

Quantitation of MicroRNA-155 in Human Cells by Heterogeneous Enzyme-Linked Oligonucleotide Assay Coupled with Mismatched Catalytic Hairpin Assembly Reaction

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Table S1. The list of oligonucleotides used in this work (5'-3').

Flu-HP1 (for microRNA-155 detection) (6-carboxyfluorescein-labeled hairpin 1)	GTGATAGGGGTGACCGTAGATAATCGT CACCCCTATCACGATTAGCATTA-FAM
B-HP2 (for microRNA-155 detection) (biotinylated hairpin 2)	CGTAGATAATCGTCTTAGGGG TGACGATTATCTACGGTCACCC- biotin
Flu-HP1 (for microRNA-39 detection)	TGTAAATCAGCTTGATGATATTG GGTGTCAAGCTGATTTACACCCGGTGA- FAM
B-HP2 (for microRNA-39 detection)	ATGATATTGGGTGTTTATCAGCTTG ACACCCAATATCATCAAGCTG-biotin
microRNA-141	UAACACUGUCUGGUAAGAUGG
microRNA-319a	UUGGACUGAAGGGUGCUC
microRNA-21	UAGCUUAUCAGACUGAUGUUGA
microRNA-205	UCCUUCAUCCACCGGAGUCUG
microRNA-155	UUAAUGCUAAUCGUGAUGGGGUU
microRNA-39	UCACCGGGUGUAAAUCAGCUUG

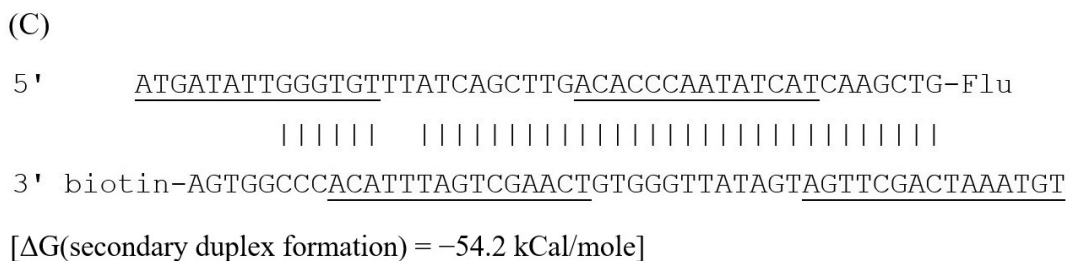
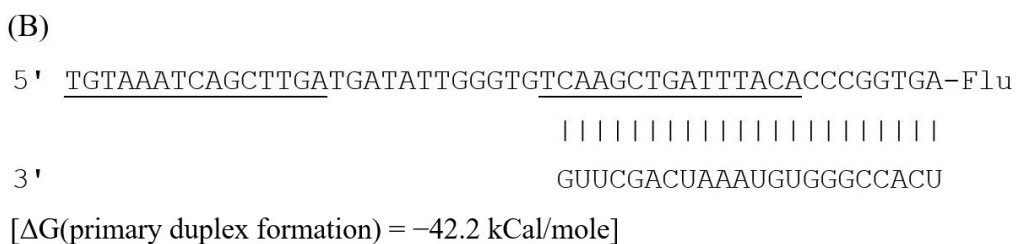
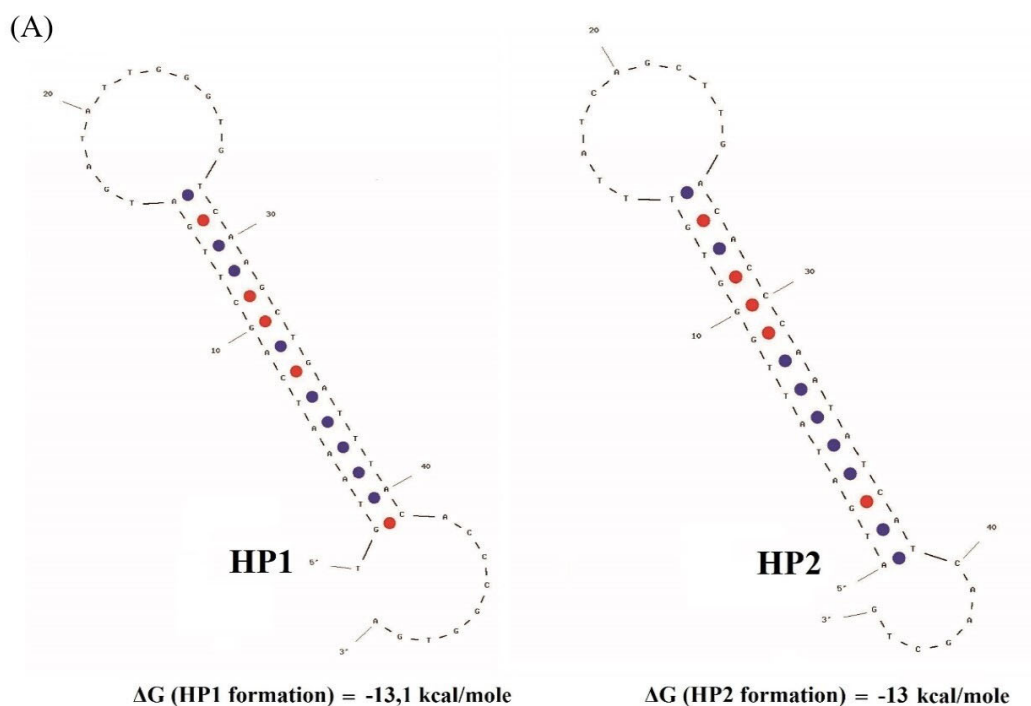


Figure S1. Secondary structures of (A) the hairpins used in chemiluminescent heterogenous mCHA-based microRNA-39 assay, (B) Flu-HP1- microRNA-39, (C) Flu-HP1- B-HP2 duplexes and ΔG values of their formation. Underlined nucleotides are involved in stem of a hairpin structures. Modeling of the hairpin structures and ΔG assessment were performed using OligoAnalyzer 3.1 software.

Optimization of the Assay Conditions for MicroRNA-39 Detection

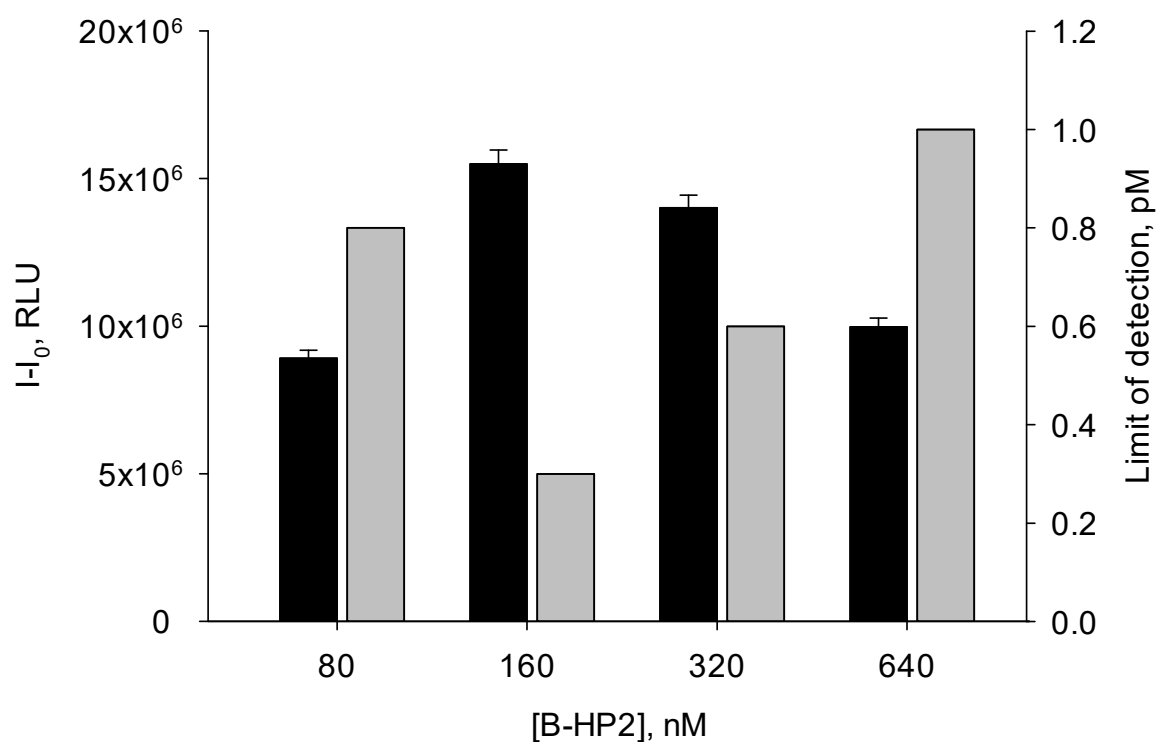


Figure S2. Effect of the B-HP2 probe concentration in the reaction solution on (A) behavior of the calibration curves, (B) chemiluminescence signal ($I - I_0$) (black columns) and limit of detection (gray columns) of the amplified microRNA-39 assay. The mismatched catalytic hairpin assembly reaction was carried out in 10 mM Tris-HCl with pH 7.2 containing 20 mM MgCl₂ at 25°C for 1 h. The concentration of the B-HP2 probe was (a) 80, (b) 160, (c) 320 and (d) 640 mM. The value of chemiluminescence signal ($I - I_0$) was calculated as a difference between the chemiluminescence intensities recorded in the presence (100 pM) and in the absence of microRNA-39. The detection limit was calculated using 3σ rule.

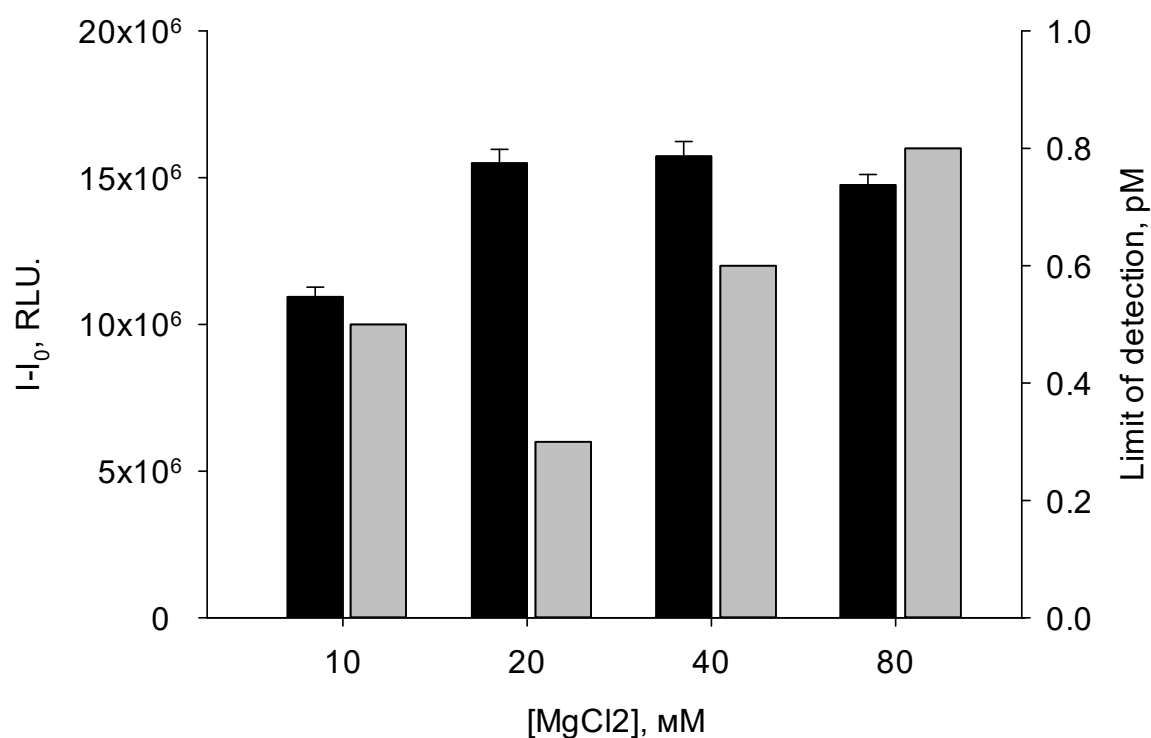


Figure S3. Effect of the MgCl_2 concentration in the reaction solution at the stage of mCHA amplification on (A) behavior of the calibration curves constructed, (B) chemiluminescence signal ($I - I_0$) (black columns) and limit of detection (gray columns) of amplified microRNA-39 assay. The mismatched catalytic hairpin assembly reaction was carried out at 25°C for 1 h using 160 nM B-HP2 probe in 10 mM Tris-HCl with pH 7.2 containing MgCl_2 concentrations of (a) 10, (b) 20, (c) 40 and (d) 80 mM. The value of chemiluminescence signal ($I - I_0$) was calculated as the difference between the chemiluminescence intensities recorded in the presence (100 pM) and in the absence of the microRNA-39. The detection limit was calculated based on 3σ rule.

Analytical Parameters of Heterogenous mCHA-Based MicroRNA-39 Assay

With the use of estimated favorable conditions calibration curve for microRNA-39 detection was obtained (Fig. 4). It is obeyed following equation:

$$Y = \frac{ax}{b+x} + \frac{cx}{d+x} (R^2 0.9976),$$

where a , b , c , d is 5.03×10^6 , 6.5, 6.3×10^{14} , 5.8×10^9 , respectively. The limit of microRNA-39 detection calculated by 3σ rule was 300 fM. The coefficient of variation of chemiluminescent signal (CV) formed upon the heterogeneous mCHA-based miRNA-39 assay within the working range was less than 11%.

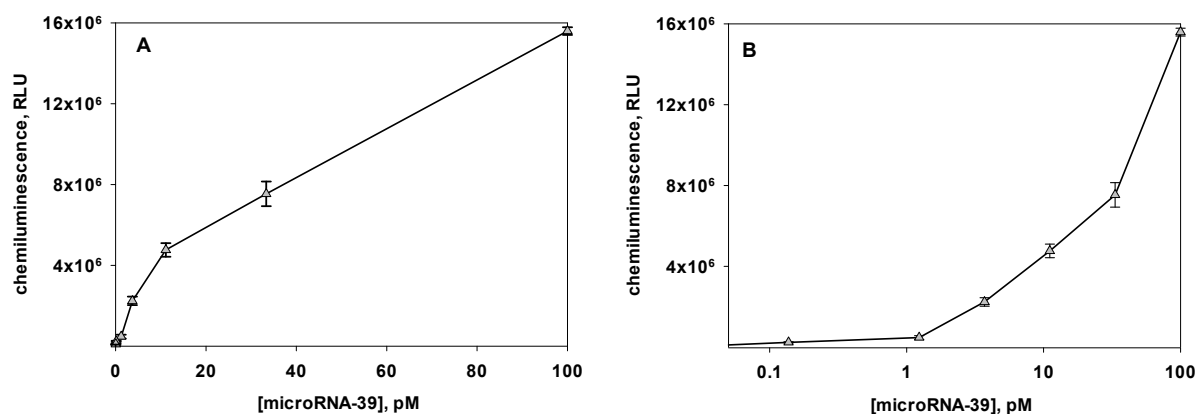


Figure S4. Calibration curve for the determination of microRNA-39 by chemiluminescent heterogenous assay based on mismatched catalytic hairpin assembly reaction ($n = 6$) presented in (A) linear and (B) semi-logarithmic coordinates. The mCHA reaction was carried out using 160 nM B-HP2 in 10 mM Tris-HCl with pH 7.2 containing 20 mM MgCl₂ at 25°C for 1 h.

Specificity of the Heterogeneous Chemiluminescent Assay of MicroRNA-39

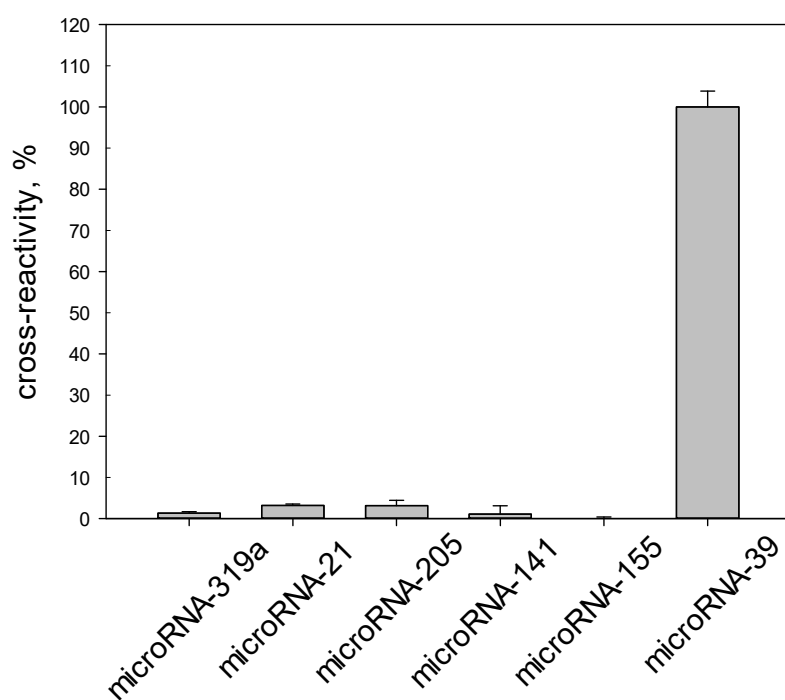


Figure S5. Specificity of the chemiluminescence heterogeneous method for the determination of microRNA-39 based on mismatched catalytic hairpin assembly reaction ($n = 3$). The concentration of the studied miRNAs was 100 pM.