

Review

# Redox Imbalance in Neurological Disorders in Adults and Children

Federica Rey <sup>1,2</sup>, Clarissa Berardo <sup>1,2</sup>, Erika Maghraby <sup>1,3</sup>, Alessia Mauri <sup>1,2</sup>, Letizia Messa <sup>2,4</sup>, Letizia Esposito <sup>1,2</sup>, Giovanna Casili <sup>5</sup>, Sara Ottolenghi <sup>6</sup>, Eleonora Bonaventura <sup>7,8</sup>, Salvatore Cuzzocrea <sup>5</sup>, Gianvincenzo Zuccotti <sup>1,9</sup>, Davide Tonduti <sup>1,7,8</sup>, Emanuela Esposito <sup>5</sup>, Irene Paterniti <sup>5</sup>, Cristina Cereda <sup>2</sup> and Stephana Carelli <sup>1,2,\*</sup>

<sup>1</sup> Pediatric Clinical Research Center “Romeo ed Enrica Invernizzi”, Department of Biomedical and Clinical Sciences, University of Milano, 20157 Milano, Italy

<sup>2</sup> Center of Functional Genomics and Rare diseases, Department of Pediatrics, Buzzi Children’s Hospital, 20154 Milano, Italy

<sup>3</sup> Department of Biology and Biotechnology “L. Spallanzani”, University of Pavia, 27100 Pavia, Italy

<sup>4</sup> Department of Electronics, Information and Bioengineering (DEIB), Politecnico di Milano, 20133 Milano, Italy

<sup>5</sup> Department of Chemical, Biological, Pharmaceutical and Environmental Sciences, University of Messina, 98166 Messina, Italy

<sup>6</sup> Department of Medicine and Surgery, University of Milano Bicocca, 20126 Milano, Italy

<sup>7</sup> Child Neurology Unit, Buzzi Children’s Hospital, 20154 Milano, Italy

<sup>8</sup> Center for Diagnosis and Treatment of Leukodystrophies and Genetic Leukoencephalopathies (COALA), Buzzi Children’s Hospital, 20154 Milano, Italy

<sup>9</sup> Department of Pediatrics, Buzzi Children’s Hospital, 20154 Milano, Italy

\* Correspondence: stephana.carelli@unimi.it; Tel.: +39-02-50319825

**Abstract:** Oxygen is a central molecule for numerous metabolic and cytophysiological processes, and, indeed, its imbalance can lead to numerous pathological consequences. In the human body, the brain is an aerobic organ and for this reason, it is very sensitive to oxygen equilibrium. The consequences of oxygen imbalance are especially devastating when occurring in this organ. Indeed, oxygen imbalance can lead to hypoxia, hyperoxia, protein misfolding, mitochondria dysfunction, alterations in heme metabolism and neuroinflammation. Consequently, these dysfunctions can cause numerous neurological alterations, both in the pediatric life and in the adult ages. These disorders share numerous common pathways, most of which are consequent to redox imbalance. In this review, we will focus on the dysfunctions present in neurodegenerative disorders (specifically Alzheimer’s disease, Parkinson’s disease and amyotrophic lateral sclerosis) and pediatric neurological disorders (X-adrenoleukodystrophies, spinal muscular atrophy, mucopolysaccharidoses and Pelizaeus–Merzbacher Disease), highlighting their underlining dysfunction in redox and identifying potential therapeutic strategies.

**Keywords:** oxygen; redox; neurodegenerative diseases; neurodevelopmental disorders; Alzheimer’s disease; Parkinson’s disease; amyotrophic lateral sclerosis; X-adrenoleukodystrophies; spinal muscular atrophy; mucopolysaccharidoses; Pelizaeus–Merzbacher disease



**Citation:** Rey, F.; Berardo, C.; Maghraby, E.; Mauri, A.; Messa, L.; Esposito, L.; Casili, G.; Ottolenghi, S.; Bonaventura, E.; Cuzzocrea, S.; et al. Redox Imbalance in Neurological Disorders in Adults and Children. *Antioxidants* **2023**, *12*, 965. <https://doi.org/10.3390/antiox12040965>

Academic Editor: Susana Solá

Received: 20 February 2023

Revised: 3 April 2023

Accepted: 14 April 2023

Published: 20 April 2023



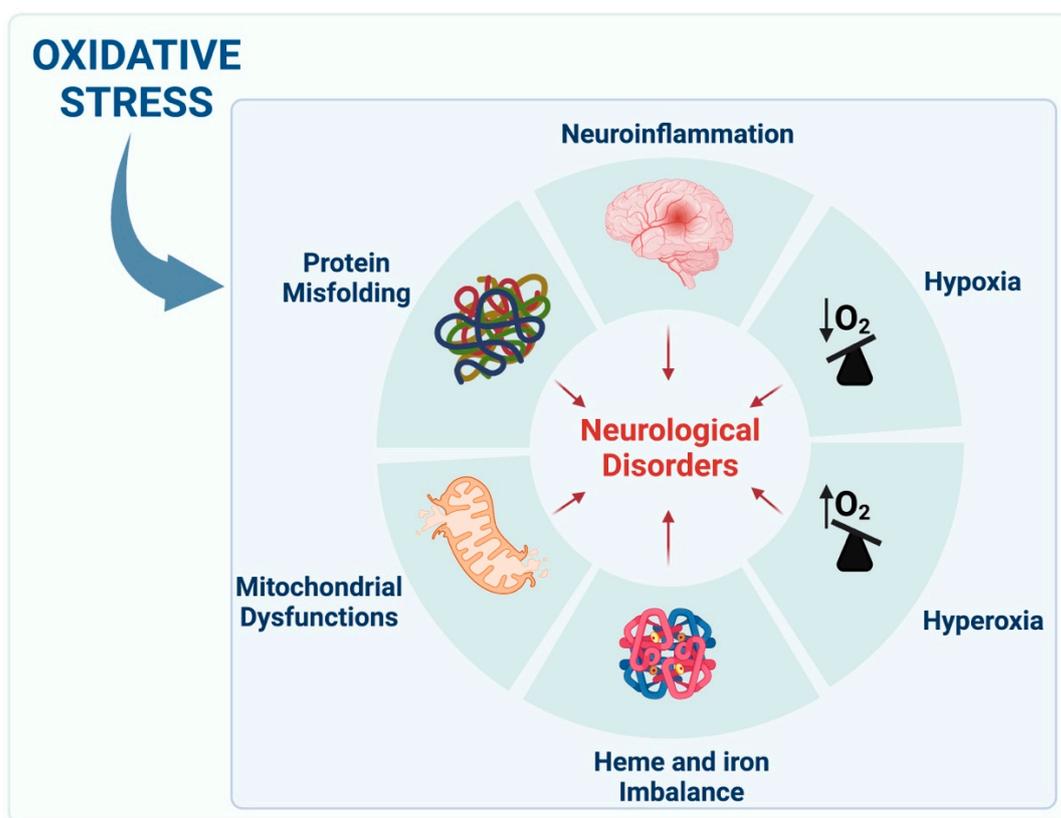
**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Oxygen is a molecule that is fundamental for life, as its action in numerous cellular processes can alter gene expression and lead to pathways’ function or dysfunction [1]. The cellular mechanisms that involve an oxidation-reduction reaction are termed redox signaling, and these are the key processes that govern most aspects of a cell’s life [2].

The brain is an entirely aerobic organ, and for this reason, it is extremely sensitive to oxygen balance [3]. The resting metabolic rate is defined as the energy needed by the organism when at rest, and 20–25% of this energy is required for brain functioning [3]. Indeed, to maintain the ionic equilibria and neurotransmitters uptake necessary for neuronal

communication, there is the need for a high rate of ATP formation and consumption [3]. Oxygen storage in the brain is limited, and without a blood supply, cerebral metabolism would be sustained for only 1 s [3,4]. The condition of low oxygen levels is known as hypoxia [5]. On the contrary, a state of excess supply of  $O_2$  in tissues and organs is known as hyperoxia [6]. Aberrations in oxygen levels can often lead to pathological states, and hypoxia is a primary cause of clinical impairment in stroke and traumatic brain injury [1,3]. Processes that are characterized by alterations in oxygen metabolism and redox signaling can be the primary causes of neurological diseases [7]. These are strictly related to the production of radical oxygen species (ROS), mitochondria abnormalities, protein misfolding and neuroinflammation [8]. Remarkably, aberrations in these processes can lead to the insurgence of neurological disorders (Figure 1).



**Figure 1.** Schematic representation of the alterations in redox mechanisms that can lead to numerous aberrant processes, and ultimately lead to neurological disorders.

## 2. Cellular Pathways of Redox Imbalance

Redox signaling processes concern all these pathways, which present an increase or decrease of reactive species [9]. Herein, we present a brief overview of the main cellular pathways affected by these processes, which will be subsequently addressed more in-depth for their correlation with the pathogenesis of neurological disorders.

### 2.1. Hypoxia

Hypoxia is defined as a condition of decreased oxygen levels, and it is a common pathophysiological condition that can affect brain metabolism [6]. With long term exposure to hypoxia, the mitochondrial volume density decreases and the expression of the electron transport chain complexes is decreased, with a subsequent effect on neuronal metabolism [10]. A significant increase of the exploration of oxygen sensing started from the discovery of erythropoietin (EPO), a kidney-derived glycoprotein mainly induced in hypoxia conditions by both hypoxia-inducible factor-1 and -2 (HIF-1 and HIF-2,

respectively) [11]. EPO has been found to mediate its neuroprotective effect through its receptor EPOR, especially in hypoxic conditions [6,12,13]. Indeed, even if the roles of EPO are not limited to the brain, its action in this organ is surely fundamental, as it has been found to promote and stimulate neurogenesis [14–16]. Its neuroprotective role can be also correlated with its action as an inhibitor of ROS production, as well as its function in counteracting oxidative stress [11].

Mild or important cognitive impairment is also known to be a consequence of the hypoxic damage that takes place during neurological impairments, such as ischemic strokes and traumatic brain injuries. The role of the relative hyperoxia induced by reperfusion in those cases is an interesting subject of ongoing studies [17,18]. Interestingly, oxidative damage due to cervical spinal cord injury (SCI) can also exacerbate muscle atrophy and weakness in the disease. In this case, hyperbaric oxygen and subsequent ROS generation has been found to stimulate adaptations of the diaphragm oxidative capacity, and this results in a reduction of oxidative stress and inflammation [19].

An important aspect to consider when describing hypoxia is the biochemical mechanism that leads to redox imbalance. Specifically, if the oxygen supply becomes inadequate, the tricarboxylic acid cycle's (TCA) functions are inhibited and pyruvate derived from glycolysis accumulates [20]. In this condition, pyruvate is converted to lactate through a reaction that provides the regeneration of nicotinamide adenine dinucleotide (NAD) for the continuance of glycolysis [21]. Since the ratio of lactate to pyruvate increases, this ratio has been proposed as a mechanism for assessing hypoxia [20]. Glycolysis produces ATP from ADP but in an inefficient way, as 1 mole of glucose-6-phosphate metabolized to pyruvate produces just 2 moles of ATP instead of the 36 moles that could be produced by the subsequent metabolism of pyruvate in the TCA [22]. All of these events culminate with the failure of electrons transport and the subsequent accumulation of hydrogen ions, which cause redox imbalance and tissue acidosis [23].

## 2.2. Hyperoxia and ROS Production

Opposite to hypoxia, hyperoxia is defined as a condition of increased oxygen levels. It stimulates ROS formation, such as hydrogen peroxide, hydroxyl radicals and superoxide anions, which can affect signaling pathways by directly modifying regulatory proteins through carbonylation, formation of disulphide bonds, glutathionylation and nitrosylation [24]. Molecular oxygen has two unpaired electrons in separate orbitals in its outer electron shell. An increase in oxygen availability leads to incomplete reduction of  $O_2$ , which produces  $O_2^{\bullet-}$ ,  $HO^{\bullet}$  and  $H_2O_2$ . Although ROS are essential regulators of multiple physiological functions of living organisms, such as signal transduction and gene transcription, their imbalance is a major cause of oxidative damage to cellular macromolecules, resulting in numerous neuronal pathologies [25].

ROS are produced by multiple cellular processes and can be overproduced in response to different stimuli, including xenobiotic compounds, cytokines and bacterial invasion [26]. Mitochondria represent major sources in ROS production due to their role in oxidative ATP production. Indeed, the superoxide radical ( $O_2^{\bullet-}$ ) is produced in various sites of the mitochondria, such as complex I and complex III, which release this molecule into the mitochondrial matrix (MM) [27]. Manganese superoxide dismutase (Mn-SOD) then converts the  $O_2^{\bullet-}$  superoxide radical in MM to hydrogen peroxide ( $H_2O_2$ ), which can be further converted by mitochondrial aconitase to a hydroxyl radical ( $^{\bullet}OH$ ) via a Fenton reaction [28]. Other intracellular structures involved in the production of ROS are the peroxisomes and the endoplasmic reticulum (ER), which generate a wide range of ROS and RNS. Specifically, peroxisomes produce  $O_2^{\bullet-}$  in both the matrix and membrane through the intervention of two enzymes: xanthine oxidoreductase (XOR) and urate oxidase (UO) [29]. XOR catalyzes the formation of uric acid during purine metabolism, which is further converted to allantoin by UO [30]. Regarding the role of ER in ROS production, it has been demonstrated that the accumulation of misfolded proteins in ER during hyperoxia initiates the unfolded protein response (UPR) [31]. UPR triggers ROS production, which

can promote ER stress through the intervention of endoplasmic reticulum oxidoreductin-1 (ERO1). ERO1 catalyzes a double reaction because it leads to the production of  $H_2O_2$  and the conversion of glutathione (GSH) to glutathione disulfide (GSSG) [32]. Accumulation of both  $H_2O_2$  and oxidized glutathione further increase ER stress. The ratio between GSH and GSSG is an essential indicator of the redox status in the ER lumen [33].

### 2.3. Role of the Antioxidant System of Cells in Redox Balance

The antioxidant system is fundamental to maintain cellular homeostasis and counteract oxidative stress. The antioxidant system works by exploiting two mechanisms: non-enzymatic and enzymatic [34]. Among the non-enzymatic defenses, GSH and  $\alpha$ -lipoic acid (ALA) must be mentioned. GSH is a thiol tripeptide formed by L- $\gamma$ -glutamyl-L-cysteinyl-glycine, and is present in every cell, especially in the brain [35]. Besides its scavenging role, GSH assists several processes, such as cellular differentiation and proliferation, apoptosis and neurotransmission [36]. Intracellular glutathione is mostly present in its reduced form, even if it can be oxidized into GSSG by glutathione peroxidase (GPx), while the inverse conversion is performed by the glutathione reductase (GR) [37]. With respect to other organelles, mitochondria contain higher levels of GSH: even if they lack the enzymes required by its biosynthesis, they are able to import the tripeptide thanks to dicarboxylate and 2-oxoglutarate carriers [38]. Under physiological conditions, the ratio of GSH to GSSG hangs in favor of GSH; on the contrary, in the presence of oxidative stress, such as in neurological disorders, the GSH/GSSG ratio sharply decreases. In this context, significant evidence has been reported that the GSH levels in the blood and brain decreased in AD patients, as well as in PD, ALS and autism patients [36]. Interestingly, CNS cells show a different sensitivity to oxidative stress based on their GSH mitochondrial concentration; in fact, astrocytes are more vulnerable to oxidative stress [39].

ALA is an endogenous antioxidant that contains two thiol groups. It is naturally synthesized in mitochondria, where it functions as a cofactor in mitochondrial metabolism and biogenesis, but it can be supplemented by food [40]. Differently from glutathione, in which only the reduced form acts as an antioxidant, in this case, both the oxidized form (LA) and the reduced form of dihydrolipoic acid (DHLA) can neutralize free radicals, chelating heavy metals. Moreover, DHLA can regenerate other antioxidant molecules with low molecular weight, such as GSH. In addition, ALA exhibits anti-inflammatory properties, reducing proinflammatory mediators, such as TNF- $\alpha$ , and increasing anti-inflammatory cytokines, such as IL-10 [41]. Thanks to ALA's amphiphilic properties, it can easily cross the blood–brain barrier, leading to beneficial effects in the CNS. In rat models of PD, it has been demonstrated that ALA was able to reduce 6-OHDA and  $H_2O_2$ , along with nitric oxide, preventing neuronal damage [42].

Besides the GPx mentioned before, superoxide dismutase (SOD) and catalase (CAT) also belong to the first line of enzymatic antioxidant defenses. Three types of SODs can be distinguished in humans, depending on the metal cofactors to which they bind: SOD1, present in the cytosol and in the mitochondrial intermembrane, binds to copper/zinc (Cu/Zn); SOD2 is present at the mitochondrial level and interacts with manganese (Mn); and SOD3, the extracellular form, complexed with copper and zinc [43]. SODs catalyze a dismutase reaction, converting highly reactive superoxide radicals into hydrogen peroxide and molecular oxygen, opposing oxidative and nitrosative stress. However, in neurological disorders such as PD, low levels of SOD in erythrocytes have been observed [44]. Moreover, in the spinal cord mitochondria of ALS mice, accumulation of misfolded mutant SOD1 can occur, causing mitochondrial damage [45].

CAT is a tetrameric enzyme containing four iron–heme groups and it is mainly located in peroxisomes. It breaks down two hydrogen peroxide molecules into one molecule of oxygen and two molecules of water in a two-step reaction [46]. Catalase deficiency or malfunctioning is associated with oxidative stress and human diseases [47]. For example,  $\beta$ -amyloid has been observed to inhibit catalase activity, leading to an increase in oxygen

peroxide. On the contrary, the addition of catalase exhibited protective effects against A $\beta$  by the reduction of oxygen peroxide levels and, ultimately, protein and lipid peroxidation [48].

#### 2.4. Heme Proteins and Iron

Amongst the molecules implicated in oxygen metabolism, heme proteins are fundamental for their role in oxygen delivery, as can be seen with the following formula:

$$\text{Oxygen delivery} = \text{cardiac output} \times \text{systemic oxygen saturation} \times \text{hemoglobin}$$

A biological response to hypoxia to limit the damage to the neuronal system is to increase the demand for iron, as this is a central component of hemoglobin. Hypoxia-inducible factor (HIF)-induced EPO production can increase intraerythrocytic hemoglobin and thus protect neurons from excessive hypoxia and hyperoxia [49]. During hypoxia, hemoglobin undergoes conformational changes that allow the release of oxygen to the tissues, thus increasing oxygen delivery to the cells and preventing damage due to oxygen deprivation. On the contrary, during hyperoxia, hemoglobin alters its affinity for oxygen by binding to oxygen more strongly, thus preventing excessive oxygen delivery and avoiding damage due to oxygen toxicity [50]. Even so, extracellular free hemoglobin and its degradation products (such as heme and free iron) can also lead to an excessive inflammatory immune response and oxidative stress, subsequently interacting with pathological processes in neurodegenerative disorders [51] and iron-induced apoptosis (ferroptosis).

Multiple ischemic and/or hemorrhagic events are the most known situations in which hemoglobin catabolism can play a key role in the damage. From excess heme catabolism, the released iron enhances the Fenton reaction, which increases the oxidative damage induced by hypoxia and ischemia-reperfusion damage [52].

#### 2.5. Mitochondria and ROS

Mitochondria are fundamental organelles that are especially relevant for their role in the production of cell energy, ROS, control of calcium homeostasis and cell death [53,54]. These processes acquire a specific relevance in the brain, as neurons require energy production and calcium availability for synaptic transmission [53,54]. Mitochondria present their own life cycle, which comprises four stages: biogenesis, fusion, fission and degradation [54]. Specifically, an increase in the cell's energetic demands leads to mitochondrial biogenesis. These organelles can then undergo fusion or fission in response to alterations in energy requirements [55]. Lastly, mitochondria that are found to be dysfunctional are removed through the autophagic process termed "mitophagy", where the mitochondria are fragmented and degraded in the lysosomes [56]. In multiple neurological diseases, both gene mutations and environmental factors can influence mitochondrial biogenesis and health.

It is well known that mitochondrial ROS (mROS) are required for a plethora of physiological functions, such as signal transduction, aging, stem cell differentiation and proliferation, wound healing, hypoxic adaptation and insulin signaling [57,58]. The production and neutralization by endogenous antioxidants of mitochondrial ROS (mROS) are tightly regulated processes in order to maintain low levels of mROS [59]. Even if ROS are formed during ATP synthesis by complexes I and III of the electron transport chain [60], they can alter all the complexes of the mitochondrial respiratory chain when they are overproduced [61], generating more ROS. Consequently, the lipids of the inner mitochondrial membrane can undergo lipid peroxidation, leading to permeability of the inner mitochondrial membrane [62]. Moreover, since mitochondrial DNA lacks histones, it tends to be more prone to mutations mediated by ROS [63,64].

#### 2.6. Protein Misfolding

Protein folding has been defined as a set of physical processes by which a newly synthesized polypeptide sequence transforms itself into a highly organized three-dimensional structure, which represents, among all possible conformations, the most stable with the

lowest free-energy [65]. This mechanism depends on the amino-acid sequence intrinsic properties, but it is also due to influences from the cellular environment (e.g., ribosomal protein synthesis, interactions with other proteins or nucleic acids and chaperone-mediated folding) [66]. An increasing number of evidence has demonstrated that the main events in many neurodegenerative diseases include the misfolding, oligomerization and accumulation of cerebral proteins. This process usually leads to cellular dysfunction, loss of synaptic connections and brain damage [67]. Protein misfolding and aggregation seem to be remarkably similar steps in most neurological disorders, even if the multiple disease-associated proteins do not exhibit complementarity in terms of sequence, size, structure or function [68].

The idea that neurotoxicity exclusively depends on insoluble fibrils is still under investigation [69]. Indeed, they may represent a mechanism to escape and limit the pre-fibrillar oligomeric intermediates formation [69]. In Alzheimer's disease (AD), the main constituents of amyloid plaques are  $\beta$ A (1–40) and  $\beta$ A (1–42) peptides, which are deposited extracellularly, whilst neurofibrillary tangles (NFTs) are intracellular aggregates [70]. In Parkinson's disease (PD), the intrinsically disordered  $\alpha$ -synuclein misfolds and leads to the formation of Lewy bodies, particularly in dopaminergic and noradrenergic neurons [71]. Moreover, misfolded disease-associated proteins, mainly  $\alpha$ -syn and  $\beta$ A peptides, exhibit prion-like behavior [72]. In amyotrophic lateral sclerosis (ALS), several polypeptides are inclined to misfold and aggregate, such as Tar DNA binding protein-43 (TDP-43), superoxide dismutase 1 (SOD1) and ubiquilin-2 [73]. Protein aggregation in these diseases will be explained in further detail in the following section. Furthermore, as a result of the poliQ expansion, mutated huntingtin (mHtt) is prone to aggregate and accumulate [74].

In recent years, the destructive effect of oligomers and misfolded proteins has been studied in-depth in order to understand the molecular mechanisms at its base. In this context, new findings have reported that ROS and reactive nitrogen species (RNS) may play a crucial role in amino acid chains' aggregation and protein misfolding, resulting in an another pathological trigger of NDs [75]. Particularly, these molecules can cause post-translational modification (PTMs) in proteins, leading to structural changes that lead to protein destabilization and aggregation. An example of this is represented by both S-nitrosylation and S-glutathionylation in reactive thiols by the protein disulfide isomerase [76]. S-nitrosylation consists of the addition of a nitric oxide to the thiol group of a cysteine, while S-glutathionylation is referred to a glutathione group (GSH) added at the thiol group of a cysteine [77]. These redox PTMs of cysteine residues can affect protein stability and structure, leading to an aberrant neuronal signal transduction pathway [78]. Moreover, biological processes can be affected by  $H_2O_2$ , which is a secondary messenger implicated in redox PMTs, capable of inducing oxidation in cysteine residues of the target protein [79].

### 2.7. Neuroinflammation

The increase in ROS production from endogenous or exogenous sources can induce an aberrant signaling inside the cells, ultimately leading to an excessive inflammatory response in the body. While in physiological conditions, the inflammatory processes represent a defense mechanism and are self-limited, in chronic oxidative stress, pro-inflammatory responses are predominant, leading to neurological dysfunctions [80]. Cellular and immune factors, such as specialized macrophages and lymphocytes, cytokines and pattern-recognition receptors, are the contributing players of neuroinflammation; these proinflammatory mediators are either released locally within the central nervous system (CNS) or engaged from the peripheral system, as a result of the destruction of the blood–brain barrier. This in turn leads to the activation of the glial cells, such as microglia and astrocytes. Therefore, considering neuroinflammation, the instantaneous focus turns to the “specialized” immune system, however, glia, mast cells, infiltrating leukocytes and ROS can lead to inflammatory responses following an injury [81].

Among the cells that participate in the inflammation of the CNS, microglia accounts for 1–16% of the total cell population in the brain [82]. Microglial cells are the main immune brain effectors, supervising their environment in forecast of an insult or damage and exerting immunosurveillance [83]. In homeostatic terms, microglial cells control and regulate both cell death and neurogenesis, contributing to synaptic maturation; when activated, they phagocytose cellular debris, present antigens to T cells and subsequently release cytokines/chemokines [84]. Microglial cells' activation can be induced by several molecules, such as matrix metalloproteinase 3 (MMP-3),  $\alpha$ -synuclein, amyloid beta peptide (A $\beta$ ), neuromelanin and ATP, as well as from cell damage signals and an increase of ROS [85].

Astrocytes account for 20–30% of the brain glial cells. They are crucial contributors of neuroinflammation and they have a role in a wide number of neurodegenerative diseases [86]. Physiologically, astrocytes can exert activities that are essential for neural survival, releasing several neurotrophic factors, maintaining the blood–brain barrier (BBB) integrity and regulating axonal outgrowth and myelination [87]. Injury leads to an increase in astrocyte reactivity, and they can then release chemokines, cytokines and trophic factors [88].

Lastly, the redox alterations can also impact the activation and differentiation of CD4+T cells. In the healthy brain, CD4+T cells are necessary for homeostasis, playing different roles in neuroprotection and neuronal destruction [89]. Th1 and Th17 produce pro-inflammatory cytokines, thus contributing to neuroinflammation through the secretion of pro-inflammatory cytokines and through the enhancement of microglia-mediated neurotoxicity up-regulating ROS and NO microglia release [90].

### 3. Neurodegenerative Diseases

Neurodegenerative diseases (NDs) represent a heterogeneous class of disorders typically characterized by the progressive degeneration on of the CNS or peripheral nervous system. Amongst the most studied NDs affecting the CNS, there are Alzheimer's disease (AD), Parkinson's disease (PD) and amyotrophic lateral sclerosis (ALS). Interestingly, most of these diseases present some common pathways, which all lead to neuronal impairment and, ultimately, cell death [91,92]. Disrupted axonal transport, loss of metabolic support from myelin sheaths and neuroinflammation, along with mitochondrial impairment [93], radical oxygen species and protein aggregation all seem to contribute to neuronal damage [94].

#### 3.1. Alzheimer's Disease

AD is an irreversible, progressive neurodegenerative disease that alters memory and thinking skills, ultimately leading to the inability to carry out simple tasks [95]. It is the most common cause of dementia and typically occurs in patients over 60 years of age [95]. The main pathological hallmarks of AD include the abnormal extracellular accumulation of beta-amyloid peptide (A $\beta$ 42), the intracellular depositions of abnormally phosphorylated tau-tangles and defects in synaptic function; this all leads to increased neuronal death [95,96], as reported in Table 1.

Chronic hypoxia was found to affect multiple pathological aspects of AD, such as amyloid  $\beta$  metabolism, tau phosphorylation, autophagy, neuroinflammation, oxidative stress, endoplasmic reticulum stress and mitochondrial and synaptic dysfunction. All of this can lead to the neurodegenerative process [97]. In response to hypoxia, the HIF-1 $\alpha$  pathway plays a neuroprotective role, limiting this damage through the activation of neuroprotective pathways, such as the ones activated by EPO [1,11]. Moreover, a decrease in hemoglobin transcription (for example, due to clinically silent mutations) was theorized to influence intraerythrocytic and neural hemoglobin, and reduce oxygen, carbon monoxide and nitric oxide transport, all of which are aspects that could be involved in the pathogenesis of AD [50].

**Table 1.** Summarized pathological features related to oxidative stress in adult neurodegenerative disorders.

Pathological Feature	Alzheimer's Disease	Parkinson's Disease	Amyotrophic Lateral Sclerosis
Hypoxia	Aberrant amyloid $\beta$ metabolism [97]	$\beta$ -synuclein accumulation [98]	/
Hyperoxia	Tau polymerization [99,100]	NADPH oxidase activation [101]	Increased markers for lipid and DNA oxidation [102]
Heme proteins and iron	Decrease in hemoglobin transcription [50]	Altered iron metabolism, low SN hemoglobin [103]	/
Mitochondrial dysfunctions	Krebs cycle enzymes deficiencies [104]	Increased mitophagy [105,106]	Impaired mitophagy, h altered mitochondria morphology [107,108]
Neuroinflammation	Microglia activation [109]	Microglia activation, lymphocytes infiltration [110–112]	Inflammatory cytokine production [113]
Protein misfolding	Amyloid plaques formation [114,115]	$\alpha$ -syn aggregation [116]	Cytoplasmic inclusions [117]

Oxidative stress has also been found to be implicated in the pathogenesis of AD and, indeed, ROS production can be related to A $\beta$  plaques. Methionine 35 of the A $\beta$  peptide could be strictly linked to oxidative stress and methionine often serves as a shield against oxidation for enzymes' active sites [118,119]. Peptide–methionine sulfoxide reductase activity appears to be decreased in AD-affected brains, leading to ROS accumulation [120]. Moreover, the oxidation of fatty acids operated by ROS accelerates tau polymerization, and this could serve as a possible link between oxidative stress and the development of fibrillar pathology in AD [99,100]. Tau polymerization also has a genetic-related mechanism, which strongly depends on SOD1 and SOD2 expression. SOD1 is a powerful and ubiquitous antioxidant enzyme expressed in human cells whose main role consists of the protection of cells from the damaging effects of superoxide radicals [121].

Another well-characterized redox-related dysfunction in AD is that concerning mitochondrial health. Alterations in these organelles in AD appear early in the disease, indicating their crucial role in the pathogenesis [104,122]. An impairment in bioenergetics in AD brains was found with the identification of deficiencies in enzymes related to bioenergetic fluxes, such as the pyruvate dehydrogenase complex,  $\alpha$ -ketoglutarate dehydrogenase complex and Krebs cycle enzymes [104]. Moreover, in vivo studies with positron emission tomography (PET) identified a reduced consumption of oxygen in AD brains, along with a reduction in cytochrome oxidase activity [123]. Mitochondria are also necessary for synaptic functioning and they localize at this site in order to buffer calcium levels and provide energy to sustain neurotransmitter release [124]. Even so, it is unclear whether mitochondrial dysfunction is a consequence of amyloidogenic pathology or rather a primary cause for AD [107]. The first hypothesis is based on evidence detailing A $\beta$  aggregation as the primary cause of cellular degeneration through its interaction with specific cellular compartments (including the mitochondria) [70,107]. A $\beta$  plaques can specifically target mitochondria, reducing the activity of electron transport chain enzymes and respiratory chain functions [108]. Interestingly, A $\beta$  administration to cells lacking mitochondrial DNA did not lead to any toxicity, suggesting mitochondria could be crucial in the mediation of amyloidogenic pathology [125]. The hypothesis of mitochondrial cascade dysfunction as the primary cause of AD pathology is based on three cornerstones: firstly, gene inheritance defines the individual's baseline mitochondrial functions; secondly, inherited and environmental factors lead to age-associated mitochondrial changes and degeneration; and lastly, the two previous factors influence the development of AD [107,126]. According to this, mitochondrial function affects APP expression, processing and the accumulation of amyloid plaques [126].

As previously mentioned, amyloid plaques are a primary hallmark of the disease, therefore, it is easy to highlight why protein misfolding can contribute to a redox imbalance. These plaques are formed by multiple proteins, the main one being amyloid- $\beta$  ( $A\beta$ ) [114,115].  $A\beta$  is a ~4-kDa protein that is processed from the amyloid precursor protein (APP) by two different aspartyl proteases, called  $\beta$ - and  $\gamma$ -secretases [127]. The  $\gamma$ -secretases can cleave APP at different sites, and this leads to the production of either 40 or 42 amino acid residues. The 42 amino acid long protein is typically less abundant, but it is more prone to oligomerize and form fibrils. The excessive production of this form of  $A\beta$  could be sufficient to cause early-onset AD [128]. Even so, the exact conformation of soluble  $A\beta$  is unclear and experiments are still needed to determine what conformations of  $A\beta$  aggregates are pathogenic [129]. To this end, an electron microscopy analysis of a post-mortem brain was performed, and the results show that all forms of amyloid plaques can be linked to the neuropathology [130]. Interestingly, both  $A\beta_{40}$  and  $A\beta_{42}$  are present in the fibrillar deposits of neuritic amyloid plaques, whilst diffuse plaques are not considered fibrillar and are mainly composed of  $A\beta_{42}$  [131]. Multiple studies demonstrated that abnormal  $A\beta$  accumulation can trigger tau pathology through the formation of neurofibrillary tangles [132]. Tau protein is encoded by the MAPT gene, localized on chromosome 17, and it is typically produced as a hydrophilic protein that presents large natively unfolded regions. It is especially enriched in the axons of developing and mature neurons [133,134]. Tau protein can be affected by an alternative splicing process that involves the N-terminal projection region and microtubule-binding domain (MBD), and this leads to the production of 4-repeat (4R) and 3-repeat (3R) tau. 3R tau expression is predominantly produced during brain development, whilst the expression of these two isoforms is strictly balanced with a 1:1 ratio in the adult brain [135]. Interestingly, maintenance of this ratio is fundamental and alterations in the 3R and 4R ratio have been implicated in AD, even if the results are conflicting and certain tau tangles-containing brain areas show increased 4R tau protein in some cases [136,137] and 3R tau in others [138]. Tau phosphorylation has also been proposed as the limiting factor in  $A\beta$ -induced neurotoxicity, as loss of either 1 or 2 tau genes protected hybrids against learning and memory deficits and excitotoxicity, which were contrary to those present in hAPPJ20 parental mice strains [139].

Lastly, neuroinflammation has been shown to play a key role in the pathogenesis of AD, mainly through microglia activation [109]. Indeed, these activated cells can interact with amyloid peptides and tau species, influencing disease progression [109]. Recent studies have focused on the potential of microglia targeting in AD, as these could represent an innovative approach to modulate disease progression [140].

### 3.2. Parkinson's Disease

PD is the second most common neurodegenerative disease, with a progressive pathology ultimately leading to motor dysfunction defined as involuntary shaking, weakness and an altered posture [141]. It is characterized by two main hallmarks, which are Lewy bodies (LB), composed of  $\alpha$ -synuclein ( $\alpha$ -syn), and dopaminergic neuronal loss in the substantia nigra pars compacta (SNpc) [142]. Even so, along with these two main processes, dysfunction is also noted in mitochondrial health, oxidative balance, RNA biology and even synaptic functioning and calcium metabolism [142–146] (Table 1).

Amongst the processes concerning redox signaling related to PD pathogenesis, the hypoxia/HIF-1 $\alpha$  signaling pathway was linked to the disease through gene mutations, risk factors, mitochondrial dysfunction, oxidative stress and metabolism impairment [147]. HIF-1 can impact the expression of LRRK2, and hypoxia can induce beta-synuclein accumulation [98]. Moreover, the cytokine EPO is neuroprotective in the disease [11,110,148–152].

Excess oxygen levels can also worsen the development of the disease, with an increase in oxidized lipids, proteins and DNA being present in PD, along with a reduction of glutathione in the SN [153]. Early-stage PD patients present a strong implication of oxidative stress, suggesting this is a crucial aspect even before the insurgence of neuronal loss [154]. ROS can be produced through numerous pathways, including activation of

NADPH oxidase (NOX), cytochrome P-450 oxidase, inflammatory responses and hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) breakdown. NOX could indeed be a major player in the connection between ROS formation and PD pathogenesis, as its function and activity was found impaired in the dopaminergic neurons of PD patients, with NOX1 being increased in PD patients leading to ROS accumulation [101,155]. Interestingly, NOX2-deficient mice demonstrated protected from the neurotoxic treatment of 1-methyl-4-phenyl-1,2,5,6-tetrahydropyridine (MPTP) [156]. Amongst the other proteins involved in oxidative stress in PD, DJ-1 acts as a chaperon protein for redox changes, as well as a marker and sensor for oxidative stress [157]. LRRK2 mutations in neurons also lead to higher vulnerability to toxicity induced by ROS [158]. Indeed, LRRK2 protein was found to increase ROS generation and causes enhanced neurotoxicity [159]. Another crucial player is the transcription factor nuclear factor erythroid 2-related factor 2 (Nrf2), known as the main protein that defends against oxidative stress [160]. This transcription factor acts by binding the promoter at the antioxidant response element (ARE) in neuroprotective genes, which include antioxidant enzymes, such as heme oxygenase-1 (HO-1), glutathione cysteine ligase regulatory subunit (GCLC) and glutathione cysteine ligase modulatory subunit (GLCM) [161]. Ultimately, evidence shows that the activation of Nrf2 leads to a decrease in neurodegenerative events, whereas its inhibition exacerbates it [162].

As neural cells have been found to contain hemoglobin, it was found that heme metabolism also plays a role in PD, and this was due to an increase in hemoglobin concentration in neurons of the SN [163]. Even so, the role of heme in the pathogenesis of the disease requires further elucidation, and the degeneration of dopamine neurons in the SNpc in PD is associated with siderotic foci [164]. While some published studies revealed no relation between new PD onset and hemoglobin concentrations [165], the severity of the disease correlates with both iron metabolism and low hemoglobin [103].

PD is one of the NDs with the most implication for mitochondria's health. The involvement of this organelle in PD was first proved in 1983 after drug users self-injected an intravenous solution of MPTP, an inhibitor of mitochondrial chain complex I, which led to the development of parkinsonism [166]. Toxins such as MPTP and rotenone impair the mitochondrial electron transport complex and hinder mitochondrial movement, whilst they also lead to an increase in the mitochondrial permeability transition, ROS generation and nitric oxide synthase activity. The complex I activity results demonstrated impairment not only in the SN, but also in the skeletal muscles, platelets and leukocytes of PD patients [107]. Mitochondrial dysfunctions are present in both sporadic and familial PD, and they can occur from early on in PD pathogenesis [167]. Common PD-causing genes, such as Parkin, DJ, PINK1,  $\alpha$ -SYN and LRRK2, can have pathogenic mutations that directly or indirectly support that mitochondrial dysfunction is implicated in familial PD [167]. PINK1 and Parkin are crucial in this process, and when PINK1 is activated, it recruits Parkin, resulting in the ubiquitination of the outer mitochondrial membrane proteins, which leads to mitophagy [107]. Moreover,  $\alpha$ -syn can impact the mitochondrial protein import machinery and affect complex I activity, DJ-1 can protect neurons against oxidative stress, and LRRK2 can potentiate the pro-fission activity of DRP1 [53,94]. Mitochondrial dysfunction is also relevant for sporadic forms of PD, as evidence obtained in fibroblasts and peripheral blood mononuclear cells (PBMCs) from PD patients identified mitochondrial and lysosomal dysfunctions [105,106]. Interestingly, mitochondria could be crucial regulators of the selective dopaminergic neuronal susceptibility in the SNpc [167]. Indeed, SNpc neurons present an extensive axonal harbor, which leads to a high bioenergetic demand [168]. Moreover, SNpc neurons appear to undergo an increased rate of mitophagy as opposed to dopaminergic neurons present in other areas, such as the ventral tegmental area [115]. High fluxes of cytosolic calcium during neuronal activity are also present in dopaminergic neurons of the SNpc, and modulation of calcium channel CaV1 appears to decrease mitochondrial oxidative stress without altering pacemaker activity [115].

The study of protein misfolding and aggregation in PD has become crucial in the disease, as mutations in the gene encoding for  $\alpha$ -syn (SNCA) cause familial forms of

the disease, and moreover, the protein is a main component of LB, a key driver in PD pathology [116].  $\alpha$ -syn is a member of the synuclein family and is composed of three proteins (including  $\beta$ - and  $\gamma$ -synucleins) all encoded by three different genes. In humans, the predominant cerebral form of  $\alpha$ -syn is a small, soluble protein of 140 residues, representing about 1% of the total neuronal proteins, where it localizes in presynaptic nerve terminals.  $\alpha$ -syn is composed of three domains: an N-terminal region (composed of an 11-amino acid sequence repeat resembling the apolipoproteins  $\alpha$ -helical lipid-binding motif), a central region composed of hydrophobic amino acids (that tends to favor aggregation and, ultimately, amyloid fibrils formation) and a hydrophilic C-terminal region rich in proline, glutamate and aspartate, which could confer chaperone activity to  $\alpha$ -syn [169]. Furthermore, LB are characterized by phosphorylated, nitrated and oxidized  $\alpha$ -syn, and dysregulated post-translational modifications of  $\alpha$ -syn may lead to the formation of pathological inclusions [170]. Genetic evidence also implicates several other proteins related to PD pathogenesis in protein misfolding mechanisms, some of which have been found to be associated with mitochondria [171]. One of these is the enzyme parkin, a ubiquitin E3-ligase encoded by the PARK2 locus. Impaired parkin could lead to a toxic accumulation of its substrates, and missense mutations can impact protein solubility, leading to its aggregation in aggresome-like inclusions [172]. Parkin loss of solubility is also linked to  $\alpha$ -syn accumulation/aggregation, but the precise mechanism is yet to be characterized [173]. The development of all these redox-related phenomenon can also exacerbate neuroinflammation, as this has been reported to act in dopaminergic neural cell death in PD. Indeed, postmortem SN from human PD brains reveal deep activation of microglia and lymphocytes infiltration when compared to controls [110–112].

### 3.3. Amyotrophic Lateral Sclerosis

ALS is a neurodegenerative disease characterized by upper and lower motor neuron loss in the spinal cord, brainstem and motor cortex [174]. The disease presents with motor symptoms that include muscular atrophy and spasticity, leading to paralysis within 3–5 years of onset, with the primary cause of death being respiratory failure [175]. ALS is a complex disease and most of the molecular pathways leading to neuronal degeneration are yet to be characterized [113,175–179]. Disrupted axonal transport, loss of metabolic support from myelin sheaths and neuroinflammation, along with mitochondrial impairment, ROS formation and protein aggregation all seem to contribute to motor neuronal damage [180,181], as summarized in Table 1.

Multiple studies have inferred the role of ROS in ALS, the main evidence being that there is increased oxidative stress in ALS postmortem tissue with respect to control samples [182]. Lipid oxidation markers were also found in spinal cords from sALS patients, whilst they were undetected in controls [102]. 8-hydroxy-2'-deoxyguanosine (8-OHdG), a marker of oxidized DNA, presented an increased abundance in ALS patients' whole cervical spinal cord [183]. Oxidative stress markers were also analyzed in the cerebrospinal fluid (CSF) of early-stage ALS patients; these markers include increased levels of 4-hydroxynonenal (for lipid peroxidation) and ascorbate free radicals [184,185]. Specifically, D90A, A4V and G93A mutations in the SOD1 gene impact SOD1 activity, resulting in the increase of highly toxic hydroxyl radicals, which lead to neurotoxicity and, consequently, neuronal death [100]. It is widely accepted that mutant SOD1 toxicity is the result of an unknown gain of function [186]. When investigating the impact of mutant SOD1 expression in motor neuronal cells with a microarray approach, the results highlighted the downregulation of genes involved in the antioxidant response, such as transcription factor Nrf2, members of the glutathione S-transferase family and peroxiredoxins [187].

Moreover, in ALS, evidence shows that axonal mitochondria have peculiar morphological features, which consist of rounding up of the mitochondria, networks fragmentation and swelling of internal cristae [188,189]. Moreover, high intracellular calcium concentration in degenerating motor neurons interferes with mitochondrial transport, causing fragmentation or morphological changes, bioenergetics dysfunctions and mitochondrial

apoptosis [189,190]. Mitochondria autophagy, known as mitophagy, is impaired in ALS, leading to the irreversible degradation of mitochondria. Physiologically, the autophagy receptor OPTN and its kinase TBK1 are responsible for autophagosome engulfment of damaged mitochondria [191]. Mutations in both OPTN and TBK1 have been linked to ALS, and a loss of function of these genes leads to impaired mitophagy and accumulation of damaged mitochondria [191,192]. In both sALS and fALS patients, the impairment of axonal transport is one of the earliest pathological events in the pathogenesis of the disease [193]. Interestingly, in murine models harboring the SOD1G85R and SOD1G37R mutations, the number of axonal mitochondria is reduced and their distribution is inhomogeneous throughout the axon [194].

ALS also presents, as a main hallmark, cytoplasmic inclusions or aggregates in neuronal cells and oligodendrocytes. Although the spinal cord is the main affected area of the pathology, these inclusions are also found in the brain regions, such as the frontal and temporal cortices, hippocampus and cerebellum [113]. ALS inclusions can be defined as hyaline- or skein-like and they are organized as filaments with a random orientation and covered by fine granules [195]. Another subclass of ALS inclusions is that of Bunina bodies, small ubiquitin-negative inclusions [196]. The main protein present in these aggregates is TDP-43, a DNA- and RNA-binding protein that can impact nuclear RNA metabolism through an alteration of splicing, transcriptional repression, miRNA synthesis, mRNA nucleo-cytoplasmic shuttling and RNA transport [197,198]. As TDP-43 exerts a nuclear function, its localization in healthy neurons is in the nucleus, but ALS pathology leads to a cytoplasmic aggregation of TDP43 and a decrease in its nuclear levels [199]. The TDP43-encoding gene, TARDBP, is mutated in 1–2% of fALS and sALS cases, with mutations being localized in the region that encodes for the C-terminal glycine-rich domain of the protein. Interestingly, this domain is implicated in the splicing activity of TDP43 and its ability to interact with other proteins [200]. Furthermore, in ALS patients, the protein is cleaved in hyperphosphorylated C-terminal fragments of 18–26 and 35 kDa, which are aggregation-prone and toxic [201]. fALS-causing mutations were also found in the RNA-binding protein FUS [202]. In contrast to TDP-43, mutant FUS protein does not present post-translational modifications, but it is enriched in the insoluble fraction in the pathology [203]. FUS is also implicated in processes that involve nucleic acids' metabolism, such as DNA repair, transcription, splicing and miRNA processing [197]. Moreover, it is responsible for shuttling mRNA from the nucleus to the cytoplasm, and most ALS-linked FUS mutations reported are localized in the nuclear localization sequence (NLS) of the protein, which ultimately results in the inability of FUS to shuttle back to the nucleus [204,205]. Both FUS and TDP-43 contain prion-like domains enriched in asparagine, glutamine, tyrosine and glycine residues. These regions can be present in two states: an unfolded and an aggregated state, and the aggregation of FUS and TDP-43 appears to be due to alterations in these domains [206,207]. The RNA-binding properties of FUS and TDP-43 are also responsible for the proteins' toxicity, as this activity is essential to create RNA–protein complexes, such as stress granules [208].

The Immune system is also involved in ALS, as it stimulates the production of pro-inflammatory cytokines, such as IFN- $\gamma$  and TNF $\alpha$ , expressing cyclooxygenase-2 (COX-2), which can be found in CSF, serum and urine samples of both types of ALS patients [117].

#### 4. Pediatric Neurological Disorders

Oxidative damage related to an oxygen imbalance can be considered a hallmark of many adult-onset neurodegenerative diseases, including AD, PD and ALS. However, this condition is also observed in many pediatric onset neurodegenerative disorders, including genetic white matter disorders, such as adrenoleukodystrophy (X-ALD) and Pelizaeus–Merzbacher disease (PMD); primary neuronal disorders, such as spinal muscular atrophy (SMA); and neurometabolic conditions, such as mucopolysaccharidoses (MPS) and Pelizaeus–Merzbacher disease (PMD) [209–211].

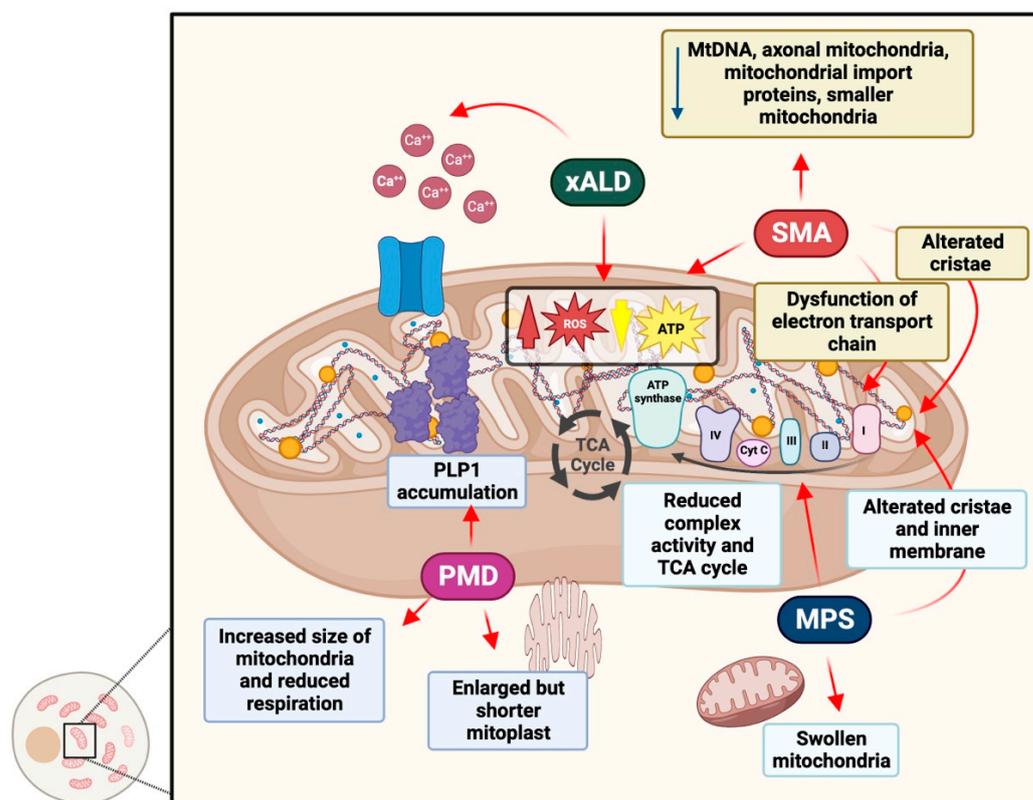
#### 4.1. Adrenoleukodystrophy

X-ALD is a genetic disorder that mostly follows an X-linked inheritance pattern, and it is the most common peroxisomal disorder that can affect both males and females. It presents an estimated birth incidence of about 1/14,700 [212,213]. This disorder is caused by mutations in the ABCD1 gene, encoding for a member of the superfamily of ATP-binding cassette (ABC), transporters also known as ALD [214]. Specifically, ALDP is implicated in the transport of straight-chain very long-chain fatty acid (VLCFA) as CoA esters from the cytosol to the peroxisome lumen, where they are metabolized via  $\beta$ -oxidation [215]. Defects in the activity of the ABCD1 transporter lead to VLCFA excess and the subsequent accumulation in many areas, especially in the brain, spinal cord and adrenal cortex [216]. Clinical presentation of X-ALD phenotypes depends on a progressive central demyelination in the brain and/or slowly progressing axonal degeneration resulting in cognitive decline, spastic quadriparesis and visual and hearing impairment [217]. The exact molecular pathway involved in X-ALD pathogenesis is only partially known, but many contributors of this disease have been identified. These include oxidative stress and mitochondrial dysfunction [218]. Indeed, excess VLCFA induces oxidative stress acting on the unfolded protein response (UPR), a cellular mechanism necessary to maintain oxygen homeostasis [219]. This process is highly regulated by an ER-located transmembrane receptor called protein kinase RNA-like endoplasmic reticulum kinase (PERK), as it has recently emerged that an increase of VLCFA, especially C26:0, induces ER stress in human ALD fibroblasts with a subsequent activation of the PERK pathway [219]. PERK activation leads to the increase of ROS generation and to the decrease of mitochondrial membrane potential [218]. Recent research has demonstrated that this condition is a major contributor of X-ALD pathogenesis in the brain, fibroblasts, erythrocytes and peripheral mononuclear cells (PBMCs) of x-ALD patients [218,220]. Specifically, once produced, mitochondrial ROS may oxidize and inhibit the oxidative phosphorylation system (OXPHOS), inducing a decline in ATP production [221]. ROS promote the opening of the mitochondrial permeability transition pore (mtPTP), leading to a massive efflux of calcium from the mitochondria to the cytosol (Figure 2). As a consequence, calcium overload in the brain leads to the activation of calpains, which dismantle the microtubule structure [222]. In the brain, all these events impair the correct axonal transport along the axon with a loss of axonal continuity and, eventually, axonal disruption [223]. In X-ALD, there is an unbalanced ROS/ATP/Ca<sup>2+</sup> homeostasis, but this mechanism has not been fully characterized yet. However, it is hypothesized that excess in VLCFA, especially C26:0, could alter the permeability of the inner mitochondrial membrane through the increase of the membrane microviscosity, provoking its disruption [224].

#### 4.2. Pelizaeus–Merzbacher Disease

Pelizaeus–Merzbacher disease (PMD; OMIM #312080) is an X-linked recessive hypomyelinating leukodystrophy [225]. PMD is caused by hemizygous mutations in males or heterozygous variants in females with skewed X inactivation in the PLP1 gene, which encodes for both proteolipid protein-1 and the alternative splicing variant DM20, two of the most abundant proteins in the CNS myelin [226,227]. According to the age of onset of symptoms, PMD can be classified as congenital, transitional, classical or intermediate [228]. In any case, when the myelination process fails, axonal damage and dysfunction arise, with deleterious consequences on cognition deficit and motor abilities [229]. From the genetic point of view, PLP1 can be altered in several ways: it can be subjected to gene duplication (triplication or even quintuplication in rare cases), deletions, point mutations or null mutations [228,230]. These mutations reflect different pathological molecular mechanisms and correlate with a wide variety of clinical phenotypes [228]. For example, point mutations, known to cause mutant protein misfolding, lead to PLP1 accumulation in ER, ER stress and, ultimately, cellular toxicity. Differently, PLP1 duplication, associated with a more severe phenotype, is characterized by an abnormal accumulation of PLP1, which fatally affects myelinating oligodendrocytes. Recent evidence suggests that mitochondria are involved in

the promotion of cellular stress and damage [227,231]. In fact, in the PLP-tg66/66 brain, the mouse model that recapitulates the duplication PMD phenotype, a reduction in ATP levels and mitochondrial membrane potential, along with an increase in cytochrome c oxidase, have been reported [231] (Figure 2). In addition, besides an increase in number and size of the mitochondria in oligodendrocytes and axons [231], PLP1 has been observed to co-localize in the mitochondrial membrane, destabilizing extracellular pH [232,233]. Moreover, in both the PLP-tg66/66 spinal cord and in human PMD fibroblasts, Ruiz and collaborators observed increased free radical generation and oxidative damage, with a concomitant decrease in antioxidant defenses, such as glutathione, while the mitochondrial dysfunction was prevented by the administration of the antioxidant NAC [231].



**Figure 2.** Schematic representation of mitochondrial damage mechanisms in some examples of pediatric-onset neurodegenerative diseases.

#### 4.3. Spinal Muscular Atrophy

SMA is a neuromuscular disorder characterized by progressive degeneration of the motor neurons of the anterior horns of the spinal cord, which results in muscle weakness and atrophy [234]. This pathology is caused by biallelic deletions or mutations in the survival motor neuron 1 gene (*SMN1*; 5q13.2), leading to the deficiency of the SMN protein [235]. Due to an inverted duplication on chromosome 5q13, the human genome contains an almost identical gene, *SMN2*, that mainly produces a transcript lacking exon 7, resulting in production of a truncated, less stable SMN protein (85–90%). Approximately 10–15% of the full-length protein is encoded by the *SMN2* gene, thus generating a very small amount of functional SMN protein [236,237]. SMN is a ubiquitous RNA-binding protein, which is involved in collaboration with partner proteins, such as Gemin2–8 [238], in the biogenesis of spliceosomal small nuclear ribonucleoproteins, snRNPs, (which are fundamental for pre-mRNA splicing) and trafficking of mRNAs to axon terminals [239].

Recent evidence suggests that SMA pathology is linked to a decrease in mitochondrial respiration and in the activity of oxidative phosphorylation enzymes, and a concomitant increase in ROS levels [240,241]. This energetic impairment has been observed in various

animal and cell culture models [241–243]. Specifically, the proteome analysis of motor neurons isolated from an SMA mouse model showed disturbed energy homeostasis due to dysfunction of the electron transport chain in the mitochondrial complex I [244]. This resulted in a higher ROS production and lower basal ATP levels that cause an increased protein carbonylation and impaired mRNA translation at the initiation step, contributing to reduced protein synthesis efficiency in SMA motor neurons [244]. Interestingly, supplementation of pyruvate led to an increase in SMN protein levels by reduction of ROS via the mTOR pathway [244].

Moreover, morphological abnormalities of mitochondria, including alterations in cristae, were found in animal models of SMA where smaller mitochondria were linked to reduced SMN protein levels [245,246]. Additionally, the number and density of axonal mitochondria are reduced in murine [247,248] and in vitro [246] models of SMA compared to wild-type ones (Figure 2). Reduced mitochondrial DNA (mtDNA) was also detected in patient muscle tissue [249], along with a decreased expression of mitochondrial import proteins found in patient-derived spinal motor neurons [250] and patient muscle biopsies [249]. In this context, a protein involved in synaptic transmission and already implicated in the SMN pathway, stasimon, interacts with VDAC1, an outer mitochondrial membrane import protein. VDAC1–stasimon interactions may be affected by reduced SMN levels, implicating a mitochondrial protein import in the SMA pathology [251,252]. Interestingly, the typical elongated mitochondrial network may also be reduced in SMA models [246]. This is supported by patient-derived cell culture models of SMA showing reduced mitochondrial trafficking due to the dissociation of SMN interactions between ARX-2 and actin filaments [210,246].

Studies have also reported an alteration in apoptosis regulatory proteins in patients and models of SMA [253]. SMN protein interacts with Bcl2 (the anti-apoptotic and outer mitochondrial membrane protein) to exert a synergistic effect against apoptosis [254]. There is evidence implicating Bcl-2 family members in SMA in cell culture models, animal models and patient tissue [255–257]. Another SMN interactor is p53, a transcription factor involved in cell stress that can both induce apoptosis (nuclear p53) and repress autophagy (cytosolic p53). In a mouse model of SMA, nuclear p53 activity was upregulated [258], while the SMN/p53 interaction correlates with SMA disease severity in patient-derived fibroblasts [259]. Finally, another apoptosis-inducing protein is ZPR1, which may impact disease severity [260].

Overall, these changes indicate mitochondrial impairment in SMA disease.

Evidence for neuroinflammation in SMA pathogenesis remains elusive, despite its likely contribution to disease onset and/or progression. Increased astrogliosis was observed in autopsies of patients and in a murine model, while microglia and T-cells have never been thoroughly investigated [261–263]. Studies have shown abnormalities in immune organs (e.g., spleen) and T-cell alterations in SMA mouse models, resulting in an abnormal neuroinflammatory response and exacerbation of the disease [264]. Importantly, functional studies investigating the status of T-cells and their protective or cytotoxic function in SMA may provide us insight on neuroinflammation in the pathogenesis of SMA.

#### 4.4. Mucopolysaccharidoses

Mucopolysaccharidoses (MPS) are a group of inherited disorders belonging to the lysosomal storage diseases. Eleven subtypes of MPS are currently known, distinguishable for the defective or absent enzyme involved in the abnormal lysosomal accumulation of glycosaminoglycans (GAGs) [265]. Physiologically, GAGs, present in every mammalian tissue [266], play a pivotal role not only in structural scaffolding, but also in several metabolic pathways: regulation of cell growth, proliferation, cell adhesion, anticoagulation and wound repair [265]. However, even if the pathophysiology of MPS has not been fully clarified, inflammation, oxidative stress and mitochondrial dysfunction contribute to the disease progression in a cross-linked cascade of events.

It has been reported that a GAG structure, mimicking that of lipopolysaccharide (LPS), was able to activate Toll-like receptor 4 (TLR-4), promoting the secretion of pro-inflammatory cytokines in an animal model of MPS [267]. GAG storage leads to abnormal morphology in lysosomes, leading to lysosomal swelling and vacuolation, as reported in MPS mice neurons and glia, as well as in MPS human neural stem cells [268]. Along with altered lysosomal homeostasis, another important source of ROS in MPS diseases is caused by mitochondrial dysfunction [211]. In particular, in MPSIIIC mice brains, an increased number of swollen mitochondria was observed 5 months after birth. These mitochondria were characterized by morphological aberrations, such as the disorganization of the cristae in the inner membrane [269]. In addition, mitochondrial respiration has also been reported to be impaired in the MPSIIIC mouse model. In fact, the activity of complexes II, III and IV and citrate synthase was reduced [269] (Figure 2). Mitochondrial-mediated oxidative stress has been associated with the worsening of the inflammatory process through the release of ROS and RNS in MPS. The NADPH-oxidase complex, specifically expressed by microglia, increases in the MPSIIIB mouse model, inducing macrophage inflammatory protein-1 $\alpha$  and caspase 11, suggesting that neuroinflammation could generate oxidative stress, leading to apoptosis [270]. Similarly, elevated levels of TNF- $\alpha$  were observed in MPSI, II and III patients [271]. A global impairment in the redox status has also been observed in MPS patients: an increase in plasmatic lipid peroxidation and protein oxidation, as well as alterations in erythrocyte SOD and CAT activities have been reported for MPSI, II and IIIB patients [272,273].

Besides GAG accumulation, secondary storage of substances such as glycosphingolipids, phospholipids and cholesterol has been documented [274]. Generally, the abnormal accumulation of metabolites that become toxic in cells and tissues is considered the main cause of the increased generation of ROS [275]. In both MPS animal models and patients' brains, histological analyses revealed that an impaired accumulation of GM2 and GM3 gangliosides occurred [276,277]. Secondary lipid accumulation has also been detected by Nile red staining in stem cells-iPSC derived from MPSIIIB fibroblasts [278]. The storage of gangliosides in the CNS has been linked to compromised neuronal function and survival, preventing calcium uptake within the endoplasmic reticulum [279]. In addition, the sequestration of cholesterol in neurons and glia observed in the MPSIIIA mouse model was associated with an impairment of endosomal transport [280]. It has been demonstrated that abnormal trafficking also has an impact on the autophagy-lysosomal pathway (ALP), the process by which intracellular macromolecules or damaged organelles are degraded [281]. ALP also plays a pivotal role in protein homeostasis and in the removal of protein aggregates. Mice lacking ALP components showed neuronal accumulation of aggregate-prone proteins, leading to neurodegeneration and oxidative stress [282]. Thus, consequent to the degeneration of the autophagy-lysosomal pathway (as described above for PD), insoluble aggregation of  $\alpha$ -syn occurred in neurons of a mouse model of MPSIIIA [283]. Monaco and collaborators reported that  $\alpha$ -syn gradually accumulates together with other amyloid proteins, including prion protein, phosphorylated tau and Ab, mostly into the lysosomes of neuronal cell bodies in MPSIIIA and MPSIIIC mice [284].  $\alpha$ -syn was also found to be accumulated in several brain regions of human MPSIIIB patients [285] and in the cerebral cortex of post-mortem tissues of MPSIIIA patients [286].

## 5. Therapeutic Prospects

Oxygen metabolism is an essential biological step for every vertebrate organism and for this reason, as previously discussed, oxidative stress and processes affected by an oxygen imbalance are the main features of many NDs and PNDs. Indeed, pathological conditions in the brain have been related with increased oxidative stress, which leads to redox imbalance and inflammation [287]. Researchers have tried for years to develop a new strategy to prevent or cure oxidative stress, but only few successes have been reported.

### 5.1. Antioxidants

Antioxidants are exogenous or endogenous molecules that can neutralize ROS and other kinds of free radicals. These molecules are contained in numerous foods, such as flavonoids and phenolic compounds, lipoic acid (thioctic acid), ubiquinone and idebenone,  $\beta$ -carotene and vitamin C [288]. The typical mechanism of antioxidants' action involves direct and indirect pathways. These include scavenging and metal chelating effects, mimicking or upregulating antioxidant enzymes, activation of Nrf2, increasing the activity of sirtuins and inhibition of pro-oxidant enzymes, among others. Recent findings refer to polyphenolics, thiolics and SOD mimetics not only as inducers of heme oxygenase-1 and nitric oxide synthase, but also as activators of Nrf2 and NADPH oxidase inhibitors [288].

These molecules have been widely studied in neurological disorders and have been found to give promising results in animal models of AD [289,290]. Amongst them, vitamin E (alpha tocopherol) acts as an antioxidant molecule partially restoring cognitive functions in individuals with AD [291,292]. Inconclusive results were also obtained in clinical trials with curcumin, which is a polyphenolic compound with antioxidant and anti-inflammatory effects [293]. The compounds that directly target mitochondria include coenzyme Q10, idebenone, creatine, latrepirdine, methylene blue, triterpenoids, curcumin, Ginkgo biloba extract and omega-3 polyunsaturated fatty acids [288]. Multiple studies have tested these compounds using both in vivo and in vitro models of AD, demonstrating their effectiveness in protecting the brain against A $\beta$ -induced oxidative stress, synaptic loss, mitochondrial dysfunction and abnormal calcium homeostasis [294]. Moreover, curcuminoids have been found to attenuate SMA-related processes in patient-derived fibroblasts [295], and, indeed, the use of these molecules has been proposed as therapy for the disease [296]. Antioxidant molecules (specifically N-acetyl-cysteine,  $\alpha$ -lipoic acid and  $\alpha$ -tocopherol) were also found to attenuate symptoms in a murine model of X-adrenoleukodystrophy [222], and this combination was also used in a phase-II pilot study, with positive outcomes [297].

Among the antioxidants, GSH is the most abundant; its nuclear accumulation plays an important role in cell proliferation, oxidative signaling and in the control of the redox state of critical protein sulfhydryls, which are necessary for DNA repair and expression [298]. In the brain, GSH levels decrease with age, which could impact cognitive function. Moreover, a decrease in GSH levels is associated with microglial activation and endothelial dysfunction, both of which can contribute to the impairment of brain function [298]. Since GSH is implicated in different pathways related to AD, different strategies to improve plasma and brain availability have been studied to restore intracellular levels and prevent the alterations observed in AD. GSH synthesis is often restricted by the availability of L-cysteine, which can be obtained by the breakdown of GSH or from proteins, synthesized endogenously, or from diet. As a result, several approaches were explored, including investigating the correlation between diet and GSH tissue levels [299], developing pharmacological L-cysteine prodrugs [300] and administering  $\gamma$ -GC orally [301]. However, to date, clinical trials did not highlight significant restoration of GSH levels [302]. Recently, supplementation with  $\gamma$ -GCS for 3 months was found to be associated with a reduction of brain oxidative stress, neuroinflammation and maintenance of the antioxidant status in an AD mouse model [303]. Thus, given the important role of GSH in biological activities, new pharmacological approaches to increase or to maintain its levels should be addressed. To date, there is only one clinical trial (start: February 2021, end: May 2025) that aims to restore GSH levels and examine the effects on cognitive improvement (NCT04740580) [304]. Specifically, since it has been observed that the depletion of GSH can be restored by using glycine and cysteine (provided as N-acetylcysteine NAC), this early phase 1 trial will compare the effects of 24 weeks of GlyNAC supplementation with respect to an alanine placebo in patients with AD. The trial aims to assess changes in cognition, GSH levels, oxidative stress, brain glucose uptake, brain inflammation and insulin resistance (NCT04740580) [304].

### 5.2. Mitochondria-Targeting Drugs

Recently, mitochondrial malfunction and oxidative damage have gained much more attention, as significant evidence suggests their role in the pathogenic mechanism of NDs and new possible targets of novel treatments for these disorders [305]. For these reasons, the cytokine EPO has also been studied as a neuroprotective potential therapy, as its effects were studied in numerous *in vitro* and *in vivo* ND models and in two clinical trials [152,306]. Several other molecules display mitochondrial protective activity, such as analogues of SOD-CAT, which present beneficial effects in various models of oxidative stress. For example, silent Mn complexes were effective when directly testing mammalian mitochondrial oxidative stress in SOD2 KO mice [307] and they were found to have protective effects against oxidative stress in animal models of several ND, such as PD [308], ALS [309], AD [310] and stroke [311].

Among the new therapeutic approaches, mitochondrial proteins, such as sirtuins, seem to be favorable drug targets, as they actively regulate cell survival during various physiological and pathological conditions [312]. Specifically, sirtuin 3 (SIRT3) is a mitochondrial NAD<sup>+</sup>-dependent protein deacetylase encoded by the nuclear genome, which is most commonly expressed in tissues with high oxidative capacity, such as the brain. This protein is involved in the regulation of all the complexes of the mitochondrial respiratory chain (MRC), thus modulating the typical mitochondrial bioenergetic failure implicated in PD [313]. Upregulation of SIRT3 confers neuroprotective effects in PD and other NDs [314]. Interestingly, activation of SIRT3 by icariin (ICA), a natural flavonoid glucoside isolated from the herb *Epimedium grandiflorum*, was found to be neuroprotective against dopaminergic neuronal cell loss in the rotenone-induced PD rat and cell models [315,316]. Several natural products are capable of stimulating SIRT3 activities. Particularly, resveratrol, a phytoalexin produced by many herbs, upregulates SIRT3 activity by activating SOD2 [317]. It has been demonstrated that resveratrol leads to dopaminergic neuroprotection activity and a decrease of oxidative stress in mitochondrial complex I-deficient PD models [318]. Honokiol is another poly-phenolic compound that modulates SIRT3 activity ameliorating motor impairment and progressive dopaminergic damage in 6-OHDA-lesioned PD mice [319]. Other natural products that modulate SIRT3 activity are kaempferol (which prevents dopaminergic neuron loss by increasing superoxide dismutase and glutathione peroxidase activities) and salidroside [320]. Given the difficulty of diffusion through the mitochondrial membranes, mitochondria-targeted compounds need to have different strategies for delivery, these being active and passive targeting [321]. In active targeting, there are interactions taking place at mitochondrial sites, such as ligand–receptor associations and antigen–antibody binding, and these can be exploited in order to take advantage of the compatibility between the physicochemical properties of the carrier molecules and those of the mitochondrial compartment. Indeed, small molecules can be efficiently targeted in mitochondria with several strategies, such as enclosure in liposomes, conjugation to lipophilic cations [321,322] and incorporation into mitochondria-targeted peptides [323].

In SMA, the molecule olesoxime, which was studied in phase III clinical trials, exerts a neuroprotective role through mitochondria targeting, with improved cellular survival in *in vitro* and *in vivo* models [210,324]. Furthermore, metformin was found to induce mitochondrial biogenesis with anti-inflammatory effects in fibroblasts obtained from patients with adrenoleukodystrophy [325].

### 5.3. Metal Protein Attenuating Compounds

Therapies aimed at targeting transition metal dysregulation are a potential treatment for specific neurological disorders. Metal protein attenuating compounds (MPACs) appear to be an emerging therapeutic approach that can lead to restoration of metal homeostasis, decreased oxidative stress and reversing or slowing disease progression. MPACs compete for binding with redox active metal ions, subsequently preventing oligomerization [326]. Many studies suggest an important role for transition metal ions in AD, enhancing A $\beta$  aggregation and facilitating oxidative stress. Clioquinol (PBT1) is a small lipophilic com-

pound with promising potential in the treatment of NDs due to the demonstrated ability to readily cross the BBB. Specifically, PBT1 oral administration in Tg2576 transgenic mice provoked a 49% decrease in brain A $\beta$  burden compared with non-treated controls [327]. Moreover, the effect of the oral PBT1 treatment in a pilot phase II clinical trial of AD patients showed a statistically substantial prevention of cognitive deterioration [328]. A novel second generation MPAC PBT2 has been synthesized, characterized by a higher solubility and increased BBB permeability as compared with PBT1. The treatment with this new compound in a mouse model of AD counteracted A $\beta$  accumulation, significantly improving cognitive performance [329]. The copper/zinc/calcium chelators, DP-109 and DP-460, were shown to be protective in ameliorating symptomatology related to ALS in a mouse transgenic model expressing G93A-mutated SOD1 [330]. Both chelators were found to improve motor activity associated with a decreased loss of lumbar spinal neurons, reduced reactive gliosis and markers of oxidative stress, as well as extend survival times of the treated mice by approximately 10% when compared with controls [331].

#### 5.4. Opioids or Cannabinoids

The endocannabinoid system is fundamental in regulating myenteric neuron activity and vagal and sympathetic nerve function. For this reason, alterations in this system are found in both experimental models and post-mortem brain samples from patients with several neurological diseases [332]. Cannabinoids can primarily act by counteracting the infiltration of peripheral immune cells to the CNS, which are directly involved in the shift of the phenotypes of microglia and infiltrating macrophages from pro-inflammatory to anti-inflammatory [332]. For these anti-inflammatory effects, cannabinoids have been studied in the context of neurodegenerative diseases. In vitro and in vivo models of A $\beta$ -induced neurotoxicity indicated that CBD can protect against A $\beta$ -induced insults, as it reduces oxidative stress, tau phosphorylation and expression of the inducible nitric oxide synthase via the WNT- $\beta$ -catenin pathway [333]. Studies performed in 6-hydroxydopamine (6-OHDA)-treated rats and lipopolysaccharide-treated rats highlighted the anti-parkinsonian effects of THC and CBD [334], probably due to the antioxidant properties of these cannabinoids. It was also demonstrated in a double-blind trial of CBD use in patients with PD that the highest dose tested (300 mg/day) improved the patients' quality of life and reduced levodopa-induced dyskinesia in PD [335]. Regarding ALS, a selective CB2 agonist slowed disease progression in SOD1 mice, suggesting that CB2 has a protective role in ALS [336]. However, since clinical tests of cannabinoids in patients with neurological disorders are limited, and most of the mechanisms of action behind these results are still unclear [332].

#### 5.5. Non-Pharmacological Interventions

Non-pharmacological treatments and lifestyle interventions, including exercise and caloric restriction, are becoming more and more relevant for their overall positive effect on health and lifespan [337]. Indeed, the Alzheimer's Association found that regular physical exercise is a key strategy to reduce the risk of cognitive decline and the development of dementia, as regular physical activity is positively correlated with reduced oxidative stress, increased antioxidant capacity, increased anti-inflammatory effects, reduced levels of ceramides that are elevated in AD, improved A $\beta$  clearance associated with the upregulating A $\beta$  transporters and induced neurogenesis [338]. Even so, research is still needed to clarify the molecular mechanisms implicated in this positive effect, and a better understanding of lifestyle modifications is needed to develop efficacious therapeutic strategies for AD.

In the last few years, major advances in gene therapy research for neurological disorders have also been realized. Through the overexpression of pro-survival growth factors or targeting endogenous mutant or wild-type genes associated with disease pathogenesis, various research has highlighted the opportunity to resort to gene therapy against neurological diseases [339]. Interestingly, lentivirus-mediated expression of the lysosomal cysteine protease cathepsin B in APP transgenic mice reduced pre-existing A $\beta$  deposits with promising results [340]. Regarding PD, it has been known that lentivirus-delivered siRNA

against APP beta secretase 1 decreased amyloid plaque levels and neurodegeneration in APP transgenic mice [341]. Gene therapy has also been approved for SMA treatment as a one-time intravenous injection of onasemnogene abeparvovec, which introduces the SMN1 transgene into motor neurons to replace the non-functional SMN1 gene [342]. Moreover, the antisense oligonucleotide targeting the SMN2 gene nusinersen has also been approved. Both therapies have proved beneficial in the treatment of SMA [343,344]. A gene therapy approach has also been proposed in X-ALD using corrected hematopoietic stem cells CD34+, but this alternative treatment can only be applied to a small subset of patients with no severe symptoms [345].

## 6. Conclusions

In conclusion, oxygen presents a critical role in multiple metabolic and physiological processes in the human body, and its imbalance can lead to several pathological consequences, especially in the brain, which is highly sensitive to oxygen equilibrium. Dysfunctions caused by oxygen imbalance can result in hypoxia, hyperoxia, protein misfolding, mitochondria dysfunctions, alterations in heme metabolism and neuroinflammation, ultimately causing neurological alterations in both pediatric and adult life. These disorders share common pathways related to redox imbalance. With this review, we have highlighted the dysfunctions present in adult and pediatric neurodegenerative disorders, emphasizing their underlying redox dysfunction and summarizing the potential therapeutic strategies. Further research is still needed to explore these mechanisms and potentially identify therapeutic strategies effective in the treatment of these disorders.

**Author Contributions:** F.R.: Conceptualization, Writing—original draft, Writing—review & editing, Investigation; C.B., E.M., A.M., L.E. and G.C.: Investigation, Writing—original draft, Writing—review and editing; L.M., S.O., E.B. and D.T.: Investigation, Writing—original draft; C.C., S.C. (Salvatore Cuzzocrea) and G.Z.: Funding acquisition, Writing—review & editing; E.E. and I.P.: Project coordination, Writing—review & editing; S.C. (Stephana Carelli): Conceptualization, Writing—original draft, Writing—review and editing, Project coordination. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was supported by a grant from the Pediatric Clinical Research Center Fondazione “Romeo and Enrica Invernizzi” to G.Z. and S.C. (Stephana Carelli) and by the Italian Ministry of Health (GR-2019-12368701) to D.T. and C.C.

**Acknowledgments:** S.C. (Stephana Carelli) acknowledges Neurogel en Marche Association for supporting the research on neurodegenerative diseases.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Wilson, J.W.; Shakir, D.; Batie, M.; Frost, M.; Rocha, S. Oxygen-Sensing Mechanisms in Cells. *FEBS J.* **2020**, *287*, 3888–3906. [[CrossRef](#)] [[PubMed](#)]
2. Forman, H.J.; Ursini, F.; Maiorino, M. An Overview of Mechanisms of Redox Signaling. *J. Mol. Cell. Cardiol.* **2014**, *73*, 2–9. [[CrossRef](#)] [[PubMed](#)]
3. Bailey, D.M. Oxygen, Evolution and Redox Signalling in the Human Brain; Quantum in the Quotidian. *J. Physiol.* **2018**, *597*, 15–28. [[CrossRef](#)]
4. Leithner, C.; Rojl, G. The Oxygen Paradox of Neurovascular Coupling. *J. Cereb. Blood Flow Metab.* **2014**, *34*, 19–29. [[CrossRef](#)] [[PubMed](#)]
5. Sharp, F.R.; Bernaudin, M. HIF1 and Oxygen Sensing in the Brain. *Nat. Rev. Neurosci.* **2004**, *5*, 437–448. [[CrossRef](#)]
6. Terraneo, L.; Paroni, R.; Bianciardi, P.; Giallongo, T.; Carelli, S.; Gorio, A.; Samaja, M. Brain Adaptation to Hypoxia and Hyperoxia in Mice. *Redox Biol.* **2017**, *11*, 12–20. [[CrossRef](#)]
7. Rey, F.; Messa, L.; Maghraby, E.; Casili, G.; Ottolenghi, S.; Barzaghini, B.; Raimondi, M.T.; Cereda, C.; Cuzzocrea, S.; Zuccotti, G.; et al. Oxygen Sensing in Neurodegenerative Diseases: Current Mechanisms, Implication of Transcriptional Response and Pharmacological Modulation. *Antioxid. Redox Signal.* **2022**, *38*, 160–182. [[CrossRef](#)]
8. Fatokun, A.A.; Stone, T.W.; Smith, R.A. Oxidative Stress in Neurodegeneration and Available Means of Protection. *Front. Biosci.* **2008**, *13*, 3288–3311. [[CrossRef](#)]

9. Margaritelis, N.V.; Chatzinikolaou, P.N.; Chatzinikolaou, A.N.; Paschalis, V.; Theodorou, A.A.; Vrabas, I.S.; Kyparos, A.; Nikolaidis, M.G. The Redox Signal: A Physiological Perspective. *IUBMB Life* **2022**, *74*, 29–40. [[CrossRef](#)]
10. Murray, A.J.; Horscroft, J.A. Mitochondrial Function at Extreme High Altitude. *J. Physiol.* **2016**, *594*, 1137–1149. [[CrossRef](#)]
11. Rey, F.; Balsari, A.; Giallongo, T.; Ottolenghi, S.; Di Giulio, A.M.; Samaja, M.; Carelli, S. Erythropoietin as a Neuroprotective Molecule: An Overview of Its Therapeutic Potential in Neurodegenerative Diseases. *ASN Neuro* **2019**, *11*, 1759091419871420. [[CrossRef](#)] [[PubMed](#)]
12. Fantacci, M.; Bianciardi, P.; Caretti, A.; Coleman, T.R.; Cerami, A.; Brines, M.; Samaja, M. Carbamylated Erythropoietin Ameliorates the Metabolic Stress Induced in Vivo by Severe Chronic Hypoxia. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 17531–17536. [[CrossRef](#)] [[PubMed](#)]
13. Terraneo, L.; Samaja, M. Comparative Response of Brain to Chronic Hypoxia and Hyperoxia. *Int. J. Mol. Sci.* **2017**, *18*, 1914. [[CrossRef](#)]
14. Shingo, T.; Sorokan, S.T.; Shimazaki, T.; Weiss, S. Erythropoietin Regulates the in Vitro and in Vivo Production of Neuronal Progenitors by Mammalian Forebrain Neural Stem Cells. *J. Neurosci.* **2001**, *21*, 9733–9743. [[CrossRef](#)] [[PubMed](#)]
15. Lombardero, M.; Kovacs, K.; Scheithauer, B.W. Erythropoietin: A Hormone with Multiple Functions. *Pathobiology* **2011**, *78*, 41–53. [[CrossRef](#)]
16. Carelli, S.; Giallongo, T.; Rey, F.; Colli, M.; Tosi, D.; Bulfamante, G.; Di Giulio, A.M.; Gorio, A. Neuroprotection, Recovery of Function and Endogenous Neurogenesis in Traumatic Spinal Cord Injury Following Transplantation of Activated Adipose Tissue. *Cells* **2019**, *8*, 329. [[CrossRef](#)] [[PubMed](#)]
17. Sun, L.; Wolferts, G.; Veltkamp, R. Oxygen Therapy Does Not Increase Production and Damage Induced by Reactive Oxygen Species in Focal Cerebral Ischemia. *Neurosci. Lett.* **2014**, *577*, 1–5. [[CrossRef](#)]
18. Wu, M.-Y.; Yiang, G.-T.; Liao, W.-T.; Tsai, A.P.-Y.; Cheng, Y.-L.; Cheng, P.-W.; Li, C.-Y.; Li, C.-J. Current Mechanistic Concepts in Ischemia and Reperfusion Injury. *Cell. Physiol. Biochem.* **2018**, *46*, 1650–1667. [[CrossRef](#)]
19. Smuder, A.J.; Turner, S.M.; Schuster, C.M.; Morton, A.B.; Hinkley, J.M.; Fuller, D.D. Hyperbaric Oxygen Treatment Following Mid-Cervical Spinal Cord Injury Preserves Diaphragm Muscle Function. *Int. J. Mol. Sci.* **2020**, *21*, 7219. [[CrossRef](#)] [[PubMed](#)]
20. Mcdowall, D.G. II: Biochemical Derangements Associated with Hypoxia and Their Measurement. *Br. J. Anaesth.* **1969**, *41*, 251–256. [[CrossRef](#)]
21. Chen, P.-S.; Chiu, W.-T.; Hsu, P.-L.; Lin, S.-C.; Peng, I.-C.; Wang, C.-Y.; Tsai, S.-J. Pathophysiological Implications of Hypoxia in Human Diseases. *J. Biomed. Sci.* **2020**, *27*, 63. [[CrossRef](#)] [[PubMed](#)]
22. Burtcher, J.; Mallet, R.T.; Burtcher, M.; Millet, G.P. Hypoxia and Brain Aging: Neurodegeneration or Neuroprotection? *Ageing Res. Rev.* **2021**, *68*, 101343. [[CrossRef](#)] [[PubMed](#)]
23. Sbodio, J.I.; Snyder, S.H.; Paul, B.D. Redox Mechanisms in Neurodegeneration: From Disease Outcomes to Therapeutic Opportunities. *Antioxid. Redox Signal.* **2019**, *30*, 1450–1499. [[CrossRef](#)] [[PubMed](#)]
24. England, K.; Cotter, T.G. Direct Oxidative Modifications of Signalling Proteins in Mammalian Cells and Their Effects on Apoptosis. *Redox. Rep.* **2005**, *10*, 237–245. [[CrossRef](#)]
25. Liochev, S.I. Reactive Oxygen Species and the Free Radical Theory of Aging. *Free Radic. Biol. Med.* **2013**, *60*, 1–4. [[CrossRef](#)]
26. Ray, P.D.; Huang, B.-W.; Tsuji, Y. Reactive Oxygen Species (ROS) Homeostasis and Redox Regulation in Cellular Signaling. *Cell. Signal.* **2012**, *24*, 981–990. [[CrossRef](#)]
27. Okado-Matsumoto, A.; Fridovich, I. Subcellular Distribution of Superoxide Dismutases (SOD) in Rat Liver. *J. Biol. Chem.* **2001**, *276*, 38388–38393. [[CrossRef](#)]
28. Snezhkina, A.V.; Kudryavtseva, A.V.; Kardymon, O.L.; Savvateeva, M.V.; Melnikova, N.V.; Krasnov, G.S.; Dmitriev, A.A. ROS Generation and Antioxidant Defense Systems in Normal and Malignant Cells. *Oxid. Med. Cell. Longev.* **2019**, *2019*, 6175804. [[CrossRef](#)]
29. Di Meo, S.; Reed, T.T.; Venditti, P.; Victor, V.M. Role of ROS and RNS Sources in Physiological and Pathological Conditions. *Oxid. Med. Cell. Longev.* **2016**, *2016*, 1245049. [[CrossRef](#)]
30. Rokka, A.; Antonenkov, V.D.; Soininen, R.; Immonen, H.L.; Pirilä, P.L.; Bergmann, U.; Sormunen, R.T.; Weckström, M.; Benz, R.; Hiltunen, J.K. Pxm2 Is a Channel-Forming Protein in Mammalian Peroxisomal Membrane. *PLoS ONE* **2009**, *4*, e5090. [[CrossRef](#)]
31. Schröder, M.; Kaufman, R.J. ER Stress and the Unfolded Protein Response. *Mutat. Res./Fundam. Mol. Mech. Mutagen.* **2005**, *569*, 29–63. [[CrossRef](#)] [[PubMed](#)]
32. Scriven, P.; Brown, N.J.; Pockley, A.G.; Wyld, L. The Unfolded Protein Response and Cancer: A Brighter Future Unfolding? *J. Mol. Med.* **2007**, *85*, 331–341. [[CrossRef](#)] [[PubMed](#)]
33. Zeeshan, H.; Lee, G.; Kim, H.-R.; Chae, H.-J. Endoplasmic Reticulum Stress and Associated ROS. *Int. J. Mol. Sci.* **2016**, *17*, 327. [[CrossRef](#)] [[PubMed](#)]
34. Salim, S. Oxidative Stress and the Central Nervous System. *J. Pharmacol. Exp. Ther.* **2017**, *360*, 201–205. [[CrossRef](#)]
35. Zitka, O.; Skalickova, S.; Gumulec, J.; Masarik, M.; Adam, V.; Hubalek, J.; Trnkova, L.; Kruseova, J.; Eckschlagler, T.; Kizek, R. Redox Status Expressed as GSH:GSSG Ratio as a Marker for Oxidative Stress in Paediatric Tumour Patients. *Oncol. Lett.* **2012**, *4*, 1247–1253. [[CrossRef](#)]
36. Iskusnykh, I.Y.; Zakharova, A.A.; Pathak, D. Glutathione in Brain Disorders and Aging. *Molecules* **2022**, *27*, 324. [[CrossRef](#)] [[PubMed](#)]

37. Booty, L.M.; King, M.S.; Thangaratnarajah, C.; Majd, H.; James, A.M.; Kunji, E.R.S.; Murphy, M.P. The Mitochondrial Dicarboxylate and 2-Oxoglutarate Carriers Do Not Transport Glutathione. *FEBS Lett.* **2015**, *589*, 621–628. [[CrossRef](#)]
38. Wilkins, H.M.; Kirchoff, D.; Manning, E.; Joseph, J.W.; Linseman, D.A. Mitochondrial Glutathione Transport Is a Key Determinant of Neuronal Susceptibility to Oxidative and Nitrosative Stress. *J. Biol. Chem.* **2013**, *288*, 5091–5101. [[CrossRef](#)]
39. Solmonson, A.; DeBerardinis, R.J. Lipoic Acid Metabolism and Mitochondrial Redox Regulation. *J. Biol. Chem.* **2018**, *293*, 7522–7530. [[CrossRef](#)]
40. Dos Santos, S.M.; Romeiro, C.F.R.; Rodrigues, C.A.; Cerqueira, A.R.L.; Monteiro, M.C. Mitochondrial Dysfunction and Alpha-Lipoic Acid: Beneficial or Harmful in Alzheimer’s Disease? *Oxid. Med. Cell Longev.* **2019**, *2019*, 8409329. [[CrossRef](#)]
41. de Araújo, D.P.; De Sousa, C.N.S.; Araújo, P.V.P.; Menezes, C.E.D.S.; Sousa Rodrigues, F.T.; Escudeiro, S.S.; Lima, N.B.C.; Patrocínio, M.C.A.; Aguiar, L.M.V.; Viana, G.S.D.B.; et al. Behavioral and Neurochemical Effects of Alpha-Lipoic Acid in the Model of Parkinson’s Disease Induced by Unilateral Stereotaxic Injection of 6-Ohda in Rat. *Evid.-Based Complement. Altern. Med.* **2013**, *2013*, 571378. [[CrossRef](#)] [[PubMed](#)]
42. Wang, Y.; Branicky, R.; Noë, A.; Hekimi, S. Superoxide Dismutases: Dual Roles in Controlling ROS Damage and Regulating ROS Signaling. *J. Cell Biol.* **2018**, *217*, 1915–1928. [[CrossRef](#)] [[PubMed](#)]
43. Baillet, A.; Chantepredrix, V.; Trocmé, C.; Casez, P.; Garrel, C.; Besson, G. The Role of Oxidative Stress in Amyotrophic Lateral Sclerosis and Parkinson’s Disease. *Neurochem. Res.* **2010**, *35*, 1530–1537. [[CrossRef](#)] [[PubMed](#)]
44. Pedrini, S.; Sau, D.; Guareschi, S.; Bogush, M.; Brown, R.H.; Naniche, N.; Kia, A.; Trotti, D.; Pasinelli, P. ALS-Linked Mutant SOD1 Damages Mitochondria by Promoting Conformational Changes in Bcl-2. *Hum. Mol. Genet.* **2010**, *19*, 2974–2986. [[CrossRef](#)] [[PubMed](#)]
45. Siegbahn, P.E.M. Quantum Chemical Studies of Manganese Centers in Biology. *Curr. Opin. Chem. Biol.* **2002**, *6*, 227–235. [[CrossRef](#)] [[PubMed](#)]
46. Nandi, A.; Yan, L.-J.; Jana, C.K.; Das, N. Role of Catalase in Oxidative Stress- and Age-Associated Degenerative Diseases. *Oxid. Med. Cell Longev.* **2019**, *2019*, 9613090. [[CrossRef](#)]
47. Nell, H.J.; Au, J.L.; Giordano, C.R.; Terlecky, S.R.; Walton, P.A.; Whitehead, S.N.; Cechetto, D.F. Targeted Antioxidant, Catalase-SKL, Reduces Beta-Amyloid Toxicity in the Rat Brain. *Brain Pathol.* **2017**, *27*, 86–94. [[CrossRef](#)]
48. van der Kooij, M.A.; Groenendaal, F.; Kavelaars, A.; Heijnen, C.J.; van Bel, F. Neuroprotective Properties and Mechanisms of Erythropoietin in in Vitro and in Vivo Experimental Models for Hypoxia/Ischemia. *Brain Res. Rev.* **2008**, *59*, 22–33. [[CrossRef](#)]
49. Altinoz, M.A.; Guloksuz, S.; Schmidt-Kastner, R.; Kenis, G.; Ince, B.; Rutten, B.P.F. Involvement of Hemoglobins in the Pathophysiology of Alzheimer’s Disease. *Exp. Gerontol.* **2019**, *126*, 110680. [[CrossRef](#)]
50. Atamna, H. Heme Binding to Amyloid-Beta Peptide: Mechanistic Role in Alzheimer’s Disease. *J. Alzheimers Dis.* **2006**, *10*, 255–266. [[CrossRef](#)]
51. Daglas, M.; Adlard, P.A. The Involvement of Iron in Traumatic Brain Injury and Neurodegenerative Disease. *Front. Neurosci.* **2018**, *12*, 981. [[CrossRef](#)] [[PubMed](#)]
52. Rey, F.; Ottolenghi, S.; Zuccotti, G.V.; Samaja, M.; Carelli, S. Mitochondrial Dysfunctions in Neurodegenerative Diseases: Role in Disease Pathogenesis, Strategies for Analysis and Therapeutic Prospects. *Neural. Regen. Res.* **2022**, *17*, 754–758. [[CrossRef](#)] [[PubMed](#)]
53. Gao, J.; Wang, L.; Liu, J.; Xie, F.; Su, B.; Wang, X. Abnormalities of Mitochondrial Dynamics in Neurodegenerative Diseases. *Antioxidants* **2017**, *6*, 25. [[CrossRef](#)]
54. Burté, F.; Carelli, V.; Chinnery, P.F.; Yu-Wai-Man, P. Disturbed Mitochondrial Dynamics and Neurodegenerative Disorders. *Nat. Rev. Neurol.* **2015**, *11*, 11–24. [[CrossRef](#)]
55. Pickles, S.; Vigié, P.; Youle, R.J. Mitophagy and Quality Control Mechanisms in Mitochondrial Maintenance. *Curr. Biol.* **2019**, *28*, R170–R185. [[CrossRef](#)] [[PubMed](#)]
56. Zhang, B.; Pan, C.; Feng, C.; Yan, C.; Yu, Y.; Chen, Z.; Guo, C.; Wang, X. Role of Mitochondrial Reactive Oxygen Species in Homeostasis Regulation. *Redox. Rep.* **2022**, *27*, 45–52. [[CrossRef](#)]
57. Antonucci, S.; Di Lisa, F.; Kaludercic, N. Mitochondrial Reactive Oxygen Species in Physiology and Disease. *Cell Calcium.* **2021**, *94*, 102344. [[CrossRef](#)]
58. Angelova, P.R.; Abramov, A.Y. Role of Mitochondrial ROS in the Brain: From Physiology to Neurodegeneration. *FEBS Lett.* **2018**, *592*, 692–702. [[CrossRef](#)]
59. Selivanov, V.A.; Votyakova, T.V.; Pivtoraiko, V.N.; Zeak, J.; Sukhomlin, T.; Trucco, M.; Roca, J.; Cascante, M. Reactive Oxygen Species Production by Forward and Reverse Electron Fluxes in the Mitochondrial Respiratory Chain. *PLoS Comput. Biol.* **2011**, *7*, e1001115. [[CrossRef](#)]
60. Guo, C.; Sun, L.; Chen, X.; Zhang, D. Oxidative Stress, Mitochondrial Damage and Neurodegenerative Diseases. *Neural. Regen. Res.* **2013**, *8*, 2003–2014. [[CrossRef](#)]
61. Ademowo, O.S.; Dias, H.K.I.; Burton, D.G.A.; Griffiths, H.R. Lipid (per) Oxidation in Mitochondria: An Emerging Target in the Ageing Process? *Biogerontology* **2017**, *18*, 859–879. [[CrossRef](#)] [[PubMed](#)]
62. Nissanka, N.; Moraes, C.T. Mitochondrial DNA Damage and Reactive Oxygen Species in Neurodegenerative Disease. *FEBS Lett.* **2018**, *592*, 728–742. [[CrossRef](#)]
63. Kalra, J. Crosslink between Mutations in Mitochondrial Genes and Brain Disorders: Implications for Mitochondrial-Targeted Therapeutic Interventions. *Neural Regen. Res.* **2023**, *18*, 94. [[CrossRef](#)] [[PubMed](#)]

64. Dobson, C.M. Protein Folding and Misfolding. *Nature* **2003**, *426*, 884–890. [[CrossRef](#)] [[PubMed](#)]
65. Anfinsen, C.B. The Formation and Stabilization of Protein Structure. *Biochem. J.* **1972**, *128*, 737–749. [[CrossRef](#)] [[PubMed](#)]
66. Ross, C.A.; Poirier, M.A. Protein Aggregation and Neurodegenerative Disease. *Nat. Med.* **2004**, *10* (Suppl. S7), S10–S17. [[CrossRef](#)] [[PubMed](#)]
67. Soto, C.; Pritzkow, S. Protein Misfolding, Aggregation, and Conformational Strains in Neurodegenerative Diseases. *Nat. Neurosci.* **2018**, *21*, 1332–1340. [[CrossRef](#)]
68. Cascella, R.; Bigi, A.; Cremades, N.; Cecchi, C. Effects of Oligomer Toxicity, Fibril Toxicity and Fibril Spreading in Synucleinopathies. *Cell. Mol. Life Sci.* **2022**, *79*, 174. [[CrossRef](#)]
69. Hardy, J.; Selkoe, D.J. The Amyloid Hypothesis of Alzheimer’s Disease: Progress and Problems on the Road to Therapeutics. *Science* **2002**, *297*, 353–356. [[CrossRef](#)]
70. Mehra, S.; Sahay, S.; Maji, S.K.  $\alpha$ -Synuclein Misfolding and Aggregation: Implications in Parkinson’s Disease Pathogenesis. *Biochim. Biophys. Acta (BBA)-Proteins Proteom.* **2019**, *1867*, 890–908. [[CrossRef](#)]
71. Busquets, M.A.; Espargaró, A.; Estelrich, J.; Sabate, R. Could  $\alpha$ -Synuclein Amyloid-Like Aggregates Trigger a Prionic Neuronal Invasion? *BioMed Res. Int.* **2015**, *2015*, 172018. [[CrossRef](#)] [[PubMed](#)]
72. McAlary, L.; Plotkin, S.S.; Yerbury, J.J.; Cashman, N.R. Prion-Like Propagation of Protein Misfolding and Aggregation in Amyotrophic Lateral Sclerosis. *Front. Mol. Neurosci.* **2019**, *12*, 262. [[CrossRef](#)] [[PubMed](#)]
73. Shacham, T.; Sharma, N.; Lederkremer, G.Z. Protein Misfolding and ER Stress in Huntington’s Disease. *Front. Mol. Biosci.* **2019**, *6*, 20. [[CrossRef](#)] [[PubMed](#)]
74. Dias, V.; Junn, E.; Mouradian, M.M. The Role of Oxidative Stress in Parkinson’s Disease. *J. Park. Dis.* **2013**, *3*, 461–491. [[CrossRef](#)] [[PubMed](#)]
75. Nakamura, T.; Oh, C.; Zhang, X.; Lipton, S.A. Protein S-Nitrosylation and Oxidation Contribute to Protein Misfolding in Neurodegeneration. *Free. Radic. Biol. Med.* **2021**, *172*, 562–577. [[CrossRef](#)]
76. Vrettou, S.; Wirth, B. S-Glutathionylation and S-Nitrosylation in Mitochondria: Focus on Homeostasis and Neurodegenerative Diseases. *Int. J. Mol. Sci.* **2022**, *23*, 15849. [[CrossRef](#)]
77. Nakamura, T.; Tu, S.; Akhtar, M.W.; Sunico, C.R.; Okamoto, S.; Lipton, S.A. Aberrant Protein S-Nitrosylation in Neurodegenerative Diseases. *Neuron* **2013**, *78*, 596–614. [[CrossRef](#)]
78. Lee, Y.M.; He, W.; Liou, Y.-C. The Redox Language in Neurodegenerative Diseases: Oxidative Post-Translational Modifications by Hydrogen Peroxide. *Cell Death Dis.* **2021**, *12*, 58. [[CrossRef](#)]
79. Jayaraj, R.L.; Rodriguez, E.A.; Wang, Y.; Block, M.L. Outdoor Ambient Air Pollution and Neurodegenerative Diseases: The Neuroinflammation Hypothesis. *Curr. Environ. Health Rep.* **2017**, *4*, 166–179. [[CrossRef](#)]
80. Carson, M.J.; Thrash, J.C.; Walter, B. The Cellular Response in Neuroinflammation: The Role of Leukocytes, Microglia and Astrocytes in Neuronal Death and Survival. *Clin. Neurosci. Res.* **2006**, *6*, 237–245. [[CrossRef](#)]
81. Bachiller, S.; Jiménez-Ferrer, I.; Paulus, A.; Yang, Y.; Swanberg, M.; Deierborg, T.; Boza-Serrano, A. Microglia in Neurological Diseases: A Road Map to Brain-Disease Dependent-Inflammatory Response. *Front. Cell. Neurosci.* **2018**, *12*, 488. [[CrossRef](#)] [[PubMed](#)]
82. Becher, B.; Spath, S.; Goverman, J. Cytokine Networks in Neuroinflammation. *Nat. Rev. Immunol.* **2017**, *17*, 49–59. [[CrossRef](#)] [[PubMed](#)]
83. Paolicelli, R.C.; Bolasco, G.; Pagani, F.; Maggi, L.; Scianni, M.; Panzanelli, P.; Giustetto, M.; Ferreira, T.A.; Guiducci, E.; Dumas, L.; et al. Synaptic Pruning by Microglia Is Necessary for Normal Brain Development. *Science* **2011**, *333*, 1456–1458. [[CrossRef](#)]
84. Tang, Y.; Le, W. Differential Roles of M1 and M2 Microglia in Neurodegenerative Diseases. *Mol. Neurobiol.* **2016**, *53*, 1181–1194. [[CrossRef](#)] [[PubMed](#)]
85. Pelvig, D.P.; Pakkenberg, H.; Stark, A.K.; Pakkenberg, B. Neocortical Glial Cell Numbers in Human Brains. *Neurobiol. Aging* **2008**, *29*, 1754–1762. [[CrossRef](#)]
86. Kiray, H.; Lindsay, S.L.; Hosseinzadeh, S.; Barnett, S.C. The Multifaceted Role of Astrocytes in Regulating Myelination. *Exp. Neurol.* **2016**, *283*, 541–549. [[CrossRef](#)]
87. Sofroniew, M.V.; Vinters, H.V. Astrocytes: Biology and Pathology. *Acta Neuropathol.* **2010**, *119*, 7–35. [[CrossRef](#)]
88. Chen, M.-L.; Yan, B.-S.; Bando, Y.; Kuchroo, V.K.; Weiner, H.L. Latency-Associated Peptide Identifies a Novel CD4+CD25+ Regulatory T Cell Subset with TGFbeta-Mediated Function and Enhanced Suppression of Experimental Autoimmune Encephalomyelitis. *J. Immunol.* **2008**, *180*, 7327–7337. [[CrossRef](#)]
89. Weiner, H.L. A Shift from Adaptive to Innate Immunity: A Potential Mechanism of Disease Progression in Multiple Sclerosis. *J. Neurol.* **2008**, *255* (Suppl. S1), 3–11. [[CrossRef](#)]
90. Chiurchiù, V.; Orlacchio, A.; Maccarrone, M. Is Modulation of Oxidative Stress an Answer? The State of the Art of Redox Therapeutic Actions in Neurodegenerative Diseases. *Oxid. Med. Cell. Longev.* **2016**, *2016*, 7909380. [[CrossRef](#)]
91. Garofalo, M.; Pandini, C.; Bordoni, M.; Pansarasa, O.; Rey, F.; Costa, A.; Minafra, B.; Diamanti, L.; Zucca, S.; Carelli, S.; et al. Alzheimer’s, Parkinson’s Disease and Amyotrophic Lateral Sclerosis Gene Expression Patterns Divergence Reveals Different Grade of RNA Metabolism Involvement. *Int. J. Mol. Sci.* **2020**, *21*, 9500. [[CrossRef](#)]
92. Pessoa, J.; Duarte, A. Overcoming Mitochondrial Dysfunction in Neurodegenerative Diseases. *Neural Regen. Res.* **2023**, *18*, 1486. [[CrossRef](#)]

93. Kodavati, M.; Wang, H.; Hegde, M.L. Altered Mitochondrial Dynamics in Motor Neuron Disease: An Emerging Perspective. *Cells* **2020**, *9*, 1065. [[CrossRef](#)]
94. Blennow, K.; de Leon, M.J.; Zetterberg, H. Alzheimer's Disease. *Lancet* **2006**, *368*, 387–403. [[CrossRef](#)]
95. Arroyo-García, L.E.; Bachiller, S.; Ruiz, R.; Boza-Serrano, A.; Rodríguez-Moreno, A.; Deierborg, T.; Andrade-Talavera, Y.; Fisahn, A. Targeting Galectin-3 to Counteract Spike-Phase Uncoupling of Fast-Spiking Interneurons to Gamma Oscillations in Alzheimer's Disease. *Transl. Neurodegener.* **2023**, *12*, 6. [[CrossRef](#)] [[PubMed](#)]
96. Zhang, F.; Niu, L.; Li, S.; Le, W. Pathological Impacts of Chronic Hypoxia on Alzheimer's Disease. *ACS Chem. Neurosci.* **2019**, *10*, 902–909. [[CrossRef](#)] [[PubMed](#)]
97. Butterfield, D.A.; Boyd-Kimball, D. The Critical Role of Methionine 35 in Alzheimer's Amyloid Beta-Peptide (1-42)-Induced Oxidative Stress and Neurotoxicity. *Biochim. Biophys. Acta* **2005**, *1703*, 149–156. [[CrossRef](#)]
98. Murray, I.V.; Sindoni, M.E.; Axelsen, P.H. Promotion of Oxidative Lipid Membrane Damage by Amyloid Beta Proteins. *Biochemistry* **2005**, *44*, 12606–12613. [[CrossRef](#)] [[PubMed](#)]
99. Gabbita, S.P.; Aksenov, M.Y.; Lovell, M.A.; Markesbery, W.R. Decrease in Peptide Methionine Sulfoxide Reductase in Alzheimer's Disease Brain. *J. Neurochem.* **1999**, *73*, 1660–1666. [[CrossRef](#)] [[PubMed](#)]
100. Schweers, O.; Mandelkow, E.M.; Biernat, J.; Mandelkow, E. Oxidation of Cysteine-322 in the Repeat Domain of Microtubule-Associated Protein Tau Controls the in Vitro Assembly of Paired Helical Filaments. *Proc. Natl. Acad. Sci. USA* **1995**, *92*, 8463–8467. [[CrossRef](#)]
101. Sau, D.; De Biasi, S.; Vitellaro-Zuccarello, L.; Riso, P.; Guarnieri, S.; Porrini, M.; Simeoni, S.; Crippa, V.; Onesto, E.; Palazzolo, I.; et al. Mutation of SOD1 in ALS: A Gain of a Loss of Function. *Hum. Mol. Genet.* **2007**, *16*, 1604–1618. [[CrossRef](#)] [[PubMed](#)]
102. Melov, S.; Adlard, P.A.; Morten, K.; Johnson, F.; Golden, T.R.; Hinerfeld, D.; Schilling, B.; Mavros, C.; Masters, C.L.; Volitakis, I.; et al. Mitochondrial Oxidative Stress Causes Hyperphosphorylation of Tau. *PLoS ONE* **2007**, *2*, e536. [[CrossRef](#)] [[PubMed](#)]
103. Wang, W.; Zhao, F.; Ma, X.; Perry, G.; Zhu, X. Mitochondria Dysfunction in the Pathogenesis of Alzheimer's Disease: Recent Advances. *Mol. Neurodegener.* **2018**, *15*, 30. [[CrossRef](#)] [[PubMed](#)]
104. Swerdlow, R.H. Mitochondria and Mitochondrial Cascades in Alzheimer's Disease. *J. Alzheimers Dis.* **2018**, *62*, 1403–1416. [[CrossRef](#)] [[PubMed](#)]
105. Fukuyama, H.; Ogawa, M.; Yamauchi, H.; Yamaguchi, S.; Kimura, J.; Yonekura, Y.; Konishi, J. Altered Cerebral Energy Metabolism in Alzheimer's Disease: A PET Study. *J. Nucl. Med.* **1994**, *35*, 1–6.
106. Cai, Q.; Tammineni, P. Mitochondrial Aspects of Synaptic Dysfunction in Alzheimer's Disease. *J. Alzheimers Dis.* **2017**, *57*, 1087–1103. [[CrossRef](#)]
107. Monzio Compagnoni, G.; Di Fonzo, A.; Corti, S.; Comi, G.P.; Bresolin, N.; Masliah, E. The Role of Mitochondria in Neurodegenerative Diseases: The Lesson from Alzheimer's Disease and Parkinson's Disease. *Mol. Neurobiol.* **2020**, *57*, 2959–2980. [[CrossRef](#)]
108. Casley, C.S.; Canevari, L.; Land, J.M.; Clark, J.B.; Sharpe, M.A. Beta-Amyloid Inhibits Integrated Mitochondrial Respiration and Key Enzyme Activities. *J. Neurochem.* **2002**, *80*, 91–100. [[CrossRef](#)]
109. Cardoso, S.M.; Santos, S.; Swerdlow, R.H.; Oliveira, C.R. Functional Mitochondria Are Required for Amyloid Beta-Mediated Neurotoxicity. *FASEB J.* **2001**, *15*, 1439–1441. [[CrossRef](#)]
110. Swerdlow, R.H.; Burns, J.M.; Khan, S.M. The Alzheimer's Disease Mitochondrial Cascade Hypothesis: Progress and Perspectives. *Biochim. Biophys. Acta* **2014**, *1842*, 1219–1231. [[CrossRef](#)]
111. Glenner, G.G.; Wong, C.W. Alzheimer's Disease: Initial Report of the Purification and Characterization of a Novel Cerebrovascular Amyloid Protein. *Biochem. Biophys. Res. Commun.* **1984**, *120*, 885–890. [[CrossRef](#)] [[PubMed](#)]
112. Masters, C.L.; Simms, G.; Weinman, N.A.; Multhaup, G.; McDonald, B.L.; Beyreuther, K. Amyloid Plaque Core Protein in Alzheimer Disease and Down Syndrome. *Proc. Natl. Acad. Sci. USA* **1985**, *82*, 4245–4249. [[CrossRef](#)] [[PubMed](#)]
113. Haass, C.; Schlossmacher, M.G.; Hung, A.Y.; Vigo-Pelfrey, C.; Mellon, A.; Ostaszewski, B.L.; Lieberburg, I.; Koo, E.H.; Schenk, D.; Teplow, D.B. Amyloid Beta-Peptide Is Produced by Cultured Cells during Normal Metabolism. *Nature* **1992**, *359*, 322–325. [[CrossRef](#)] [[PubMed](#)]
114. Bentahir, M.; Nyabi, O.; Verhamme, J.; Tolia, A.; Horré, K.; Wiltfang, J.; Esselmann, H.; De Strooper, B. Presenilin Clinical Mutations Can Affect Gamma-Secretase Activity by Different Mechanisms. *J. Neurochem.* **2006**, *96*, 732–742. [[CrossRef](#)] [[PubMed](#)]
115. Gouras, G.K.; Olsson, T.T.; Hansson, O.  $\beta$ -Amyloid Peptides and Amyloid Plaques in Alzheimer's Disease. *Neurotherapeutics* **2015**, *12*, 3–11. [[CrossRef](#)] [[PubMed](#)]
116. Takahashi, R.H.; Almeida, C.G.; Kearney, P.F.; Yu, F.; Lin, M.T.; Milner, T.A.; Gouras, G.K. Oligomerization of Alzheimer's Beta-Amyloid within Processes and Synapses of Cultured Neurons and Brain. *J. Neurosci.* **2004**, *24*, 3592–3599. [[CrossRef](#)]
117. Iwatsubo, T.; Odaka, A.; Suzuki, N.; Mizusawa, H.; Nukina, N.; Ihara, Y. Visualization of A Beta 42(43) and A Beta 40 in Senile Plaques with End-Specific A Beta Monoclonals: Evidence That an Initially Deposited Species Is A Beta 42(43). *Neuron* **1994**, *13*, 45–53. [[CrossRef](#)]
118. Götz, J.; Chen, F.; van Dorpe, J.; Nitsch, R.M. Formation of Neurofibrillary Tangles in P3011 Tau Transgenic Mice Induced by A $\beta$  42 Fibrils. *Science* **2001**, *293*, 1491–1495. [[CrossRef](#)]
119. Alexander, G.E.; Chen, K.; Pietrini, P.; Rapoport, S.I.; Reiman, E.M. Longitudinal PET Evaluation of Cerebral Metabolic Decline in Dementia: A Potential Outcome Measure in Alzheimer's Disease Treatment Studies. *Am. J. Psychiatry* **2002**, *159*, 738–745. [[CrossRef](#)]

120. Park, S.Y.; Ferreira, A. The Generation of a 17 KDa Neurotoxic Fragment: An Alternative Mechanism by Which Tau Mediates Beta-Amyloid-Induced Neurodegeneration. *J. Neurosci.* **2005**, *25*, 5365–5375. [[CrossRef](#)]
121. Shoji, M.; Golde, T.E.; Ghiso, J.; Cheung, T.T.; Estus, S.; Shaffer, L.M.; Cai, X.D.; McKay, D.M.; Tintner, R.; Frangione, B. Production of the Alzheimer Amyloid Beta Protein by Normal Proteolytic Processing. *Science* **1992**, *258*, 126–129. [[CrossRef](#)] [[PubMed](#)]
122. Kumar-Singh, S.; Theuns, J.; Van Broeck, B.; Pirici, D.; Vennekens, K.; Corsmit, E.; Cruts, M.; Dermaut, B.; Wang, R.; Van Broeckhoven, C. Mean Age-of-Onset of Familial Alzheimer Disease Caused by Presenilin Mutations Correlates with Both Increased Aβ<sub>42</sub> and Decreased Aβ<sub>40</sub>. *Hum. Mutat.* **2006**, *27*, 686–695. [[CrossRef](#)] [[PubMed](#)]
123. Corder, E.H.; Saunders, A.M.; Strittmatter, W.J.; Schmechel, D.E.; Gaskell, P.C.; Small, G.W.; Roses, A.D.; Haines, J.L.; Pericak-Vance, M.A. Gene Dose of Apolipoprotein E Type 4 Allele and the Risk of Alzheimer’s Disease in Late Onset Families. *Science* **1993**, *261*, 921–923. [[CrossRef](#)] [[PubMed](#)]
124. Prasher, V.P.; Farrer, M.J.; Kessling, A.M.; Fisher, E.M.; West, R.J.; Barber, P.C.; Butler, A.C. Molecular Mapping of Alzheimer-Type Dementia in Down’s Syndrome. *Ann. Neurol.* **1998**, *43*, 380–383. [[CrossRef](#)]
125. Roberson, E.D.; Searce-Levie, K.; Palop, J.J.; Yan, F.; Cheng, I.H.; Wu, T.; Gerstein, H.; Yu, G.Q.; Mucke, L. Reducing Endogenous Tau Ameliorates Amyloid Beta-Induced Deficits in an Alzheimer’s Disease Mouse Model. *Science* **2007**, *316*, 750–754. [[CrossRef](#)]
126. Leng, F.; Edison, P. Neuroinflammation and Microglial Activation in Alzheimer Disease: Where Do We Go from Here? *Nat. Rev. Neurol.* **2021**, *17*, 157–172. [[CrossRef](#)]
127. Althafar, Z.M. Targeting Microglia in Alzheimer’s Disease: From Molecular Mechanisms to Potential Therapeutic Targets for Small Molecules. *Molecules* **2022**, *27*, 4124. [[CrossRef](#)]
128. Parkinson, J. An Essay on the Shaking Palsy. 1817. *J. Neuropsychiatry. Clin. Neurosci.* **2002**, *14*, 223–236. [[CrossRef](#)]
129. Poewe, W.; Seppi, K.; Tanner, C.M.; Halliday, G.M.; Brundin, P.; Volkman, J.; Schrag, A.E.; Lang, A.E. Parkinson Disease. *Nat. Rev. Dis. Prim.* **2017**, *3*, 17013. [[CrossRef](#)]
130. Blesa, J.; Trigo-Damas, I.; Quiroga-Varela, A.; Jackson-Lewis, V.R. Oxidative Stress and Parkinson’s Disease. *Front. Neuroanat.* **2015**, *9*, 91. [[CrossRef](#)]
131. Rey, F.; Pandini, C.; Messa, L.; Launi, R.; Barzaghini, B.; Zangaglia, R.; Raimondi, M.T.; Gagliardi, S.; Cereda, C.; Zuccotti, G.V.; et al. α-Synuclein Antisense Transcript SNCA-AS1 Regulates Synapses- and Aging-Related Genes Suggesting Its Implication in Parkinson’s Disease. *Aging Cell* **2021**, *20*, e13504. [[CrossRef](#)] [[PubMed](#)]
132. Rodriguez, M.; Rodriguez-Sabate, C.; Morales, I.; Sanchez, A.; Sabate, M. Parkinson’s Disease as a Result of Aging. *Aging Cell* **2015**, *14*, 293–308. [[CrossRef](#)] [[PubMed](#)]
133. Hirsch, E.C.; Jenner, P.; Przedborski, S. Pathogenesis of Parkinson’s Disease. *Mov. Disord.* **2013**, *28*, 24–30. [[CrossRef](#)] [[PubMed](#)]
134. Gibson, G.E.; Starkov, A.; Blass, J.P.; Ratan, R.R.; Beal, M.F. Cause and Consequence: Mitochondrial Dysfunction Initiates and Propagates Neuronal Dysfunction, Neuronal Death and Behavioral Abnormalities in Age-Associated Neurodegenerative Diseases. *Biochim. Biophys. Acta* **2010**, *1802*, 122–134. [[CrossRef](#)]
135. Sims, N.R.; Finegan, J.M.; Blass, J.P.; Bowen, D.M.; Neary, D. Mitochondrial Function in Brain Tissue in Primary Degenerative Dementia. *Brain Res.* **1987**, *436*, 30–38. [[CrossRef](#)]
136. Carelli, S.; Giallongo, T.; Viaggi, C.; Gombalova, Z.; Latorre, E.; Mazza, M.; Vaglini, F.; Di Giulio, A.M.; Gorio, A. Grafted Neural Precursors Integrate Into Mouse Striatum, Differentiate and Promote Recovery of Function Through Release of Erythropoietin in MPTP-Treated Mice. *ASN Neuro* **2016**, *8*, 1759091416676147. [[CrossRef](#)]
137. Carelli, S.; Giallongo, T.; Viaggi, C.; Latorre, E.; Gombalova, Z.; Raspa, A.; Mazza, M.; Vaglini, F.; Di Giulio, A.M.; Gorio, A. Recovery from Experimental Parkinsonism by Intrastratial Application of Erythropoietin or EPO-Releasing Neural Precursors. *Neuropharmacology* **2017**, *119*, 76–90. [[CrossRef](#)]
138. Erbaş, O.; Çınar, B.P.; Solmaz, V.; Çavuşoğlu, T.; Ateş, U. The Neuroprotective Effect of Erythropoietin on Experimental Parkinson Model in Rats. *Neuropeptides* **2015**, *49*, 1–5. [[CrossRef](#)]
139. Jang, W.; Park, J.; Shin, K.J.; Kim, J.S.; Youn, J.; Cho, J.W.; Oh, E.; Ahn, J.Y.; Oh, K.W.; Kim, H.T. Safety and Efficacy of Recombinant Human Erythropoietin Treatment of Non-Motor Symptoms in Parkinson’s Disease. *J. Neurol. Sci.* **2014**, *337*, 47–54. [[CrossRef](#)]
140. Rey, F.; Ottolenghi, S.; Giallongo, T.; Balsari, A.; Martinelli, C.; Rey, R.; Allevi, R.; Giulio, A.M.D.; Zuccotti, G.V.; Mazzucchelli, S.; et al. Mitochondrial Metabolism as Target of the Neuroprotective Role of Erythropoietin in Parkinson’s Disease. *Antioxidants* **2021**, *10*, 121. [[CrossRef](#)]
141. Carelli, S.; Giallongo, T.; Gombalova, Z.; Rey, F.; Gorio, M.C.F.; Mazza, M.; Di Giulio, A.M. Counteracting Neuroinflammation in Experimental Parkinson’s Disease Favors Recovery of Function: Effects of Er-NPCs Administration. *J. Neuroinflamm.* **2018**, *15*, 333. [[CrossRef](#)] [[PubMed](#)]
142. Nakabeppu, Y.; Tsuchimoto, D.; Yamaguchi, H.; Sakumi, K. Oxidative Damage in Nucleic Acids and Parkinson’s Disease. *J. Neurosci. Res.* **2007**, *85*, 919–934. [[CrossRef](#)]
143. Ferrer, I.; Martinez, A.; Blanco, R.; Dalfó, E.; Carmona, M. Neuropathology of Sporadic Parkinson Disease before the Appearance of Parkinsonism: Preclinical Parkinson Disease. *J. Neural Transm.* **2011**, *118*, 821–839. [[CrossRef](#)] [[PubMed](#)]
144. Hemmati-Dinarvand, M.; Saedi, S.; Valilo, M.; Kalantary-Charvadeh, A.; Alizadeh Sani, M.; Kargar, R.; Safari, H.; Samadi, N. Oxidative Stress and Parkinson’s Disease: Conflict of Oxidant-Antioxidant Systems. *Neurosci. Lett.* **2019**, *709*, 134296. [[CrossRef](#)] [[PubMed](#)]
145. Choi, D.H.; Cristóvão, A.C.; Guhathakurta, S.; Lee, J.; Joh, T.H.; Beal, M.F.; Kim, Y.S. NADPH Oxidase 1-Mediated Oxidative Stress Leads to Dopamine Neuron Death in Parkinson’s Disease. *Antioxid. Redox Signal.* **2012**, *16*, 1033–1045. [[CrossRef](#)] [[PubMed](#)]

146. Zawada, W.M.; Banninger, G.P.; Thornton, J.; Marriott, B.; Cantu, D.; Rachubinski, A.L.; Das, M.; Griffin, W.S.; Jones, S.M. Generation of Reactive Oxygen Species in 1-Methyl-4-Phenylpyridinium (MPP+) Treated Dopaminergic Neurons Occurs as an NADPH Oxidase-Dependent Two-Wave Cascade. *J. Neuroinflamm.* **2011**, *8*, 129. [[CrossRef](#)]
147. Canet-Avilés, R.M.; Wilson, M.A.; Miller, D.W.; Ahmad, R.; McLendon, C.; Bandyopadhyay, S.; Baptista, M.J.; Ringe, D.; Petsko, G.A.; Cookson, M.R. The Parkinson's Disease Protein DJ-1 Is Neuroprotective Due to Cysteine-Sulfinic Acid-Driven Mitochondrial Localization. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 9103–9108. [[CrossRef](#)]
148. Cooper, O.; Seo, H.; Andrabi, S.; Guardia-Laguarta, C.; Graziotto, J.; Sundberg, M.; McLean, J.R.; Carrillo-Reid, L.; Xie, Z.; Osborn, T.; et al. Pharmacological Rescue of Mitochondrial Deficits in iPSC-Derived Neural Cells from Patients with Familial Parkinson's Disease. *Sci. Transl. Med.* **2012**, *4*, 141ra90. [[CrossRef](#)]
149. Heo, H.Y.; Park, J.M.; Kim, C.H.; Han, B.S.; Kim, K.S.; Seol, W. LRRK2 Enhances Oxidative Stress-Induced Neurotoxicity via Its Kinase Activity. *Exp. Cell Res.* **2010**, *316*, 649–656. [[CrossRef](#)]
150. Senger, D.R.; Li, D.; Jaminet, S.C.; Cao, S. Activation of the Nrf2 Cell Defense Pathway by Ancient Foods: Disease Prevention by Important Molecules and Microbes Lost from the Modern Western Diet. *PLoS ONE* **2016**, *11*, e0148042. [[CrossRef](#)]
151. Mostafavi-Pour, Z.; Ramezani, F.; Keshavarzi, F.; Samadi, N. The Role of Quercetin and Vitamin C in Nrf2-Dependent Oxidative Stress Production in Breast Cancer Cells. *Oncol. Lett.* **2017**, *13*, 1965–1973. [[CrossRef](#)] [[PubMed](#)]
152. Zhang, M.; An, C.; Gao, Y.; Leak, R.K.; Chen, J.; Zhang, F. Emerging Roles of Nrf2 and Phase II Antioxidant Enzymes in Neuroprotection. *Prog. Neurobiol.* **2013**, *100*, 30–47. [[CrossRef](#)] [[PubMed](#)]
153. Graham, J.; Hobson, D.; Ponnampalam, A. High Affinity Hemoglobin and Parkinson's Disease. *Med. Hypotheses* **2014**, *83*, 819–821. [[CrossRef](#)] [[PubMed](#)]
154. Ma, L.; Gholam Azad, M.; Dharmasivam, M.; Richardson, V.; Quinn, R.J.; Feng, Y.; Pountney, D.L.; Tonissen, K.F.; Mellick, G.D.; Yanatori, I.; et al. Parkinson's Disease: Alterations in Iron and Redox Biology as a Key to Unlock Therapeutic Strategies. *Redox Biol.* **2021**, *41*, 101896. [[CrossRef](#)]
155. Umehara, T.; Oka, H.; Nakahara, A.; Shiraiishi, T.; Sato, T.; Matsuno, H.; Komatsu, T.; Omoto, S.; Murakami, H.; Iguchi, Y. Sympathetic Nervous Activity and Hemoglobin Levels in de Novo Parkinson's Disease. *Clin. Auton. Res.* **2020**, *30*, 273–278. [[CrossRef](#)]
156. Deng, Q.; Zhou, X.; Chen, J.; Pan, M.; Gao, H.; Zhou, J.; Wang, D.; Chen, Q.; Zhang, X.; Wang, Q.; et al. Lower Hemoglobin Levels in Patients with Parkinson's Disease Are Associated with Disease Severity and Iron Metabolism. *Brain Res.* **2017**, *1655*, 145–151. [[CrossRef](#)]
157. Langston, J.W.; Ballard, P.; Tetrud, J.W.; Irwin, I. Chronic Parkinsonism in Humans Due to a Product of Meperidine-Analog Synthesis. *Science* **1983**, *219*, 979–980. [[CrossRef](#)]
158. Malpartida, A.B.; Williamson, M.; Narendra, D.P.; Wade-Martins, R.; Ryan, B.J. Mitochondrial Dysfunction and Mitophagy in Parkinson's Disease: From Mechanism to Therapy. *Trends Biochem. Sci.* **2021**, *46*, 329–343. [[CrossRef](#)]
159. Carling, P.J.; Mortiboys, H.; Green, C.; Mihaylov, S.; Sandor, C.; Schwartzenuber, A.; Taylor, R.; Wei, W.; Hastings, C.; Wong, S.; et al. Deep Phenotyping of Peripheral Tissue Facilitates Mechanistic Disease Stratification in Sporadic Parkinson's Disease. *Prog. Neurobiol.* **2020**, *187*, 101772. [[CrossRef](#)]
160. Smith, A.M.; Depp, C.; Ryan, B.J.; Johnston, G.I.; Alegre-Abarrategui, J.; Evetts, S.; Rolinski, M.; Baig, F.; Ruffmann, C.; Simon, A.K.; et al. Mitochondrial Dysfunction and Increased Glycolysis in Prodromal and Early Parkinson's Blood Cells. *Mov. Disord.* **2018**, *33*, 1580–1590. [[CrossRef](#)]
161. Bolam, J.P.; Pissadaki, E.K. Living on the Edge with Too Many Mouths to Feed: Why Dopamine Neurons Die. *Mov. Disord.* **2012**, *27*, 1478–1483. [[CrossRef](#)] [[PubMed](#)]
162. Atik, A.; Stewart, T.; Zhang, J. Alpha-Synuclein as a Biomarker for Parkinson's Disease. *Brain Pathol.* **2016**, *26*, 410–418. [[CrossRef](#)] [[PubMed](#)]
163. George, J.M.; Jin, H.; Woods, W.S.; Clayton, D.F. Characterization of a Novel Protein Regulated during the Critical Period for Song Learning in the Zebra Finch. *Neuron* **1995**, *15*, 361–372. [[CrossRef](#)] [[PubMed](#)]
164. Giasson, B.I.; Duda, J.E.; Murray, I.V.; Chen, Q.; Souza, J.M.; Hurtig, H.I.; Ischiropoulos, H.; Trojanowski, J.Q.; Lee, V.M. Oxidative Damage Linked to Neurodegeneration by Selective Alpha-Synuclein Nitration in Synucleinopathy Lesions. *Science* **2000**, *290*, 985–989. [[CrossRef](#)]
165. El-Agnaf, O.M.; Jakes, R.; Curran, M.D.; Wallace, A. Effects of the Mutations Ala30 to Pro and Ala53 to Thr on the Physical and Morphological Properties of Alpha-Synuclein Protein Implicated in Parkinson's Disease. *FEBS Lett.* **1998**, *440*, 67–70. [[CrossRef](#)]
166. Wang, C.; Tan, J.M.; Ho, M.W.; Zaiden, N.; Wong, S.H.; Chew, C.L.; Eng, P.W.; Lim, T.M.; Dawson, T.M.; Lim, K.L. Alterations in the Solubility and Intracellular Localization of Parkin by Several Familial Parkinson's Disease-Linked Point Mutations. *J. Neurochem.* **2005**, *93*, 422–431. [[CrossRef](#)]
167. Kawahara, K.; Hashimoto, M.; Bar-On, P.; Ho, G.J.; Crews, L.; Mizuno, H.; Rockenstein, E.; Imam, S.Z.; Masliah, E. Alpha-Synuclein Aggregates Interfere with Parkin Solubility and Distribution: Role in the Pathogenesis of Parkinson Disease. *J. Biol. Chem.* **2008**, *283*, 6979–6987. [[CrossRef](#)]
168. Hirsch, E.C.; Hunot, S. Neuroinflammation in Parkinson's Disease: A Target for Neuroprotection? *Lancet Neurol.* **2009**, *8*, 382–397. [[CrossRef](#)]

169. Brochard, V.; Combadière, B.; Prigent, A.; Laouar, Y.; Perrin, A.; Beray-Berthet, V.; Bonduelle, O.; Alvarez-Fischer, D.; Callebert, J.; Launay, J.M.; et al. Infiltration of CD4+ Lymphocytes into the Brain Contributes to Neurodegeneration in a Mouse Model of Parkinson Disease. *J. Clin. Invest.* **2009**, *119*, 182–192. [[CrossRef](#)]
170. Recabarren-Leiva, D.; Alarcón, M. New Insights into the Gene Expression Associated to Amyotrophic Lateral Sclerosis. *Life Sci.* **2018**, *193*, 110–123. [[CrossRef](#)]
171. Robberecht, W.; Philips, T. The Changing Scene of Amyotrophic Lateral Sclerosis. *Nat. Rev. Neurosci.* **2013**, *14*, 248–264. [[CrossRef](#)] [[PubMed](#)]
172. Al-Chalabi, A.; Jones, A.; Troakes, C.; King, A.; Al-Sarraj, S.; van den Berg, L.H. The Genetics and Neuropathology of Amyotrophic Lateral Sclerosis. *Acta Neuropathol.* **2012**, *124*, 339–352. [[CrossRef](#)] [[PubMed](#)]
173. Galán, L.; Gómez-Pinedo, U.; Guerrero, A.; García-Verdugo, J.M.; Matías-Guiu, J. Amyotrophic Lateral Sclerosis Modifies Progenitor Neural Proliferation in Adult Classic Neurogenic Brain Niches. *BMC Neurol.* **2017**, *17*, 173. [[CrossRef](#)]
174. Pandini, C.; Garofalo, M.; Rey, F.; Garau, J.; Zucca, S.; Sproviero, D.; Bordoni, M.; Berzero, G.; Davin, A.; Poloni, T.E.; et al. MINCR: A Long Non-Coding RNA Shared between Cancer and Neurodegeneration. *Genomics* **2021**, *113*, 4039–4051. [[CrossRef](#)]
175. Rey, F.; Marcuzzo, S.; Bonanno, S.; Bordoni, M.; Giallongo, T.; Malacarne, C.; Cereda, C.; Zuccotti, G.V.; Carelli, S. LncRNAs Associated with Neuronal Development and Oncogenesis Are Deregulated in SOD1-G93A Murine Model of Amyotrophic Lateral Sclerosis. *Biomedicines* **2021**, *9*, 809. [[CrossRef](#)] [[PubMed](#)]
176. Andrus, P.K.; Fleck, T.J.; Gurney, M.E.; Hall, E.D. Protein Oxidative Damage in a Transgenic Mouse Model of Familial Amyotrophic Lateral Sclerosis. *J. Neurochem.* **1998**, *71*, 2041–2048. [[CrossRef](#)] [[PubMed](#)]
177. Kiernan, M.C.; Vucic, S.; Cheah, B.C.; Turner, M.R.; Eisen, A.; Hardiman, O.; Burrell, J.R.; Zoing, M.C. Amyotrophic Lateral Sclerosis. *Lancet* **2011**, *377*, 942–955. [[CrossRef](#)]
178. Wijesekera, L.C.; Leigh, P.N. Amyotrophic Lateral Sclerosis. *Orphanet. J. Rare Dis.* **2009**, *4*, 3. [[CrossRef](#)]
179. Islam, M.T. Oxidative Stress and Mitochondrial Dysfunction-Linked Neurodegenerative Disorders. *Neurol. Res.* **2017**, *39*, 73–82. [[CrossRef](#)]
180. Shibata, N.; Nagai, R.; Uchida, K.; Horiuchi, S.; Yamada, S.; Hirano, A.; Kawaguchi, M.; Yamamoto, T.; Sasaki, S.; Kobayashi, M. Morphological Evidence for Lipid Peroxidation and Protein Glycoxidation in Spinal Cords from Sporadic Amyotrophic Lateral Sclerosis Patients. *Brain Res.* **2001**, *917*, 97–104. [[CrossRef](#)]
181. Ferrante, R.J.; Browne, S.E.; Shinobu, L.A.; Bowling, A.C.; Baik, M.J.; MacGarvey, U.; Kowall, N.W.; Brown, R.H.; Beal, M.F. Evidence of Increased Oxidative Damage in Both Sporadic and Familial Amyotrophic Lateral Sclerosis. *J. Neurochem.* **1997**, *69*, 2064–2074. [[CrossRef](#)] [[PubMed](#)]
182. Smith, R.G.; Henry, Y.K.; Mattson, M.P.; Appel, S.H. Presence of 4-Hydroxynonenal in Cerebrospinal Fluid of Patients with Sporadic Amyotrophic Lateral Sclerosis. *Ann. Neurol.* **1998**, *44*, 696–699. [[CrossRef](#)] [[PubMed](#)]
183. Ihara, Y.; Nobukuni, K.; Takata, H.; Hayabara, T. Oxidative Stress and Metal Content in Blood and Cerebrospinal Fluid of Amyotrophic Lateral Sclerosis Patients with and without a Cu, Zn-Superoxide Dismutase Mutation. *Neurol. Res.* **2005**, *27*, 105–108. [[CrossRef](#)]
184. Deng, H.X.; Hentati, A.; Tainer, J.A.; Iqbal, Z.; Cayabyab, A.; Hung, W.Y.; Getzoff, E.D.; Hu, P.; Herzfeldt, B.; Roos, R.P. Amyotrophic Lateral Sclerosis and Structural Defects in Cu,Zn Superoxide Dismutase. *Science* **1993**, *261*, 1047–1051. [[CrossRef](#)] [[PubMed](#)]
185. Kirby, J.; Halligan, E.; Baptista, M.J.; Allen, S.; Heath, P.R.; Holden, H.; Barber, S.C.; Loynes, C.A.; Wood-Allum, C.A.; Lunec, J.; et al. Mutant SOD1 Alters the Motor Neuronal Transcriptome: Implications for Familial ALS. *Brain* **2005**, *128*, 1686–1706. [[CrossRef](#)] [[PubMed](#)]
186. De Vos, K.J.; Chapman, A.L.; Tennant, M.E.; Manser, C.; Tudor, E.L.; Lau, K.F.; Brownlee, J.; Ackerley, S.; Shaw, P.J.; McLoughlin, D.M.; et al. Familial Amyotrophic Lateral Sclerosis-Linked SOD1 Mutants Perturb Fast Axonal Transport to Reduce Axonal Mitochondria Content. *Hum. Mol. Genet.* **2007**, *16*, 2720–2728. [[CrossRef](#)]
187. Sasaki, S.; Iwata, M. Mitochondrial Alterations in the Spinal Cord of Patients with Sporadic Amyotrophic Lateral Sclerosis. *J. Neuropathol. Exp. Neurol.* **2007**, *66*, 10–16. [[CrossRef](#)]
188. Carri, M.T.; D’Ambrosi, N.; Cozzolino, M. Pathways to Mitochondrial Dysfunction in ALS Pathogenesis. *Biochem. Biophys. Res. Commun.* **2017**, *483*, 1187–1193. [[CrossRef](#)]
189. Moore, A.S.; Holzbaur, E.L. Dynamic Recruitment and Activation of ALS-Associated TBK1 with Its Target Optineurin Are Required for Efficient Mitophagy. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, E3349–E3358. [[CrossRef](#)]
190. Evans, C.S.; Holzbaur, E.L.F. Autophagy and Mitophagy in ALS. *Neurobiol. Dis.* **2018**, *122*, 35–40. [[CrossRef](#)]
191. De Vos, K.J.; Hafezparast, M. Neurobiology of Axonal Transport Defects in Motor Neuron Diseases: Opportunities for Translational Research? *Neurobiol. Dis.* **2017**, *105*, 283–299. [[CrossRef](#)] [[PubMed](#)]
192. Vande Velde, C.; McDonald, K.K.; Boukhedimi, Y.; McAlonis-Downes, M.; Lobsiger, C.S.; Bel Hadj, S.; Zandona, A.; Julien, J.P.; Shah, S.B.; Cleveland, D.W. Misfolded SOD1 Associated with Motor Neuron Mitochondria Alters Mitochondrial Shape and Distribution Prior to Clinical Onset. *PLoS ONE* **2011**, *6*, e22031. [[CrossRef](#)] [[PubMed](#)]
193. Lin, W.L.; Dickson, D.W. Ultrastructural Localization of TDP-43 in Filamentous Neuronal Inclusions in Various Neurodegenerative Diseases. *Acta Neuropathol.* **2008**, *116*, 205–213. [[CrossRef](#)] [[PubMed](#)]
194. Okamoto, K.; Mizuno, Y.; Fujita, Y. Bunina Bodies in Amyotrophic Lateral Sclerosis. *Neuropathology* **2008**, *28*, 109–115. [[CrossRef](#)]

195. Arai, T.; Hasegawa, M.; Akiyama, H.; Ikeda, K.; Nonaka, T.; Mori, H.; Mann, D.; Tsuchiya, K.; Yoshida, M.; Hashizume, Y.; et al. TDP-43 Is a Component of Ubiquitin-Positive Tau-Negative Inclusions in Frontotemporal Lobar Degeneration and Amyotrophic Lateral Sclerosis. *Biochem. Biophys. Res. Commun.* **2006**, *351*, 602–611. [[CrossRef](#)] [[PubMed](#)]
196. Lagier-Tourenne, C.; Polymenidou, M.; Cleveland, D.W. TDP-43 and FUS/TLS: Emerging Roles in RNA Processing and Neurodegeneration. *Hum. Mol. Genet.* **2010**, *19*, R46–R64. [[CrossRef](#)]
197. Mackenzie, I.R.; Bigio, E.H.; Ince, P.G.; Geser, F.; Neumann, M.; Cairns, N.J.; Kwong, L.K.; Forman, M.S.; Ravits, J.; Stewart, H.; et al. Pathological TDP-43 Distinguishes Sporadic Amyotrophic Lateral Sclerosis from Amyotrophic Lateral Sclerosis with SOD1 Mutations. *Ann. Neurol.* **2007**, *61*, 427–434. [[CrossRef](#)]
198. D'Ambrogio, A.; Buratti, E.; Stuani, C.; Guarnaccia, C.; Romano, M.; Ayala, Y.M.; Baralle, F.E. Functional Mapping of the Interaction between TDP-43 and HnRNP A2 in Vivo. *Nucleic Acids Res.* **2009**, *37*, 4116–4126. [[CrossRef](#)]
199. Van Deerlin, V.M.; Leverenz, J.B.; Bekris, L.M.; Bird, T.D.; Yuan, W.; Elman, L.B.; Clay, D.; Wood, E.M.; Chen-Plotkin, A.S.; Martinez-Lage, M.; et al. TARDBP Mutations in Amyotrophic Lateral Sclerosis with TDP-43 Neuropathology: A Genetic and Histopathological Analysis. *Lancet Neurol.* **2008**, *7*, 409–416. [[CrossRef](#)]
200. Kwiatkowski, T.J.; Bosco, D.A.; Leclerc, A.L.; Tamrazian, E.; Vanderburg, C.R.; Russ, C.; Davis, A.; Gilchrist, J.; Kasarskis, E.J.; Munsat, T.; et al. Mutations in the FUS/TLS Gene on Chromosome 16 Cause Familial Amyotrophic Lateral Sclerosis. *Science* **2009**, *323*, 1205–1208. [[CrossRef](#)]
201. Neumann, M.; Rademakers, R.; Roeber, S.; Baker, M.; Kretschmar, H.A.; Mackenzie, I.R. A New Subtype of Frontotemporal Lobar Degeneration with FUS Pathology. *Brain* **2009**, *132*, 2922–2931. [[CrossRef](#)] [[PubMed](#)]
202. Zinszner, H.; Sok, J.; Immanuel, D.; Yin, Y.; Ron, D. TLS (FUS) Binds RNA in Vivo and Engages in Nucleo-Cytoplasmic Shuttling. *J. Cell Sci.* **1997**, *110 Pt 15*, 1741–1750. [[CrossRef](#)] [[PubMed](#)]
203. Dormann, D.; Rodde, R.; Edbauer, D.; Bentmann, E.; Fischer, I.; Hruscha, A.; Than, M.E.; Mackenzie, I.R.; Capell, A.; Schmid, B.; et al. ALS-Associated Fused in Sarcoma (FUS) Mutations Disrupt Transportin-Mediated Nuclear Import. *EMBO J.* **2010**, *29*, 2841–2857. [[CrossRef](#)] [[PubMed](#)]
204. Polymenidou, M.; Cleveland, D.W. The Seeds of Neurodegeneration: Prion-like Spreading in ALS. *Cell* **2011**, *147*, 498–508. [[CrossRef](#)]
205. Johnson, B.S.; McCaffery, J.M.; Lindquist, S.; Gitler, A.D. A Yeast TDP-43 Proteinopathy Model: Exploring the Molecular Determinants of TDP-43 Aggregation and Cellular Toxicity. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 6439–6444. [[CrossRef](#)] [[PubMed](#)]
206. Daigle, J.G.; Lanson, N.A.; Smith, R.B.; Casci, I.; Maltare, A.; Monaghan, J.; Nichols, C.D.; Kryndushkin, D.; Shewmaker, F.; Pandey, U.B. RNA-Binding Ability of FUS Regulates Neurodegeneration, Cytoplasmic Mislocalization and Incorporation into Stress Granules Associated with FUS Carrying ALS-Linked Mutations. *Hum. Mol. Genet.* **2013**, *22*, 1193–1205. [[CrossRef](#)] [[PubMed](#)]
207. Liu, J.; Wang, F. Role of Neuroinflammation in Amyotrophic Lateral Sclerosis: Cellular Mechanisms and Therapeutic Implications. *Front. Immunol.* **2017**, *8*, 1005. [[CrossRef](#)]
208. Baarine, M.; Beeson, C.; Singh, A.; Singh, I. ABCD1 Deletion-Induced Mitochondrial Dysfunction Is Corrected by SAHA: Implication for Adrenoleukodystrophy. *J. Neurochem.* **2015**, *133*, 380–396. [[CrossRef](#)]
209. James, R.; Chaytow, H.; Ledahawsky, L.M.; Gillingwater, T.H. Revisiting the Role of Mitochondria in Spinal Muscular Atrophy. *Cell. Mol. Life Sci.* **2021**, *78*, 4785–4804. [[CrossRef](#)]
210. Heon-Roberts, R.; Nguyen, A.L.A.; Pshezhetsky, A.V. Molecular Bases of Neurodegeneration and Cognitive Decline, the Major Burden of Sanfilippo Disease. *JCM* **2020**, *9*, 344. [[CrossRef](#)]
211. Engelen, M.; Kemp, S.; Poll-The, B.-T. X-Linked Adrenoleukodystrophy: Pathogenesis and Treatment. *Curr. Neurol. Neurosci. Rep.* **2014**, *14*, 486. [[CrossRef](#)] [[PubMed](#)]
212. Moser, A.; Jones, R.; Hubbard, W.; Tortorelli, S.; Orsini, J.; Caggana, M.; Vogel, B.; Raymond, G. Newborn Screening for X-Linked Adrenoleukodystrophy. *Int. J. Neonatal Screen.* **2016**, *2*, 15. [[CrossRef](#)] [[PubMed](#)]
213. Roerig, P.; Mayerhofer, P.; Holzinger, A.; Gärtner, J. Characterization and Functional Analysis of the Nucleotide Binding Fold in Human Peroxisomal ATP Binding Cassette Transporters. *FEBS Lett.* **2001**, *492*, 66–72. [[CrossRef](#)] [[PubMed](#)]
214. Roermund, C.W.T.; Visser, W.F.; IJlst, L.; Cruchten, A.; Boek, M.; Kulik, W.; Waterham, H.R.; Wanders, R.J.A. The Human Peroxisomal ABC Half Transporter ALDP Functions as a Homodimer and Accepts Acyl-CoA Esters. *FASEB J.* **2008**, *22*, 4201–4208. [[CrossRef](#)]
215. Powers, J.M. Adreno-Leukodystrophy: A Personal Historical Note. *Acta Neuropathol.* **2005**, *109*, 124–127. [[CrossRef](#)]
216. Ferrer, I.; Aubourg, P.; Pujol, A. General Aspects and Neuropathology of X-Linked Adrenoleukodystrophy: X-Linked Adrenoleukodystrophy Neuropathology. *Brain Pathol.* **2009**, *20*, 817–830. [[CrossRef](#)]
217. Fourcade, S.; Lopez-Erauskin, J.; Galino, J.; Duval, C.; Naudi, A.; Jove, M.; Kemp, S.; Villarroja, F.; Ferrer, I.; Pamplona, R.; et al. Early Oxidative Damage Underlying Neurodegeneration in X-Adrenoleukodystrophy. *Hum. Mol. Genet.* **2008**, *17*, 1762–1773. [[CrossRef](#)]
218. van de Beek, M.-C.; Ofman, R.; Dijkstra, I.; Wijburg, F.; Engelen, M.; Wanders, R.; Kemp, S. Lipid-Induced Endoplasmic Reticulum Stress in X-Linked Adrenoleukodystrophy. *Biochim. Biophys. Acta (BBA)-Mol. Basis Dis.* **2017**, *1863*, 2255–2265. [[CrossRef](#)]
219. Launay, N.; Aguado, C.; Fourcade, S.; Ruiz, M.; Grau, L.; Riera, J.; Guilera, C.; Giròs, M.; Ferrer, I.; Knecht, E.; et al. Autophagy Induction Halts Axonal Degeneration in a Mouse Model of X-Adrenoleukodystrophy. *Acta Neuropathol.* **2015**, *129*, 399–415. [[CrossRef](#)]

220. Kruska, N.; Schönfeld, P.; Pujol, A.; Reiser, G. Astrocytes and Mitochondria from Adrenoleukodystrophy Protein (ABCD1)-Deficient Mice Reveal That the Adrenoleukodystrophy-Associated Very Long-Chain Fatty Acids Target Several Cellular Energy-Dependent Functions. *Biochim. Biophys. Acta (BBA)-Mol. Basis Dis.* **2015**, *1852*, 925–936. [[CrossRef](#)]
221. López-Erauskin, J.; Fourcade, S.; Galino, J.; Ruiz, M.; Schlüter, A.; Naudi, A.; Jove, M.; Portero-Otin, M.; Pamplona, R.; Ferrer, I.; et al. Antioxidants Halt Axonal Degeneration in a Mouse Model of X-adrenoleukodystrophy. *Ann. Neurol.* **2011**, *70*, 84–92. [[CrossRef](#)] [[PubMed](#)]
222. Court, F.A.; Coleman, M.P. Mitochondria as a Central Sensor for Axonal Degenerative Stimuli. *Trends Neurosci.* **2012**, *35*, 364–372. [[CrossRef](#)] [[PubMed](#)]
223. Whitcomb, R.W.; Linehan, W.M.; Knazek, R.A. Effects of Long-Chain, Saturated Fatty Acids on Membrane Microviscosity and Adrenocorticotropin Responsiveness of Human Adrenocortical Cells in Vitro. *J. Clin. Investig.* **1988**, *81*, 185–188. [[CrossRef](#)] [[PubMed](#)]
224. Osório, M.J.; Goldman, S.A. Neurogenetics of Pelizaeus-Merzbacher Disease. *Handb. Clin. Neurol.* **2018**, *148*, 701–722. [[CrossRef](#)]
225. Harding, B.N. Pelizaeus-Merzbacher Disease. In *Developmental Neuropathology*; Adle-Biassette, H., Harding, B.N., Golden, J., Eds.; John Wiley & Sons, Ltd.: Oxford, UK, 2018; pp. 417–425, ISBN 978-1-119-01311-2.
226. Duan, R.; Li, L.; Yan, H.; He, M.; Gao, K.; Xing, S.; Ji, H.; Wang, J.; Cao, B.; Li, D.; et al. Novel Insight into the Potential Pathogenicity of Mitochondrial Dysfunction Resulting from PLP1 Duplication Mutations in Patients with Pelizaeus-Merzbacher Disease. *Neuroscience* **2021**, *476*, 60–71. [[CrossRef](#)]
227. Inoue, K. Pelizaeus-Merzbacher Disease: Molecular and Cellular Pathologies and Associated Phenotypes. *Adv. Exp. Med. Biol.* **2019**, *1190*, 201–216. [[CrossRef](#)]
228. Edgar, J.M.; McCulloch, M.C.; Montague, P.; Brown, A.M.; Thilemann, S.; Pratola, L.; Gruenenfelder, F.I.; Griffiths, I.R.; Nave, K.-A. Demyelination and Axonal Preservation in a Transgenic Mouse Model of Pelizaeus-Merzbacher Disease. *EMBO Mol. Med.* **2010**, *2*, 42–50. [[CrossRef](#)]
229. Woodward, K.J.; Cundall, M.; Sperle, K.; Sistermans, E.A.; Ross, M.; Howell, G.; Gribble, S.M.; Burford, D.C.; Carter, N.P.; Hobson, D.L.; et al. Heterogeneous Duplications in Patients with Pelizaeus-Merzbacher Disease Suggest a Mechanism of Coupled Homologous and Nonhomologous Recombination. *Am. J. Hum. Genet.* **2005**, *77*, 966–987. [[CrossRef](#)]
230. Ruiz, M.; Bégou, M.; Launay, N.; Ranea-Robles, P.; Bianchi, P.; López-Erauskin, J.; Morató, L.; Guilera, C.; Petit, B.; Vaur-Barriere, C.; et al. Oxidative Stress and Mitochondrial Dynamics Malfunction Are Linked in Pelizaeus-Merzbacher Disease. *Brain Pathol.* **2018**, *28*, 611–630. [[CrossRef](#)]
231. Hüttemann, M.; Zhang, Z.; Mullins, C.; Bessert, D.; Lee, I.; Nave, K.-A.; Appikatla, S.; Skoff, R.P. Different Proteolipid Protein Mutants Exhibit Unique Metabolic Defects. *ASN Neuro* **2009**, *1*, AN20090028. [[CrossRef](#)]
232. Appikatla, S.; Bessert, D.; Lee, I.; Hüttemann, M.; Mullins, C.; Somayajulu-Nitu, M.; Yao, F.; Skoff, R.P. Insertion of Proteolipid Protein into Oligodendrocyte Mitochondria Regulates Extracellular PH and Adenosine Triphosphate: Functions of PLPs Insertion into Mitochondria. *Glia* **2014**, *62*, 356–373. [[CrossRef](#)] [[PubMed](#)]
233. Prior, T.W.; Leach, M.E.; Finanger, E. Spinal Muscular Atrophy. In *GeneReviews*<sup>®</sup>; Adam, M.P., Everman, D.B., Mirzaa, G.M., Pagon, R.A., Wallace, S.E., Bean, L.J., Gripp, K.W., Amemiya, A., Eds.; University of Washington, Seattle: Seattle, WA, USA, 1993.
234. Lefebvre, S.; Bürglen, L.; Reboullet, S.; Clermont, O.; Burlet, P.; Viollet, L.; Benichou, B.; Cruaud, C.; Millasseau, P.; Zeviani, M. Identification and Characterization of a Spinal Muscular Atrophy-Determining Gene. *Cell* **1995**, *80*, 155–165. [[CrossRef](#)] [[PubMed](#)]
235. Anhof, D.; Eggermann, T.; Rudnik-Schöneborn, S.; Zerres, K. Determination of SMN1 and SMN2 Copy Number Using TaqMan Technology. *Hum. Mutat.* **2003**, *22*, 74–78. [[CrossRef](#)] [[PubMed](#)]
236. Calucho, M.; Bernal, S.; Aliás, L.; March, F.; Venceslá, A.; Rodríguez-Álvarez, F.J.; Aller, E.; Fernández, R.M.; Borrego, S.; Millán, J.M.; et al. Correlation between SMA Type and SMN2 Copy Number Revisited: An Analysis of 625 Unrelated Spanish Patients and a Compilation of 2834 Reported Cases. *Neuromuscul. Disord.* **2018**, *28*, 208–215. [[CrossRef](#)]
237. Gubitz, A.K.; Feng, W.; Dreyfuss, G. The SMN Complex. *Exp. Cell Res.* **2004**, *296*, 51–56. [[CrossRef](#)]
238. Fallini, C.; Zhang, H.; Su, Y.; Silani, V.; Singer, R.H.; Rossoll, W.; Bassell, G.J. The Survival of Motor Neuron (SMN) Protein Interacts with the mRNA-Binding Protein HuD and Regulates Localization of Poly(A) mRNA in Primary Motor Neuron Axons. *J. Neurosci.* **2011**, *31*, 3914–3925. [[CrossRef](#)]
239. Miller, N.; Shi, H.; Zelikovich, A.S.; Ma, Y.-C. Motor Neuron Mitochondrial Dysfunction in Spinal Muscular Atrophy. *Hum. Mol. Genet.* **2016**, *25*, 3395–3406. [[CrossRef](#)]
240. Boyd, P.J.; Tu, W.-Y.; Shorrock, H.K.; Groen, E.J.N.; Carter, R.N.; Powis, R.A.; Thomson, S.R.; Thomson, D.; Graham, L.C.; Motyl, A.A.L.; et al. Bioenergetic Status Modulates Motor Neuron Vulnerability and Pathogenesis in a Zebrafish Model of Spinal Muscular Atrophy. *PLoS Genet.* **2017**, *13*, e1006744. [[CrossRef](#)]
241. Acsadi, G.; Lee, I.; Li, X.; Khaidakov, M.; Pecinova, A.; Parker, G.C.; Hüttemann, M. Mitochondrial Dysfunction in a Neural Cell Model of Spinal Muscular Atrophy. *J. Neurosci. Res.* **2009**, *87*, 2748–2756. [[CrossRef](#)]
242. Hellbach, N.; Peterson, S.; Haehnke, D.; Shankar, A.; LaBarge, S.; Pivaroff, C.; Saenger, S.; Thomas, C.; McCarthy, K.; Ebeling, M.; et al. Impaired Myogenic Development, Differentiation and Function in HESC-Derived SMA Myoblasts and Myotubes. *PLoS ONE* **2018**, *13*, e0205589. [[CrossRef](#)]
243. Thelen, M.P.; Wirth, B.; Kye, M.J. Mitochondrial Defects in the Respiratory Complex I Contribute to Impaired Translational Initiation via ROS and Energy Homeostasis in SMA Motor Neurons. *Acta Neuropathol. Commun.* **2020**, *8*, 223. [[CrossRef](#)] [[PubMed](#)]

244. Wang, Y.; Xu, C.; Ma, L.; Mou, Y.; Zhang, B.; Zhou, S.; Tian, Y.; Trinh, J.; Zhang, X.; Li, X.-J. Drug Screening with Human SMN2 Reporter Identifies SMN Protein Stabilizers to Correct SMA Pathology. *Life Sci. Alliance* **2019**, *2*, e201800268. [[CrossRef](#)] [[PubMed](#)]
245. Xu, C.-C.; Denton, K.R.; Wang, Z.-B.; Zhang, X.; Li, X.-J. Abnormal Mitochondrial Transport and Morphology as Early Pathological Changes in Human Models of Spinal Muscular Atrophy. *Dis. Model. Mech.* **2016**, *9*, 39–49. [[CrossRef](#)] [[PubMed](#)]
246. Kariya, S.; Park, G.-H.; Maeno-Hikichi, Y.; Leykekhman, O.; Lutz, C.; Arkovitz, M.S.; Landmesser, L.T.; Monani, U.R. Reduced SMN Protein Impairs Maturation of the Neuromuscular Junctions in Mouse Models of Spinal Muscular Atrophy. *Hum. Mol. Genet.* **2008**, *17*, 2552–2569. [[CrossRef](#)]
247. Torres-Benito, L.; Neher, M.F.; Cano, R.; Ruiz, R.; Tabares, L. SMN Requirement for Synaptic Vesicle, Active Zone and Microtubule Postnatal Organization in Motor Nerve Terminals. *PLoS ONE* **2011**, *6*, e26164. [[CrossRef](#)]
248. Ripolone, M.; Ronchi, D.; Violano, R.; Vallejo, D.; Fagiolari, G.; Barca, E.; Lucchini, V.; Colombo, I.; Villa, L.; Berardinelli, A.; et al. Impaired Muscle Mitochondrial Biogenesis and Myogenesis in Spinal Muscular Atrophy. *JAMA Neurol.* **2015**, *72*, 666–675. [[CrossRef](#)]
249. Ando, S.; Funato, M.; Ohuchi, K.; Inagaki, S.; Sato, A.; Seki, J.; Kawase, C.; Saito, T.; Nishio, H.; Nakamura, S.; et al. The Protective Effects of Levetiracetam on a Human iPSCs-Derived Spinal Muscular Atrophy Model. *Neurochem. Res.* **2019**, *44*, 1773–1779. [[CrossRef](#)]
250. Chacinska, A.; Koehler, C.M.; Milenkovic, D.; Lithgow, T.; Pfanner, N. Importing Mitochondrial Proteins: Machineries and Mechanisms. *Cell* **2009**, *138*, 628–644. [[CrossRef](#)]
251. Van Alstyne, M.; Lotti, F.; Dal Mas, A.; Area-Gomez, E.; Pellizzoni, L. Stasimon/Tmem41b Localizes to Mitochondria-Associated ER Membranes and Is Essential for Mouse Embryonic Development. *Biochem. Biophys. Res. Commun.* **2018**, *506*, 463–470. [[CrossRef](#)]
252. Anderton, R.S.; Meloni, B.P.; Mastaglia, F.L.; Boulos, S. Spinal Muscular Atrophy and the Antiapoptotic Role of Survival of Motor Neuron (SMN) Protein. *Mol. Neurobiol.* **2013**, *47*, 821–832. [[CrossRef](#)]
253. Sato, K.; Eguchi, Y.; Kodama, T.S.; Tsujimoto, Y. Regions Essential for the Interaction between Bcl-2 and SMN, the Spinal Muscular Atrophy Disease Gene Product. *Cell Death Differ.* **2000**, *7*, 374–383. [[CrossRef](#)] [[PubMed](#)]
254. Garcera, A.; Mincheva, S.; Gou-Fabregas, M.; Caraballo-Miralles, V.; Lladó, J.; Comella, J.X.; Soler, R.M. A New Model to Study Spinal Muscular Atrophy: Neurite Degeneration and Cell Death Is Counteracted by BCL-X(L) Overexpression in Motoneurons. *Neurobiol. Dis.* **2011**, *42*, 415–426. [[CrossRef](#)] [[PubMed](#)]
255. Piras, A.; Schiaffino, L.; Boido, M.; Valsecchi, V.; Guglielmotto, M.; De Amicis, E.; Puyal, J.; Garcera, A.; Tamagno, E.; Soler, R.M.; et al. Inhibition of Autophagy Delays Motoneuron Degeneration and Extends Lifespan in a Mouse Model of Spinal Muscular Atrophy. *Cell Death Dis.* **2017**, *8*, 3223. [[CrossRef](#)]
256. Simic, G.; Seso-Simic, D.; Lucassen, P.J.; Islam, A.; Krsnik, Z.; Cviko, A.; Jelasic, D.; Barisic, N.; Winblad, B.; Kostovic, I.; et al. Ultrastructural Analysis and TUNEL Demonstrate Motor Neuron Apoptosis in Werdnig-Hoffmann Disease. *J. Neuropathol. Exp. Neurol.* **2000**, *59*, 398–407. [[CrossRef](#)]
257. Simon, C.M.; Dai, Y.; Van Alstyne, M.; Koutsoumpa, C.; Pagiazitis, J.G.; Chalif, J.I.; Wang, X.; Rabinowitz, J.E.; Henderson, C.E.; Pellizzoni, L.; et al. Converging Mechanisms of P53 Activation Drive Motor Neuron Degeneration in Spinal Muscular Atrophy. *Cell Rep.* **2017**, *21*, 3767–3780. [[CrossRef](#)] [[PubMed](#)]
258. Young, P.J.; Day, P.M.; Zhou, J.; Androphy, E.J.; Morris, G.E.; Lorson, C.L. A Direct Interaction between the Survival Motor Neuron Protein and P53 and Its Relationship to Spinal Muscular Atrophy. *J. Biol. Chem.* **2002**, *277*, 2852–2859. [[CrossRef](#)]
259. Helmken, C.; Hofmann, Y.; Schoenen, F.; Oprea, G.; Raschke, H.; Rudnik-Schöneborn, S.; Zerres, K.; Wirth, B. Evidence for a Modifying Pathway in SMA Discordant Families: Reduced SMN Level Decreases the Amount of Its Interacting Partners and Htra2-Beta1. *Hum. Genet.* **2003**, *114*, 11–21. [[CrossRef](#)]
260. Rindt, H.; Feng, Z.; Mazzasette, C.; Glascock, J.J.; Valdivia, D.; Pyles, N.; Crawford, T.O.; Swoboda, K.J.; Patitucci, T.N.; Ebert, A.D.; et al. Astrocytes Influence the Severity of Spinal Muscular Atrophy. *Hum. Mol. Genet.* **2015**, *24*, 4094–4102. [[CrossRef](#)]
261. Kuru, S.; Sakai, M.; Konagaya, M.; Yoshida, M.; Hashizume, Y.; Saito, K. An Autopsy Case of Spinal Muscular Atrophy Type III (Kugelberg-Welander Disease). *Neuropathology* **2009**, *29*, 63–67. [[CrossRef](#)]
262. McGivern, J.V.; Patitucci, T.N.; Nord, J.A.; Barabas, M.-E.A.; Stucky, C.L.; Ebert, A.D. Spinal Muscular Atrophy Astrocytes Exhibit Abnormal Calcium Regulation and Reduced Growth Factor Production. *Glia* **2013**, *61*, 1418–1428. [[CrossRef](#)]
263. Deguise, M.-O.; Kothary, R. New Insights into SMA Pathogenesis: Immune Dysfunction and Neuroinflammation. *Ann. Clin. Transl. Neurol.* **2017**, *4*, 522–530. [[CrossRef](#)] [[PubMed](#)]
264. Pierzynowska, K.; Gaffke, L.; Cyske, Z.; Węgrzyn, G.; Buttari, B.; Profumo, E.; Saso, L. Oxidative Stress in Mucopolysaccharidoses: Pharmacological Implications. *Molecules* **2021**, *26*, 5616. [[CrossRef](#)] [[PubMed](#)]
265. Afratis, N.; Gialeli, C.; Nikitovic, D.; Tseggenidis, T.; Karousou, E.; Theocharis, A.D.; Pavão, M.S.; Tzanakakis, G.N.; Karamanos, N.K. Glycosaminoglycans: Key Players in Cancer Cell Biology and Treatment: GAG Targeting in Cancer Cell Biology. *FEBS J.* **2012**, *279*, 1177–1197. [[CrossRef](#)] [[PubMed](#)]
266. Simonaro, C.M.; Ge, Y.; Eliyahu, E.; He, X.; Jepsen, K.J.; Schuchman, E.H. Involvement of the Toll-like Receptor 4 Pathway and Use of TNF- $\alpha$  Antagonists for Treatment of the Mucopolysaccharidoses. *Proc. Natl. Acad. Sci. USA.* **2010**, *107*, 222–227. [[CrossRef](#)] [[PubMed](#)]
267. Mandolfo, O.; Parker, H.; Bigger, B. Innate Immunity in Mucopolysaccharide Diseases. *Int. J. Mol. Sci.* **2022**, *23*, 1999. [[CrossRef](#)]

268. Martins, C.; Hůlková, H.; Dridi, L.; Dormoy-Raclet, V.; Grigoryeva, L.; Choi, Y.; Langford-Smith, A.; Wilkinson, F.L.; Ohmi, K.; DiCristo, G.; et al. Neuroinflammation, Mitochondrial Defects and Neurodegeneration in Mucopolysaccharidosis III Type C Mouse Model. *Brain* **2015**, *138*, 336–355. [[CrossRef](#)]
269. Villani, G.R.D.; Gargiulo, N.; Faraonio, R.; Castaldo, S.; Gonzalez y Reyero, E.; Di Natale, P. Cytokines, Neurotrophins, and Oxidative Stress in Brain Disease from Mucopolysaccharidosis IIIB. *J. Neurosci. Res.* **2007**, *85*, 612–622. [[CrossRef](#)]
270. Polgreen, L.E.; Vehe, R.K.; Rudser, K.; Kunin-Batson, A.; Utz, J.J.; Dickson, P.; Shapiro, E.; Whitley, C.B. Elevated TNF- $\alpha$  Is Associated with Pain and Physical Disability in Mucopolysaccharidosis Types I, II, and VI. *Mol. Genet. Metab.* **2016**, *117*, 427–430. [[CrossRef](#)]
271. Pereira, V.G.; Martins, A.M.; Micheletti, C.; D’Almeida, V. Mutational and Oxidative Stress Analysis in Patients with Mucopolysaccharidosis Type I Undergoing Enzyme Replacement Therapy. *Clin. Chim. Acta* **2008**, *387*, 75–79. [[CrossRef](#)]
272. Filippin, L.; Vanzin, C.S.; Biancini, G.B.; Pereira, I.N.; Manfredini, V.; Sitta, A.; Peralba, M.d.C.R.; Schwartz, I.V.D.; Giugliani, R.; Vargas, C.R. Oxidative Stress in Patients with Mucopolysaccharidosis Type II before and during Enzyme Replacement Therapy. *Mol. Genet. Metab.* **2011**, *103*, 121–127. [[CrossRef](#)]
273. McGlynn, R.; Dobrenis, K.; Walkley, S.U. Differential Subcellular Localization of Cholesterol, Gangliosides, and Glycosaminoglycans in Murine Models of Mucopolysaccharide Storage Disorders. *J. Comp. Neurol.* **2004**, *480*, 415–426. [[CrossRef](#)]
274. Jacques, C.E.D.; Donida, B.; Mescka, C.P.; Rodrigues, D.G.B.; Marchetti, D.P.; Bitencourt, F.H.; Burin, M.G.; de Souza, C.F.M.; Giugliani, R.; Vargas, C.R. Oxidative and Nitrate Stress and Pro-Inflammatory Cytokines in Mucopolysaccharidosis Type II Patients: Effect of Long-Term Enzyme Replacement Therapy and Relation with Glycosaminoglycan Accumulation. *Biochim. Biophys. Acta (BBA)-Mol. Basis Dis.* **2016**, *1862*, 1608–1616. [[CrossRef](#)] [[PubMed](#)]
275. Breiden, B.; Sandhoff, K. Mechanism of Secondary Ganglioside and Lipid Accumulation in Lysosomal Disease. *Int. J. Mol. Sci.* **2020**, *21*, 2566. [[CrossRef](#)] [[PubMed](#)]
276. Constantopoulos, G.; Dekaban, A.S. Neurochemistry of The Mucopolysaccharidoses: Brain Lipids and Lysosomal Enzymes in Patients with Four Types of Mucopolysaccharidosis and in Normal Controls. *J. Neurochem.* **1978**, *30*, 965–973. [[CrossRef](#)] [[PubMed](#)]
277. Huang, W.; Cheng, Y.-S.; Yang, S.; Swaroop, M.; Xu, M.; Huang, W.; Zheng, W. Disease Modeling for Mucopolysaccharidosis Type IIIB Using Patient Derived Induced Pluripotent Stem Cells. *Exp. Cell Res.* **2021**, *407*, 112785. [[CrossRef](#)] [[PubMed](#)]
278. Ginzburg, L.; Futerman, A.H. Defective Calcium Homeostasis in the Cerebellum in a Mouse Model of Niemann-Pick A Disease. *J. Neurochem.* **2005**, *95*, 1619–1628. [[CrossRef](#)]
279. Fraldi, A.; Annunziata, F.; Lombardi, A.; Kaiser, H.-J.; Medina, D.L.; Spanpanato, C.; Fedele, A.O.; Polishchuk, R.; Sorrentino, N.C.; Simons, K.; et al. Lysosomal Fusion and SNARE Function Are Impaired by Cholesterol Accumulation in Lysosomal Storage Disorders. *EMBO J.* **2010**, *29*, 3607–3620. [[CrossRef](#)]
280. Finkbeiner, S. The Autophagy Lysosomal Pathway and Neurodegeneration. *Cold Spring Harb. Perspect. Biol.* **2020**, *12*, a033993. [[CrossRef](#)]
281. Chai, P.; Ni, H.; Zhang, H.; Fan, X. The Evolving Functions of Autophagy in Ocular Health: A Double-Edged Sword. *Int. J. Biol. Sci.* **2016**, *12*, 1332–1340. [[CrossRef](#)]
282. Sambri, I.; D’Alessio, R.; Ezhova, Y.; Giuliano, T.; Sorrentino, N.C.; Cacace, V.; De Risi, M.; Cataldi, M.; Annunziato, L.; De Leonibus, E.; et al. Lysosomal Dysfunction Disrupts Presynaptic Maintenance and Restoration of Presynaptic Function Prevents Neurodegeneration in Lysosomal Storage Diseases. *EMBO Mol. Med.* **2017**, *9*, 112–132. [[CrossRef](#)]
283. Monaco, A.; Maffia, V.; Sorrentino, N.C.; Sambri, I.; Ezhova, Y.; Giuliano, T.; Cacace, V.; Nusco, E.; De Risi, M.; De Leonibus, E.; et al. The Amyloid Inhibitor CLR01 Relieves Autophagy and Ameliorates Neuropathology in a Severe Lysosomal Storage Disease. *Mol. Ther.* **2020**, *28*, 1167–1176. [[CrossRef](#)] [[PubMed](#)]
284. Hamano, K.; Hayashi, M.; Shioda, K.; Fukatsu, R.; Mizutani, S. Mechanisms of Neurodegeneration in Mucopolysaccharidoses II and IIIB: Analysis of Human Brain Tissue. *Acta Neuropathol.* **2008**, *115*, 547–559. [[CrossRef](#)] [[PubMed](#)]
285. Winder-Rhodes, S.E.; Garcia-Reitböck, P.; Ban, M.; Evans, J.R.; Jacques, T.S.; Kempainen, A.; Foltynie, T.; Williams-Gray, C.H.; Chinnery, P.F.; Hudson, G.; et al. Genetic and Pathological Links between Parkinson’s Disease and the Lysosomal Disorder Sanfilippo Syndrome: Parkinson’s Disease and Sanfilippo Syndrome. *Mov. Disord.* **2012**, *27*, 312–315. [[CrossRef](#)] [[PubMed](#)]
286. Maurya, S.K.; Bhattacharya, N.; Mishra, S.; Bhattacharya, A.; Banerjee, P.; Senapati, S.; Mishra, R. Microglia Specific Drug Targeting Using Natural Products for the Regulation of Redox Imbalance in Neurodegeneration. *Front. Pharmacol.* **2021**, *12*, 654489. [[CrossRef](#)] [[PubMed](#)]
287. Chen, X.; Guo, C.; Kong, J. Oxidative Stress in Neurodegenerative Diseases. *Neural. Regen. Res.* **2012**, *7*, 376–385. [[CrossRef](#)]
288. Rajasekar, N.; Dwivedi, S.; Tota, S.K.; Kamat, P.K.; Hanif, K.; Nath, C.; Shukla, R. Neuroprotective Effect of Curcumin on Okadaic Acid Induced Memory Impairment in Mice. *Eur. J. Pharmacol.* **2013**, *715*, 381–394. [[CrossRef](#)]
289. Sancheti, H.; Kanamori, K.; Patil, I.; Díaz Brinton, R.; Ross, B.D.; Cadenas, E. Reversal of Metabolic Deficits by Lipoic Acid in a Triple Transgenic Mouse Model of Alzheimer’s Disease: A  $^{13}\text{C}$  NMR Study. *J. Cereb Blood Flow Metab.* **2014**, *34*, 288–296. [[CrossRef](#)]
290. Sano, M.; Ernesto, C.; Thomas, R.G.; Klauber, M.R.; Schafer, K.; Grundman, M.; Woodbury, P.; Growdon, J.; Cotman, C.W.; Pfeiffer, E.; et al. A Controlled Trial of Selegiline, Alpha-Tocopherol, or Both as Treatment for Alzheimer’s Disease. The Alzheimer’s Disease Cooperative Study. *N. Engl. J. Med.* **1997**, *336*, 1216–1222. [[CrossRef](#)]

291. Dysken, M.W.; Sano, M.; Asthana, S.; Vertrees, J.E.; Pallaki, M.; Llorente, M.; Love, S.; Schellenberg, G.D.; McCarten, J.R.; Malphurs, J.; et al. Effect of Vitamin E and Memantine on Functional Decline in Alzheimer Disease: The TEAM-AD VA Cooperative Randomized Trial. *JAMA* **2014**, *311*, 33–44. [CrossRef]
292. Ringman, J.M.; Frautschy, S.A.; Teng, E.; Begum, A.N.; Bardens, J.; Beigi, M.; Gylys, K.H.; Badmaev, V.; Heath, D.D.; Apostolova, L.G.; et al. Oral Curcumin for Alzheimer's Disease: Tolerability and Efficacy in a 24-Week Randomized, Double Blind, Placebo-Controlled Study. *Alzheimers Res. Ther.* **2012**, *4*, 43. [CrossRef]
293. Kumar, A.; Singh, A. A Review on Mitochondrial Restorative Mechanism of Antioxidants in Alzheimer's Disease and Other Neurological Conditions. *Front. Pharmacol.* **2015**, *6*, 206. [CrossRef] [PubMed]
294. Köstel, A.S.; Bora-Tatar, G.; Erdem-Yurter, H. Spinal Muscular Atrophy: An Oxidative Stress Response Counteracted with curcumin. *Biomed. Aging Pathol.* **2012**, *2*, 61–66. [CrossRef]
295. Grygiel-Górniak, B.; Puszczewicz, M. Oxidative Damage and Antioxidative Therapy in Systemic Sclerosis. *Mediat. Inflamm.* **2014**, *2014*, 1–11. [CrossRef] [PubMed]
296. Casanovas, C.; Ruiz, M.; Schlüter, A.; Naudí, A.; Fourcade, S.; Veciana, M.; Castañer, S.; Albertí, A.; Bargalló, N.; Johnson, M.; et al. Biomarker Identification, Safety, and Efficacy of High-Dose Antioxidants for Adrenomyeloneuropathy: A Phase II Pilot Study. *Neurotherapeutics* **2019**, *16*, 1167–1182. [CrossRef] [PubMed]
297. Haddad, M.; Hervé, V.; Ben Khedher, M.R.; Rabanel, J.-M.; Ramassamy, C. Glutathione: An Old and Small Molecule with Great Functions and New Applications in the Brain and in Alzheimer's Disease. *Antioxid. Redox Signal.* **2021**, *35*, 270–292. [CrossRef]
298. Taylor, S.; Wheeler, L.C.; Taylor, J.R.; Griffin, H.C. Nutrition: An Issue of Concern for Children with Disabilities. *Nurse Pract.* **1996**, *21*, 17–18, 20.
299. Pocernich, C.B.; Butterfield, D.A. Elevation of Glutathione as a Therapeutic Strategy in Alzheimer Disease. *Biochim. Biophys. Acta (BBA)-Mol. Basis Dis.* **2012**, *1822*, 625–630. [CrossRef]
300. Zarka, M.H.; Bridge, W.J. Oral Administration of  $\gamma$ -Glutamylcysteine Increases Intracellular Glutathione Levels above Homeostasis in a Randomised Human Trial Pilot Study. *Redox Biol.* **2017**, *11*, 631–636. [CrossRef]
301. Schmitt, B.; Vicenzi, M.; Garrel, C.; Denis, F.M. Effects of N-Acetylcysteine, Oral Glutathione (GSH) and a Novel Sublingual Form of GSH on Oxidative Stress Markers: A Comparative Crossover Study. *Redox Biol.* **2015**, *6*, 198–205. [CrossRef]
302. Liu, Y.; Chen, Z.; Li, B.; Yao, H.; Zarka, M.; Welch, J.; Sachdev, P.; Bridge, W.; Braidy, N. Supplementation with  $\gamma$ -Glutamylcysteine ( $\gamma$ -GC) Lessens Oxidative Stress, Brain Inflammation and Amyloid Pathology and Improves Spatial Memory in a Murine Model of AD. *Neurochem. Int.* **2021**, *144*, 104931. [CrossRef]
303. U.S. National Library of Medicine. ClinicalTrials.Gov. Available online: <https://clinicaltrials.gov/ct2/home> (accessed on 1 January 2021).
304. Brieger, K.; Schiavone, S.; Miller, F.J.; Krause, K.H. Reactive Oxygen Species: From Health to Disease. *Swiss Med. Wkly.* **2012**, *142*, w13659. [CrossRef] [PubMed]
305. Pedroso, I.; Bringas, M.L.; Aguiar, A.; Morales, L.; Alvarez, M.; Valdés, P.A.; Alvarez, L. Use of Cuban Recombinant Human Erythropoietin in Parkinson's Disease Treatment. *MEDICC Rev.* **2012**, *14*, 11–17. [PubMed]
306. Langan, A.R.; Khan, M.A.; Yeung, I.W.T.; Van Dyk, J.; Hill, R.P. Partial Volume Rat Lung Irradiation: The Protective/Mitigating Effects of Eukarion-189, a Superoxide Dismutase-Catalase Mimetic. *Radiother. Oncol.* **2006**, *79*, 231–238. [CrossRef] [PubMed]
307. Peng, J.; Stevenson, F.F.; Doctrow, S.R.; Andersen, J.K. Superoxide Dismutase/Catalase Mimetics Are Neuroprotective against Selective Paraquat-Mediated Dopaminergic Neuron Death in the Substantial Nigra: Implications for Parkinson Disease. *J. Biol. Chem.* **2005**, *280*, 29194–29198. [CrossRef] [PubMed]
308. Jung, C.; Rong, Y.; Doctrow, S.; Baudry, M.; Malfroy, B.; Xu, Z. Synthetic Superoxide Dismutase/Catalase Mimetics Reduce Oxidative Stress and Prolong Survival in a Mouse Amyotrophic Lateral Sclerosis Model. *Neurosci. Lett.* **2001**, *304*, 157–160. [CrossRef]
309. Melov, S.; Wolf, N.; Strozyk, D.; Doctrow, S.R.; Bush, A.I. Mice Transgenic for Alzheimer Disease Beta-Amyloid Develop Lens Cataracts That Are Rescued by Antioxidant Treatment. *Free Radic. Biol. Med.* **2005**, *38*, 258–261. [CrossRef] [PubMed]
310. Baker, K.; Marcus, C.B.; Huffman, K.; Kruk, H.; Malfroy, B.; Doctrow, S.R. Synthetic Combined Superoxide Dismutase/Catalase Mimetics Are Protective as a Delayed Treatment in a Rat Stroke Model: A Key Role for Reactive Oxygen Species in Ischemic Brain Injury. *J. Pharmacol. Exp. Ther.* **1998**, *284*, 215–221.
311. Hallows, W.C.; Albaugh, B.N.; Denu, J.M. Where in the Cell Is SIRT3?—Functional Localization of an NAD<sup>+</sup>-Dependent Protein Deacetylase. *Biochem. J.* **2008**, *411*, e11–e13. [CrossRef]
312. Shen, Y.; Wu, Q.; Shi, J.; Zhou, S. Regulation of SIRT3 on Mitochondrial Functions and Oxidative Stress in Parkinson's Disease. *Biomed. Pharm.* **2020**, *132*, 110928. [CrossRef]
313. Anamika; Khanna, A.; Acharjee, P.; Acharjee, A.; Trigun, S.K. Mitochondrial SIRT3 and Neurodegenerative Brain Disorders. *J. Chem. Neuroanat.* **2019**, *95*, 43–53. [CrossRef]
314. Zeng, R.; Wang, X.; Zhou, Q.; Fu, X.; Wu, Q.; Lu, Y.; Shi, J.; Klaunig, J.E.; Zhou, S. Icarin Protects Rotenone-Induced Neurotoxicity through Induction of SIRT3. *Toxicol. Appl. Pharmacol.* **2019**, *379*, 114639. [CrossRef] [PubMed]
315. Jin, J.; Wang, H.; Hua, X.; Chen, D.; Huang, C.; Chen, Z. An Outline for the Pharmacological Effect of Icarin in the Nervous System. *Eur. J. Pharmacol.* **2019**, *842*, 20–32. [CrossRef] [PubMed]
316. Metsämuuronen, S.; Sirén, H. Bioactive Phenolic Compounds, Metabolism and Properties: A Review on Valuable Chemical Compounds in Scots Pine and Norway Spruce. *Phytochem. Rev.* **2019**, *18*, 623–664. [CrossRef]

317. Mathieu, L.; Lopes Costa, A.; Le Bachelier, C.; Slama, A.; Lebre, A.S.; Taylor, R.W.; Bastin, J.; Djouadi, F. Resveratrol Attenuates Oxidative Stress in Mitochondrial Complex I Deficiency: Involvement of SIRT3. *Free Radic. Biol. Med.* **2016**, *96*, 190–198. [[CrossRef](#)]
318. Chen, H.H.; Chang, P.C.; Wey, S.P.; Chen, P.M.; Chen, C.; Chan, M.H. Therapeutic Effects of Honokiol on Motor Impairment in Hemiparkinsonian Mice Are Associated with Reversing Neurodegeneration and Targeting PPAR $\gamma$  Regulation. *Biomed. Pharm.* **2018**, *108*, 254–262. [[CrossRef](#)]
319. Li, S.; Pu, X.P. Neuroprotective Effect of Kaempferol against a 1-Methyl-4-Phenyl-1,2,3,6-Tetrahydropyridine-Induced Mouse Model of Parkinson's Disease. *Biol. Pharm. Bull.* **2011**, *34*, 1291–1296. [[CrossRef](#)]
320. Serviddio, G.; Bellanti, F.; Sastre, J.; Vendemiale, G.; Altomare, E. Targeting Mitochondria: A New Promising Approach for the Treatment of Liver Diseases. *Curr. Med. Chem.* **2010**, *17*, 2325–2337. [[CrossRef](#)]
321. Murphy, M.P.; Smith, R.A. Drug Delivery to Mitochondria: The Key to Mitochondrial Medicine. *Adv. Drug Deliv. Rev.* **2000**, *41*, 235–250. [[CrossRef](#)]
322. Szeto, H.H. Cell-Permeable, Mitochondrial-Targeted, Peptide Antioxidants. *AAPS J.* **2006**, *8*, E277–E283. [[CrossRef](#)]
323. Bordet, T.; Berna, P.; Abitbol, J.-L.; Pruss, R.M. Olesoxime (TRO19622): A Novel Mitochondrial-Targeted Neuroprotective Compound. *Pharmaceuticals* **2010**, *3*, 345–368. [[CrossRef](#)]
324. Singh, J.; Olle, B.; Suhail, H.; Felicella, M.M.; Giri, S. Metformin-Induced Mitochondrial Function and ABCD2 up-Regulation in X-Linked Adrenoleukodystrophy Involves AMP-Activated Protein Kinase. *J. Neurochem.* **2016**, *138*, 86–100. [[CrossRef](#)]
325. Kenche, V.B.; Barnham, K.J. Alzheimer's Disease & Metals: Therapeutic Opportunities. *Br. J. Pharmacol.* **2011**, *163*, 211–219. [[CrossRef](#)] [[PubMed](#)]
326. Cherny, R.A.; Atwood, C.S.; Xilinas, M.E.; Gray, D.N.; Jones, W.D.; McLean, C.A.; Barnham, K.J.; Volitakis, I.; Fraser, F.W.; Kim, Y.; et al. Treatment with a Copper-Zinc Chelator Markedly and Rapidly Inhibits Beta-Amyloid Accumulation in Alzheimer's Disease Transgenic Mice. *Neuron* **2001**, *30*, 665–676. [[CrossRef](#)] [[PubMed](#)]
327. Ritchie, C.W.; Bush, A.I.; Mackinnon, A.; Macfarlane, S.; Mastwyk, M.; MacGregor, L.; Kiers, L.; Cherny, R.; Li, Q.-X.; Tammer, A.; et al. Metal-Protein Attenuation with Idochlorhydroxyquin (Clioquinol) Targeting Abeta Amyloid Deposition and Toxicity in Alzheimer Disease: A Pilot Phase 2 Clinical Trial. *Arch. Neurol.* **2003**, *60*, 1685–1691. [[CrossRef](#)] [[PubMed](#)]
328. Faux, N.G.; Ritchie, C.W.; Gunn, A.; Rembach, A.; Tsatsanis, A.; Bedo, J.; Harrison, J.; Lannfelt, L.; Blennow, K.; Zetterberg, H.; et al. PBT2 Rapidly Improves Cognition in Alzheimer's Disease: Additional Phase II Analyses. *J. Alzheimers Dis.* **2010**, *20*, 509–516. [[CrossRef](#)]
329. Petri, S.; Calingasan, N.Y.; Alsaied, O.A.; Wille, E.; Kiaei, M.; Friedman, J.E.; Baranova, O.; Chavez, J.C.; Beal, M.F. The Lipophilic Metal Chelators DP-109 and DP-460 Are Neuroprotective in a Transgenic Mouse Model of Amyotrophic Lateral Sclerosis. *J. Neurochem.* **2007**, *102*, 991–1000. [[CrossRef](#)]
330. Watanabe, S.; Nagano, S.; Duce, J.; Kiaei, M.; Li, Q.-X.; Tucker, S.M.; Tiwari, A.; Brown, R.H.; Beal, M.F.; Hayward, L.J.; et al. Increased Affinity for Copper Mediated by Cysteine 111 in Forms of Mutant Superoxide Dismutase 1 Linked to Amyotrophic Lateral Sclerosis. *Free Radic. Biol. Med.* **2007**, *42*, 1534–1542. [[CrossRef](#)]
331. Cristino, L.; Bisogno, T.; Di Marzo, V. Cannabinoids and the Expanded Endocannabinoid System in Neurological Disorders. *Nat. Rev. Neurol.* **2019**, *16*, 9–29. [[CrossRef](#)]
332. Esposito, G.; De Filippis, D.; Carnuccio, R.; Izzo, A.A.; Iuvone, T. The Marijuana Component Cannabidiol Inhibits Beta-Amyloid-Induced Tau Protein Hyperphosphorylation through Wnt/Beta-Catenin Pathway Rescue in PC12 Cells. *J. Mol. Med.* **2006**, *84*, 253–258. [[CrossRef](#)]
333. García, C.; Palomo-Garo, C.; García-Arencibia, M.; Ramos, J.; Pertwee, R.; Fernández-Ruiz, J. Symptom-Relieving and Neuroprotective Effects of the Phytocannabinoid  $\Delta^9$ -THCV in Animal Models of Parkinson's Disease. *Br. J. Pharm.* **2011**, *163*, 1495–1506. [[CrossRef](#)]
334. Sieradzan, K.A.; Fox, S.H.; Hill, M.; Dick, J.P.; Crossman, A.R.; Brotchie, J.M. Cannabinoids Reduce Levodopa-Induced Dyskinesia in Parkinson's Disease: A Pilot Study. *Neurology* **2001**, *57*, 2108–2111. [[CrossRef](#)] [[PubMed](#)]
335. Shoemaker, J.L.; Seely, K.A.; Reed, R.L.; Crow, J.P.; Prather, P.L. The CB2 Cannabinoid Agonist AM-1241 Prolongs Survival in a Transgenic Mouse Model of Amyotrophic Lateral Sclerosis When Initiated at Symptom Onset. *J. Neurochem.* **2007**, *101*, 87–98. [[CrossRef](#)] [[PubMed](#)]
336. Mendiola-Precoma, J.; Berumen, L.C.; Padilla, K.; Garcia-Alcocer, G. Therapies for Prevention and Treatment of Alzheimer's Disease. *BioMed Res. Int.* **2016**, *2016*, 2589276. [[CrossRef](#)] [[PubMed](#)]
337. Baumgart, M.; Snyder, H.M.; Carrillo, M.C.; Fazio, S.; Kim, H.; Johns, H. Summary of the Evidence on Modifiable Risk Factors for Cognitive Decline and Dementia: A Population-Based Perspective. *Alzheimers Dement.* **2015**, *11*, 718–726. [[CrossRef](#)]
338. Navarro-Yepes, J.; Zavala-Flores, L.; Anandhan, A.; Wang, F.; Skotak, M.; Chandra, N.; Li, M.; Pappa, A.; Martinez-Fong, D.; Del Razo, L.M.; et al. Antioxidant Gene Therapy against Neuronal Cell Death. *Pharm. Ther.* **2014**, *142*, 206–230. [[CrossRef](#)]
339. Mueller-Steiner, S.; Zhou, Y.; Arai, H.; Roberson, E.D.; Sun, B.; Chen, J.; Wang, X.; Yu, G.; Esposito, L.; Mucke, L.; et al. Anti-amyloidogenic and Neuroprotective Functions of Cathepsin B: Implications for Alzheimer's Disease. *Neuron* **2006**, *51*, 703–714. [[CrossRef](#)]
340. Singer, O.; Marr, R.A.; Rockenstein, E.; Crews, L.; Coufal, N.G.; Gage, F.H.; Verma, I.M.; Masliah, E. Targeting BACE1 with siRNAs Ameliorates Alzheimer Disease Neuropathology in a Transgenic Model. *Nat. Neurosci.* **2005**, *8*, 1343–1349. [[CrossRef](#)]

341. Day, J.W.; Finkel, R.S.; Chiriboga, C.A.; Connolly, A.M.; Crawford, T.O.; Darras, B.T.; Iannaccone, S.T.; Kuntz, N.L.; Peña, L.D.M.; Shieh, P.B.; et al. Onasemnogene Apeparvovec Gene Therapy for Symptomatic Infantile-Onset Spinal Muscular Atrophy in Patients with Two Copies of SMN2 (STR1VE): An Open-Label, Single-Arm, Multicentre, Phase 3 Trial. *Lancet Neurol.* **2021**, *20*, 284–293. [[CrossRef](#)]
342. Mirea, A.; Shelby, E.-S.; Axente, M.; Badina, M.; Padure, L.; Leanca, M.; Dima, V.; Sporea, C. Combination Therapy with Nusinersen and Onasemnogene Apeparvovec-Xioi in Spinal Muscular Atrophy Type I. *J. Clin. Med.* **2021**, *10*, 5540. [[CrossRef](#)]
343. Gidaro, T.; Servais, L. Nusinersen Treatment of Spinal Muscular Atrophy: Current Knowledge and Existing Gaps. *Dev. Med. Child. Neurol.* **2019**, *61*, 19–24. [[CrossRef](#)]
344. Cartier, N.; Hacein-Bey-Abina, S.; Bartholomae, C.C.; Veres, G.; Schmidt, M.; Kutschera, I.; Vidaud, M.; Abel, U.; Dal-Cortivo, L.; Caccavelli, L.; et al. Hematopoietic Stem Cell Gene Therapy with a Lentiviral Vector in X-Linked Adrenoleukodystrophy. *Science* **2009**, *326*, 818–823. [[CrossRef](#)] [[PubMed](#)]
345. Larochelle, A.; Dunbar, C.E. Hematopoietic Stem Cell Gene Therapy: Assessing the Relevance of Preclinical Models. *Semin. Hematol.* **2013**, *50*, 101–130. [[CrossRef](#)] [[PubMed](#)]

**Disclaimer/Publisher’s Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.