



Supplementary Materials

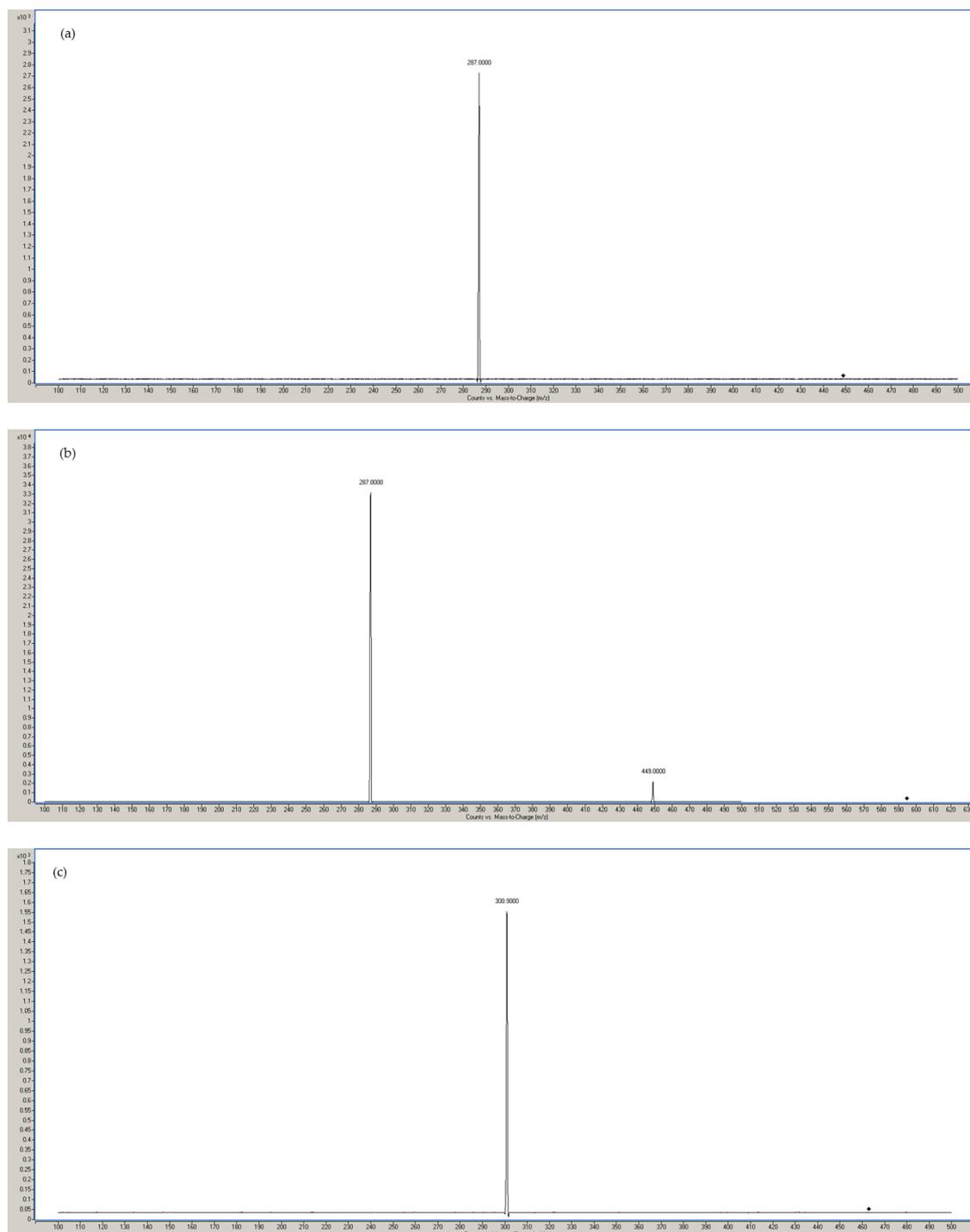


Figure S1. Mass spectra of the four identified peaks in *Cannabis* samples. Second-order mass spectra, obtained with ESI ionization in positive mode, of cyanidin 3-glucoside (peak 1), according to the standard mass spectra as reported by Olivas-Aguirre et al., 2016 (a), second-order mass spectra, obtained with ESI ionization in positive mode, of cyanidin 3-rutinoside (peak 2), according to the standard mass spectra as reported by de Rosso et al., 2008 (b), second-order mass spectra, obtained with ESI ionization in positive mode, of peonidin 3-glucoside (peak 3), according to the standard mass spectra as reported by Castañeda-Ovando et al. (2012) and De Pascual-Teresa et al. (2002)

(c), second-order mass spectra, obtained with ESI ionization in positive mode, of peonidin 3-rutino-side (peak 4) according to the standard mass spectra as reported by de Rosso et al. (2008) and Pra-dhan and Saha (2016).

Table S1. List of primer pairs used for RT-Qpcr. Forward (Fw) and reverse (Rv) primer sequences, the accession number of cs10 transcript used for primer design, amplicon length expressed in base pairs (bp), PCR efficiency (Eff) are given.

Gene	NCBI transcript	Primer sequences	Effi	Amplicon length
<i>CsF3'H</i>	XM_030637390.1	FW:CGACACATCATCAAGCACAGT RV:CAGCATCACTTACAAGCCTATCTC	0.85	125 bp
<i>CsF3'5'H</i>	XM_030644276.1	FW:ATTGCGTGTACATCCACCTG RV:GGATTTTCCCAAAGGCTAGG	1.07	147 bp
<i>CsANS</i>	XM_030645652.1	FW:GCAATGTTTTTCGAGGAGGAG RV:TCAACTCCTCTCGGCATTTTC	1.34	114 bp
<i>CsDFR</i>	XM_030638485.1	Fw:TTCTCCACCTCGGTTACA Rv:TCAGCCTTCCACAGAGTA	1	118 bp
<i>CsOMT</i>	XM_030640905.1	Fw:ACACGAGCAGTTGAAGGAGC Rv:IGTCCTTCATCCACAGGCAC	1.46	87 bp
<i>CsDTX35</i>	XM_030648430.1	Fw:CAATCTTAACGGGTGGGAAG Rv:TTGGACGTTTCCTACCAAGC	1.64	92 bp
<i>Cs3GT</i>	XM_030645093.1	Fw:CCGAGATTGGGAAGGAGGTG Rv:AAGGCCCAGCAGTCCAAAAT	1.54	114 bp
<i>CsTTG1</i>	XM_030645708.1	Fw:ATAGCTTGGGGTGAAGCAAG Rv:AGCAAAGGGGTATCAGTTG	1.18	128 bp
<i>CsMYB82</i>	XM_030636983.1	Fw:ACCAGGAAGGACAGCCAATG Rv:AGGTCCGAGGACGAGGTTTA	1.21	167 bp
<i>CsMYB87</i>	XM_030622841.1	FW:CGTGGAGAATTTGAGGAGGAC RV:TCTACCGCAATCAATGACC	1.45	87 bp
<i>CsbHLH112</i>	XM_030646090.1	FW:GCCACTTCATCAGACAAC RV:CTCCTCATCCTCTTCTTCAT	1.47	165 bp
<i>CsbHLH114</i>	XM_030630819.1	FW:GCCACTTCATCAGACAAC RV:CTCCTCATCCTCTTCTTCAT	1.27	162 bp

Table S2. Amino acid sequences used for Neighbour-Joining phylogenetic analysis reported in Figure 2.

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>XP030499266.1

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>XP030486933.1

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>XP030488163.1

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>XP030488086.1

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>XP030487456.1

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>XP030500350.1

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>XP030487307.1

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>XP030508430.1

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>XP030480622.1

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>XP030495192.1

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>XP030498085.1

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CINVGFSYGVGFPVAMFVGFKMKGKGLIGLWVGLVGAQLVCACVMVVVLVTTDWNVQAERA
KELTRSTSNDVDDGLISLVLEN

Previously described flavonoid transporter belonging to MATEs were included according to [35]: *Arabidopsis thaliana* FFT, (AT4G25640.2) and AtTT12 (AT3G59030.1); *Vitis vinifera* VvanthoMATE1, (NP_001290007.1) and VvanthoMATE3 (NP_001268037.1); *Medicago truncatula* MtMATE1, (XP_003592215.2) and MtMATE2 (; *Solanum lycopersicum* MTP77 (Solyc03g025220.2.1).

>SIMTP77

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 SISICMNINGWESMLFIGINA AISIRVSNELGQGHPRATKYSVYITVFQSLIGILCMVI
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>AtTT12

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 TTVLISSVLCVIVLVFRVGLSKAFTSDAEVIAAVSDLFLLAVSIFLNGIQPILSGVAIG
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>VvanthoMATE1

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 RSWIILLVSCIILLPIYIFATPILKALGQEDEIADLAGQFTLETIPQLFSLAIIPTQKF
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 WCKEGWTGLTWSAFREIWAFAVRLSLASAVMLCLEIWFMIILLTGHLQNAVIAVGSLSI
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 GQRIEADDV

>VvanthoMATE3

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>MtMATE2

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 MGVLVTKDYFAIVFTNSKTLQVAVADLGNLLAVTMVLNSVQPVISGVAVGGGWQALVAY
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>AtFFT

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 AEEI AVPAGQFTLLTIPQLFSLAFNFPTSKFLQAQSKVVAIAWIGFVALSLHVIMLWL-
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VIAVDSLICMNINGLEAMFIGINA AIS-
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WQGLVAYINLGCYYIFGLPFGYLLGYIANFGVMGLWSGMIAGTALQTLILLI-
VLYKTNWNKEVEETMERMKKWGGSETTSKDILASGWPSGLRRQTQVLVVRGRGFKPHF