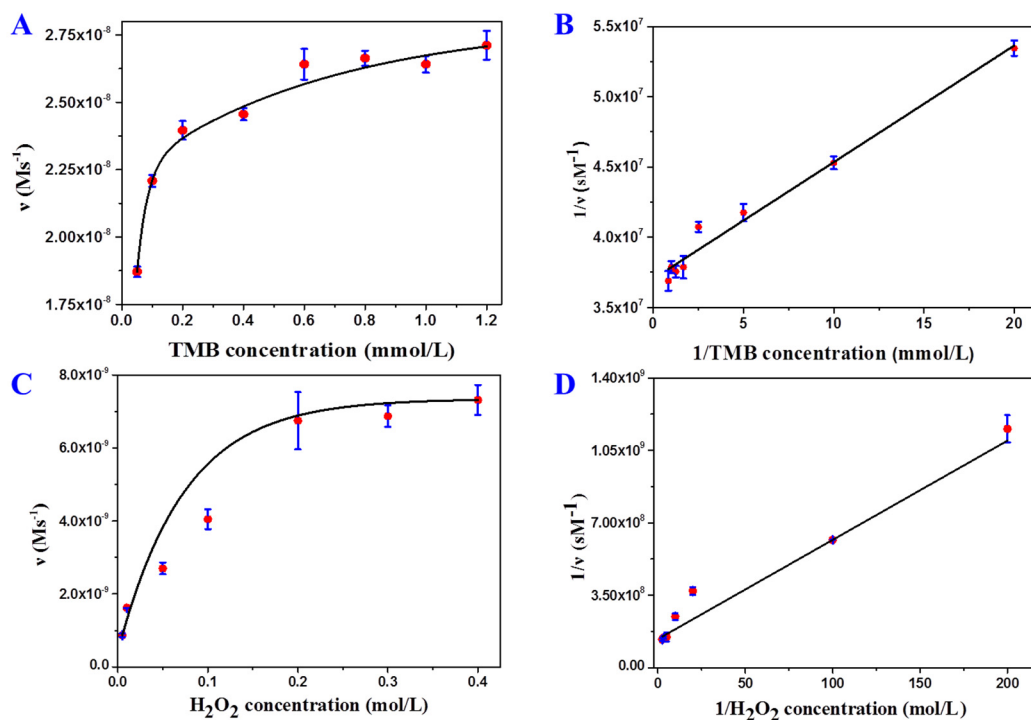
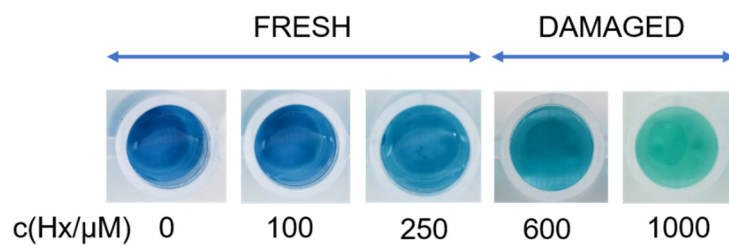


**Figure S1.** (A) TEM image of PVP-PtNCs to show its uniformity and cube shape, (B) Image of single PVP-PtNC taken by a high-resolution TEM



**Figure S2.** The kinetic assays of PVP-PtNC as artificial peroxidase-like catalysts for oxidation of TMB by  $\text{H}_2\text{O}_2$ . (A) Graphic representation of the plots of initial rate ( $v$ ) vs. TMB concentration; (B) double-reciprocal plot generated from (A); (C) Graphic representation of  $v$  vs.  $\text{H}_2\text{O}_2$  concentration; (D) double-reciprocal plot generated from (C)



**Figure S3.** The colorimetric images of as-proposed method under different concentrations of Hx.

**Table S1** Comparison of PVP-PtNC kinetic parameters and HRP

Catalyst	[E] (M) <sup>a</sup>	Substrate	$V_{\max}$ (Ms <sup>-1</sup> ) <sup>b</sup>	$K_m$ (M) <sup>c</sup>	$K_{\text{cat}}$ (s <sup>-1</sup> ) <sup>d</sup>
PVP-PtNC	$1.67 \times 10^{-11}$	TMB	$1.05 \times 10^{-6}$	$2.22 \times 10^{-5}$	$6.29 \times 10^4$
PVP-PtNC	$1.67 \times 10^{-11}$	H <sub>2</sub> O <sub>2</sub>	$3.61 \times 10^{-7}$	$3.92 \times 10^{-3}$	$2.16 \times 10^4$
HRP[1]	$2.5 \times 10^{-11}$	TMB	$10.00 \times 10^{-8}$	$4.34 \times 10^{-4}$	$4.00 \times 10^3$
HRP[1]	$2.5 \times 10^{-11}$	H <sub>2</sub> O <sub>2</sub>	$8.71 \times 10^{-8}$	$3.70 \times 10^{-3}$	$3.48 \times 10^3$

<sup>a</sup>[E] is the enzyme (or PVP-PtNC) concentration

<sup>b</sup> $K_m$  is the Michaelis constant

<sup>c</sup> $V_{\max}$  is the maximal reaction velocity

<sup>d</sup> $K_{\text{cat}}$  is the catalytic constant, where  $K_{\text{cat}}=V_{\max}/[E]$ .

**Table S2** Comparison of reported methods with the current one for Hx detection

Method	Linear range ( $\mu\text{M}$ )	LOD ( $\mu\text{M}$ )	Time (min)	Reference
Fluorescent assay	5-120	1.2	15	[2]
Fluorescent assay	8-2500	2.88	60	[3]
Paper-based device	10-700	3.1	30	[4]
Paper-based device	2-10	0.65	30	[5]
Electrochemical assay	20-512	9.5	>60	[6]
Electrochemical assay	5-60	2	>60	[7]
Colorimetric assay	0.5-10000	0.16	10	This work

## References

1. Gao L, Zhuang J, Nie L, Zhang J, Zhang Y, Gu N, et al. Intrinsic peroxidase-like activity of ferromagnetic nanoparticles. *Nat Nanotechnol.* 2007;2(9):577-83.
2. Zhang Z, Kwok RTK, Yu Y, Tang BZ, Ng KM. Aggregation-induced emission luminogen-based fluorescence detection of hypoxanthine: a probe for biomedical diagnosis of energy metabolism-related conditions. *J Mater Chem B.* 2018;6(28):4575-8.
3. Chen J, Lu Y, Yan F, Wu Y, Huang D, Weng Z. A fluorescent biosensor based on catalytic activity of platinum nanoparticles for freshness evaluation of aquatic products. *Food Chem.* 2020;310:125922.
4. Mustafa F, Andreescu S. Paper-Based Enzyme Biosensor for One-Step Detection of Hypoxanthine in Fresh and Degraded Fish. *ACS Sens.* 2020;5(12):4092-100.
5. Chen P-C, Li Y-C, Ma J-Y, Huang J-Y, Chen C-F, Chang H-T. Size-tunable copper nanocluster aggregates and their application in hydrogen sulfide sensing on paper-based devices. *Sci Rep.* 2016;6(1):1-9.
6. Albelda JAV, Uzunoglu A, Santos GNC, Stanciu LA. Graphene-titanium dioxide nanocomposite based hypoxanthine sensor for assessment of meat freshness. *Biosens Bioelectron.* 2017;89(Pt 1):518-24.
7. Liao L, Xing Y, Xiong X, Gan L, Hu L, Zhao F, et al. An electrochemical biosensor for hypoxanthine detection in vitreous humor: A potential tool for estimating the post-mortem interval in forensic cases. *Microchem J.* 2020;155:104760.