38) MPa). Intragroup heterogeneity was observed for μ SBS ($\tau^2 = 10.38$, (Cochrane's Q value $p = 0.009$) and $I^2 = 67\%$) and for μ TBS ($\tau^2 = 51.06$ (Cochrane's Q value $p < 0.00001$) and $I^2 = 92\%$). The test for subgroup differences showed no heterogeneity between them ($I^2 = 0\%$, Cochrane's Q value = 0.34).

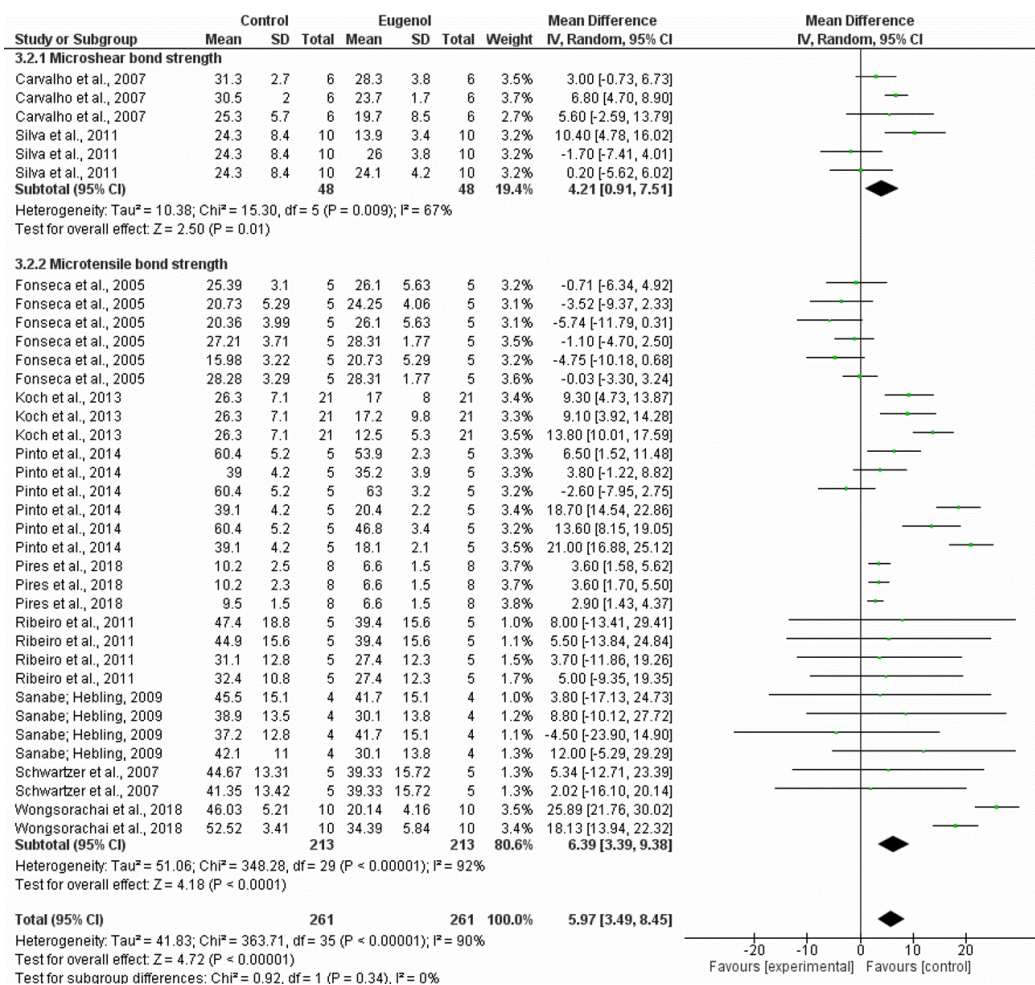


Figure 4. Forest plot of global and subgroups (μ SBS or μ TBS) meta-analyses. The bond strength of adhesive systems/resin-based restorative materials to coronal dentin after contact with eugenol-based materials and without contact were compared via two tests used to classify the studies as μ SBS or μ TBS.

Table 3 shows a qualitative description of the methods applied to remove the cements and to clean the surfaces before performing any bonding procedure. There was high variability among the techniques. For instance, some of them used Al_2O_3 sandblasting, and others a pumice–water slurry, a scaler, or a combination of these methods. Moreover, these techniques were based on different parameters, such as times of application, distance, and presence or absence of water irrigation, in different studies.

Table 3. Identification of the study and detailed method apply to remove the eugenol-containing cement and to clean the dentin surfaces prior to the bonding procedures.

| Author | Adhesive | Eugenol Material | Details of Cleaness Method Applied |
|-------------------------------|------------------|------------------|--|
| Fonseca et al. 2005 [39] | Single Bond | Dycal | <ul style="list-style-type: none"> • Hand scaler for 10 s, or • Pumice-water slurry for 10 s, or • Sandblasting with 50 µm aluminum oxide particles for 5 s at a pressure of 4 bars and source-to-samples distance of 2 cm. |
| | | Provy | <ul style="list-style-type: none"> • Hand scaler for 10 s, or • Pumice-water slurry for 10 s, or • Sandblasting with 50 µm aluminum oxide particles for 5 s at a pressure of 4 bars and source-to-samples distance of 2 cm. |
| | | TempBond NE | <ul style="list-style-type: none"> • Hand scaler for 10 s, or • Pumice-water slurry for 10 s, or • Sandblasting with 50 µm aluminum oxide particles for 5 s at a pressure of 4 bars and source-to-samples distance of 2 cm. |
| Carvalho et al., 2007 [18] | Clearfil SE Bond | IRM | <ul style="list-style-type: none"> • Mechanically removed with a scaler until the dentin surfaces were visually macroscopically free of material; • Cleaned with a pumice-water slurry in a slow-speed handpiece for 60 s and rinsed off with air-water stream 60 s. |
| | | None | <ul style="list-style-type: none"> • Not reported |
| | iBond | IRM | <ul style="list-style-type: none"> • Mechanically removed with a scaler until the dentin surfaces were visually macroscopically free of material; • Cleaned with a pumice-water slurry in a slow-speed handpiece for 60 s and rinsed off with air-water stream 60 s. |
| | | None | <ul style="list-style-type: none"> • Not reported |

Table 3. Cont.

| Author | Adhesive | Eugenol Material | Details of Cleanness Method Applied |
|-----------------------------------|-------------------|------------------|--|
| Schwartzter, et al., 2007 [17] | Single Bond | IRM | <ul style="list-style-type: none"> • Mechanically removed with a scaler until the dentin surfaces were visually macroscopically free of material; • Cleaned with a pumice-water slurry in a slow-speed handpiece for 60 s and rinsed off with air-water stream 60 s. |
| | | None | <ul style="list-style-type: none"> • Not reported |
| | One-UP Bond F | TempCem | <ul style="list-style-type: none"> • Mechanically removed using periodontal instruments and Robinson brushes equipped with pumice; • Rinsing with distilled water. |
| | | None | <ul style="list-style-type: none"> • Not reported |
| Sanabe; Hebling, 2009 [40] | Adper Single Bond | TempCem NE | <ul style="list-style-type: none"> • Mechanically removed using periodontal instruments and Robinson brushes equipped with pumice; • Rinsing with distilled water. |
| | | Cavit | <ul style="list-style-type: none"> • Mechanically removed with dentin spoon up to a visually clean surface; • Pumice-water and running water. |
| | | IRM | <ul style="list-style-type: none"> • Mechanically removed with dentin spoon up to a visually clean surface; • Pumice-water and running water. |
| | | None | <ul style="list-style-type: none"> • Not reported |

Table 3. Cont.

| Author | Adhesive | Eugenol Material | Details of Cleanness Method Applied |
|---------------------------|------------------|------------------|--|
| | Clearfil SE Bond | Cavit | <ul style="list-style-type: none"> • Mechanically removed with dentin spoon up to a visually clean surface; • Pumice-water and running water. |
| | | IRM | <ul style="list-style-type: none"> • Mechanically removed with dentin spoon up to a visually clean surface; • Pumice-water and running water. |
| | | None | <ul style="list-style-type: none"> • Not reported |
| Ribeiro et al., 2011 [19] | Single Bond | TempBond | <ul style="list-style-type: none"> • Scraped off with a dental instrument until unaided visual inspection indicated that the surface was free of cement; • Rinsed with water spray, cleaned with cotton pellets soaked with pumice-water slurry for 20 s; • Rinsed again and air-dried. |
| | | None | <ul style="list-style-type: none"> • Not reported |
| | | Freegenol | <ul style="list-style-type: none"> • Scraped off with a dental instrument until unaided visual inspection indicated that the surface was free of cement; • Rinsed with water spray, cleaned with cotton pellets soaked with pumice-water slurry for 20 s; • Rinsed again and air-dried. |

Table 3. Cont.

| Author | Adhesive | Eugenol Material | Details of Cleanness Method Applied |
|-------------------------|---------------|------------------|--|
| | Adper Prompt | TempBond | <ul style="list-style-type: none"> Scraped off with a dental instrument until unaided visual inspection indicated that the surface was free of cement; Rinsed with water spray, cleaned with cotton pellets soaked with pumice-water slurry for 20 s/ Rinsed again and air-dried. |
| | | None | <ul style="list-style-type: none"> Not reported |
| | | Freegenol | <ul style="list-style-type: none"> Scraped off with a dental instrument until unaided visual inspection indicated that the surface was free of cement; Rinsed with water spray, cleaned with cotton pellets soaked with pumice-water slurry for 20 s; Rinsed again and air-dried. |
| Silva et al., 2011 [20] | Adper SE Plus | IRM | <ul style="list-style-type: none"> Mechanically removed with a scaler until the dentin surface was visually (macroscopically) free of the material; Cleaned with pumice-water slurry in a slow-speed handpiece and rinsed with an air-water stream. |
| | | | <ul style="list-style-type: none"> Mechanically removed with a scaler until the dentin surface was visually (macroscopically) free of the material; Cleaned with pumice-water slurry in a slow-speed handpiece and rinsed with an air-water stream. |
| | | | <ul style="list-style-type: none"> Mechanically removed with a scaler until the dentin surface was visually (macroscopically) free of the material; Cleaned with pumice-water slurry in a slow-speed handpiece and rinsed with an air-water stream. |

Table 3. Cont.

| Author | Adhesive | Eugenol Material | Details of Cleaness Method Applied |
|-------------------------|---------------------|------------------|---|
| Koch et al., 2013 [1] | Optibond FL | None | <ul style="list-style-type: none"> Not reported. |
| | | IRM | <ul style="list-style-type: none"> Macroscopically cleaned of the ZOE using a dental scaler. |
| | | | <ul style="list-style-type: none"> Macroscopically cleaned of the ZOE using a dental scaler. |
| | | | <ul style="list-style-type: none"> Macroscopically cleaned of the ZOE using a dental scaler. |
| Pinto et al., 2014 [41] | Adper Single Bond 2 | None | <ul style="list-style-type: none"> Not reported. |
| | | IRM | <ul style="list-style-type: none"> Removed using a stainless-steel spatula; Cleaned with pumice- water slurry using a slow speed handpiece for 60 s and rinsed with an air-water stream (60 s). |
| | | | <ul style="list-style-type: none"> Removed using a stainless-steel spatula; Cleaned with pumice- water slurry using a slow speed handpiece for 60 s and rinsed with an air-water stream (60 s). |
| | | | <ul style="list-style-type: none"> Removed using a stainless-steel spatula; Cleaned with pumice- water slurry using a slow speed handpiece for 60 s and rinsed with an air-water stream (60 s). |
| | | None | <ul style="list-style-type: none"> Cleaned with pumice- water slurry using a slow speed handpiece for 60 s and rinsed with an air-water stream (60 s). |

Table 3. Cont.

| Author | Adhesive | Eugenol Material | Details of Cleanness Method Applied |
|------------------------|---------------------|---|---|
| Pires et al., 2018 [6] | Clearfil S3 Bond | IRM | <ul style="list-style-type: none"> Removed using a stainless-steel spatula; Cleaned with pumice- water slurry using a slow speed handpiece for 60 s and rinsed with an air-water stream (60 s). |
| | | | <ul style="list-style-type: none"> Removed using a stainless-steel spatula; Cleaned with pumice- water slurry using a slow speed handpiece for 60 s and rinsed with an air-water stream (60 s). |
| | | | <ul style="list-style-type: none"> Removed using a stainless-steel spatula; Cleaned with pumice- water slurry using a slow speed handpiece for 60 s and rinsed with an air-water stream (60 s). |
| | Adper Single Bond 2 | None | <ul style="list-style-type: none"> Cleaned with pumice- water slurry using a slow speed handpiece for 60 s and rinsed with an air-water stream (60 s). |
| | | Zinc oxide and eugenol | <ul style="list-style-type: none"> Mechanically removed with a spatula and the dentine surfaces rubbed with a dry cotton pellet until visually free of material; Water rinsing for 15 s and drying with oil-free airstream for 5 s. |
| | | Iodoform-based Guedes-Pinto paste | <ul style="list-style-type: none"> Mechanically removed with a spatula and the dentine surfaces rubbed with a dry cotton pellet until visually free of material; Water rinsing for 15 s and drying with oil-free airstream for 5 s. |
| | | Calcium hydroxide paste thickened with zinc oxide | <ul style="list-style-type: none"> Mechanically removed with a spatula and the dentine surfaces rubbed with a dry cotton pellet until visually free of material; Water rinsing for 15 s and drying with oil-free airstream for 5 s. |
| | | None | <ul style="list-style-type: none"> Not reported. |

Table 3. Cont.

| Author | Adhesive | Eugenol Material | Details of Cleanliness Method Applied |
|-----------------------------------|------------------|------------------|---|
| Wongsorachai et al., 2018 [42] | Optibond FL | IRM | <ul style="list-style-type: none"> • Mechanically removed with an ultrasonic scaler at the frequency of 28 kHz until the dentin surfaces were visually free of material; • cleaned with pumice and water slurry using a slow-speed handpiece for 60 s and rinsed off with an air-water stream for 30 s. |
| | | None | <ul style="list-style-type: none"> • cleaned with pumice and water slurry using a slow-speed handpiece for 60 s and rinsed off with an air-water stream for 30 s. |
| | Clearfil SE Bond | IRM | <ul style="list-style-type: none"> • Mechanically removed with an ultrasonic scaler at the frequency of 28 kHz until the dentin surfaces were visually free of material. • Cleaned with pumice and water slurry using a slow-speed handpiece for 60 s and rinsed off with an air-water stream for 30 s. |
| | | None | <ul style="list-style-type: none"> • cleaned with pumice and water slurry using a slow-speed handpiece for 60 s and rinsed off with an air-water stream for 30 s. |

4. Discussion

The bonding performance of dental adhesives and luting agents to dentin “contaminated” with eugenol [1,17–19,30,31,39] has been the subject of a plethora of investigations during the last two decades. Nevertheless, the controversial results, inconsistencies, and inaccuracies of these studies have not provided the dentists with any evidence to support a judicious clinical decision considering the adverse effect of eugenol on adhesive dentistry. In order to increase the quality of the current systematic review and meta-analysis, we systematized all the selected studies into subgroups: (1) conventional ER adhesive system or SE adhesive system and, (2) evaluation through μ SBS or μ TBS. For both subgroups analyses as well as for the global analysis, it was found that the presence of residual eugenol jeopardized the bond strength of resin-based materials.

Previous *in vitro* studies highlighted the detrimental effect of eugenol-containing materials on dental bonding outcomes of resin-based materials [1,20], although there was no general agreement in the literature about such issue. Several studies have suggested that the effect of eugenol on the bond strength depended on the adhesive system employed [17]. While ER adhesive systems require the use of phosphoric acid [43], SE adhesives act through acid monomers, which can demineralize and consecutively infiltrate the dentin surface. Accordingly, the conventional ER bonding approach involves a water-rinsing step after the application of phosphoric acid to remove the residual acid containing a dissolved smear layer [43], while SE adhesives modify and incorporate the smear layer within the adhesive layer [44]. Therefore, after the removal of eugenol-containing provisional cement, the following cleaning steps for dentinal surface treatment can change the concentration of eugenol present on dentin and affect the final bond strength of resin-based materials [17–19]. In a previous study [18], it was emphasized that the use of phosphoric acid during the application of the conventional ER system could remove the smear layer contaminated with eugenol. Nevertheless, the pooled outcome of the meta-analysis performed here showed that the eugenol-contaminated dentin presented lower bond strength, regardless of the adhesive system applied.

Surface treatments such as orthophosphoric acid or ethylenediaminetetraacetic acid (EDTA) were investigated, and both significantly reduced the quantity of eugenol on the dentin surface [1]. However, it was previously observed that, even after acid application, residues of provisional cements could remain on the dentin surface [22]; hence, this justifies the outcomes of the current study. The phenolic component of eugenol can react with free radicals in the monomeric conversion from C = C bonds to C–C bonds, reducing the reactivity of the resin monomers and their polymerization. In this way, the remaining eugenol can bind to hydroxyl groups of the monomers [23,24,45,46] and reduce the degree of conversion of the adhesive system [47]. A reliable degree of conversion is essential for polymer-based materials, as it is related to the achievement of proper physical properties (e.g., high tensile strength [48] and microhardness [49]) and lower water sorption and solubility [48,50]. Thus, resin adhesive systems that show a high polymerization rate achieve better bonding performance and stability over time [51]. Here, by analyzing only immediate μ TBS and μ SBS, it was possible to observe that eugenol could reduce the adhesion values significantly. Therefore, it is possible to consider that, in terms of long-term results, this scenario could be even worse due to water sorption and degradation of the hybrid layer over time, mainly for a system with its monomer conversion impaired.

Bonding tests, mainly those through μ TBS, are important for the study of resin-based restorative materials. The μ TBS results have been associated with *in vivo* outcomes, mainly when the specimens are subjected to storage and longitudinal assessment [52]. The longer the specimens of bonding procedures are stored, the higher is the correlation with *in vivo* results [53]. Indeed, μ TBS after six months of storage in water correlates with marginal discoloration of restorations. Unfortunately, during literature screening, we have noticed that there was no longitudinal assessment of bonding after eugenol application on dentin. The most extended time of specimens’ storage was 45 days [41] but this study did not perform a longitudinal analysis for dentin bonding strength. Thus, a further *in vitro* longitudinal analysis, possibly including *in vivo* outcomes, would be beneficial in order to avoid overestimating the results found in immediate μ TBS. However, to the best of our knowledge,

the present review provides the most complete view of the impact of eugenol in resin restorations on coronal dentin. In this context, bond strength tests using a reduced specimen size were preferred based on Griffith's defect theory, which suggests a decrease in the tensile strength of large materials in comparison to small ones [54]. The bond strength of large specimens could be low or involve too many defects. Such test could not show the proper performance of the tested materials [54–56]. The subgroup analysis considered for the type of method applied (μ TBS or μ SBS) showed $I^2 = 0$ in the test for subgroup difference and supported the findings of the pooled results. Thus, eugenol jeopardized the bonding to coronal dentin regardless of the adhesive systems used and the method of application.

Through this meta-analysis, it was possible to combine data across multiple studies and better estimate the differences between single studies. The percentage value defined by I^2 showed the magnitude of the difference among the studies, which can be explained by heterogeneity and not by chance [57]. Although eugenol reduced the bond strength in all meta-analyses performed here, high heterogeneity could be observed in the subgroups and in the pooled data analysis. Therefore, other factors presented here should explain the model and the obtained results. Among 10 studies included, 4 showed a high risk of bias, and 5 a moderate risk of bias. The resulting bias can be accredited to the lack of a proper report of methodologies. It can also be hypothesized that a link exists between the lack of a proper report of methodologies and the high heterogeneity found here.

Moreover, the studies differed mainly regarding the method used for the removal of the temporary eugenol-based material (Table 3). The different instruments applied and the duration of each step (e.g., time for cleaning with a pumice–water slurry) may have influenced the results. Some studies reported similar cleaning steps for the control group (no contact with eugenol prior to bonding), while other papers did not mention cleaning (Table 3).

Variation in the cleaning steps was also found among several studies. Different cleaning steps can lead to changes in the dentin's surface, which might impact the bonding performance. Finally, the type of ZOE-based material could also be an influencing factor for high heterogeneity. Different powder/liquid ratios for the ZOE cements classifications could increase eugenol release if a higher concentration of liquid with respect to that of powder is used [18]. One of the studies included in our analysis tested the effect of a root canal filling paste with eugenol instead of provisional eugenol–cement on the μ SBS of an adhesive system [6]. We have included this study because its methodological design was very similar to those applied for testing eugenol-containing cements. Regardless of the high heterogeneity observed, it is possible to conclude that this study corroborates a previous one showing that eugenol jeopardizes not only the bonding to root dentin [29] but also the bonding to coronal dentin.

5. Conclusions

Within the limitations of an in vitro systematic review, the included studies have an acceptable methodological quality, contributing to a fact-based pathway of information regarding this subject. We found moderate-quality evidence supporting the adverse effect of eugenol on the bonding of resin-based materials (e.g., adhesive systems) to coronal dentin. Finally, we believe that further clinical investigations can improve the quality of the evidence of the detrimental effect of eugenol and guide dentists to better treatment decisions that foster the longevity of adhesive restorations.

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