



Cosmetic Ingredients as Emerging Pollutants of Environmental and Health Concern. A Mini-Review

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Abstract: Cosmetic and personal care products are used in huge quantities throughout the world; as a result of their regular use, they are continuously released into the environment in very large amounts. Many of these products are biologically active and are characterized by persistence and bioaccumulation potential, posing a threat to ecosystem and human health. On the basis of the most recent scientific literature available on this subject, this paper provides an overview of some cosmetic ingredients that are considered environmental emerging pollutants of particular concern such as UV filters, some preservatives (parabens, triclosan), and microplastics.

Keywords: cosmetics; UV filters; parabens; triclosan; microplastics; environmental pollutants

1. Introduction

According to EU Regulation 1223/2009 (article 2, 1.a), "cosmetic product means any substance or mixture intended to be placed in contact with the external parts of the human body (epidermis, hair system, nails, lips and external genital organs) or with the teeth and the mucous membranes of the oral cavity with a view exclusively or mainly to cleaning them, perfuming them, changing their appearance, protecting them, keeping them in good condition or correcting body odours" [1]. According to a widely accepted classification, cosmetics can be divided into leave-on and rinse-off products. A leave-on cosmetic is a product that for its functions is intended to stay on the skin for a rather extended period; examples are perfumes, decorative cosmetics, body and face creams, and antiperspirants. On the contrary, a rinse-off cosmetic is a product designed to be rinsed off after a short stay on the skin or mucous membranes, such as shampoos, soaps, shower gels, and toothpastes.

In the last few years, cosmetics, as well as pharmaceuticals and many other products for personal care that do not fall within cosmetic regulation (disinfectants, insect repellents, dietary supplements), have raised significant concerns as one of the most important classes of emerging pollutants because they are continually released into the aquatic environment; their ecological and environmental impact is associated with the large amounts used and with the fact that sometimes they are environmentally persistent, bioactive, and potentially able to bioaccumulate [2]. Moreover, sewage treatment plants are not always effective in removing chemicals used as cosmetic ingredients, as shown, for example, with synthetic musks [3,4], perfluoroalkyls compounds [5], some organic UV-filters [6] and microplastics [7]. Another object of concern is that some of these products can accumulate in sewage sludge during wastewater treatment and then enter the environment because of the common practice of using sludge as a fertilizer on crops [8]. Cosmetics pose the most pressing ecological problems compared to pharmaceuticals because they are used in much larger quantities and throughout the course of life and, being intended for external application, are not subjected to metabolic transformation; therefore they are introduced unaltered into the environment in large amounts during washing, showering, or bathing [9]. Since relatively little is known about the fate and the toxicity of personal care products

released into the environment, increasing attention is being placed on their occurrence, persistence, and potential threat to ecosystems and human health.

The role of some cosmetic ingredients in environmental pollution is well-known; this is the case, for example, with surfactants. As a result of their presence in a broad range of personal care products intended for cleaning purposes (body wash, shampoo, hand cleanser), huge amounts of surfactants are discharged every day into aquatic and terrestrial environments, with widely recognised ecological risk and toxic effects on plant and animal species and human health [10]. The objective of this review was instead to provide an overview of some commonly used cosmetic ingredients that have not historically been considered of ecological concern but are now being monitored because of their suspected adverse ecological effects; in particular, we focused our attention on UV filters, parabens, triclosan, and microplastics, which are being considered as emerging environmental contaminants. The stylized world map of Figure 1 shows the sampling sites cited in research papers used to compile this review and allows the reader to know at a glance where the research efforts are focused.



Figure 1. Graphic overview of the sampling locations for waters, sediments, and/or organisms cited in the References. \bullet = UV filters (References [5–31]); * = parabens (References [32–42]); \blacklozenge = triclosan (References [34,42–51]); **X** = plastic microbeads (References [52–57]).

2. UV Filters

UV-filters are chemicals that absorb or reflect ultraviolet radiation in sunlight; they are widely used in a variety of personal care products to protect the skin from UV-induced damage and sometimes to stabilize the colour and the fragrance of cosmetic formulations. In the European Union, the UV-filters approved for use in cosmetic products number 28 and are listed in Annex VI of the European Regulation on Cosmetic Products [1]. UV filters can provide protection against UV-A rays (400–320 nm), UV-B rays (320–280 nm), or both (so-called broad-spectrum filters). Currently, the active ingredients of sunscreens are classified as organic or inorganic chemical absorbers. Organic filters (formerly known as "chemical filters") absorb radiation and then dissipate the absorbed energy through photophysical and photochemical pathways; inorganic mineral filters (once named "physical filters") act partly by absorbing and partly reflecting and scattering UV radiation [11,12]. UV filters, which sometimes can account for up to 20% of sunscreen formulations [13], are being considered emerging pollutants due to their widespread presence in the environment. They can reach the aquatic environment directly, via wash-off from the skin surface during recreational activities (e.g., swimming, bathing) as well as indirectly through wastewater from the use of personal care products, washing clothes, and industrial

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discharges; probably the most important sources of pollution are represented by the effluents from wastewater treatment plants and sewage sludge, often used as a fertilizer in agriculture [6,14].

The occurrence of UV filters is widely documented in different substrates and many different geographical locations. UV filters have been detected, for example, in surface waters such as seas, oceans and coastal waters [15–17], rivers [15,18], lakes [19,20,58], groundwater [59], and sediments [21]. Organic and inorganic UV filters will be discussed below separately because they raise distinct environmental and health issues.

2.1. Organic Filters

The distribution and the fate of organic UV filters entering the aquatic environment depend on their physico-chemical properties and their stability toward bio-and photodegradation. Some organic UV filters are characterized by stability in the aquatic environment and lipophilicity, with an organic partition coefficient in general greater than three [60]; therefore they can accumulate in sediment of body waters, a matrice with high organic carbon content, where they can act as a contaminant reserve posing high risk to benthic and sediment-dwelling organisms [22,60]. The presence of UV filters in riverine and marine sediment is widely documented in the literature. The filters most frequently identified in sediment are EHMC (ethylhexyl methoxycinnamate), OC (octocrylene), BMDM (butyl methoxydibenzoylmethane), OD-PABA (octyldimethyl-*p*-aminobenzoic acid), and benzophenone derivatives [21–23,60–63], with concentrations included in very broad ranges and, in some cases, amounting to hundreds or thousands of ng/g of dry weight. Other sun blockers (4-methylbenzyliden camphor, 4-MBC; isoamyl *p*-methoxycinnamate, IAMC; ethylhexyl salicylate, EHS; homosalate HMS), although detectable in surface waters, are not recovered in sediment, probably due to their reduced use in sunscreen formulations or to their degradation by microorganisms [64].

The occurrence of common organic UV filters has been documented in water samples collected in different countries, such as China, USA, Japan, Thailand and the North American Arctic region [15,60], Hong Kong [60], the Canary Islands [17], Switzerland [20], Korea [18], Taiwan [16], and Norway [24], to mention just the most recent investigations. Surprisingly, Tsui et al. [15] reported the occurrence of UV filters even in the Arctic region, where benzophenone-3 (BP-3), EHMC, and OC were detected at relatively low concentrations (median concentrations 16.6, 25.4, and 25.8 ng/L, respectively). The pathways by which organic UV filters arrive in the Arctic region are yet to be elucidated. Oceanic transport via ocean currents seems to be the most likely, especially for filters (e.g., BP-3 and OC) that are highly photostable towards UV irradiation, though atmospheric transport cannot be excluded. The concentrations of UV organic filters detected in these studies were comprised of a wide range of values, reaching in some cases hundreds or thousands of ng/L([15], [25]) and the literature therein). Variability in the composition profiles of UV filters was detected, and the concentrations reported depended on different factors; for example, the sampling location, differences in population densities, and the development level at each location, as well as differences in each national regulatory body for the cosmetic sector. UV filter concentrations were in general subjected to predictably large seasonal fluctuations (some filters exceeded 1000 ng/L in sea water samples collected at a popular beach in Hong Kong on hot summer days with strong UV radiation; [15]). An exception was constituted by Canary Island Gran Canaria, where no clear seasonal pattern was detected, probably due to an assiduous frequentation of its beaches during the year [17].

The Literature reports the occurrence of sunscreen organic compounds also in the water of swimming pools. The presence and concentration of UV filters in these facilities depends basically on their washing off from the skin of bathers during swimming and on the bather load. In swimming pools, UV filters occur at higher concentrations compared to other water bodies (rivers, lakes, seas), where chemical pollutants undergo high dilution, while in swimming pools the water is recycled [65]. Interestingly, [66] compared the sunscreen concentration in children's and adult's swimming pools and detected the highest concentrations of these compounds in baby pools, with amounts of sun blockers being ten times higher for the same number of users. Although the fate of UV filters in these closed

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systems is not fully understood, it has been shown that they can undergo photodegradation and can react with the chlorine used to disinfect the water, with the formation of by-products being potentially harmful [67–70].

In aqueous environments, organic UV-filters can degrade by direct or indirect photolysis; the first process depends on the absorption of UV radiation by the organic compound, while the second one implies the participation of different photosensitizer agents (such as dissolved organic matter or reactive oxygen species (ROS)); the ecotoxicity of these by-products remains to be elucidated. Under UV radiation, some organic filters are also able to produce ROS [25]. Organic UV filters have been found in several aquatic organisms (marine invertebrates, fish, marine mammals, aquatic birds); often they were detected at significantly high concentrations (e.g., OC at up to 712 ng/g in the liver of the Brazilian dolphin *Pontoporia blainvillei*, [26]; octyl methoxycinnamate (OMC) at up to 256 ng/g dry weight, and OC at up to 7112 ng/g dry weight of *Mytilus* sp. collected along French coasts, [27]; EHMC at up to 701 ng/g lipids of the cormorant *Phalacrocorax* sp., [28]). Gago-Ferrero et al. [29] reported that several fish species from Iberian rivers contained detectable concentrations of lipophilic UV filters (BP-3, EHMC, 4-MBC, and OC) in the range of ng/g of dry weight. Overall these data show that some organic UV filters are able to bioaccumulate in the muscle and lipids of these organisms and are likely to enter marine food chains.

The toxicity of organic UV filters was evaluated in several aquatic organisms; some frequently used UV filters have proven to be toxic for phytoplankton species, microalgae, protozoa, and crustaceans ([25] and the literature therein). Danovaro et al. [71] demonstrated that some UV filters (EHMC, BP-3, and 4-MBC) can cause coral bleaching at very low concentrations; the results of their experiments, jointly with the estimated release of sunscreens in reef areas, strongly suggest that at least 10% of the reefs are likely to meet coral bleaching due to UV filters pollution. A number of studies also suggest that some organic UV filters possess, similarly to many other xenobiotics, endocrine-disruption capacities, in both in vitro and in vivo models. BP-3, for example, displayed a weak estrogenicity in vitro and strong anti-androgenic and weak estrogenic (but anti-estrogenic activity as well) activity in murine models [63]; moreover, adverse effects of this compound on reproduction of aquatic organisms were reported [63]. Kunz et al. [72] investigated the estrogenic activity of a panel of organic UV filters both in vitro, with a receptor-based assay, and in vivo on fish; the experiments demonstrated the potential endocrine-disrupting activity in vivo of 3-benzylidene camphor (3-BC), benzophenone-1 (BP-1), and benzophenone-2 (BP-2). In a rodent model of sexual brain differentiation, two UV filters with a known estrogenic-like activity, 4-MBC and 3-BC, were able to impair female sexual behaviour [73]. It is noteworthy that human milk samples analysed by Schlumpf et al. [74] contained UV filters in over 75% of the cases, indicating a possible exposure of the suckling neonate to these chemicals.

2.2. Inorganic Filters

The inorganic compounds approved by European Regulation as sunscreen are TiO_2 and ZnO. TiO_2 was authorized as a non-nano UV filter by the EU Cosmetic Regulation, Annex VI, while TiO_2 (nano) was introduced in this Annex by Commission Regulation (EU) 2016/1143 [75]. In May 2016, the European Commission recognized the use of ZnO as a UV filter in cosmetic products as safe, adding both in its nano and non-nano forms to Annex VI of the EU Cosmetic Regulation [76]. TiO_2 and ZnO are almost invariably present in sun blocker formulations as nanoparticles because the nanoscale improves skin retention, cosmetic acceptance for consumers, and the UV attenuation properties of these oxides when compared to their bulk counterparts [11]. The release of these compounds from cosmetics and personal care products applied on skin into the environment can occur due to immersion or abrasion with sand [77]. TiO_2 is also released into aquatic environments from sources other than sunscreens (anthropogenic input due to its use as a food ingredient, textile washing, leaching from facade coatings); wastewater treatment plants (WWTPs) remove the majority of TiO₂ present in the influent sewage, but a small amount is released into natural water bodies [30,77]. TiO_2 residues

released from sunscreen produce submicronic aggregates that remain in suspension in fresh water, while in seawater conditions (i.e., in presence of high salt concentrations) they aggregate and gradually settle, being confined to the sediment [77].

Nano TiO₂ and ZnO can undergo photocatalitic reactions when exposed to ultraviolet radiation, with the generation of ROS such as O_2^{-} , OH, and H_2O_2 [78]; TiO₂ and ZnO extracted from commercial sunscreens can react with some chemicals and cause corrosion to paint surfaces when irradiated with UV rays [79,80]. This photoactivity can present the risk of cellular damage and environmental toxicity. To prevent the production of ROS, TiO₂ nanoparticles present in sunscreen are coated with silica and alumina and/or doped with manganese or vanadium, even though sometimes these coatings do not withstand contact with water [81].

To quantify the impact of inorganic UV filters release in coastal areas, Sánchez-Quiles and Tovar-Sánchez [31] performed laboratory experiments by suspending 1 g·L⁻¹ of a commercial sunscreen containing nano TiO₂ (77.6 mg \cdot g⁻¹) in coastal seawater collected from Palmira Beach (Majorca Island, Spain); in 22 h, under controlled UV irradiation, $10,202 \pm 606$ nM of H₂O₂ was produced in seawater, i.e., 447 times the initial levels detected (22.8 \pm 2.6 nm). The authors also investigated H₂O₂ and Ti⁴⁺ concentration in the water column at the same beach during 24 h of a summer day; the highest concentrations were measured at noon (278.0 \pm 11.6 nM for H_2O_2 and 833.7 ± 50.1 for Ti⁴⁺). Finally, Sánchez-Quiles and Tovar-Sánchez [31] estimated the production of H_2O_2 ascribable to TiO₂ in the coastal waters of Palmira beach. Taking into account the average content of TiO_2 in commercial sunscreens (46 mg/g), the recommended amount of product that an adult should use for a typical application (36 g), the percentage of sunscreen that is removed by water during recreational activities (about 25%), and the average number of beachgoers per summer day at that location (approximately 10,000), the authors estimated a summer daily release of nano-TiO₂ of approximately 4 kg and an increment in the concentration of H_2O_2 of 270 nM/day, with the highest values at noon. These results suggest that the normal recreational activities in coastal resorts can result in the production of significant amount of H_2O_2 and consequent damage to or death of marine coastal phytoplankton; this could have reverberations on the marine food web, which relies on these microorganisms.

Studies on the toxic effects of nano-TiO₂ on algae, freshwater and marine invertebrates, and fish were reported in recent years in several papers ([82] and the literature therein, [83]). The partial information available on the physicochemical characteristics of the nanoparticles tested in different investigations and the different endpoints examined does not allow a reasonable comparison of the data obtained and thus does not permit firm conclusions to be drawn about the ecotoxicity of nano-TiO₂. Therefore, even though in some papers nano-TiO₂ has been reported to have negligible effects on various aquatic organisms [84–86], a dramatic increase of the toxicity of this compound occurs in the presence of exposition to strong sunlight or UV radiation due to the production of ROS [87–89]. As far as ZnO is concerned, its toxicity for aquatic organisms is well documented; for example, for zebrafish [90,91], marine algae [92,93], sea urchins [94], and other marine organisms [95,96], nano-ZnO was classified "extremely toxic" [95]. In contrast to TiO₂, the toxicity of nano-ZnO and the corresponding bulk particles is quite similar [83]; experimental data demonstrated that the toxic effect of ZnO is partly ascribable to the release of Zn⁺⁺ ions and partly to oxidative stress due to ROS generation [83,97].

Jeon et al. [98] evaluated the potential risk for human health of TiO₂ and ZnO nanoparticles release from commercial sunscreens into the water of an outdoor swimming pool in Seoul. The authors demonstrated that significant amounts nanoparticles were released from sunscreen-applied skin into the water during swimming activity, producing hydrogen peroxide; however, the estimated concentrations of H_2O_2 generated as a consequence of sunlight and UV irradiation in a typical day of pool activities (0.011–0.139 μ M) were lower than those required to cause adverse effects on human health.

3. Parabens

Parabens are a class of preservatives that have been used for nearly a century for their antimicrobial properties in a wide range of products, such as cosmetics, foodstuffs, and pharmaceuticals; chemically they are esters of the *para*-hydroxybenzoic acid, with alkyl (methyl, ethyl, propyl, isopropyl, butyl, isobutyl, pentyl, heptyl) or aryl (benzyl, phenyl) substituents [99]. Although some of these compounds occur naturally since they are synthesized by bacteria or plants [100,101], all commercially available parabens are produced synthetically. In the European Union the use of parabens in cosmetics is regulated by the European Regulation on Cosmetic Products (2009) [1]; they are listed in Annex V and are allowed at a maximum concentration of 0.4% (w/w) for single ester and of 0.8% (w/w), expressed as *p*-hydroxybenzoic acid, for paraben mixtures. It is worth mentioning that the European Union has recently placed some restrictions on the parabens allowed in cosmetic formulations; in particular Regulation (EU) No 358/2014 recently amended Annex V of Regulation (EU) No 1223/2009 to prohibit the use of isopropylparaben, isobutylparaben, phenylparaben, benzylparaben, and pentylparaben in cosmetics [102]. Moreover, Regulation (EU) No 1004/2014 lowered the maximum concentration allowed for propylparaben and butylparaben, and prohibited their use in products for children [103].

In spite of these recent restrictions, parabens are still extensively employed in cosmetics and personal care products because of their broad spectrum of activity against yeasts, molds, and bacteria and their chemical stability, low toxicity, and low cost [99].

Parabens can be detected in air, dust, soil, and water [99]. They can be discharged in the environment from their production factories, but more importantly they are continuously released at relatively high levels into urban and hospital wastewaters, as a result of the wide use of personal care products, pharmaceuticals, and household chemicals containing them. In raw wastewaters, parabens can reach values up to 30,000 ng/L for methylparaben and 20,000 ng/L for propylparaben [104]. Parabens are present almost exclusively in the aqueous phase, due to their moderate water solubility (e.g., 2500 mg/L for methylparaben, 885 mg/L for ethylparaben; [104]); when the occurrence of five parabens was assessed in influent and effluent samples from three Tunisian wastewater treatment plants, they were detected in water at concentrations of mg/L, whereas their concentrations in suspended matter were some orders of magnitude lower ($\mu g/L$; [32]). Wastewater treatment plants effectively remove most parabens present in the aqueous phase, with an average removal rate of >90% [104,105]. In these plants, parabens, can adhere to the sewage sludge or can be degraded; in particular, they can undergo fast biodegradation, with the production of *p*-hydroxybenzoic acid as a main degradation product, or can react easily with free chlorine, forming several chlorinated by-products [106]. Halogenated parabens are removed as efficiently as their parent compounds in wastewater treatment plants, although they require more time to be degraded [105]. These halogenated derivatives can also be generated when paraben-containing cosmetics are mixed with chlorinated tap water at home; moreover, if traces of bromide are present in tap water, brominated parabens are preferentially formed [106]. Unfortunately, little is known about the potential risk of these halogenated derivatives for human health [104].

Despite their efficient removal in wastewater treatment plants, parabens are still detected in surface waters, especially rivers [33–36]. Methylparaben and propylparaben, the most common parabens present in cosmetics, are also the parabens most frequently detected in surface waters [99], with a wide range of concentrations that reach the highest values in Chinese rivers (1062 ng/L for methylparaben and 3142 ng/L for propylparaben) [33]). To the best of our knowledge there are no specific studies regarding the presence of parabens in marine and ocean waters; however, the occurrence of these preservatives and their metabolites (especially *para*-hydroxybenzoic acid) is well documented in several animals living in the sea or in coastal areas. Paraben occurrence was in fact reported in fish tissues [37–39], in marine mammals [40], and in the tissues of marine birds and their eggs [39]. The presence of parabens and their metabolite, *p*-hydroxybenzoic acid, even in animals from remote marine locations, such as polar bears from Alaska [40] and albatrosses from Midway

Atoll (Northwestern Pacific Ocean) [39], suggest that these compounds are ubiquitous in the oceanic environment. The occurrence of parabens in drinking water was investigated in a limited number of studies [104]. Generally parabens were not observed or were detected at very low concentrations in tap water; however, a recent investigation recorded the presence of parabens at concentrations of ng/L in bottled water [41].

Parabens have been detected in human urine, serum, adipose tissue, milk, amniotic fluid and placental tissue [107–113], and in human breast cancer tissues [114,115]. In humans, dermal exposure to paraben-containing products is important for their absorption because some parabens penetrate the human skin [116]; the systemic absorption of parabens after environmental exposure has also been confirmed, although it seems to be a less important paraben source [38]. Oral ingestion seems to be less important, as orally ingested parabens are metabolized by esterases to *p*-hydroxybenzoic acid [116].

For a long time, parabens were considered to have a very low toxicity and an excellent safety record; however, recently, concerns have been raised about their harmlessness, as parabens display endocrine-disrupting activity both in vitro and in vivo, and they have been associated with alterations of the reproductive system of male experimental animals [116]. In in vitro investigations, parabens exhibited a weak estrogenic activity, with a binding affinity for the estrogen receptors about five orders of magnitude lower than that of diethylstilboestrol [117]. Studies on the potential endocrine disruption of parabens on aquatic wildlife demonstrated that these compounds are able to interfere with the vitellogenin plasma concentration in some organisms, such as rainbow trout and Japanese medaka fish, although at concentrations largely exceeding the maximum concentrations detected in surface waters [99]. As far as the human species is concerned, several studies have been published claiming a link between the endocrine-disrupting activity of parabens and breast cancer [114,118]. These studies were subsequently rejected by a number of scientists and international health authorities, which concluded that there is no scientific evidence linking cosmetics or parabens with human breast cancer [119–122]. However, some recent studies seem to suggest a more cautious attitude. Pan et al. [123] described a potential synergy between parabens and epidermal growth factor, suggesting that parabens can be active at concentrations lower than those previously considered toxicologically relevant. Similarly, a recent paper reported that in immature Sprague-Dawley rats, after intragastric administration of paraben doses close to the acceptable daily intake, the in vivo estrogenicity of parabens is not as low as established by in vitro experiments [124].

Overall, current scientific knowledge is inadequate to demonstrate a clear correlation between parabens and human breast cancer risk, and further research would be necessary to definitively exclude this association; therefore, this controversy is still ongoing. In the light of current knowledge, it seems reasonable not to underestimate the possible consequences for human health of the continuous introduction of parabens into the environment.

4. Triclosan

Triclosan (TCS, 5-chloro-2-(2,4-dichlorophenoxy)phenol) is a lipid-soluble, broad-spectrum, antimicrobial agent, allowed as a preservative in personal care products such as hand soaps, shampoos, detergents, toothpastes, sunscreen, and deodorants, where it can be used at concentrations up to 0.3% (Annex V of European Regulation on Cosmetic Products) [1]. Due to its antibacterial and antifungal properties, TCS is also widely employed as an antiseptic, in medical devices, in household items, and as an additive in packaging, textiles, and functional clothing [125]. The most important pollution source of TCS is domestic sewage, where this compound is released with the rinsing of products containing it; the extensive use, the large amounts emitted and its incomplete removal in sewage treatment plants lead to the often considerable presence of this compound in the environment, with concentrations detected ranging from nanograms to micrograms per liter in sediments and waters [43,44]. TCS is found ubiquitously in the aquatic environment, where it partitions between sediment and water and can be thus detected in both phases [43,45], with a preferential accumulation in sediments, due to its lipophilicity. TCS is among the top 10 most commonly detected organic wastewater

compounds for frequency and concentration [2]. Its presence is ubiquitous; it has been found in surface water throughout the United States [46–48], the United Kingdom [34], South Korea [49], and China [126,127]. The occurrence and the concentration of TCS were recently examined in the Parisian sewer network [42], and high levels were found in wastewater (2140–5260 ng·L⁻¹). The results of this study were extrapolated at the national scale to give a national mass load of 18.8–22.2 tons year⁻¹. In a study performed on 13 sites in Japanese rivers contaminated by domestic wastewater, sewage treatment plant effluent, or industrial wastewater, TCS was detected ubiquitously with concentrations up to 177 ng L^{-1} and little difference in concentration between sites with and without a sewerage service [36]. TCS presence has been also documented in terrestrial environments, due to the application of sewage sludge as fertilizer on farm land [128]. TCS persists in the environment but, during the wastewater treatment process, can also be converted into chlorinated derivatives, which are more toxic and more persistent than their parent compound [44]. Due to its stability and lipophilicity, TCS has a potential for bioaccumulation in algae [50], plants [129], and animals such as earthworms [130], marine mussels [51], snails [131], amphibian larvae [132], fish [133], and marine mammals [134]; moreover, methyl triclosan, an environmental transformation product of TCS, has been detected in fish from various lakes in Switzerland [135]. TCS present in the aquatic environment causes adverse ecological effects: it exhibits toxicity to algae species [136], alters benthic bacterial communities composition (fostering cyanobacteria over algae) [137], exhibits teratogenic responses and mortality in the embryos and larvae of zebrafish [138], and exhibits endocrine disruption in fish [139]. Epidemiological studies reported the presence of TCS in a wide range of concentrations in human body fluids such as urine, blood, and breast milk [44]. According to some researchers, the presence of TCS in human fluids is mostly correlated with the use of TCS-containing personal care products [140]; other authors believe that oral absorption can be significant, considering that TCS has been detected in drinking water [141,142]. Exposure to TCS can produce negative effects on human health, including thyroid function impairment, endocrine disruption, oxidative stress, and liver carcinogenesis ([44] and the literature therein). All data available on the environmental presence of TCS and its by-products and its health implications suggest that this compound should be considered a priority pollutant [143].

5. Plastic Microbeads

Plastic pollution of the marine environment represents a growing environmental concern, addressed at both macroplastic and microplastic debris; microplastic is classified as being material <5 mm, according to a scheme proposed by Moore [144]. The majority of small plastic particles in the seas come from the fragmentation of larger items; however, there is growing evidence that microplastic also enters the marine environment directly from a variety of sources, including cosmetics, clothing, and industrial processes. Plastic microbeads are present as abrasive scrubbers in a number of personal care and cosmetic products such as hand-cleansers, toothpaste, face scrubs, bubblebath, shampoos, and soaps [145,146], where they replace natural exfoliating materials previously utilized (pumice, oatmeal, apricot husks). In addition to the function of dead skin removal and deep cleansing, plastic microbeads can also play a decorative role in personal care products. Around 93% of the microbeads used in personal care products are made of polyethylene [147], but sometimes the material that they are made from is polypropylene, nylon, polyethylene terephthalate, or polymethyl methacrylate [148]. The microplastics present in cosmetics are too small to be trapped in sewage treatment plants, so they enter waterways via domestic drainage systems and are transported to seas and oceans [52,149], although some authors believe that the current treatment processes are effective for the removal of microplastic contaminants in wastewater treatment plants [53].

Although there is a large amount of literature that documents the presence of microplastics in the marine environment, organisms, and sediments [54,55,146,150], not as many works have been made to quantify the emission of microplastics from personal care products in different geographical areas. Gouin et al. [151] estimated that the per capita consumption of microplastics used in personal care products for the U.S. population is approximately 2.4 mg·person⁻¹·day⁻¹, so that the U.S. population

is emitting 263 tons year⁻¹ of polyethylene microplastic. Quantification of cosmetic sources of plastic microbeads was attempted by Gouin et al. [147] on the basis of the results of a survey conducted in the European Union countries Norway and Switzerland by Cosmetics Europe, the European personal care association that represents more than 4000 personal care product manufacturers. All members of Cosmetics Europe were requested to complete a survey on the use of plastic microbeads in their cosmetic products for the year 2012. The survey results reported a total amount of 4360 tons of plastic microbeads used across the European Union market, including Norway and Switzerland. Polyethylene microbeads were reported to be the principal type of plastic material used as a scrubber in detergents (4073 tons), while the remaining 287 tons were made up of miscellaneous polymers. Some authors characterised the microbeads present in cosmetic products with respect to their shapes and size range [145,148]. In their recent study [148] didn't just characterize the plastic microbeads contained in six brands of facial scrubs present in supermarkets in the United Kingdom, but also investigated their capacity to accumulate and transport chemicals by using a mixture of phenanthrene (Phe) and dichlorodiphenyltrichloroetane (DDT) in seawater. The numbers of total particles in each product package varied greatly, ranging between 18,906 and 919 microparticles/mL; they also showed a wide size range (mean diameters between 327 and 164 μ m). The authors of this study estimated that the quantity of microbeads released in a single use of these cosmetics could range between 4594 and 94,500. The results of adsorption experiments conducted with mixtures of ³H-Phe and ¹⁴C-DDT in seawater showed that PE microparticles extracted from facial scrubs were able to hold these contaminants onto their surface, with sorption capacity in all cases being significantly higher for DDT; "rough" microbeads were more efficient than "smooth" ones, probably due to the increased surface area. Other studies confirmed that microplastics can act as a transport medium for chemical pollutants in the marine environment [152,153].

Once in the environment, microbeads made of high-density plastics (polyester, polyvinyl chloride) settle out of the water column and accumulate in the sediment, while low-density microplastics (polyethylene, polystyrene) float on the sea surface [150]. Since there is no way of effectively removing microplastic contaminants from the marine environment, and they are highly resistant to degradation, microparticles can be ingested by aquatic organisms. Ingestion and accumulation of microplastics has been well documented in different marine organisms, including copepods [146], bivalves [52,54,154,155], fish [56], and seabirds [57].

Microplastics negatively impact upon marine biota. Many marine organisms may adsorb microplastics on their surfaces and may ingest small plastic fragments, mistaking them for their natural food source [156]; when exposed to polystyrene microparticles, some organisms, like copepods, are able to ingest them via filter-feeding and then egest them within hours in faecal pellets [157]. However, organisms can also be at risk from starvation or intestinal blockage, impairment of reproduction and feeding ability, and reduced growth rates consequent to microplastic ingestion [149,158]. A recent study [159] provided evidence that microplastics induce feeding modification and reproductive disruption in oysters; oyster experimentally exposed to polystyrene microspheres for two months during a reproductive cycle showed, in particular, significant decrease in oocyte number, diameter, and sperm velocity. Moreover, there is the potential for the microplastics to accumulate at higher levels of the food chain [156,160]. Another concern is the potential of microplastics to aid the delivery (after ingestion) of POPs (persistent organic pollutants); for example, polychlorinated biphenyls, polycyclic aromatic hydrocarbons, organochlorine pesticides, and alkylphenols, added during plastics manufacture or, more often, adsorbed and concentrated from the surrounding seawater [153,161,162], which can be transferred along the food chain and can harm fish and other aquatic organisms.

Considering the increasing concern expressed by the scientific community over microplastic pollution of the marine environment, several European countries (Belgium, Netherlands, Austria, and Sweden) have issued a joint call to outlaw microplastics in cosmetic formulations [163]; Cosmetics Europe [164] has recommended its membership to discontinue, in wash-off cosmetic products placed on the market as of 2020, the use of synthetic, solid, plastic particles used for exfoliating and cleansing

that are non-biodegradable in the marine environment. In December 2015, the U.S. President signed a bill that will phase out the manufacturing of cosmetics containing plastic microbeads by 1 July 2017 and their sale by 1 July 2019 [165]. In recent times, the major cosmetic companies have made commitments to phase out the use of microbeads in their products and are promoting natural alternatives such as pumice, walnut and coconut shells, ground palm nut, bamboo powder, apricot kernel shell, biodegradable hydrogenated oils waxes like jojoba beads, and diatomaceous earth.

6. Conclusions

The number of emerging contaminants released in the environment as a consequence of human activities is increasing day by day and reflects the growing consumption of a wide range of products, including cosmetics and personal care products. Chemical compounds that compose cosmetic formulations number in the several thousands, and the annual production and consumption of personal care products exceeds thousands of tons. The hazard of the continuous release of these huge amounts of chemicals into waters should not be underestimated. The environmental fate of these products is largely unknown, and, if in some cases they are removed in wastewater treatment plants, in other cases, they can escape conventional treatment processes, persist in the environment at unexpected levels, undergo bioaccumulation, and even react with other pollutants to originate new unpredictable contaminants.

Banning the products responsible for these problems is an impracticable option, except in particular circumstances (for instance, the use of sunscreen is banned in some marine ecoparks in Mexico [71]). Addressing this issue realistically requires different approaches and strategies. To some extent, our increased awareness of the pollution potential of these products is the result of the advanced technologies of analytical chemistry. Therefore, the development of improved extraction and analytical methods would allow a more comprehensive and accurate evaluation of environmental pollutants in complex matrices. On the other hand, further studies on the acute and chronic toxicity of these contaminants should be conducted to allow a more precise assessment of their actual ecological and health risk. Finally, information about the environmental impact of cosmetics on their packaging could encourage consumers to employ a more responsible and informed use of these products.

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References

- Regulation (EC) No 1223/2009 of the European Parliament and of the Council of 30 November 2009 on cosmetic products. Available online: http://eur-lex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L:2009: 342:0059:0209:en:PDF (accessed on 10 March 2017).
- 2. Brausch, J.M.; Rand, G.M. A review of personal care products in the aquatic environment: Environmental concentrations and toxicity. *Chemosphere* **2011**, *82*, 1518–1532. [CrossRef] [PubMed]
- Carballa, M.; Omil, F.; Lema, J.M.; Llompart, M.; García-Jares, C.; Rodríguez, I.; Gómez, M.; Ternes, T. Behavior of pharmaceuticals, cosmetics and hormones in a sewage treatment plant. *Water Res.* 2004, *38*, 2918–2926. [CrossRef] [PubMed]
- Liu, N.; Shi, Y.; Li, W.; Xu, L.; Cai, Y. Concentrations and distribution of synthetic musks and siloxanes in sewage sludge of wastewater treatment plants in China. *Sci. Total Environ.* 2014, 476–477, 65–72. [CrossRef] [PubMed]
- Campo, J.; Masiá, A.; Picó, Y.; Farré, M.; Barceló, D. Distribution and fate of perfluoroalkyl substances in Mediterranean Spanish sewage treatment plants. *Sci. Total Environ.* 2014, 472, 912–922. [CrossRef] [PubMed]
- Ramos, S.; Homem, V.; Alves, A.; Santos, L. A review of organic UV-filters in wastewater treatment plants. *Environ. Int.* 2016, *86*, 24–44. [CrossRef] [PubMed]

- Browne, M.A.; Galloway, T.; Thompson, R. Microplastics—An emerging contaminant of potential concern? *Integr. Environ. Assess. Manag.* 2009, *3*, 559–561. [CrossRef]
- 8. Díaz-Cruz, M.S.; García-Galán, M.J.; Guerra, P.; Jelic, A.; Postigo, C.; Eljarrat, E.; Farré, M.; López de Alda, M.J.; Petrovic, M.; Barceló, D. Analysis of selected emerging contaminants in sewage sludge. *Trends Anal. Chem.* **2009**, *28*, 1263–1275. [CrossRef]
- 9. Ternes, T.A.; Joss, A.; Siegrist, H. Scrutinizing pharmaceuticals and personal care products in wastewater treatment. *Environ. Sci. Technol.* **2004**, *38*, 392A–399A. [CrossRef] [PubMed]
- 10. Jardak, K.; Drogui, P.; Daghrir, R. Surfactants in aquatic and terrestrial environment: Occurrence, behavior, and treatment processes. Environ. *Sci. Pollut. Res.* **2016**, *23*, 3195–3216. [CrossRef] [PubMed]
- 11. Serpone, N.; Dondi, D.; Albini, A. Inorganic and organic UV filters: their role and efficacy in sunscreens and suncare products. *Inorganica Chim. Acta* 2007, *360*, 794–802. [CrossRef]
- 12. Volkova, K.; Bilanicova, D.; Bartonova, A.; Letašová, S.; Dusinska, M. Association between environmental factors and incidence of cutaneous melanoma. Review. *Environ. Health* **2012**, *11*, 1–13. [CrossRef] [PubMed]
- Chisvert, A.; Salvador, A. UV filters in sunscreen and other cosmetics. Regulatory aspects and analytical methods. In *Analysis of Cosmetic Products*; Chisvert, A., Salvador, A., Eds.; Elsevier: Amsterdam, The Netherlands, 2007; pp. 83–120.
- 14. Giokas, D.L.; Salvador, A.; Chisvert, A. UV filters: From sunscreens to human body and the environment. *Trends Anal. Chem.* **2007**, *26*, 360–374. [CrossRef]
- 15. Tsui, M.M.P.; Leung, H.W.; Wai, T.-C.; Yamashita, N.; Taniyasu, S.; Liu, W.; Lam, P.K.S.; Murphy, M.B. Occurrence, distribution and ecological risk assessment of multiple classes of UV filters in surface waters from different countries. *Water Res.* **2014**, *67*, 55–65. [CrossRef] [PubMed]
- 16. Jiang, J.-J.; Lee, C.-L.; Fang, M.-D. Emerging organic contaminants in coastal waters: anthropogenic impact, environmental release and ecological risk. *Mar. Pollut. Bull.* **2014**, *85*, 391–399. [CrossRef] [PubMed]
- 17. Sánchez Rodríguez, A.; Rodrigo Sanz, M.; Betancort Rodríguez, J.R. Occurrence of eight UV filters in beaches of Gran Canaria (Canary Islands). An approach to environmental risk assessment. *Chemosphere* **2015**, *131*, 85–90. [CrossRef] [PubMed]
- 18. Ekpeghere, K.I.; Kim, U.-J.; O, S.-H.; Kim, H.-Y.; Oh, J.-E. Distribution and seasonal occurrence of UV filters in rivers and wastewater treatment plants in Korea. *Sci. Total Environ.* **2016**, *542*, 121–128. [CrossRef] [PubMed]
- Poiger, T.; Buser, H.R.; Balmer, M.E.; Bergqvist, P.A.; Müller, M.D. Occurrence of UV filter compounds from sunscreens in surface waters: regional mass balance in two Swiss lakes. *Chemosphere* 2004, 55, 951–963. [CrossRef] [PubMed]
- 20. Balmer, M.E.; Buser, H.R.; Muller, M.D.; Poiger, T. Occurrence of some organic UV filters in wastewater, in surface waters, and in fish from Swiss lakes. *Environ. Sci. Technol.* 2005, *39*, 953–962. [CrossRef] [PubMed]
- 21. Amine, H.; Gomez, E.; Halwani, J.; Casella, C.; Fenet, H. UV filters, ethylhexyl methoxycinnamate, octocrylene and ethylhexyl dimethyl PABA from untreated wastewater in sediment from Eastern Mediterranean river transition and coastal zones. *Mar. Pollut. Bull.* **2012**, *64*, 2435–2442. [CrossRef] [PubMed]
- 22. Barón, E.; Gago-Ferrero, P.; Gorga, M.; Rudolph, I.; Mendoza, G.; Zapata, A.M.; Díaz-Cruz, S.; Barra, R.; Ocampo-Duque, W.; Páez, M.; et al. Occurrence of hydrophobic organic pollutants (BFRs and UV-filters) in sediments from South America. *Chemosphere* **2013**, *92*, 309–316. [CrossRef] [PubMed]
- Kameda, Y.; Kimura, K.; Miyazak, M. Occurrence and profiles of organic sun-blocking agents in surface waters and sediments in Japanese rivers and lakes. *Environ. Pollut.* 2011, 159, 1570–1576. [CrossRef] [PubMed]
- 24. Langford, K.H.; Reid, M.J.; Fjeld, E.; Øxnevad, S.; Thomas, K.V. Environmental occurrence and risk of organic UV filters and stabilizers in multiple matrices in Norway. *Environ. Int.* **2015**, *80*, 1–7. [CrossRef] [PubMed]
- 25. Sánchez-Quiles, D.; Tovar-Sánchez, A. Are sunscreens a new environmental risk associated with coastal tourism? *Environ. Int.* **2015**, *83*, 158–170. [CrossRef] [PubMed]
- 26. Gago-Ferrero, P.; Alonso, M.B.; Bertozzi, C.P.; Marigo, J.; Barbosa, L.; Cremer, M.; Secchi, E.R.; Azevedo, A.; Lailson-Brito, J., Jr.; Torres, J.P.M.; et al. First determination of UV filters in marine mammals. Octocrylene levels in Franciscana dolphins. *Environ. Sci. Technol.* **2013**, *47*, 5619–5625. [CrossRef] [PubMed]
- 27. Bachelot, M.; Munaron, D.; Le Gall, P.; Casellas, C.; Fenet, H.; Gomez, E. Organic UV filter concentrations in marine mussels from French coastal regions. *Sci. Tot. Environ.* **2012**, *420*, 273–279. [CrossRef] [PubMed]

- 28. Fent, K.; Zenker, A.; Rapp, M. Widespread occurrence of estrogenic UV-filters in aquatic ecosystems in Switzerland. *Environ. Pollut.* **2010**, *158*, 1817–1824. [CrossRef] [PubMed]
- Gago-Ferrero, P.; Díaz-Cruz, M.S.; Barceló, D. UV filters bioaccumulation in fish from Iberian river basins. *Sci. Total Environ.* 2015, 518–519, 518–525. [CrossRef] [PubMed]
- Shi, X.; Li, Z.; Chen, W.; Qiang, L.; Xia, J.; Chen, M.; Zhu, L.; Alvarez, P.J.J. Fate of TiO₂ nanoparticles entering sewage treatment plants and bioaccumulation in fish in the receiving streams. *NanoImpact* 2016, 3–4, 96–103. [CrossRef]
- 31. Sánchez-Quiles, D.; Tovar-Sánchez, A. Sunscreens as a source of hydrogen peroxide production in coastal waters. *Environ. Sci. Technol.* **2014**, *48*, 9037–9042. [CrossRef] [PubMed]
- 32. Aziza, I.H.H.; Ingrid, B.; Khémary, U.; Aghleb, B.; Gonzalez, C. Estrogenic activity and parabens occurrence in three Tunisian wastewater treatment plants. *Eur. J. Water Qual.* **2011**, *42*, 91–103. [CrossRef]
- 33. Peng, X.; Yu, Y.; Tang, C.; Tan, J.; Huang, Q.; Wang, Z. Occurrence of steroid estrogens, endocrine-disrupting phenols, and acid pharmaceutical residues in urban riverine water of the Pearl River Delta, South China. *Sci. Total Environ.* **2008**, *397*, 158–166. [CrossRef] [PubMed]
- Kasprzyk-Hordern, B.; Dinsdale, R.M.; Guwy, A.J. The occurrence of pharmaceuticals, personal care products, endocrine disruptors and illicit drugs in surface water in South Wales, UK. *Water Res.* 2008, 42, 3498–3518. [CrossRef] [PubMed]
- 35. Yamamoto, H.; Tamura, I.; Hirata, Y.; Kato, J.; Kagota, K.; Katsuki, S.; Yamamoto, A.; Kagami, Y.; Tatarazako, N. Aquatic toxicity and ecological risk assessment of seven parabens: Individual and additive approach. *Sci. Total Environ.* **2011**, 410–411, 102–111. [CrossRef] [PubMed]
- Kimura, K.; Kameda, Y.; Yamamoto, H.; Nakada, N.; Tamura, I.; Miyazaki, M.; Masunaga, S. Occurrence of preservatives and antimicrobials in Japanese rivers. *Chemosphere* 2014, 107, 393–399. [CrossRef] [PubMed]
- 37. Kim, J.-W.; Ramaswamy, B.R.; Chang, K.-H.; Isobe, T.; Tanabe, S. Multiresidue analytical method for the determination of antimicrobials, preservatives, benzotriazole UV stabilizers, flame retardants and plasticizers in fish using ultra high performance liquid chromatography coupled with tandem mass spectrometry. *J. Chromatogr. A* 2011, 1218, 3511–3520. [CrossRef] [PubMed]
- Ramaswamy, B.R.; Kim, J.-W.; Isobe, T.; Chang, K.-H.; Amano, A.; Miller, T.W.; Siringan, F.P.; Tanabe, S. Determination of preservative and antimicrobial compounds in fish from Manila Bay, Philippines using ultra high performance liquid chromatography tandem mass spectrometry, and assessment of human dietary exposure. J. Hazard. Mater. 2011, 192, 1739–1745. [CrossRef] [PubMed]
- 39. Xue, J.; Kannan, K. Accumulation profiles of parabens and their metabolites in fish, black bear, and birds, including bald eagles and albatrosses. *Environ. Int.* **2016**, *94*, 546–553. [CrossRef] [PubMed]
- Xue, J.; Sasaki, N.; Elangovan, M.; Diamond, G.; Kannan, K. Elevated accumulation of parabens and their metabolites in marine mammals from the United States coastal waters. *Environ. Sci. Technol.* 2015, 49, 12071–12079. [CrossRef] [PubMed]
- 41. Carmona, E.; Andreu, V.; Picó, Y. Occurrence of acidic pharmaceuticals and personal care products in Turia River Basin: from waste to drinking water. *Sci. Total Environ.* **2014**, *484*, 53–63. [CrossRef] [PubMed]
- 42. Gasperi, J.; Geara, D.; Lorgeoux, C.; Bressy, A.; Zedek, S.; Rocher, V.; El Samrani, A.; Chebbo, G.; Moilleron, R. First assessment of triclosan, triclocarban and paraben mass loads at a very large regional scale: Case of Paris conurbation (France). *Sci. Total Environ.* **2014**, *493*, 854–861. [CrossRef] [PubMed]
- 43. Chen, Z.-F.; Ying, G.-G.; Liu, Y.-S.; Zhang, Q.-Q.; Zhao, J.-L.; Liu, S.-S.; Chen, J.; Peng, F.-J.; Lai, H.-J.; Pan, C.-G. Triclosan as a surrogate for household biocides: An investigation into biocides in aquatic environments of a highly urbanized region. *Water Res.* 2014, *58*, 269–279. [CrossRef] [PubMed]
- 44. Yueh, M.-F.; Tukey, R.H. Triclosan: a widespread environmental toxicant with many biological effects. *Annu. Rev. Pharmacol. Toxicol.* **2016**, *56*, 251–272. [CrossRef] [PubMed]
- 45. Roberts, J.; Kumar, A.; Du, J.; Hepplewhite, C.; Ellis, D.J.; Christy, A.G.; Beavis, S.G. Pharmaceuticals and personal care products (PPCPs) in Australia's largest inland sewage treatment plant, and its contribution to a major Australian river during high and low flow. *Sci. Total Environ.* **2016**, *541*, 1625–1637. [CrossRef] [PubMed]
- Kolpin, D.W.; Skopec, M.; Meyer, M.T.; Furlong, E.T.; Zaugg, S.D. Urban contamination of pharmaceuticals and other organic wastewater contaminants to streams during differing flow conditions. *Sci. Total Environ.* 2004, 328, 119–130. [CrossRef] [PubMed]

- Benotti, M.J.; Trenholm, R.A.; Vanderford, B.J.; Holady, J.C.; Stanford, B.D.; Snyder, S.A. Pharmaceuticals and endocrine disrupting compounds in US drinking water. *Environ. Sci. Technol.* 2009, 43, 597–603. [CrossRef] [PubMed]
- 48. Dougherty, J.A.; Swarzenski, P.W.; Pinicola, R.S.; Reinhard, M. Occurrence of herbicides and pharmaceuticals and personal care products in surface water and groundwater around Liberty Bay, Puget Sound, Washington. *J. Environ. Qual.* **2010**, *39*, 1173–1180. [CrossRef] [PubMed]
- 49. Kim, J.W.; Jang, H.-S.; Kim, J.-G.; Ishibashi, H.; Hirano, M.; Nasu, K.; Ichikawa, N.; Takao, Y.; Shinohara, R.; Arizono, K. Occurrence of pharmaceuticals and personal care products (PPCPs) in surface water from Mankyung River, South Korea. *J. Health Sci.* **2009**, *55*, 249–258. [CrossRef]
- 50. Coogan, M.A.; Edziyie, R.E.; La Point, T.W.; Venables, B.J. Algal bioaccumulation of triclocarban, triclosan, and methyl-triclosan in a North Texas wastewater treatment plant receiving stream. *Chemosphere* **2007**, *67*, 1911–1918. [CrossRef] [PubMed]
- 51. Kookana, R.S.; Shareef, A.; Fernandes, M.B.; Hoare, S.; Gaylard, S.; Kumar, A. Bioconcentration of triclosan and methyl-triclosan in marine mussels (Mytilus galloprovincialis) under laboratory conditions and in metropolitan waters of Gulf St Vincent, South Australia. *Mar. Pollut. Bull.* 2013, 15, 66–72. [CrossRef] [PubMed]
- Browne, M.A.; Dissanayake, A.; Galloway, T.S.; Lowe, D.M. Ingested microscopic plastic translocates to the circulatory system of the mussel, Mytilus edulis. *Environ. Sci. Technol.* 2008, 42, 5026–5031. [CrossRef] [PubMed]
- 53. Carr, S.A.; Liu, J.; Tesoro, A.G. Transport and fate of microplastic particles in wastewater treatment plants. *Water Res.* **2016**, *91*, 174–182. [CrossRef] [PubMed]
- 54. Vandermeersch, G.; Van Cauwenberghe, L.; Janssen, C.R.; Marques, A.; Granby, K.; Fait, G.; Kotterman, M.J.J.; Diogène, J.; Bekaert, K.; Robbens, J.; Devriese, L. A critical view on microplastic quantification in aquatic organisms. *Environ. Res.* **2015**, *143*, 46–55. [CrossRef] [PubMed]
- 55. Isobe, A.; Uchiyama-Matsumoto, K.; Uchida, K.; Tokai, T. Microplastics in the Southern Ocean. *Mar. Pollut. Bull.* **2017**, *14*, 623–626. [CrossRef] [PubMed]
- 56. Davison, P.; Asch, R.G. Plastic ingestion by mesopelagic fishes in the North Pacific Subtropical Gyre. *Mar. Ecol. Prog. Ser.* **2011**, 432, 173–180. [CrossRef]
- 57. Van Franeker, J.A.; Blaize, C.; Danielsen, J.; Fairclough, K.; Gollan, J.; Guse, N.; Hansen, P.-L.; Heubeck, M.; Jensen, J.-K.; Le Guillou, G.; et al. Monitoring plastic ingestion by the northern fulmar Fulmarus glacialis in the North Sea. *Environ. Pollut.* **2011**, *159*, 2609–2615. [CrossRef] [PubMed]
- Zhang, P.-P.; Shi, Z.-G.; Yu, Q.-W.; Feng, Y.-Q. A new device for magnetic stirring-assisted dispersive liquid–liquid microextraction of UV filters in environmental water samples. *Talanta* 2011, *83*, 1711–1715. [CrossRef] [PubMed]
- Jurado, A.; Gago-Ferrero, P.; Vàzquez-Suñé, E.; Carrera, J.; Pujades, E.; Díaz-Cruz, M.S.; Barceló, D. Urban groundwater contamination by residues of UV filters. *J. Hazard. Mater.* 2014, 271, 141–149. [CrossRef] [PubMed]
- 60. Tsui, M.M.P.; Leung, H.W.; Kwan, B.K.Y.; Ng, K.-Y.; Yamashita, N.; Taniyasu, S.; Lam, P.K.S.; Murphy, M.B. Occurrence, distribution and ecological risk assessment of multiple classes of UV filters in marine sediments in Hong Kong and Japan. *J. Hazard. Mater.* **2015**, *292*, 180–187. [CrossRef] [PubMed]
- 61. Gago-Ferrero, P.; Díaz-Cruz, M.S.; Barceló, D. Fast pressurized liquid extraction with in-cell purification analysis by liquid chromatography tandem mass spectrometry for the determination of UV filters and their degradation products in sediments. *Anal. Bioanal. Chem.* **2011**, *400*, 2195–2204. [CrossRef] [PubMed]
- 62. Kaiser, D.; Wappelhorst, O.; Oekten, M.; Oehlmann, M. Occurrence of widely used organic UV filters in lake and river sediments. *Environ. Chem.* **2012**, *9*, 139–147. [CrossRef]
- 63. Kim, S.; Choi, K. Occurrences, toxicities, and ecological risks of benzophenone-3, a common component of organic sunscreen products: a mini-review. *Environ. Int.* **2014**, *70*, 143–157. [CrossRef] [PubMed]
- 64. Oie, C.S.I.; Albaugh, C.E.; Peyton, B.M. Benzoate and salicylate degradation by Halomonas campisalis, an alkaliphilic and moderately halophilic microorganism. *Water Res.* **2007**, *41*, 1235–1242. [CrossRef] [PubMed]
- 65. Teo, T.L.L.; Coleman, H.M.; Khan, S.J. Chemical contaminants in swimming pools: Occurrence, implications and control. *Environ. Int.* **2015**, *76*, 16–31. [CrossRef] [PubMed]
- Zwiener, C.; Richardson, S.D.; DeMarini, D.; Frimmel, F.H. Drowning in disinfection byproducts? Assessing swimming pool water. *Environ. Sci. Technol.* 2007, 41, 363–372. [CrossRef] [PubMed]

- 67. Díaz-Cruz, M.S.; Barceló, D. Chemical analysis and ecotoxicological effects of organic UV-absorbing compounds in aquatic ecosystems. *Trends Anal. Chem.* **2009**, *28*, 708–717. [CrossRef]
- 68. Lakind, J.S.; Richardson, S.D.; Blount, B.C. The good, the bad, and the volatile: can we have both healthy pools and healthy people? *Environ. Sci. Technol.* **2010**, *44*, 3205–3210. [CrossRef] [PubMed]
- 69. Kockler, J.; Oelgemöller, M.; Robertson, S.; Glass, B.D. Photostability of sunscreens. *J. Photoch. Photobio. C* **2012**, *13*, 91–110. [CrossRef]
- 70. Santos, A.J.M.; Miranda, M.S.; Esteves da Silva, J.C.G. The degradation products of UV filters in aqueous and chlorinated aqueous solutions. *Water Res.* **2012**, *46*, 3167–3176. [CrossRef] [PubMed]
- Danovaro, R.; Bongiorni, L.; Corinaldesi, C.; Giovannelli, D.; Damiani, E.; Astolfi, P.; Greci, L.; Pusceddu, A. Sunscreens cause coral bleaching by promoting viral infections. *Environ. Health Perspect.* 2008, 116, 441–447. [CrossRef] [PubMed]
- 72. Kunz, P.Y.; Galicia, H.F.; Fent, K. Comparison of in vitro and in vivo estrogenic activity of UV filters in fish. *Toxicol. Sci.* **2006**, *90*, 349–361. [CrossRef] [PubMed]
- 73. Faass, O.; Schlumpf, M.; Reolon, S.; Henseler, M.; Maerkel, K.; Durrer, S.; Lichtensteiger, W. Female sexual behavior, estrous cycle and gene expression in sexually dimorphic brain regions after pre- and postnatal exposure to endocrine active UV filter. *Neurotoxicology* **2009**, *30*, 249–260. [CrossRef] [PubMed]
- 74. Schlumpf, M.; Kypke, K.; Vökt, C.C.; Birchler, M.; Durrer, S.; Faass, O.; Ehnes, C.; Fuetsch, M.; Gaille, C.; Henseler, M.; et al. Endocrine active UV filters: developmental toxicity and exposure through breast milk. *Chimia* **2008**, *62*, 345–351. [CrossRef]
- 75. Commission Regulation (EU) 2016/1143 of 13 July 2016 amending Annex VI to Regulation (EC) No 1223/2009 of the European Parliament and of the Council on cosmetic products. Available online: http://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32016R1143&from=EN (accessed on 10 March 2017).
- 76. Commission Regulation (EU) 2016/621 of 21 April 2016, amending Annex VI to Regulation (EC) No 1223/2009 of the European Parliament and of the Council on cosmetic products. Available online: http://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32016R0621&from=EN (accessed on 10 March 2017).
- 77. Botta, C.; Labille, J.; Auffan, M.; Borschneck, D.; Miche, H.; Cabié, M.; Masion, A.; Rose, J.; Bottero, J.-Y. TiO₂-based nanoparticles released in water from commercialized sunscreens in a life-cycle perspective: Structures and quantities. *Environ. Pollut.* **2011**, *159*, 1543–1550. [CrossRef] [PubMed]
- 78. Lewicka, Z.A.; Yu, W.W.; Oliva, B.L.; Contreras, E.Q.; Colvin, V.L. Photochemical behavior of nanoscale TiO₂ and ZnO sunscreen ingredients. *J. Photoch. Photobio. A* **2013**, 263, 24–33. [CrossRef]
- Buchalska, M.; Kras, G.; Oszajca, M.; Łasocha, W.; Macyk, W. Singlet oxygen generation in the presence of titanium dioxide materials used as sunscreens in suntan lotions. *J. Photoch. Photobio. A* 2010, 213, 158–163. [CrossRef]
- 80. Barker, P.J.; Branch, A. The interaction of modern sunscreen formulations with surface coatings. *Prog. Org. Coat.* **2008**, *62*, 313–320. [CrossRef]
- Auffan, M.; Pedeutour, M.; Rose, J.; Masion, A.; Ziarelli, F.; Borschneck, D.; Chaneac, C.; Botta, C.; Chaurand, P.; Labille, J.; et al. Structural degradation at the surface of a TiO₂-based nanomaterial used in cosmetics. *Environ. Sci. Technol.* 2010, 44, 2689–2694. [CrossRef] [PubMed]
- Menard, A.; Drobne, D.; Jemec, A. Ecotoxicity of nanosized TiO₂. Review of in vivo data. *Environ. Pollut.* 2011, 159, 677–684. [CrossRef] [PubMed]
- Xiong, D.; Fang, T.; Yu, L.; Sima, X.; Zhu, W. Effects of nano-scale TiO₂, ZnO and their bulk counterparts on zebrafish: Acute toxicity, oxidative stress and oxidative damage. *Sci. Total Environ.* 2011, 409, 1444–1452. [CrossRef] [PubMed]
- 84. Blaise, C.; Gagne, F.; Ferard, J.F.; Eullaffoy, P. Ecotoxicity of selected nano-materials to aquatic organisms. *Environ. Toxicol.* **2008**, *23*, 591–598. [CrossRef] [PubMed]
- Ma, H.; Brennan, A.; Diamond, S.A. Phototoxicity of TiO₂ nanoparticles under solar radiation to two aquatic species: Daphnia magna and Japanese medaka. *Environ. Toxicol. Chem.* 2012, *31*, 1621–1629. [CrossRef] [PubMed]
- Clemente, Z.; Castro, V.L.; Feitosa, L.O.; Lima, R.; Jonsson, C.M.; Maia, A.H.; Fraceto, L.F. Fish exposure to nano-TiO₂ under different experimental conditions: Methodological aspects for nanoecotoxicology investigations. *Sci. Total Environ.* 2013, 463–464, 647–656. [CrossRef] [PubMed]

- 87. Hund-Rinke, K.; Simon, M. Ecotoxic effect of photocatalytic active nanoparticles (TiO₂) on algae and daphnids. Environ. *Sci. Pollut. Res.* **2006**, *13*, 225–232. [CrossRef]
- Zhang, J.; Wages, M.; Cox, S.B.; Maul, J.D.; Li, Y.; Barnes, M.; Hope-Weeks, L.; Cobb, G.P. Effect of titanium dioxide nanomaterials and ultraviolet light coexposure on African clawed frogs (Xenopus laevis). *Environ. Toxicol. Chem.* 2012, *31*, 176–183. [CrossRef] [PubMed]
- Clemente, Z.; Castro, V.L.; Moura, M.A.M.; Jonsson, C.M.; Fraceto, L.F. Toxicity assessment of TiO₂ nanoparticles in zebrafish embryos under different exposure conditions. *Aquat. Toxicol.* 2014, 147, 129–139. [CrossRef] [PubMed]
- Zhao, X.; Wang, S.; Wu, Y.; You, H.; Lv, L. Acute ZnO nanoparticles exposure induces developmental toxicity, oxidative stress and DNA damage in embryo-larval zebrafish. *Aquat. Toxicol.* 2013, 136–137, 49–59. [CrossRef] [PubMed]
- 91. Chen, T.-H.; Lin, C.-C.; Meng, P.-J. Zinc oxide nanoparticles alter hatching and larval locomotor activity in zebrafish (Danio rerio). *J. Hazard. Mater.* **2014**, 277, 134–140. [CrossRef] [PubMed]
- Suman, T.Y.; Radhika Rajasree, S.R.; Kirubagaran, R. Evaluation of zinc oxide nanoparticles toxicity on marine algae Chlorella vulgaris through flow cytometric, cytotoxicity and oxidative stress analysis. *Ecotoxicol. Environ. Saf.* 2015, 113, 23–30. [CrossRef] [PubMed]
- 93. Zhang, C.; Wang, J.; Tan, L.; Chen, X. Toxic effects of nano-ZnO on marine microalgae Skeletonema costatum: Attention to the accumulation of intracellular Zn. *Aquat. Toxicol.* **2016**, *178*, 158–164. [CrossRef] [PubMed]
- Manzo, S.; Miglietta, M.L.; Rametta, G.; Buono, S.; Di Francia, G. Embryotoxicity and spermiotoxicity of nanosized ZnO for Mediterranean sea urchin Paracentrotus lividus. *J. Hazard. Mater.* 2013, 254–255, 1–9. [CrossRef] [PubMed]
- 95. Kahru, A.; Dubourguier, H.-C. From ecotoxicology to nanoecotoxicology. *Toxicology* **2010**, *269*, 105–119. [CrossRef] [PubMed]
- Ma, H.; Kabengi, N.J.; Bertsch, P.M.; Unrine, J.M.; Glenn, T.C.; Williams, P.L. Comparative phototoxicity of nanoparticulate and bulk ZnO to a free-living nematode Caenorhabditis elegans: The importance of illumination mode and primary particle size. *Environ. Pollut.* 2011, 159, 1473–1480. [CrossRef] [PubMed]
- 97. Zhu, X.; Wang, J.; Zhang, X.; Chang, Y.; Chen, Y. The impact of ZnO nanoparticle aggregates on the embryonic development of zebrafish (Danio rerio). *Nanotechnology* **2009**, *20*, 195103. [CrossRef] [PubMed]
- Jeon, S.; Kim, E.; Lee, J.; Lee, S. Potential risks of TiO2 and ZnO nanoparticles released from sunscreens into outdoor swimming pools. J. Hazard. Mater. 2016, 317, 312–318. [CrossRef] [PubMed]
- 99. Błędzka, D.; Gromadzińska, J.; Wąsowicz, W. Parabens. From environmental studies to human health. *Environ. Int.* **2014**, 67, 27–42. [CrossRef] [PubMed]
- 100. Baardseth, P.; Russwurm, H., Jr. Content of some organic acids in cloudberry (Rubus chamaemorus L.). *Food Chem.* **1978**, *3*, 43–46. [CrossRef]
- Peng, X.; Adachi, K.; Chen, C.; Kasai, H.; Kanoh, K.; Shizuri, Y.; Misawa, N. Discovery of a marine bacterium producing 4-hydroxybenzoate and its alkyl esters, parabens. *Appl. Environ. Microbiol.* 2006, 72, 5556–5561. [CrossRef] [PubMed]
- 102. Regulation (EU) No 358/2014 amending Annexes II and V to Regulation (EC) No 1223/2009 of the European Parliament and of the Council of 30 November 2009 on cosmetic products. Available online: http://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32014R0358&from=EN (accessed on 10 March 2017).
- 103. Regulation (EU) No 1004/2014 amending Annex V to Regulation (EC) No 1223/2009 of the European Parliament and of the Council of 30 November 2009 on cosmetic products. Available online: http://eur-lex. europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32014R1004&from=EN (accessed on 10 March 2017).
- 104. Haman, C.; Dauchy, X.; Rosin, C.; Munoz, J.-F. Occurrence, fate and behavior of parabens in aquatic environments: A review. *Water Res.* 2015, *68*, 1–11. [CrossRef] [PubMed]
- 105. González-Mariño, I.; Quintana, J.B.; Rodríguez, I.; Cela, R. Evaluation of the occurrence and biodegradation of parabens and halogenated by-products in wastewater by accurate mass liquid chromatography-quadrupole-time-of-flight-mass spectrometry (LC-QTOF-MS). *Water Res.* 2011, 45, 6770–6780. [CrossRef] [PubMed]
- 106. Canosa, P.; Rodríguez, I.; Rubí, E.; Negreira, N.; Cela, R. Formation of halogenated by-products of parabens in chlorinated water. *Anal. Chim. Acta* 2006, 575, 106–113. [CrossRef] [PubMed]

- 107. Smith, K.W.; Braun, J.M.; Williams, P.L.; Ehrlich, S.; Correia, K.F.; Calafat, A.M.; Ye, X.; Ford, J.; Keller, M.; Meeker, J.D.; et al. Predictors and variability of urinary paraben concentrations in men and women, including before and during pregnancy. *Environ. Health Perspect.* **2012**, *120*, 1538–1543. [CrossRef] [PubMed]
- 108. Philippat, C.; Wolff, M.S.; Calafat, A.M.; Ye, X.; Bausell, R.; Meadows, M.; Stone, J.; Slama, R.; Engel, S.M. Prenatal exposure to environmental phenols: concentrations in amniotic fluid and variability in urinary concentrations during pregnancy. *Environ. Health Perspect.* 2013, 121, 1225–1231. [CrossRef] [PubMed]
- 109. Hines, E.P.; Mendola, P.; von Ehrenstein, O.S.; Ye, X.; Calafat, A.M.; Fenton, S.E. Concentrations of environmental phenols and parabens in milk, urine and serum of lactating North Carolina women. *Reprod. Toxicol.* 2015, 54, 120–128. [CrossRef] [PubMed]
- 110. Moos, K.; Koch, H.M.; Angerer, J.; Apel, P.; Schröter-Kermani, C.; Brüning, T.; Kolossa-Gehring, M. Parabens in 24 h urine samples of the German Environmental Specimen Bank from 1995 to 2012. *Int. J. Hyg. Environ. Health* 2015, 218, 666–674. [CrossRef] [PubMed]
- 111. Wang, L.; Asimakopoulos, A.G.; Kannan, K. Accumulation of 19 environmental phenolic and xenobiotic heterocyclic aromatic compounds in human adipose tissue. *Environ. Int.* **2015**, *78*, 45–50. [CrossRef] [PubMed]
- 112. Valle-Sistac, J.; Molins-Delgado, D.; Díaz, M.; Ibáñez, L.; Barceló, D.; Díaz-Cruz, M.S. Determination of parabens and benzophenone-type UV filters in human placenta. First description of the existence of benzyl paraben and benzophenone-4. *Environ. Int.* 2016, *88*, 243–249. [CrossRef] [PubMed]
- Guo, J.; Wu, C.; Lu, D.; Jiang, S.; Liang, W.; Chang, X.; Xu, H.; Wang, G.; Zhou, Z. Urinary paraben concentrations and their associations with anthropometric measures of children aged 3 years. *Environ. Pollut.* 2017, 222, 307–314. [CrossRef] [PubMed]
- 114. Darbre, P.D.; Aljarrah, A.; Miller, W.R.; Coldham, N.G.; Sauer, M.J.; Pope, G.S. Concentrations of parabens in human breast tumours. *J. Appl. Toxicol.* **2004**, *24*, 5–13. [CrossRef] [PubMed]
- Barr, L.; Metaxas, G.; Harbach, C.A.J.; Savoy, L.A.; Darbre, P.D. Measurement of paraben concentrations in human breast tissue at serial locations across the breast from axilla to sternum. *J. Appl. Toxicol.* 2012, 32, 219–232. [CrossRef] [PubMed]
- 116. Darbre, P.D.; Harvey, P.W. Paraben esters: review of recent studies of endocrine toxicity, absorption, esterase and human exposure, and discussion of potential human health risks. J. Appl. Toxicol. 2008, 28, 561–568. [CrossRef] [PubMed]
- 117. Routledge, E.J.; Parker, J.; Odum, J.; Ashby, J.; Sumpter, J.P. Some alkyl hydroxy benzoate preservatives (parabens) are estrogenic. *Toxicol. Appl. Pharmacol.* **1998**, *153*, 12–19. [CrossRef] [PubMed]
- Darbre, P.D. Environmental oestrogens, cosmetics and breast cancer. *Best Pract. Res. Clin. Endocrinol. Metab.* 2006, 20, 121–143. [CrossRef] [PubMed]
- 119. National Cancer Institute Fact Sheets. Antiperspirants/Deodorants and Breast Cancer. 2008. Available online: http://www.cancer.gov/about-cancer/causes-prevention/risk/myths/antiperspirants-fact-sheet (accessed on 9 August 2016).
- 120. Witorsch, R.J.; Thomas, J.A. Personal care products and endocrine disruption: A critical review of the literature. *Crit. Rev. Toxicol.* **2010**, *3*, 1–30. [CrossRef] [PubMed]
- 121. Scientific Committee on Consumer Safety. Opinion on Parabens. 2011 SCCS/1348/10. Available online: https://ec.europa.eu/health/scientific_committees/consumer_safety/docs/sccs_o_069.pdf (accessed on 10 March 2017).
- 122. Nohynek, G.J.; Borgert, C.J.; Dietrich, D.; Rozman, K.K. Endocrine disruption: Fact or urban legend? *Toxicol. Lett.* **2013**, *223*, 295–305. [CrossRef] [PubMed]
- Pan, S.; Yuan, C.; Tagmount, A.; Rude, R.A.; Ackerman, J.M.; Yaswen, P.; Vulpe, C.D.; Leitman, D.C. Parabens and human epidermal growth factor receptor ligand cross-talk in breast cancer cells. *Environ. Health Perspect.* 2016, 124, 563–569. [CrossRef] [PubMed]
- 124. Sun, L.; Yu, T.; Guo, J.; Zhang, Z.; Hu, Y.; Xiao, X.; Sun, Y.; Xiao, H.; Li, J.; Zhu, D.; et al. The estrogenicity of methylparaben and ethylparaben at doses close to the acceptable daily intake in immature Sprague-Dawley rats. *Sci. Rep.* **2016**, *6*, 25173. [CrossRef] [PubMed]
- 125. Dann, A.B.; Hontela, A. Triclosan: environmental exposure, toxicity and mechanisms of action. *J. Appl. Toxicol.* **2011**, *31*, 285–311. [CrossRef] [PubMed]
- 126. Zhao, J.-L.; Ying, G.-G.; Liu, Y.-S.; Chen, F.; Yang, J.-F.; Wang, L. Occurrence and risks of triclosan and triclocarban in the Pearl River system, South China: from source to the receiving environment. *J. Hazard. Mater.* 2010, 179, 215–222. [CrossRef] [PubMed]

- 127. Liu, J.-L.; Wong, M.-H. Pharmaceuticals and personal care products (PPCPs): A review on environmental contamination in China. *Environ. Int.* 2013, *59*, 208–224. [CrossRef] [PubMed]
- 128. Chalew, T.E.; Halden, R.U. Environmental exposure of aquatic and terrestrial biota to triclosan and triclocarban. *J. Am. Water Works Assoc.* **2009**, *45*, 4–13. [CrossRef] [PubMed]
- Prosser, R.S.; Lissemore, L.; Topp, E.; Sibley, P.K. Bioaccumulation of triclosan and triclocarban in plants grown in soils amended with municipal dewatered biosolids. *Environ. Toxicol. Chem.* 2014, 33, 975–984. [CrossRef] [PubMed]
- Higgins, C.P.; Paesani, Z.J.; Chalew, T.E.; Halden, R.U.; Hundal, L.S. Persistence of triclocarban and triclosan in soils after land application of biosolids and bioaccumulation in Eisenia foetida. *Environ. Toxicol. Chem.* 2011, 30, 556–563. [CrossRef] [PubMed]
- Coogan, M.A.; La Point, T.W. Snail bioaccumulation of triclocarban, triclosan, and methyltriclosan in a North Texas, USA, stream affected by wastewater treatment plant runoff. Environ. *Toxicol. Chem.* 2008, 27, 1788–1793. [CrossRef] [PubMed]
- 132. Palenske, N.M.; Nallani, G.C.; Dzialowski, E.M. Physiological effects and bioconcentration of triclosan on amphibian larvae. *Comp. Biochem. Physiol. C* 2010, 152, 232–240. [CrossRef] [PubMed]
- Houtman, C.J.; Van Oostveen, A.M.; Brouwer, A.; Lamoree, M.H.; Legler, J. Identification of estrogenic compounds in fish bile using bioassay-directed fractionation. *Environ. Sci. Technol.* 2004, *38*, 6415–6423. [CrossRef] [PubMed]
- 134. Fair, P.A.; Lee, H.B.; Adams, J.; Darling, C.; Pacepavicius, G.; Alaee, M.; Bossart, G.D.; Henry, N.; Muir, D. Occurrence of triclosan in plasma of wild Atlantic bottlenose dolphins (Tursiops truncatus) and in their environment. *Environ. Pollut.* 2009, 157, 2248–2254. [CrossRef] [PubMed]
- 135. Balmer, M.E.; Poiger, T.; Droz, C.; Romanin, K.; Bergqvist, P.-A.; Müller, M.D.; Buser, H.-R. Occurrence of methyl triclosan, a transformation product of the bactericide triclosan, in fish from various lakes in Switzerland. *Environ. Sci. Technol.* **2004**, *38*, 390–395. [CrossRef] [PubMed]
- 136. Yang, L.-H.; Ying, G.-G.; Su, H.-C.; Stauber, J.; Adams, M.S.; Binet, M.T. Growth-inhibiting effects of 12 antibacterial agents and their mixtures on the freshwater microalga Pseudokirchneriella subcapitata. Environ. *Toxicol. Chem.* 2008, 27, 1201–1208. [CrossRef] [PubMed]
- Drury, B.; Scott, J.; Rosi-Marshall, E.J.; Kelly, J.J. Triclosan exposure increases triclosan resistance and influences taxonomic composition of benthic bacterial communities. *Environ. Sci. Technol.* 2013, 47, 8923–8930. [CrossRef] [PubMed]
- 138. Oliveira, R.; Domingues, I.; Koppe, G.C.; Soares, A.M. Effects of triclosan on zebrafish early-life stages and adults. *Environ. Sci. Pollut. Res. Int.* 2009, *16*, 679–688. [CrossRef] [PubMed]
- Crofton, K.M.; Paul, K.B.; DeVito, M.J.; Hedge, J.M. Short-term in vivo exposure to the water contaminant triclosan: evidence for disruption of thyroxine. *Environ. Toxicol. Pharmacol.* 2007, 24, 194–197. [CrossRef] [PubMed]
- Allmyr, M.; Adolfsson-Erici, M.; McLachlan, M.S.; Sandborgh-Englund, G. Triclosan in plasma and milk from Swedish nursing mothers and their exposure via personal care products. *Sci. Total Environ.* 2006, 372, 87–93. [CrossRef] [PubMed]
- 141. Boyd, G.R.; Reemtsma, H.; Grimm, D.A.; Mitra, S. Pharmaceuticals and personal care products (PPCPs) in surface and treated waters of Louisiana, USA and Canada. *Sci. Total Environ.* **2003**, *311*, 135–149. [CrossRef]
- 142. Stackelberg, P.E.; Furlong, E.T.; Meyer, M.T.; Zaugg, S.D.; Hendersen, A.K.; Reissman, D.B. Persistence of pharmaceutical and other organic wastewater contaminants in a conventional drinking-water-treatment plant. *Sci. Total Environ.* **2004**, *329*, 99–113. [CrossRef] [PubMed]
- Dhillon, G.S.; Kaur, S.; Pulicharla, R.; Brar, S.K.; Cledón, M.; Verma, M.; Surampalli, R.Y. Triclosan: current status, occurrence, environmental risks and bioaccumulation potential. *Int. J. Environ. Res. Public Health* 2015, 12, 5657–5684. [CrossRef] [PubMed]
- 144. Moore, C.J. Synthetic polymers in the marine environment: A rapidly increasing, long term threat. *Environ. Res.* **2008**, *108*, 131–139. [CrossRef] [PubMed]
- 145. Fendall, L.S.; Sewell, M.A. Contributing to marine pollution by washing your face: Microplastics in facial cleanser. *Mar. Pollut. Bull.* **2009**, *58*, 1225–1228. [CrossRef] [PubMed]
- 146. Cole, M.; Lindeque, P.; Halsband, C.; Galloway, T. Microplastics as contaminants in the marine environment: A review. *Mar. Pollut. Bull.* **2011**, *62*, 2588–2597. [CrossRef] [PubMed]

- 147. Gouin, T.; Avalos, J.; Brunning, I.; Brzuska, K.; de Graaf, J.; Kaumanns, J.; Konong, T.; Meyberg, M.; Rettinger, K.; Schlatter, H.; et al. Use of micro-plastic beads in cosmetic products in Europe and their estimated emissions to the North Sea environment. *SOFW J.* **2015**, *3*, 1–33.
- Napper, I.E.; Bakir, A.; Rowland, S.J.; Thompson, R.C. Characterisation, quantity and sorptive properties of microplastics extracted from cosmetics. *Mar. Pollut. Bull.* 2015, 99, 178–185. [CrossRef] [PubMed]
- 149. Derraik, J.G.B. The pollution of the marine environment by plastic debris: a review. *Mar. Pollut. Bull.* 2002, 44, 842–852. [CrossRef]
- 150. Van Cauwenberghe, L.; Devriese, L.; Galgani, F.; Robbens, J.; Janssen, C.R. Microplastics in sediments: A review of techniques, occurrence and effects. *Mar. Environ. Res.* **2015**, *111*, 5–17. [CrossRef] [PubMed]
- 151. Gouin, T.; Roche, N.; Lohmann, R.; Hodges, G. A thermodynamic approach for assessing the environmental exposure of chemical absorbed to microplastic. *Environ. Sci. Technol.* **2011**, *45*, 1466–1472. [CrossRef] [PubMed]
- 152. Mato, Y.; Isobe, T.; Takada, H.; Kanehiro, H.; Ohtake, C.; Kaminuma, T. Plastic resin pellets as a transport medium for toxic chemicals in the marine environment. *Environ. Sci. Technol.* 2001, 35, 318–324. [CrossRef] [PubMed]
- 153. Teuten, E.L.; Saquing, J.M.; Knappe, D.R.U.; Morton, A.M.A.; Jonsson, S.; Björn, A.; Rowland, S.J.; Thompson, R.C.; Galloway, T.S.; Yamashita, R.; et al. Transport and release of chemicals from plastics to the environment and to wildlife. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2009**, *364*, 2027–2045. [CrossRef] [PubMed]
- 154. Von Moos, N.; Burkhardt-Holm, P.; Koehler, A. Uptake and effects of micro-plastics on cells and tissues of the blue mussel Mytilus edulis L. after experimental exposure. *Environ. Sci. Technol.* 2012, 46, 11327–11335. [CrossRef] [PubMed]
- 155. Van Cauwenberghe, L.; Janssen, C.R. Microplastics in bivalves cultured for human consumption. *Environ. Pollut.* **2014**, 193, 65–70. [CrossRef] [PubMed]
- 156. Boerger, C.M.; Lattin, G.L.; Moore, S.L.; Moore, C.J. Plastic ingestion by planktivorous fishes in the North Pacific Central Gyre. *Mar. Pollut. Bull.* **2010**, *60*, 2275–2278. [CrossRef] [PubMed]
- 157. Cole, M.; Lindeque, P.; Fileman, E.; Halsband, C.; Goodhead, R.; Moger, J.; Galloway, T.S. Microplastic ingestion by zooplankton. *Environ. Sci. Technol.* **2013**, *47*, 6646–6655. [CrossRef] [PubMed]
- 158. Wright, S.L.; Thompson, R.C.; Galloway, T.S. The physical impacts of microplastics on marine organisms: A review. *Environ. Pollut.* **2013**, *178*, 483–492. [CrossRef] [PubMed]
- 159. Sussarellu, R.; Suquet, M.; Thomas, Y.; Lambert, C.; Fabioux, C.; Pernet, M.E.J.; Le Goïc, N.; Quillien, V.; Mingant, C.; Epelboin, Y.; et al. Oyster reproduction is affected by exposure to polystyrene microplastics. *Proc. Natl. Acad. Sci. U.S.A.* 2016, 113, 2430–2435. [CrossRef] [PubMed]
- Eriksson, C.; Burton, H. Origins and biological accumulation of small plastic particles in fur seals from Macquarie Islands. *Ambio* 2003, *32*, 380–384. [CrossRef] [PubMed]
- Andrady, A.L. Microplastics in the marine environment. *Mar. Pollut. Bull.* 2011, 62, 1596–1605. [CrossRef]
 [PubMed]
- Wardrop, P.; Shimeta, J.; Nugegoda, D.; Morrison, P.D.; Miranda, A.; Tang, M.; Clarke, B.O. Chemical pollutants sorbed to ingested microbeads from personal care products accumulate in fish. *Environ. Sci. Technol.* 2016, 50, 4037–4044. [CrossRef] [PubMed]
- 163. Council of European Union 2014. Note 16263/14. Elimination of micro-plastics in products an urgent need. Available online: http://register.consilium.europa.eu/doc/srv?l=EN&f=ST%2016263%202014%20INIT (accessed on 10 March 2017).
- 164. Cosmetics Europe 2015. Available online: https://www.cosmeticseurope.eu/news-a-events/news/ 822-cosmetics-europe-issues-a-recommendation-on-solid-plastic-particles-plastic-micro-particles.html (accessed on 3 April 2017).
- 165. Congress of the United States of America 2015. Microbead-Free Waters Act of 2015. Available online: https://www.congress.gov/114/plaws/publ114/PLAW-114publ114.pdf (accessed on 10 March 2017).



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