Lifetime and Fluorescence Quantum Yield of Two Fluorescein-Amino Acid-Based Compounds in Different Organic Solvents and Gold Colloidal Suspensions

Synthesis of Compounds 1 and 2

Scheme S1 shows the synthetic route of fluorescein, the process started with Fisher's esterification, followed by coupling with the amino acids derivatives: (a) Boc-ser(TMS)-OH and (b) Boc-cys(4-methylbenzyl)-OH, using N,N⁻-dicyclohexylcarboiimide (DCC) as carboxylic activator group, which resulted in the esters *I* and *2*. The complete characterization of compounds *I* and *2* by ¹H and ¹³C NMR, elemental analysis, mass spectrometry (MALDI-TOF-MS), UV–vis absorption, emission spectroscopy, and IR spectra are presented in reference [1].



Scheme S1 Synthesis of compounds 1 and 2 in 86% and 90% yields are presented respectively.

Spectroscopy Characterization of compound 2 in different solvents

Figure S1 shows absorbance (a) and emission (b) of compound 2 suspended in different solvents.



Figure S1 (a) Absorbance and (b) normalized emission spectra of compound 2 in different solvents (concentration of 1×10^{-5} mol/L): (1) THF, (2) ethanol, (3) acetonitrile and (4) DMSO.

Spectroscopy Characterization of Fluorescein, 1 and 2 in Alkaline Solutions

Figure S2 presents the absorbance and normalized emission spectra of compounds *1*, *2* and fluorescein in alkaline aqueous solutions (pH 12.8), and similar results were observed for all compounds.



Figure S2 Absorbance and emission spectra for compound **1** (a), compound 2 (b) and fluorescein (c) in alkaline water pH 12.8 (λ_e = 430 nm).

Time-Resolved Photoluminescence Measurements

Experimental Setup

Figure S3 shows the time-resolved photoluminescence setup. In this case, a NanoLed at λ_{e} = 460 nm (1 MHz and 1.3 ns) was used as light source. The cuvette of liquid sample was placed in a sample holder positioned inside of the Horiba Jobin-Yvon Tempro equipment. The time-resolved photoluminescence signal was acquired at photomultiplier detector using a time-correlated single-photon counting (TCSPC) [2,3]. The TCSPC assumes that the probability distribution of a single luminescence photon is equivalent to the intensity as a function of the time distribution for all the photons emitted [3]. The incidence of the luminescent photon after the excitation by a NanoLed is converted in voltage or

time-to-pulse height conversion (TPHC), and the TPHC signal is digitized and stored in a multi-channel analyzer (MCA). Finally, the MCA produces a histogram of photon time distribution (Counts versus Channels) equivalent to the actual photoluminescence decay (Figure S3). In this form, the photon time distribution $\mathcal{F}(t)$ is obtained from the photomultiplier detector using time-correlated single-photon counting. For all compounds I and 2, $\mathcal{F}(t)$ results were fitted using an appropriate sum of exponentials as A+B1*Exp(-t/ τ_1) + B2*Exp(-t/ τ_2). Here A, B1 and B2 are constants, τ_1 and τ_2 are lifetimes values.



Figure S3 Time-resolved photoluminescence setup. $\mathcal{F}(t)$ is photon time distribution obtained from photomultiplier detector using a time-correlated single-photon counting (TCSPC).

Figure S4 shows the results of counts versus channels for compound I in alkaline aqueous solutions and Ludox solutions, obtained from time-resolved photoluminescence signal using a TCSPC. In this case, $\mathcal{F}(t)$ obtained from the photomultiplier detector was fitted using just one exponential. For all time-resolved photoluminescence measurements, a scattering sample is necessary for prompt measurement $\mathcal{P}(t)$. Being prompt is very important because $\mathcal{P}(t)$ is used for iterative comparison with $\mathcal{F}(t)$ for determination of the fit parameters.



Figure S4 Lifetime measurements of (a) compound *I* in alkaline aqueous solutions (concentration of 1 x 10⁻⁵ mol/L) and (b) Ludox (Time calibration is 5.487x 10⁻¹¹ sec/ch). Fitting the experimental results, the parameters $\tau_1 = (4.057 \pm 0.005)$ ns, B1= 100 Rel. Ampl., and $\chi^2 = 1.01$ were obtained.

References

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