

## Article

# Detection of AFB<sub>1</sub> via TiO<sub>2</sub> Nanotubes/Au Nanoparticles/Enzyme Photoelectrochemical Biosensor

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**Abstract:** TiO<sub>2</sub> nanotubes/Au nanoparticles/enzyme photoelectrochemical biosensor is developed by the chemical bonding of acetylcholinesterase with Au nanoparticles-modified TiO<sub>2</sub> photoactive electrode, based on the inhibitory effect of aflatoxin B<sub>1</sub> on acetylcholinesterase activity. In this method, AuNPs were deposited on the surface of the electrode by potentiostatic deposition and the acetylcholinesterase was chemically crosslinked to the surface for determination of aflatoxin B<sub>1</sub>. Enzymatic hydrolysate is generated to capture the photogenerated holes of UV-sensitized TiO<sub>2</sub> nanotube arrays, causing magnification of the photoelectrochemical signal. The photoelectrochemical biosensor morphological and structural details were evaluated, applying different techniques, such as X-ray Diffraction (XRD) and Scanning Electron Microscopy (SEM). Aflatoxin B<sub>1</sub> competitively inhibits acetylcholinesterase, leading to a decrease in photocurrent that should have been increased. The detection performance of biosensors for different concentrations of AFB<sub>1</sub> is discussed. The linear response range of the biosensor is from 1–6 nM with detection limitation of 0.33 nM, the linear equation is  $I (\mu A) = -0.13C (nM) + 9.98 (\mu A)$ , with a correlation coefficient of 0.988. This new biosensor could be used to detect Aflatoxin B<sub>1</sub> in foods.

**Keywords:** photoelectrochemical; aflatoxin B<sub>1</sub>; acetylcholinesterase; TiO<sub>2</sub> nanotubes

## 1. Introduction

Aflatoxin B<sub>1</sub> (AFB<sub>1</sub>) has a strong toxicity, carcinogenicity, mutagenicity, and teratogenic toxicity, existing extensively in natural food. Previous studies have shown that AFB<sub>1</sub> is 10 times more toxic than potassium cyanide, 68 times more toxic than arsenic [1], and its carcinogenicity is 70-fold that of dimethylnitrosamine. It was also defined by the IARC (international agency for research on cancer) [2,3]. The chronic exposure of human to AFB<sub>1</sub>, even at a low concentration level, will cause the drastic health problems, such as nausea, fever, jaundice, lower extremity edema, and even fulminant hepatic failure. What is worse is that it has very stable physical and chemical properties under high temperature during cooking process. Thus, it is very important to develop a simple and rapid method to detect AFB<sub>1</sub> in the environment to protect public health and ensure

food safety. To date, a number of analysis methods are usually used to interrogate AFB<sub>1</sub> levels in foods, including liquid chromatography-mass spectrometry (LC-MS), enzyme-linked immunosorbent assay (ELISA), and immune colloidal gold technique (GICT) [4]. Other immunological-based technologies are also available to quickly detect aflatoxins, such as radioimmunoassay, time-resolved fluorescence immunoassay, fluorescence polarization immunoassay, and lateral flow immunoassay [5]. The traditional analytical tests often require expensive instruments, tedious sample preparation, pre-treatment procedures and trained testers.

The photoelectrochemical (PEC) test, a newly developed detection method, uses light and electricity energy for the sensor excitation and determination, which can effectively reduce unwanted background noise and improve sensitivity. A molecular imprinted polymer thin film for photoelectrochemical (PEC) sensing of chlorpyrifos molecules could be successfully applied to the detection of reduced chlorpyrifos in green vegetables [6]. Based on the nanocomposite of CdSe@ZnS quantum dots (QDs) and graphene deposited on the ITO coated glass electrode as a photoactive electrode, a sensitive photoelectrochemical (PEC) biosensor had been applied in the detection of organophosphorus pesticides (OPs) [7]. The fabricated derivative photoelectrochemical sensor based on the perylene-3,4,9,10-tetracarboxylic acid/titanium dioxide (PTCA/TiO<sub>2</sub>) heterojunction had been successfully applied to the detection of parathion-methyl in green vegetables [8]. The photonic charge process of the photoactive materials, such as organic dyes and inorganic semiconductor materials, are highly sensitive to the surface chemistry and microenvironment fluctuation. TiO<sub>2</sub> is a very attractive candidate for PEC detection, because of its strong light absorption, high chemical stability, environmental benignity, and low cost [9]. Among the matrixes, TiO<sub>2</sub>-based nanostructures feature a high degree of vertically oriented geometry and unidirectional charge transfer channel, thereby making them prime candidates for photocatalytic and PEC applications [10], and their semiconducting nanostructures have proven to be potential electrode materials that can immobilize biomolecules. Nonetheless, poor light absorption in the visible light spectrum and fast recombination of photoexcited electron-hole charge carriers remarkably hinder the potential applications of TiO<sub>2</sub>. To overpass these obstacles, various synthetic strategies have been explored, including metal or nonmetal doping [11,12], metal deposition [13], and heterocoupling with narrow-band gap semiconductors [14]. In our study, the introduction of precious metal Au nanoparticles on the surface of titanium dioxide, excited electrons flow from the semiconductor to the metal under light irradiation, and then the Schottky barrier between the titanium oxide and the metal nanoparticles prevents electrons from flowing to the titanium dioxide and prevents electron-hole recombination, acting as an electron trap, thereby improving the stability of the photocatalyst [15,16]. We mainly explore the simple, sensitive, and rapid photoelectrochemical method for detecting aflatoxin B<sub>1</sub> in food. In the photoelectrochemical (PEC) test of TiO<sub>2</sub>, the use of electronic detection makes the optoelectrochemical apparatus easier and low-cost when compared with traditional optical methods.

In this manuscript, a photoelectrochemical biosensor is developed by the chemical bonding of acetylcholinesterase (AChE) on TiO<sub>2</sub> photoactive electrode modified with Au nanoparticles (AuNPs), based on the inhibitory effect of AFB<sub>1</sub>. By this means, enzymatic hydrolysate (sulfhydryl) was generated to capture the photogenerated holes UV-sensitized TiO<sub>2</sub> nanotube arrays (TiO<sub>2</sub>NTs), resulting in an amplification of the photocurrent signal. AFB<sub>1</sub> could competitively inhibit AChE activity, leading to a decrease in photocurrent. In addition, we found that the bandgap effect of doped AuNPs on TiO<sub>2</sub>NTs contributed significantly to the enhanced PEC and photocatalytic performances of ternary nanostructures. This detection technique could be applied to detect aflatoxins B<sub>1</sub> in foods.

## 2. Materials and Methods

Ammonium fluoride ( $\geq 99.99\%$  trace metal basis), glycerol (anhydrous, 99.8%), dibasic sodium phosphate ( $\geq 99.0\%$ , powder), potassium dihydrogen phosphate ( $\geq 99.0\%$ , powder), acetone ( $\geq 99.0\%$ ), anhydrous ethanol ( $\geq 99.0\%$ ), chloroauric acid, chitosan (CS, deacetylation, 95%), glutaraldehyde (CAD  $\geq 99.0\%$ ), acetylcholinesterase (AChE, specific activity 1000 U/mg), 2-acetylsulfanylethyl

(trimethyl) azanium, chloride (ATCl), aflatoxin B<sub>1</sub> (AFB<sub>1</sub>), and methanol ( $\geq 99.0\%$ ) were purchased by Sigma-Aldrich St. Louis, MO, USA. Titanium tablets ( $\geq 99.7\%$ ) and aluminum sheet ( $\geq 99.7\%$ ) were used as a substrate material. All of the solutions were prepared with ultrapure water (Milli-Q, Merck, Darmstadt, Germany).

All electrochemical experiments we reported were executed with CHI660E electrochemical workstation (CH Instruments Ins., Austin, TX, USA) and three-electrode arrangement. Constant potential deposition and amperometric *i-t* method was performed with Ag/AgCl (3 M NaCl type) and platinum wire as reference and auxiliary electrodes, separately. The exposed or modified form of TiO<sub>2</sub>NTs was used as the working electrode. The potential was 0 V and sample interval is 0.1 s. The working electrode surface was irradiated by a focused UV light beam from a high-pressure mercury lamp (365 nm, F8T5). Scanning electron microscopy studies were performed with S-4800 (Hitachi S-4800, Tokyo, Japan). Samples were coated with platinum (5 nm) prior to imaging with SEM. X-ray diffraction spectra (XRD) of the samples were performed using Rigaku MiniFlex 600, Tokyo, Japan. All of the samples were detected at a scan rate of 1°/min and a step of 0.03° in the 2 $\theta$  range of 10°–90°.

## 2.1. Biosensors Preparation

### 2.1.1. Fabrication of TiO<sub>2</sub>NTs Electrode

TiO<sub>2</sub>NTs electrode was fabricated using an anodization process described elsewhere [17]. Specifically, titanium foils of thickness 0.25 mm and 99.8% purity were used to fabricate titania nanotubes. The electrolyte consisted of 0.5 wt % NH<sub>4</sub>F-glycerol and water (1:1), a platinum electrode served as a cathode, and titanium served as anode. Anodization was carried out at a constant voltage of 20 V for 2 h. Then, the anodized samples were ultrasonically cleaned in absolute ethanol and deionized water for 15 s to remove surface debris. The nanotubes were then annealed at 500 °C for 1 h in dry oxygen because these environmental conditions are known to affect the phase transition of titanium dioxide. Titanium dioxide with pore size of 100 nm was obtained ultimately.

### 2.1.2. Fabrication of Au/TiO<sub>2</sub>NTs Composite Electrode

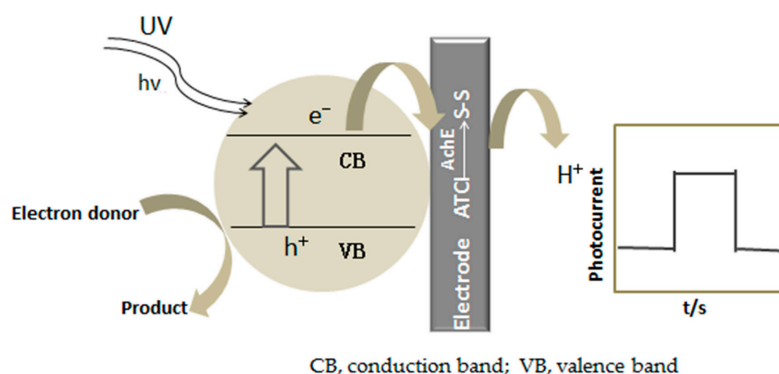
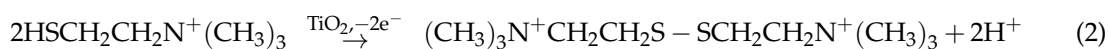
Au/TiO<sub>2</sub>NTs composite electrode was prepared by potentiostatic electrodeposition of chloroauric acid. Three-electrode arrangement was used for electrochemical deposition. The TiO<sub>2</sub>NTs electrode was immersed in a 0.1 mM HAuCl<sub>4</sub>·4H<sub>2</sub>O electrodeposition solution. After the electrochemical deposited at 0 V for 5 min using a current time profile (*i-t*), during this period, the Au nanoparticles was reduced to the electrode surface, then the electrode was rinsed with water and dried to get a Au/TiO<sub>2</sub>NTs composite electrode.

### 2.1.3. Fabrication of Au/AchE/TiO<sub>2</sub>NTs Modified Electrode

The Au/TiO<sub>2</sub>NTs composite electrode was immersed in 5% chitosan solution overnight, then dried at room temperature and immersed in 5% glutaraldehyde solution overnight. After drying at room temperature, 10  $\mu$ L of AchE (500 U/mL) was added dropwise to the electrode surface and placed in a refrigerator (4 °C).

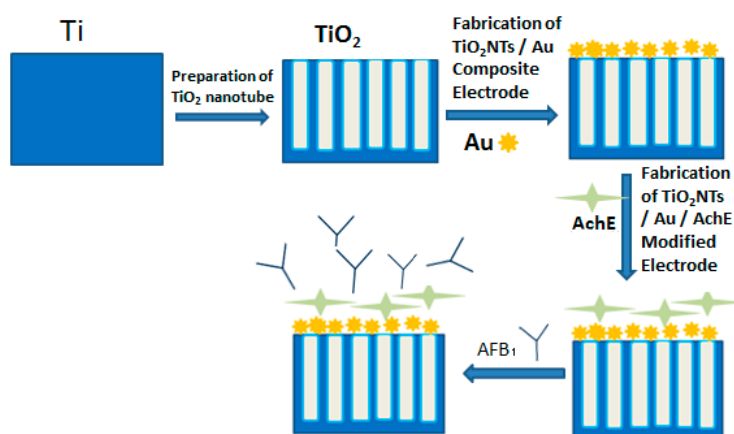
## 2.2. Procedure for Electrochemical Biosensor

The schematic diagram of the biosensor is shown in Scheme 1. TiO<sub>2</sub>NTs form an oxidation-reduction system of electron-hole pairs under the irradiation of ultraviolet light. Acetylcholinesterase catalyzes the formation of mercapto-containing cholinechloride by ATCl. The mercapto groups are easily oxidized and the photocurrent response increases as a result of the effect of electron-hole pairs system. AFB<sub>1</sub> has a strong inhibitory effect on acetylcholinesterase and irreversibly noncompetitive inhibition by altering the site of action of acetylcholinesterase [18–22]. The reaction formula is as follows:



**Scheme 1.** The schematic diagram of the biosensor photoelectrochemical process.

The schematic diagram of the biosensors assembly procedure is shown in Scheme 2. Experiments were performed by placing 100  $\mu\text{L}$  of different concentrations AFB<sub>1</sub> on top of the biosensor (i.e., Au/AchE/TiO<sub>2</sub>NTs) and allowed to stand still for 5 min. The surface of the electrode was gently washed with water and then placed in 0.1 M PBS ( $c(\text{ATCI}) = 0.1 \text{ mM}$ ,  $\text{pH} = 7.4$ ). Then, the time-current ( $i$ - $t$ ) curve was used to obtain the peak current value of different concentrations of AFB<sub>1</sub> standard solution.



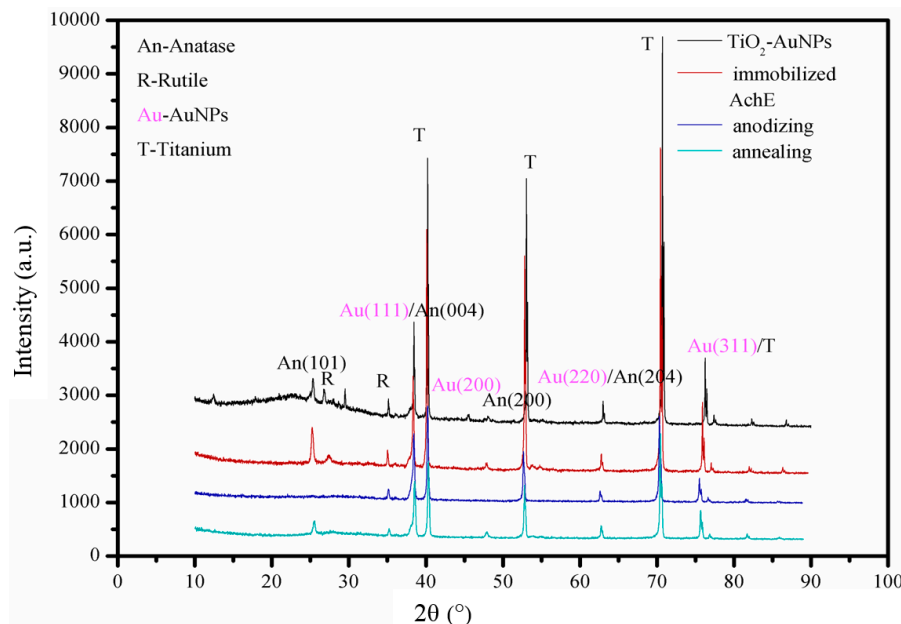
**Scheme 2.** The schematic diagram of the biosensors assembly procedure.

### 3. Results and Discussion

#### 3.1. Characterization

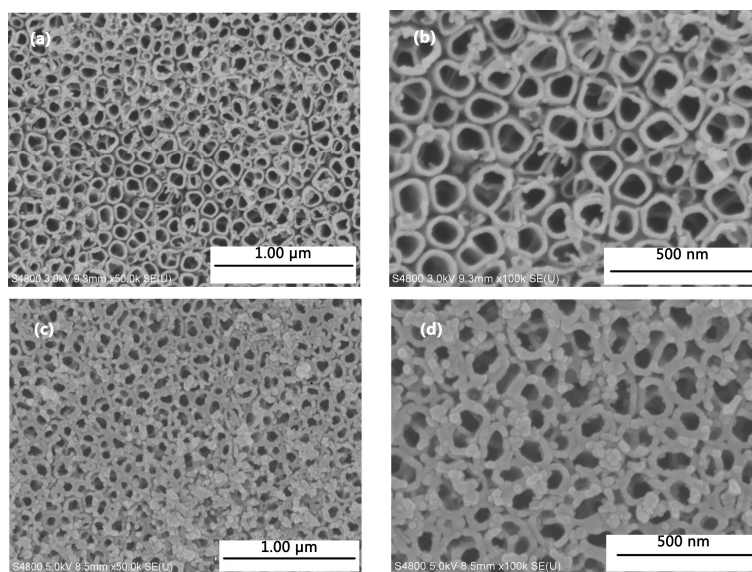
The morphology of TiO<sub>2</sub> nanotube array was also useful for separating and directing electrons to the collecting electrode surface, making it an ideal candidate for photocurrent response [23]. However, the large band gap of TiO<sub>2</sub> nanotube array (3.2 eV) determined its inherent low photoelectric conversion efficiency in the visible region, limiting its further applications [24]. Hence, Au/TiO<sub>2</sub>NTs could be employed as the photoelectric transducer in this work, because doping Au has been proved to narrow the band gap and greatly sensitize to the visible light photoresponse [25–27]. Figure 1 shows X-ray diffraction (XRD) patterns of the samples. When comparing with pure TiO<sub>2</sub>NTs

as-prepared (blue), characteristic peaks of anatase are appearing at  $2\theta$   $25.3^\circ$  and  $47.0^\circ$  after annealing (green), deposited with AuNPs (black) and immobilized AchE (red), respectively. In addition, Au characteristic peaks could be observed at  $2\theta$   $38.20^\circ$  (111),  $44.40^\circ$  (200),  $64.50^\circ$  (220), and  $77.50^\circ$  (311) after electrochemical deposition.



**Figure 1.** X-ray Diffraction (XRD) patterns of samples including anodized  $\text{TiO}_2$  nanotube arrays ( $\text{TiO}_2\text{NTs}$ ),  $\text{TiO}_2\text{NTs}$  annealed under  $500^\circ\text{C}$ , deposited AuNPs and immobilized with AchE.

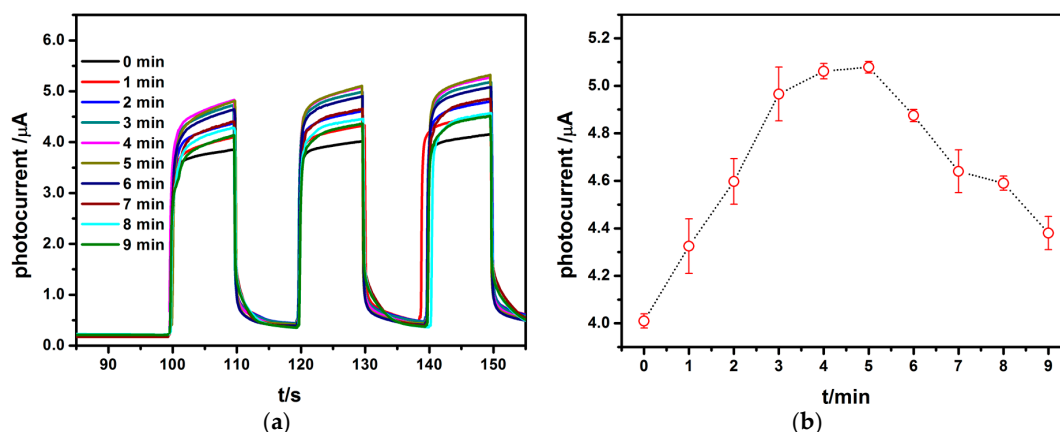
Figure 2 shows SEM images of samples.  $\text{TiO}_2\text{NTs}$  after annealed are highly self-ordered porous tubular structure, and the nozzle is more regular smooth. The average pore diameter is approximately 100 nm and the average pore spacing is approximately 150 nm (Figure 2a,b). After constant potential deposition, a mass of AuNPs is observed on the surface of  $\text{TiO}_2\text{NTs}$  (Figure 2c,d).



**Figure 2.** Scanning Electron Microscopy (SEM) micrographs of samples. Top surface view of anodized  $\text{TiO}_2\text{NTs}$  ((a)  $\times 50,000$ ; (b)  $\times 100,000$ ). Top surface view of anodized  $\text{TiO}_2\text{NTs}$  deposited with AuNPs ((c)  $\times 50,000$ ; (d)  $\times 100,000$ ).

### 3.2. Effect of AuNPs on Photocatalytic Effect of TiO<sub>2</sub>NTs

Constant potential deposition was conducted by placing the electrode (i.e., TiO<sub>2</sub>NTs) in 0.1 mM HAuCl<sub>4</sub>·4H<sub>2</sub>O (initial potential 0 V). When the energy of light is greater than the band gap of TiO<sub>2</sub>, the photoexcited electrons transit to the conduction band to form conduction band electrons, while holes leaving in the valence band. The PEC detection system is exposed to visible light. As the electrode, TiO<sub>2</sub>NTs/Au absorbs the energy of its band gap, resulting in Au valence band photoelectrons to its conduction band excitation to form electron-hole pairs. The holes in the valence band are transferred to the surface of the Au and are captured by the electron donor. At the same time, the electron enters into the TiO<sub>2</sub> and transfers to the external circuit through the conductive Ti foil to produce the photoelectric current. For improving the biosensors performance, electrochemical deposition time was investigated at first. Figure 3a is photocurrent responses of TiO<sub>2</sub>NTs/AuNPs after a different deposition time (0, 1, 2, 3, 4, 5, 6, 7, 8, 9 min). Figure 3b presents temporal evolution of peak photocurrents of the electrode (i.e., TiO<sub>2</sub>NTs). The photocurrent increases first as deposition time increases, and then decreases slightly, and a maximum value of the photocurrent as deposited appears after 5 min. The main results are summarized as follows. First, the dopant Au is introduced into the intermediate energy level for the TiO<sub>2</sub>NTs, and its larger electronegativity leads to a lower position of its conduction band, resulting in narrowing the band gap of the TiO<sub>2</sub>NTs. Thus, it is easier for the generation and transition process of photo-generated electrons. Meanwhile, it can be associated with trapping wells of photocatalytic carriers generated by the intrinsic excitation of TiO<sub>2</sub>. The shallow trapping facilitates the diffusion process of the excited carriers in the TiO<sub>2</sub>NTs, prolongs the life of the excited carriers, greatly reduces the surface recombination of the electron hole pairs, and enhances the photocatalytic activity of the photocatalysts. However, the photocurrent increases first and then decreases with the increasing of deposition time, because that excess doping AuNPs inhibit the light absorption of TiO<sub>2</sub> and reduce the photocatalytic activity of TiO<sub>2</sub> [28].

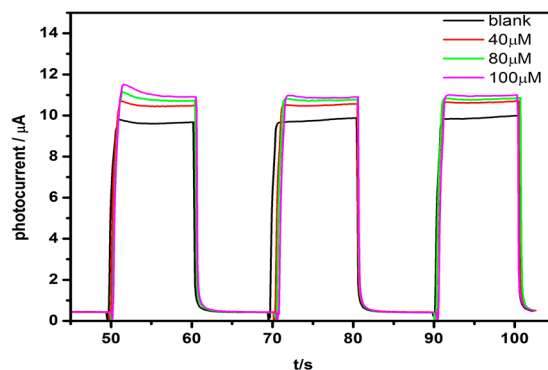


**Figure 3.** Photocurrent responses of TiO<sub>2</sub>NTs/Au nanoparticles (AuNPs) after different deposition time (0, 1, 2, 3, 4, 5, 6, 7, 8, 9 min): (a) the photocurrent curves of the TiO<sub>2</sub>NTs deposited for different times in 0.1 mM HAuCl<sub>4</sub>·4H<sub>2</sub>O under deposition potential 0 V; (b) peak currents of the TiO<sub>2</sub> NTs for different deposition times ( $N = 3$ ).

### 3.3. Effect of Acetylcholine Concentration on Photocurrent Response of Enzyme Biosensor

For bioinhibitory sensors, substrate concentration is one of the important influence parameters. AuNPs/AChE/TiO<sub>2</sub>NTs electrode was placed in different concentrations of ATCl (40 μM, 80 μM, 100 μM) with 0.1 mM, pH = 7.4 phosphate buffered saline (PBS) in order to investigate the effects on the response of the sensor. Embedded AChE biological activity will be affected by the solution pH. According to the literature, the optimal pH of acetylcholinesterase is closed to 7 [29,30]. As shown in Figure 4, the photocatalyses of the enzyme biosensor are found to become more obvious as the concentration of ATCl solution increased. This experimental phenomenon is ascribed to the catalytic

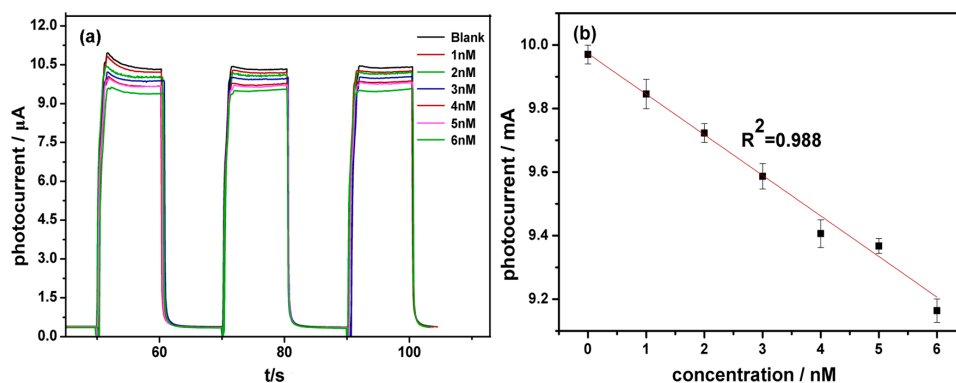
reaction of AChE to ATCl. The mercapto that is generated by the catalytic reaction is easily oxidized by the electron-hole pair system that is produced by TiO<sub>2</sub>NTs, so that the photocurrent response increases. However, when concentration of ATCl is increased more than 80  $\mu$ M, the change of photocurrent response is very slight, indicating that the catalytic reaction is approached to equilibration. Therefore, the later experiments were performed with ATCl concentrations of 0.1 mM.



**Figure 4.** The photocurrent responses of the AuNPs/AChE/TiO<sub>2</sub>NTs modified electrode in different concentration of ATCl solutions.

### 3.4. Performance of the Biosensor

As mixed, AFB<sub>1</sub> can inhibit AChE enzyme activity by blocking access of the substrate to the active site or by inducing a defective conformational change in the enzyme through non-covalent bonding interacting with the AChE peripheral binding site. When AFB<sub>1</sub> is added, the activity of AChE is inhibited, thereby inhibiting the hydrolysis of ATCl and reducing the generated electroactive substance choline and acetic acid, reducing the oxidation current [31]. When comparing the change of the photocurrent of the enzymatic reaction, the concentration of AFB<sub>1</sub> could be obtained via the inhibitory rate of AFB<sub>1</sub> to AChE. The responses of the biosensor to AFB<sub>1</sub> were recorded with the current-time curve as shown in Figure 5. The attenuation of the photocurrent response is dependent on AFB<sub>1</sub> concentration. The peak current is inversely proportional to the analyte concentration, which demonstrates that AFB<sub>1</sub> indeed inhibits the catalytic reaction of AChE by the two mechanisms described previously. The response time of the sensor is about 5 s. The time for the calculation curve is selected as the current approached steady state. For example, it is about 55, 75, or 95 s. The linear response range of the biosensor is from 1–6 nM with detection limitation of 0.33 nM, the linear equation is  $I$  ( $\mu$ A) =  $-0.13C$  (nM) + 9.98 ( $\mu$ A) with a correlation coefficient of 0.988. The results show that TiO<sub>2</sub> nanotubes/Au nanoparticles/enzyme photoelectrochemical biosensor has a certain feasibility.



**Figure 5.** The photocurrent responses of AFB<sub>1</sub>: (a) the photocurrent responses of AuNPs/AChE/TiO<sub>2</sub>NTs in 0.1 M PBS solution (pH 7.4) containing 0.1 mM ATCl after adding different concentration of AFB<sub>1</sub>; (b) standard curve line of different concentration of AFB<sub>1</sub>,  $N = 3$ ).

The comparison of aflatoxin B<sub>1</sub> detection methods by several different biosensors is displayed in Table 1. The detection line of TiO<sub>2</sub> nanotubes/Au nanoparticles/enzyme photoelectrochemical biosensor is second only to the optical biosensor based on gold nanorods (GNRs) [32–46]. However, TiO<sub>2</sub> nanotubes/Au nanoparticles/enzyme photoelectrochemical biosensor do not require the purchase of expensive antibodies, so that the detection steps are simpler and less costly [43,44].

**Table 1.** Several Biosensors for aflatoxin B<sub>1</sub> detection.

Methods	Detection Limit or IC <sub>50</sub>	Ref.
Au/AchE/TiO <sub>2</sub> NTs	0.33 nM	This experiment
AFO/MWCNTS/Pt	1.6 nM	[32–34]
Amperometric, screen printed electrode modified with CoPc	302 µM	[35–37]
AchE-based Conductometric Biosensor	0.05 µg/mL	[38–41]
Amperometric, screen printed electrode	IC <sub>50</sub> = 100 ppb	[42]
Aflatoxin B <sub>1</sub> based on aggregation of gold nanorods	0.04 ppb	[43,44]
Detection of aflatoxin B <sub>1</sub> in corn and nut products using the array biosensor	0.3 ng/mL	[45,46]

#### 4. Conclusions

A new photoelectrochemical biosensor is developed via chemical bonding of AchE with Au NPs-modified TiO<sub>2</sub>NTs from the present work. Two steps were used to synthesize the Au/AchE/TiO<sub>2</sub> electrode. In Step 1, immobilizing the nanogold to CS-GAD mixed film that was applied to chemical deposition using nano-gold constant potential deposition. In Step 2, AchE was modified to the electrode surface of CS-GAD mixed membrane by chemical cross-linking. It was shown that TiO<sub>2</sub>NTs electrode deposited Au nanoparticles obtained more obvious photocurrent response signal. AFB<sub>1</sub> can inhibit the enzyme activity of AchE, leading to a decrease in photocurrent. Through the electrochemical detection, the linear response range of the biosensor is from 1–6 nM with detection limitation of 0.33 nM, which is significantly more sensitive than other methods, the linear equation is  $I (\mu A) = -0.13C (nM) + 9.98 (\mu A)$  with a correlation coefficient of 0.988. This photoelectrochemical biosensor could be applied to detect aflatoxins B<sub>1</sub> in foods.

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**Author Contributions:** Chengyong Li proposed the research topic and revised the paper; Qiong Yuan designed the experiments and wrote the paper, Chuxian He, Rijian Mo and Lei He performed the experiments; Chunxia Zhou measured XRD patterns; Pengzhi Hong observed SEM images; Shengli Sun analyzed the data.

**Conflicts of Interest:** The authors declare no conflict of interest.

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