



Testing Alpha-1 Antitrypsin Deficiency in Black Populations

Pascale Lafortune¹, Kanza Zahid¹, Magdalena Ploszaj¹, Emilio Awadalla¹, Tomás P. Carroll^{2,3,*} and Patrick Geraghty^{1,*}

- ¹ Department of Medicine, State University of New York Downstate Medical Center, Brooklyn, NY 11203, USA; pascale.lafortune@downstate.edu (P.L.); kanza.s.zahid@gmail.com (K.Z.);
- magdalena.ploszaj@nyulangone.org (M.P.); emilio.awadalla@bath.edu (E.A.)
- ² Irish Centre for Genetic Lung Disease, Royal College of Surgeons in Ireland, D02 YN77 Dublin, Ireland ³ Alpha 1 Foundation Ireland, Payal College of Surgeons in Ireland, D02 YN77 Dublin, Ireland
 - Alpha-1 Foundation Ireland, Royal College of Surgeons in Ireland, D02 YN77 Dublin, Ireland
- * Correspondence: tcarroll@rcsi.ie (T.P.C.); patrick.geraghty@downstate.edu (P.G.); Tel.: +353-1-809-3876 (T.P.C.); +001-718-270-3141 (P.G.)

Highlights:

What are the main findings?

- Alpha-1 antitrypsin deficiency is extensively studied in populations of European ancestry but other ethnic populations also carry *SERPINA1* mutations that may be harmful to these populations.
- The majority of studies undertaken in non-European populations screen for *SERPINA1* mutations in small subject numbers and not from the general population.

What is the implication of the main finding?

- Insufficient alpha-1 antitrypsin deficiency testing is performed in Black populations that already
 experience poor health outcomes.
- Diagnosis of severe *SERPINA1* mutations and counseling assists patients in their health education and a diagnosis of alpha-1 antitrypsin deficiency is a stronger motivator to quit smoking, improves the frequency of regular health checks, and lung and liver scans in patients.

Abstract: Alpha-1 antitrypsin (AAT) deficiency (AATD) is an under-recognized hereditary disorder and a significant cause of chronic obstructive pulmonary disease (COPD), a disease that contributes to global mortality. AAT is encoded by the *SERPINA1* gene, and severe mutation variants of this gene increase the risk of developing COPD. AATD is more frequently screened for in non-Hispanic White populations. However, AATD is also observed in other ethnic groups and very few studies have documented the mutation frequency in these other ethnic populations. Here, we review the current literature on AATD and allele frequency primarily in Black populations and discuss the possible clinical outcomes of low screening rates in a population that experiences poor health outcomes and whether the low frequency of AATD is related to a lack of screening in this population or a truly low frequency of mutations causing AATD. This review also outlines the harmful *SERPINA1* variants, the current epidemiology knowledge of AATD, health inequity in Black populations, AATD prevalence in Black populations, the clinical implications of low screening of AATD in this population, and the possible dangers of not diagnosing or treating AATD.

Keywords: alpha-1 antitrypsin deficiency; chronic obstructive pulmonary disease; Black populations; genetic screening

1. Introduction

Black populations experience significant health inequities that lead to poor outcomes, such as higher rates of chronic diseases, maternal and infant mortality, infectious diseases, mental health issues, and lower life expectancy [1]. Some of these outcomes can be addressed by improving access to healthcare, addressing social determinants of health, and



Citation: Lafortune, P.; Zahid, K.; Ploszaj, M.; Awadalla, E.; Carroll, T.P.; Geraghty, P. Testing Alpha-1 Antitrypsin Deficiency in Black Populations. *Adv. Respir. Med.* **2024**, *92*, 1–12. https://doi.org/10.3390/ arm92010001

Academic Editor: Monika Franczuk

Received: 9 November 2023 Revised: 11 December 2023 Accepted: 15 December 2023 Published: 19 December 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). tackling discrimination and bias in healthcare and within our society. However, this excess burden is well documented and is not completely explained by socioeconomic status or access to care [2]. With the recent COVID-19 pandemic, one study reported that 27% of associated deaths within the USA were Black patients, while Black patients account for only 12.5% of the population in the USA [3]. In 2020, chronic liver disease was the ninth leading cause of death for African Americans, ages 45–64 years old [4], and in a single-center study, African American patients with antineutrophilic cytoplasmic antibody (ANCA)associated vasculitis were diagnosed at a younger age than Caucasian patients [5]. Studies looking at ethnic differences in COPD diagnosis also demonstrate the underdiagnosis of the disease in Black populations [6]. In 2003, the American Thoracic Society (ATS) and European Respiratory Society (ERS) recommended that all individuals with a diagnosis of chronic obstructive pulmonary disease (COPD), refractory asthma, unexplained chronic liver disease, or panniculitis, irrespective of age or ethnicity, should be tested for alpha-1 antitrypsin deficiency (AATD) [7]. Since AAT is an acute phase protein, it is recommended to perform iso-electric focusing testing for the common alleles of AATD in these individuals. While AATD continues to be underdiagnosed in all populations, the frequency and nature of harmful serine protein inhibitor-A1 (SERPINA1 gene) mutations that occur in Black populations are unclear. Since Black populations experience significant health inequities, one could expect similar disparities in AATD diagnosis and treatment. In this mini-review, we want to outline the current literature on AATD and SERPINA1 mutation frequency in Black populations and discuss the consequences of low screening rates in a population that already experiences disproportionately poor health outcomes compared to other ethnic groups. Equally, we discuss whether the reported low prevalence of AATD is related to a lack of screening in this population or simply a low frequency of SERPINA1 mutations causing AATD.

2. AATD: A Historical and Biological Perspective

AAT was first characterized as a protease inhibitor, with loss in this activity associated with lung disease by Swedish researchers in 1963 [8]. AAT is a 52 kDa glycoprotein produced mainly by hepatocytes and secreted into the blood. AAT primarily inhibits neutrophil elastase, and in the absence of AAT, unregulated active proteases cleave the structural proteins of the lungs. AAT also has a plethora of anti-inflammatory and immunemodulatory properties. AAT deficiency is the most common genetic cause of COPD. Without sufficient concentrations of biologically active AAT, tissue destruction and airspace enlargement can occur, leading to progressive emphysema. This process is accelerated by exposure to cigarette smoke or other environmental factors. COPD development is common in AATD, especially in combination with cigarette smoke exposure. AATD patients with a smoking history typically present with emphysema on a chest computed tomography and with obstruction determined by spirometry. Chronic bronchitis or asthma is also observed but less frequently [9]. However, AATD subjects who do not smoke tend to get radiographic emphysema after 60 years old [10]. Asthma may be more prevalent in AATD individuals, as wheezing and dyspnea are some of the first pulmonary symptoms in AATD [11]. Bronchiectasis is also increased in AATD [12], and this is associated with atypical mycobacterial infection [13]. Therefore, AAT testing is recommended for subjects with emphysema, COPD, bronchiectasis, chronic bronchitis, and asthma where spirometry fails to return to normal upon the treatment of asthma [7]. Approximately 2–3% of patients diagnosed with COPD will be AATD [14]. AATD is also a significant cause of liver disease through the polymerization and accumulation of misfolded Z AAT protein within hepatocytes and is a common cause of liver transplantation. Since lung disease in AATD is almost indistinguishable from nonhereditary lung disease, AATD is an under-recognized hereditary disorder and screening typically occurs after disease establishment. AATD is often suspected only following the diagnosis of early-onset obstructive lung disease in individuals with minimal or no cigarette consumption or panacinar emphysema affecting mainly the lower lobes.

AAT is encoded by the SERPINA1 gene. The most common mutation known to cause severe AAT deficiency is Z (p.Glu342Lys, rs28929474). Individuals homozygous or heterozygous for the Z mutation are at increased risk of developing COPD, although heterozygotes require a second insult such as smoking before developing COPD [4,15-17]. Classically, severe AATD is more frequently screened for in non-Hispanic White populations. Therefore, AATD is primarily documented in Northern, Western, and Central Europe. However, AATD is also observed in other ethnic groups [18]. This is important as COPD is the fourth leading cause of death in the United States of America, with over 16 million (6.6%) people reporting a diagnosis of COPD [19]. In 2019, COPD was the third leading cause of death worldwide, causing 3.23 million deaths [20]. Self-reported COPD is estimated to be 6.1% in Black populations [21]. Though COPD is typically diagnosed in patients who are current and or former smokers, one in four people who are nonsmokers develop COPD [22]. In addition to environmental factors, individuals who have a genetic predisposition, such as AATD, can develop COPD. A recent study demonstrated that the primary factors for favoring AAT testing were whether the patient was of the White race and had concomitant COPD and liver disease [23]. This same study also observed that increasing age, being non-White, current tobacco use, and being a male with COPD reduced the odds of AAT testing being performed [23]. Therefore, there is bias in performing AAT testing and screening in certain populations.

3. AATD Mutations

SERPINA1 is a pleiomorphic gene with alleles inherited in an autosomal co-dominant fashion. The most frequent clinically significant alleles are Z and S, with M being the normal/non-mutated allele. It is important to note that the S allele (p.Glu264Val, rs17580) is less polymerogenic and causes mild serum deficiency. Thus, the SZ genotype results in a phenotype similar to the MZ phenotype and is deemed less severe than ZZ [24]. There are a few published unbiased studies that address the allele frequencies in the general population. Most studies perform AAT testing on cohorts with a high number of COPD subjects or other subjects with already diagnosed pulmonary diseases, which do not reflect the true numbers in the general population. In a recent study from the Canary Islands (Spain), the estimated frequency of S and Z alleles in the general population was 8.2% and 2.1%, respectively [25]. An Irish study found an estimated prevalence of 1/25 (4%) for the Z allele and 1/10 (10%) for the S allele in DNA collected from 1100 individuals randomly sampled from the general population [26]. In a large genetic testing study, the allele frequency for the Z and S variants among 195,014 study participants was 6.5% and 15.1%, respectively. Notably, this cohort included 1443 African Americans [27]. Unfortunately, allele frequencies in the African American group were not reported. Finally, the Genome Aggregation Database (gnomAD), which is a shared aggregate exome and genome sequencing database from a variety of largescale sequencing projects [28], is a useful tool for looking at SERPINA1 variants in multiple ethnic populations, including African and African American populations. These datasets may represent a better overall frequency for SERPINA1 variants in multiple populations.

Recent advances in sequencing have identified large numbers of harmful new *SER*-*PINA1* variants, with over 200 identified to date [29]. There are many rare *SERPINA1* variants that could be population specific [30] but require further investigation. Table 1 outlines some of these harmful *SERPINA1* variants. All these variants are predicted to be observed in Black populations and require further study. A recent study that compiled a comprehensive database of *SERPINA1* coding mutations reported that 2.59% of an African cohort carry harmful SERPINA1 mutations [31].

SERPINA1 Variant	Molecular Basis	SNP Number	Cellular Effect	Disease Association	Reference	Observed in Non-European Populations?
S	p.Glu264Val	rs17580	Polymerization, impaired secretion, reduced antiprotease activity	Lung & liver if inherited with other severe AATD variant(s) (e.g., Z)	Lomas et al., 1999 [32]	Yes
Z	p.Glu342Lys	rs28929474	Polymerization, impaired secretion, reduced antiprotease activity	Lung & liver	Laurell & Eriksson, 1963 [33]	Yes
I	p.Arg39Cys	rs28931570	Polymerization, impaired secretion, reduced antiprotease activity	Lung & liver	Lomas et al., 1999 [32]	Yes
F	p.Arg223Cys	rs28929470	Reduced antiprotease activity	Lung	Fagerhol & Tenfjord, 1968 [34]	Yes
M _{malton}	p.Phe52del	rs775982338	Polymerization, impaired secretion, reduced antiprotease activity	Lung and liver	Curiel et al., 1989 [35]	Yes
M _{wurzburg}	p.Pro369Ser	rs61761869	Intracellular accumulation & polymerization	Lung and liver	Poller et al., 1999 [36]	Yes
M _{heerlen}	p.Pro369Leu	rs199422209	Block in production	Lung	Poller et al., 1999 [36]	Yes
S _{iiyama}	p.Ser53Phe	rs55819880	Polymerization	Lung and liver	Lomas et al., 1993 [37]	Yes
Null (Q0)	Premature termination codon	N/A	Family of mutations that produce no detectable AAT	Lung	Talamo et al., 1973 [38]	Yes

|--|

Single nucleotide polymorphism (SNP), not available (N/A).

Severe AATD affects about 1 in 1500 to 3500 individuals with European ancestry. While several studies do show lower frequencies in other ethnic groups, the Z and S alleles are documented to be detected in countries in the Caribbean, North and South America, Asia, and Africa [39]. Historically, detection has focused on ZZ AATD, but it is now accepted that Z heterozygotes are also at risk of COPD. A study in Ireland found that MZ smokers were at a higher risk of developing COPD when compared to MM siblings who smoked [15]. The finding of increased risk and severity of COPD in Z heterozygotes has been replicated in larger, multi-ethnic cohorts [4,16,17]. One of these studies showed that African American MZ subjects had lower lung function, observed with low FEV₁ percent predicted and FEV₁/FVC compared to African American MM subjects [16]. It is clear that MZ smokers are at risk for lung function changes in both White and Black populations.

Regulation of the SERPINA1 Gene

The regulation of the *SERPINA1* gene is quite complex. An epigenome-wide association study (EWAS) in 2012 was performed on peripheral blood mononuclear cells from adults who were smokers and suggested a positive correlation between hypomethylation at two CpGs in the *SERPINA1* gene promoter and COPD risk [40]. However, another study performed on samples from smoke-exposed children and adults observed no correlation between *SERPINA1* gene methylation and lung function [41]. A recent study demonstrated that the *SERPINA1* gene promoter is differentially methylated in peripheral blood mononuclear cells from healthy subjects [42]. However, further studies are needed to assess the direct link between AAT circulating levels and *SERPINA1* promoter methylation in blood cells. There are 11 known *SERPINA* mRNA isoforms, which are generated through alternative splicing involving the 5'-UTR of the pre-mRNA [43,44]. Mutations in *SERPINA1* 5'-UTR non-coding regions can lead to altered translation, as observed in a large-scale clinical study looking at AAT serum levels in patients [17]. One of these 5'-UTR (NM_000295.4) can reduce AAT translation [45]. This may be another means of observing altered AATD levels in different populations.

4. Health Outcomes in Black Populations

The Black population in the United States of America has worse health outcomes in comparison to other ethnic groups [1]. A significant number of physicians are unaware of the current guidelines for screening for AATD in patients and may not be aware of possible treatment available for this form of COPD [46]. Therefore, this may further lead to a reduced urgency of making a diagnosis of AATD. A recent whole genome sequence (WGS) study examined gene variants and lung function and COPD and identified two common variant signals unique to lung function in African Americans [47]. There is likely heterogeneity in genetic effects when investigating race/ethnicity and lung function. Therefore, we cannot presume genetic variants associated with lung function identified by GWAS may be applicable to all populations. Unique variant signaling associated with certain ethnic populations requires further investigation.

AATD Prevalence in Black Populations

Several studies estimate the global frequency of AATD and *SERPINA1* variants in different populations worldwide [48]. Studies looking at 94 countries encompassing 75% of the global population estimated that 173,430 individuals possess the ZZ genotype and 1,011,069 the SZ genotype [49]. Importantly, a recent study predicted that more than 35 million people in 74 countries possess the MZ genotype [50].

A study conducted comparing Black and White populations with emphysema demonstrated that Black subjects had a similar degree of lung impairment compared with Whites but developed emphysema younger despite smoking less [51]. As seen in Table 2, consisting of studies reporting Black population screening, only a small number of studies report actual numbers detected during screening, and AAT mutations are detected in this population. Some studies do state that non-White subjects were tested but do not provide screening data based on race [52,53]. Estimated frequencies exist in Z and S alleles in Caribbean and African countries; the most significant, in Cuba, Dominican Republic, Puerto Rico, Nigeria, Somalia, Angola, and Namibia, are reported [50]. It is important to note that many estimates are based on studies from the 1970s and 1980s in small cohorts and not necessarily the general population. This could warrant further AAT screening in larger Black cohorts with newer diagnostic techniques.

Chard and	.	Study Population	AAT Genotype					
Study	Location	Number (N) MM MS MZ S		SS	SZ	ZZ		
Spinola et al. [54]	Cape Verde Islands	202	191	7	1	3	0	0
Foreman et al. [16]	USA; COPDGene cohort ^a	2803	2731	49	22	0	1	0
Miskoff et al. [55]	Neptune Township, New Jersey, USA	18	16	1	1	0	0	0
Ashenhurst et al. [27]	23andMe customers	1443	N/A	N/A	N/A	N/A	N/A	N/A
Ortega et al. [17]	USA; SPIROMICS cohort	385 ^b	N/A	N/A	N/A	N/A	N/A	N/A

Table 2. Studies reporting AATD screening in Black populations.

Table 2. Cont.

Chu la	.	Study Population	AAT Genotype						
Study	Location	Number (N)	MM	MS	MZ	SS	SZ	ZZ	
Denden et al. [56]	Tunisia; obstructive lung disease cohort	120	119	0	1	0	0	0	
Webb et al. [57]	Rochester, Monroe County, NY, USA	53 c	N/A	N/A	N/A	N/A	N/A	N/A	
Young et al. [58]	Washington, DC, USA	94 ^d	N/A	N/A	2	N/A	1	1	
Pierce et al. [59]	St. Louis, MO, USA	204	196	4	2	N/A	N/A	N/A	
Massi et al. [60]	Mogadishu, Somalia	347	333	9	1	0	1	3	
Vandeville et al. [61]	Zaire	132 ^e	124	0	0	0	0	0	
Welch et al. [62]	Gambia, West Africa	701 ^f	700	0	0	0	0	0	
Pascali et al. [63]	Congo, West Africa	278	243	35	0	0	0	0	
Chaabani et al. [64]	Tunisia	310	260	50	0	0	0	0	
Giacopuzzi et al. [31]	Multiple public databases	5203 ^g	N/A	81	35	N/A	116	N/A	
Lieberman et al. [65]	High school students from Long Beach, CA, USA	186 ^h	182	3	0	0	0	0	

^a Denotes studies with smokers with and without COPD only, i.e., not the general population. Data only for MM and MZ groups; ^b Study looked at missense and frameshift exonic *SERPINA1* variants and identified four variants unique to African Americans. Exact numbers not outlined in the manuscript for African American data breakdown; ^c Study reported that 3 of the 53 African Americans were not MM. However, no additional information given; ^d Study consisted of 559 African American ambulatory and hospitalized patients with high-risk cardiopulmonary disorders. However, AAT screening is only outlined in 94 COPD-positive patients. Other data are combined with data from 115 control Caucasian patients; ^e Other mutation phenotypes detected: LM = 7, MV = 1; ^f PiGAM variant detected, n = 1; ^g 2.59% of African cohort carry harmful *SERPINA1* mutations, ^h One subject was EM phenotype. N/A, not available or data not detailed in manuscript.

Finally, one must also consider that spirometry reference values differ by race/ethnicity, which could result in the underestimation of COPD in Black populations [66]. Applying spirometry reference equations used for Caucasian populations may produce normal lung function values (% predicted) in Black populations [67]. Equally, the smoking habits between populations differ, with cigarette products containing menthol and other flavorings frequently being targeted at Black smokers and vaping device users [68]. Equally, there is some evidence to suggest that racial/ethnic minority populations and younger smokers find it harder to quit menthol versus nonmenthol cigarettes [69].

5. Screening and Diagnosis of AATD

A new ERS statement on AATD outlines an extended algorithm for family screening of individuals diagnosed with severe AATD, including their close relatives and spouses [70]. Even with guidelines in place, there are still significant delays between the appearance of symptoms and the correct diagnosis of AATD. The recommended approach to testing for AATD is to first measure plasma or serum AAT levels. However, it should be noted that AAT levels have weak intra-individual reproducibility due to the acute phase nature of AAT [71]. This could result in a missed AATD diagnosis. Thus, CRP should be ordered in combination with AAT to rule out falsely elevated AAT levels due to illness or inflammation. If AAT is abnormally low, further testing should be performed by either AAT phenotyping with isoelectric focusing or AAT mutation-specific genotyping [72]. The gold standard to detect AATD is DNA sequencing, especially for rarer variants extended molecular techniques are required, such as whole exome sequencing [73]. A recent study in Greece observed several new rare variants by sequencing and these variants appear to be pathogenic as they were detected in patients with early emphysema and lower than normal AAT levels [74]. Non-coding DNA may be an important area to be assessed [75]. For example, the integrative deep sequencing of SERPINA1 identified a 5' untranslated region

insertion (rs568223361) in African Americans that is associated with lower AAT levels and an increased risk of small airway disease [17]. Commercially available direct-to-consumer genetic tests have also allowed people to explore the possibilities of genetic screening. However, one needs to be cautious with over-the-counter genetic testing as this results in the customer paying for the service, and these tests are not federally regulated in regard to quality control and provide no counseling service upon the identification of possible genetic diseases. A recent study using data from the U.S. Bronchiectasis Research Registry found that non-Hispanic Black patients were tested less frequently for AATD compared to other groups [76]. This was not the case for screening for cystic fibrosis, immunoglobulin deficiency, and mycobacteria [76].

Clinical Implications of Low Screening Rates

Early recognition of AATD is critical as it permits interventions, including education (e.g., smoking cessation and avoidance), genetic counseling, testing family members, and specific treatment options. The cessation of smoking is strongly recommended following diagnosis of AATD. Equally, the initiation of early additional clinical interventions is paramount, such as bronchodilator and inhaler therapies, pulmonary rehabilitation, and lung volume reduction or lung transplantation in severe cases [70]. Delayed diagnosis is also associated with a negative psychosocial impact, which could be mitigated [77]. For each additional year of diagnostic delay in AATD, FEV1% predicted decreases by 0.3%, the St. George Respiratory Questionnaire total score increases by 1.6 points, and the COPD Assessment Test score increases by 0.7 points [77]. Equally, determining the precise AATD genotype is a major indicator of disease risk, with the ZZ and rare ZZ-equivalent genotypes associated with a higher risk of COPD and liver disease.

Without neonatal screening, or at the very least systematic targeted screening for those with obstructive lung disease and unexplained liver disease, the true prevalence of AATD will remain undetermined, and the disease will continue to be underdiagnosed. Currently, the ATS/ERS does not recommend screening neonates or adolescents [7], but this would facilitate early education and intervention.

6. Benefits of Diagnosing AATD

A correct diagnosis of AATD provides patients and physicians with a variety of management and treatment options. These include consultation with a genetic counselor to discuss the diagnosis, treatment options, lifestyle changes, and the screening of other family members. Diagnosis is particularly important as it permits the screening of relatives, as they need to consider AATD in their clinical history and maintain surveillance and risk-reduction for liver and lung diseases (such as alcohol consumption and smoke cessation). A recent study demonstrated that the greater the severity of the AATD genotype, the lower the smoking rates among ever-smokers, with a diagnosis of AATD shown to be a stronger motivator to quit smoking than a diagnosis of COPD [78]. Diagnosed AATD individuals also should consider vaccination strategies, undergo regular health checks, and lung and liver scans.

In those with emphysema caused by severe AATD, weekly intravenous (IV) infusions of plasma-purified AAT, known as augmentation therapy, are an effective treatment option. A recent trial demonstrated that patients with emphysema caused by severe AATD treated with AAT augmentation therapy showed a slower radiologic progression of the disease as compared to placebo [79]. In addition, the discontinuation of AAT augmentation therapy in an Irish ZZ population prompted a deterioration of lung disease, including increased exacerbations following the abrupt cessation of treatment [80].

It is important to note that the costs of AAT augmentation therapy are high within the USA and throughout the world. One study published in 2018 found the total medical cost of patients on augmentation therapy to be USD 127,000, while the cost of patients not on therapy was USD 15,874 annually for insurance companies [81]. Disparities already exist in

the diagnosis of AATD, and the high cost of treatment may have an impact on populations unable to access this expensive therapy.

7. Conclusions

AATD is substantially underdiagnosed, with an estimated 10% of predicted severe cases of AATD diagnosed. Most of the available data regarding the mutation frequency of SERPINA1 variants in Black populations are estimates. A small number of studies provide actual data on AATD in Black populations, but these cohorts are very small. The increasing frequency of transnational or multiracial relationships may also affect the presence of AAT alleles in these populations. Comparing the prior clinical epidemiological studies from earlier studies to large existing datasets (such as the gnomAD database) would likely increase our knowledge of the true frequencies of *SERPINA1* variants and AATD in Black populations. The recent improvement in diagnosis and treatment further emphasizes the importance of identifying AATD in Black populations, thereby providing this neglected population with appropriate and more effective medical care and treatment options. See Figure 1 for possible AATD testing, management, and treatment strategies for subjects with a suspicion of AATD independent of their age or ethnicity.



Figure 1. Flowchart for the testing of AATD and management/treatment options.

Author Contributions: Conceptualization, P.L., T.P.C. and P.G.; writing—original draft preparation, P.L., K.Z., M.P., E.A., T.P.C. and P.G.; writing—review and editing, P.L., T.P.C. and P.G.; supervision, T.P.C. and P.G.; project administration, P.G. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by grants made available to P.G. (the Alpha-1 Foundation (614218)).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare that there is no conflict of interest regarding the publication of this article.

Abbreviations

Alpha-1 antitrypsin, AAT; Alpha-1 antitrypsin deficiency, AATD; chronic obstructive pulmonary disease, COPD; American Thoracic Society, ATS; European Respiratory Society, ERS; Antineutrophilic cytoplasmic antibody, ANCA; whole genome sequence, WGS.

References

- Cunningham, T.J.; Croft, J.B.; Liu, Y.; Lu, H.; Eke, P.I.; Giles, W.H. Vital Signs: Racial Disparities in Age-Specific Mortality Among Blacks or African Americans—United States, 1999–2015. MMWR Morb. Mortal. Wkly. Rep. 2017, 66, 444–456. [CrossRef] [PubMed]
- Williams, D.R. Miles to go before we sleep: Racial inequities in health. *J. Health Soc. Behav.* 2012, *53*, 279–295. [CrossRef] [PubMed]
 Azar, K.M.J.; Shen, Z.; Romanelli, R.J.; Lockhart, S.H.; Smits, K.; Robinson, S.; Brown, S.; Pressman, A.R. Disparities in Outcomes
- Among COVID-19 Patients in A Large Health Care System in California. *Health Aff.* 2020, 39, 1253–1262. [CrossRef] [PubMed]
 Ghosh, A.J.; Hobbs, B.D.; Moll, M.; Saferali, A.; Boueiz, A.; Yun, J.H.; Sciurba, F.; Barwick, L.; Limper, A.H.; Flaherty, K.; et al. Alpha-1 Antitrypsin MZ Heterozygosity Is an Endotype of Chronic Obstructive Pulmonary Disease. *Am. J. Respir. Crit. Care Med.* 2022, 205, 313–323. [CrossRef] [PubMed]
- Palomino, L.; Gaffo, A.; Sun, D.; Sattui, S.E. Clinical Features of ANCA-Associated Vasculitis in African American Patients in the United States: A Single-Center Medical Records Review Study. J. Clin. Rheumatol. 2022, 28, 212–216. [CrossRef] [PubMed]
- Mamary, A.J.; Stewart, J.I.; Kinney, G.L.; Hokanson, J.E.; Shenoy, K.; Dransfield, M.T.; Foreman, M.G.; Vance, G.B.; Criner, G.J.; Investigators, C.O. Race and Gender Disparities are Evident in COPD Underdiagnoses Across all Severities of Measured Airflow Obstruction. *Chronic Obstr. Pulm. Dis.* 2018, 5, 177–184. [CrossRef] [PubMed]
- American Thoracic, S.; European Respiratory, S. American Thoracic Society/European Respiratory Society statement: Standards for the diagnosis and management of individuals with alpha-1 antitrypsin deficiency. *Am. J. Respir. Crit. Care Med.* 2003, 168, 818–900. [CrossRef]
- 8. Laurell, C.B.; Eriksson, S. Hypo-Alpha-1-Antitrypsinemia. Verh. Dtsch. Ges. Inn. Med. 1964, 70, 537–539.
- McElvaney, N.G.; Stoller, J.K.; Buist, A.S.; Prakash, U.B.; Brantly, M.L.; Schluchter, M.D.; Crystal, R.D. Baseline characteristics of enrollees in the National Heart, Lung and Blood Institute Registry of alpha 1-antitrypsin deficiency. Alpha 1-Antitrypsin Deficiency Registry Study Group. *Chest* 1997, 111, 394–403. [CrossRef]
- 10. Strange, C. Alpha-1 Antitrypsin Deficiency Associated COPD. Clin. Chest Med. 2020, 41, 339–345. [CrossRef]
- 11. Bernspang, E.; Sveger, T.; Piitulainen, E. Respiratory symptoms and lung function in 30-year-old individuals with alpha-1antitrypsin deficiency. *Respir. Med.* 2007, 101, 1971–1976. [CrossRef]
- 12. Parr, D.G.; Guest, P.G.; Reynolds, J.H.; Dowson, L.J.; Stockley, R.A. Prevalence and impact of bronchiectasis in alpha1-antitrypsin deficiency. *Am. J. Respir. Crit. Care Med.* 2007, 176, 1215–1221. [CrossRef]
- Adair-Kirk, T.L.; Senior, R.M. Fragments of extracellular matrix as mediators of inflammation. Int. J. Biochem. Cell Biol. 2008, 40, 1101–1110. [CrossRef]
- 14. Lascano, J.E.; Campos, M.A. The important role of primary care providers in the detection of alpha-1 antitrypsin deficiency. *Postgrad. Med.* **2017**, *129*, 889–895. [CrossRef]
- Molloy, K.; Hersh, C.P.; Morris, V.B.; Carroll, T.P.; O'Connor, C.A.; Lasky-Su, J.A.; Greene, C.M.; O'Neill, S.J.; Silverman, E.K.; McElvaney, N.G. Clarification of the risk of chronic obstructive pulmonary disease in alpha1-antitrypsin deficiency PiMZ heterozygotes. *Am. J. Respir. Crit. Care Med.* 2014, 189, 419–427. [CrossRef] [PubMed]
- Foreman, M.G.; Wilson, C.; DeMeo, D.L.; Hersh, C.P.; Beaty, T.H.; Cho, M.H.; Ziniti, J.; Curran-Everett, D.; Criner, G.; Hokanson, J.E.; et al. Alpha-1 Antitrypsin PiMZ Genotype Is Associated with Chronic Obstructive Pulmonary Disease in Two Racial Groups. *Ann. Am. Thorac. Soc.* 2017, 14, 1280–1287. [CrossRef] [PubMed]
- 17. Ortega, V.E.; Li, X.; O'Neal, W.K.; Lackey, L.; Ampleford, E.; Hawkins, G.A.; Grayeski, P.J.; Laederach, A.; Barjaktarevic, I.; Barr, R.G.; et al. The Effects of Rare SERPINA1 Variants on Lung Function and Emphysema in SPIROMICS. *Am. J. Respir. Crit. Care Med.* **2020**, *201*, 540–554. [CrossRef] [PubMed]
- Blanco, I.; Bueno, P.; Diego, I.; Perez-Holanda, S.; Casas-Maldonado, F.; Esquinas, C.; Miravitlles, M. Alpha-1 antitrypsin Pi*Z gene frequency and Pi*ZZ genotype numbers worldwide: An update. *Int. J. Chron. Obstruct Pulmon Dis.* 2017, 12, 561–569. [CrossRef] [PubMed]

- Association, A.L. COPD Prevalence. Available online: https://www.lung.org/research/trends-in-lung-disease/copd-trendsbrief/copd-prevalence (accessed on 9 September 2023).
- World Health Organization. Chronic Obstructive Pulmonary Disease (COPD). Available online: https://www.who.int/data/ global-health-estimates (accessed on 9 September 2023).
- Ejike, C.O.; Dransfield, M.T.; Hansel, N.N.; Putcha, N.; Raju, S.; Martinez, C.H.; Han, M.K. Chronic Obstructive Pulmonary Disease in America's Black Population. *Am. J. Respir. Crit. Care Med.* 2019, 200, 423–430. [CrossRef] [PubMed]
- Tan, W.C.; Sin, D.D.; Bourbeau, J.; Hernandez, P.; Chapman, K.R.; Cowie, R.; FitzGerald, J.M.; Marciniuk, D.D.; Maltais, F.; Buist, A.S.; et al. Characteristics of COPD in never-smokers and ever-smokers in the general population: Results from the CanCOLD study. *Thorax* 2015, *70*, 822–829. [CrossRef]
- 23. Riley, L.; Sriram, A.; Brantly, M.; Lascano, J. Testing Patterns and Disparities for Alpha-1 Antitrypsin Deficiency. *Am. J. Med.* 2023, 136, 1011–1017. [CrossRef] [PubMed]
- Franciosi, A.N.; Hobbs, B.D.; McElvaney, O.J.; Molloy, K.; Hersh, C.; Clarke, L.; Gunaratnam, C.; Silverman, E.K.; Carroll, T.P.; McElvaney, N.G. Clarifying the Risk of Lung Disease in SZ Alpha-1 Antitrypsin Deficiency. *Am. J. Respir. Crit. Care Med.* 2020, 202, 73–82. [CrossRef] [PubMed]
- Hernandez-Perez, J.M.; Ramos-Diaz, R.; Vaquerizo-Pollino, C.; Perez, J.A. Frequency of alleles and genotypes associated with alpha-1 antitrypsin deficiency in clinical and general populations: Revelations about underdiagnosis. *Pulmonology* 2022, 29, 214–220. [CrossRef] [PubMed]
- Carroll, T.P.; O'Connor, C.A.; Floyd, O.; McPartlin, J.; Kelleher, D.P.; O'Brien, G.; Dimitrov, B.D.; Morris, V.B.; Taggart, C.C.; McElvaney, N.G. The prevalence of alpha-1 antitrypsin deficiency in Ireland. *Respir. Res.* 2011, 12, 91. [CrossRef] [PubMed]
- Ashenhurst, J.R.; Nhan, H.; Shelton, J.F.; Wu, S.; Tung, J.Y.; Elson, S.L.; Stoller, J.K.; 23andMe Research Team. Prevalence of Alpha-1 Antitrypsin Deficiency, Self-Reported Behavior Change, and Health Care Engagement Among Direct-to-Consumer Recipients of a Personalized Genetic Risk Report. *Chest* 2022, *161*, 373–381. [CrossRef] [PubMed]
- 28. The Genome Aggregation Database (gnomAD). Available online: https://gnomad.broadinstitute.org/ (accessed on 12 August 2023).
- Seixas, S.; Marques, P.I. Known Mutations at the Cause of Alpha-1 Antitrypsin Deficiency an Updated Overview of SERPINA1 Variation Spectrum. *Appl. Clin. Genet.* 2021, 14, 173–194. [CrossRef]
- Silva, D.; Oliveira, M.J.; Guimaraes, M.; Lima, R.; Gomes, S.; Seixas, S. Alpha-1-antitrypsin (SERPINA1) mutation spectrum: Three novel variants and haplotype characterization of rare deficiency alleles identified in Portugal. *Respir. Med.* 2016, 116, 8–18. [CrossRef]
- Giacopuzzi, E.; Laffranchi, M.; Berardelli, R.; Ravasio, V.; Ferrarotti, I.; Gooptu, B.; Borsani, G.; Fra, A. Real-world clinical applicability of pathogenicity predictors assessed on SERPINA1 mutations in alpha-1-antitrypsin deficiency. *Hum. Mutat.* 2018, 39, 1203–1213. [CrossRef]
- 32. Gaillard, M.C.; Mahadeva, R.; Lomas, D.A. Identification of DNA polymorphisms associated with the V type alpha1-antitrypsin gene. *Biochim. Biophys. Acta* **1999**, 1444, 166–170. [CrossRef]
- Laurell, C.B.; Eriksson, S. The electrophoretic alpha1-globulin pattern of serum in alpha1-antitrypsin deficiency. 1963. COPD: J. Chronic Obstr. Pulm. Dis. 2013, 10, 3–8. [CrossRef]
- 34. Fagerhol, M.K.; Tenfjord, O.W. Serum Pi types in some European, American, Asian and African populations. *Acta Pathol. Microbiol. Scand.* **1968**, 72, 601–608. [CrossRef] [PubMed]
- Curiel, D.T.; Holmes, M.D.; Okayama, H.; Brantly, M.L.; Vogelmeier, C.; Travis, W.D.; Stier, L.E.; Perks, W.H.; Crystal, R.G. Molecular basis of the liver and lung disease associated with the alpha 1-antitrypsin deficiency allele Mmalton. *J. Biol. Chem.* 1989, 264, 13938–13945. [CrossRef] [PubMed]
- Poller, W.; Merklein, F.; Schneider-Rasp, S.; Haack, A.; Fechner, H.; Wang, H.; Anagnostopoulos, I.; Weidinger, S. Molecular characterisation of the defective alpha 1-antitrypsin alleles PI Mwurzburg (Pro369Ser), Mheerlen (Pro369Leu), and Q0lisbon (Thr68Ile). *Eur. J. Hum. Genet.* 1999, 7, 321–331. [CrossRef] [PubMed]
- Lomas, D.A.; Finch, J.T.; Seyama, K.; Nukiwa, T.; Carrell, R.W. Alpha 1-antitrypsin Siiyama (Ser53-->Phe). Further evidence for intracellular loop-sheet polymerization. *J. Biol. Chem.* 1993, 268, 15333–15335. [CrossRef] [PubMed]
- Talamo, R.C.; Langley, C.E.; Reed, C.E.; Makino, S. α₁-Antitrypsin deficiency: A variant with no detecTable α₁-antitrypsin. *Science* 1973, *181*, 70–71. [CrossRef] [PubMed]
- de Serres, F.J.; Blanco, I. Prevalence of alpha1-antitrypsin deficiency alleles PI*S and PI*Z worldwide and effective screening for each of the five phenotypic classes PI*MS, PI*MZ, PI*SS, PI*SZ, and PI*ZZ: A comprehensive review. *Ther. Adv. Respir. Dis.* 2012, 6, 277–295. [CrossRef] [PubMed]
- Qiu, W.; Baccarelli, A.; Carey, V.J.; Boutaoui, N.; Bacherman, H.; Klanderman, B.; Rennard, S.; Agusti, A.; Anderson, W.; Lomas, D.A.; et al. Variable DNA methylation is associated with chronic obstructive pulmonary disease and lung function. *Am. J. Respir. Crit. Care Med.* 2012, 185, 373–381. [CrossRef] [PubMed]
- Beckmeyer-Borowko, A.; Imboden, M.; Rezwan, F.I.; Wielscher, M.; Amaral, A.F.S.; Jeong, A.; Schaffner, E.; Auvinen, J.; Sebert, S.; Karhunen, V.; et al. SERPINA1 methylation and lung function in tobacco-smoke exposed European children and adults: A meta-analysis of ALEC population-based cohorts. *Respir. Res.* 2018, 19, 156. [CrossRef]
- Rotondo, J.C.; Oton-Gonzalez, L.; Selvatici, R.; Rizzo, P.; Pavasini, R.; Campo, G.C.; Lanzillotti, C.; Mazziotta, C.; De Mattei, M.; Tognon, M.; et al. SERPINA1 Gene Promoter Is Differentially Methylated in Peripheral Blood Mononuclear Cells of Pregnant Women. *Front. Cell Dev. Biol.* 2020, *8*, 550543. [CrossRef]

- Corley, M.; Solem, A.; Phillips, G.; Lackey, L.; Ziehr, B.; Vincent, H.A.; Mustoe, A.M.; Ramos, S.B.V.; Weeks, K.M.; Moorman, N.J.; et al. An RNA structure-mediated, posttranscriptional model of human alpha-1-antitrypsin expression. *Proc. Natl. Acad. Sci. USA* 2017, 114, E10244–E10253. [CrossRef]
- Lackey, L.; McArthur, E.; Laederach, A. Increased Transcript Complexity in Genes Associated with Chronic Obstructive Pulmonary Disease. PLoS ONE 2015, 10, e0140885. [CrossRef] [PubMed]
- Grayeski, P.J.; Weidmann, C.A.; Kumar, J.; Lackey, L.; Mustoe, A.M.; Busan, S.; Laederach, A.; Weeks, K.M. Global 5'-UTR RNA structure regulates translation of a SERPINA1 mRNA. *Nucleic Acids Res.* 2022, 50, 9689–9704. [CrossRef] [PubMed]
- 46. Stoller, J.K. Alpha-1 antitrypsin deficiency: An underrecognized, treatable cause of COPD. *Cleve Clin. J. Med.* **2016**, *83*, 507–514. [CrossRef] [PubMed]
- Zhao, X.; Qiao, D.; Yang, C.; Kasela, S.; Kim, W.; Ma, Y.; Shrine, N.; Batini, C.; Sofer, T.; Taliun, S.A.G.; et al. Whole genome sequence analysis of pulmonary function and COPD in 19,996 multi-ethnic participants. *Nat. Commun.* 2020, 11, 5182. [CrossRef] [PubMed]
- de Serres, F.J. Worldwide racial and ethnic distribution of alpha1-antitrypsin deficiency: Summary of an analysis of published genetic epidemiologic surveys. Chest 2002, 122, 1818–1829. [CrossRef] [PubMed]
- 49. Stoller, J.K.; Aboussouan, L.S. A review of alpha1-antitrypsin deficiency. *Am. J. Respir. Crit. Care Med.* 2012, 185, 246–259. [CrossRef] [PubMed]
- 50. Martinez-Gonzalez, C.; Blanco, I.; Diego, I.; Bueno, P.; Miravitlles, M. Estimated Prevalence and Number of PiMZ Genotypes of Alpha-1 Antitrypsin in Seventy-Four Countries Worldwide. *Int. J. Chron. Obstruct Pulmon Dis.* 2021, *16*, 2617–2630. [CrossRef]
- 51. Chatila, W.M.; Hoffman, E.A.; Gaughan, J.; Robinswood, G.B.; Criner, G.J.; National Emphysema Treatment Trial Research, G. Advanced emphysema in African-American and white patients: Do differences exist? *Chest* **2006**, *130*, 108–118. [CrossRef]
- 52. Silverman, E.K.; Miletich, J.P.; Pierce, J.A.; Sherman, L.A.; Endicott, S.K.; Broze, G.J., Jr.; Campbell, E.J. Alpha-1-antitrypsin deficiency. High prevalence in the St. Louis area determined by direct population screening. *Am. Rev. Respir. Dis.* **1989**, *140*, 961–966. [CrossRef]
- Banauch, G.I.; Brantly, M.; Izbicki, G.; Hall, C.; Shanske, A.; Chavko, R.; Santhyadka, G.; Christodoulou, V.; Weiden, M.D.; Prezant, D.J. Accelerated spirometric decline in New York City firefighters with alpha(1)-antitrypsin deficiency. *Chest* 2010, 138, 1116–1124. [CrossRef]
- 54. Spinola, C.; Brehm, A.; Spinola, H. Alpha-1-antitrypsin deficiency in the Cape Verde islands (Northwest Africa): High prevalence in a sub-Saharan population. *Respir. Med.* **2010**, *104*, 1069–1072. [CrossRef] [PubMed]
- 55. Miskoff, J.A.; Khan, B.; Chaudhri, M.; Phan, H.; Carson, M.P. Identifying Alpha-1 Antitrypsin Deficiency Based on Computed Tomography Evidence of Emphysema. *Cureus* **2019**, *11*, e3971. [CrossRef] [PubMed]
- Denden, S.; Zorzetto, M.; Amri, F.; Knani, J.; Ottaviani, S.; Scabini, R.; Gorrini, M.; Ferrarotti, I.; Campo, I.; Chibani, J.B.; et al. Screening for Alpha 1 antitrypsin deficiency in Tunisian subjects with obstructive lung disease: A feasibility report. *Orphanet J. Rare Dis.* 2009, *4*, 12. [CrossRef] [PubMed]
- 57. Webb, D.R.; Hyde, R.W.; Schwartz, R.H.; Hall, W.J.; Condemi, J.J.; Townes, P.L. Serum alpha 1-antitrypsin variants. Prevalence and clinical spirometry. *Am. Rev. Respir. Dis.* **1973**, *108*, 918–925. [CrossRef] [PubMed]
- 58. Young, R.C., Jr.; Headings, V.E.; Henderson, A.L.; Bose, S.; Hackney, R.L., Jr. Protease inhibitor profile of black Americans with and without chronic cardiopulmonary disease. *J. Natl. Med. Assoc.* **1978**, *70*, 849–856. [PubMed]
- 59. Pierce, J.A.; Eradio, B.; Dew, T.A. Antitrypsin phenotypes in St. Louis. J. Am. Med. Assoc. 1975, 231, 609-612. [CrossRef]
- 60. Massi, G.; Vecchio, F.M. Alpha-1-antitrypsin phenotypes in a group of newborn infants in Somalia. *Hum. Genet.* **1977**, *38*, 265–269. [CrossRef]
- 61. Vandeville, D.; Martin, J.P.; Ropartz, C. Alpha 1-antitrypsin polymorphism of a Bantu population: Description of a new allele PiL. *Humangenetik* **1974**, *21*, 33–38. [CrossRef]
- 62. Welch, S.G.; McGregor, I.A.; Williams, K. Alpha 1-antitrypsin (pi) phenotypes in a village population from The Gambia, West Africa. Evidence of a new variant occurring at a polymorphic frequency. *Hum. Genet.* **1980**, *53*, 233–235. [CrossRef]
- 63. Pascali, V.L.; Ranalletta, D.; Spedini, G. Antitrypsin and Gc polymorphisms in some populations of Congo: An unusual, highly frequent mutant, PIS, in Bateke and Babenga. *Ann. Hum. Biol.* **1986**, *13*, 267–271. [CrossRef]
- 64. Chaabani, H.; Martin, J.P.; Frants, R.R.; Lefranc, G. Genetic study of Tunisian Berbers. II. Alpha 1-antitrypsin (Pi) polymorphism: Report of a new allele (Pi S Berber). *Exp. Clin. Immunogenet.* **1984**, *1*, 19–24. [PubMed]
- 65. Lieberman, J.; Gaidulis, L.; Roberts, L. Racial distribution of alpha1-antitrypsin variants among junior high school students. *Am. Rev. Respir. Dis.* **1976**, *114*, 1194–1198. [CrossRef]
- 66. Brems, J.H.; Balasubramanian, A.; Psoter, K.J.; Shah, P.D.; Bush, E.; Merlo, C.A.; McCormack, M.C. Race-specific Interpretation of Spirometry: Impact on the Lung Allocation Score. *Ann. Am. Thorac. Soc.* **2023**, *20*, 1408–1415. [CrossRef] [PubMed]
- 67. Ekstrom, M.; Mannino, D. Research race-specific reference values and lung function impairment, breathlessness and prognosis: Analysis of NHANES 2007–2012. *Respir. Res.* **2022**, *23*, 271. [CrossRef] [PubMed]
- Seaman, E.L.; Corcy, N.; Chang, J.T.; Chomenko, D.; Hartman, A.M.; Kittner, D.L.; Reyes-Guzman, C.M. Menthol Cigarette Smoking Trends among United States Adults, 2003–2019. *Cancer Epidemiol. Biomark. Prev.* 2022, 31, 1959–1965. [CrossRef] [PubMed]
- 69. Foulds, J.; Hooper, M.W.; Pletcher, M.J.; Okuyemi, K.S. Do smokers of menthol cigarettes find it harder to quit smoking? *Nicotine Tob. Res.* **2010**, *12*, S102–S109. [CrossRef] [PubMed]

- 70. Patrucco, F.; Venezia, L.; Gavelli, F.; Pellicano, R.; Solidoro, P. Alpha1-antitrypsin deficiency: What's new after European Respiratory Society Statement. *Panminerva Med.* **2018**, *60*, 101–108. [CrossRef] [PubMed]
- 71. Haillot, A.; Pelland, A.A.; Bosse, Y.; Carroll, T.P.; Maltais, F.; Dandurand, R.J. IntraIndividual Variability in Serum Alpha-1 Antitrypsin Levels. *Chronic Obstr. Pulm. Dis.* **2021**, *8*, 464–473. [CrossRef]
- 72. Franciosi, A.N.; Carroll, T.P.; McElvaney, N.G. Pitfalls and caveats in alpha1-antitrypsin deficiency testing: A guide for clinicians. *Lancet Respir. Med.* **2019**, *7*, 1059–1067. [CrossRef]
- 73. Belmonte, I.; Barrecheguren, M.; Lopez-Martinez, R.M.; Esquinas, C.; Rodriguez, E.; Miravitlles, M.; Rodriguez-Frias, F. Application of a diagnostic algorithm for the rare deficient variant Mmalton of alpha-1-antitrypsin deficiency: A new approach. *Int. J. Chron. Obs. Pulmon Dis.* **2016**, *11*, 2535–2541. [CrossRef]
- 74. Papiris, S.A.; Veith, M.; Papaioannou, A.I.; Apollonatou, V.; Ferrarotti, I.; Ottaviani, S.; Tzouvelekis, A.; Tzilas, V.; Rovina, N.; Stratakos, G.; et al. Alpha1-antitrypsin deficiency in Greece: Focus on rare variants. *Pulmonology*, 2023; *Epub ahead of print*. [CrossRef]
- 75. Kueppers, F.; Sanders, C. State-of-the-art testing for alpha-1 antitrypsin deficiency. *Allergy Asthma Proc.* 2017, *38*, 108–114. [CrossRef] [PubMed]
- McShane, P.J.; Choate, R.; Johnson, M.; Maselli, D.J.; Winthrop, K.L.; Metersky, M.L.; Bronchiectasis and NTM Research Registry Investigators. Racial and ethnic differences in patients enrolled in the national bronchiectasis and nontuberculous mycobacteria research registry. *Respir. Med.* 2023, 209, 107167. [CrossRef] [PubMed]
- 77. Tejwani, V.; Nowacki, A.S.; Fye, E.; Sanders, C.; Stoller, J.K. The Impact of Delayed Diagnosis of Alpha-1 Antitrypsin Deficiency: The Association Between Diagnostic Delay and Worsened Clinical Status. *Respir. Care* **2019**, *64*, 915–922. [CrossRef] [PubMed]
- 78. Franciosi, A.N.; Alkhunaizi, M.A.; Woodsmith, A.; Aldaihani, L.; Alkandari, H.; Lee, S.E.; Fee, L.T.; McElvaney, N.G.; Carroll, T.P. Alpha-1 Antitrypsin Deficiency and Tobacco Smoking: Exploring Risk Factors and Smoking Cessation in a Registry Population. *COPD: J. Chronic Obstr. Pulm. Dis.* 2021, 18, 76–82. [CrossRef] [PubMed]
- 79. Chapman, K.R.; Burdon, J.G.; Piitulainen, E.; Sandhaus, R.A.; Seersholm, N.; Stocks, J.M.; Stoel, B.C.; Huang, L.; Yao, Z.; Edelman, J.M.; et al. Intravenous augmentation treatment and lung density in severe alpha1 antitrypsin deficiency (RAPID): A randomised, double-blind, placebo-controlled trial. *Lancet* 2015, *386*, 360–368. [CrossRef]
- McElvaney, O.J.; Carroll, T.P.; Franciosi, A.N.; Sweeney, J.; Hobbs, B.D.; Kowlessar, V.; Gunaratnam, C.; Reeves, E.P.; McElvaney, N.G. Consequences of Abrupt Cessation of Alpha1-Antitrypsin Replacement Therapy. N. Engl. J. Med. 2020, 382, 1478–1480. [CrossRef]
- 81. Sieluk, J.; Levy, J.; Sandhaus, R.A.; Silverman, H.; Holm, K.E.; Mullins, C.D. Costs of Medical Care Among Augmentation Therapy Users and Non-Users with Alpha-1 Antitrypsin Deficiency in the United States. *Chronic Obstr. Pulm. Dis.* 2018, *6*, 6–16. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.