

Article

# Prevalence of Antibiotic Resistance Genes in Multidrug-Resistant *Enterobacteriaceae* on Portuguese Livestock Manure

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## Supplementary Materials

**Table S1.** Primers used for the identification of chloramphenicol (*cat*), trimethoprim (*dfp*), quinolones (*qnr*, *aac(6′)-Ib*, *oqx*, *qep*), sulphonamides (*sul*) and tetracyclines (*tet*) resistance genes and for integron class.

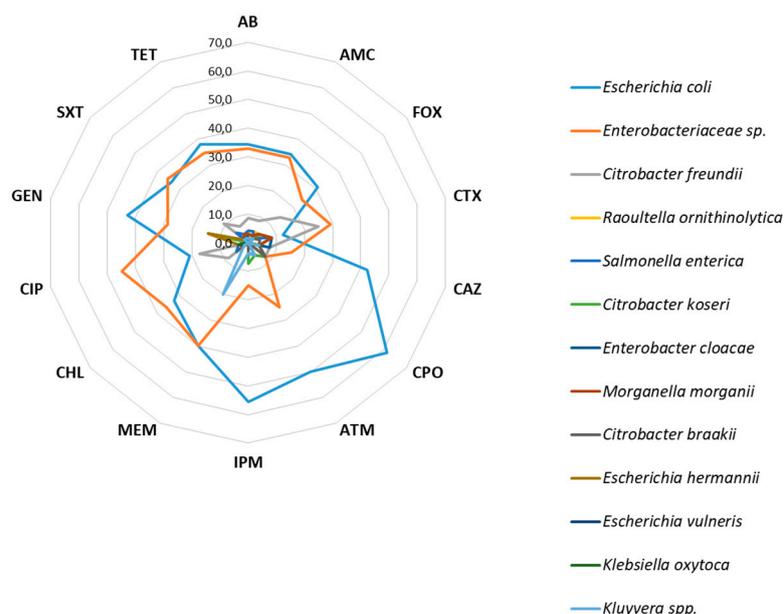
Target gene/group	Primers sequences (5′-3′) Fw/Rv	Amplicon size (bp)	Primers (μM)	Reference
<i>cat I</i>	GGTGATATGGGATAGTGT/CCATCACATACTGCATGATG	349	1.0	[1]
<i>cat II</i>	GATTGACCTGAATACCTGGAA/CCATCACATACTGCATGATG	567	1.0	[1]
<i>cat III</i>	CCATACTCATCCGATATTGA/CCATCACATACTGCATGATG	275	1.0	[1]
<i>cat IV</i>	CCGGTAAAGCGAAATTGTAT/CCATCACATACTGCATGATG	451	1.0	[1]
<i>dfp Ia</i>	GTGAAACTATCACTAATGG/ACCCTTTTGCCAGATTG	471	1.0	[2]
<i>dfp Ib</i>	TTGGGAAGGACAACGCACTT/ACCATTTCCGCCAGATCAAC	382	1.0	[3]
<i>dfp Ic</i>	GGTGAGCARAAGATYTTTCGC/TGGGAAGAAGGCGTCACCCCTC	309	1.0	[2]
<i>dfp IIa</i>	GCBAAGGDGARCAGCT/TTMCCAYATTTGATAGC	394	1.0	[2]
<i>dfp IIb</i>	AAAATTTTCATTGATTTCTGCA/TTAGCCTTTTTTCCAAATCT	471	1.0	[2]
<i>dfp IIc</i>	TTTATTGTGGTAAGCAATAC/GTATACATCTGCATCAAAAAC	201	1.0	[2]
<i>dfp IIIa</i>	ACCTGCCGATCTGCGTCAT/TCGCAGGCATAGCTGTTCTT	387	1.0	[3]
<i>dfp IIIb</i>	ACCAGAGCATTCCGTAATCA/TTGGATCACCTACCCATAGA	445	1.0	[3]
<i>dfp IIIc</i>	CACAGTCTATCGCCTTAATC/ATAGACCACAAAGCTAAACG	233	1.0	[2]
<i>dfp IVa</i>	GTTTCCGAGAATGGAGTAAT/GGTACGTGTAATCAATATTG	429	1.0	[3]
<i>dfp IVb</i>	TCACCAAGAAGTCAGAGATT/TAAAACCAGATTCGACTTTC	311	1.0	[2]
<i>dfp IVc</i>	AGAATTCCTTCTCTTTGAT/ATGCCAACAGTTGAGATTAT	218	1.0	[2]
<i>dfp Va</i>	GATCACGTRCGCAAGAARTC/GACTCGACVGCRTASCCTTC	95	1.0	[2]
<i>dfp Vb</i>	TGAACCAGAAGATTTAAAACAC/AATGGTCGGGACCTCAGAT	384	1.0	[2]
<i>dfp Vc</i>	AGTCGCTGTGGATTCTAAGT/CAATGTGAAAATTGTTCTGG	455	1.0	[2]
<i>dfp Vd</i>	ATGATTTGCTTTGGCACTTA/CCACCAATAATGAAGCATGT	250	1.0	[2]
<i>sul1</i>	CGGCGTGGGCTACCTGAACG/GCCGATCGCGTGAAGTTCCG	433	0.4	[4]
<i>sul2</i>	GCGCTCAAGGCAGATGGCATT/GCGTTTGATACCGGCTCCCGT	293	0.4	[4]
<i>sul3</i>	GAGCAAGATTTTGAATCG/CATCTGCAGCTAACCTAGGGCTTTGGA	790	0.4	[5]
<i>tet(A)</i>	GCTACATCCTGCTTGCCTTC/CATAGATCGCCGTGAAGAGG	210	1.0	[6]
<i>tet(B)</i>	TTGGTTAGGGGCAAGTTTGT/GTAATGGGCCAATAACACCG	659	0.25	[6]
<i>tet(C)</i>	CTTGAGAGCCTTCAACCCAG/ATGGTCGTCATCTACCTGCC	418	0.25	[6]
<i>tet(D)</i>	AAACCATTACGGCATTCTGC/GACCGGATACACCATCCATC	787	2.0	[6]
<i>tet(E)</i>	AAACCACATCCTCCATACGC/AAATAGGCCACAACCGTCAG	278	1.0	[6]
<i>tet(G)</i>	CAGCTTTCGGATTCTTACGG/GATTGGTGAGGCTCGTTAGC	468	1.0	[6]
<i>tet(K)</i>	TCGATAGGAACAGCAGTA/CAGCAGATCCTACTCCTT	844	1.25	[6]
<i>tet(L)</i>	TCGTTAGCGTGCTGTCATTC/GTATCCCACCAATGTAGCCG	267	1.0	[6]

Target gene/group	Primers sequences (5'-3') Fw/Rv	Amplicon size (bp)	Primers (μM)	Reference
<i>tet(M)</i>	GTGGACAAAGGTACAACGAG/CGGTAAGTTCGTACACAC	406	0.5	[6]
<i>tet(O)</i>	AACTTAGGCATTCTGGCTCAC/TCCCCTGTTCCATATCGTCA	515	1.25	[6]
<i>tet(S)</i>	CATAGACAAGCCGTTGACC/ATGTTTTTGGAAACGCCAGAG	667	0.5	[6]
<i>tetA(P)</i>	CTTGGATTGCGGAAGAAGAG/ATATGCCCATTTAACCACGC	676	1.25	[6]
<i>tet(Q)</i>	TTATACTTCTCCGGCATCG/ATCGGTTGAGAAATGTCCAC	904	1.25	[6]
<i>tet(X)</i>	CAATAATTGGTGGTGGACCC/TCTTACCTTGGACATCCCC	468	1.25	[6]
<i>qnr A</i>	AGAGGATTTCTCACGCCAGG/TGCCAGGCACAGATCTTGAC	580	0.25	[7]
<i>qnr B</i>	GGMATHGAAATTCGCCACTG/TTTGCYGYCGCCAGTCGAA	264	0.25	[7]
<i>qnr C</i>	GGGTTGTACATTTATTGAATCG/CACCTACCCATTTATTTTCA	307	0.25	[8]
<i>qnr D</i>	CGAGATCAATTTACGGGGAATA/AACAAGCTGAAGCGCCTG	465	0.25	[9]
<i>qnr S</i>	GCAAGTTCATTGAACAGGGT/TCTAAACCGTCGAGTTCGGCG	428	0.25	[7]
<i>aac(6')-Ib</i>	TTGCGATGCTCTATGAGTGGCTA/CTCGAATGCCTGGCGTGT	482	0.25	[10]
<i>oqx A</i>	CTCGGCGCATGATGCT/CCACTCTCACGGGAGACGA	392	0.25	[8]
<i>oqx B</i>	TCCTGATCTCCATTAACGCCCA/ATCCGGAACCCATCTCGATGC	131	0.25	[8]
<i>qep A</i>	CCAGCTCGGCAACTTGATAC/ATGCTCGCCTTCCAGAAAA	570	0.25	[11]
<i>intI1</i>	GGGTCAAGGATCTGGATTTTCG/ACATGCGTGAAATCATCGTCCG	465	0.3	[12]
<i>intI2</i>	CACGGATATGCGACAAAAAGGT/GTAGCAAACGAGTGACGAAATG	788	0.3	[12]
<i>intI3</i>	GCCTCCGGCAGCGACTTTTCAG/ACGGATCTGCCAAACCTGACT	979	0.3	[12]

Table S2. Multiplex PCR conditions for target genes.

Target gene/group	Cycling conditions					Final concentrations			
	1st step <sup>a</sup>	Cycles	2nd step <sup>b</sup>	3rd step <sup>c</sup>	4th step <sup>d</sup>	5th step <sup>e</sup>	MgCl <sub>2</sub> (mM)	dNTP (μM)	Taq pol. (U)
<i>cat</i> I, II, III, IV	95/3'	34x	95/60''	55/60''	72/90''	72/5'	3.0	300	1.5
<i>intI</i> 1, 2, 3	94/5'	30x	94/30''	62/30''	72/60''	72/8'	5.0	360	1.0
<i>sul</i> 1, 2	94/5'	30x	94/15''	69/30''	72/60''	72/7'	2.0	200	0.5
<i>sul</i> 3				51/30''					
<i>tet</i> A, E, G, K, L, M, O, S	94/5'	35x	94/60''	55/60''	72/90''	72/7'	3.0	300	2.5
<i>tet</i> B, C, D, A(P), Q, X							4.0		
<i>dfr</i> I	94/2'	30x	94/30''	46/30''	72/30''	72/1'	2.5	200	1.0
<i>dfr</i> II				44/30''					
<i>dfr</i> III, IV				52/30''					
<i>dfr</i> V				45/30''					
<i>qnr</i> A, B				60/45''					
<i>aac</i> (6')-Ib-cr	95/5'	35x	95/45''	52/45''	72/60''	72/10'	2.5	200	1.0
<i>qnr</i> C, D				54/45''					
<i>qep</i> A				62/45''					
<i>qnr</i> S	95/5'	35x	95/45''	52/45''	72/60''	72/10'	2.5	200	1.0
<i>qep</i> A									
<i>oqx</i> A	95/5'	35x	95/45''	52/45''	72/60''	72/10'	2.5	200	1.0
<i>oqx</i> B									

a) 1<sup>st</sup> denaturation, b) 2<sup>nd</sup> denaturation, c) annealing, d) extension, e) final extension, all expressed with temperature in °C /time in minutes, ' or seconds, ''.



**Figure S1.** Relative frequency of antibiotics resistance by species.

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