

# Supporting Information

## **Heterologous synthesis of Monacolin J by reconstructing its biosynthetic gene cluster in *Aspergillus niger***

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**Supplemental Table S1.** The plasmid construction strategy used in this study

Fragments assembled by overlapping PCR (Corresponding primers)			Generated plasmid via in-fusion cloning
Fragment A	Fragment B	Fragment C	
*Up <sub>pyrG</sub> -P <sub>gpdA</sub> (Up <sub>pyrG</sub> -F/R, P <sub>gpdA</sub> -F/R)	<i>lovC</i> ( <i>lovC</i> -F/R)	T <sub>tef</sub> -*Dn <sub>pyrG</sub> (T <sub>tef</sub> -1-F/R, Dn <sub>pyrG</sub> -F/R)	pM18T-Up <sub>pyrG</sub> -P <sub>gpdA</sub> - <i>lovC</i> -T <sub>tef</sub> -Dn <sub>pyrG</sub>
Up <sub>aamA</sub> -P <sub>tef</sub> (Up <sub>aamA</sub> -F/R, P <sub>tef</sub> -1-F/R)	<i>lovG</i> ( <i>lovG</i> -F/R)	T <sub>tef</sub> -pyrG <sub>An</sub> -Dn <sub>aamA</sub> (T <sub>tef</sub> -2-F/R, pyrG <sub>An</sub> -F/R, Dn <sub>aamA</sub> -F/R)	pM18T-Up <sub>aamA</sub> -P <sub>tef</sub> - <i>lovG</i> -T <sub>tef</sub> -pyrG <sub>An</sub> -Dn <sub>aamA</sub>
Up <sub>amyA</sub> -P <sub>tef</sub> (Up <sub>amyA</sub> -F/R, P <sub>tef</sub> -2-F/R)	<i>lovA</i> ( <i>lovA</i> -F/R)	T <sub>tef</sub> -Dn <sub>amyA</sub> (T <sub>tef</sub> -3-F/R, Dn <sub>amyA</sub> -F/R)	pM18T-Up <sub>amyA</sub> -P <sub>tef</sub> - <i>lovA</i> -T <sub>tef</sub> -Dn <sub>amyA</sub>

\*Up<sub>geneA</sub>: represents the upstream region of gene A.

Dn<sub>geneA</sub>: represents the downstream region of gene A.

**Supplemental Table S2.** Construction and identification primers used in this study

Primer	Sequences	Function description
<i>lovB</i> -up-F	ACTGAGAGCCTGAGCTTCATCCCCAGCATC ATTACACCTCAGCATCTAGAATGGCTCAAT CTATGTATCCTAATG	Amplification of the upstream fragment of <i>lovB</i> ORF
<i>lovB</i> -up-R	GCCGGTCCCTGCTCCAATCTCCAGAATAT CCATTGACTGATAGCGATGGGCGATCTGC GCCACCAATTCCCGGGCGTAGT	Amplification of the upstream fragment of <i>lovB</i> ORF
<i>lovB</i> -dn-F	ACTACGCCCCGGAATTGGTGGCGCAGATC GCCCATCGCTATCAGTCAATGGATATTCTG GAGATTGGAGCAG	Amplification of the downstream fragment of <i>lovB</i> ORF
<i>lovB</i> -dn-R	TCGTAAAGGCTTTTTTAAGGAAGTCATAA CGGCATAAATCGAATGTCCGCTCATGCCA GCTTCAGGGCGG	Amplification of the downstream fragment of <i>lovB</i> ORF
<i>Up<sub>pyrG</sub></i> -F	<u>ACCCGGGGATCCTCTAGAG</u> ACTGGTGGC AGGCGTCAAGT	Amplification of the upstream fragment of <i>pyrG<sub>An</sub></i>
<i>Up<sub>pyrG</sub></i> -R	<u>CTCTTGGGTCTCTCCCGTC</u> AGATGTTAAA GGGTTGGGATGG	Amplification of the upstream fragment of <i>pyrG<sub>An</sub></i>
<i>P<sub>gpdA</sub></i> -F	<u>CATCCCAACCCTTTAACATCT</u> GACGGGAG AGACCCAAGAG	Amplification of the fragment <i>P<sub>gpdA</sub></i>
<i>P<sub>gpdA</sub></i> -R	<u>ATGAATGGCTGGTCGCCCATTG</u> TTTAGATG TGTCTATGTGGCG	Amplification of the fragment <i>P<sub>gpdA</sub></i>
<i>lovC</i> -F	<u>CACATAGACACATCTAAACA</u> ATGGG CGACCAGCCATTCATT	Amplification of the whole <i>lovC</i> ORF
<i>lovC</i> -R	<u>TAGCGAAATGGATTGATTG</u> TTTACGG CCCCTCGAGCCGA	Amplification of the whole <i>lovC</i> ORF
<i>T<sub>tef</sub></i> -1-F	<u>TTCGGCTCGAGGGGCCGTAAG</u> CGGACATT CGATTATGCCGT	Amplification of the fragment <i>T<sub>tef</sub></i>
<i>T<sub>tef</sub></i> -1-R	<u>GACGTTGGTTTTCTTCTCCT</u> GTATTGGGAT GAATTTTGTATGCAC	Amplification of the fragment <i>T<sub>tef</sub></i>
<i>Dn<sub>pyrG</sub></i> -F	<u>TACAAAATTCATCCCAATAC</u> AGGAGAAG AAAACCAACGTCAC	Amplification of the downstream fragment of <i>pyrG<sub>An</sub></i>
<i>Dn<sub>pyrG</sub></i> -R	<u>CTTGCATGCCTGCAGGTCGAC</u> GATAACTC AACCGAGCGTGCCGT	Amplification of the downstream fragment of <i>pyrG<sub>An</sub></i>
<i>Up<sub>aamA</sub></i> -F	<u>ACCCGGGGATCCTCTAGAG</u> AGCGTT GTTTCTTACCCCC	Amplification of the upstream fragment of <i>Up<sub>aamA</sub></i>
<i>Up<sub>aamA</sub></i> -R	<u>GCGGTGATTCTGCTGTCTC</u> GGAATTG TCCGTCCTACCGAA	Amplification of the upstream fragment of <i>Up<sub>aamA</sub></i>

<i>P<sub>tef</sub></i> -1-F	<u>TTCGGTAGGACGGACAATTCCGAGA</u> CAGCAGAATCACCGC	Amplification of the fragment <i>P<sub>tef</sub></i>
<i>P<sub>tef</sub></i> -1-R	<u>GGAGATGCTTGGTAACGCATGTGAA</u> GGTTGTGTTATGTTTTGTG	Amplification of the fragment <i>P<sub>tef</sub></i>
<i>lovG</i> -F	<u>AAACATAACACAACCTTCACATGCG</u> TTACCAAGCATCTCCA	Amplification of the whole <i>lovG</i> ORF
<i>lovG</i> -R	<u>GGCATAAATCGAATGTCCGCCTACTC</u> CAATGTCTGGGCCG	Amplification of the whole <i>lovG</i> ORF
<i>T<sub>tef</sub></i> -2-F	<u>CGGCCCAGACATTGGAGTAGGCGGA</u> CATTGATTTATGCCGTT	Amplification of the fragment <i>T<sub>tef</sub></i>
<i>T<sub>tef</sub></i> -2-R	<u>CGCGTTCTCGAGGAAGTTGCGTATT</u> GGGATGAATTTTGTATGCA	Amplification of the fragment <i>T<sub>tef</sub></i>
<i>pyrG<sub>An</sub></i> -F	<u>TACAAAATTCATCCCAATACGCAACT</u> TCCTCGAGAACGCG	Amplification of the fragment <i>pyrG<sub>An</sub></i>
<i>pyrG<sub>An</sub></i> -R	<u>CCAACGTACTTGGTCACCCGCCCTTT</u> TAGTCAATACCGTTAC	Amplification of the fragment <i>pyrG<sub>An</sub></i>
<i>D<sub>naamA</sub></i> -F	<u>AACGGTATTGACTAAAAGGGCGGGT</u> GACCAAGTACGTTG	Amplification of the downstream fragment of <i>aamA</i>
<i>D<sub>naamA</sub></i> -R	<u>ATGCCTGCAGGTCGACGATAGCCCT</u> GGTACAGTTAGCG	Amplification of the downstream fragment of <i>aamA</i>
<i>Up<sub>amyA</sub></i> -F	<u>ACCCGGGGGATCCTCTAGAGAGCAAT</u> CCATTTATTTCTTCTC	Amplification of the upstream fragment of <i>Up<sub>amyA</sub></i>
<i>Up<sub>amyA</sub></i> -R	<u>GCGGTGATTCTGCTGTCTCGGAGCG</u> CCTTCAAGTCATCT	Amplification of the upstream fragment of <i>Up<sub>amyA</sub></i>
<i>P<sub>tef</sub></i> -2-F	<u>CAGATGACTTGAAGGCGCTCCGAGA</u> CAGCAGAATCACCGC	Amplification of the fragment <i>P<sub>tef</sub></i>
<i>P<sub>tef</sub></i> -2-R	<u>TGAGCGCGTCGACAGTCATGGTGAA</u> GGTTGTGTTATGTTTTGTG	Amplification of the fragment <i>P<sub>tef</sub></i>
<i>lovA</i> -F	<u>AACATAACACAACCTTCACCATGAC</u> TGTCGACGCGCTCACA	Amplification of the whole <i>lovA</i> ORF
<i>lovA</i> -R	<u>GGCATAAATCGAATGTCCGCCTATAG</u> TGAACCAGGAAGGCGG	Amplification of the whole <i>lovA</i> ORF
<i>T<sub>tef</sub></i> -3-F	<u>GCCTTCCTGGTTCACTATAGGCGGAC</u> ATTCGATTTATGCCGTT	Amplification of the fragment <i>T<sub>tef</sub></i>
<i>T<sub>tef</sub></i> -3-R	<u>CGACCATAAGATACATCCCCGTATTG</u> GGATGAATTTTGTATGCA	Amplification of the fragment <i>T<sub>tef</sub></i>
<i>D<sub>namyA</sub></i> -F	<u>TACAAAATTCATCCCAATACGGGGAT</u> GTATCTTATGGTTCG	Amplification of the downstream fragment of <i>amyA</i>
<i>D<sub>namyA</sub></i> -R	<u>ATGCCTGCAGGTCGACGATACGGAT</u>	Amplification of the

	CACCGTCGAGCAC	downstream fragment of <i>amyA</i>
<i>LovB</i> -YZ1-F	CGTACCTGGCGACCTATGACTA	Yeast transformation verification primers
<i>LovB</i> -YZ1-R	GTAGGTAGCCTTGATGAGTGCC	Yeast transformation verification primers
<i>LovB</i> -YZ2-F	AGGGAGACTACTTGAGCGGTGA	Yeast transformation verification primers
<i>LovB</i> -YZ2-R	TGAACACTCTCAGGCACAAACC	Yeast transformation verification primers
<i>LovB</i> -YZ3-F	ACTGTCTCTCGGGCAAGAATAC	Yeast transformation verification primers
<i>LovB</i> -YZ3-R	TCAACTCTCCCCTCTGTGCTAA	Yeast transformation verification primers
<i>LovB</i> -JD-1-F	GAGGATTGCCTGAACATTGACA	Location of <i>lovB</i> in the <i>glaA</i> (An03g06550) locus
<i>LovB</i> -JD-1-R	ACTCACGCCTCTGGCATAGC	Location of <i>lovB</i> in the <i>glaA</i> (An03g06550) locus
<i>LovB</i> -JD-2-F	TAACATCTAAACTCTTCTCCATCG	Location of <i>lovB</i> in the <i>glaA</i> (An03g06550) locus
<i>LovB</i> -JD-2-R	ACGGTCTGGTTCTGTAGTCC	Location of <i>lovB</i> in the <i>glaA</i> (An03g06550) locus
<i>LovC</i> -JD-1-F	CGGAAAGTGGAACGATTGGGGG	Location of <i>lovC</i> in the <i>pyrG<sub>An</sub></i> locus
<i>LovC</i> -JD-1-R	TGGCAAAGGCAGCCCAAGCAAC	Location of <i>lovC</i> in the <i>pyrG<sub>An</sub></i> locus
<i>LovC</i> -JD-2-F	GCAGTGAGGAAGAGCGGCAG	Location of <i>lovC</i> in the <i>pyrG<sub>An</sub></i> locus
<i>LovC</i> -JD-2-R	ATTGTCGATCTGCCCACTTTCT	Location of <i>lovC</i> in the <i>pyrG<sub>An</sub></i> locus
<i>LovG</i> -JD-1-F	CGTCCTTCGGTCGGAATGATG	Location of <i>lovG</i> in the <i>aamA</i> (An11g03340) locus
<i>LovG</i> -JD-1-R	CGCTCACGGAGGGTCCATTAT	Location of <i>lovG</i> in the <i>aamA</i> (An11g03340) locus
<i>LovG</i> -JD-2-F	GGAACGAAGTGATGAAGAACCA	Location of <i>lovG</i> in the <i>aamA</i> (An11g03340) locus
<i>LovG</i> -JD-2-R	CATACGAGGCGAAACGGGGA	Location of <i>LovG</i> in the <i>aamA</i> (An11g03340) locus
<i>LovA</i> -JD-1-F	ATATCAGGCCGCGCACGAAAG	Location of <i>lovA</i> in the <i>amyA</i> (An05g02100) locus
<i>LovA</i> -JD-1-R	CCTCATGGTCGTCAGTTCCC	Location of <i>lovA</i> in the <i>amyA</i> (An05g02100) locus
<i>LovA</i> -JD-2-F	TTCGGTTGGCATCCACGGGC	Location of <i>lovA</i> in the <i>amyA</i> (An05g02100) locus

<i>LovA</i> -JD-2-R	TTAACAGTTTGC GGGAGATGCA	Location of <i>lovA</i> in the <i>amyA</i> (An05g02100) locus
RT- <i>lovB</i> -F	GCGTCCAACCGCATCTCG	Semi-quantitative primers for <i>lovB</i> gene
RT- <i>lovB</i> -R	AGATTTCGCACCCGCAGCAA	Semi-quantitative primers for <i>lovB</i> gene
RT- <i>lovC</i> -F	ACCGCTGGATTGGCGATGA	Semi-quantitative primers for <i>lovC</i> gene
RT- <i>lovC</i> -R	CGATAGTCAAAGACCTCCTC	Semi-quantitative primers for <i>lovC</i> gene
RT- <i>lovG</i> -F	CGACAACAATCATAATGGACCC	Semi-quantitative primers for <i>lovG</i> gene
RT- <i>lovG</i> -R	CTTGCCTCGTCTTGATACCAG	Semi-quantitative primers for <i>lovG</i> gene
RT- <i>lovA</i> -F	CTGGTTCTCGCAAAATGATAAG	Semi-quantitative primers for <i>lovA</i> gene
RT- <i>lovA</i> -R	GTGACTTCCTTGAATCCATCG	Semi-quantitative primers for <i>lovA</i> gene
RT- <i>gpdA</i> -F	CTCTGCTCCTTCCGCTGAT	Semi-quantitative primers of internal reference gene <i>gpdA</i> (An16g01830)
RT- <i>gpdA</i> -R	ACCCTCAACGATGCCGAAC	Semi-quantitative primers of internal reference gene <i>gpdA</i> (An16g01830)

**Supplemental Table S3.** CRISPR/Cas9 plasmids used in this study

Plasmids	Description/derivation	Reference
PFC332- <i>pyrG<sub>An</sub></i> - <i>hygB</i>	PFC332 derivate with gRNA scaffold of <i>pyrG<sub>An</sub></i> under the control of promoter of <i>A. oryzae</i> U6 and <i>A. fumigatus</i> U6-2	This work
PFC332- <i>aamA</i> - <i>hygB</i>	PFC332 derivate with gRNA scaffold of <i>aamA</i> under the control of promoter of <i>A. oryzae</i> U6 and <i>A. fumigatus</i> U6-2	This work

PFC332- <i>aamA</i> - <i>amyA-hygB</i>	PFC332 derivate with gRNA scaffold of <i>aamA</i> and <i>amyA</i> under the control of promoter of <i>A. oryzae</i> U6 and <i>A. fumigatus</i> U6-2	This work
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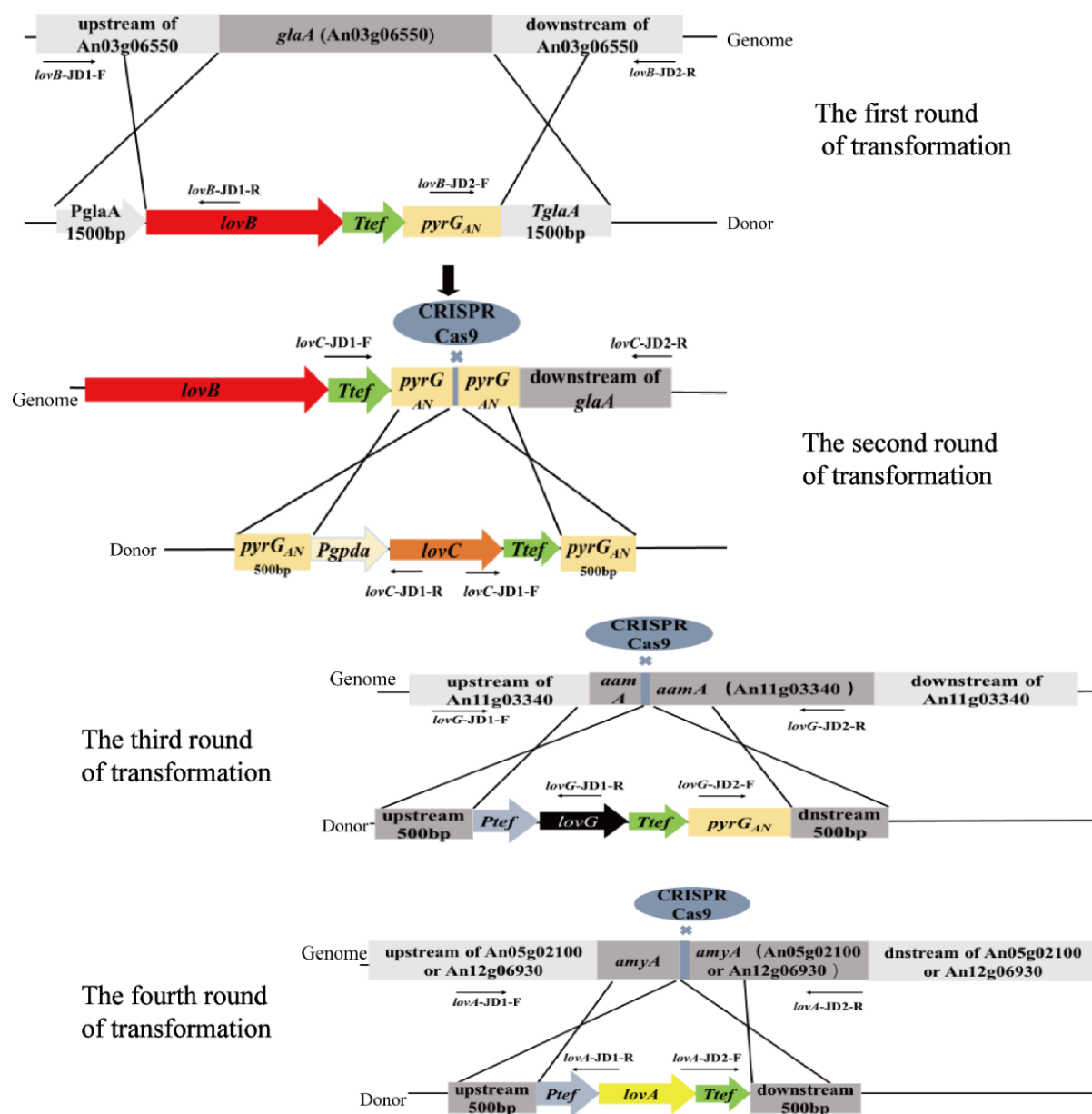


Figure S1. Schematic gene integration in *A. niger*.

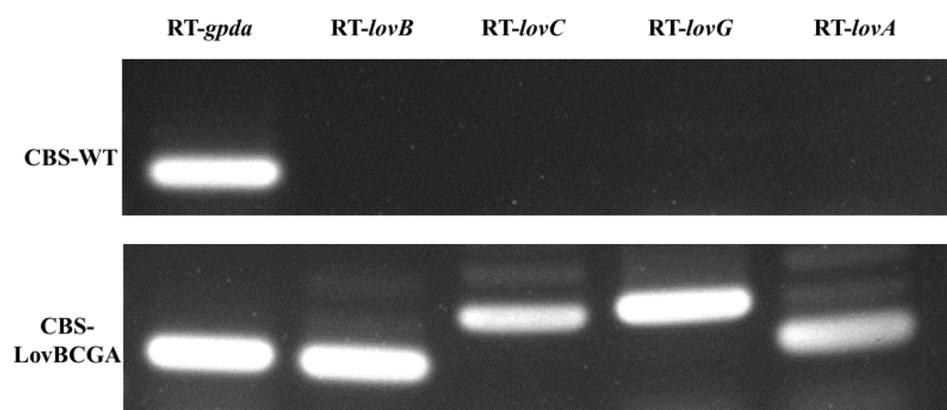
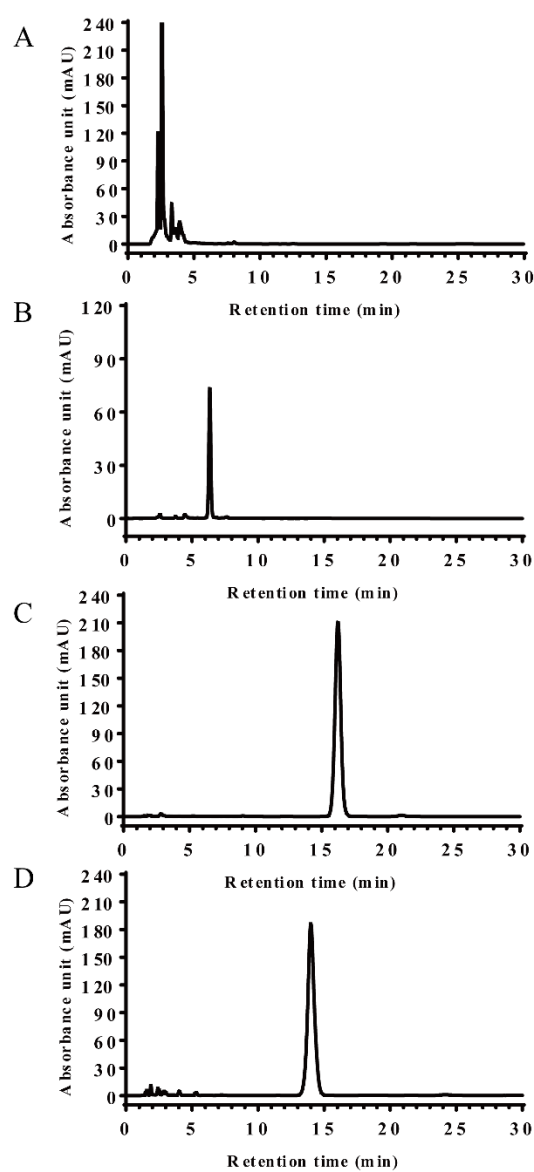
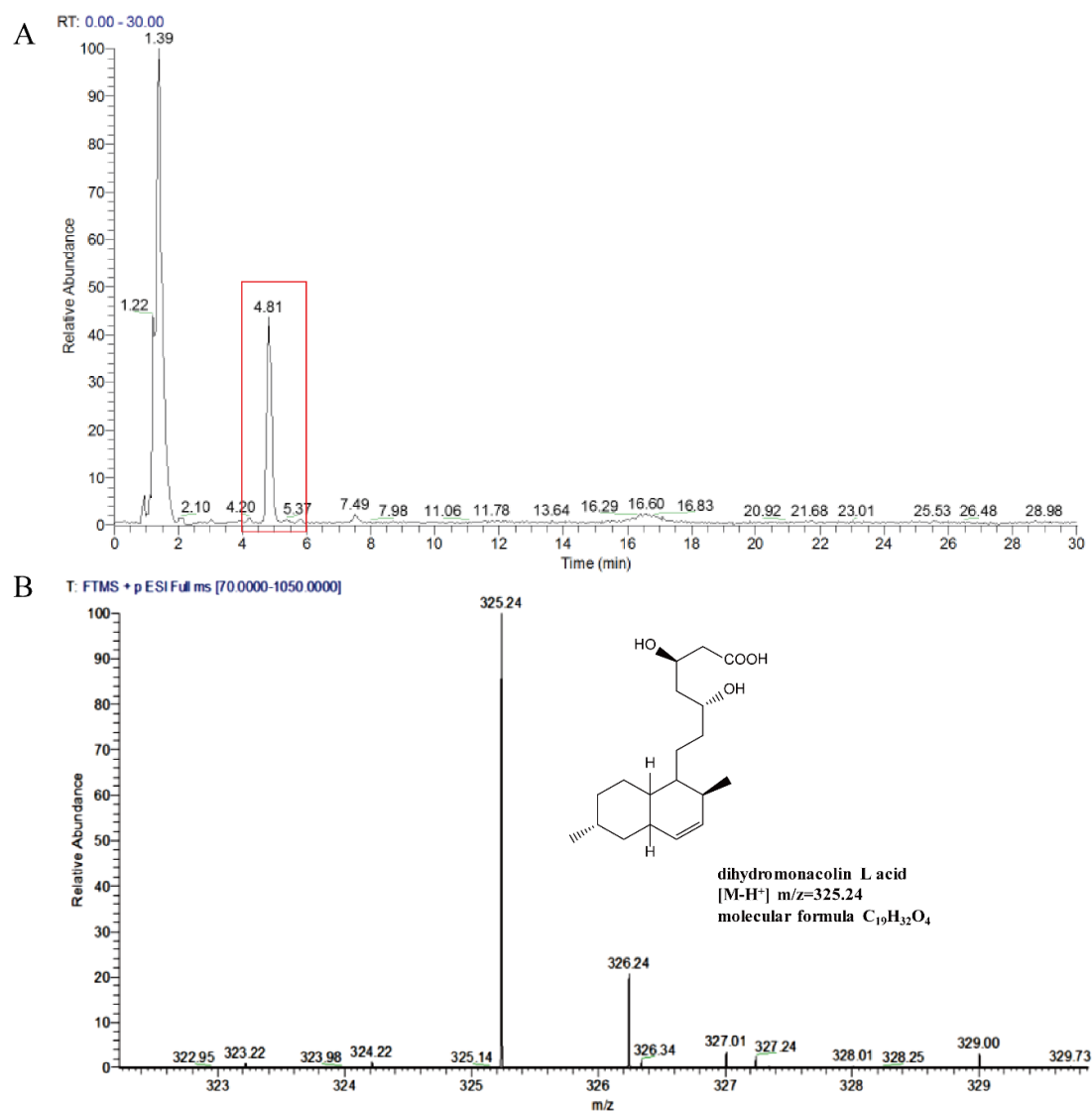


Figure S2. RT-PCR analysis of the expression of MJ genes in *A. niger* CBS-WT and CBS-LovBCGA strain. *gpda* was used as the internal reference gene.





**Figure S3.** HPLC identification of *A. niger* strain fermentation product and standard products at 237 nm wavelength. (A) *A. niger* strain CBS513.88( $\Delta kusA$ ,  $\Delta pyrG$ ) (B) Monacolin J acid standard, retention time 6.473 min (C) Lovastatin lactone standard, retention time 16.238 min (D) Lovastatin acid standard, retention time 13.999 min.



**Figure S4.** LC-MS identification DML acid of *A. niger* transformed strain CBS-LovBCG fermentation product (A) DML acid, retention time 4.81 min with the red square. (B) LC-MS identification of DML acid from *A. niger* strain CBS-LovBCG.

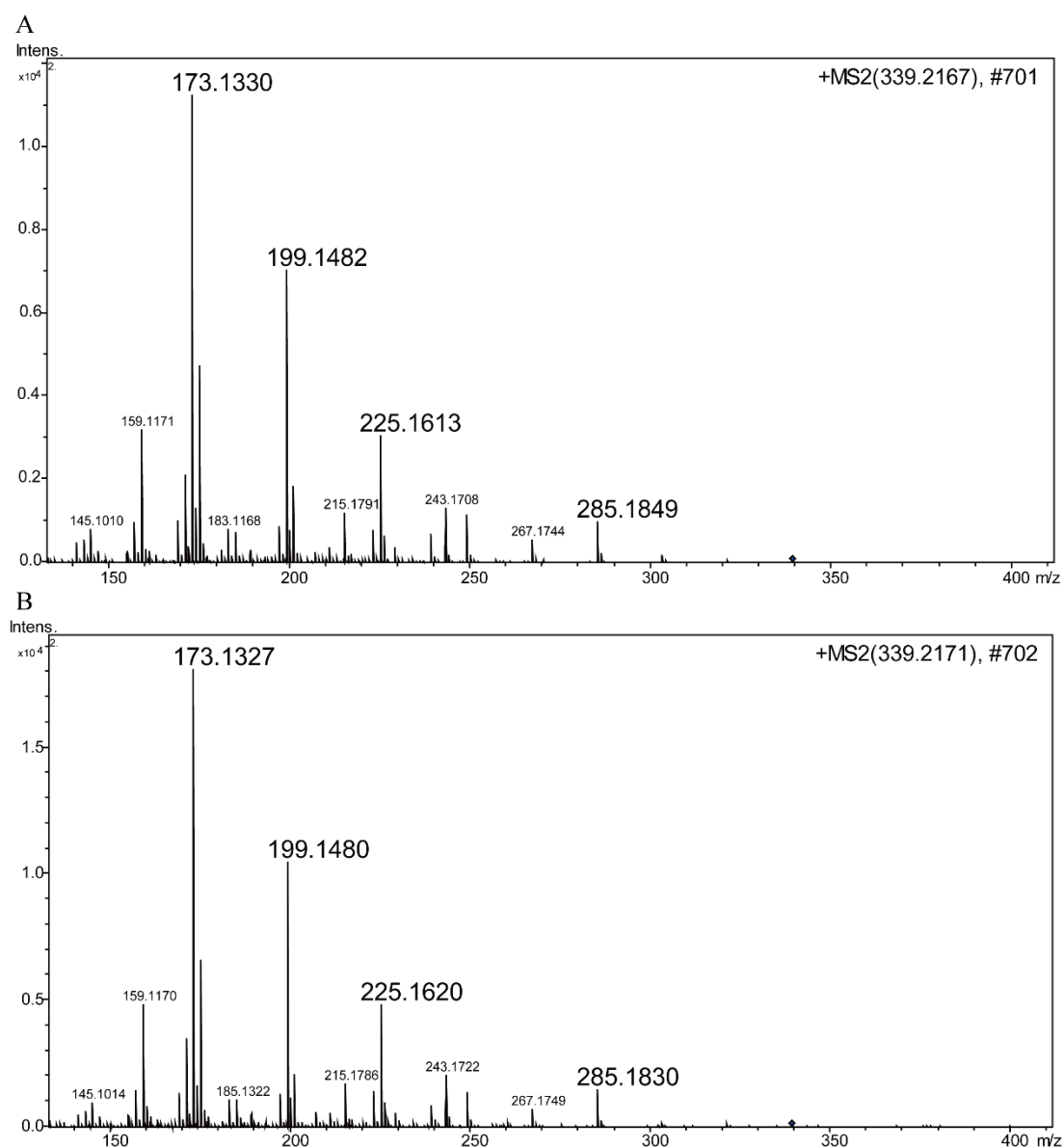


Figure S5. Tandem LC-MS/MS analysis of the mass spectra fragmentation of MJ standard (A) and MJ produced by *A. niger* strain CBS-LovBCGA (B).

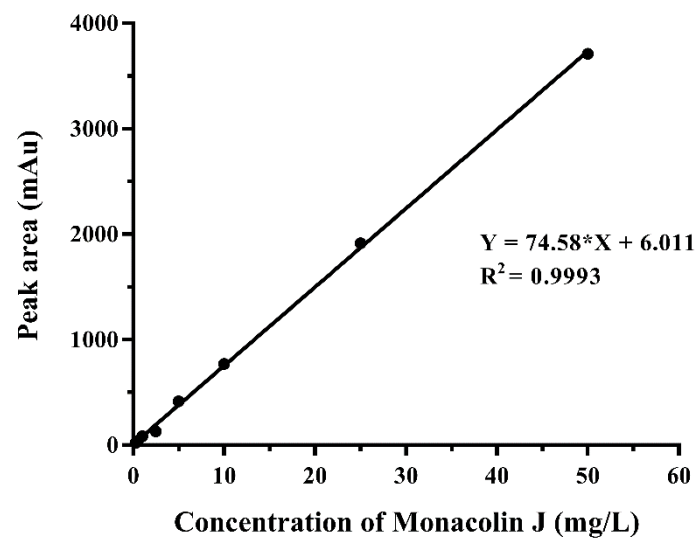


Figure S6. Calibration curves of Monacolin J developed by HPLC

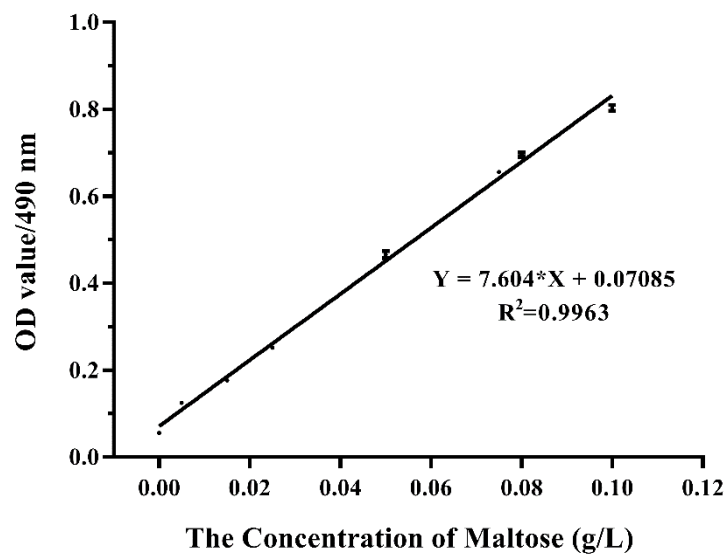


Figure S7. Calibration curves of the concentration of Maltose