

# Article

# Optimization of Extrusion and Ultrasound-Assisted Extraction of Phenolic Compounds from Jizi439 Black Wheat Bran

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**Abstract:** Jizi439, a newly developed black wheat breeding line, was reported to effectively regulate blood glucose, which may potentially be associated with its intrinsic high level of phenolic compounds (PCs). To maximize the PCs yield and thereby enhance their antioxidant activity, orthogonal experiments were designed in sequence for extrusion of Jizi439 black wheat bran (BWB) powder and followed by the extraction of PCs assisted with ultrasound technique. White wheat bran was used as a control. The optimum condition for extrusion was 110 °C, 25% feed water content, 140 rpm screw speed; meanwhile, 50 °C, 40 min, 35 kHz ultrasonic frequency, 300 W ultrasonic power for ultrasound-assisted extraction (UAE). Total phenolic content (TPC) as determined by Folin–Ciocalteu method was 2856.3  $\pm$  57.7 µg gallic acid equivalents (GAE) per gram of dry weight (DW) of phenolic extract; meanwhile, antioxidant activity (AA) in terms of DPPH radical scavenging ratio was 85.5%  $\pm$  1.1% under optimized conditions, which were both significantly higher than the control. Phenolic acids except for gallic acid, as well as flavonoids, including luteolin and apigenin were increased by extrusion and ultrasound, as suggested by HPLC results. In conclusion, our study would provide a valuable reference for processing Jizi439 BWB before making or commercially utilize it into health-related food products.

**Keywords:** extrusion; ultrasound-assisted extraction; Jizi439 black wheat; phenolic compounds; orthogonal experiment

# 1. Introduction

Wheat bran, the by-product of the wheat milling process, is commonly discarded and destined for animal feeding. However, due to the rising findings on the health benefits of its components, especially phenolic compounds, the use of bran as a food ingredient is increasing [1]. Jizi439 black wheat was recently developed by the combination of genes from four different breeding lines, namely, Australia black wheat 14,928, 95-5031, durum wheat 190,268, and Henan black wheat Luozhen No.1, by Hebei Academy of Agricultural and Forestry Sciences (Hebei, China) [2]. This breeding has been reported to effectively regulate blood glucose levels of Type 2 Diabetes patients after ingestion [3], which may (to some extent) associated with the high level of chromium and low GI index [3–6].

Except for that, phenolic compounds in the bran could be another factor in regulating the blood glucose levels of Type 2 Diabetes patients. Since diabetes is associated with not only hyperglycemia and



hyperlipidemia, but also oxidative stress caused by free radicals [7]. Phenolic compounds can serve as

natural antioxidants that act against those free radicals [8], and lower the risk of mortality from Type 2 diabetes and obesity [9–12]. It was reported that wheat bran phenolics were of notable antioxidant activity [13,14]. Among different varieties of wheat (white, red, purple, black, etc.), black wheat possesses the highest level of phenolics [15,16]. Thus, the phenolics from the newly developed breeding Jizi439 were selected in the present study, which to our knowledge, is rarely reported.

Wheat bran is often pre-processed, so it has a more desirable sensory acceptance and nutritional value, before it is put into the market. Extrusion is a versatile process comprising operations, including mixing, cooking, kneading, shearing, shaping, and forming [17]. Research on cereal products has shown that thermal processing, such as extrusion might assist in releasing bound phenolics by breaking down cellular constituents and cell walls [18]. Studies reported that the antioxidant activity could be significantly increased by extrusion of wheat bran [19,20].

Besides extrusion, ultrasound-assisted extraction is another efficient way to maximize the yield of phenolic compounds [21,22], which is well known to have a significant positive effect on the extraction rate in the chemical and food industry. Using this technique, full extraction could be completed quickly, solvent consumption and fossil energy could be reduced, which is superior to conventional extraction, such as water or enzyme-assistant extraction [22].

The present study aims to optimize the extrusion and ultrasound-assistant extraction conditions in order to obtain more phenolic compounds and improve the antioxidant activity of extracts. In this respect, two orthogonal experiments were conducted for extrusion and ultrasound-assistant extraction in sequence, while total phenolic content and the antioxidant activity of the extracts were used as responses for the orthogonal design. Phenolic profiles were compared among extracts of the control, raw, and treated Jizi439 BWB as preliminary investigation for further study of Jizi439 phenolic compounds.

#### 2. Materials and Methods

#### 2.1. Raw Materials and Chemicals

The Jizi439 black wheat bran (BWB) was obtained from Yueqing Agricultural Science and Technology Co., Ltd. (Handan, China). Gallic acid standard was purchased from Aladdin chemical Co. (Shanghai, China). Tris-HCl buffer was purchased from Solarbio Science and Technology Co. (Beijing, China). DPPH (2,2-diphenyl-1-picrylhydrazyl) free radical was purchased from Sigma-Aldrich (St. Louis, MO, USA). Folin–Ciocalteu reagent and sodium carbonate anhydrous were purchased from Hushi chemical Co. (Shanghai, China). Phenolic compounds standards were purchased from Shanghai Yuanye Bio-Technology Co., Ltd. (Shanghai, China).

#### 2.2. Experiment Design

The scheme of optimizing Jizi439 BWB extrusion and extraction processing steps are shown in Figure 1. Two orthogonal experiments were conducted, respectively, for extrusion and ultrasound-assisted extraction (UAE). Total phenolic content (TPC) and antioxidant activity (AA, in terms of DPPH scavenging activity) were used as responses. After optimization, the Jizi439 BWB was divided into four groups, including: (1) NENU group, raw bran without extrusion and phenolics were extracted without ultrasound; (2) E group, extruded bran whose phenolics were extracted without ultrasound, (3) U group, raw bran with UAE; and (4) EU group, extruded bran with UAE. White wheat bran, without extrusion and ultrasound, was used as a control. Total phenolic content and antioxidant activity were compared among each group. The profile of phenolic compounds in the extracts was compared among NENU, EU, and control groups.



Figure 1. Scheme of optimizing Jizi439 black wheat bran (BWB) processing steps.

#### 2.3. Orthogonal Experiment for Extrusion and UAE

Jizi439 BWB was extruded in a twin-screw extruder (FMHE36-24; Hunan Fumach Foodstuff Engineering and Technology Co., Ltd., Hunan, China). The length (L) of the screw was 864 mm, and its diameter (D) was 36 mm (L/D = 24:1). The feed rate was 125 g/min. To optimize extrusion condition, an L<sub>9</sub>(3<sup>4</sup>) orthogonal design was set up with three levels of screw speeds (140, 160, or 180 rpm), feed water content (21%, 23%, or 25%), and the third heating block temperature (90, 110, or 130 °C), respectively (Table 1). The temperature in the first, second, fourth, and fifth heating blocks were 60, 90, 60, and 50 °C, respectively [23]. The extrudates were dried at 60 °C for 30 min, ground and passed through a 60-mesh sieve, and then stored at -18 °C in sealed plastic bags and protected from light until further study.

**Table 1.**  $L_9(3^4)$  orthogonal factors and levels of extrusion.

T1	I	s	
Level	Α	В	С
1	90	21	140
2	110	23	160
3	130	25	180

A, the third heating block temperature (°C); B, feed water content (%); C, screw speeds (rpm).

Phenolic compounds were extracted from extruded Jizi439 BWB with 80% aqueous ethanol assisted by an ultrasound electronic instrument (THC-2B, Jining Tianhua Ultrasound Electronic Instrument Co., Ltd., Jining, China) [14]. The orthogonal design was set up with three levels of temperature (30, 50, or 70 °C), time (20, 40, or 60 min), ultrasonic frequency (20, 35, or 50 kHz), and ultrasonic power (200, 300, or 400 W), respectively (Table 2). Extracts were then centrifuged at 2800×g for 15 min. Supernatants were collected for determination of TPC, or concentrated under rotary evaporation and freeze-dried for DPPH scavenging activity assay and HPLC analysis.

**Table 2.**  $L_9(3^4)$  orthogonal factors and levels of extraction.

	Factors					
Level	Α	В	С	D		
1	30	20	20	200		
2	50	40	35	300		
3	70	60	50	400		

A, temperature (°C); B, time (min); C, ultrasonic frequency (kHz); D, ultrasonic power (Watt).

#### 2.4. Determination of TPC

The TPC of extracts were determined by Folin–Ciocalteu method, according to Zhang et al., in 2006, with slight modifications [24]. Briefly, 20  $\mu$ L of each sample solution was mixed with 100  $\mu$ L of

Folin–Ciocalteu reagent and loaded on a 96-well plate, incubated for 5 min before 80  $\mu$ L of 7.5% Na<sub>2</sub>CO<sub>3</sub> solution was added. The 96-well plate was then covered and left in a dark at room temperature for 2 h. The absorbance was measured at 750 nm with a spectrophotometric microplate reader (EnSpire<sup>®</sup> Multimode Plate reader, PerkinElmer Management (Shanghai) Co., Ltd., China). Distilled water was used as a blank. Each sample solution and the standard solution was repeated in sextuplicate. The TPC was expressed as micrograms of gallic acid equivalents per gram of dry weight BWB ( $\mu$ g GAE/g DW) calculated from a calibration curve of a standard gallic acid solution with concentrations of 0–100  $\mu$ g/mL (R<sup>2</sup> > 0.999).

# 2.5. Determination of Antioxidant Activity (AA)

The antioxidant activity (AA) of Jizi439 BWB extracts was tested by DPPH radical scavenging assay, according to Li and Shimamura [25,26] with slight modifications. Extracts were dried under a nitrogen flow and weighed before re-dissolved in 3 mL 80% methanol. The extracts were then diluted 10, 20, 50, and 100 times and analyzed for DPPH scavenging activity. Briefly, the DPPH solution was freshly made for each run, 20  $\mu$ L of sample solutions with 80  $\mu$ L of 0.1 M Tris-HCl buffer was added into each well before 100  $\mu$ L of DPPH solution was added. Left in a dark at room temperature for 30 min. The absorbance was measured at 515 nm with a spectrophotometric microplate reader (EnSpire<sup>®</sup> Multimode Plate reader, PerkinElmer Management (Shanghai) Co., Ltd., Shanghai, China). Each sample solution was repeated in sextuplicate. The inhibition ratio of the phenolic extracts was calculated according to the following Equation (1):

Inhibitioin ratio (%) = 
$$\left(1 - \frac{A_s - A_1}{A_c}\right) \times 100$$
 (1)

Determination of the DPPH. As is the absorbance of DPPH solution and Tris-HCl buffer with sample solution; Ac is the absorbance of DPPH solution Tris-HCl buffer with ethanol (80%) instead of the sample; A1 is the absorbance of Tris-HCl buffer and sample solution with methanol (80%) for background absorbance.

The  $IC_{50}$  value of each phenolic extract was determined by interpolation from linear regression analysis using the SPSS statistics program (SPSS Inc., Chicago, IL, USA) [27]. Lower  $IC_{50}$  values indicate higher antioxidant activities.

#### 2.6. HPLC Analysis of Phenolic Compounds

The profile of phenolic compounds in Jizi439 BWB extracts were analyzed by a Waters e2695 HPLC system equipped with a symmetry C18 column ( $250 \times 4.6 \text{ mm}$ , 5 µm) and a UV detector (Milford, MA, USA) using methanol (solvent A) and 0.1% formic acid solution (solvent B) as mobile phase: 5% A in 5 min, 5–15% A in 10 min, 37–38% A in 25 min, 38–50% A in 10 min, 50–95% A in 10 min, 95% A in 5 min and 95–5% A in 2 min, at a flow rate of 0.5 mL/min. The injection volume was 10 µL, and the column temperature was 30 °C. Pure compounds (*p*-hydroxybenzoic acid, gallic acid, vanillic acid, *p*-coumaric acid, ferulic acid, syringic acid, caffeic acid, chlorogenic acid, luteolin, and apigenin) were used as standards. The detection of components was monitored at 280 nm. Retention times and ultraviolet-visible spectra of obtained chromatographic peaks were matched with those of the standards. Samples were prepared and analyzed in duplicate.

#### 2.7. Statistical Analysis

The result of TPC and AA were expressed as average  $\pm$  standard deviation (SD). Data were analyzed by one-way ANOVA and Duncan's multiple range test at a 5% significance level (p < 0.05) by using the SPSS statistics program (SPSS Inc., Chicago, IL, USA).

# 3. Results

## 3.1. Orthogonal Experiment for Extrusion

The orthogonal experiment was carried out according to Table 1. Effects of the three factors, i.e., the third heating block temperature (A), feed water content (B), and screw speeds (C) on TPC was analyzed by the range and variance analysis.

*Range analysis.* The results of the range analysis were shown in Table 3. The average of responses at each level and their ranges were calculated and expressed as  $k_1$ ,  $k_2$ ,  $k_3$ , and R. The value of k could be used to assess the influence of each actor level on the response, while R values reflect the importance of each factor. A larger k or R represents a more desirable condition or a greater effect of the factor on the response. K values were calculated according to Li [28].

Experime	nt No.	Α	В	С	TPC (μg GAE/g DW)	AA DPPH Scavenging Activity (%)
1		1	1	1	$1848.0 \pm 138.1$	$69.5 \pm 2.3$
2		1	2	2	$1897.2 \pm 40.9$	$71.2 \pm 1.1$
3		1	3	3	$1951.0 \pm 88.3$	$72.3 \pm 1.6$
4		2	1	2	$2106.8 \pm 93.7$	$83.3 \pm 1.3$
5		2	2	3	$1956.5 \pm 64.6$	$75.4 \pm 1.7$
6		2	3	1	$2232.4 \pm 64.9$	$86.7 \pm 2.9$
7		3	1	3	$1951.8 \pm 30.4$	$81.1 \pm 0.6$
8		3	2	1	$2024.7 \pm 69.1$	$85.0 \pm 1.8$
9		3	3	2	$1999.3 \pm 120.3$	$82.9 \pm 1.8$
	$k_1$	1898.7	1968.9	2035.0		
TDC	k <sub>2</sub>	2098.6	1959.5	2001.1		
Irc	k <sub>3</sub>	1991.9	2060.9	1953.1		
	R	199.8	101.4	82.0		
	k <sub>1</sub>	71.0	78.0	80.4		
	k <sub>2</sub>	81.8	77.2	79.1		
AA	k <sub>3</sub>	83.0	80.6	76.2		
	R	12.0	3.4	4.1		

**Table 3.**  $L_9(3^4)$  Orthogonal experiment for extrusion of BWB.

Means ± SD, n = 3; A, the third heating block temperature (°C); B, feed water content (%); C, screw speeds (rpm); TPC, total phenolic content; AA, antioxidant activity in terms of DPPH radical scavenging activity;  $k_1$ ,  $k_2$ ,  $k_3$ , average responses at level 1, 2, 3; R, range of average responses.

For response TPC, results showed that  $R_A > R_B > R_C$ , this means A (the third heating block temperature) had the greatest effect on TPC, followed by B (feed water content) and C (screw speeds). According to k values of A, B, and C, the optimum combination was  $A_2B_3C_1$ . For response AA, results showed that  $R_A > R_C > R_B$ , and the optimum combination is  $A_3B_3C_1$ , according to k values. As shown in Figure 2, TPC reached a peak value at 110 °C; however, for response AA, the peak was at 130 °C. The impact of factors B and C on TPC and AA show similar trends. A verification experiment was conducted using combination  $A_2B_3C_1$  the best combination for both TPC and AA, higher TPC (2304.3 ± 127.4 µg GAE/g DW), and AA (86.2% ± 0.8%) was obtained. Thus, the optimum extrusion conditions should be A2B3C1 (the third heating block temperature 110 °C, feed water content 25%, and screw speeds 140 rpm).

*Variance analysis.* The variance analysis was carried out to assess the influence of each factor on TPC and AA. As shown in Table 3, SS and MS represent for the sum of the squares of deviations and the sum of mean-square error, respectively. F and p value were used to determine whether the factors significantly impact the response or not. A larger F value or a smaller p value represents a stronger impact of the factor. According to Table 4, factor A was the most significant factor (p < 0.05) that influence AA, which was consistent with the Range analysis above.



**Figure 2.** The effect of different extrusion parameters on TPC (**a**) and AA (**b**). A, the third heating block temperature (°C); B, feed water content (%); C, screw speeds (rpm).

Variance Sources	Response	Free Degree	SS	MS	F	p
٨	TPC	2	59,990	29,995	2.9	0.26
A	AA	2	316	158	26	0.04
В	TPC	2	23,760	11,880	1.1	0.47
	AA	2	8.8	4.4	0.7	0.58
С	TPC	2	1765	882	0.1	0.92
	AA	2	1.1	0.6	0.1	0.91
Error	TPC	2	20,682	10,341		
	AA	2	12.0	6		

**Table 4.** Variance analysis of  $L_9(3^4)$  orthogonal experiment for extrusion.

A, the third heating block temperature (°C); B, feed water content (%); C, screw speeds (rpm); SS, the sum of the squares of deviations; MS, the sum of mean-square error. The critical F value is 39.00 at 97.5% confidence, and 19.00 at 95.0% confidence.

#### 3.2. Orthogonal Experiment for Ultrasound-Assisted Extraction (UAE)

To further maximize the TPC yield of Jizi439 BWB, another  $L_9(3^4)$  orthogonal experiment was conducted using Jizi439 BWB, which extruded under the optimum condition mentioned in Section 3.1. Table 4 shows the assignment of the orthogonal factors and their levels. The four factors were temperature (A), time (B), ultrasonic frequency (C), and ultrasound power (D). The effects of four factors on TPC were analyzed by range (Table 5) and variance analysis (Table 6).

*Range analysis.* According to Table 5, for response TPC,  $R_A > R_C > R_B > R_D$ , this means A (temperature) had the greatest effect on TPC, followed by C (ultrasonic frequency), B (time) and D (ultrasonic power). And the optimum process combination was  $A_3B_2C_2D_2$ , according to k values. For response AA, results showed that  $R_A > R_D > R_B > R_c$ , which means A had the greatest effect on AA, followed by D and B, and the optimum combination was  $A_2B_1(B_2)C_3D_3$ , according to k values. TPC and AA reached peaks at 70 °C and 50 °C, respectively (Figure 3). The results of the verification experiment showed that both higher TPC (2856.3 ± 57.7 µg GAE/g DW) and AA (85.5% ± 1.1%) were obtained under combination  $A_2B_2C_2D_2$ . Thus,  $A_2B_2C_2D_2$  (50 °C, 40 min, ultrasonic frequency 35 kHz, and ultrasonic power 300 W) should be the optimum UAE conditions.

*Variance analysis.* The results of the variance analysis for ultrasound-assisted extraction (UAE) were listed in Table 6. Factor A was the most significant factor (p < 0.05) for both TPC and AA. Factor B and C could significantly influence TPC (p < 0.05), while factor D could significantly influence AA (p < 0.05).

#### 3.3. The Effect of Extrusion and UAE on TPC and AA of Jizi439 BWB

Jizi439 BWB powder was divided into four groups, including NENU, E, U, and EU groups. The white wheat bran was used as a control. As shown in Figure 4, the TPC and AA of Jizi439 BWB were both significantly improved by extrusion or UAE, which were up to 2856.3  $\pm$  57.7 µg GAE/g DW and 85.5%  $\pm$  1.1% when two processes were both conducted.

Experimen	t No.	Α	В	С	D	TPC (μg GAE/g DW)	AA DPPH Scavenging Activity (%)
1		1	1	1	1	$2188.2 \pm 110.0$	$79.0 \pm 1.1$
2		1	2	2	2	$2499.2 \pm 38.8$	$79.7 \pm 1.6$
3		1	3	3	3	$2301.3 \pm 145.9$	$80.3 \pm 1.1$
4		2	1	2	3	$2491.7 \pm 14.6$	$86.8 \pm 1.3$
5		2	2	3	1	$2370.9 \pm 187.4$	$85.6 \pm 1.5$
6		2	3	1	2	$2527.3 \pm 96.0$	$85.1 \pm 0.8$
7		3	1	3	2	$2682.7 \pm 26.6$	$72.4 \pm 1.0$
8		3	2	1	3	$2898.5 \pm 230.1$	$72.9 \pm 0.9$
9		3	3	2	1	$2883.2 \pm 87.6$	$71.0\pm0.6$
	k <sub>1</sub>	2393.6	2454.2	2538.0	2480.8		
TDC	$k_2$	2463.3	2589.5	2624.7	2569.8		
IFC	k <sub>3</sub>	2821.5	2570.6	2451.6	2563.8		
	R	491.9	135.3	173.1	89.0		
	k <sub>1</sub>	79.7	79.4	79.0	78.6		
	k <sub>2</sub>	85.8	79.4	79.2	79.1		
AA	$k_3$	72.1	78.8	79.4	80.0		
	R	13.7	0.6	0.4	1.43		

**Table 5.**  $L_9(3^4)$  Orthogonal experiment for extraction of BWB.

Means  $\pm$  SD, n = 3; A, temperature (°C); B, time (min); C, ultrasonic frequency (kHz); D, ultrasonic power(Watt); TPC, total phenolic content; AA, antioxidant activity; k<sub>1</sub>, k<sub>2</sub>, k<sub>3</sub>, average responses at level 1, 2, 3; R, range of average responses.

Variance Sources	Response	Free Degree	SS	MS	F	р
٨	TPC	2	1,164,365	582,182	37.2	0.00
A	AA	2	851	426	326.5	0.00
D	TPC	2	96,645	48,322	3.1	0.07
В	AA	2	2.0	1.0	0.8	0.47
С	TPC	2	134,773	67,387	4.3	0.03
	AA	2	0.7	0.4	0.3	0.76
	TPC	2	44,477	22,238	1.4	0.27
D	AA	2	9.5	4.7	3.6	0.04
Error	TPC	2	281,516	15,640		
	AA	2	23.5	1.3		

**Table 6.** Variance analysis of  $L_9(3^4)$  orthogonal experiment for UAE.

A, temperature (°C); B, time (min); C, ultrasonic frequency (kHz); D, ultrasonic power (Watt); SS, the sum of the squares of deviations; MS, the sum of mean-square error. The critical F value is 199.00 at 99.5% confidence, 99.01 at 99.0% confidence, 39.00 at 97.5% confidence, and 19.00 at 95.0% confidence.



**Figure 3.** The effect of different UAE parameters on TPC (**a**) and AA (**b**). A, temperature (°C); B, time (min); C, ultrasonic frequency (kHz); D, ultrasonic power (Watt).



**Figure 4.** Effect of different treatments on TPC and AA of Jizi439 BWB. Control, white wheat bran (without extrusion and UAE); NENU, raw black wheat bran (without extrusion and UAE); E, extruded but without UAE black wheat bran; U, UAE but without extruded black wheat bran; EU, extruded, and UAE black wheat bran. (**a**–**e**) Indicate a significant difference between different groups (n = 3; p < 0.05).

The IC<sub>50</sub> values were calculated by interpolation from linear regression analysis of the percentage of scavenging activity (AA) against sample concentration (mg/mL) using the SPSS statistics program, and lower IC<sub>50</sub> values represent higher antioxidant activity. There is no significant difference between the control and NENU groups. Extrusion or UAE could decrease IC<sub>50</sub> values significantly, with the EU group being the lowest, suggesting that extrusion and UAE could increase AA (Figure 5).



**Figure 5.** IC<sub>50</sub> values of phenolic extracts from Jizi439 BWB by different treatments. Control, white wheat bran (without extrusion and UAE); NENU, raw Jizi439 BWB (without extrusion and UAE); E, extruded but without UAE; U, UAE but without extruded; EU, extruded, and UAE; (**a**–**d**) Indicate a significant difference between different groups (n = 3; p < 0.05).

#### 3.4. Profile of Phenolic Compounds in Jizi439 BWB Extracts

The identification of phenolic compounds from Jizi439 BWB extracts was carried out by comparison of retention times with external standards. The HPLC chromatogram of 11 pure compositions mixture standard was shown in Figure 6.

The profiles of phenolic compounds from NENU, EU, and control groups were shown in Table 7. Three simple phenolic acids, including gallic acid, syringic acid, and *p*-coumaric acid, were observed in the control group. Gallic acid, chlorogenic acid, vanillic acid, caffeic acid, luteolin, and apigenin were identified in NENU and EU groups. Gallic acid in NENU and EU groups were significantly lower than that of control. Other phenolic acids, including chlorogenic acid, vanillic acid, caffeic acid, caffeic acid,

and flavonoids (including luteolin, and apigenin) in the EU group were significantly higher than the NENU group.



**Figure 6.** HPLC chromatogram of mixed standards. Peak 1, gallic acid; 2, chlorogenic acid; 3, *p*-hydroxybenzoic acid; 4, vanillic acid and caffeic acid; 5, syringic acid; 6, *p*-coumaric acid; 7, ferulic acid; 8, *o*-coumaric acid; 9, luteolin; 10, apigenin.

Table 7. Contents of identified phenolic compounds in extracts from Jizi439 BWB (µg/mg dry matter).

	Control	NENU	EU
Gallic acid	$2.80 \pm 0.09a$	$0.46 \pm 0.02b$	$0.47\pm0.001\mathrm{b}$
Chlorogenic acid	ND	$2.17 \pm 0.01b$	$2.41 \pm 0.001a$
Vanillic acid and caffeic acid	ND	$0.65 \pm 0.003b$	$0.80 \pm 0.001a$
Syringic acid	$1.16 \pm 0.002$	ND	ND
P-coumaric acid	$0.92 \pm 0.02$	ND	ND
Luteolin	ND	$1.30 \pm 0.05b$	$2.10 \pm 0.01a$
Apigenin	ND	$0.02 \pm 0.002b$	$0.03 \pm 0.001a$

Control, white wheat bran (without extrusion and UAE); NENU, raw black wheat bran (without extrusion and UAE); EU, extruded, and UAE black wheat bran. (**a–b**) Indicate a significant difference between different groups (n = 3; p < 0.05); ND, not detected.

## 4. Discussion

Extrusion has been reported to increase TPC and AA of snacks made from beans [29] or wheat bran [23]. It could potentially improve sensory acceptance and nutritional properties of many food products, including rice, sorghum, apple pomace [30]. Our study showed that the optimized extrusion processes should be 110 °C for the third heating block temperature, 25% feed water content, and 140 rpm screw speeds. Results in Tables 3 and 4 indicated that extrusion temperature was the most important factor on AA, which reached peak value at 130 °C, according to Figure 2. Similar results were found previously by Sharama et al., and the possible explanation could be the browning pigments that generated from Maillard reaction enhanced AA [31]. Increased temperature promoted the release of phenolic compounds, so that TPC reached peak value at 110 °C; however, it decreased along with the increase of temperature (Figure 2) as phenolics are unstable when the temperature is over 120 °C [32].

An increase in feed water content resulted in higher TPC and AA (Table 3), possibly because of the weakened shearing effects by water in the extruder [33]. There's controversy regarding whether screw speed could increase AA or not, our results was in accordance with Khanal et al. [34] that AA increased with screw speed, which was due to the procyanidin in the wheat bran degraded under high screw speed. On the contrary, a decrease of AA was found when the screw speed increased in Ozer's research [33]. Thus, more work should be addressed on the association between screw speed and AA during the extrusion process in the future.

Based on the optimized extrusion condition, another orthogonal experiment was designed for UAE. Results showed that the extraction temperature was the most significant factor on TPC (p < 0.05) and AA (p < 0.005). TPC reached maximum value when the temperature went up to 70 °C (Figure 3), since more phenolics diffused into the solvent at higher temperature [35]. Conversely, AA was declined after it reached the peak value at 50 °C (Figure 3). Similar phenomena were reported by Thoo et al. [36]. Unlike extrusion, whose thermal process is within a short time, heat treatment during extraction is a relatively long-time process that could cause the degradation of polyphenols, and therefore, leading to a significant loss of antioxidant capacity [36–38]. The optimal extraction time for TPC and AA was different, because DPPH scavenging ability depends not only on phenolics, but also other antioxidant compounds that may exist in the extracts [39]. Ultrasonic frequency possessed the second most significant impact on TPC (p < 0.05), and the optimal condition was at the medium level of 35 kHz. Increased frequency and energy provided by ultrasound could release phenolics from the matrix, while paradoxically accelerate the degradation of phenolics [40]. Ultrasonic power was another factor that significantly increases AA (p < 0.05) according to our results (Table 6), which was reported before in tannin extraction [41]. Thus, the optimized UAE processes should be 50  $^{\circ}$ C, 40 min, 35 kHz, and 300 W for ultrasonic frequency and power. Verification experiments were conducted under optimized extrusion and extraction conditions. As shown in Figures 2 and 3, TPC and AA of Jizi439 BWB were significantly increased by optimized extrusion, and especially UAE processing (p < 0.05). Extrusion could partially break the ester bonds between phenolics and cell wall matrix that resulted in the release of bound phenolics. Meanwhile, sonication simultaneously enhanced the hydration and fragmentation process, while facilitating the mass transfer of solutes to the extraction solvent [21].

The profile of phenolic extracts from Jizi439 BWB was investigated by HPLC analysis; results were shown in Table 7. Four phenolic acid (gallic acid, chlorogenic acid, vanillic acid, and caffeic acid), and two flavonoids (luteolin and apigenin) had been detected in the extracts. Chlorogenic acid was the predominant phenolic compound in Jizi439 BWB according to our results, which was different from other studies that ferulic acid is the most abundant phenolic compound in wheat [42,43]. Although differences of wheat varieties and growing environments could be the reason for that [44], more investigation regarding phenolic profile in Jizi439 BWB should be done in the future. It should be noted that gallic acid was not increased by either extrusion or UAE, indicating that gallic acid may not be freed from bonds with other molecules, a similar phenomenon was mentioned before by Gaxiola-Cuevas et al. [45]. Luteolin exhibited the greatest increase after extrusion and UAE, which was in accordance with Hernandez et al. [46] in black bean.

#### 5. Conclusions

Both extrusion and UAE could significantly increase the TPC and AA of Jizi439 BWB. The optimized extrusion combination was the third heating block temperature 110 °C, feed water content 25%, screw speeds 140 rpm; meanwhile, the optimum UAE combination was temperature 50 °C, time 40 min, 35 kHz, and 300 W for ultrasonic frequency and power. Preliminary investigation of the phenolic profile by HPLC suggested that by optimizing extrusion and extraction condition, phenolic acids (except for gallic acid), and flavonoid (especially luteolin) were significantly increased in Jizi439 BWB. Our study would provide a valuable reference for processing Jizi439 BWB before making or commercially utilize it into health-related food products.

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