

Supplementary Materials

Synthesis of D-Fructose-Based Bifunctional Primary Amine-Thiourea Organocatalysts and Their Applications in Asymmetric Reactions

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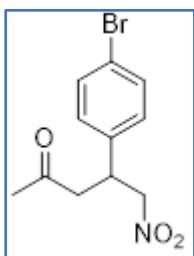
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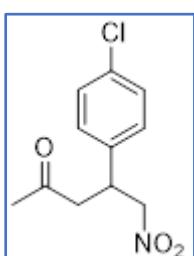
Copies of HPLC data of enantioenriched and racemic data of compound **9a–e**: Page 52–56.

Typical procedure for asymmetric Michael Addition reaction:

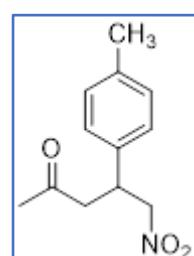
To a stirred solution of β -nitrostyrene (0.2 mmol) and ketone (3 equiv.) in dry dichloromethane (0.25 ml), 15 mol% saccharide-based amine-thiourea organocatalyst and benzoic acid were added. The reaction mixture was then stirred at room temperature for an appropriate reaction time, followed by concentration under vacuum. The reaction mixture was then subjected to purification by column chromatography using silica gel (60-120 mesh) with a hexane:EtOAc mixture as an eluent to obtain the desired product. The enantiomeric excess values of the product were determined by HPLC analysis on a chiral column using a mixture of n-hexane and iso-propanol as the mobile phase.



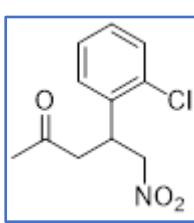
(*S*)-4-(4-bromophenyl)-5-nitropentan-2-one (**7a**): yield: 95% as yellow solid. $[\alpha]_D^{25} = -12.1$ ($c = 0.068$, CHCl₃). Chiralpak AD-H, hexane/i-PrOH=85/15, UV 220 nm, flow rate 1.0 ml/min, $t_R = 10.2$ min (major, *S*), $t_R = 12.6$ (minor, *R*), ee: >99%. ¹H NMR (400 MHz, CDCl₃): δ 7.40-7.36 (m, 2H), 7.04-7.02 (m, 2H), 4.63-4.47 (m, 2H), 3.94-3.87 (m, 1H), 2.82 (d, 2H, $J = 8$ Hz), 2.06 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 204.95, 137.88, 132.21, 131.57, 131.18, 129.15, 121.87, 79.09, 45.91, 38.47, 30.37 ppm.



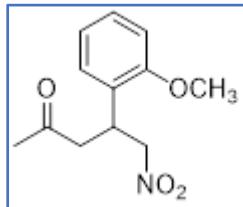
(*S*)-4-(4-chlorophenyl)-5-nitropentan-2-one (**7b**): yield: 90% as yellow solid. $[\alpha]_D^{25} = +5.9$ ($c = 0.052$, CHCl₃). Chiralpak AD-H, hexane/i-PrOH=85/15, UV 220 nm, flow rate 1.0 ml/min, $t_R = 9.8$ min (major, *S*), $t_R = 11.5$ (minor, *R*) ee: 91%. ¹H NMR (400 MHz, CDCl₃): δ 7.24-7.20 (m, 2H), 7.11-7.07 (m, 2H), 4.63-4.47 (m, 2H), 4.06-3.88 (m, 1H), 2.82 (d, 2H, $J = 8$ Hz), 2.05 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 205.00, 137.36, 133.78, 130.86, 129.25, 128.82, 128.59, 79.18, 45.97, 38.41, 30.36 ppm.



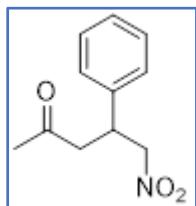
(*S*)-5-nitro-4-(*p*-tolyl)pentan-2-one (**7c**): yield: 81% as white solid. $[\alpha]_D^{25} = +1.2$ ($c = 0.045$, CHCl₃). Chiralpak AD-H, hexane/i-PrOH=85/15, UV 220 nm, flow rate 1.0 ml/min, $t_R = 7.4$ min (major, *S*), $t_R = 8.2$ (minor, *R*) ee: 85%. ¹H NMR (400 MHz, CDCl₃): δ 7.20-7.07 (m, 4H), 4.69-4.54 (m, 2H), 4.00-3.93 (m, 1H), 2.89 (d, 2H, $J = 8$ Hz), 2.31 (s, 3H), 2.10 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 204.56, 136.56, 134.75, 128.70, 127.91, 126.51, 126.21, 78.60, 45.18, 37.73, 29.34, 19.99 ppm.



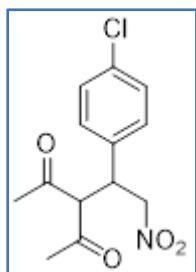
(*S*)-4-(2-chlorophenyl)-5-nitropentan-2-one (**7d**): yield: 88% as yellow oil. $[\alpha]_D^{25} = +6.8$ ($c = 0.062$, CHCl₃). Chiralpak AD-H, hexane/i-PrOH=90/10, UV 210 nm, flow rate 1.0 ml/min, $t_R = 9.817$ min (major, *S*), $t_R = 12.583$ min (minor, *R*) ee: 83%. ¹H NMR (400 MHz, CDCl₃): δ 7.34-7.12 (m, 4H), 4.73-4.64 (m, 2H), 4.43-4.36 (m, 1H), 3.02-2.86 (m, 2H), 2.09 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 204.24, 134.97, 132.71, 129.41, 128.03, 127.40, 126.38, 76.41, 43.50, 34.78, 29.18 ppm.



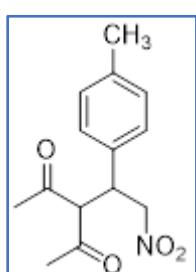
(S)-4-(2-methoxyphenyl)-5-nitropentan-2-one (**7e**): yield: 94% as pale-yellow oil. $[\alpha]_D^{25} = +15.3$ ($c = 0.034$, CHCl₃). Chiralpak AD-H, hexane/i-PrOH=48/2, UV 205 nm, flow rate 0.5 ml/min, t_R =31.2 min (major, *S*), t_R =32.7 (minor, *R*) ee: 81%. ¹H NMR (400 MHz, CDCl₃): δ 7.17-7.15 (m, 1H), 7.07-7.05 (m, 1H), 6.84-6.80 (m, 2H), 4.69-4.63 (m, 2H), 4.17-4.12 (m, 1H), 3.78 (s, 3H), 2.98-2.84 (m, 2H), 2.04 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 205.15, 156.09, 128.27, 127.97, 125.45, 119.88, 109.99, 76.84, 54.32, 43.51, 34.34, 29.18 ppm.



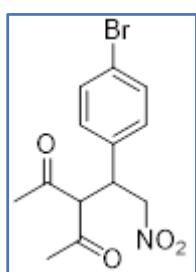
(S)-5-nitro-4-phenylpentane-2-one (**7f**): yield: 93% as white solid. $[\alpha]_D^{25} = +6.9$ ($c = 0.055$, CHCl₃). Chiralpak AD-H, hexane/i-PrOH=97/3, UV 220 nm, flow rate 1 ml/min, t_R =18.3 min (major, *S*), t_R =21.4 (minor, *R*) ee: 95%. ¹H NMR (400 MHz, CDCl₃): δ 7.37-7.23 (m, 5H), 4.74-4.60 (m, 2H), 4.07-3.99 (m, 1H), 2.94 (d, $J = 8$ Hz, 2H), 2.14 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 205.39, 138.82, 129.09, 127.92, 127.39, 79.46, 46.18, 39.05, 30.40 ppm.



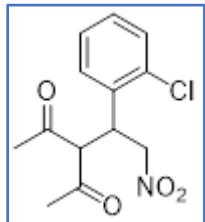
(S)-3-(1-(4-chlorophenyl)-2-nitroethyl)pentane-2,4-dione (**7g**): yield: 80% as white solid. $[\alpha]_D^{25} = +6.9$ ($c = 0.055$, CHCl₃). Chiralpak AD-H, hexane/i-PrOH=80/20, UV 205 nm, flow rate 1 ml/min, t_R =8.2 min (major, *S*), t_R =22.0 (minor, *R*) ee: 84%. ¹H NMR (400 MHz, CDCl₃): δ 7.32-7.29 (m, 2H), 7.15-7.11 (m, 2H), 4.65-4.58 (m, 2H), 4.34-4.20 (m, 2H), 2.30 (s, 3H), 1.97 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 201.38, 200.55, 134.58, 134.27, 133.22, 129.57, 129.33, 77.92, 70.56, 42.15, 30.41, 29.64 ppm.



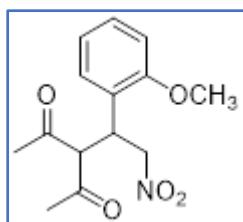
(S)-3-(2-nitro-1-(p-tolyl)ethyl)pentane-2,4-dione (**7h**): yield: 73% as white solid. $[\alpha]_D^{25} = +91.3$ ($c = 0.066$, CHCl₃). Chiralpak AD-H, hexane/i-PrOH=90/10, UV 210 nm, flow rate 1 ml/min, t_R =10.0 min (major, *S*), t_R =17.1 (minor, *R*) ee: 81%. ¹H NMR (400 MHz, CDCl₃): δ 7.13-7.05 (m, 4H), 4.64-4.56 (m, 2H), 4.35 (d, 1H, $J = 8$ Hz), 4.23-4.17 (m, 1H), 2.30 (s, 3H), 2.29 (s, 3H), 1.94 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 201.88, 201.08, 138.38, 132.85, 130.01, 127.78, 78.38, 70.88, 42.47, 30.38, 29.41, 21.05 ppm.



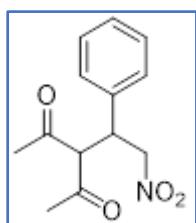
(S)-3-(1-(4-bromophenyl)-2-nitroethyl)pentane-2,4-dione (**7i**): yield: 78% as pale-yellow solid. $[\alpha]_D^{25} = +141.7$ ($c = 0.068$, CHCl₃). Chiralpak AD-H, hexane/i-PrOH=85/15, UV 210 nm, flow rate 1 ml/min, t_R =10.6 min (major, *S*), t_R =29.8 (minor, *R*) ee: 81%. ¹H NMR (400 MHz, CDCl₃): δ 7.48-7.45 (m, 2H), 7.09-7.05 (m, 2H), 4.65-4.58 (m, 2H), 4.32 (d, 1H, $J = 12$ Hz), 4.24-4.18 (m, 1H), 2.30 (s, 3H), 1.98 (s, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 201.40, 200.56, 135.11, 132.58, 129.68, 122.71, 77.87, 70.54, 42.22, 30.42, 29.64 ppm.



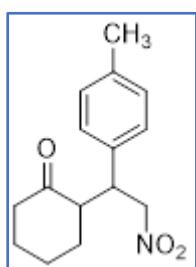
(*S*)-3-(1-(2-chlorophenyl)-2-nitroethyl)pentane-2,4-dione (**7j**): yield: 62% as pale-yellow solid. $[\alpha]_D^{25} = +200.1$ ($c = 0.048$, CHCl_3). Chiralpak AD-H, hexane/i-PrOH= 49/1, UV 205 nm, flow rate 0.5 ml/min, $t_R=33.9$ min (major, *S*), $t_R=36.1$ (minor, *R*) ee: 76%. ^1H NMR (400 MHz, CDCl_3): δ 7.38-7.07 (m, 4H), 4.79-4.75 (m, 1H), 4.70-4.65 (m, 1H), 4.61-4.52 (m, 2H), 2.23 (s, 3H), 1.97 (s, 3H) ppm. ^{13}C NMR (100 MHz, CDCl_3): δ 201.89, 200.83, 133.78, 133.45, 130.67, 129.73, 129.04, 127.67, 77.33, 69.02, 38.86, 30.88, 28.43 ppm.



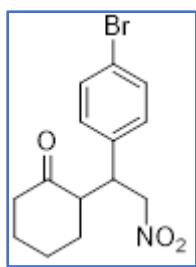
(*S*)-3-(1-(2-methoxyphenyl)-2-nitroethyl)pentane-2,4-dione (**7k**): yield: 52% as pale-yellow oil. $[\alpha]_D^{25} = +196.8$ ($c = 0.055$, CHCl_3). Chiralpak AD-H, hexane/i-PrOH= 49/1, UV 210 nm, flow rate 0.5 ml/min, $t_R=36.6$ min (major, *S*), $t_R=40.0$ (minor, *R*) ee: 71%. ^1H NMR (400 MHz, CDCl_3): δ 7.22-7.17 (m, 1H), 7.02-7.00 (m, 1H), 6.84-6.80 (m, 2H), 4.74-4.69 (m, 1H), 4.54-4.49 (m, 2H), 4.44-4.39 (m, 1H), 3.81 (s, 3H), 2.21 (s, 3H), 1.87 (s, 3H) ppm. ^{13}C NMR (100 MHz, CDCl_3): δ 203.08, 202.27, 158.11, 139.57, 134.04, 131.20, 128.98, 79.57, 60.27, 52.90, 31.58, 30.60, 22.24 ppm.



(*S*)-3-(2-nitro-1-phenylethyl)pentane-2,4-dione (**7l**): yield: 81% as pale-yellow solid. $[\alpha]_D^{25} = +173.1$ ($c = 0.048$, CHCl_3). Chiralpak AD-H, hexane/i-PrOH= 45/5, UV 210 nm, flow rate 0.5 ml/min, $t_R=21.8$ min (major, *S*), $t_R=29.5$ (minor, *R*) ee: 74%. ^1H NMR (400 MHz, CDCl_3): δ 7.40-7.10 (m, 5H), 4.59-4.52 (m, 2H), 4.32-4.16 (m, 2H), 2.22 (s, 3H), 1.87 (s, 3H) ppm. ^{13}C NMR (100 MHz, CDCl_3): δ 201.80, 201.03, 135.99, 129.35, 129.17, 128.88, 128.57, 127.94, 78.19, 42.80, 30.44, 29.55 ppm.

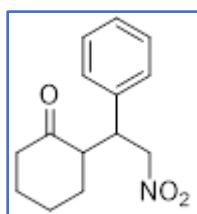


(*R*)-2-[*(S*)-1-(4-methylphenyl)-2-nitroethyl] cyclohexanone (**7m**): yield: 46% as yellow solid. $[\alpha]_D^{25} = +13.1$ ($c = 0.05$, CHCl_3). Chiralpak AD-H, hexane/i-PrOH= 48/2, UV 254 nm, flow rate 0.5 ml/min, retention time: 25.0 min (major, *syn*), 29.7 min (minor, *anti*), 31.3 min (major, *anti*) 38.8 min (minor, *syn*). ^1H NMR (400 MHz, CDCl_3): δ 7.15-7.03 (m, 4H), 4.96-4.80 (m, 1H), 4.65-4.60 (m, 1H) 4.01-3.96 (m, 1H for *anti*), 3.77-3.71 (m, 1H for *syn*), 2.76-2.66 (m, 1H), 2.52-1.20 (m, 11H). ^{13}C NMR (100 MHz, CDCl_3): δ 212.08 (*syn*), 210.58 (*anti*), 137.46, 137.22, 135.22, 134.60, 129.63, 129.45, 128.24, 128.01, 79.03, 53.82, 52.59, 43.59, 42.76, 42.74, 33.19, 30.03, 28.54, 27.36, 25.03, 25.01, 21.06 ppm.



(*S*)-2-[*(R*)-1-(4-bromophenyl)-2-nitroethyl] cyclohexanone (**7n**): yield: 55% as white solid. Chiralpak AD-H, hexane/i-PrOH= 90/10, UV 254 nm, flow rate 1 ml/min, retention time: 11.9 min (minor, *syn*), 13.6 min (major, *anti*), 15.3 min (minor, *anti*) 18.7 min (major, *syn*). ^1H NMR (400 MHz, CDCl_3): δ 7.65-7.44 (m, 2H), 7.18-7.06 (m, 2H), 4.97-4.85 (m, 1H), 4.84-4.58 (m, 1H), 3.96-3.78 (m, 1H for *anti*), 3.77-3.73 (m, 1H for *syn*), 2.75-2.63 (m, 1H), 2.51-1.25 (m, 8H). ^{13}C NMR (100 MHz, CDCl_3): δ 211.55 (*syn*), 210.30 (*anti*), 137.37,

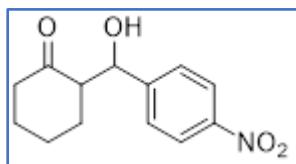
136.86, 133.75, 132.10, 131.87, 130.23, 129.93, 128.51, 121.73, 78.52, 53.55, 52.35, 43.44, 42.90, 42.74, 33.17, 30.32, 28.44, 27.35, 25.07 ppm.



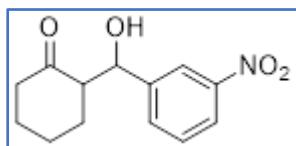
(S)-2-((R)-2-Nitro-1-phenylethyl)cyclohexan-1-one (7o): yield: 42% as white solid. $[\alpha]_D^{25} = -6.1$ ($c = 0.048$, CHCl_3). Chiralpak AD-H, hexane/i-PrOH=90/10, UV 254 nm, flow rate 1 ml/min, retention time: 12.6 min (minor, *syn*), 14.5 min (minor, *anti*), 19.1 min (major, *anti*), 25.9 min (major, *syn*). ^1H NMR (400 MHz, CDCl_3): δ 7.57-7.09 (m, 5H), 4.89-4.74 (m, 1H), 4.59-4.53 (m, 1H), 3.96-3.72 (m, 1H for *anti*), 3.71-3.66 (m, 1H for *syn*), 2.69-2.58 (m, 1H), 2.44-1.11 (m, 8H). ^{13}C NMR (100 MHz, CDCl_3): δ 212.00 (*syn*), 210.52 (*anti*), 128.95, 128.76, 128.50, 128.18, 127.79, 127.56, 78.90, 53.78, 52.55, 43.95, 43.08, 42.33, 33.22, 30.00, 28.54, 27.35, 25.04 ppm.

Typical procedure for the asymmetric Aldol reaction:

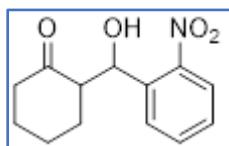
A solution of 20 mol% of the saccharide-based amine-thiourea organocatalyst **6a**, 20 mol% of benzoic acid, aldehydes (0.2 mmol), and ketone (4 equiv.) in water (0.5 ml) was stirred at 0 °C. The reaction mixture was then concentrated under vacuum, followed by purification with column chromatography using silica gel with hexane:EtOAc mixture. The enantiomeric excess values of the products were identified by HPLC analysis on a chiral column using a mixture of n-hexane and iso-propanol as the mobile phase.



(2R,1'R)-2-[Hydroxy-(4-nitro-phenyl)-methyl] cyclohexanone (8a): yield: 58% as white solid. Chiralpak OD-H, hexane/i-PrOH=95/5, UV 254 nm, flow rate 0.5 ml/min, *syn* product t_R (major) 56.6 min, t_R (minor) 66.6 min, ee: 30%, *anti* product t_R (major) 71.3 min, t_R (minor) 103.2 min, ee: 47%. ^1H NMR (400 MHz, CDCl_3): δ 8.29-8.24 (m, 2H), 7.56-7.50 (m, 2H) 5.53 (d, 1H for *syn*, $J = 2$ Hz), 4.95 (d, 1H for *anti*, $J = 8$ Hz), 3.21 (s, 1H), 2.88-1.30 (m, 9H) ppm. ^{13}C NMR (100 MHz, CDCl_3): δ 214.65 (*anti*), 213.96 (*syn*), 142.25, 139.98, 130.69, 127.89, 126.63, 74.02 (*anti*), 70.15 (*syn*), 57.21 (*anti*), 56.82 (*syn*), 42.68, 40.44, 24.71, 23.77, 23.31 ppm.

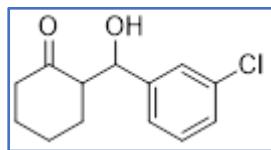


(2R,1'R)-2-[Hydroxy-(3-nitro-phenyl)-methyl] cyclohexanone (8b): yield: 52% as white solid. Chiralpak OD-H, hexane/i-PrOH=95/5, UV 254 nm, flow rate 0.5 ml/min, *syn* product t_R (major) 35.9 min, t_R (minor) 37.5 min, ee: 15%, *anti* product t_R (major) 42.6 min, t_R (minor) 61.8 min, ee: 52%. ^1H NMR (400 MHz, CDCl_3): δ 8.14-8.02 (m, 2H), 7.61-7.38 (m, 2H), 5.40 (d, 1H for *syn*, $J = 2$ Hz), 4.83 (d, 1H for *anti*, $J = 8$ Hz), 3.41 (s, 1H), 2.78-1.26 (m, 9H) ppm. ^{13}C NMR (100 MHz, CDCl_3): δ 214.82 (*anti*), 214.03 (*syn*), 143.98, 143.37, 132.44, 132.01, 124.39, 123.02, 122.84, 73.99 (*anti*), 65.90 (*syn*), 57.14 (*anti*), 56.78 (*syn*), 30.74, 25.94, 24.75, 24.66 ppm.

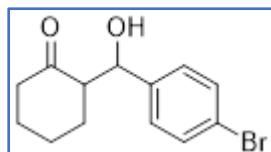


(2R,1'R)-2-[Hydroxy-(2-nitro-phenyl)-methyl] cyclohexanone (8c): 46% as pale-yellow solid. Chiralpak OD-H, hexane/i-PrOH=95/5, UV 254 nm, flow rate 0.5 ml/min, *syn* product t_R (minor) 20.3 min, t_R (major) 23.4 min, ee: 40%, *anti* product t_R (major)

32.2 min, t_R (minor) 38.6 min, ee: 67%. ^1H NMR (400 MHz, CDCl_3): δ 7.93-7.32 (m, 4H), 5.89 (s, 1H for *syn*), 5.37 (d, 1H for *anti*, $J = 8$ Hz), 3.27 (s, 1H), 2.82-0.81 (m, 9H) ppm. ^{13}C NMR (100 MHz, CDCl_3): δ 213.87 (*anti*), 212.91 (*syn*), 147.27, 146.14, 132.12, 132.02, 128.03, 127.92, 127.43, 126.91, 68.72 (*anti*), 65.59 (*syn*), 56.34 (*anti*), 53.86 (*syn*), 41.79, 41.52, 27.67, 26.93, 26.74 ppm.



(*2R,1'R*)-2-[Hydroxy-(3-chloro-phenyl)-methyl] cyclohexanone (**8d**): 54% as white solid. Chiralpak OD-H, hexane/i-PrOH=95/5, UV 220 nm, flow rate 0.5 ml/min, *syn* product t_R (minor) 18.1 min, t_R (major) 19.0 min, ee: 23%, *anti* product t_R (major) 22.7 min, t_R (minor) 30.1 min, ee: 18%. ^1H NMR (400 MHz, CDCl_3): δ 7.33-7.15 (m, 4H), 5.35 (d, 1H for *syn*, $J = 2$ Hz), 4.74 (d, 1H for *anti*, $J = 8$ Hz), 2.60-1.25 (m, 9H) ppm. ^{13}C NMR (100 MHz, CDCl_3): δ 215.20 (*anti*), 214.46 (*syn*), 143.71, 143.12, 129.86, 129.62, 128.48, 128.39, 128.07, 127.18, 74.30 (*anti*), 70.11 (*syn*), 57.30 (*anti*), 57.00 (*syn*), 30.81, 25.96, 24.73, 23.83, 23.36 ppm.



(*2R,1'R*)-2-[Hydroxy-(4-bromo-phenyl)-methyl] cyclohexanone (**8e**): yield: 50% as white solid. Chiralpak OD-H, hexane/i-PrOH=90/10, UV 254 nm, flow rate 1 ml/min, *syn* product t_R (minor) 6.6 min, t_R (major) 7.8 min, ee: 69%, *anti* product t_R (major) 9.9 min, t_R (minor) 13.5 min, ee: 73%. ^1H NMR (400 MHz, CDCl_3): δ 7.57-7.49 (m, 2H), 7.29-7.22 (m, 2H), 5.38 (d, 1H for *syn*, $J = 4$ Hz), 4.80 (d, 1H for *anti*, $J = 8$ Hz), 3.23 (s, 1H), 2.86-1.31 (m, 9H) ppm. ^{13}C NMR (100 MHz, CDCl_3): δ 215.23 (*anti*), 214.52 (*syn*), 140.58, 140.08, 134.53, 134.25, 131.76, 131.51, 128.77, 127.58, 74.21 (*anti*), 70.19 (*syn*), 57.36 (*anti*), 57.02 (*syn*), 42.68, 40.33, 27.94, 26.00, 24.74, 23.83, 23.34 ppm.

Figure S1: ^1H NMR and ^{13}C NMR of **3**.

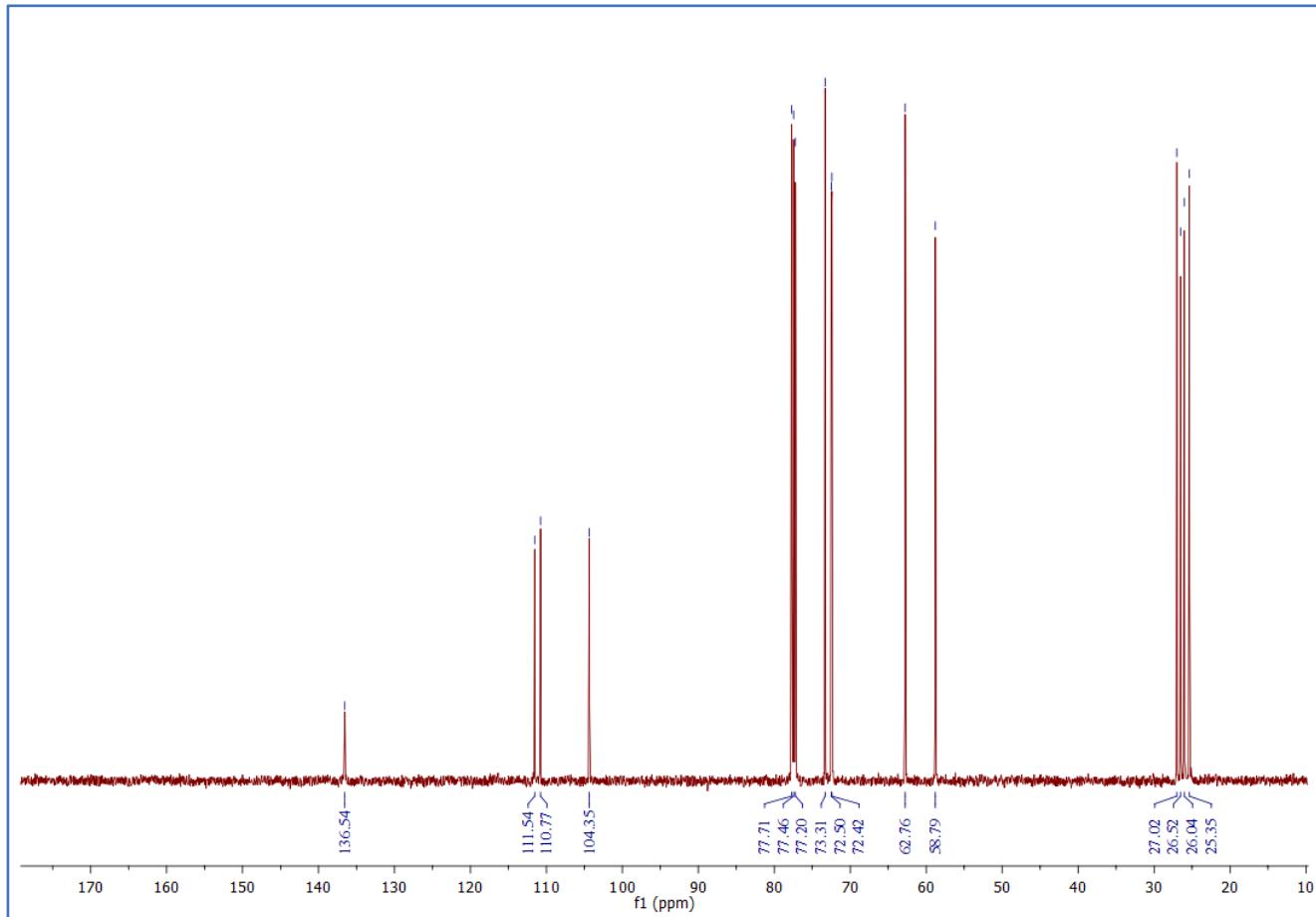
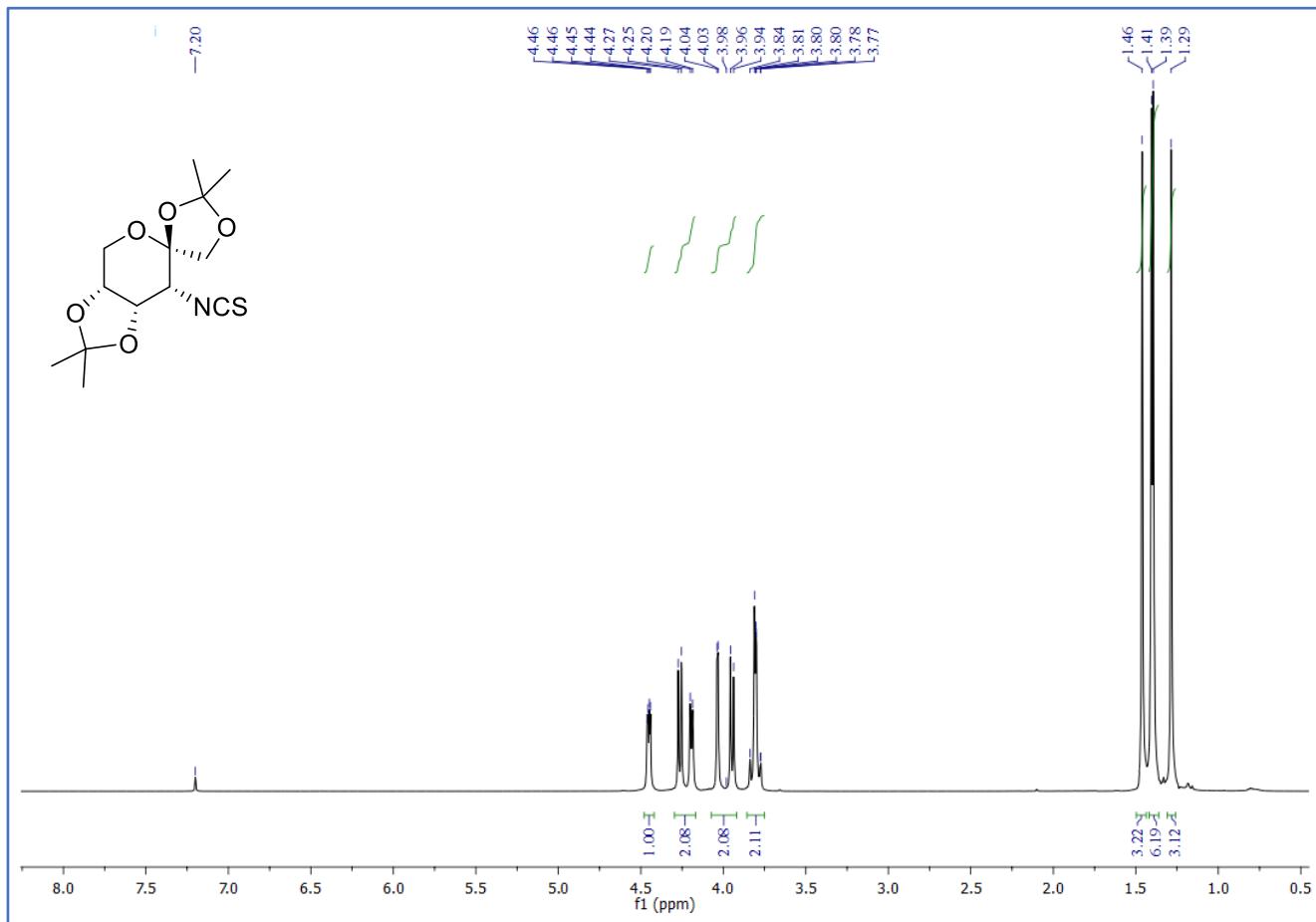


Figure S2: ^1H NMR and ^{13}C NMR of **4**.

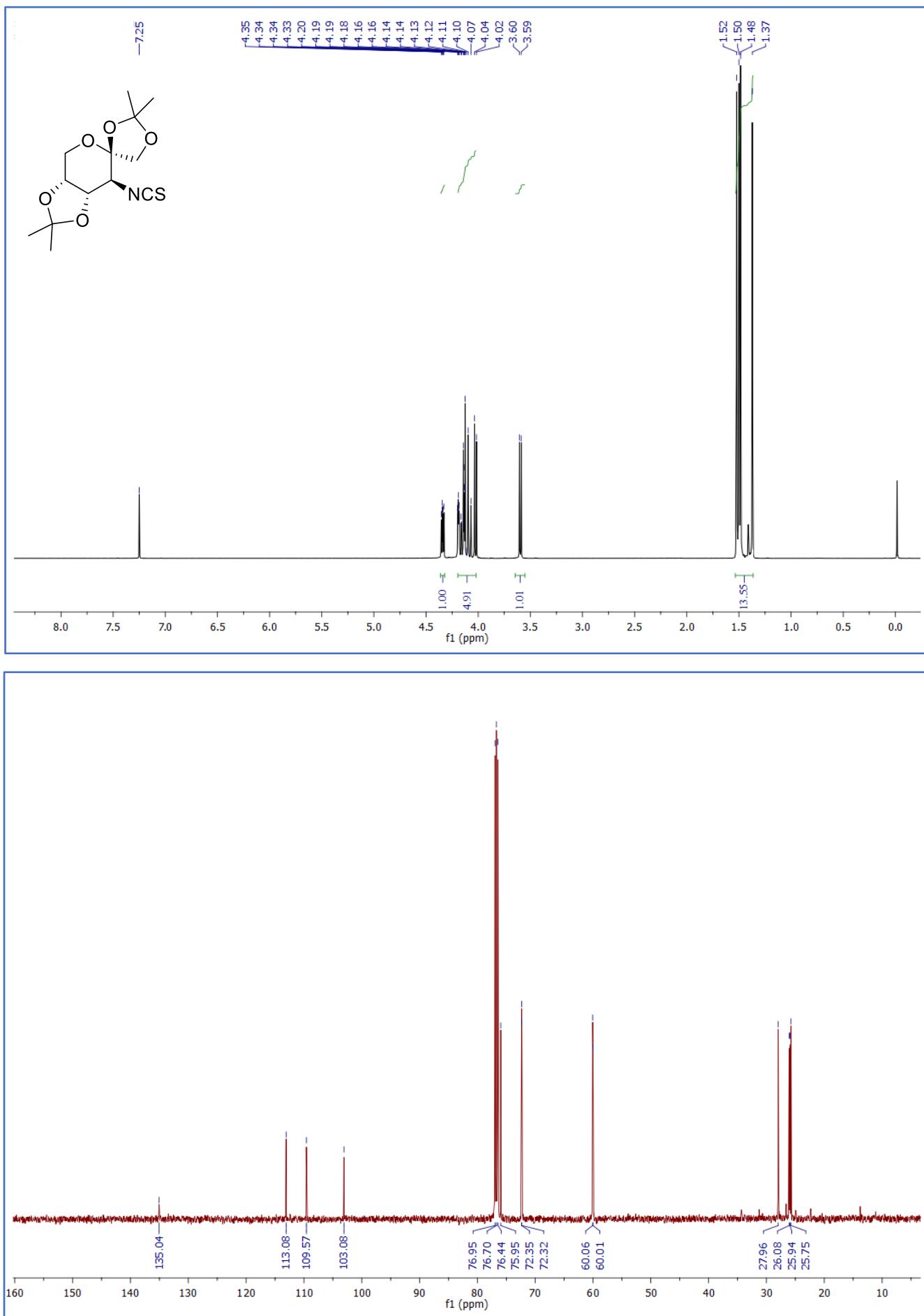


Figure S3: ^1H NMR and ^{13}C NMR of **5a**.

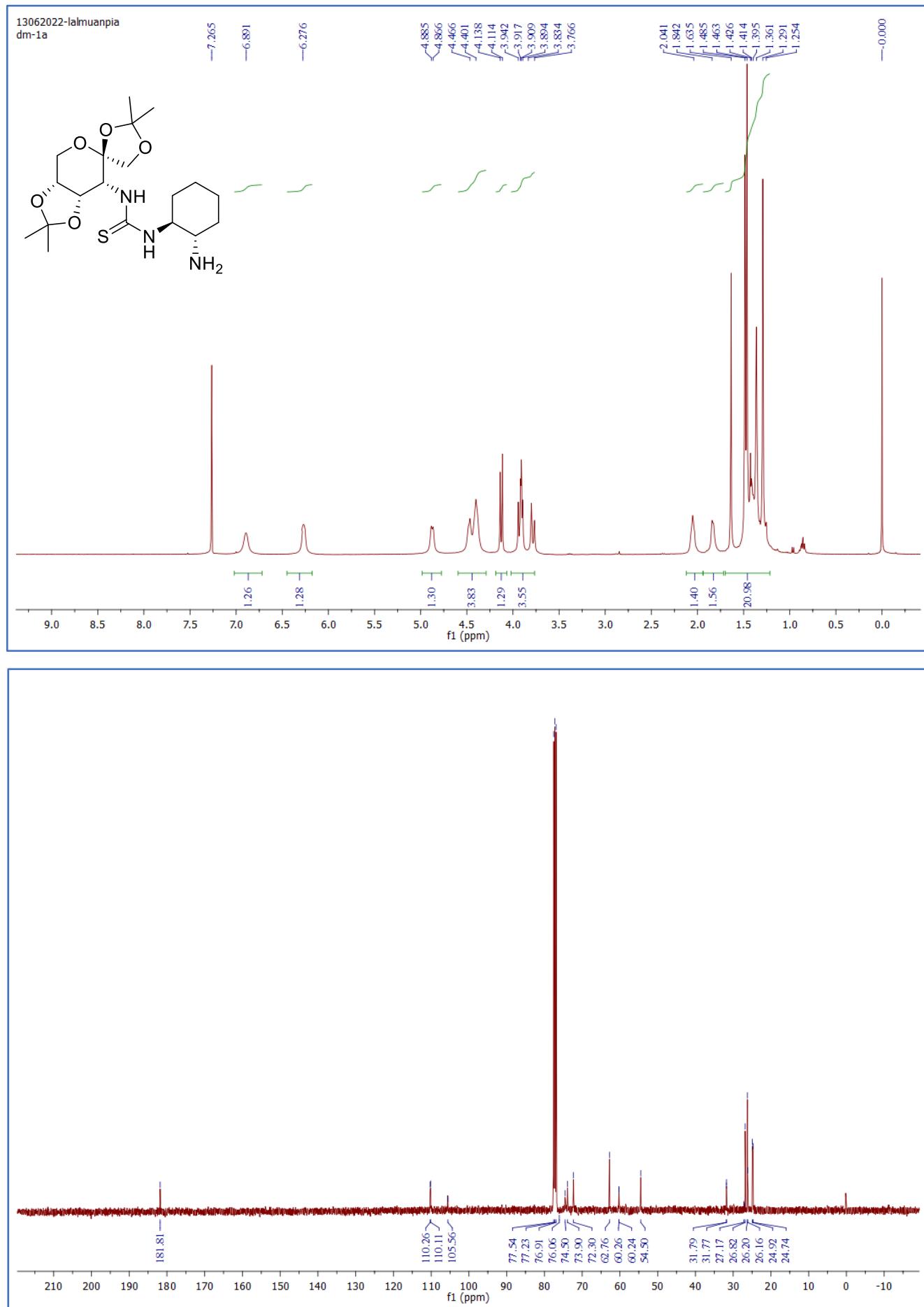


Figure S4: ^1H NMR and ^{13}C NMR of **5b**.

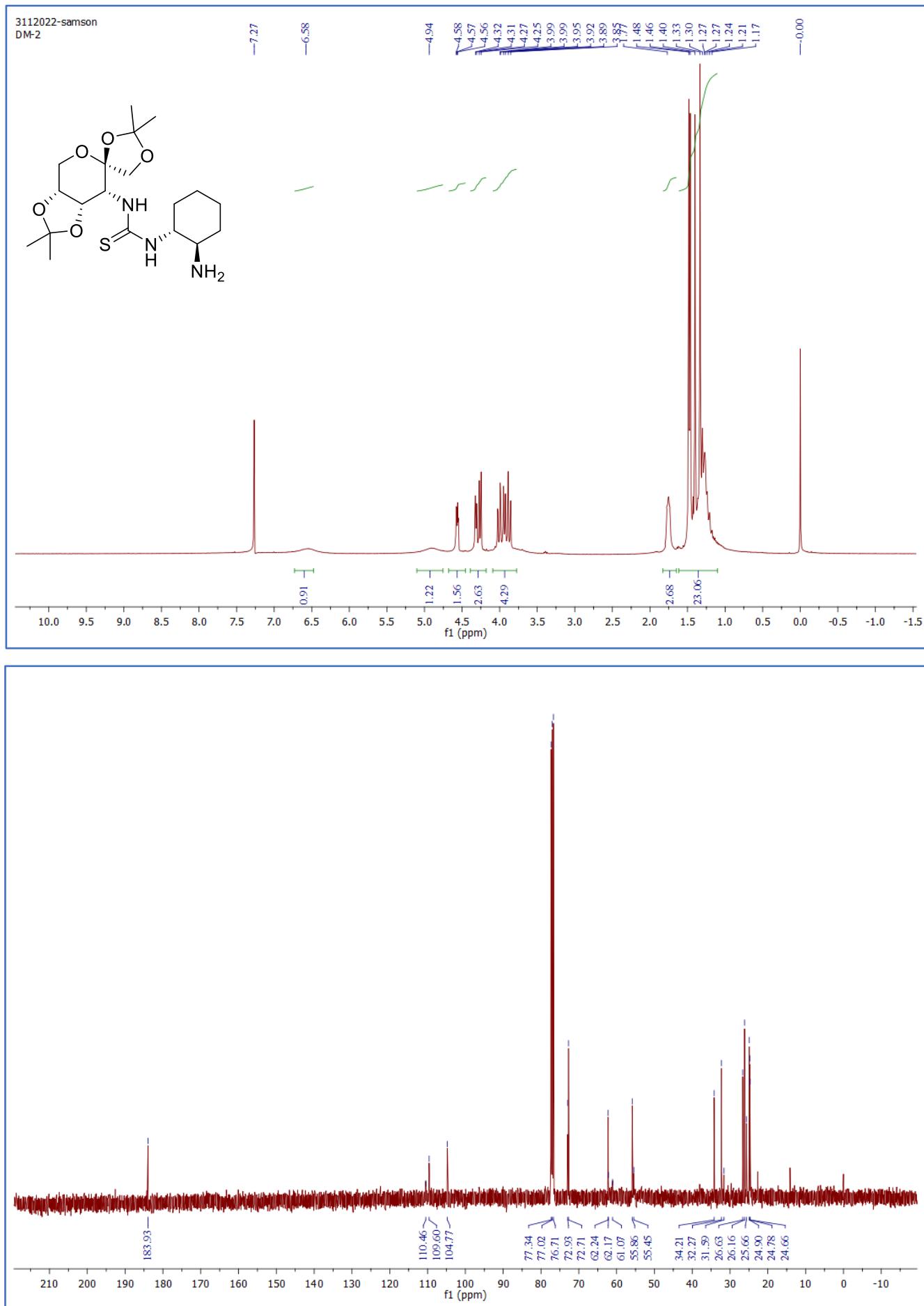


Figure S5: ^1H NMR and ^{13}C NMR of **5c**.

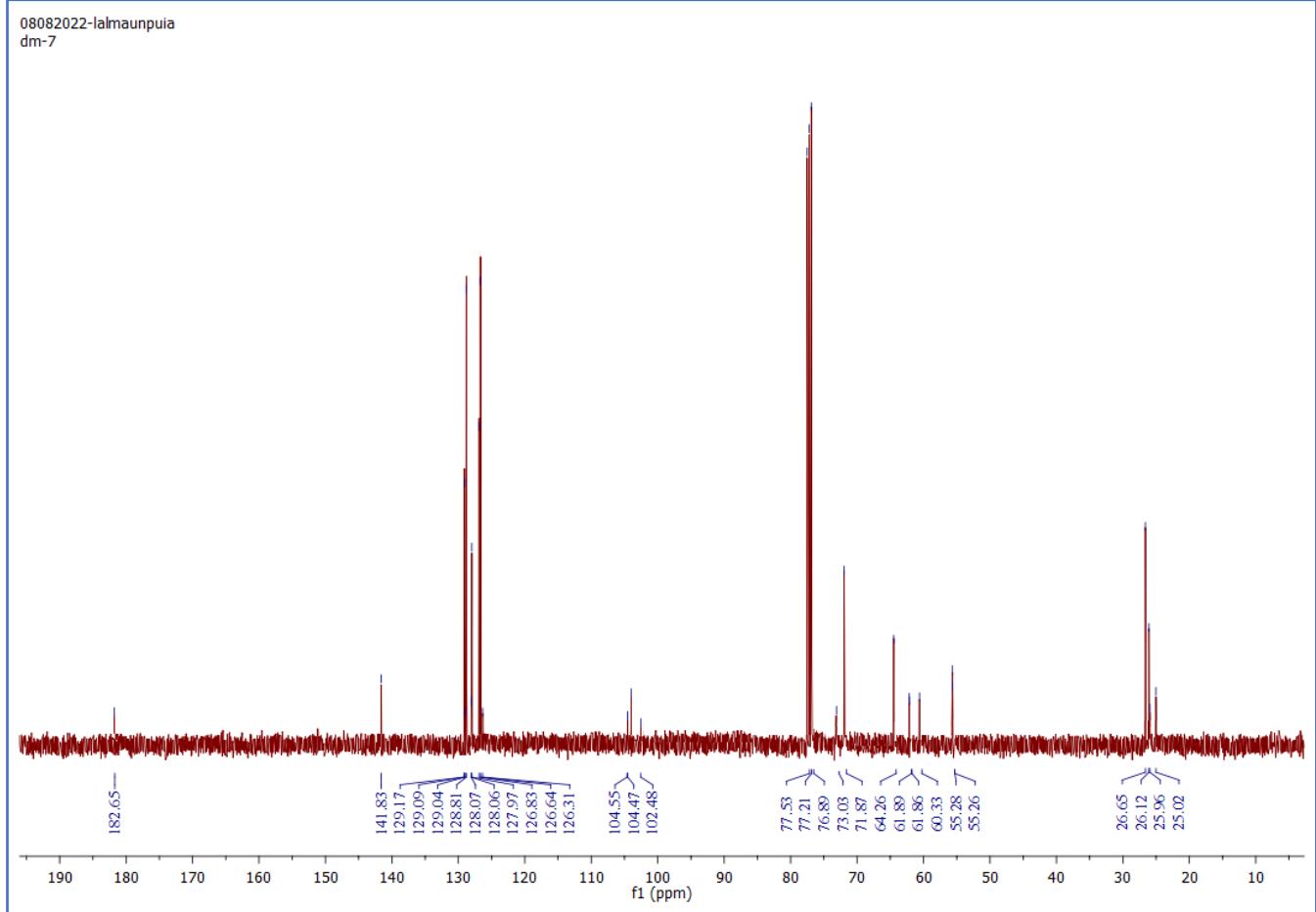
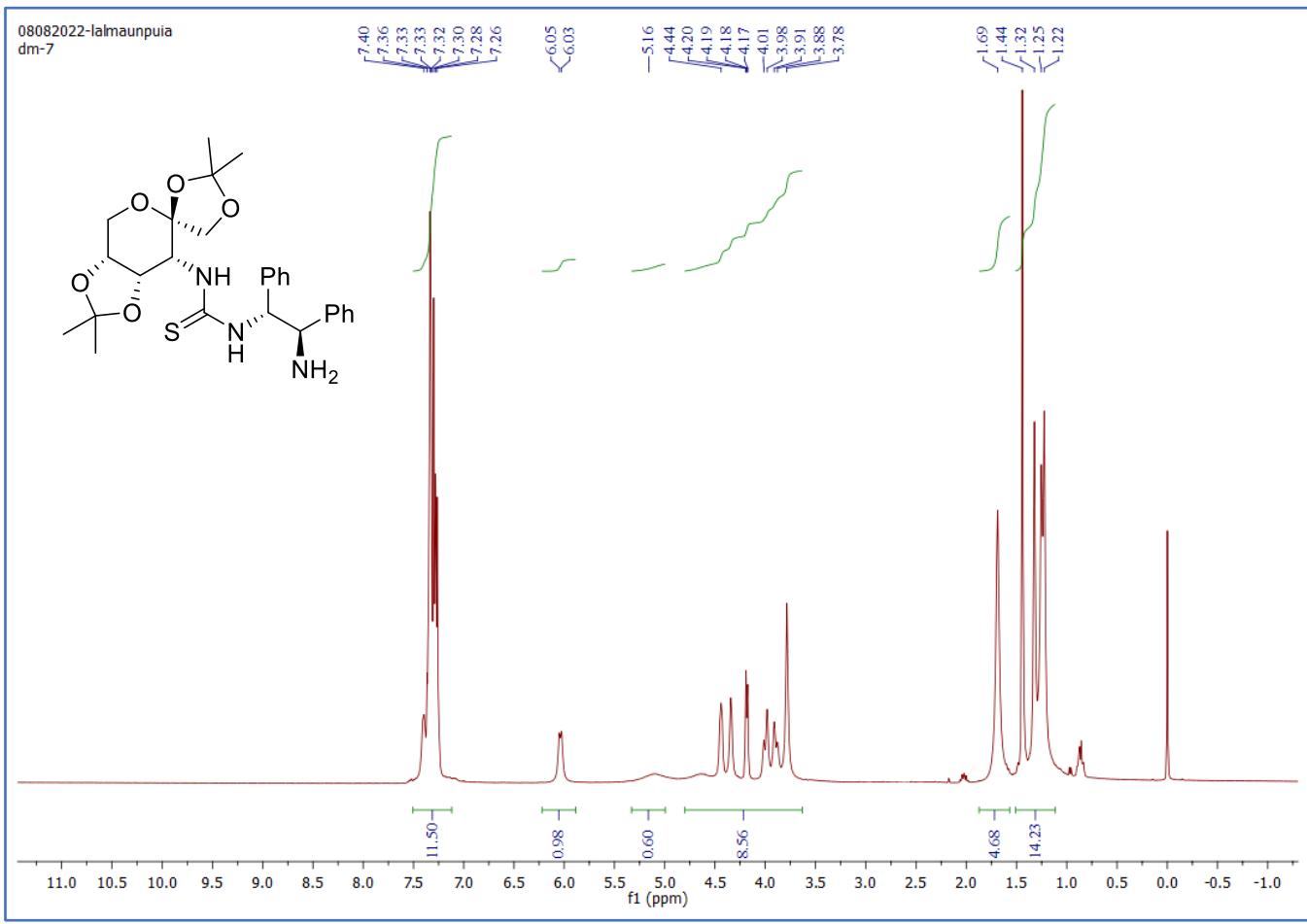


Figure S6: ^1H NMR and ^{13}C NMR of **5d**.

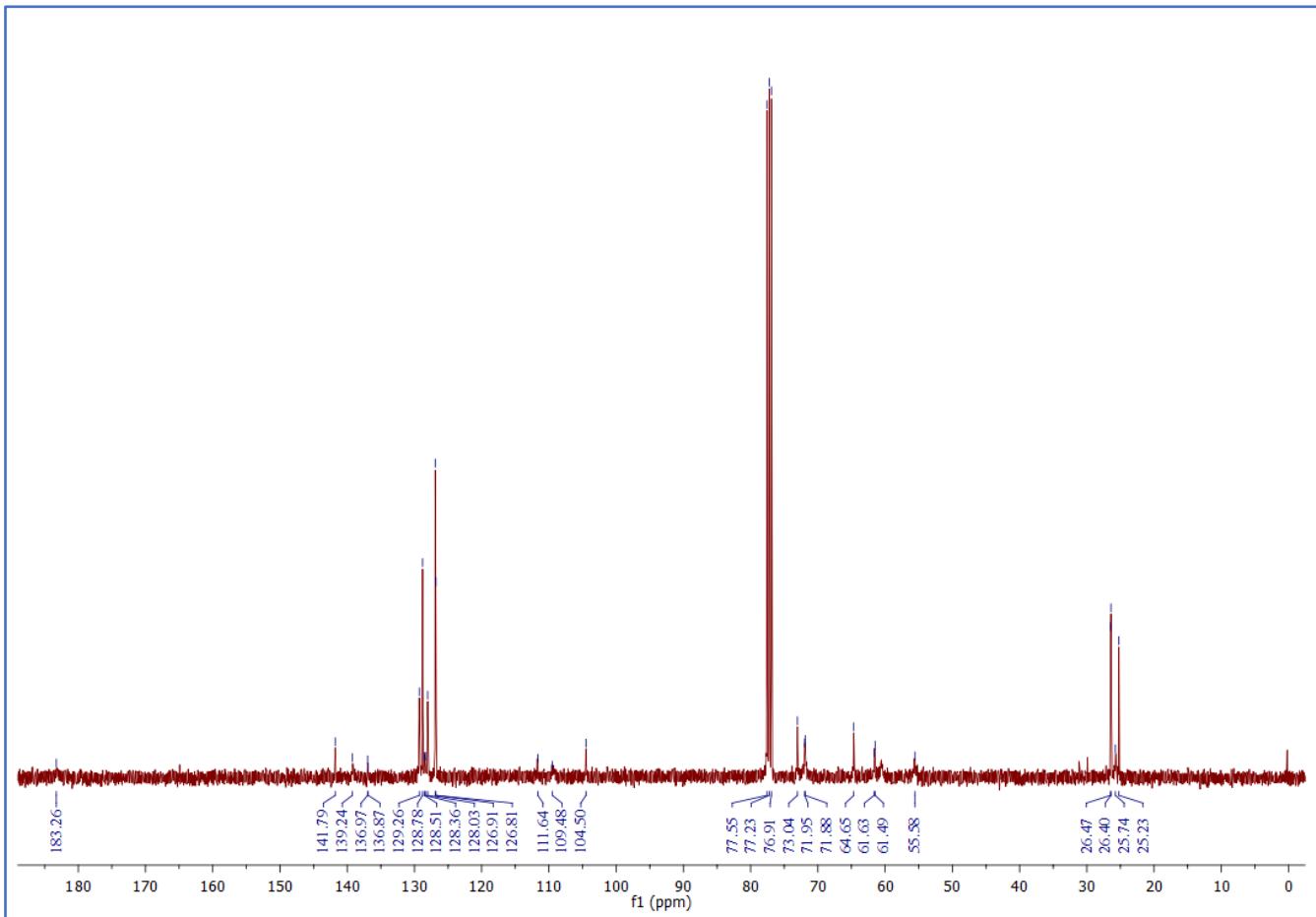
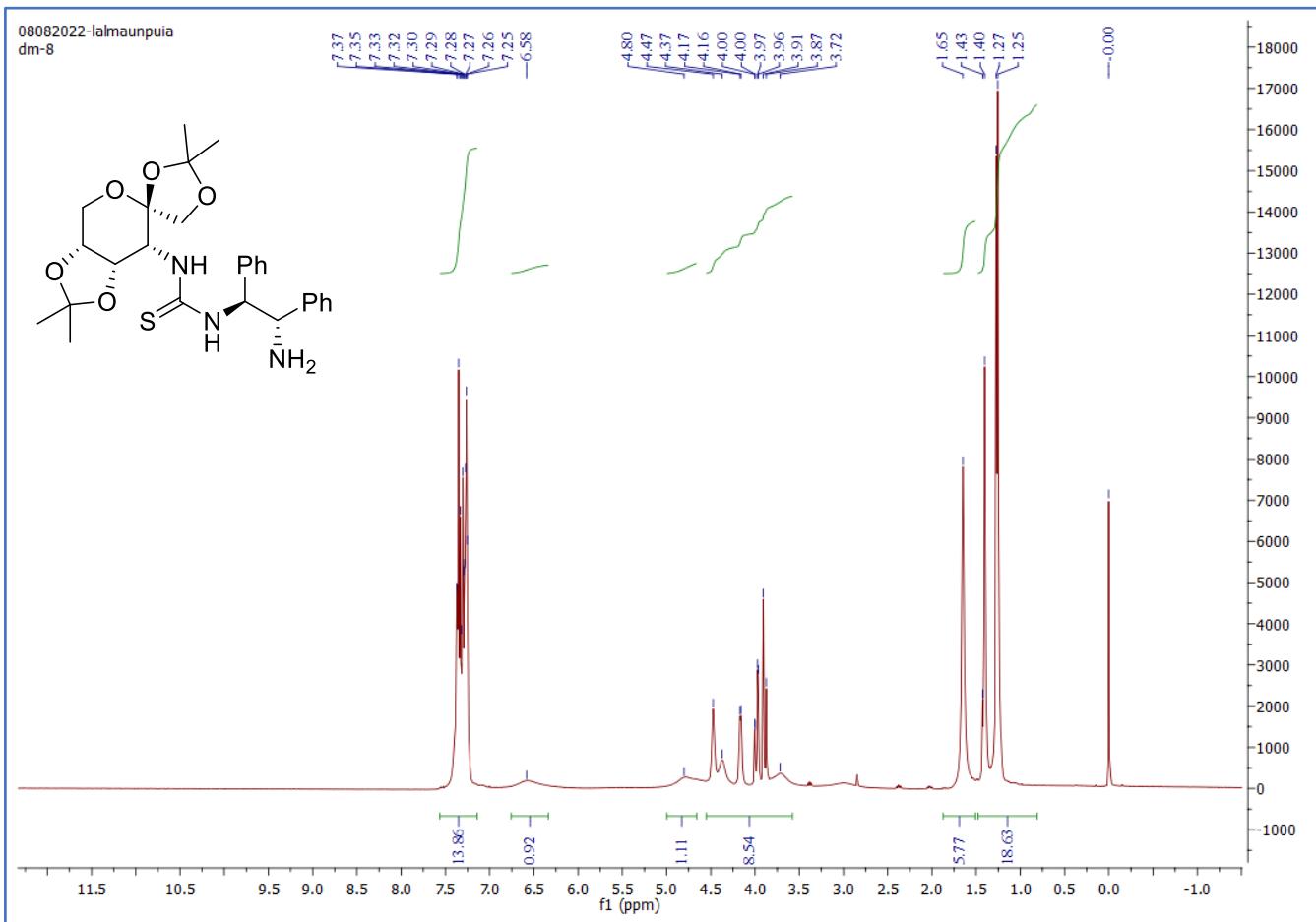


Figure S7: ^1H NMR and ^{13}C NMR of **6a**.

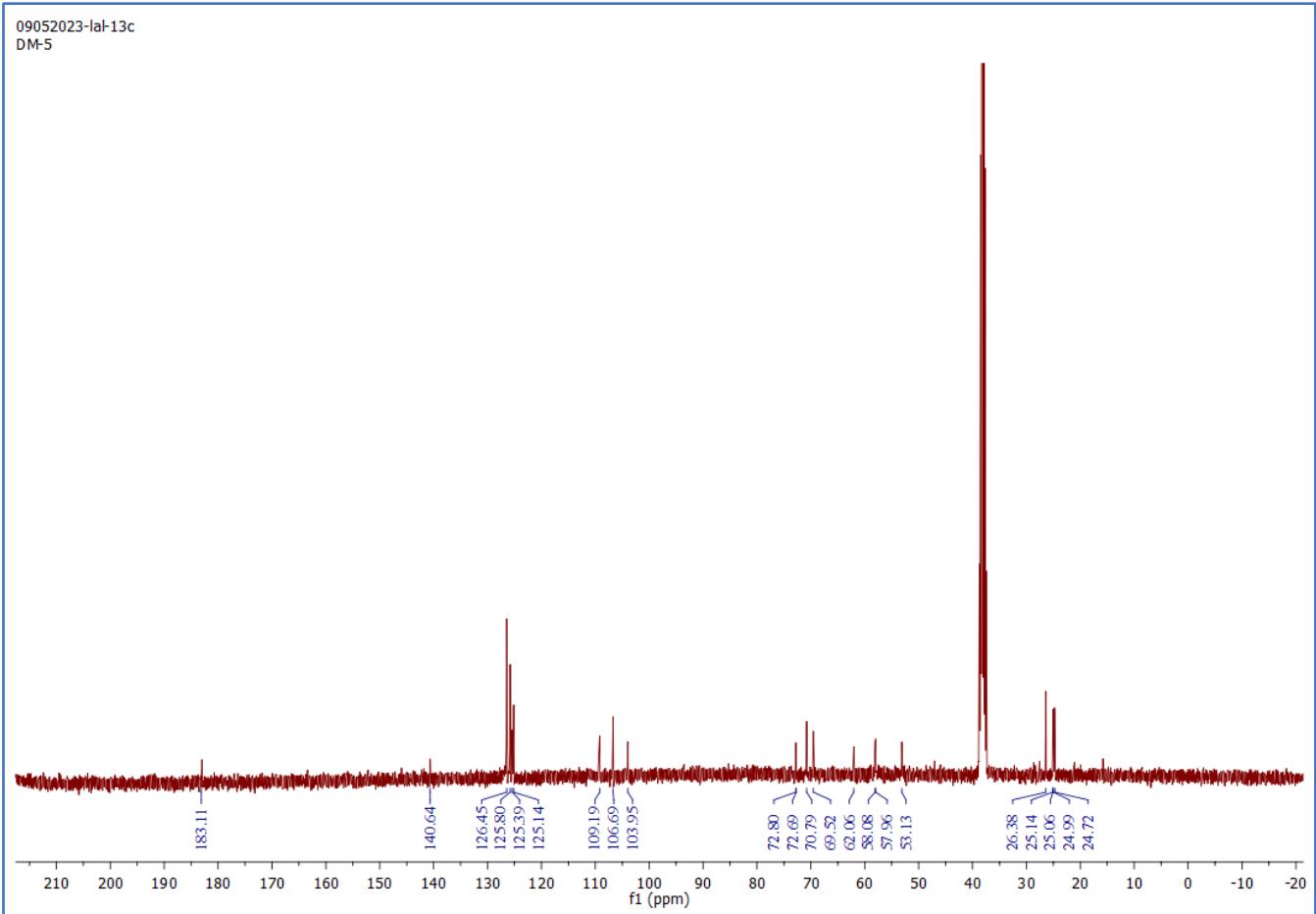
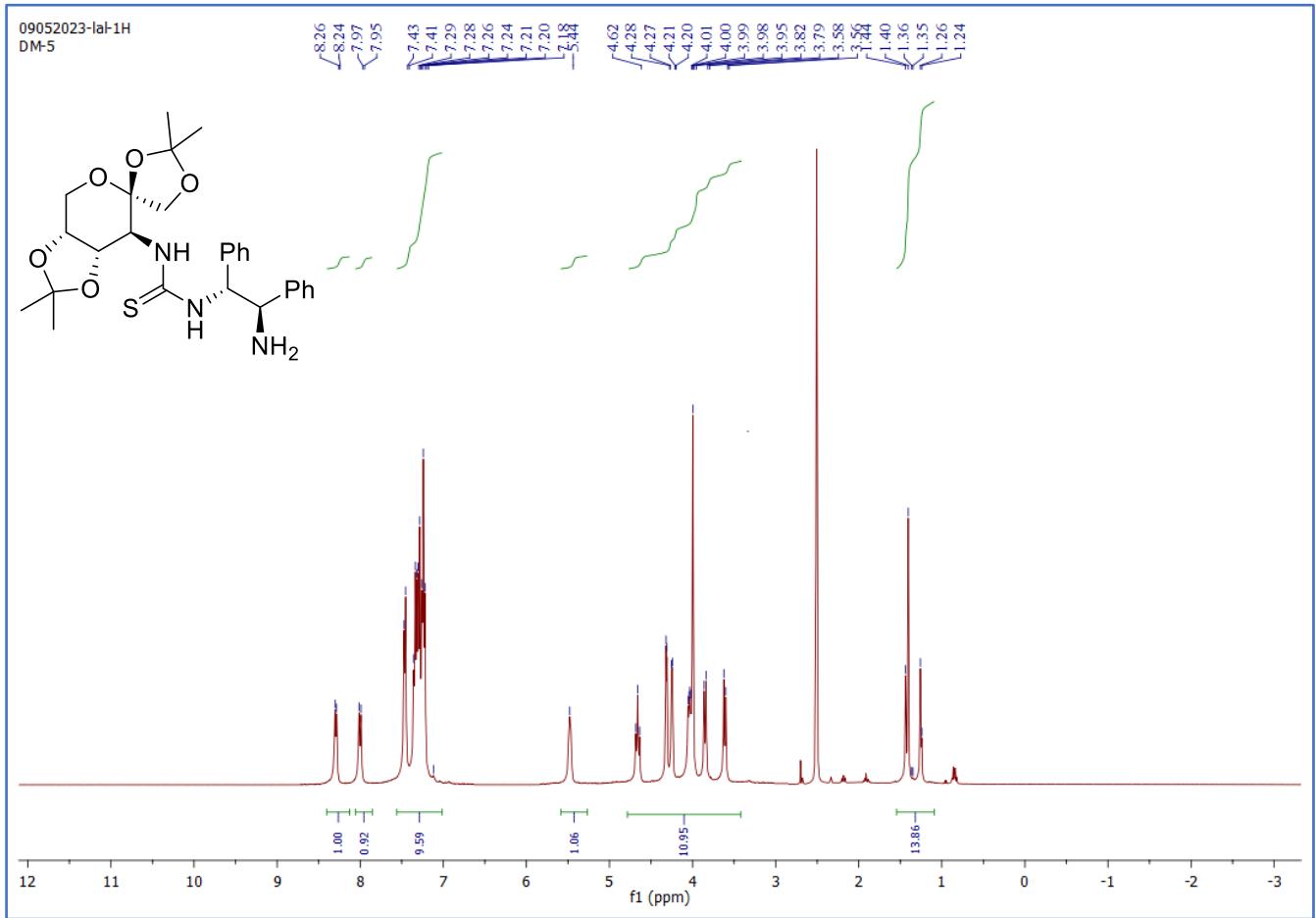


Figure S8: ^1H NMR and ^{13}C NMR of **6b**.

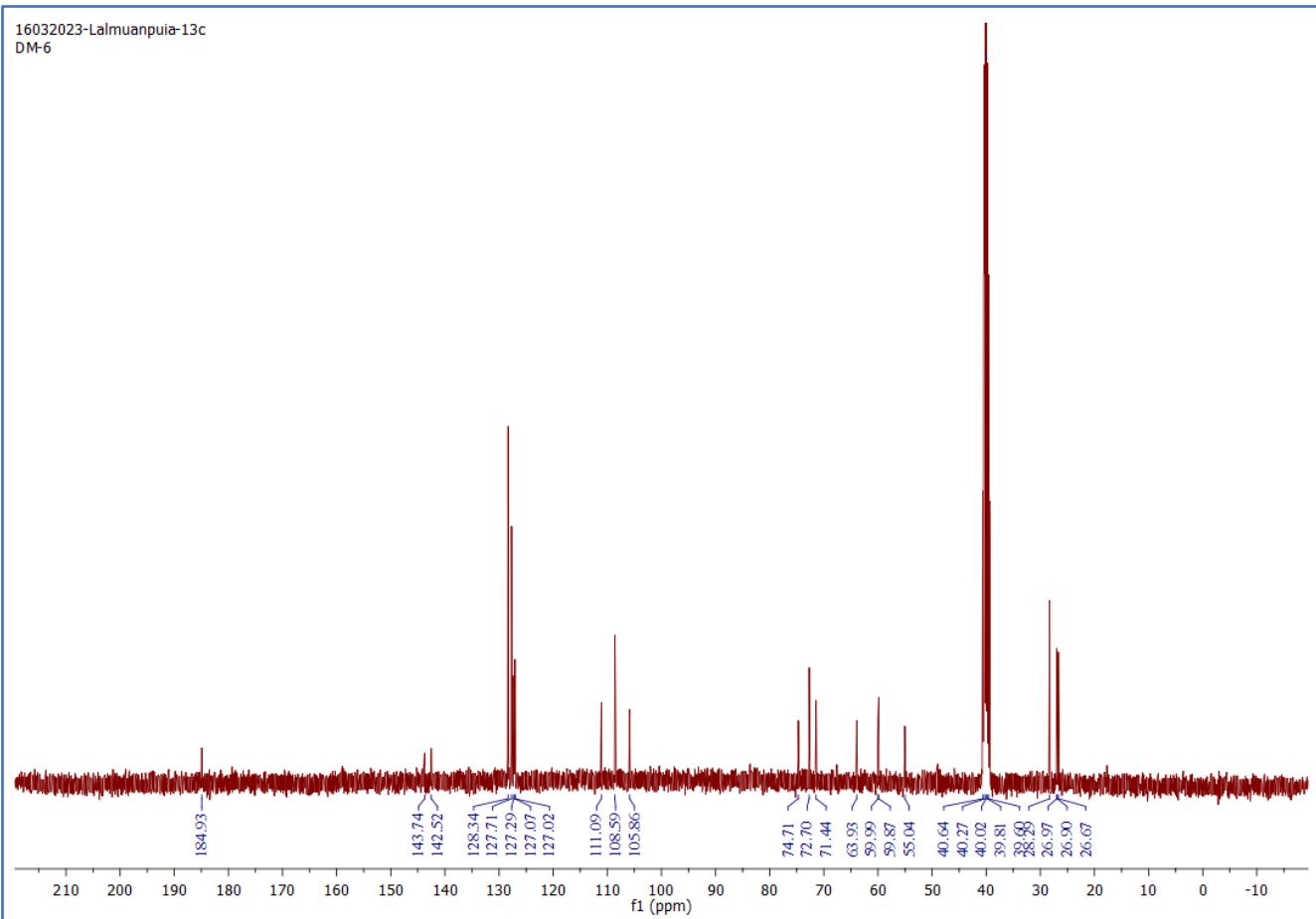
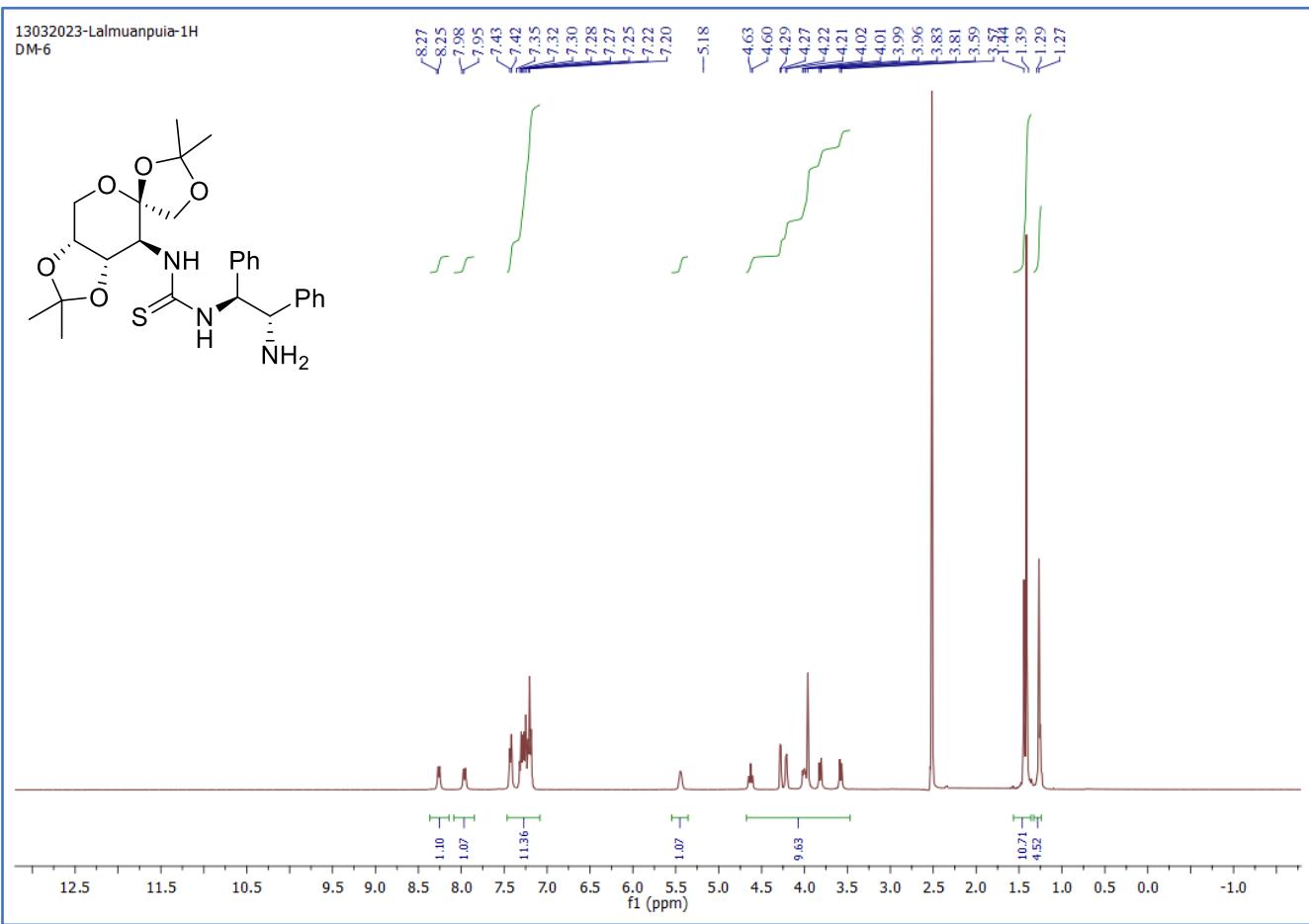


Figure S9: HRMS data of 3.

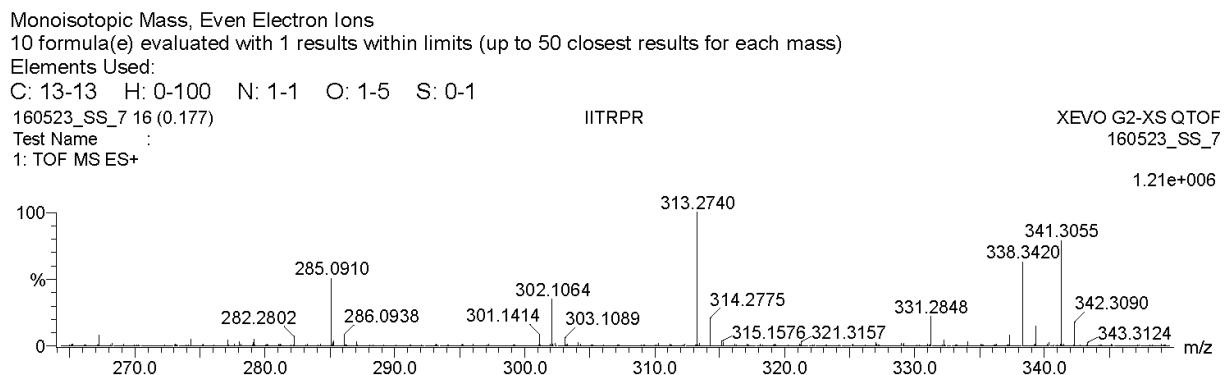


Figure S10: HRMS data of 4.

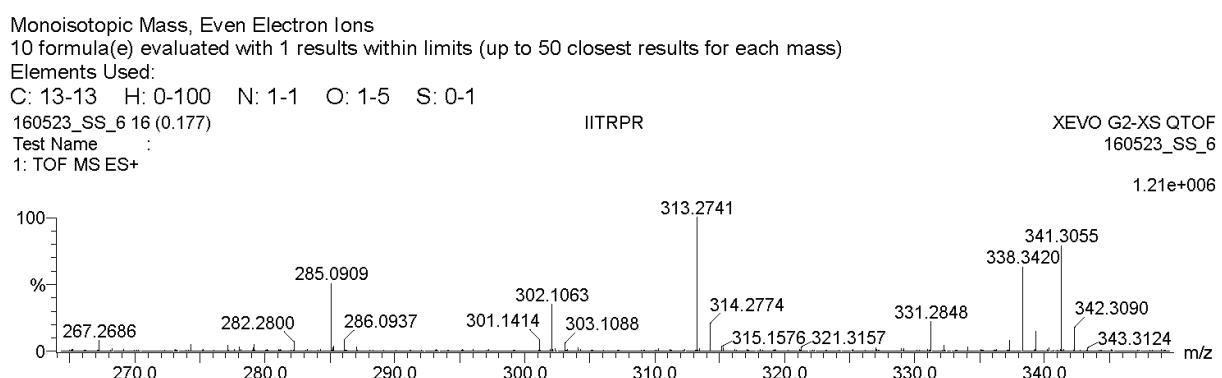


Figure S11: HRMS data of 5a.

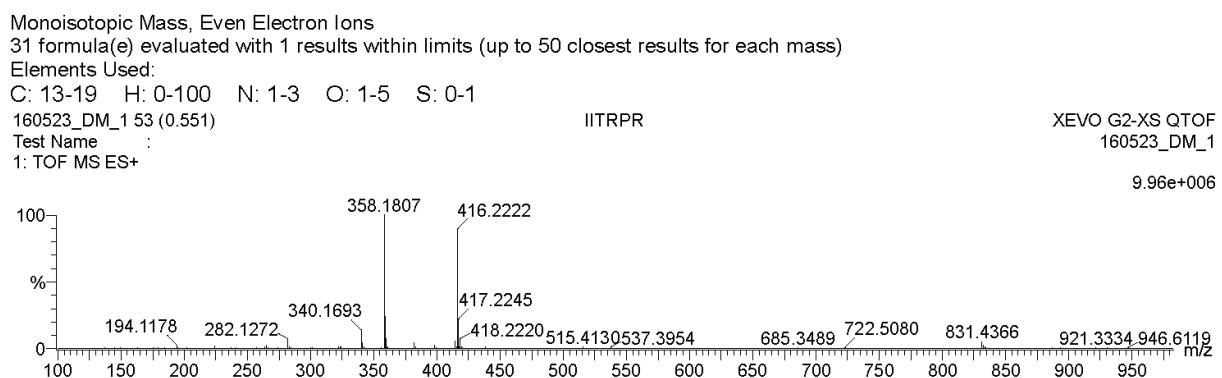


Figure S12: HRMS data of 5b.

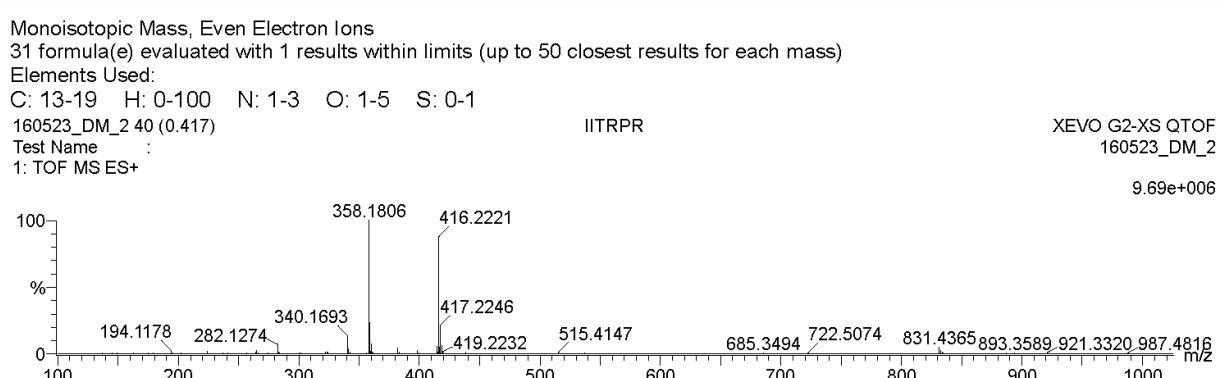


Figure S13: HRMS data of **5c**.

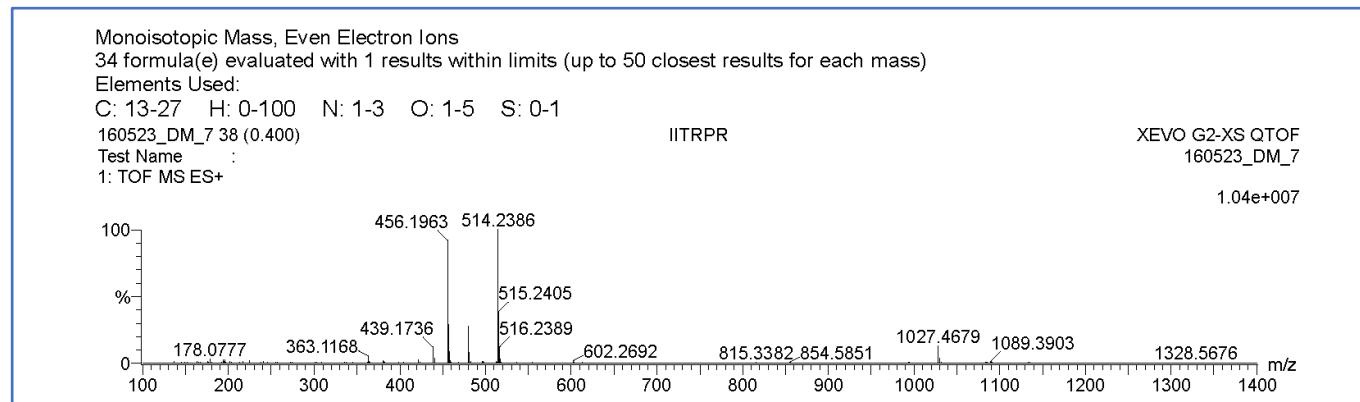


Figure S14: HRMS data of **5d**.

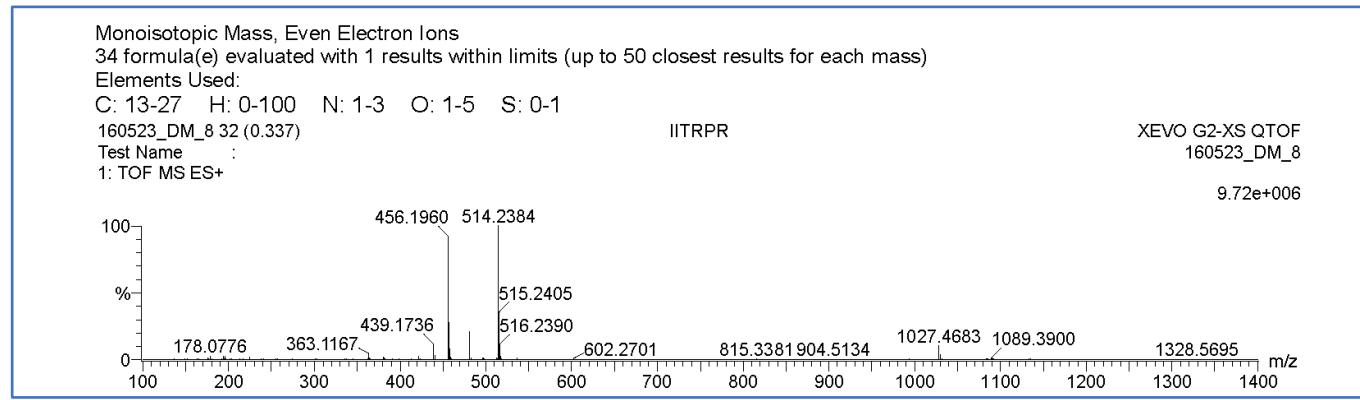


Figure S15: HRMS data of **6a**.

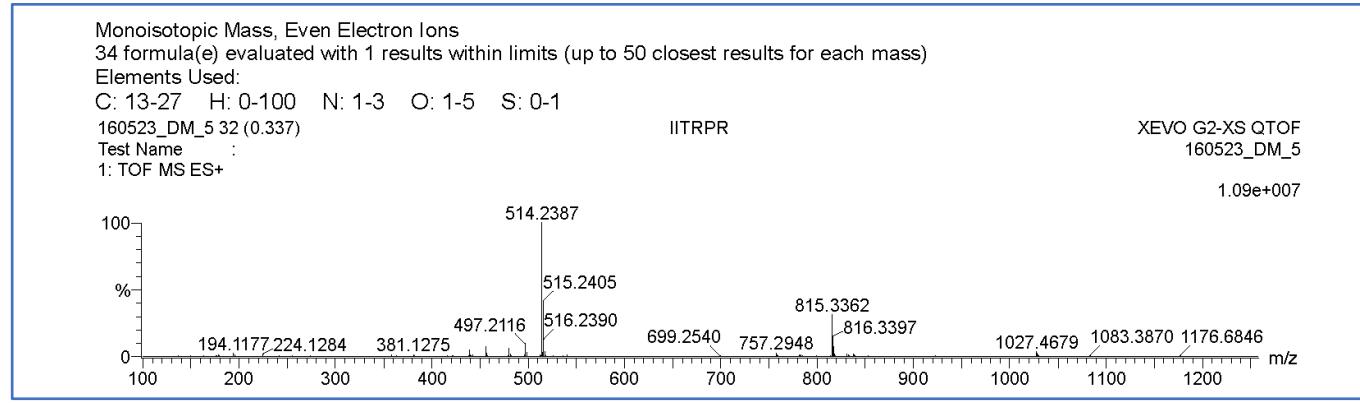


Figure S16: HRMS data of **6b**.

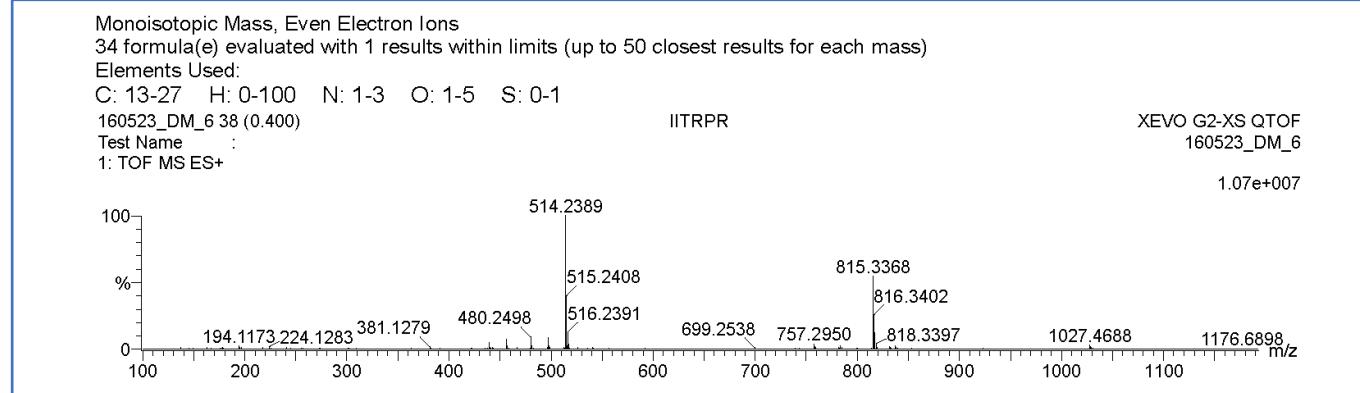


Figure S17: ^1H NMR and ^{13}C NMR of **8a**.

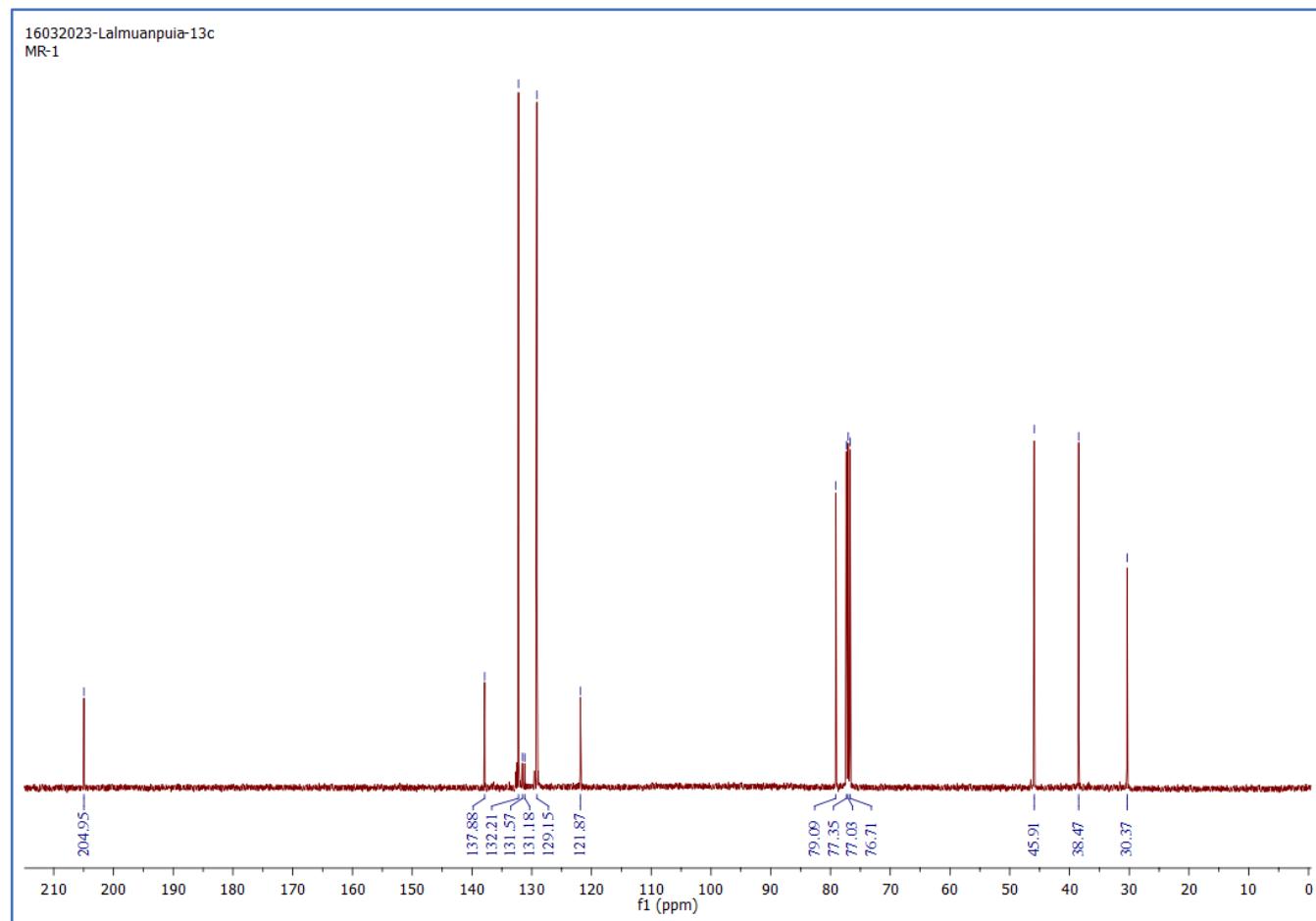
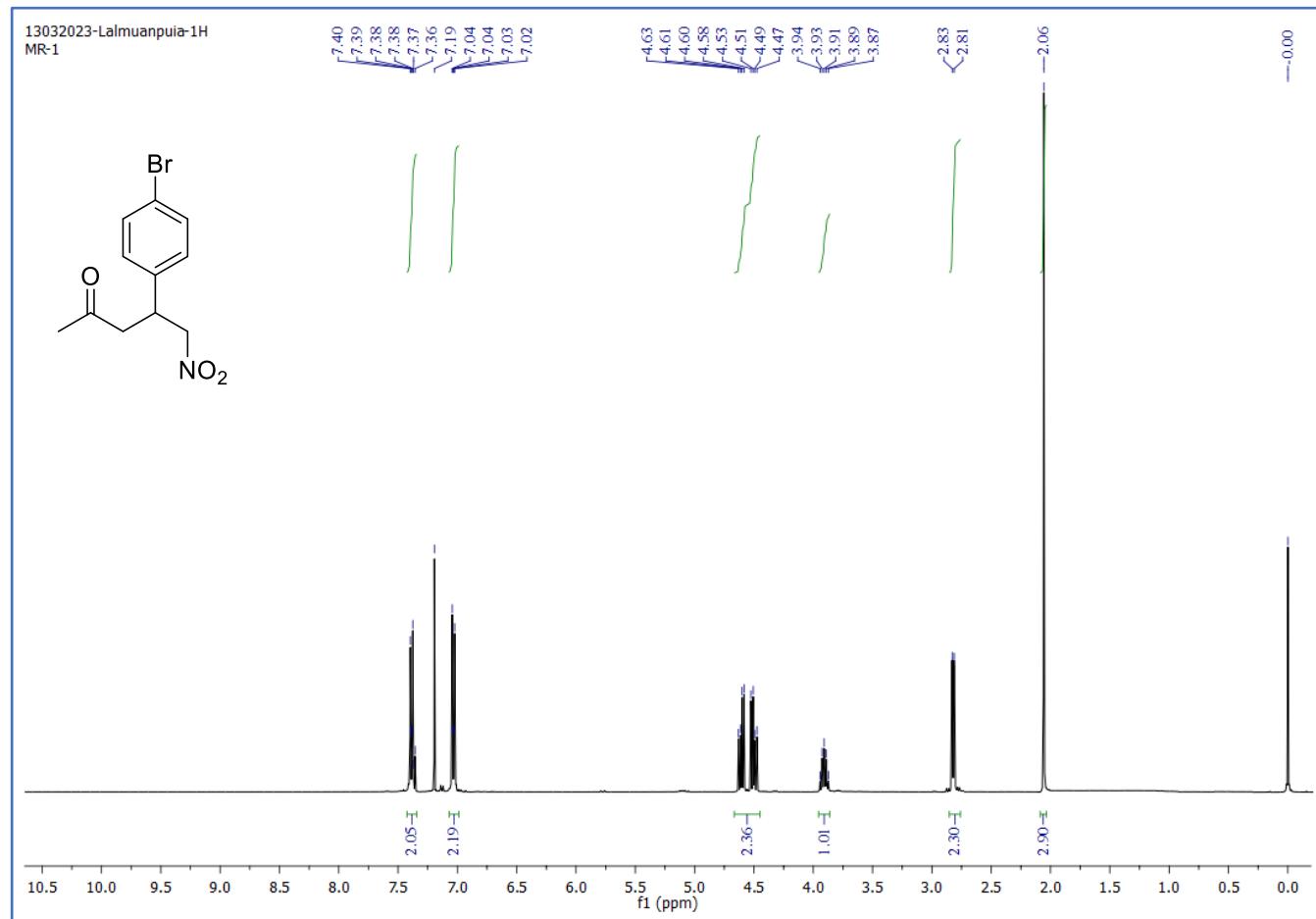


Figure S18: ^1H NMR and ^{13}C NMR of **8b**.

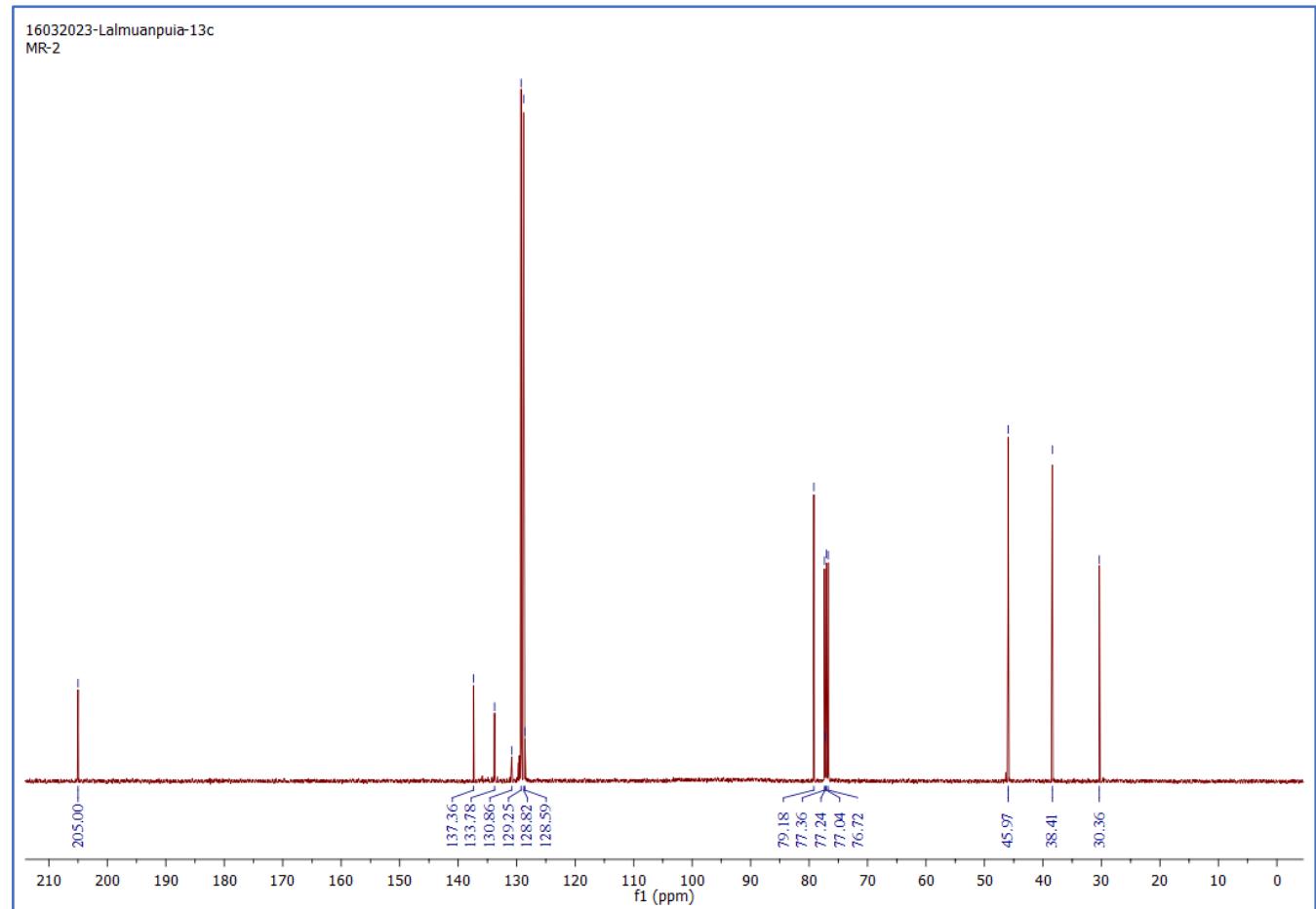
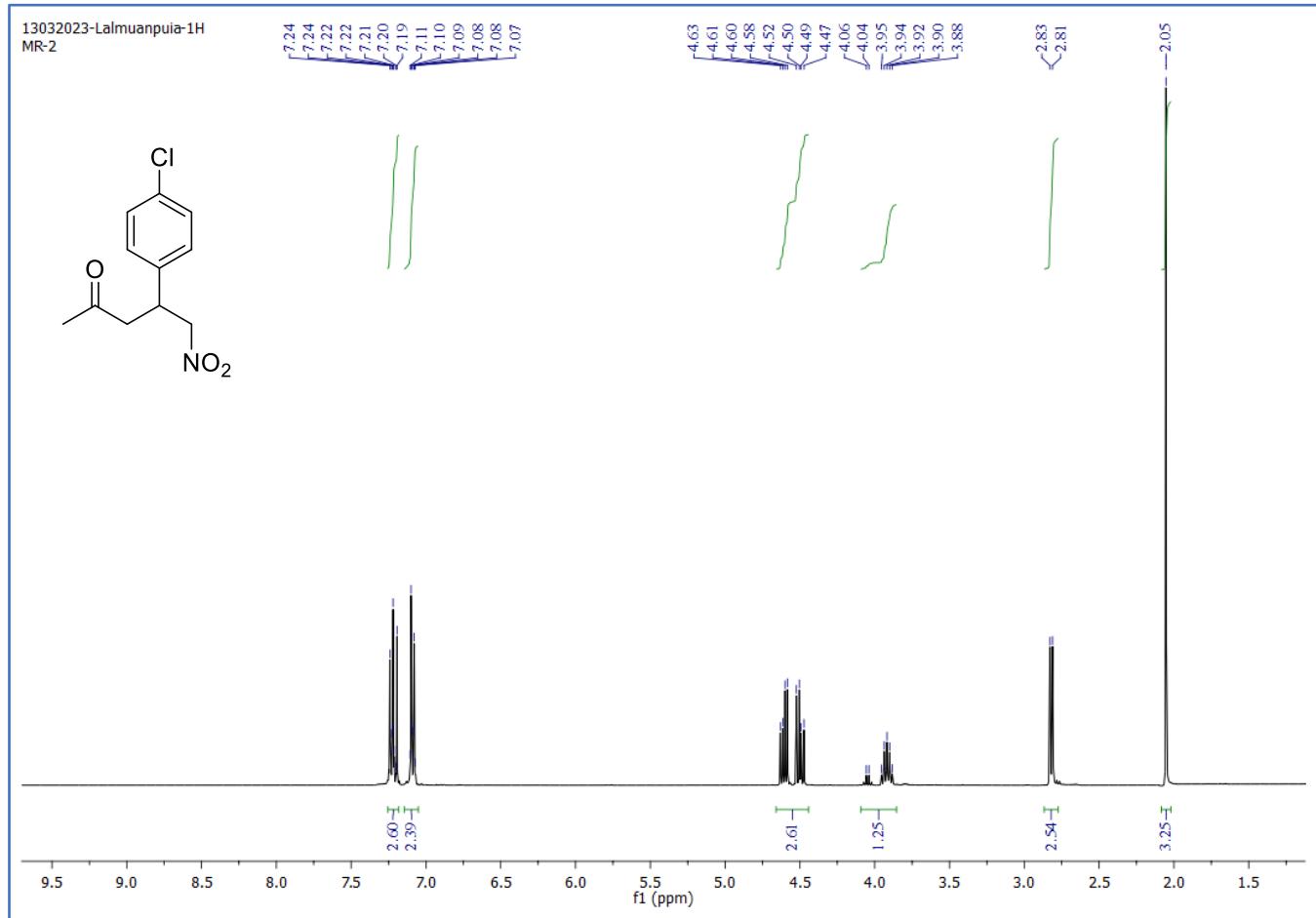


Figure S19: ^1H NMR and ^{13}C NMR of **8c**.

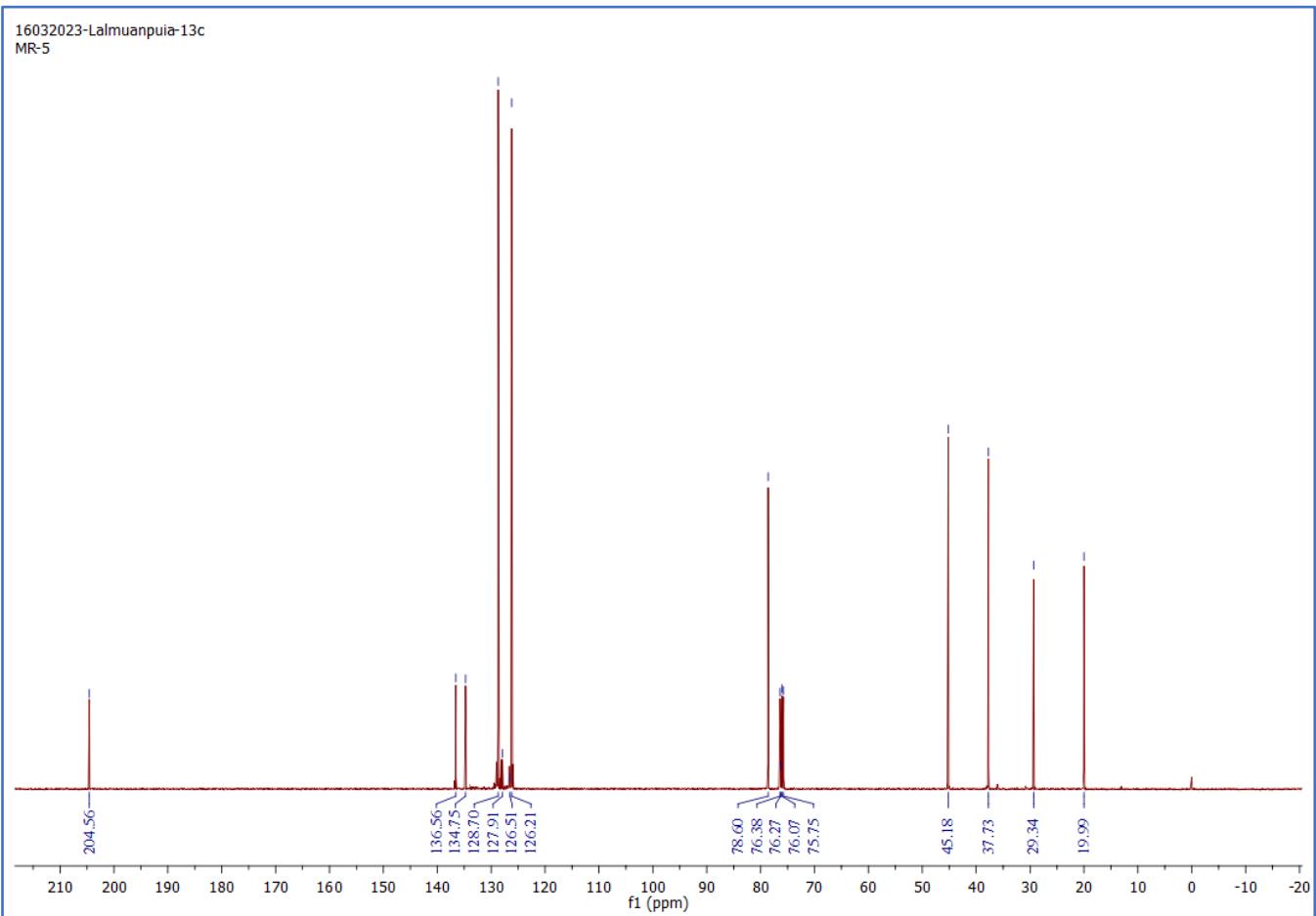
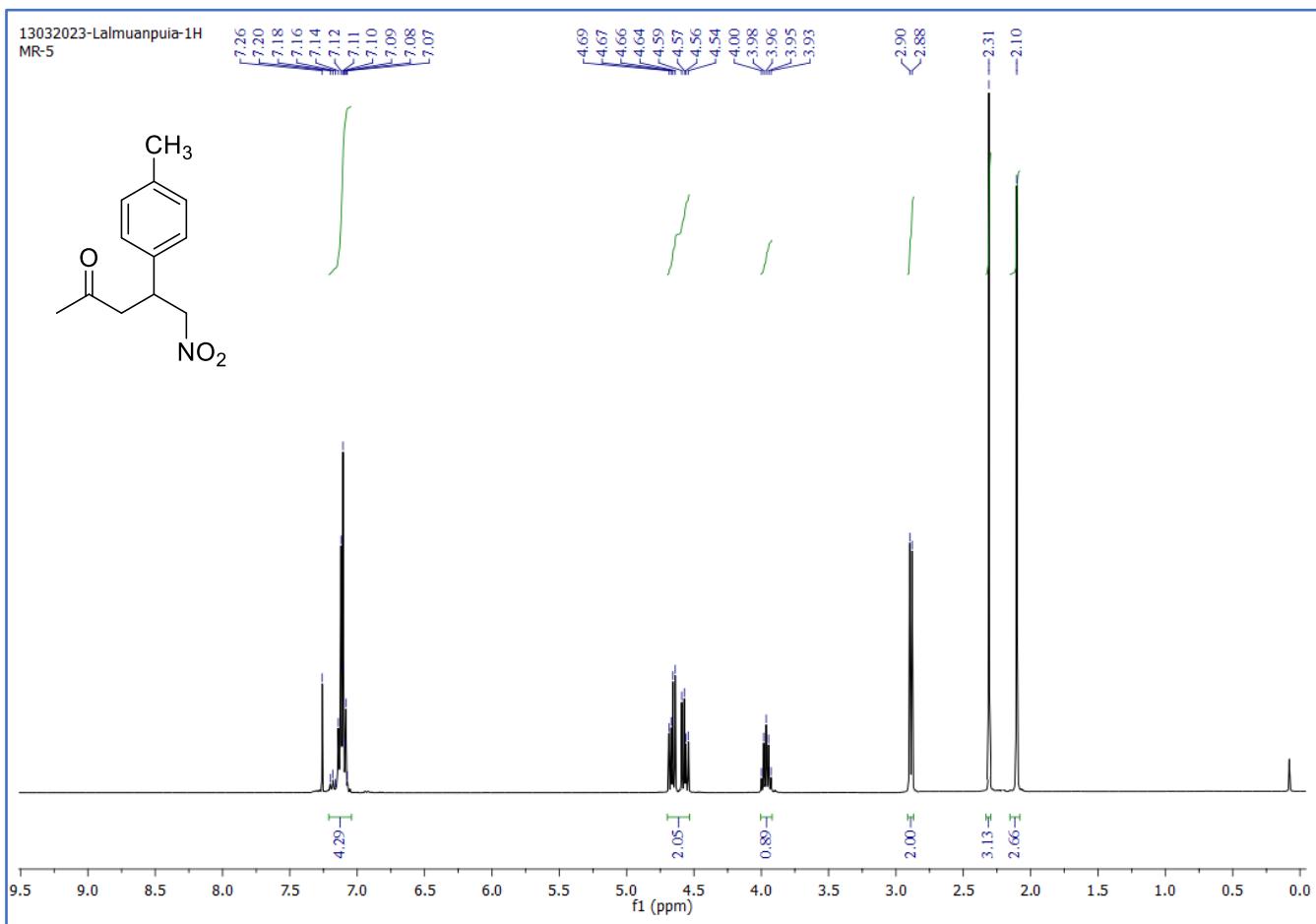


Figure S20: ^1H NMR and ^{13}C NMR of **8d**.

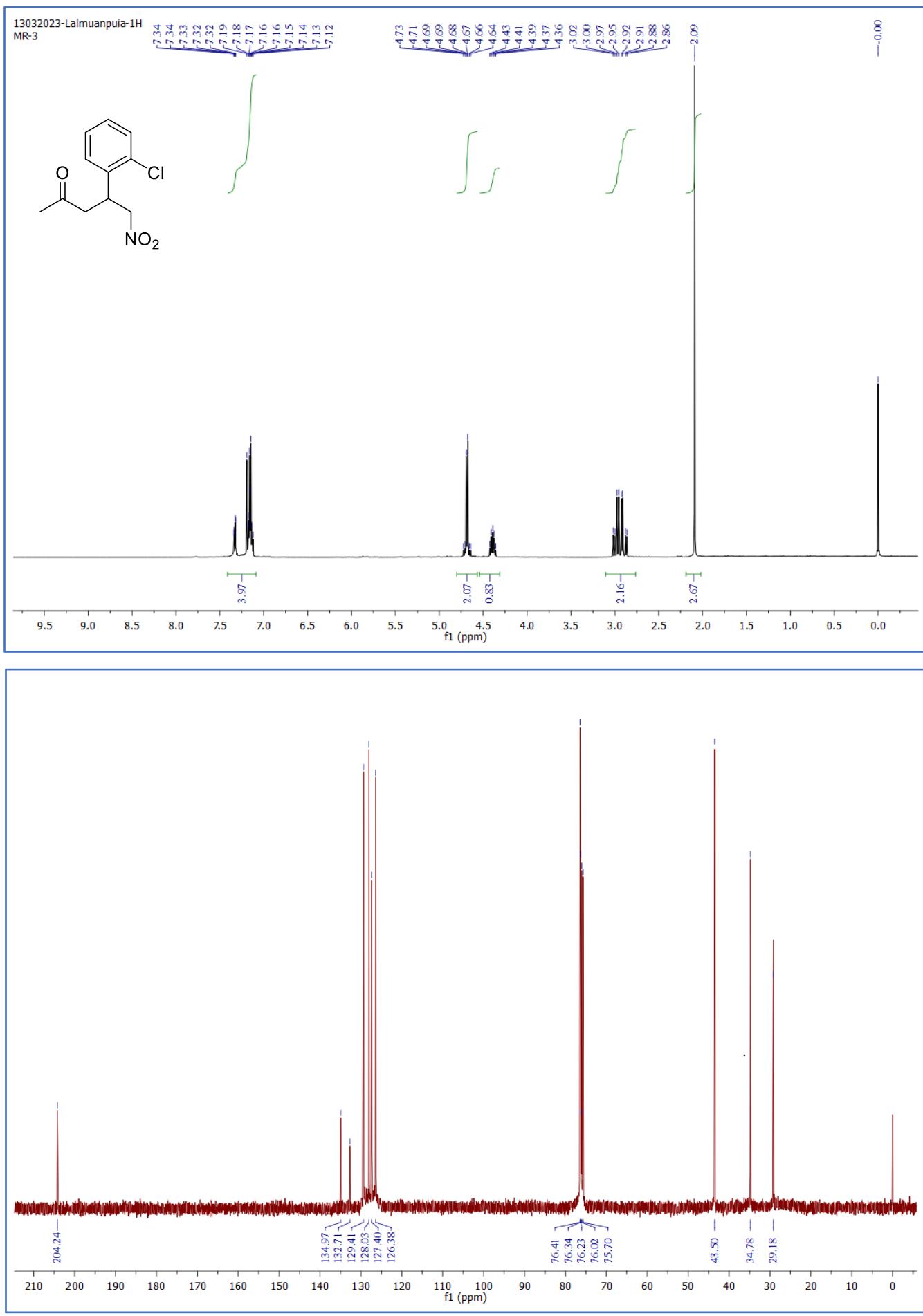


Figure S21: ^1H NMR and ^{13}C NMR of **8e**.

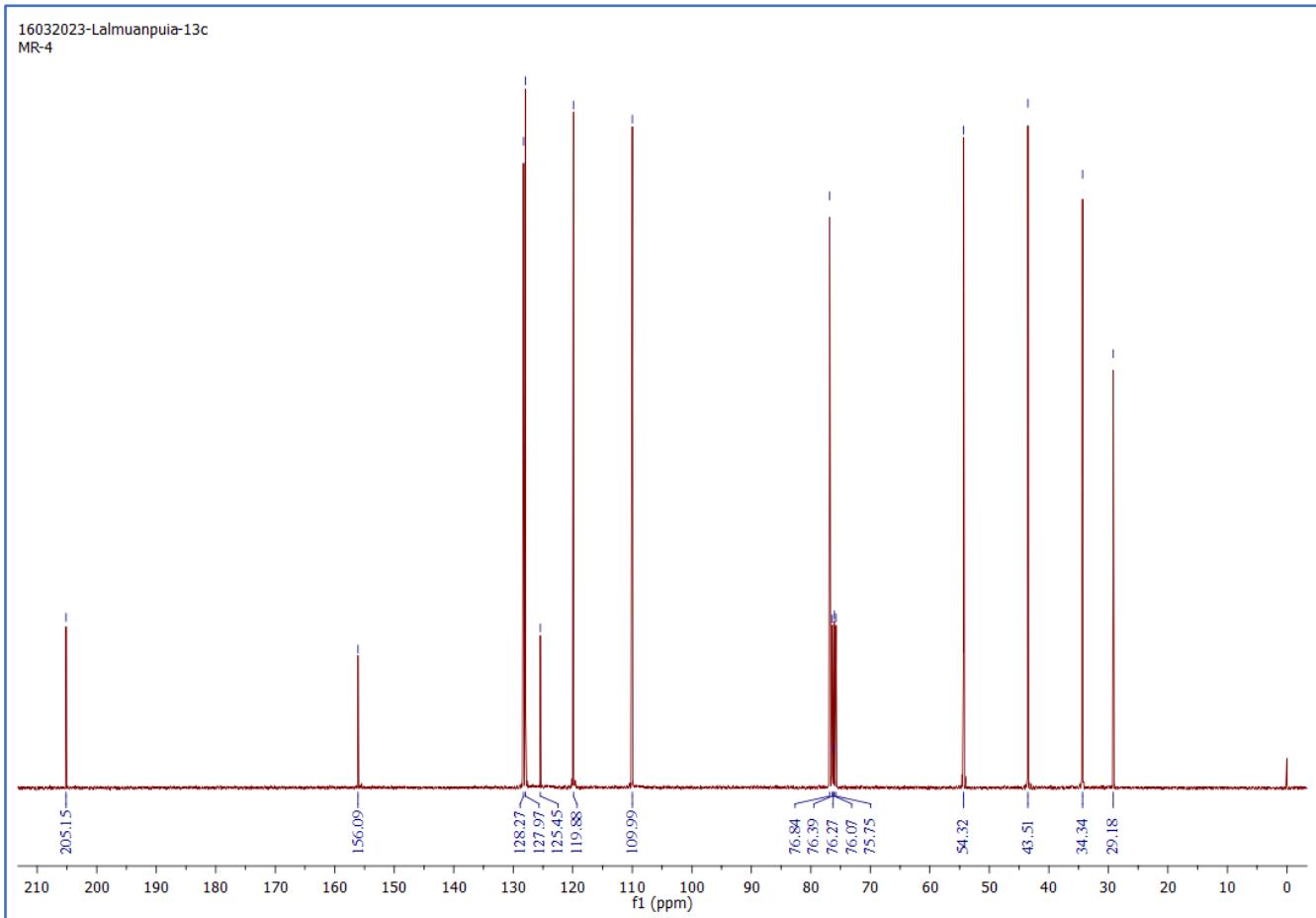
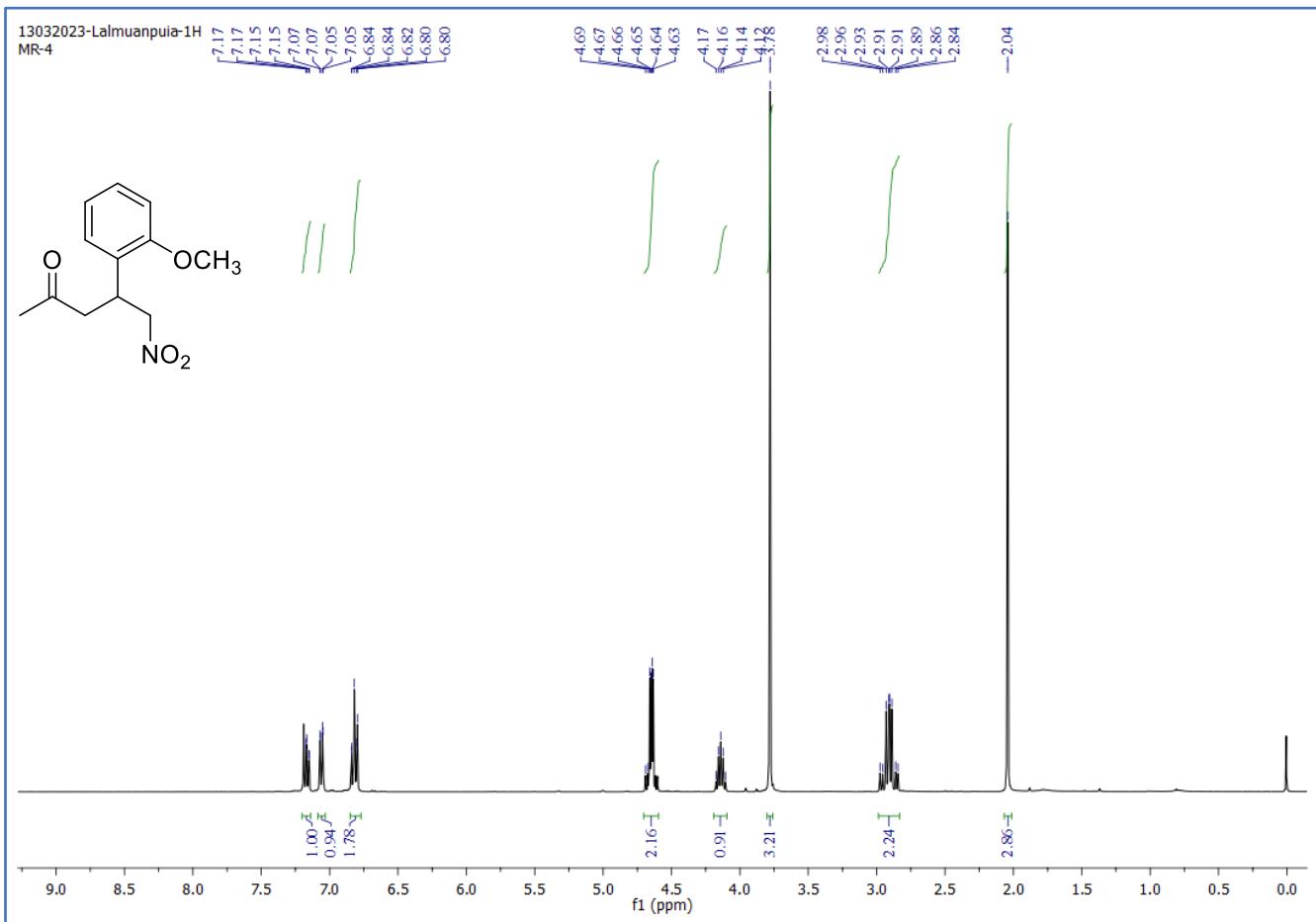


Figure S22: ^1H NMR and ^{13}C NMR of **8f**.

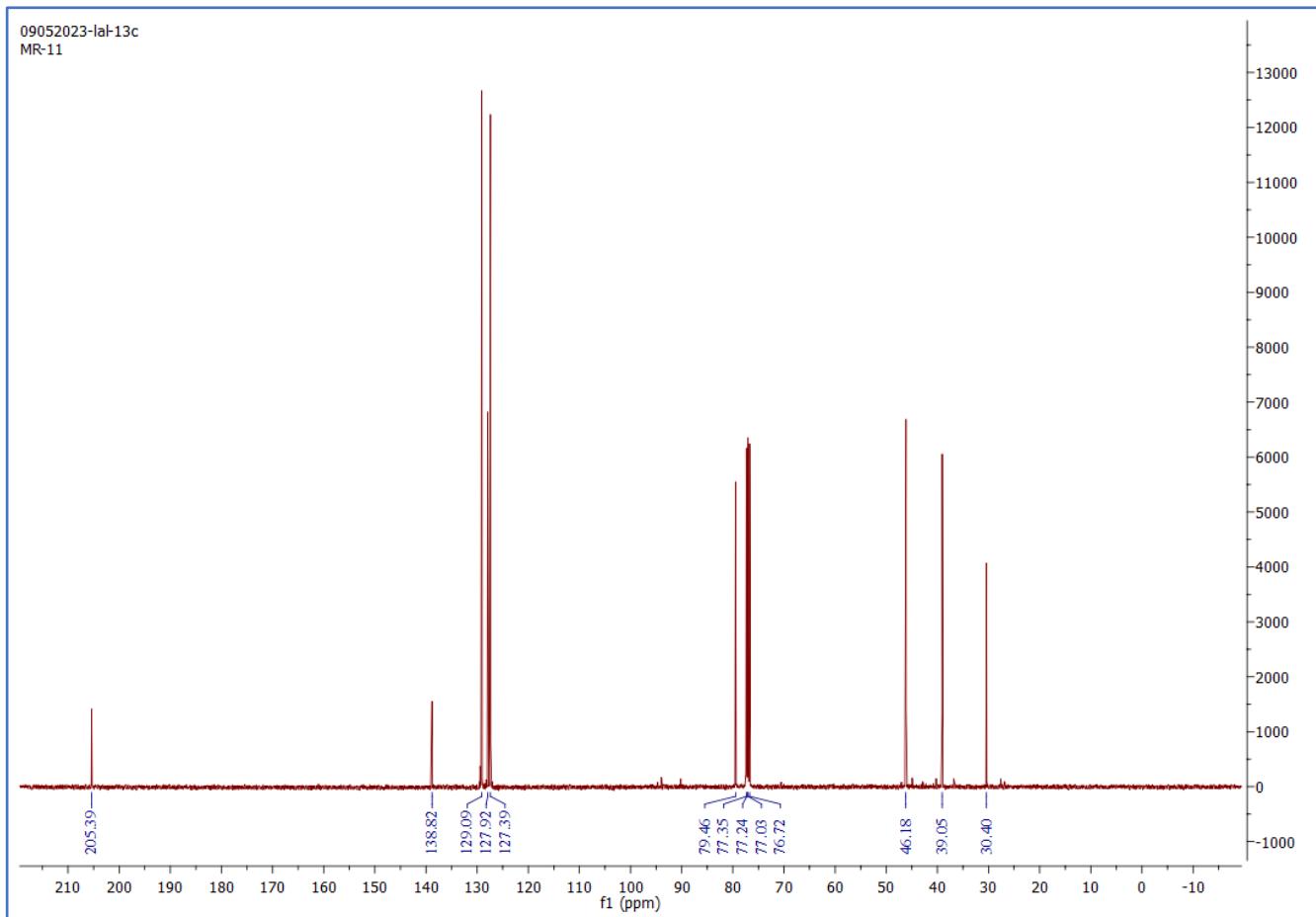
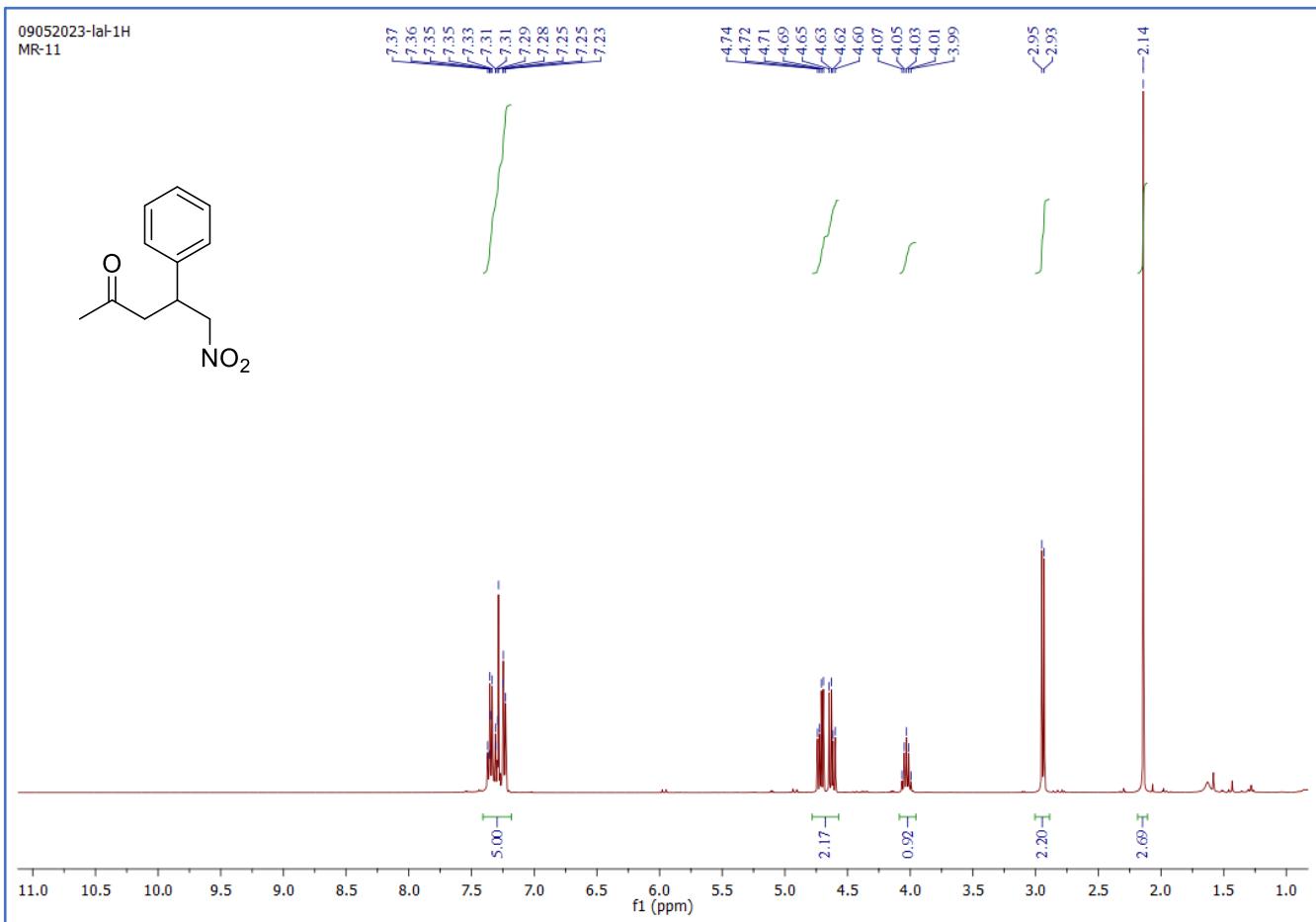


Figure S23: ^1H NMR and ^{13}C NMR of **8g**.

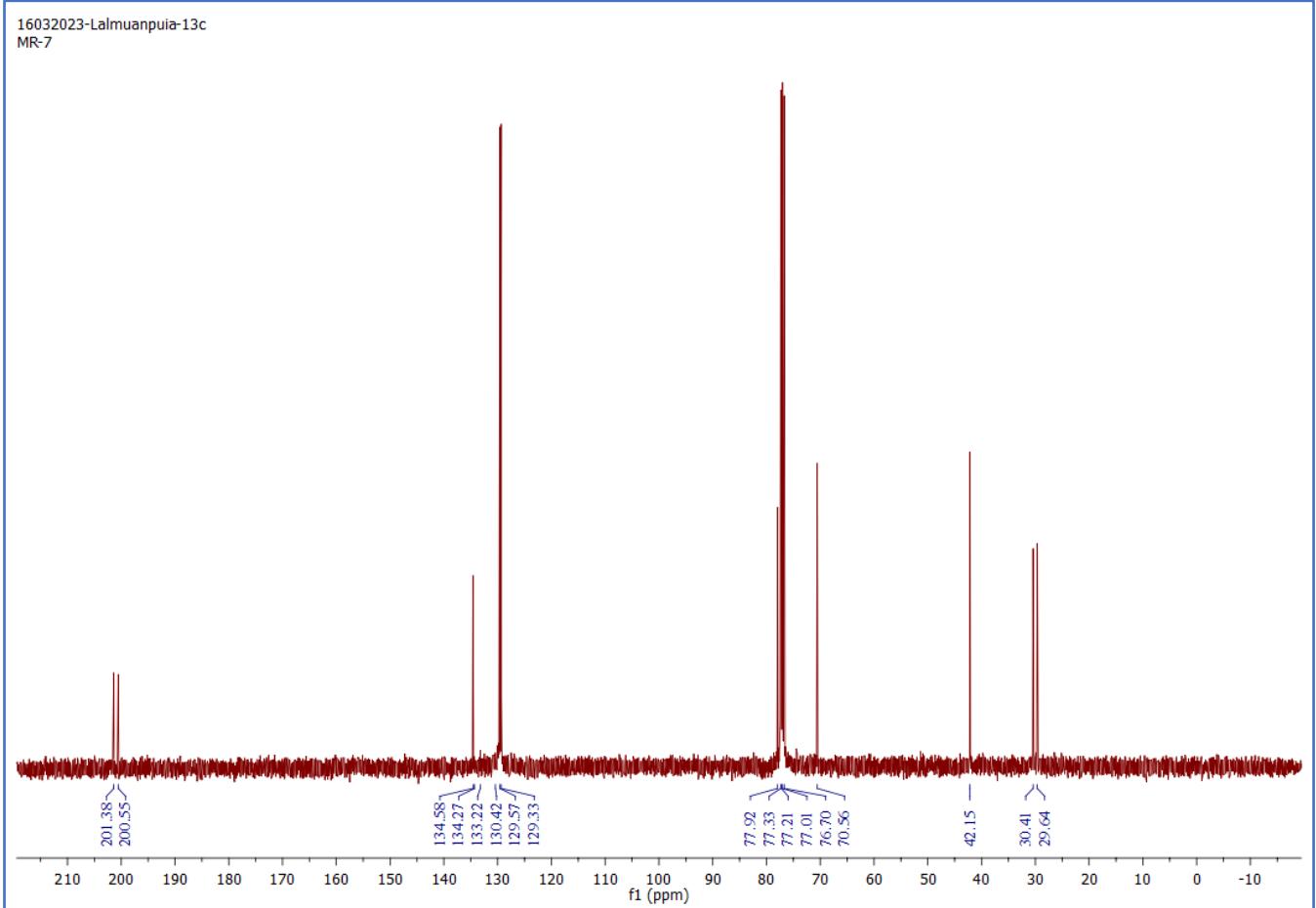
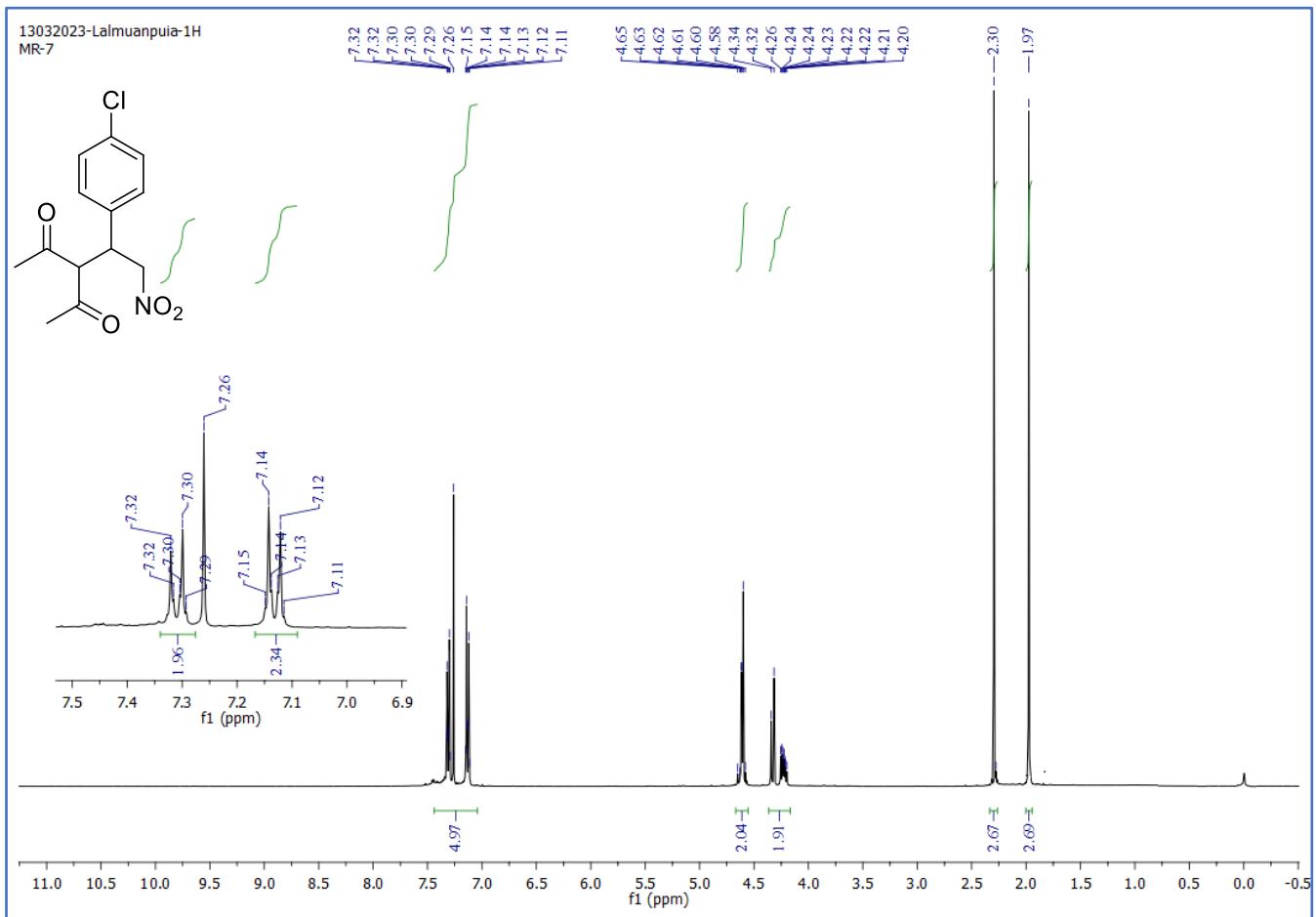


Figure S24: ^1H NMR and ^{13}C NMR of **8h**.

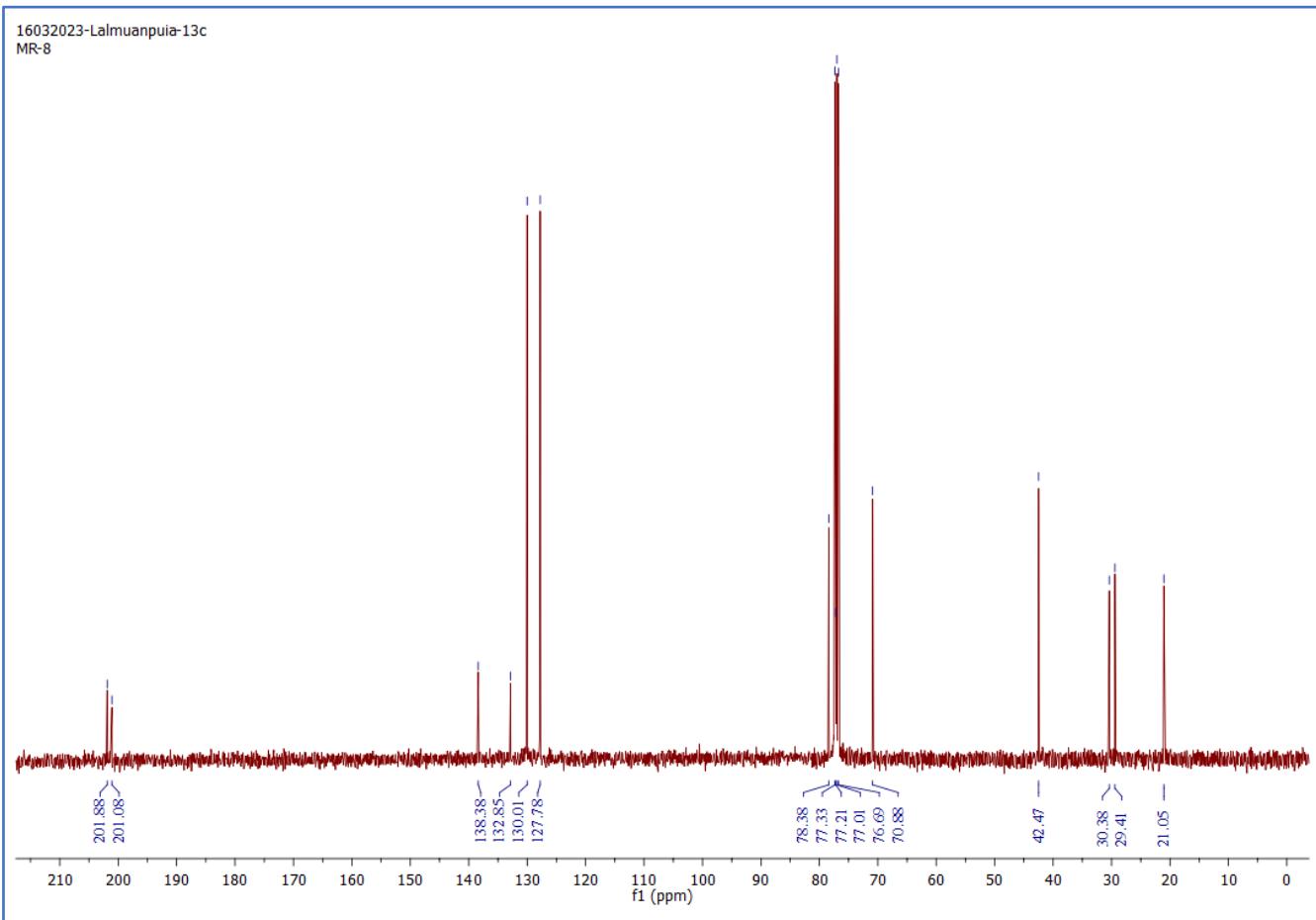
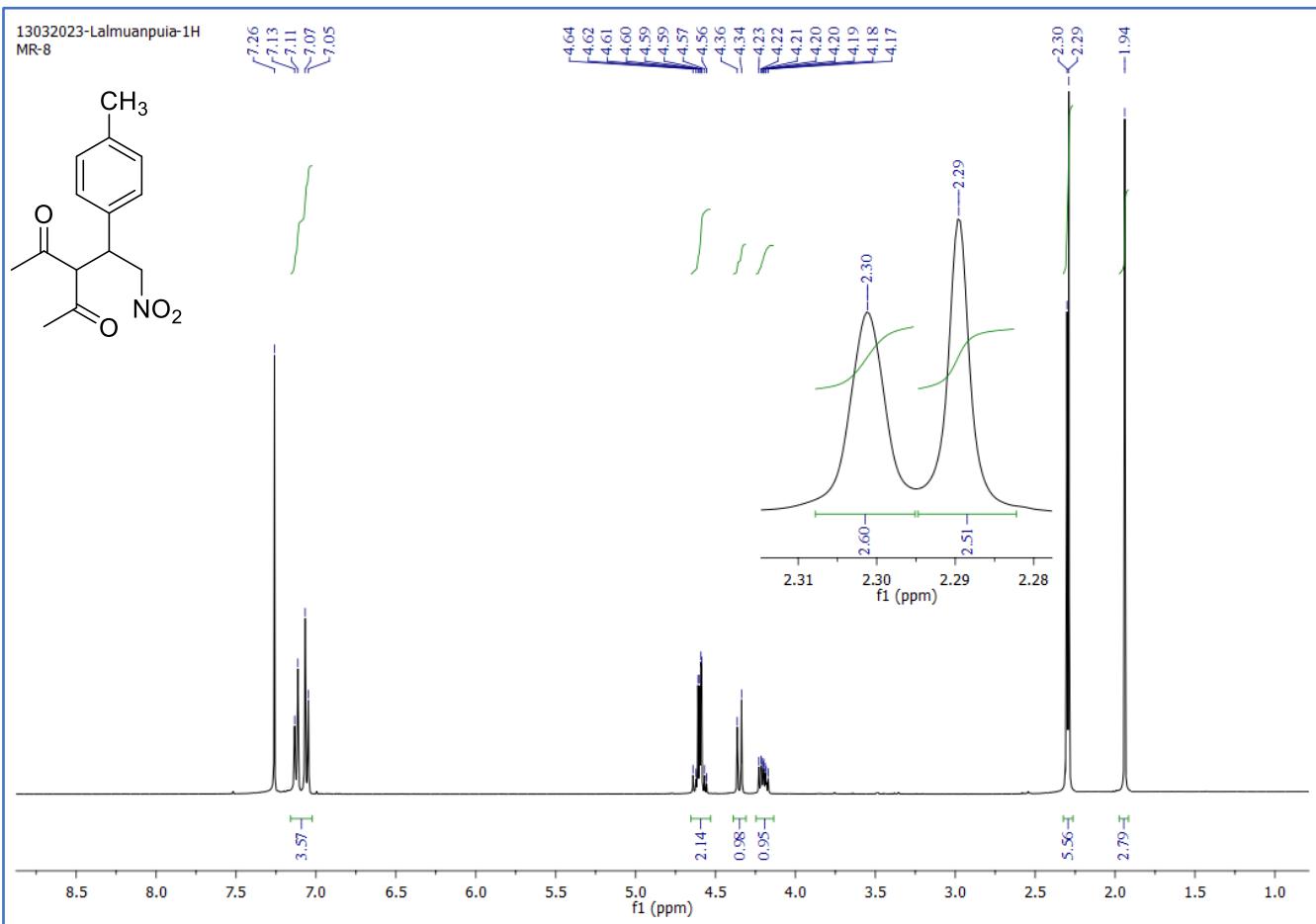


Figure S25: ^1H NMR and ^{13}C NMR of **8i**.

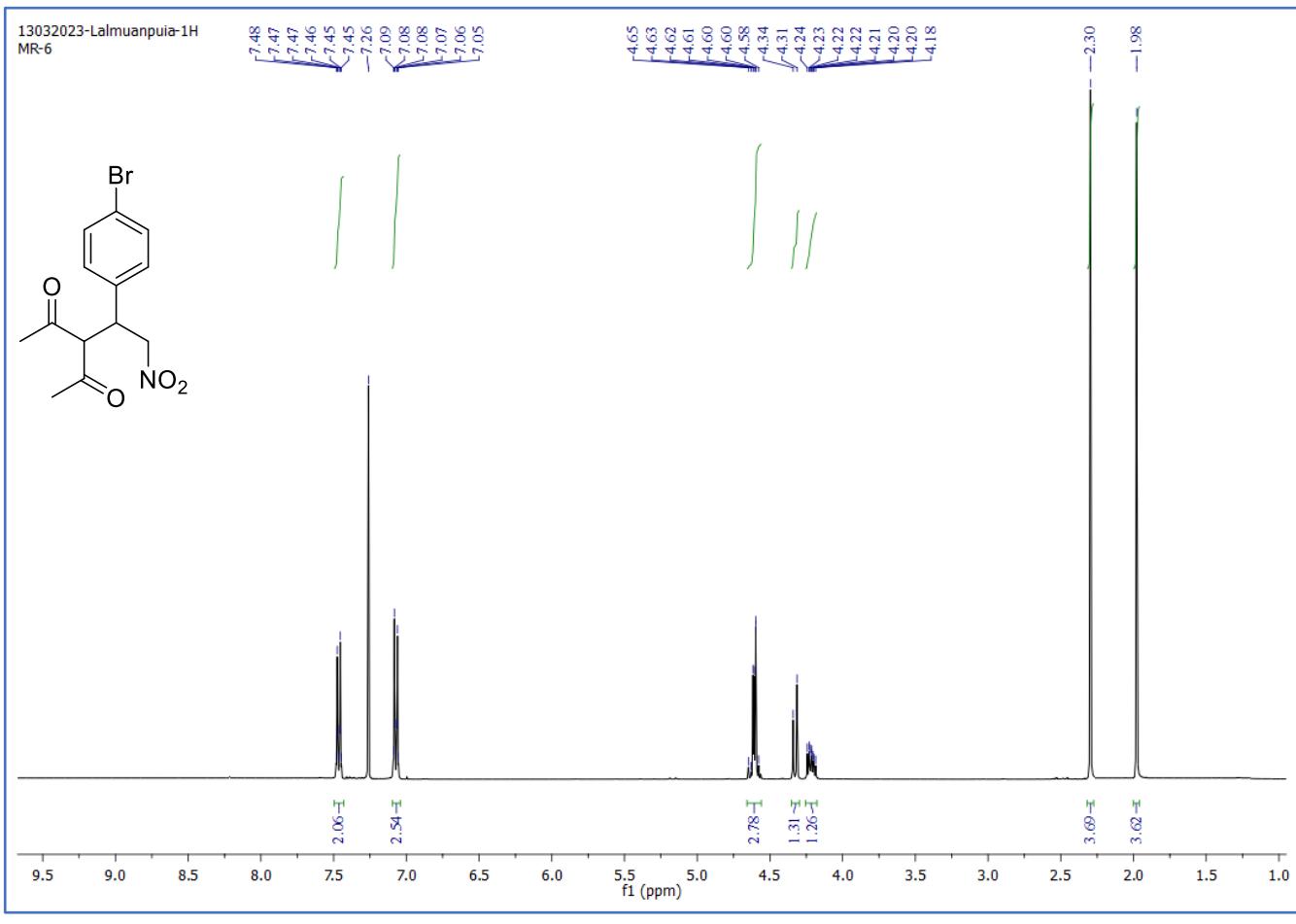


Figure S26: ^1H NMR and ^{13}C NMR of **8j**.

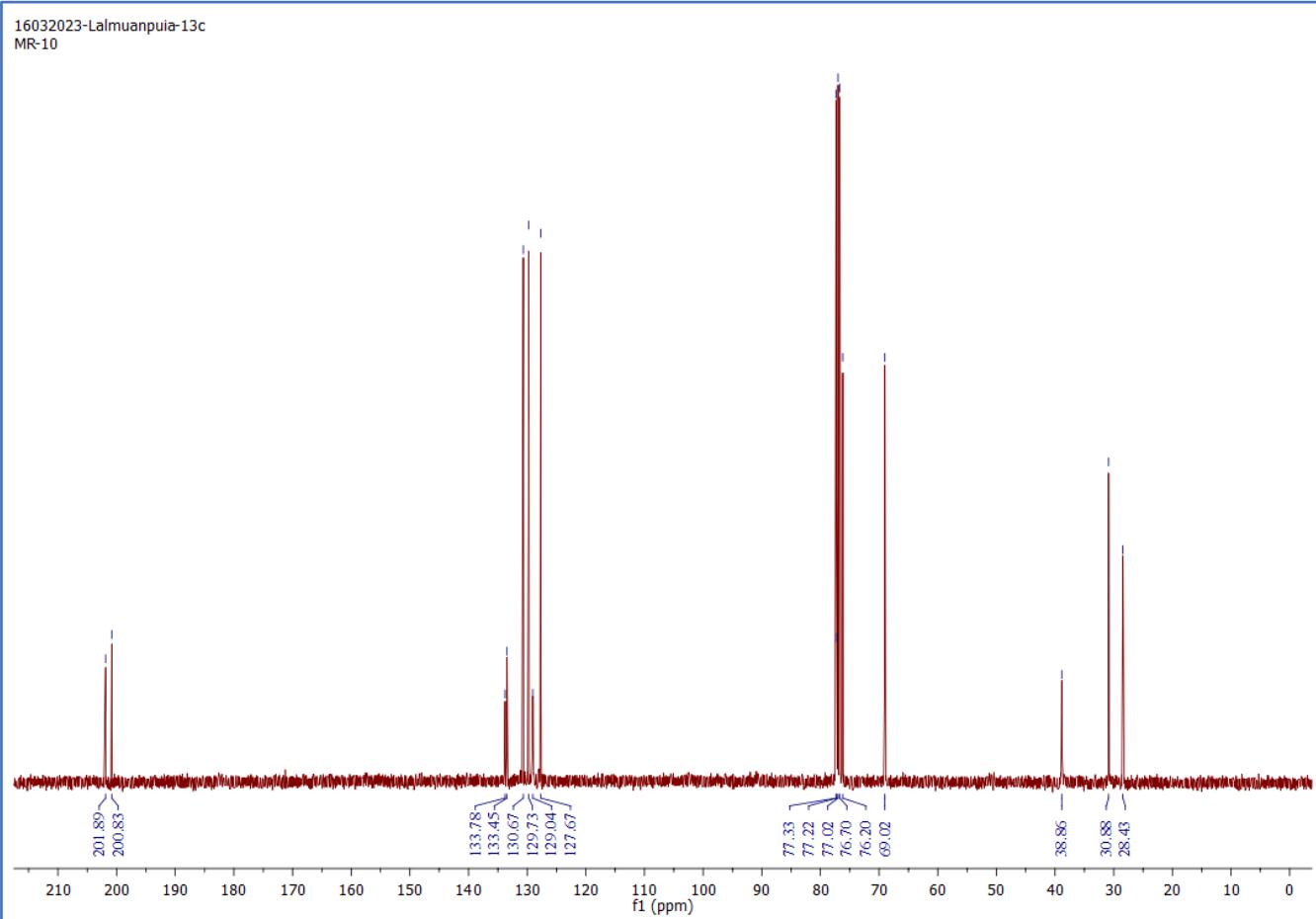
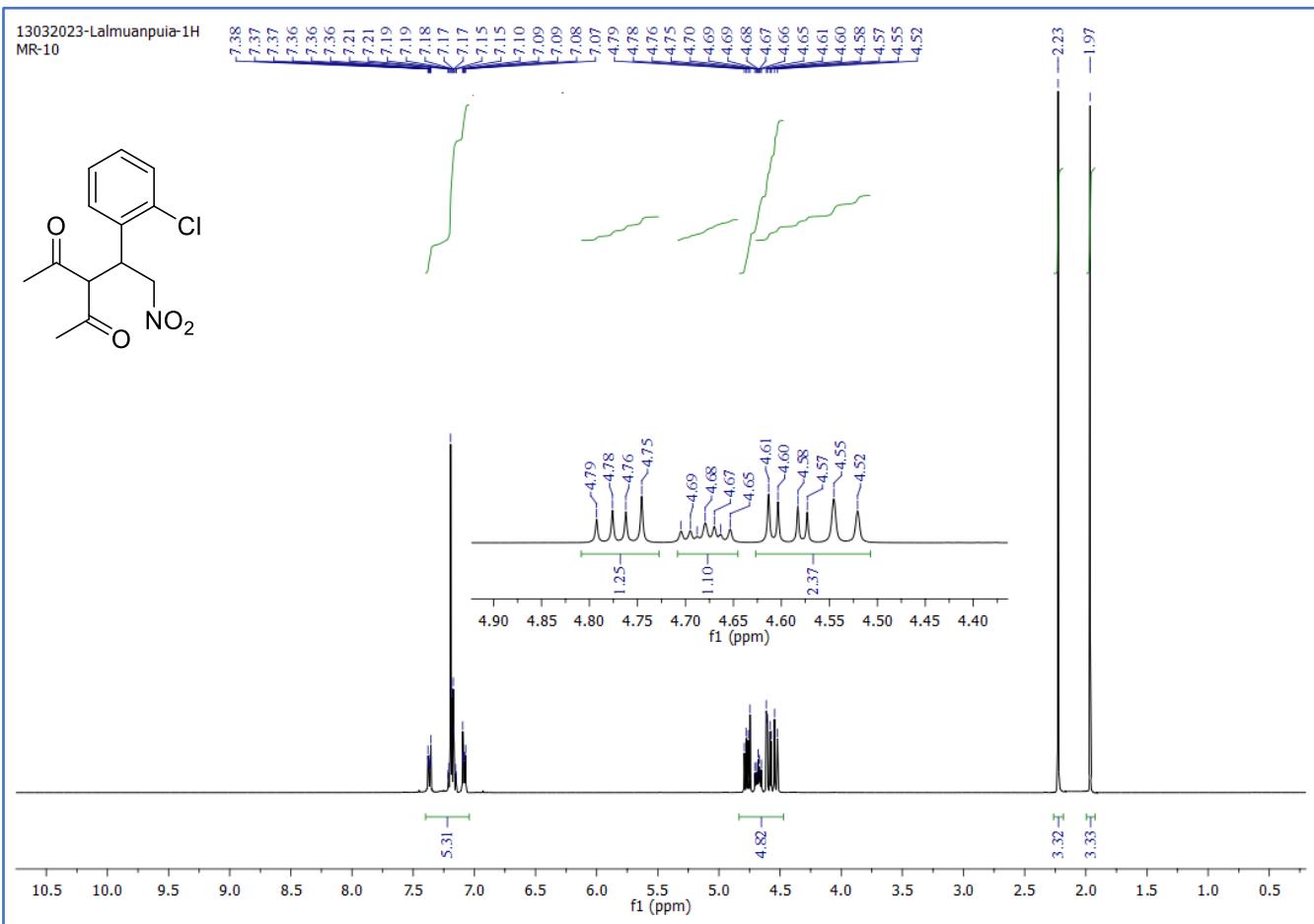


Figure S27: ^1H NMR and ^{13}C NMR of **8k**.

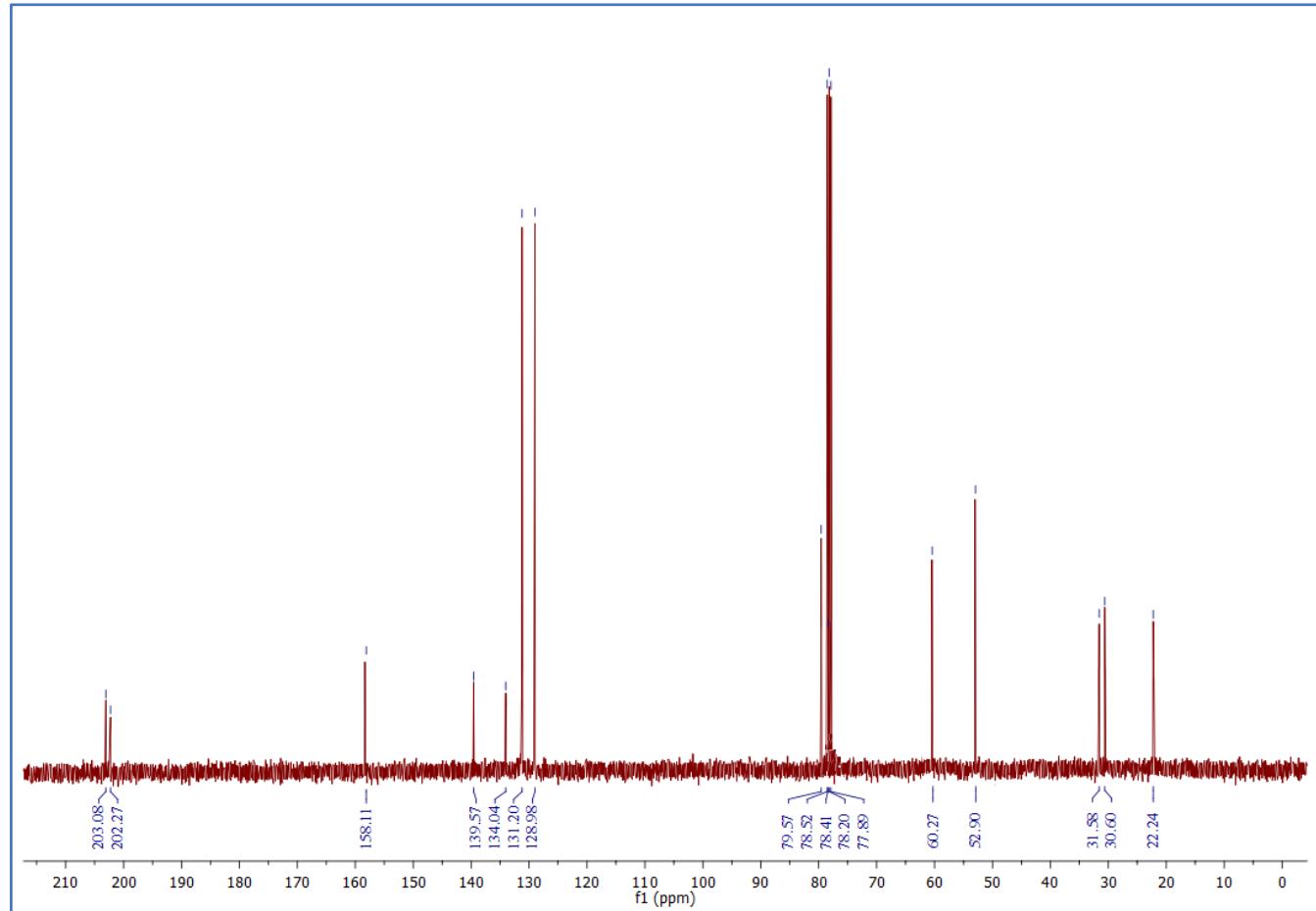
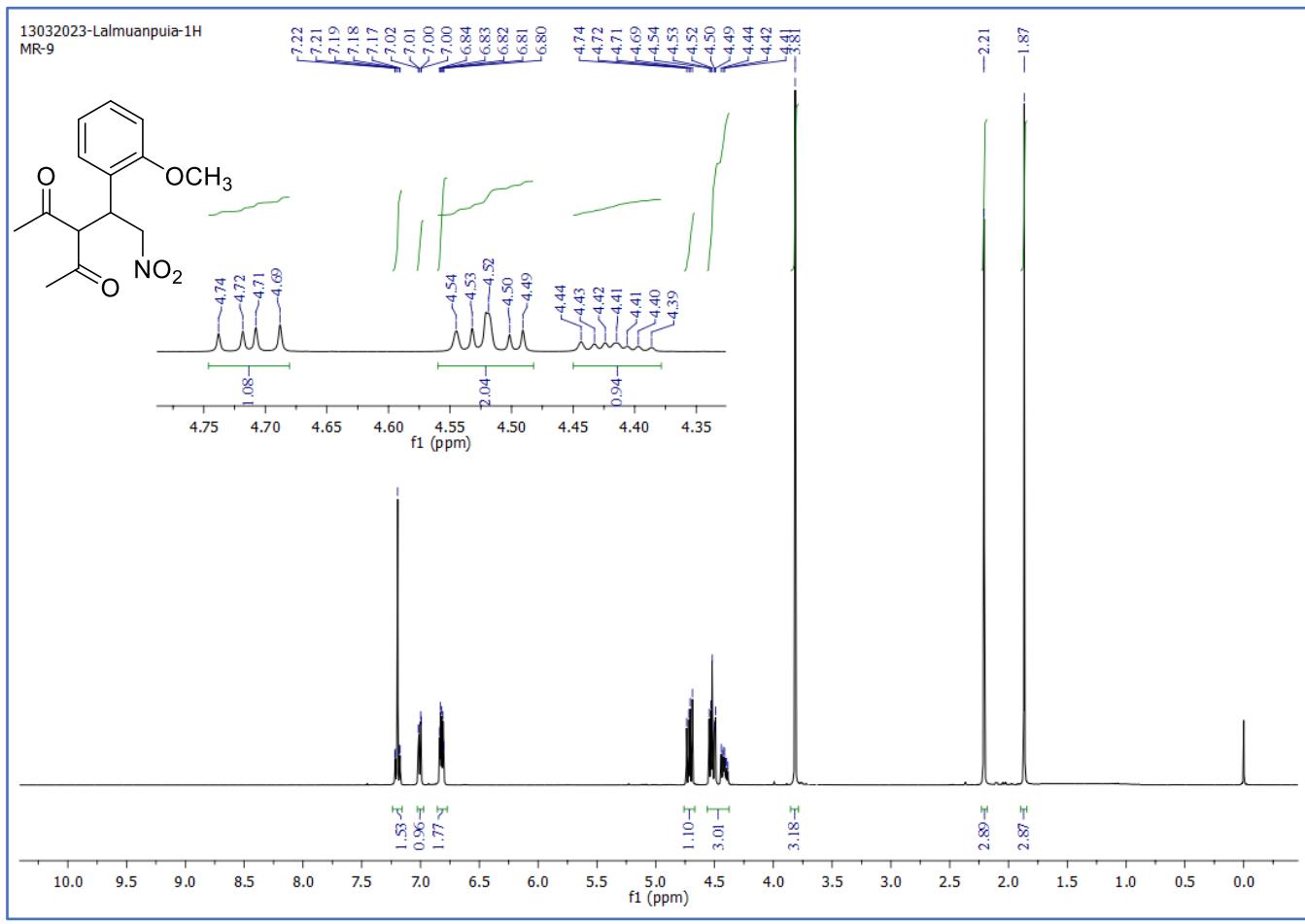


Figure S28: ^1H NMR and ^{13}C NMR of **8l**.

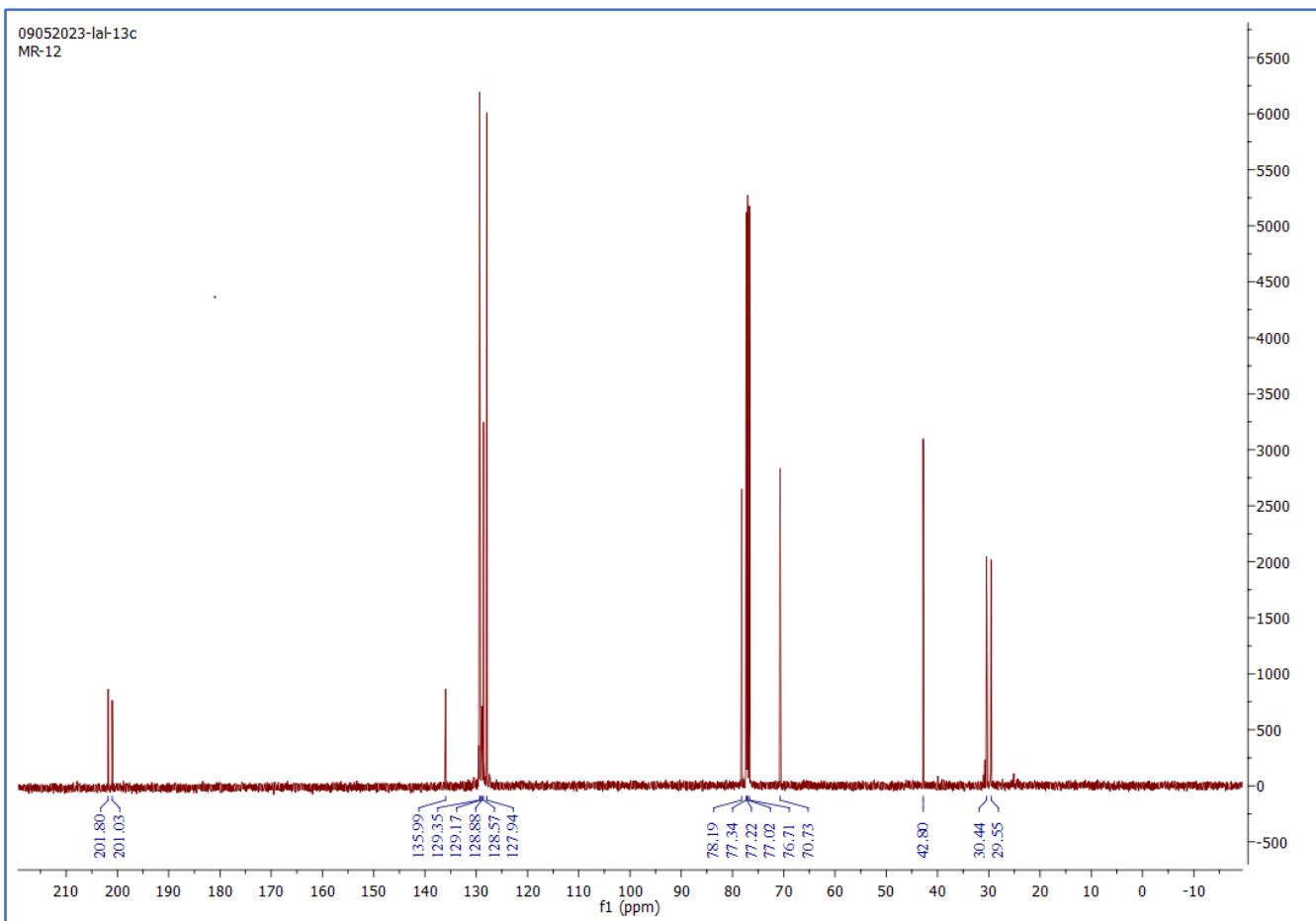
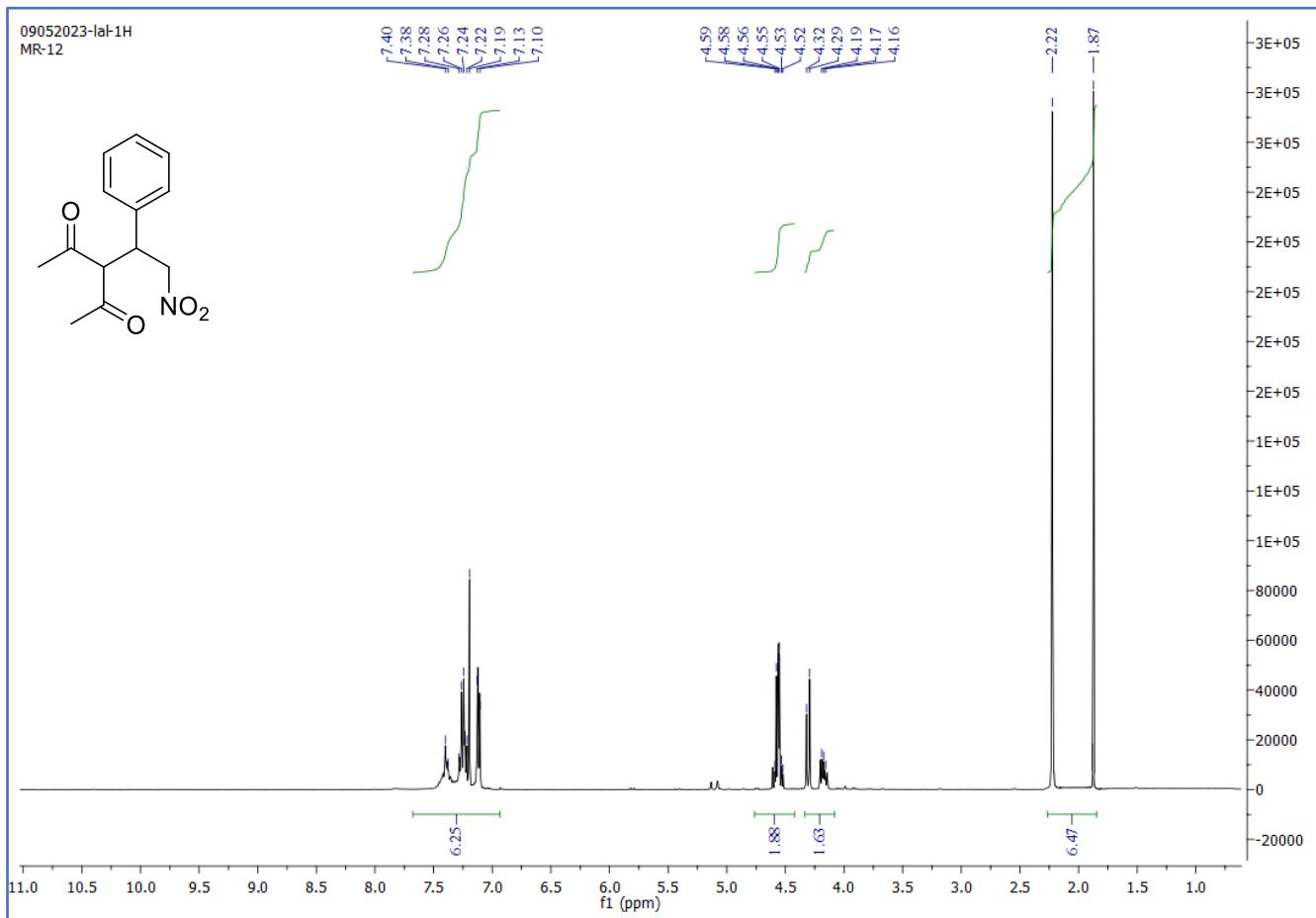


Figure S29: ^1H NMR and ^{13}C NMR of **8m**.

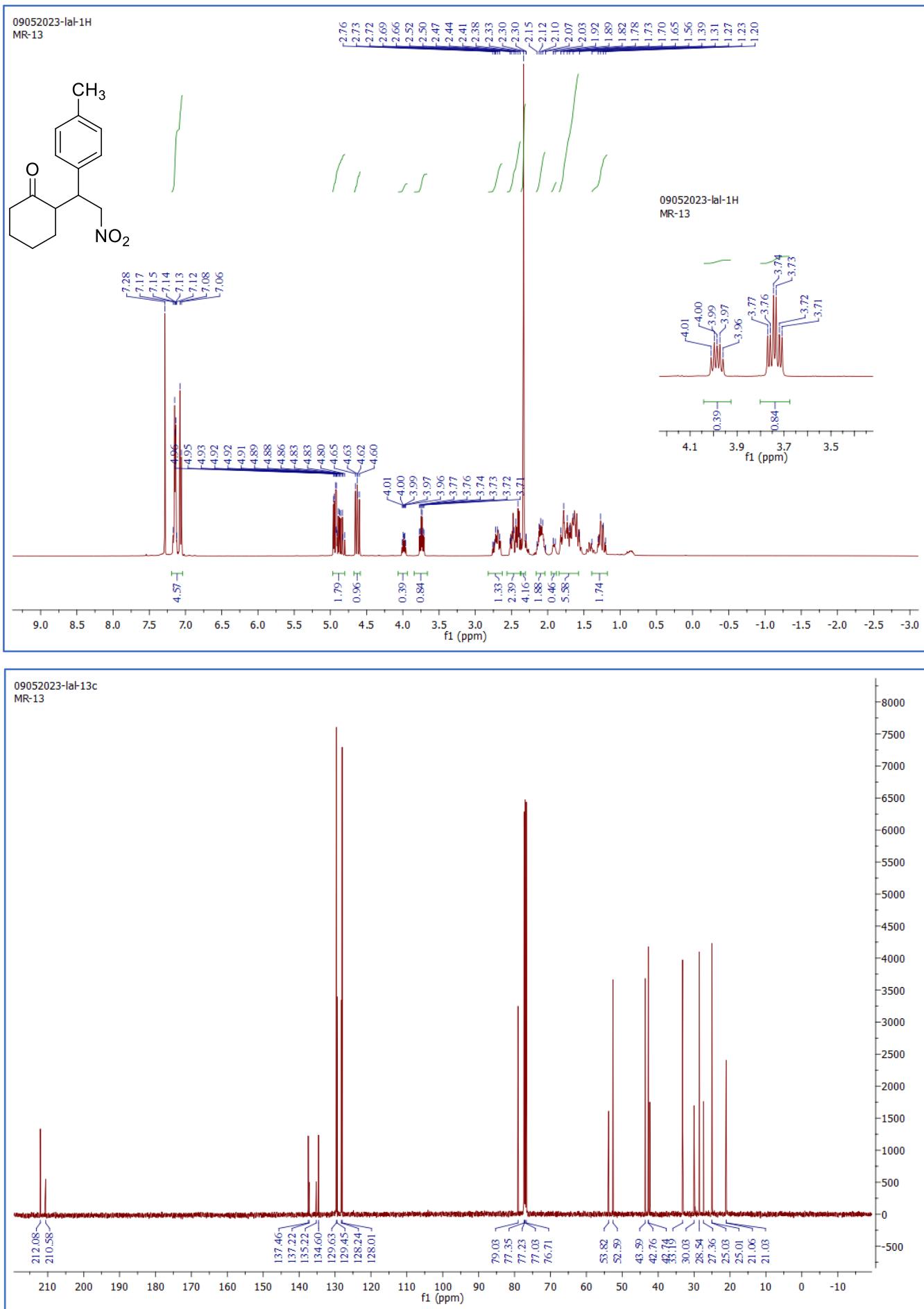


Figure 30: ^1H NMR and ^{13}C NMR of **8n**.

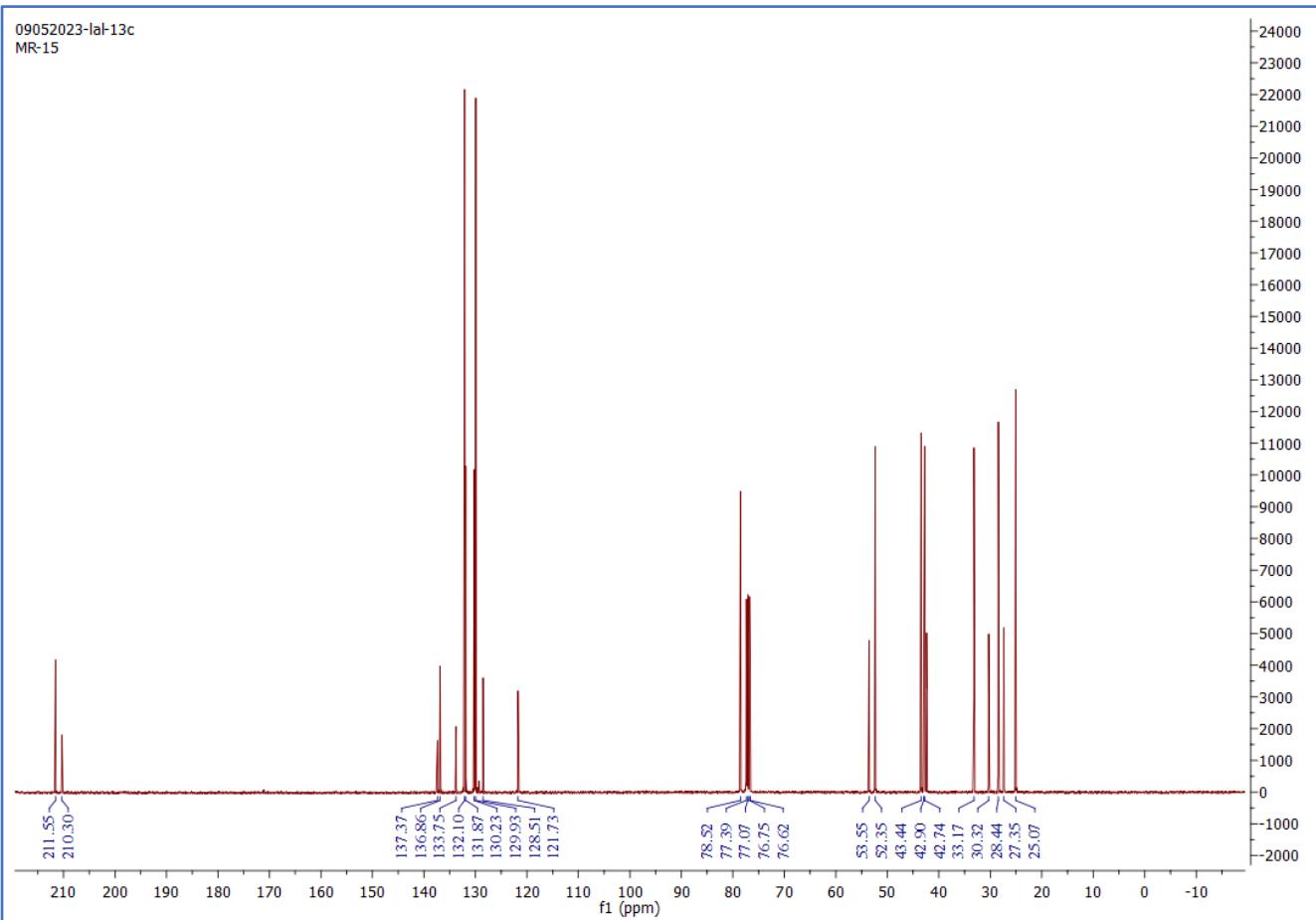
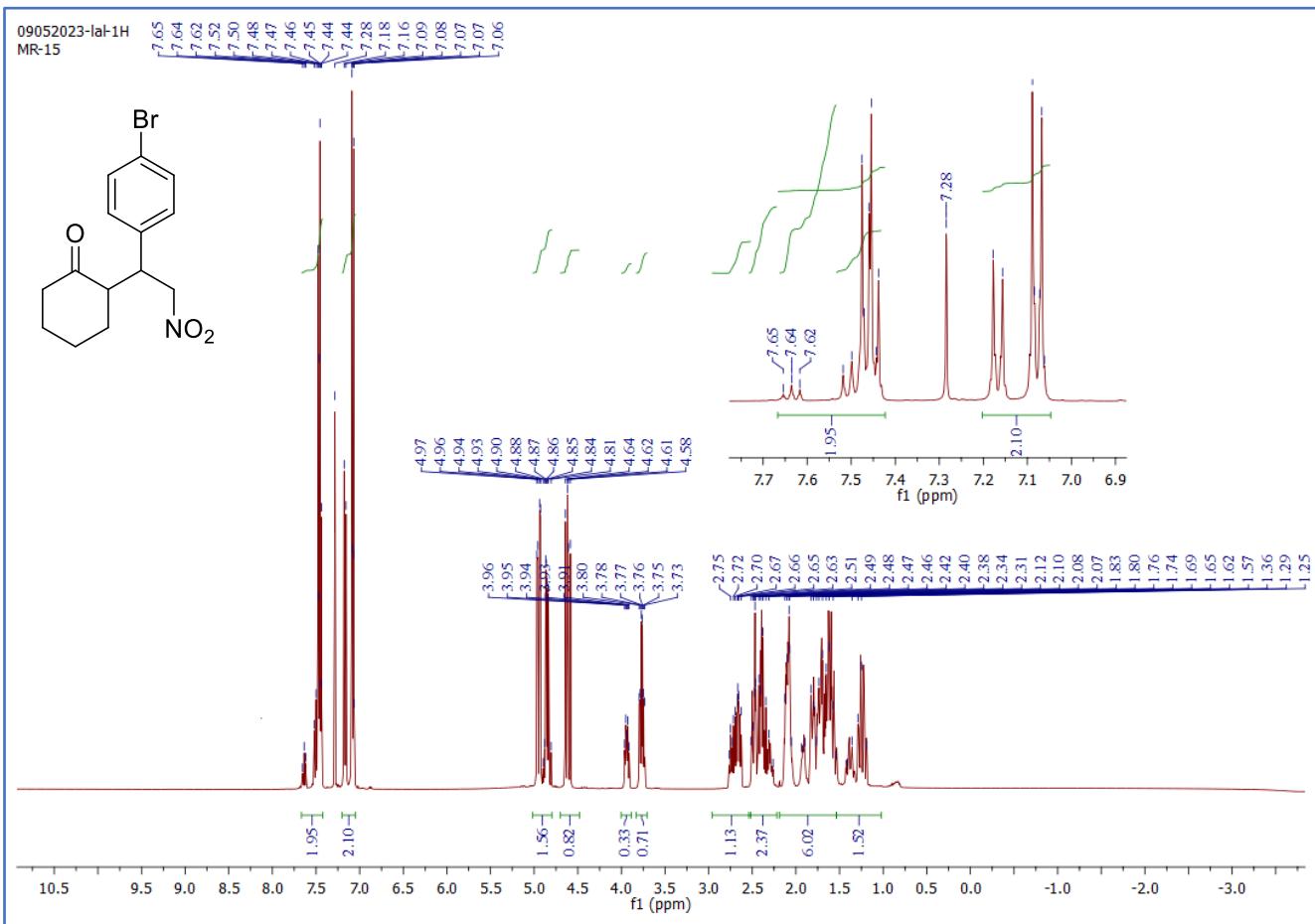


Figure S31: ^1H NMR and ^{13}C NMR of **8o**.

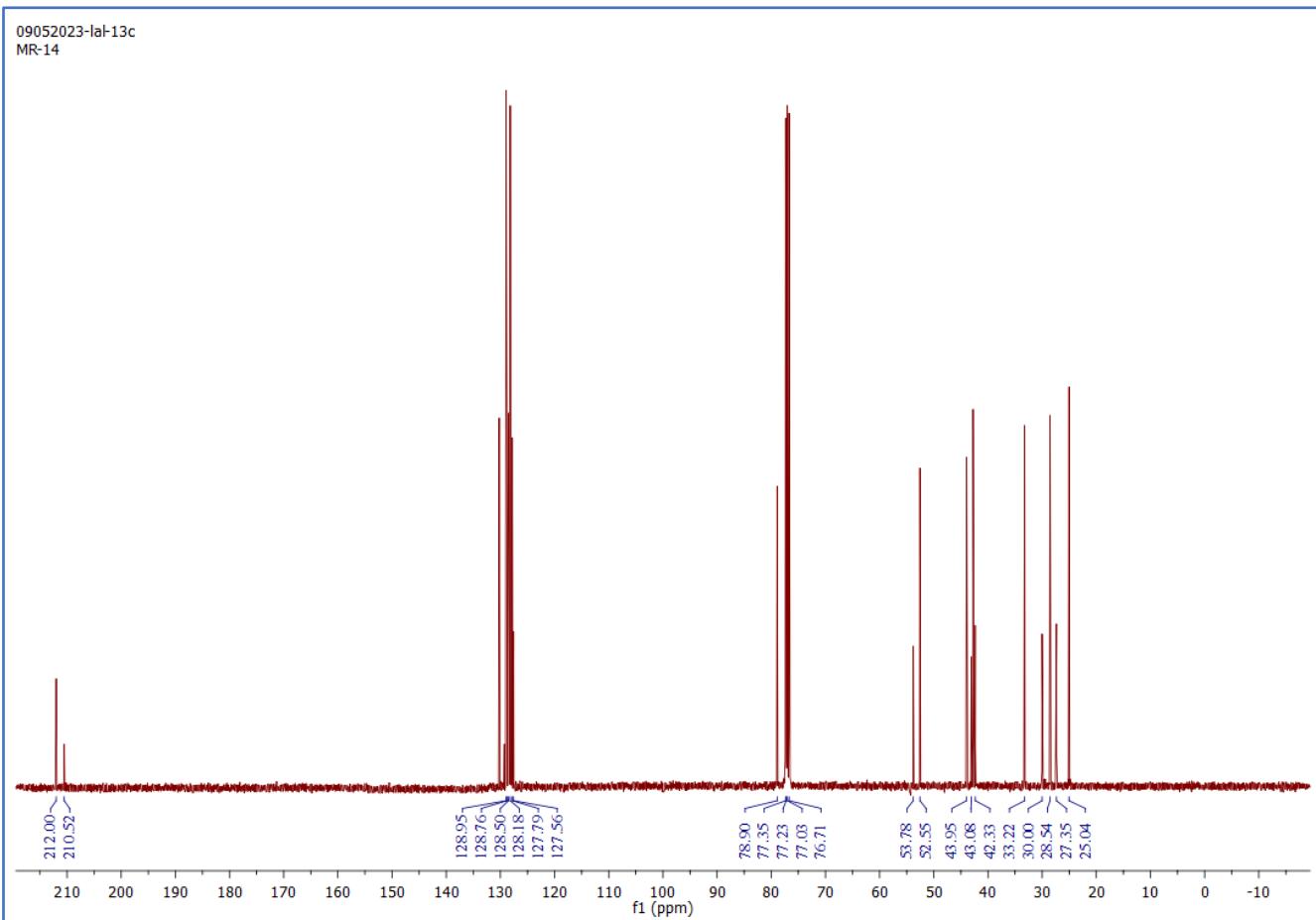
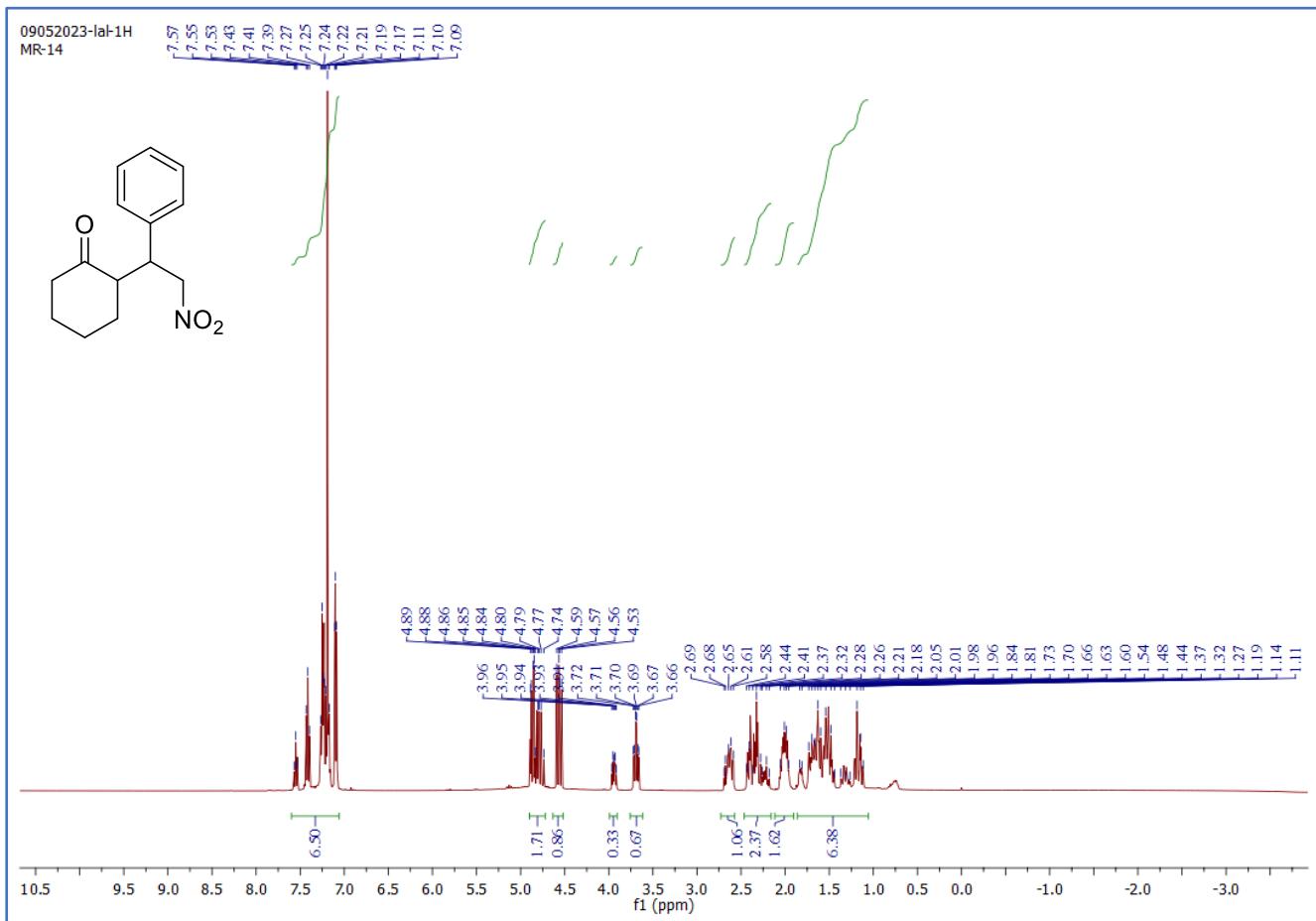


Figure S32: HPLC data of enantioenriched and racemic of **8a**.

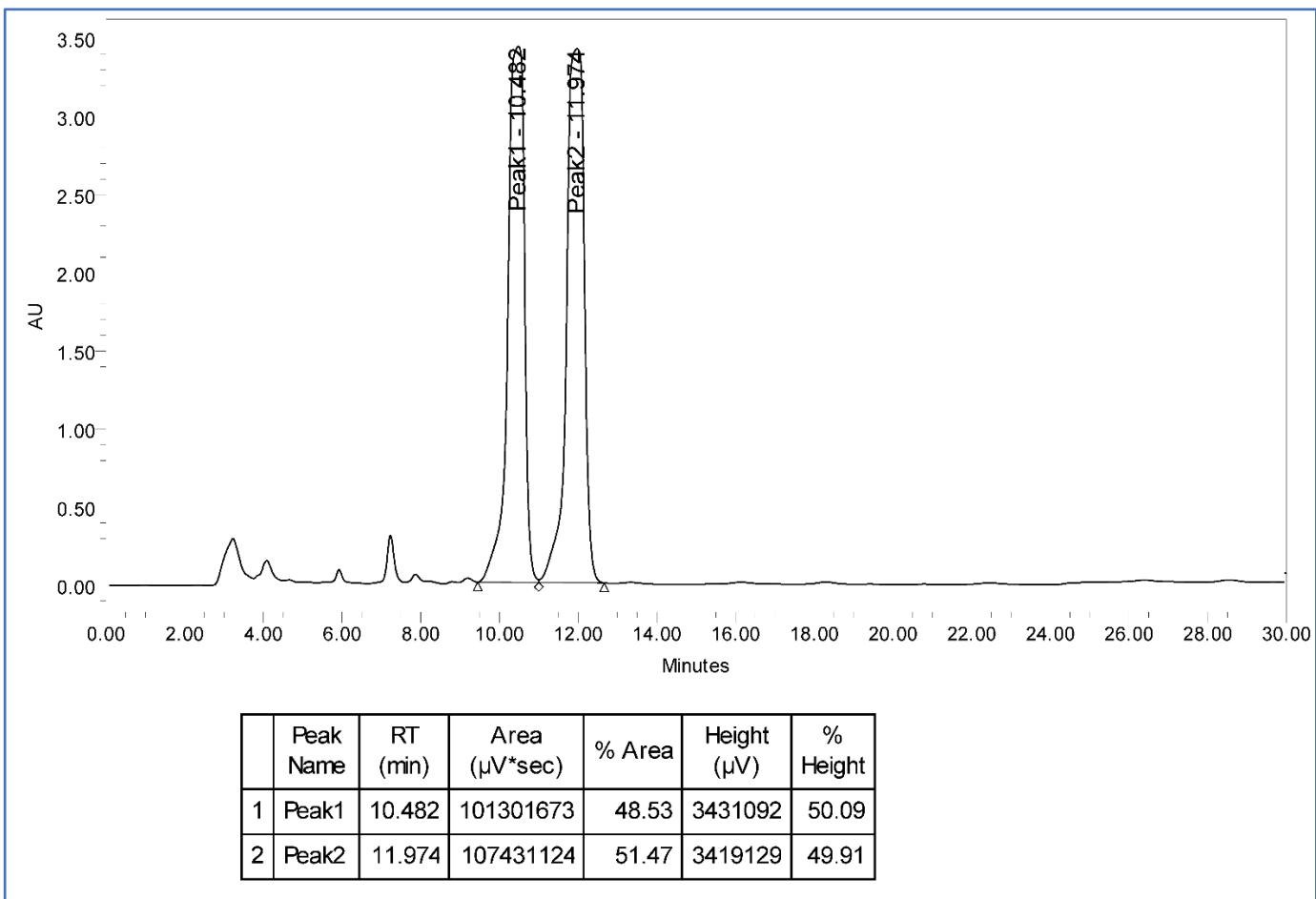
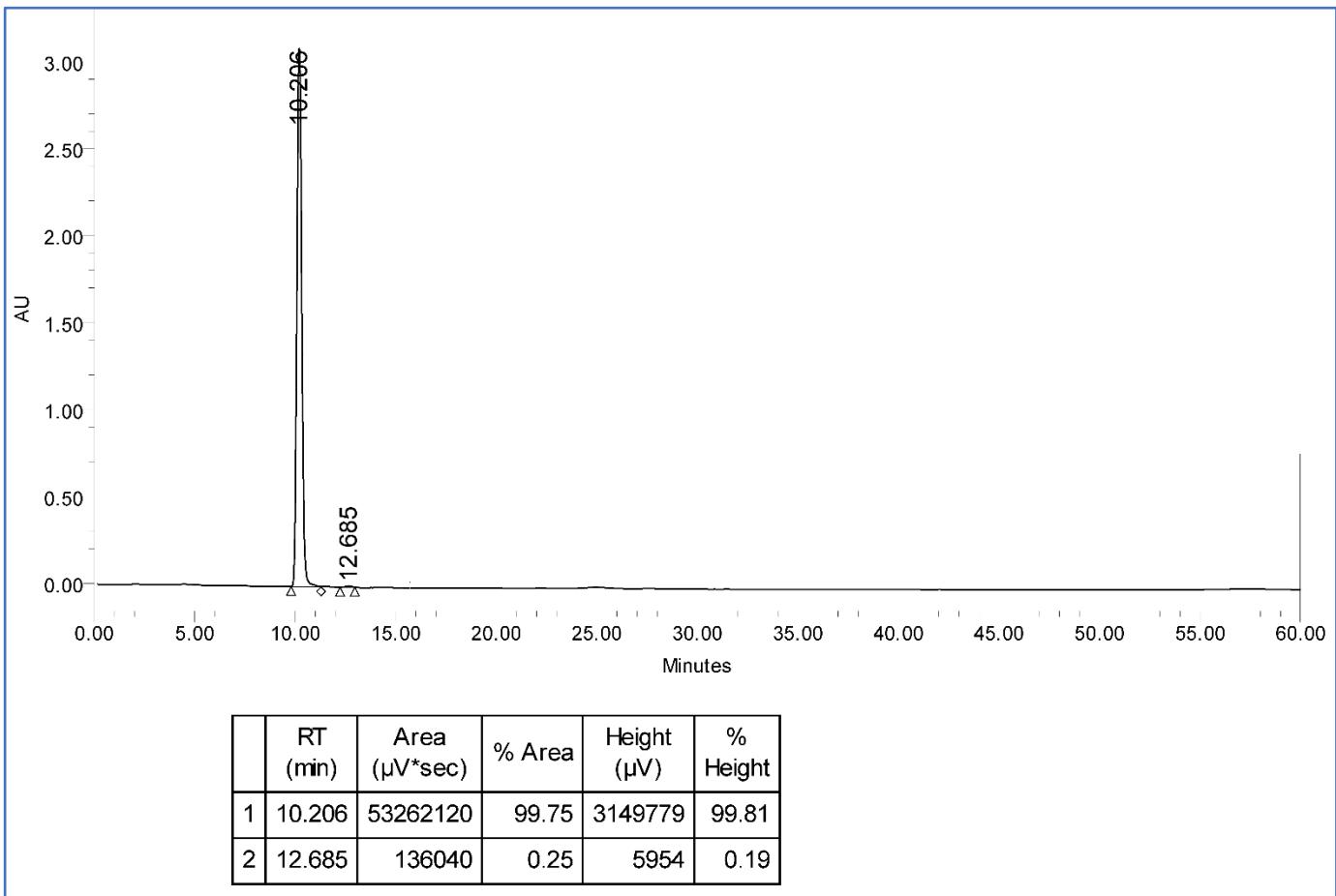


Figure S33: HPLC data of enantioenriched and racemic of **8b**.

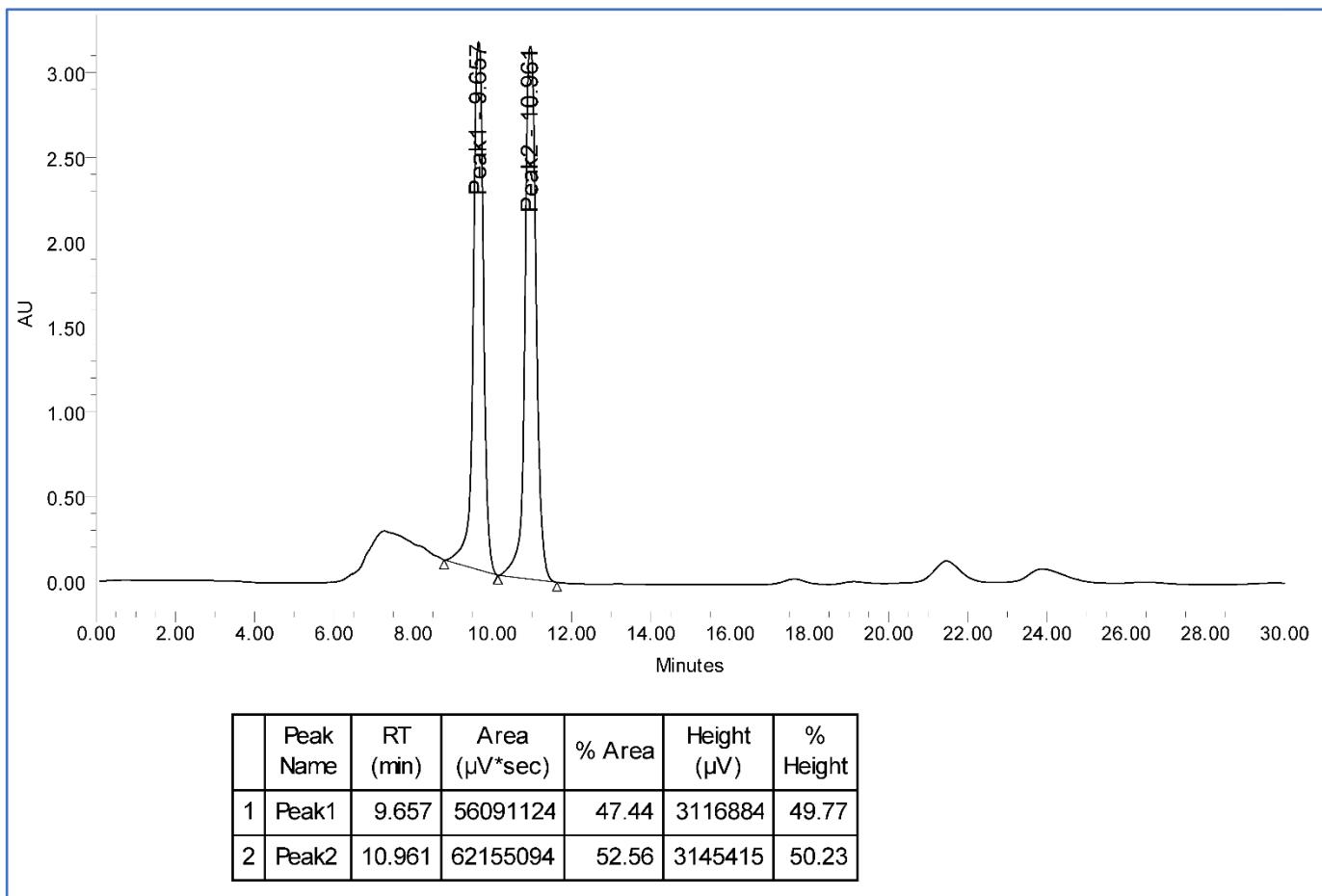
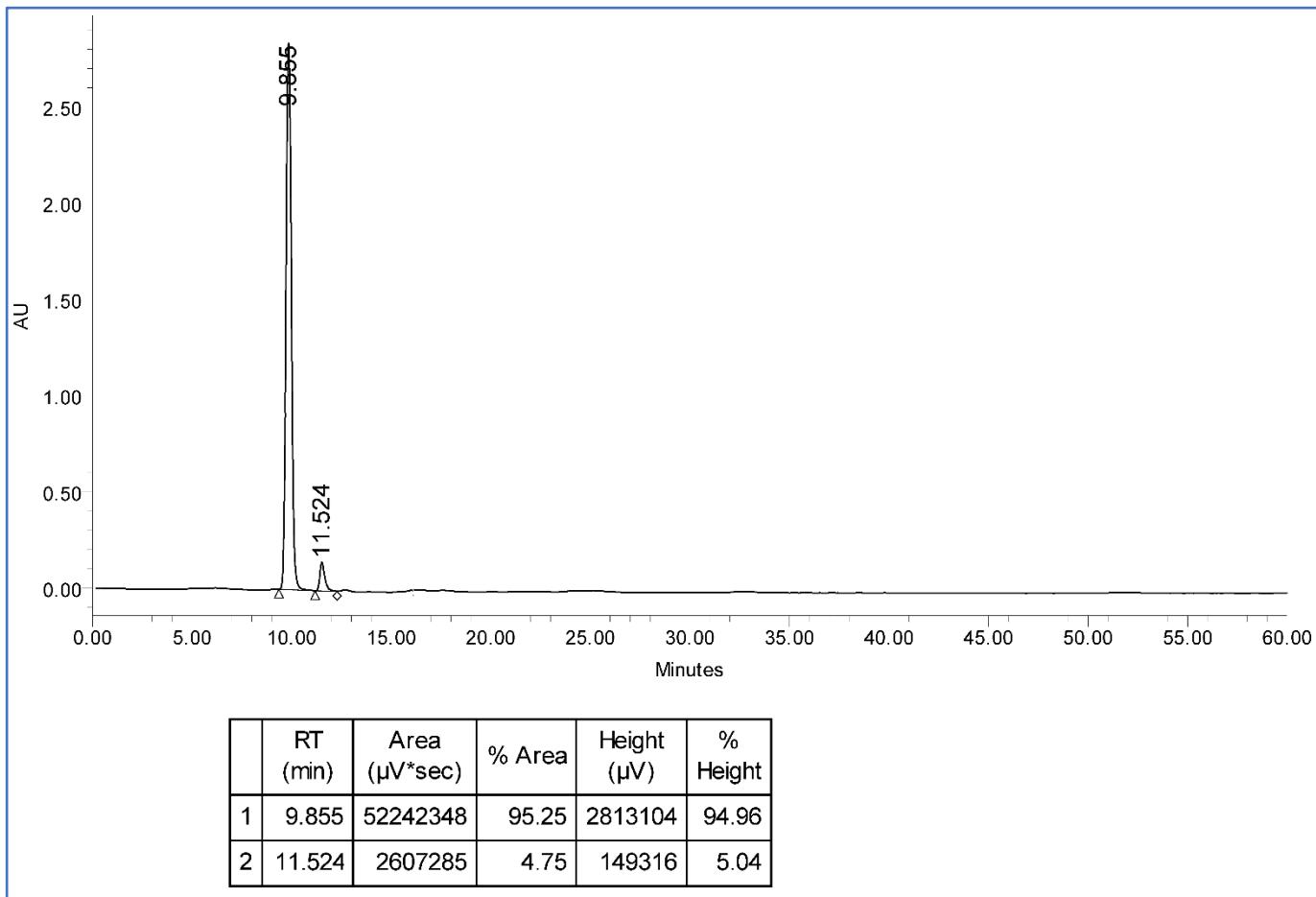


Figure S34: HPLC data of enantioenriched and racemic of **8c**.

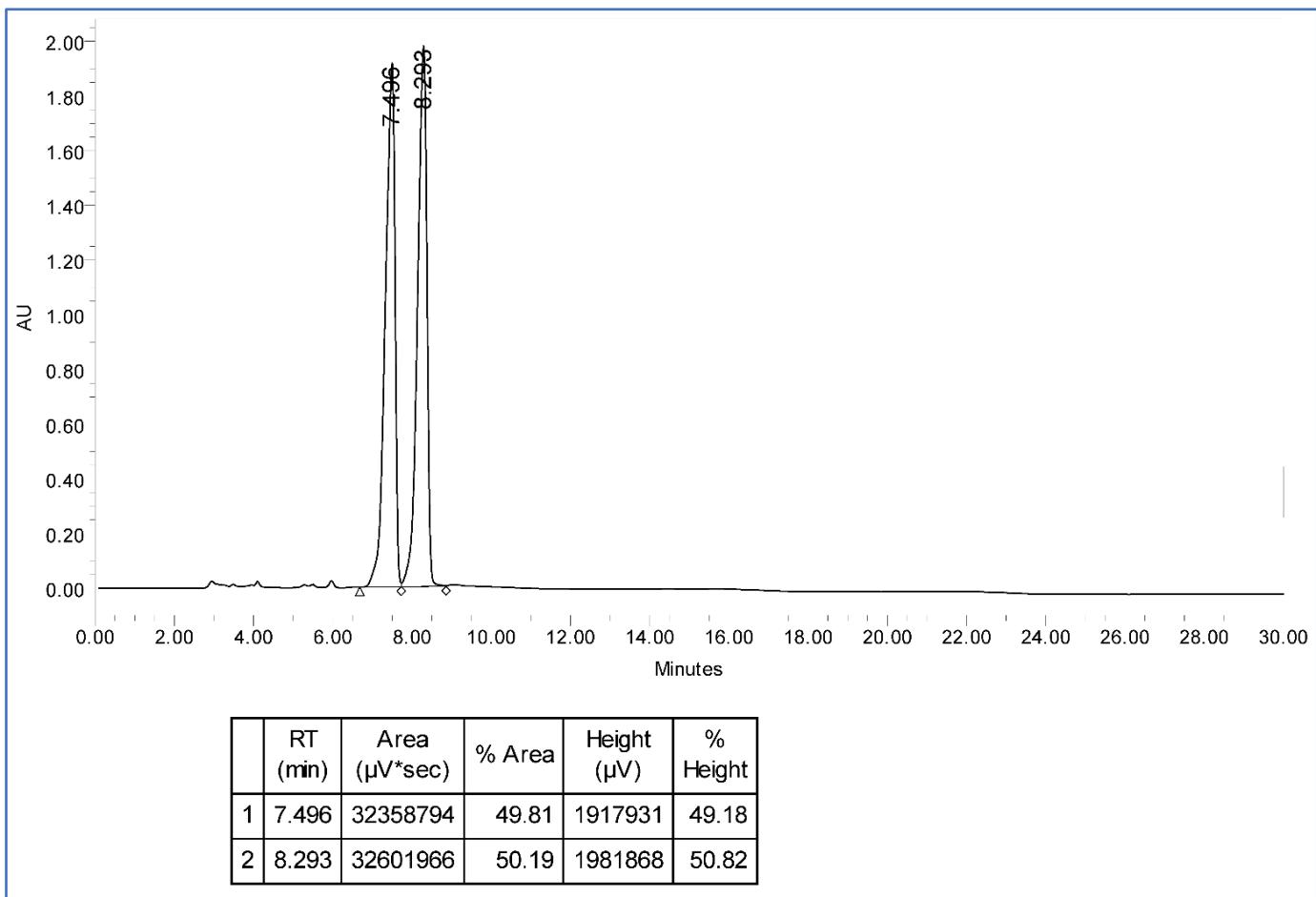
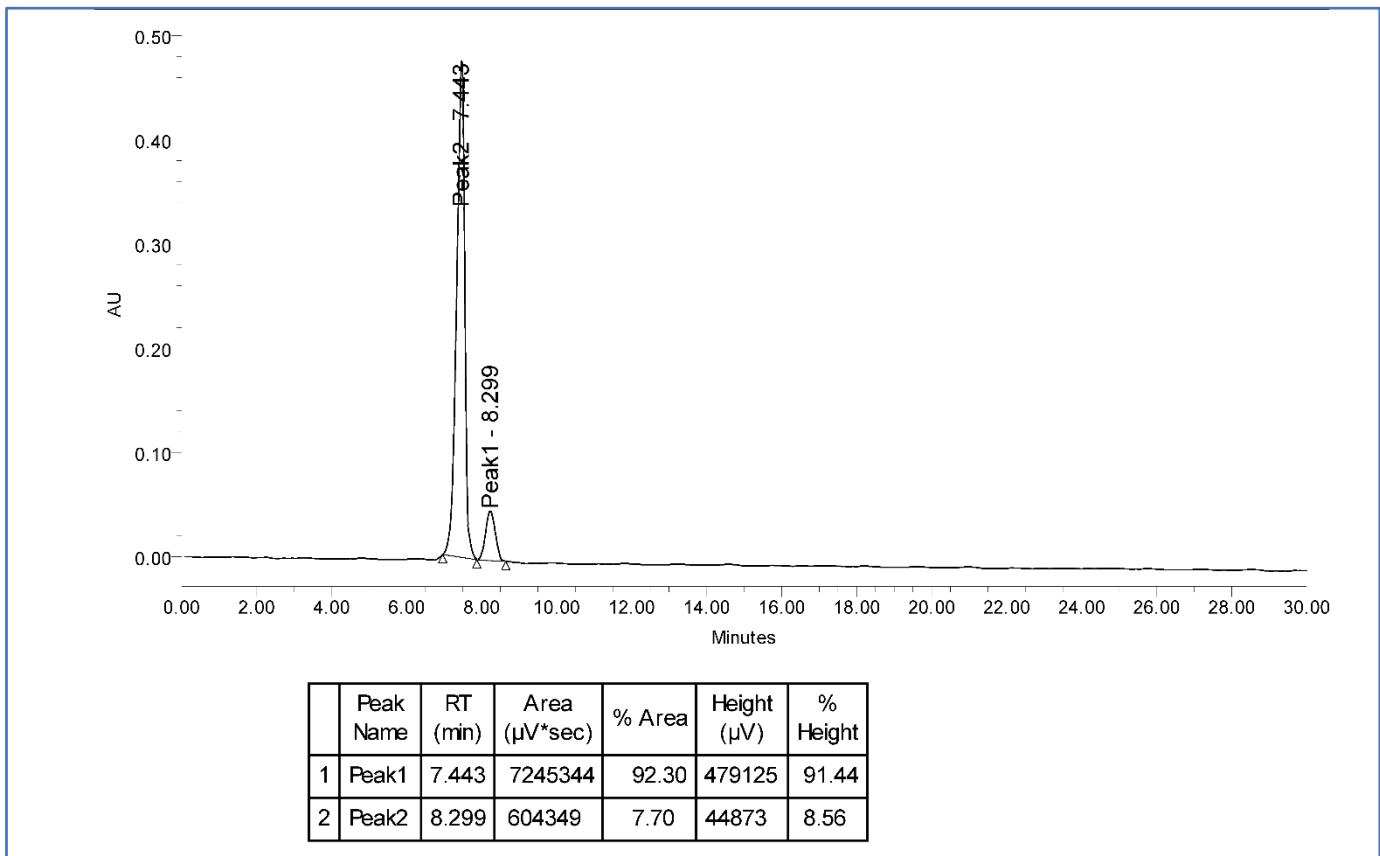


Figure S35: HPLC data of enantioenriched and racemic of **8d**.

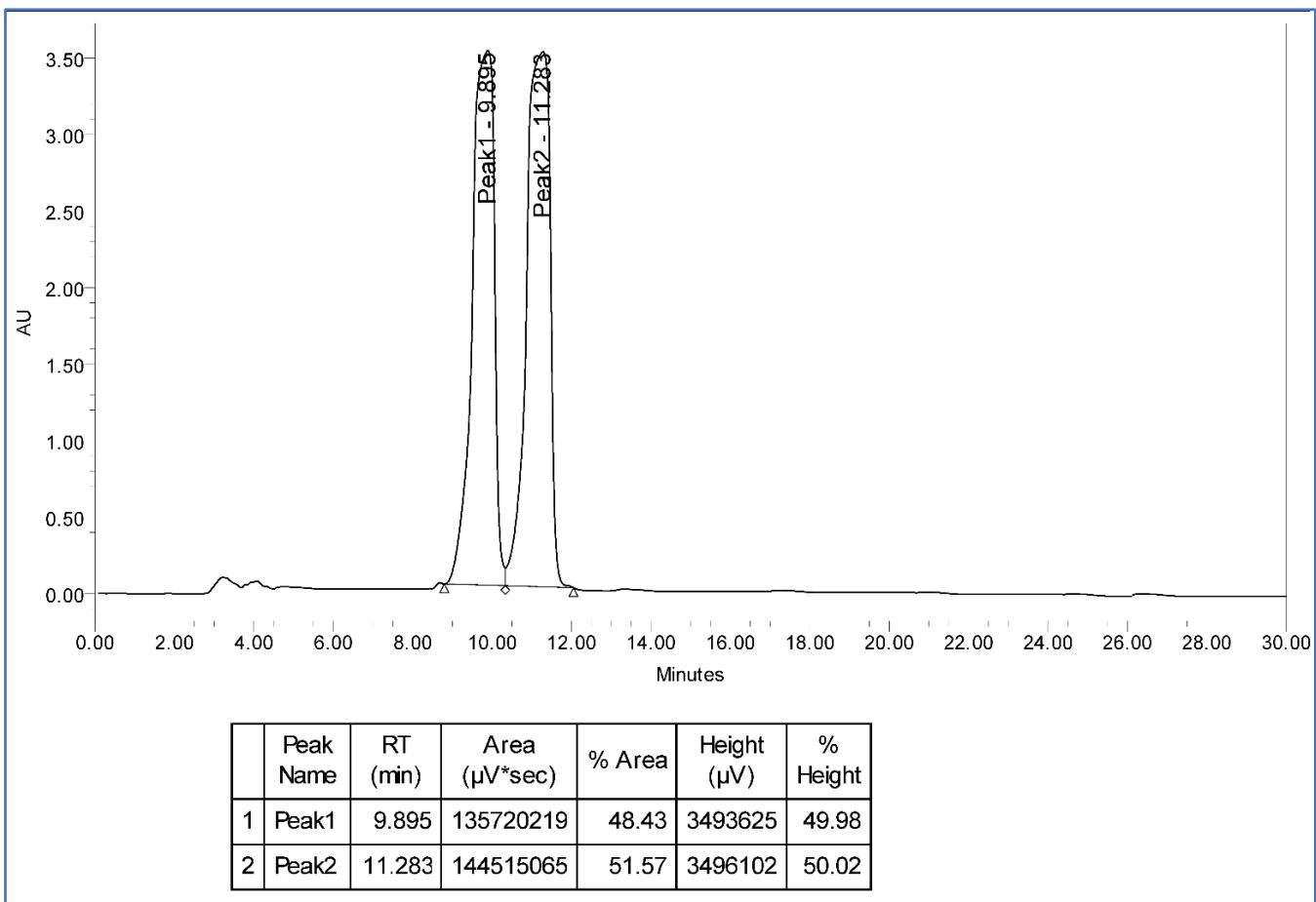
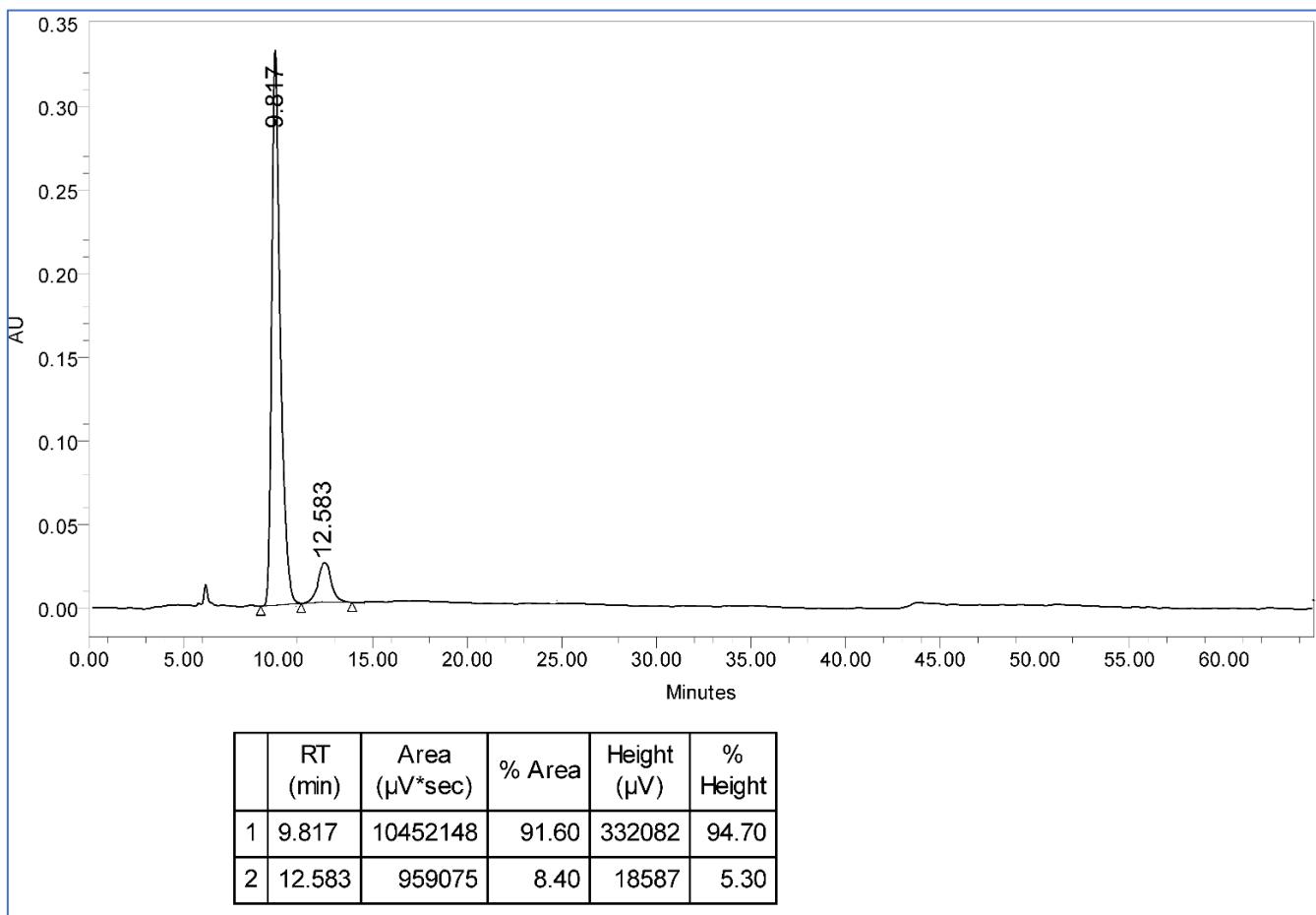


Figure S36: HPLC data of enantioenriched and racemic of **8e**.

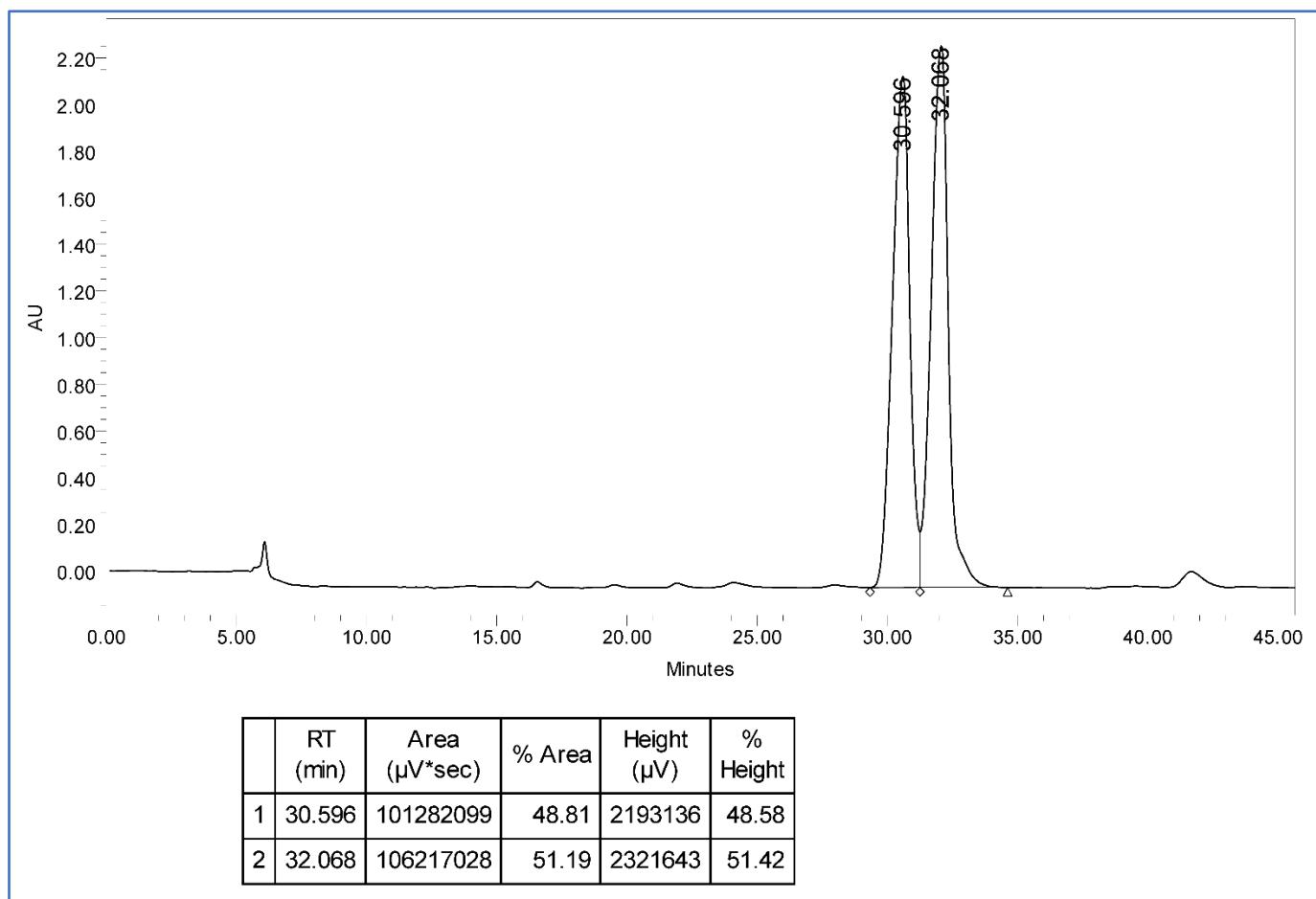
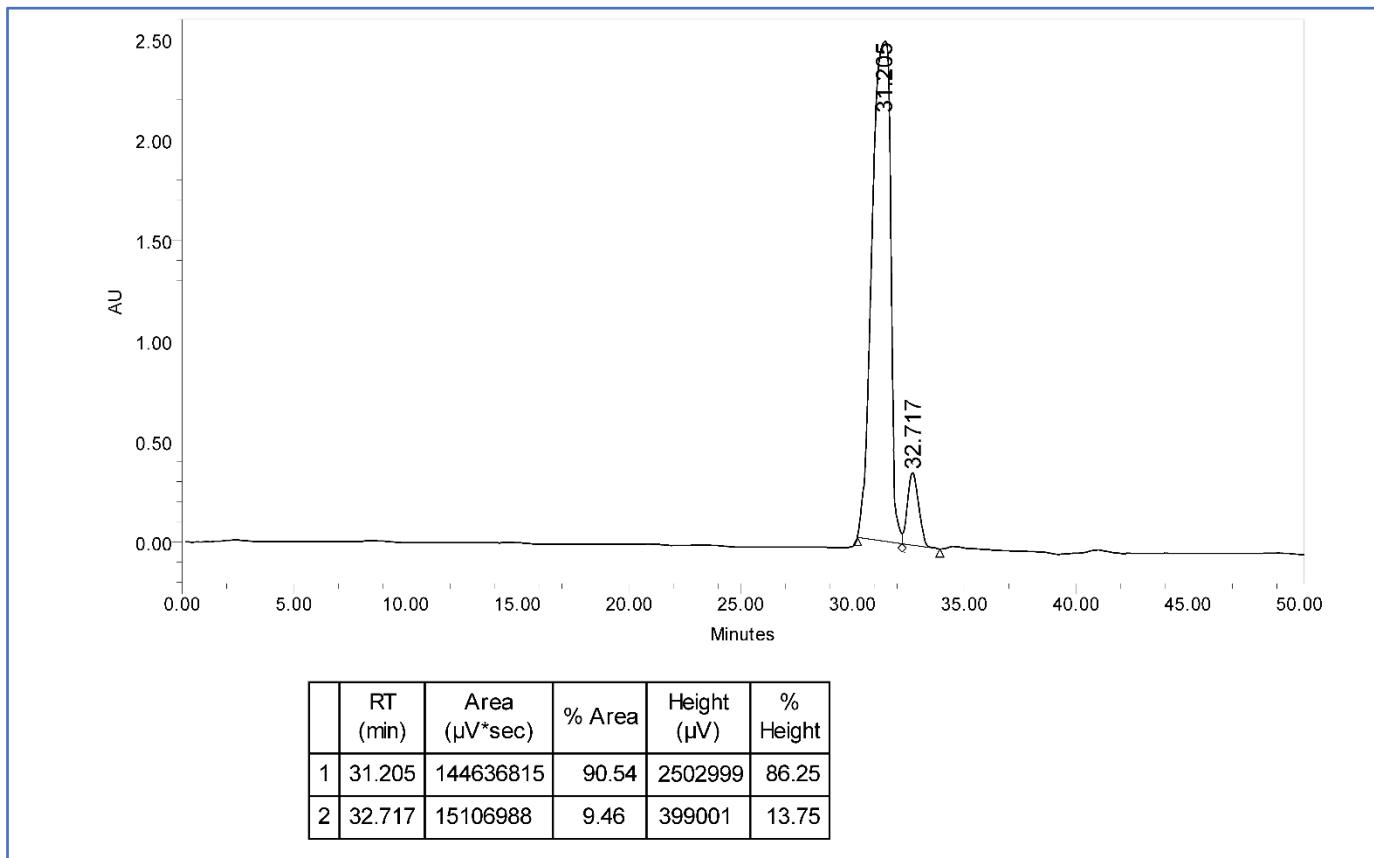


Figure S37: HPLC data of enantioenriched and racemic of **8f**.

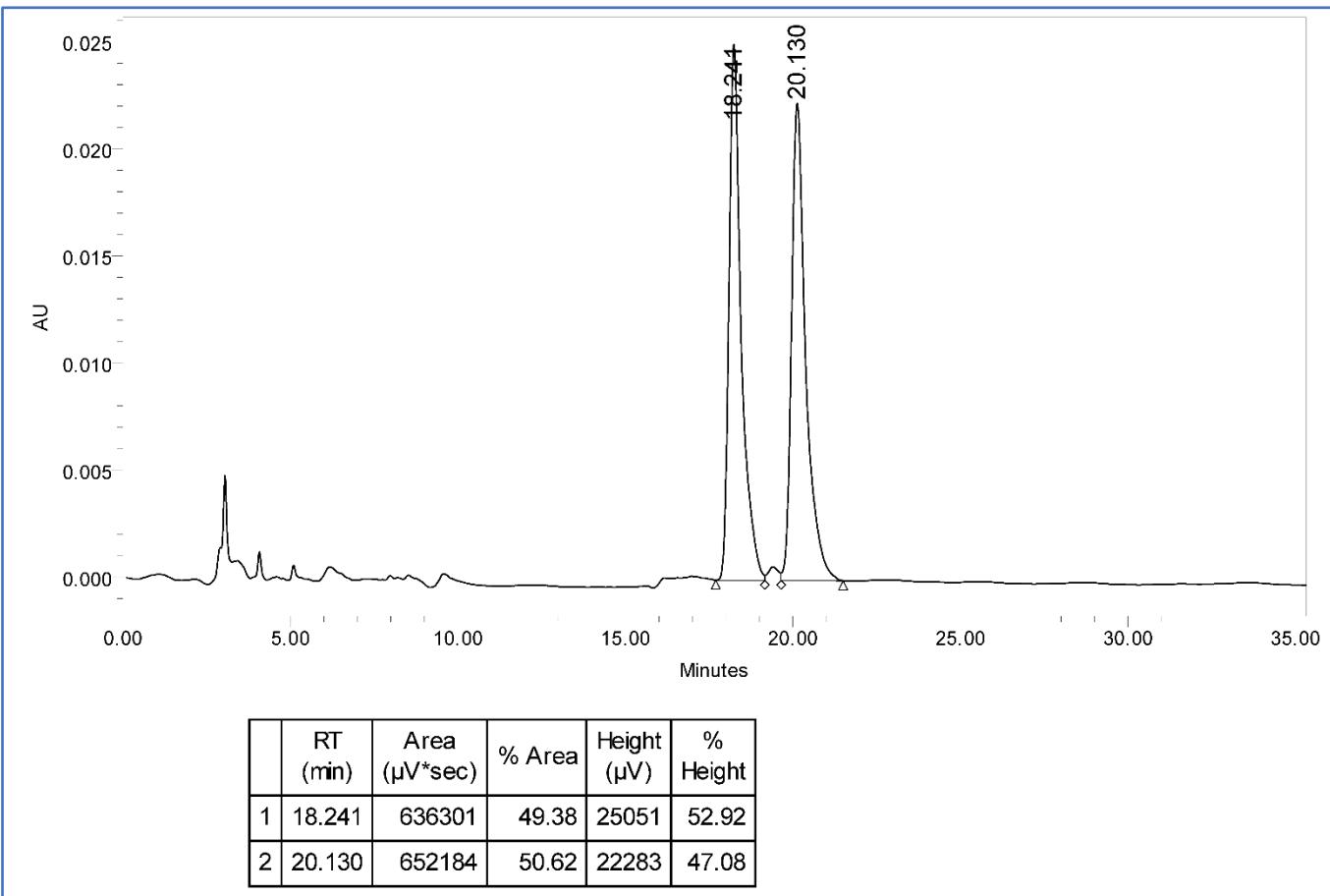
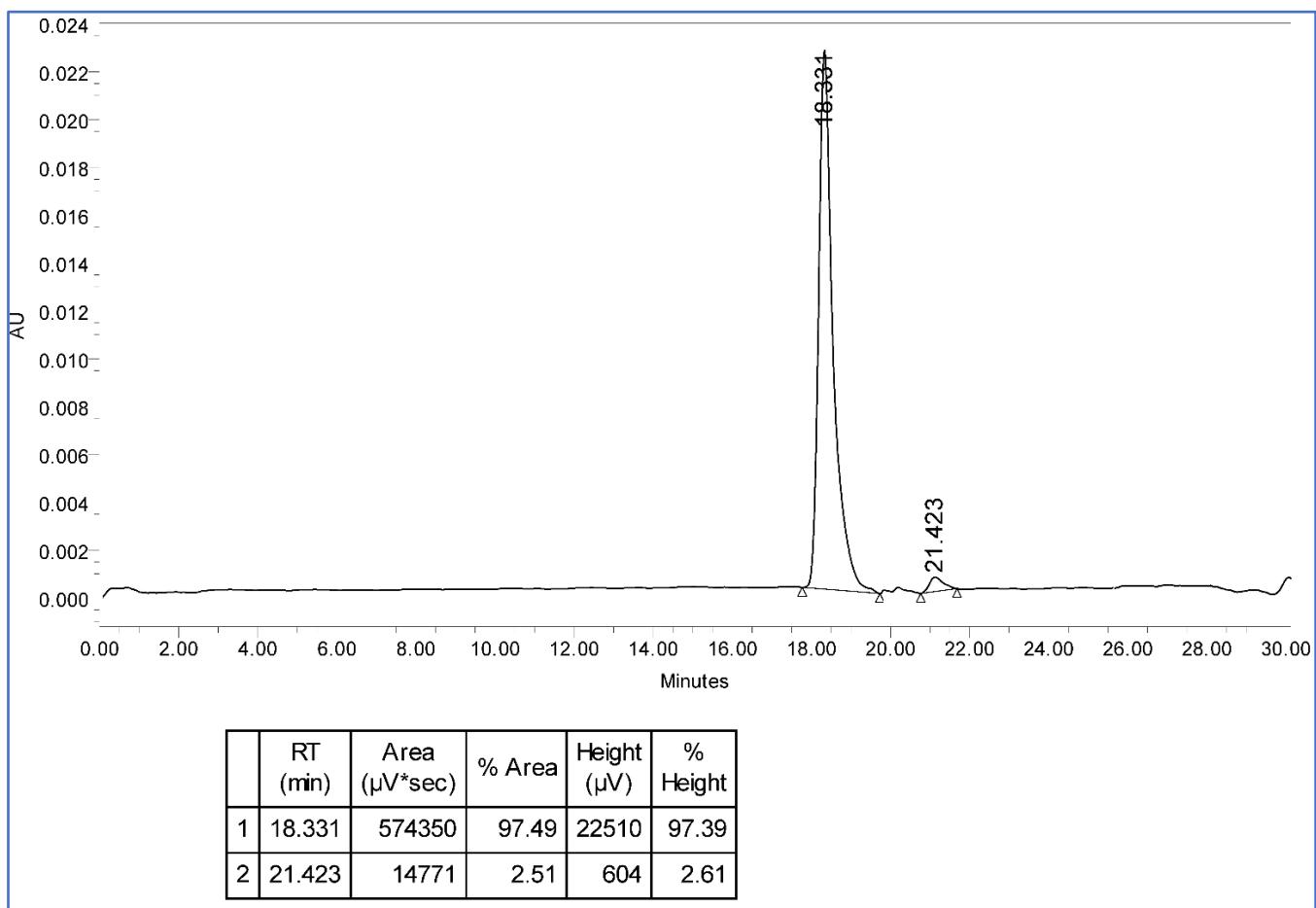


Figure S38: HPLC data of enantioenriched and racemic of **8g**.

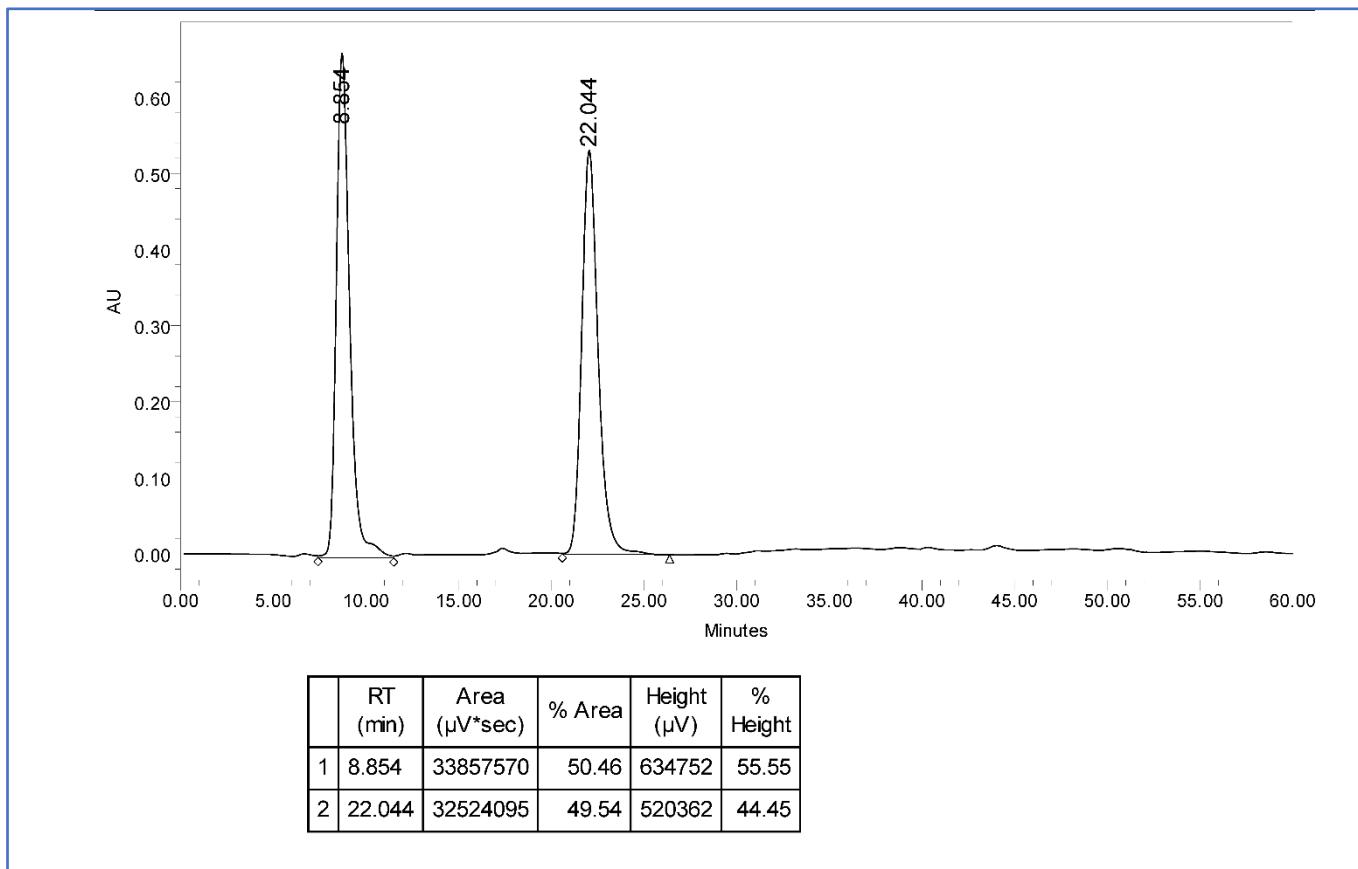
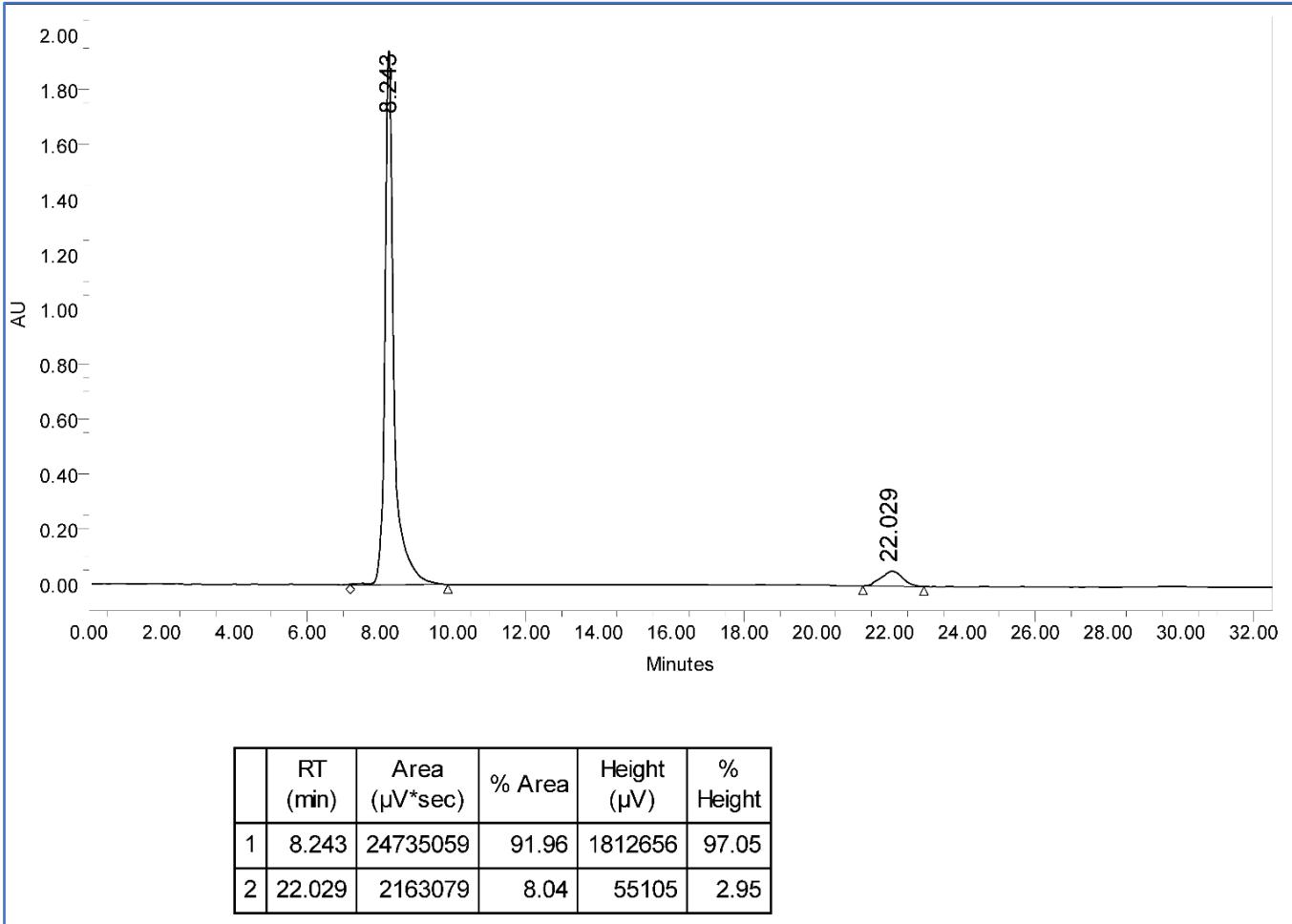


Figure S39: HPLC data of enantioenriched and racemic of **8h**.

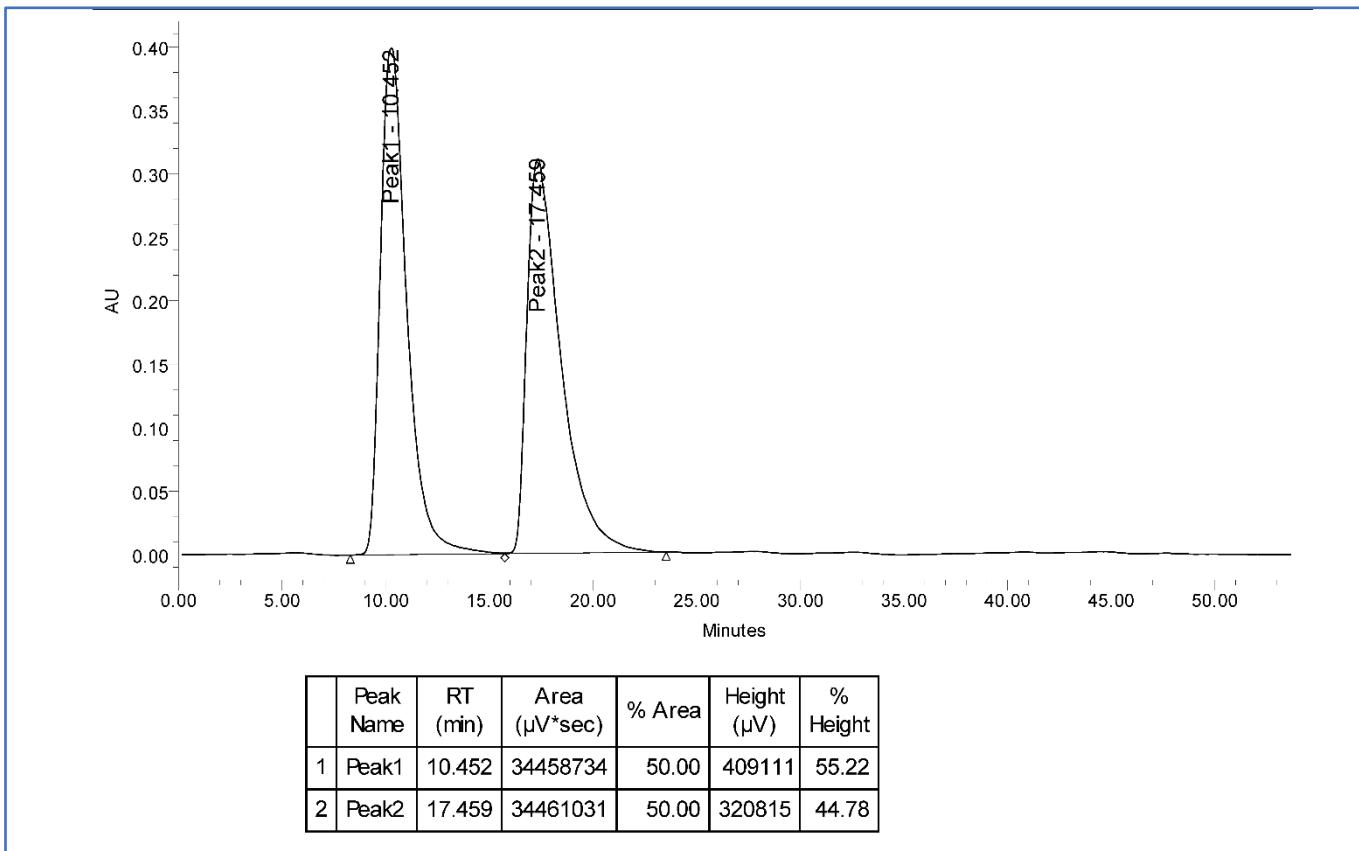
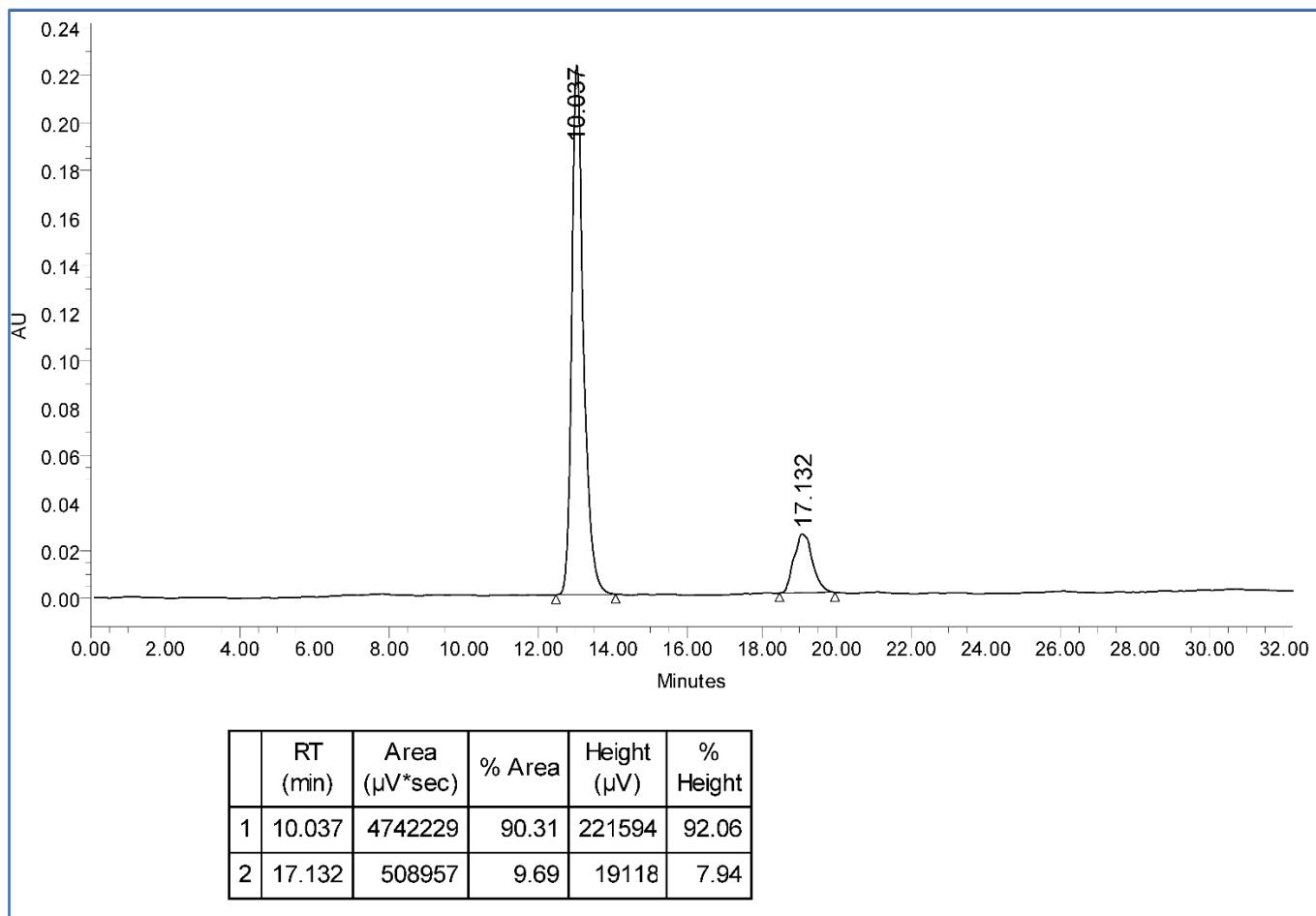


Figure S40: HPLC data of enantioenriched and racemic of **8i**.

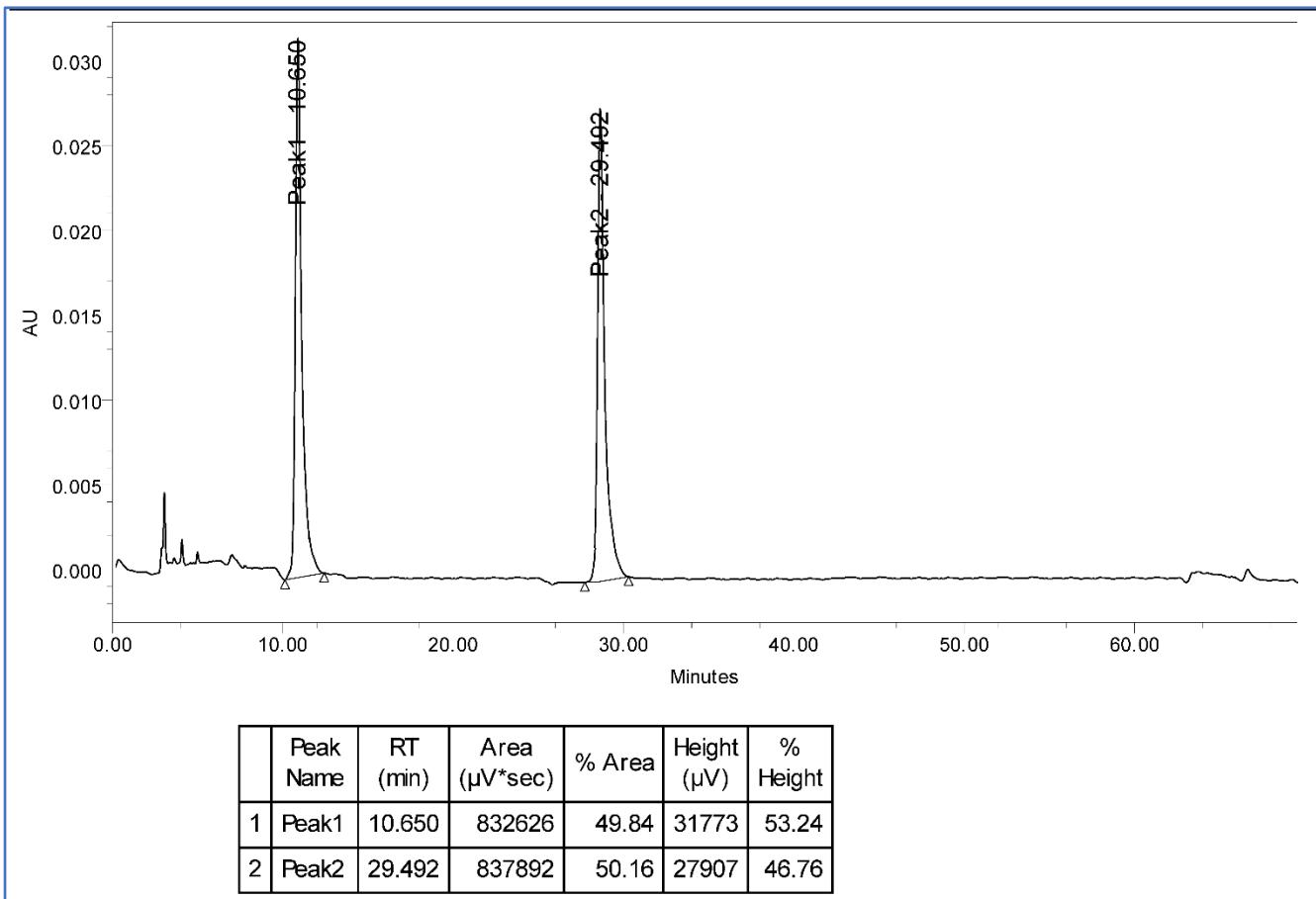
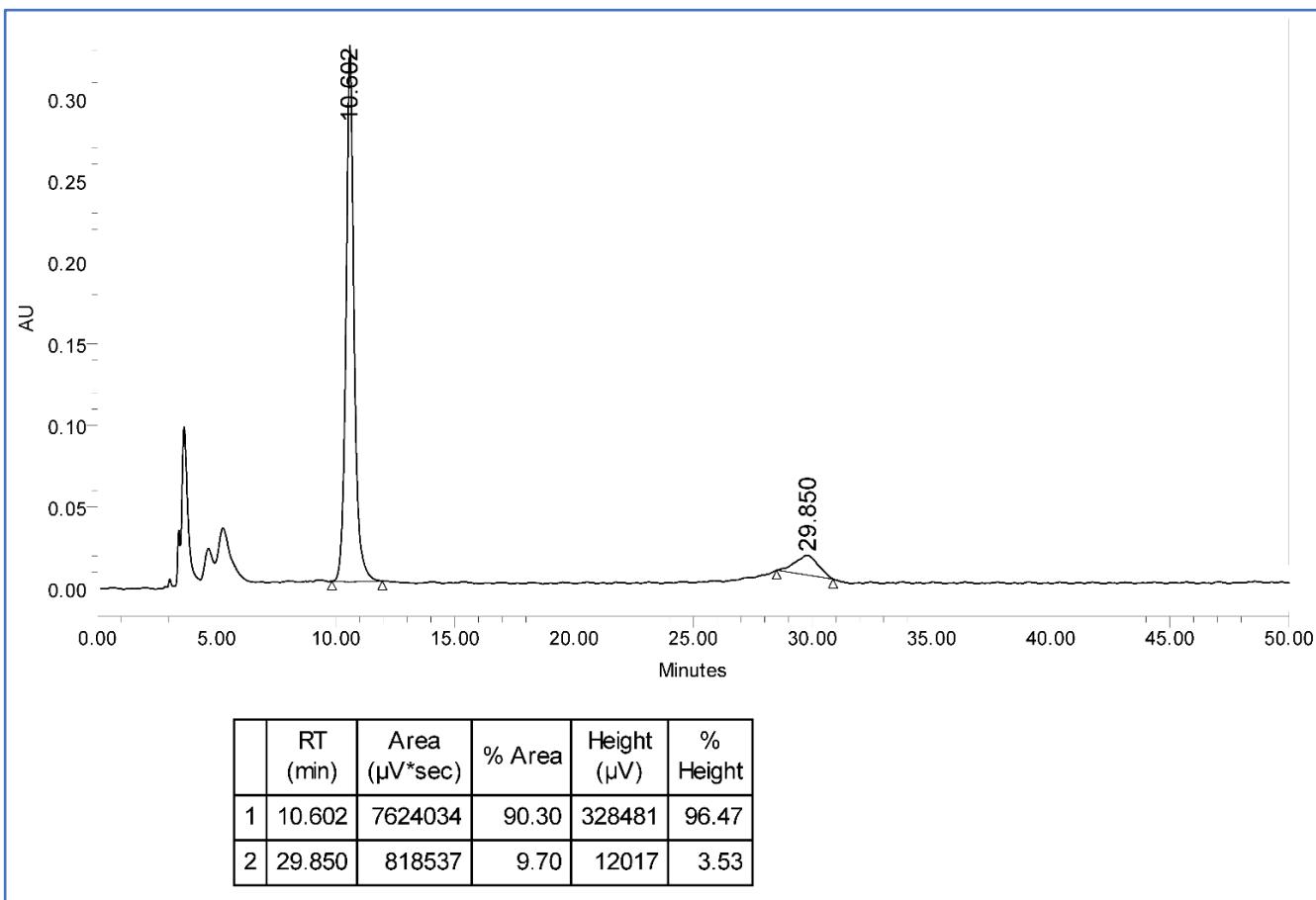


Figure S41: HPLC data of enantioenriched and racemic of **8j**.

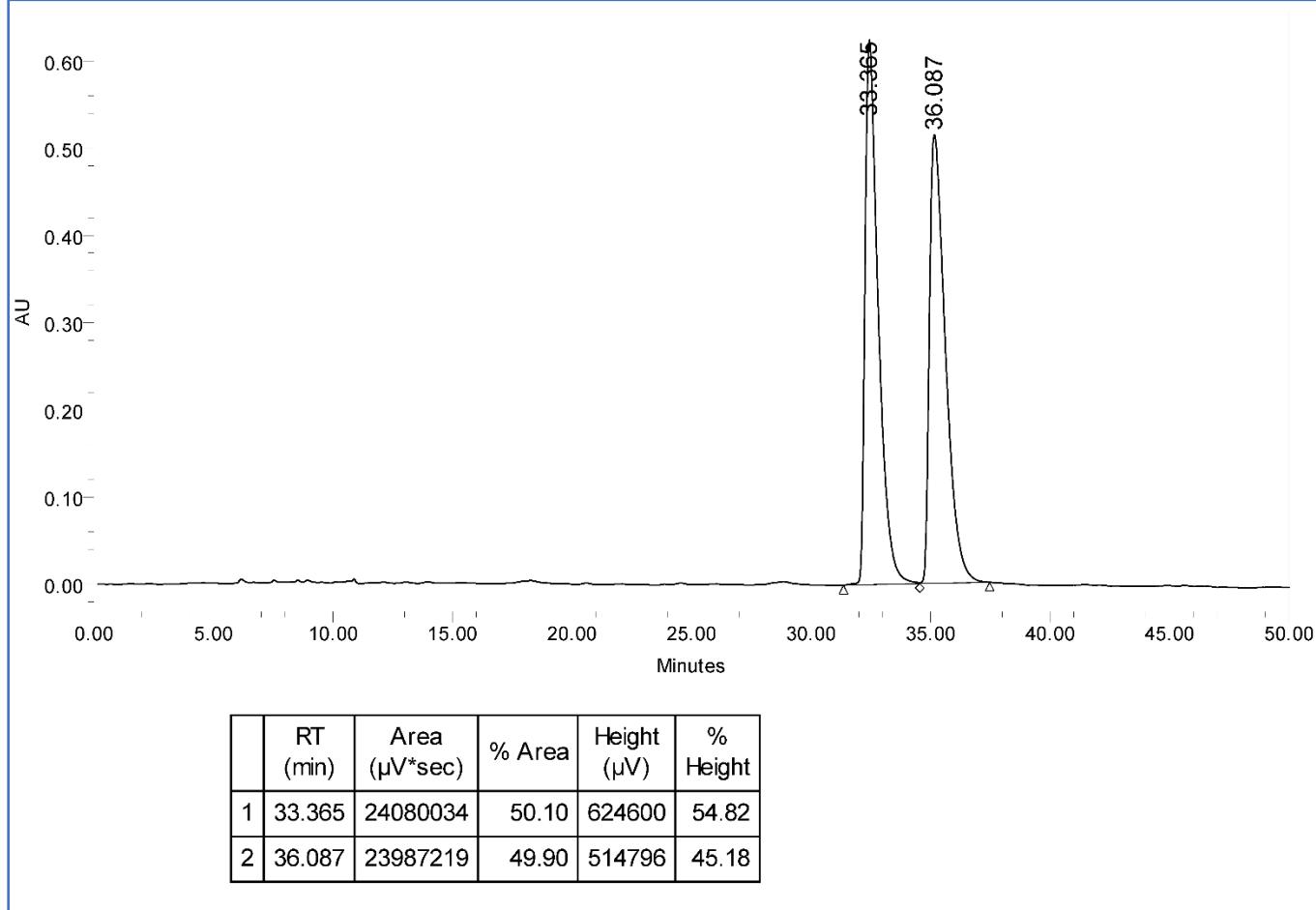
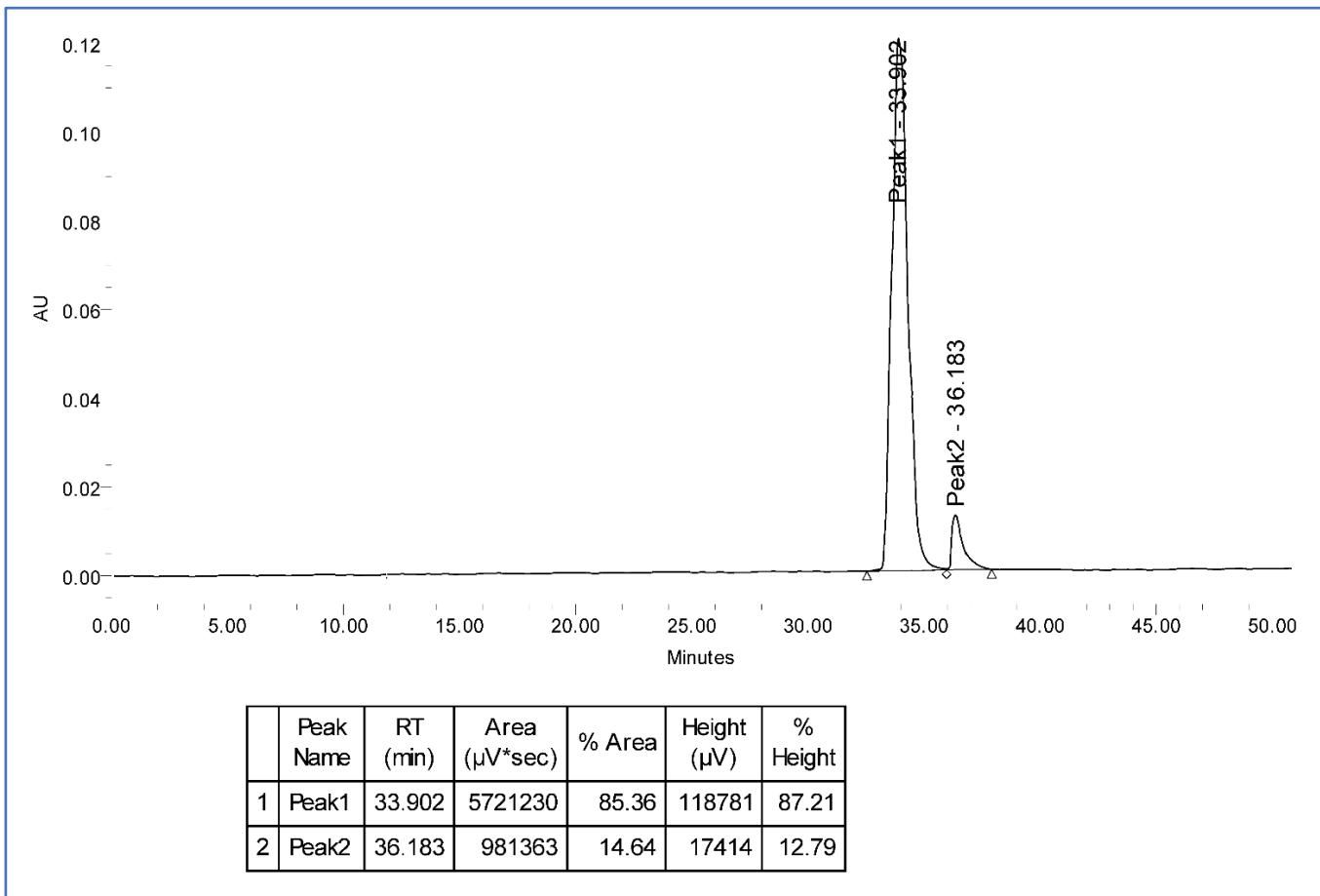


Figure S42: HPLC data of enantioenriched and racemic of **8k**.

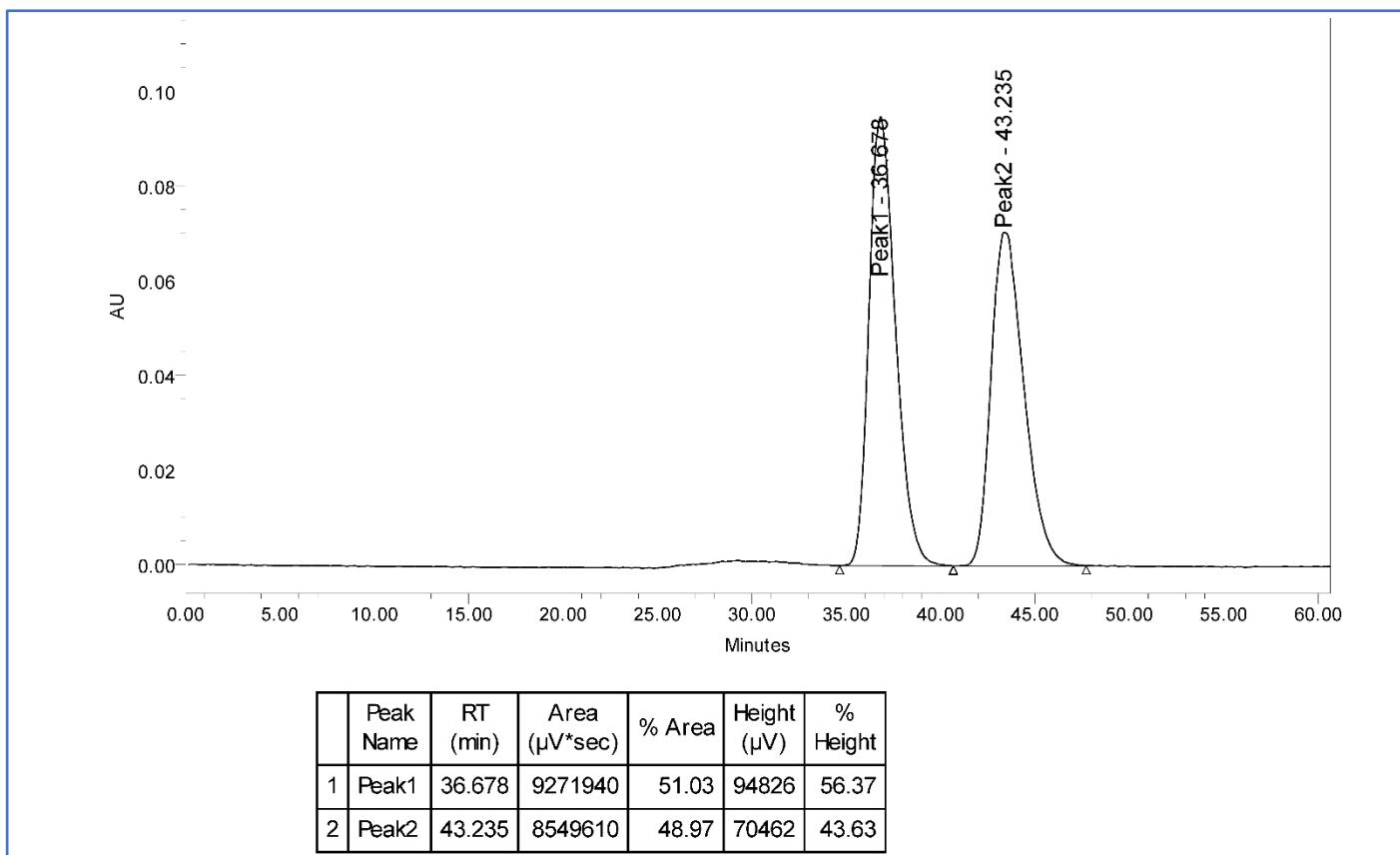
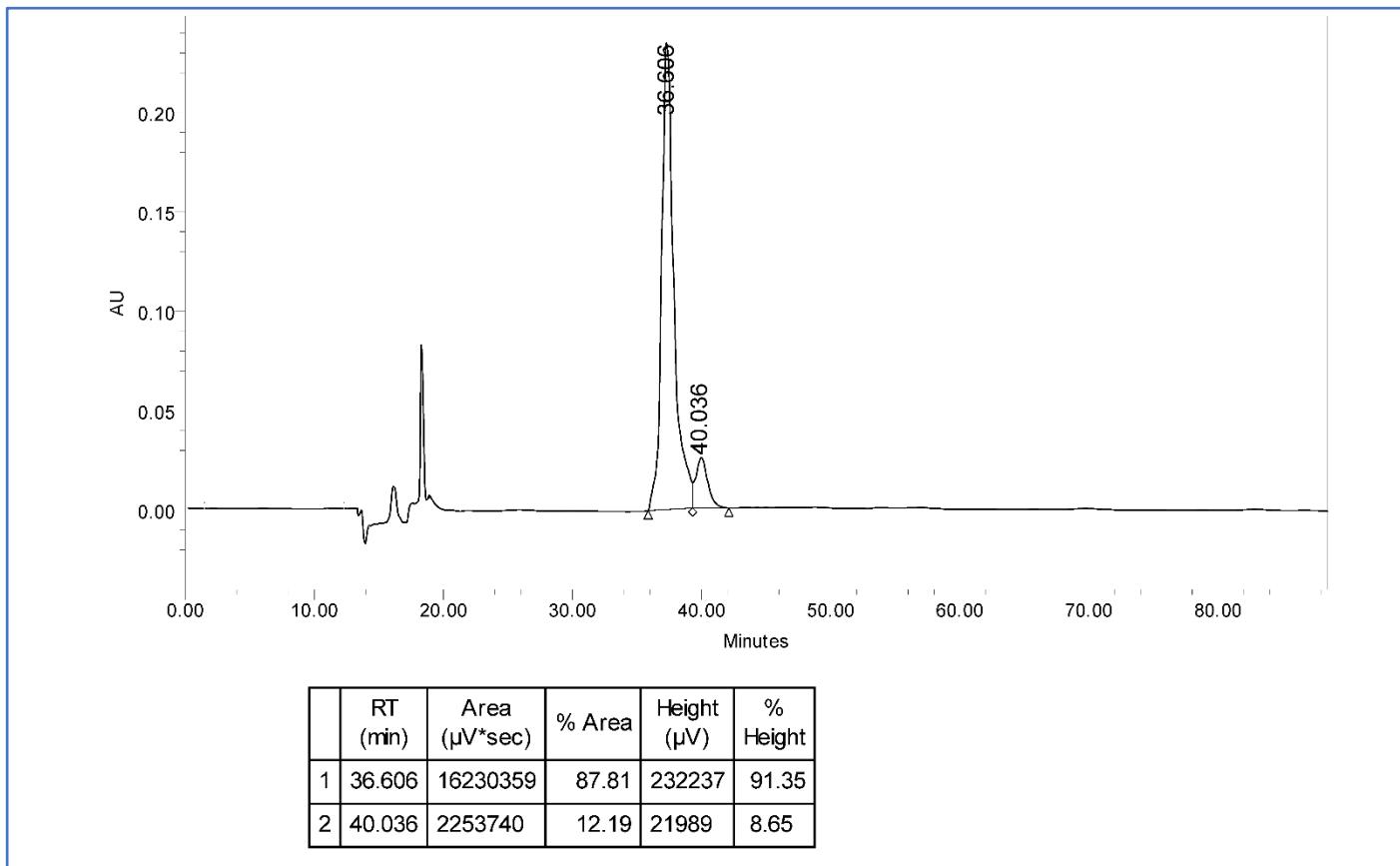


Figure S43: HPLC data of enantioenriched and racemic of **8l**.

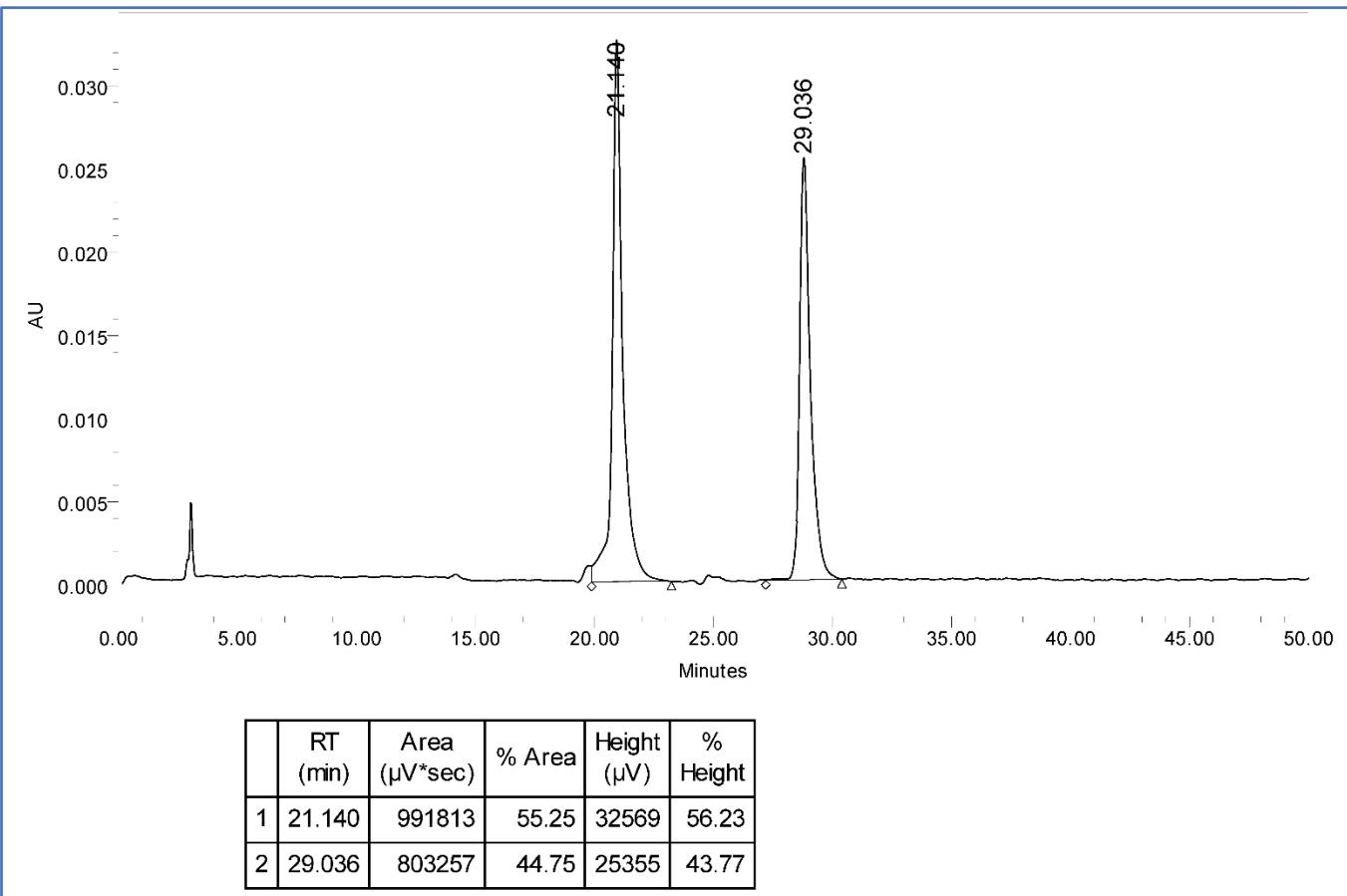
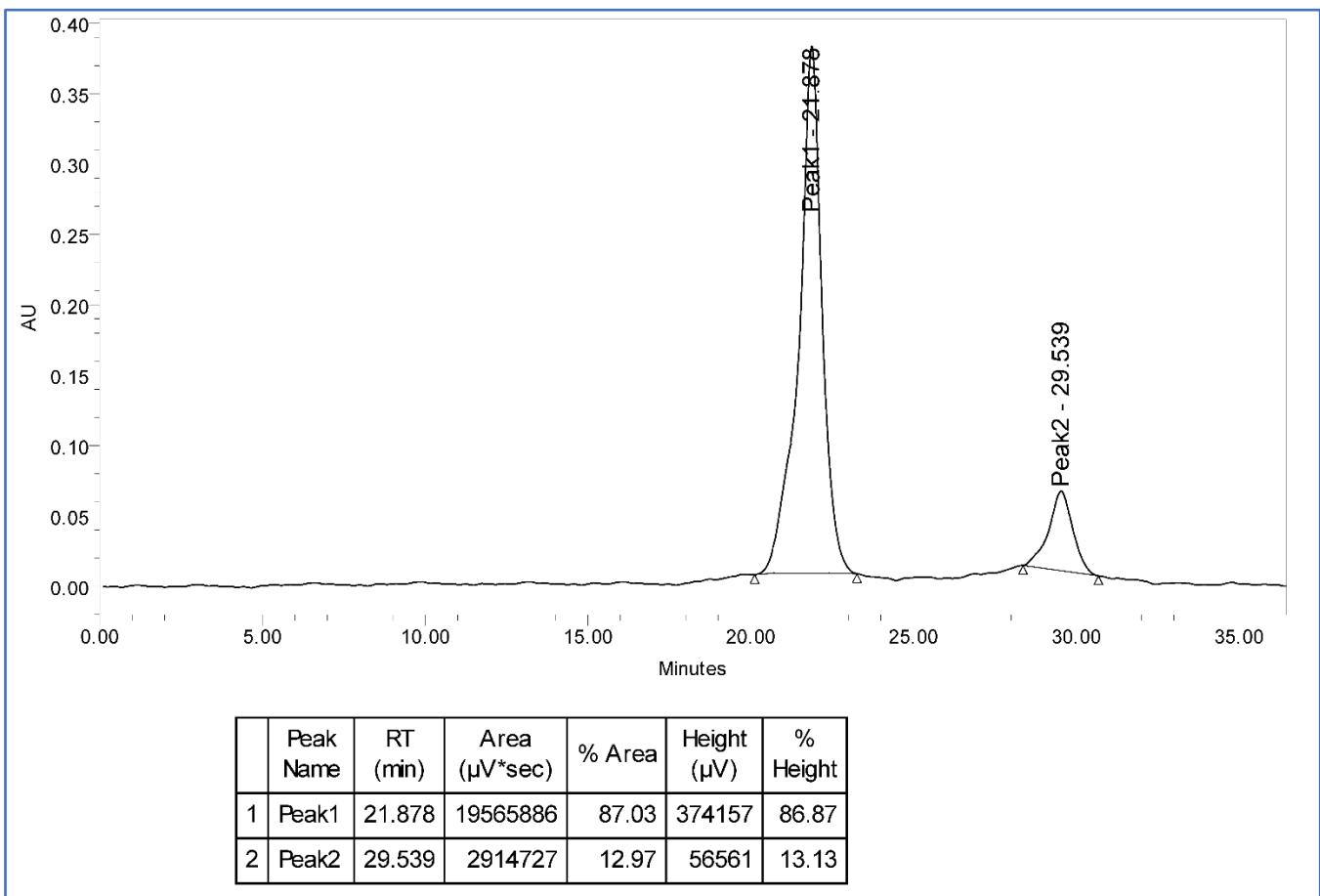


Figure S44: HPLC data of enantioenriched and racemic of **8m**.

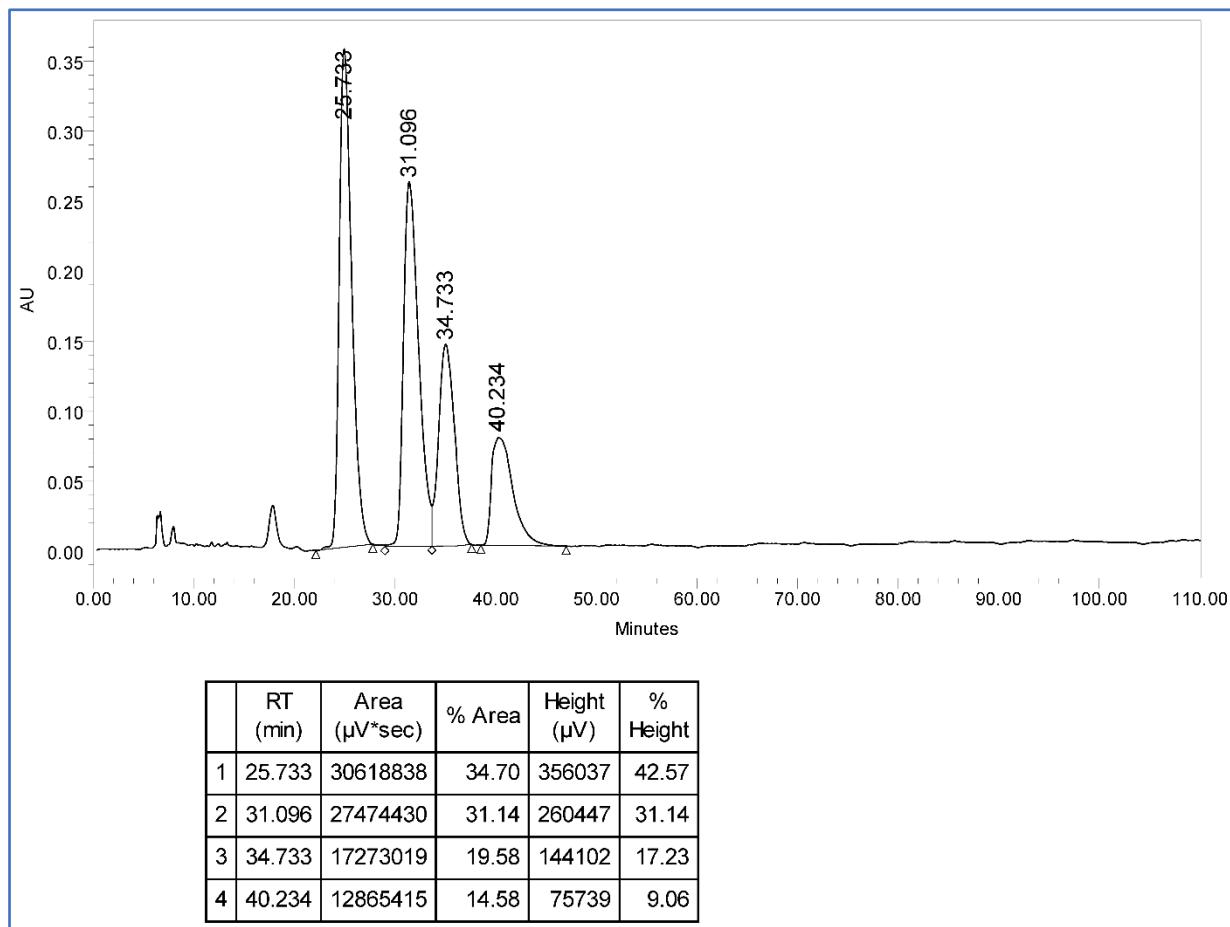
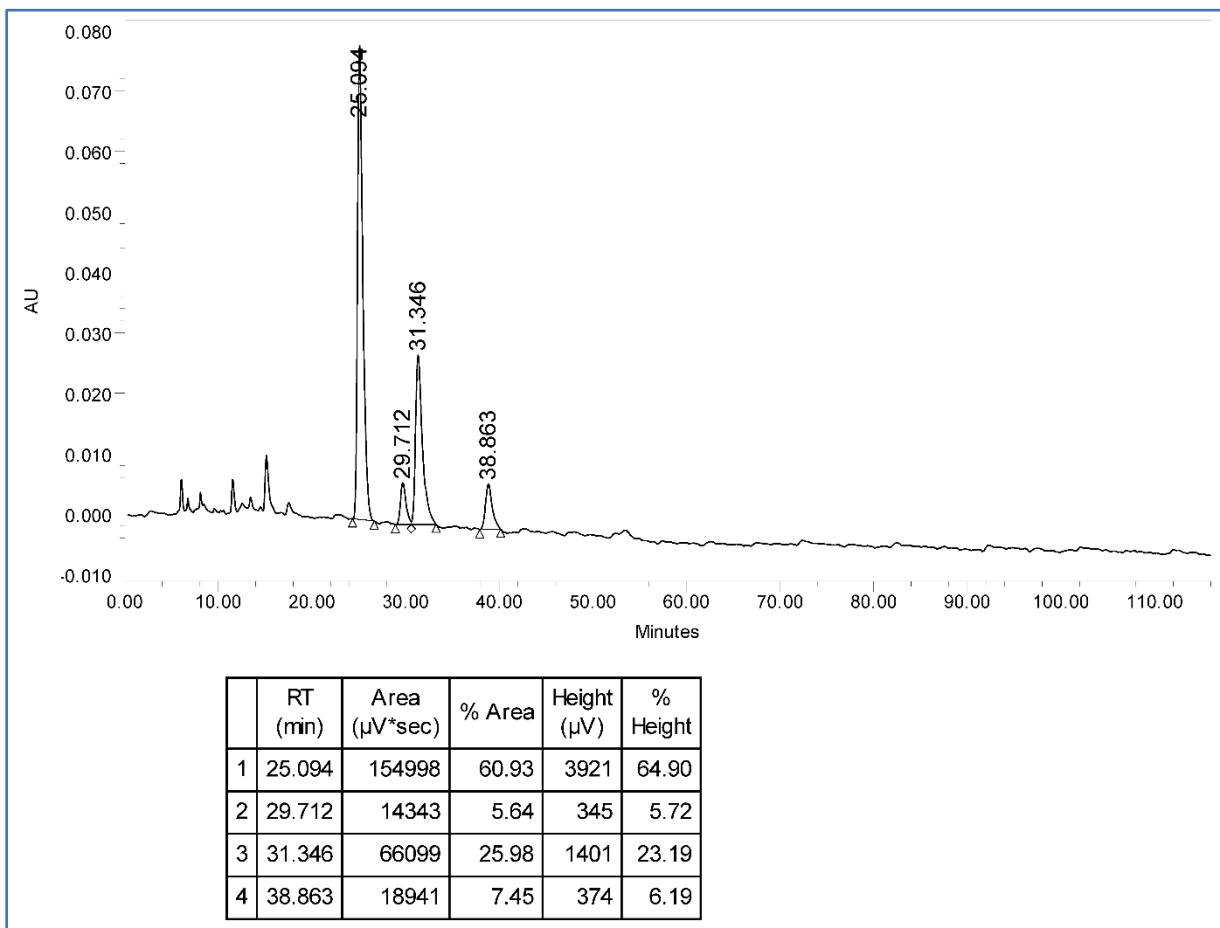


Figure S45: HPLC data of enantioenriched and racemic of **8n**.

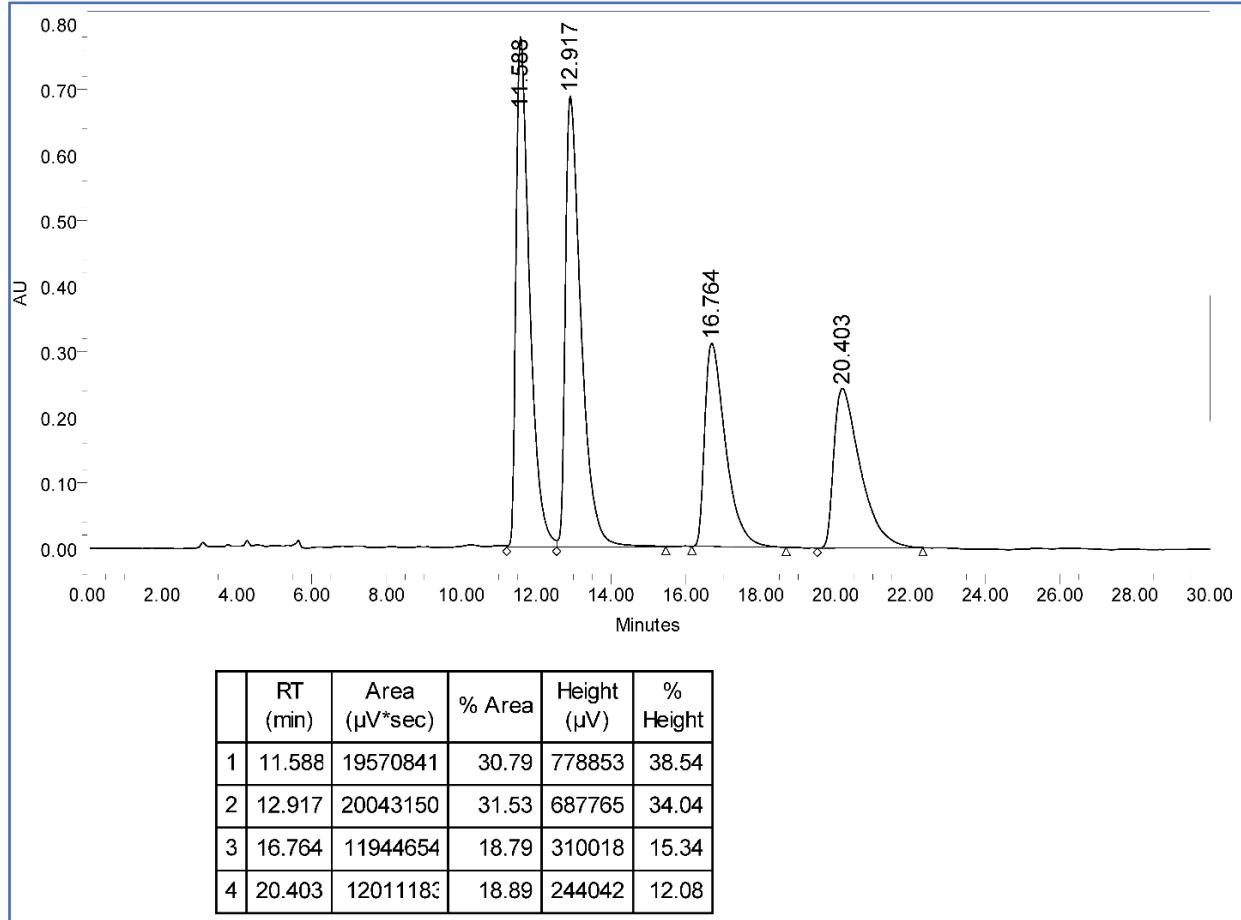
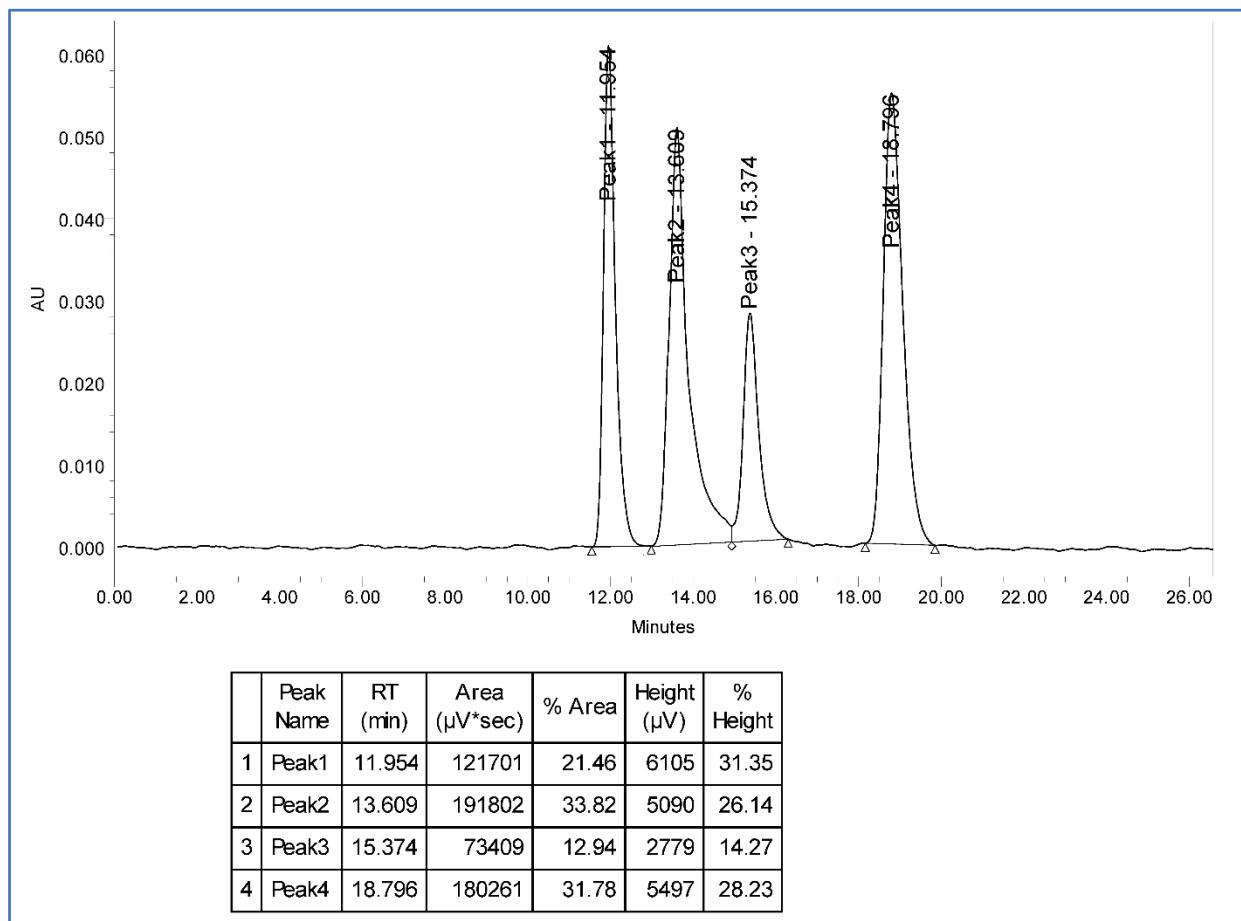


Figure S46: HPLC data of enantioenriched and racemic of **80**.

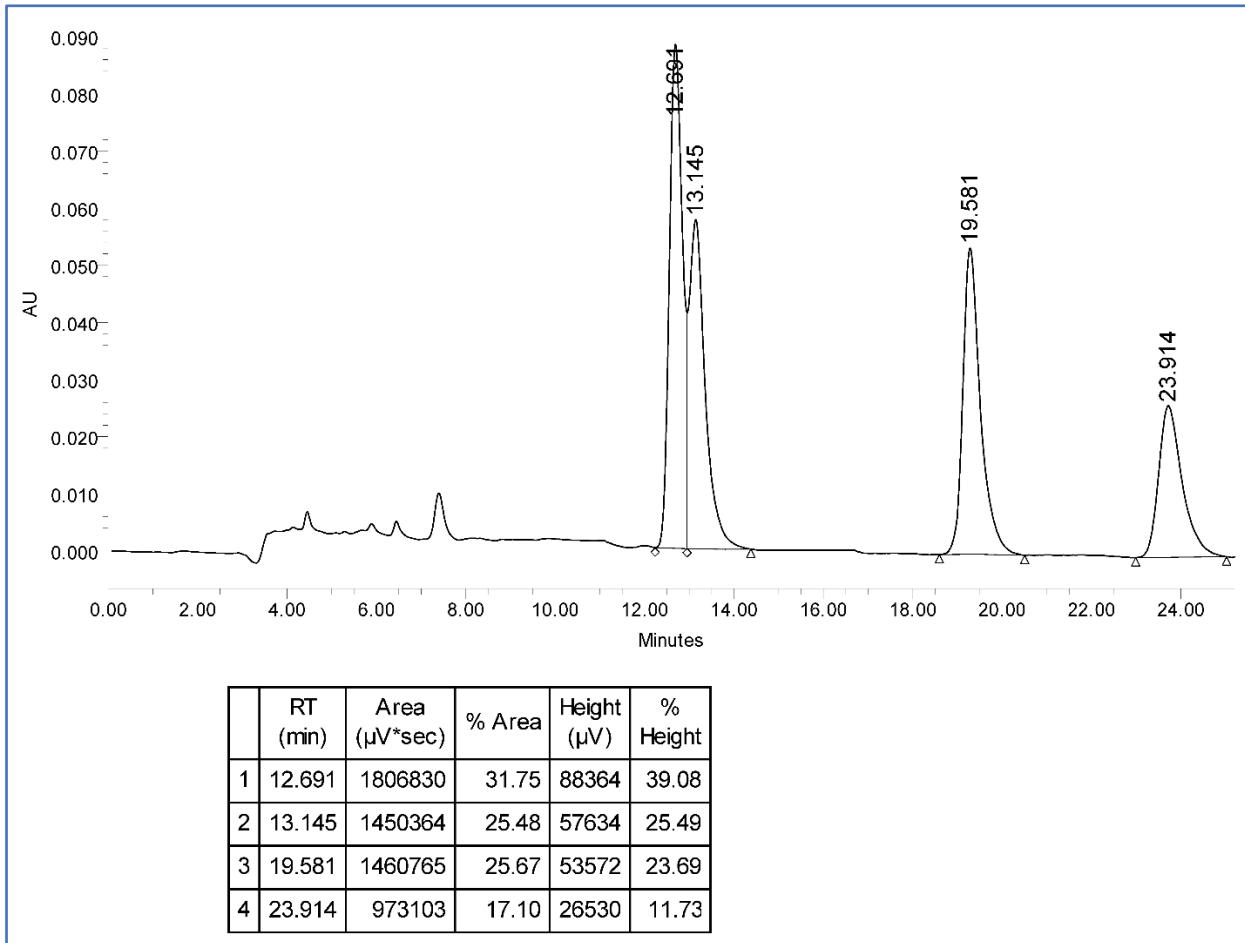
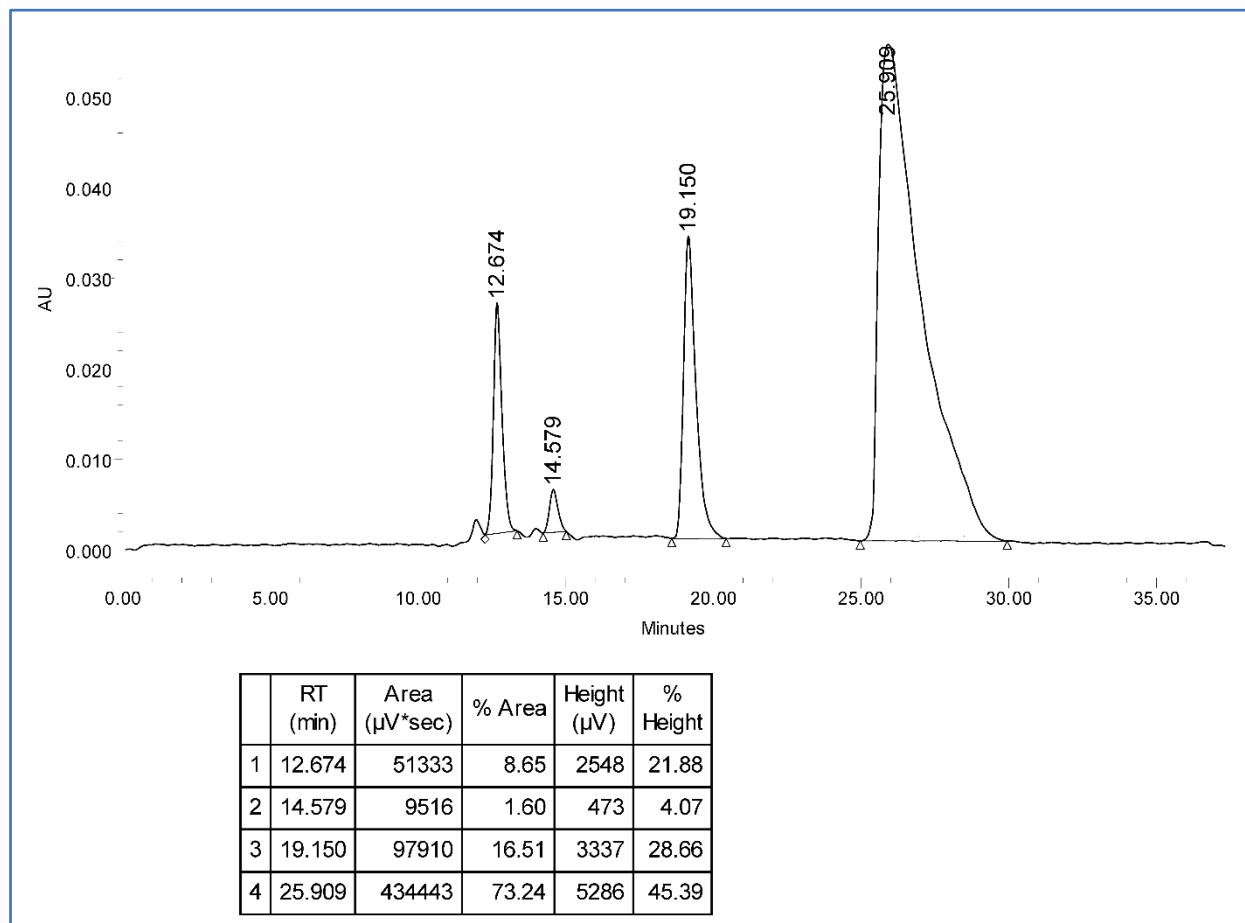


Figure S47: ^1H NMR and ^{13}C NMR of **9a**.

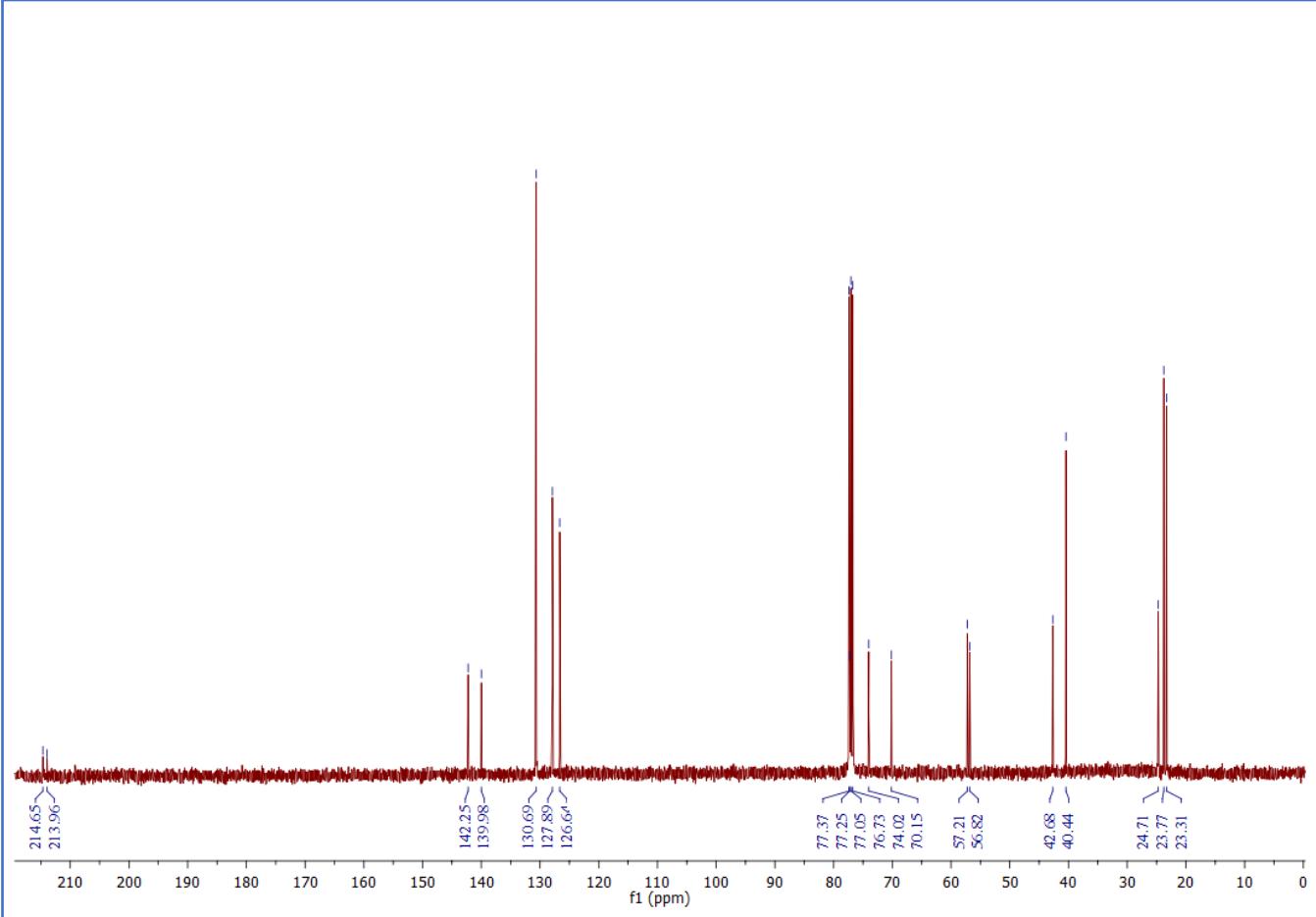
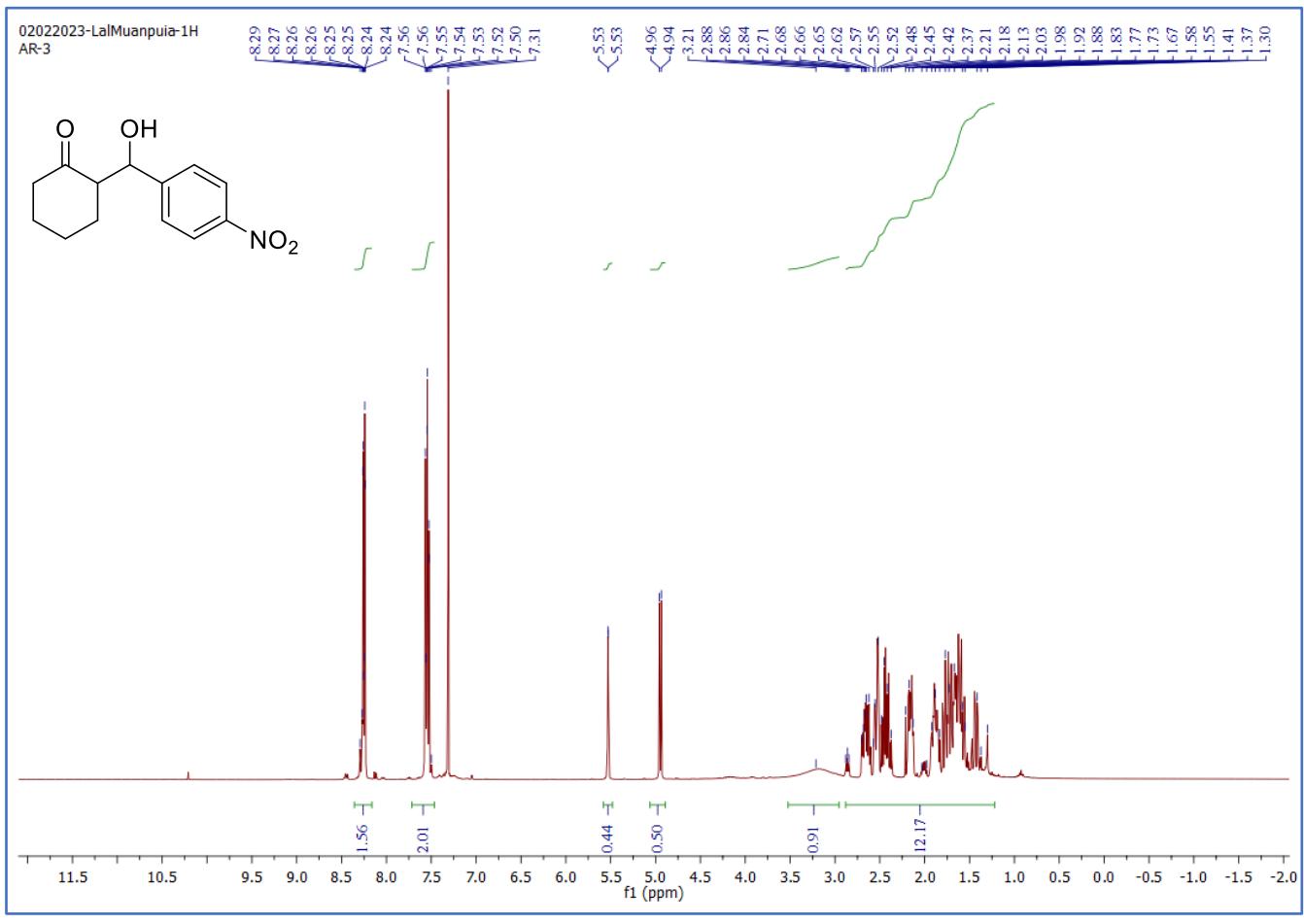


Figure S48: ^1H NMR and ^{13}C NMR of **9b**.

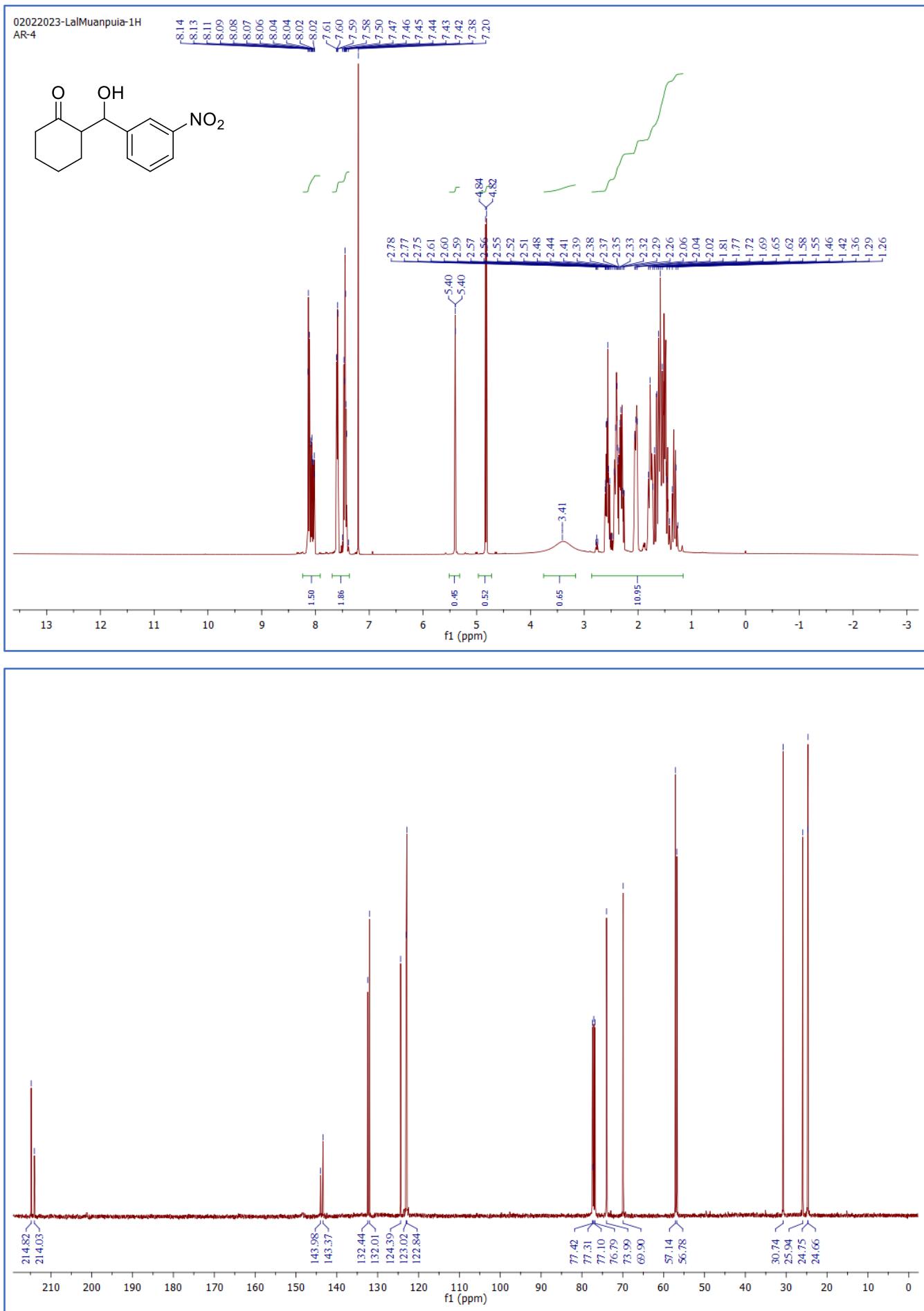


Figure S49: ^1H NMR and ^{13}C NMR of **9c**.

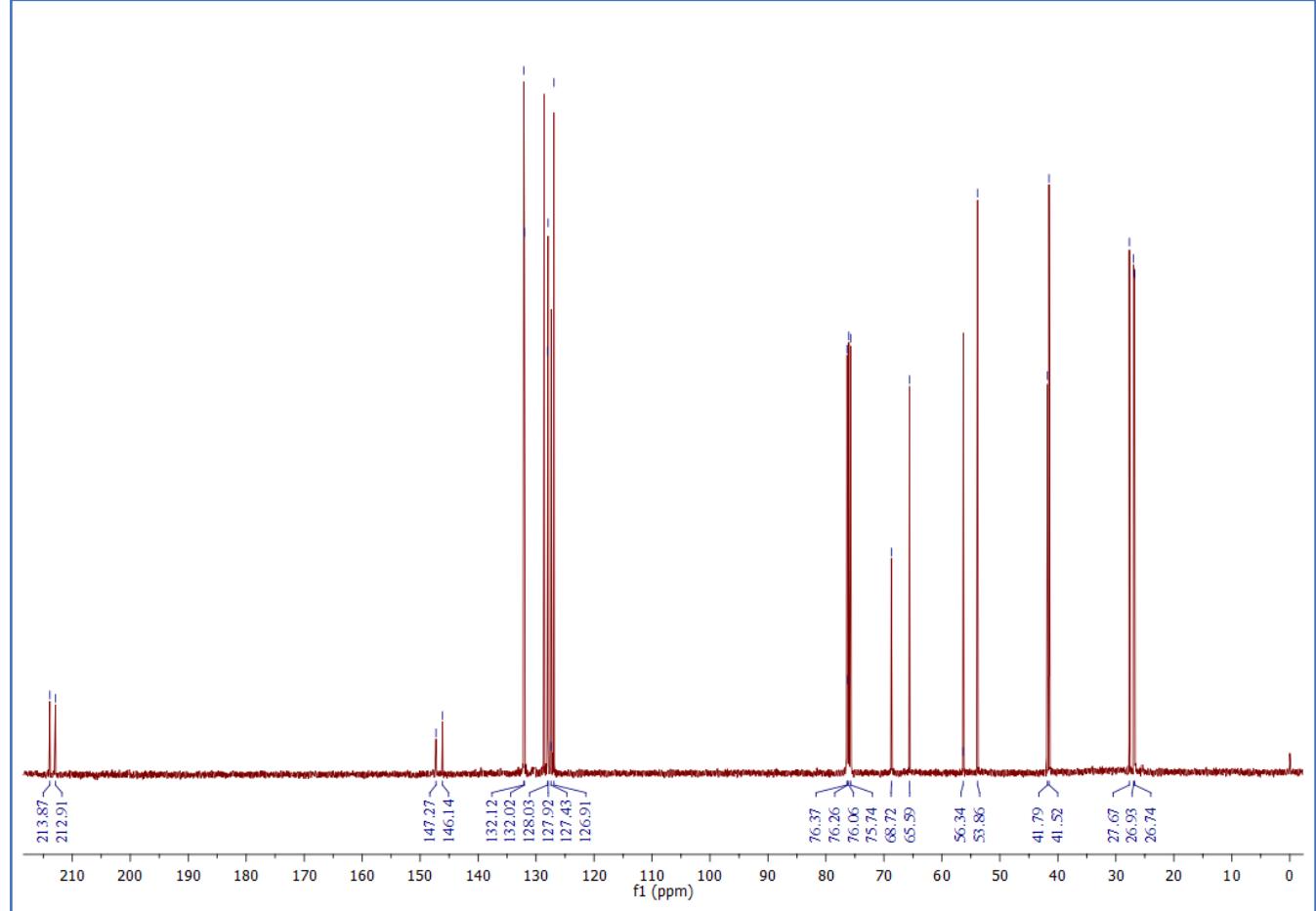
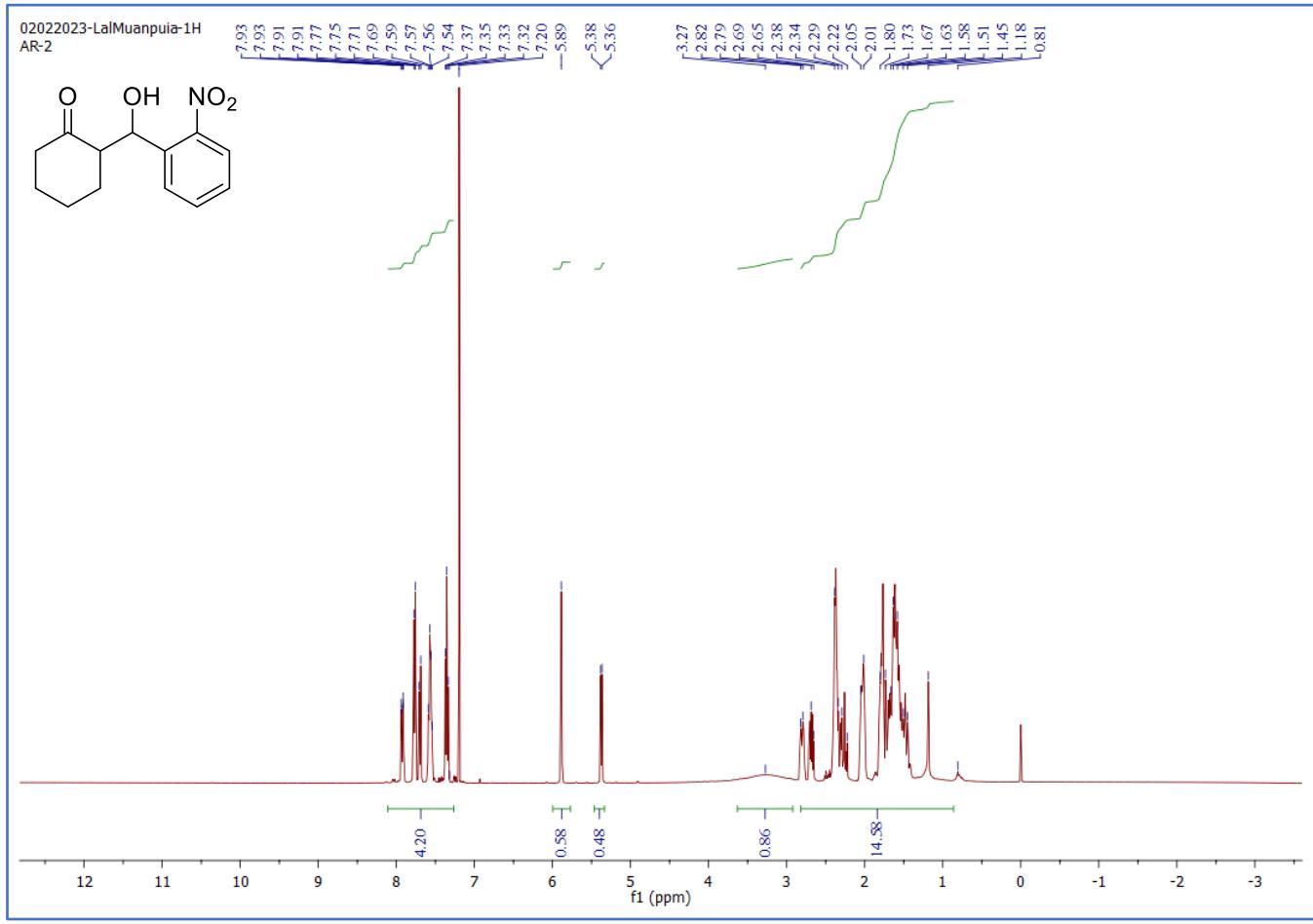


Figure S50: ^1H NMR and ^{13}C NMR of **9d**.

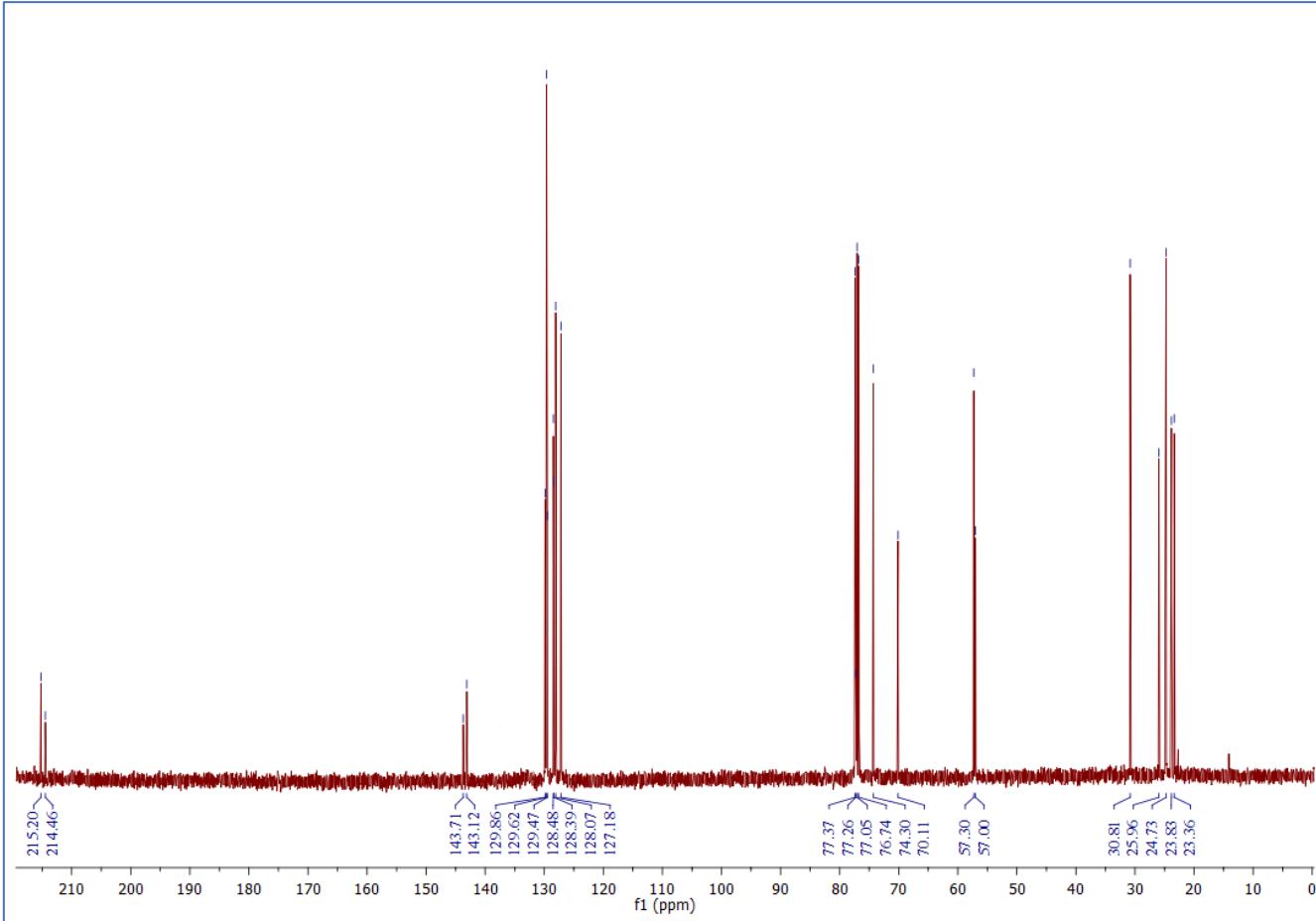
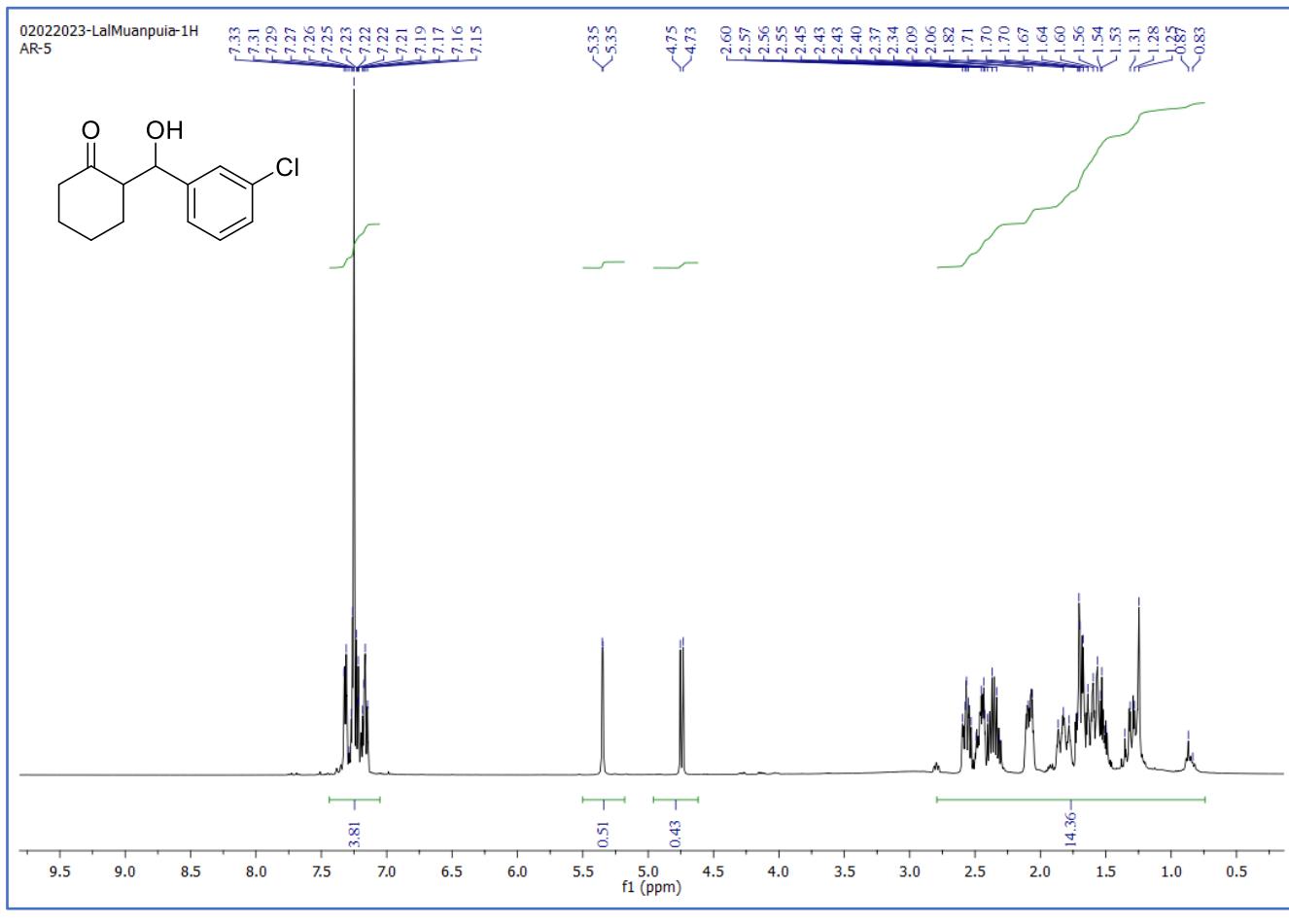


Figure S51: ^1H NMR and ^{13}C NMR of **9e**.

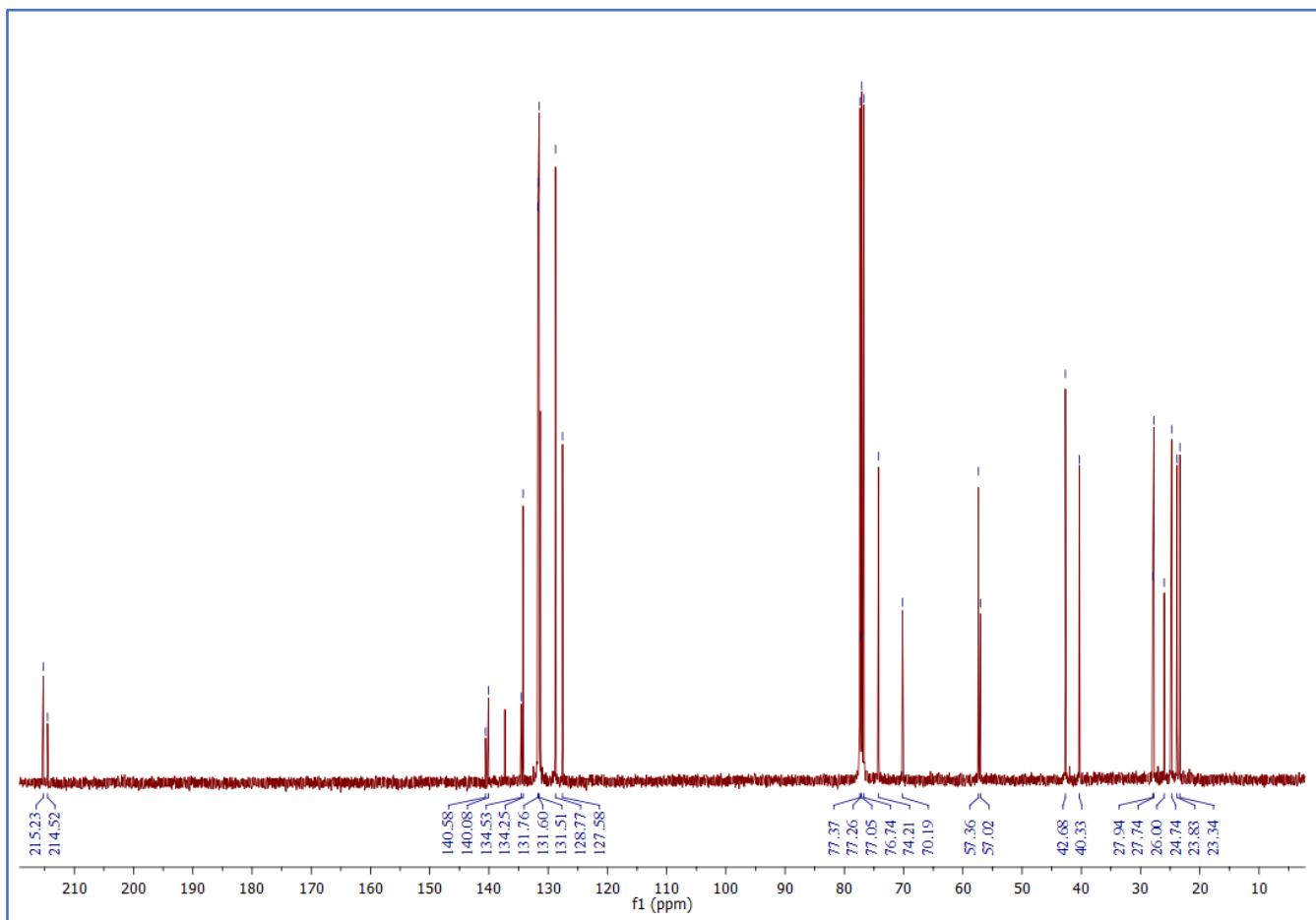
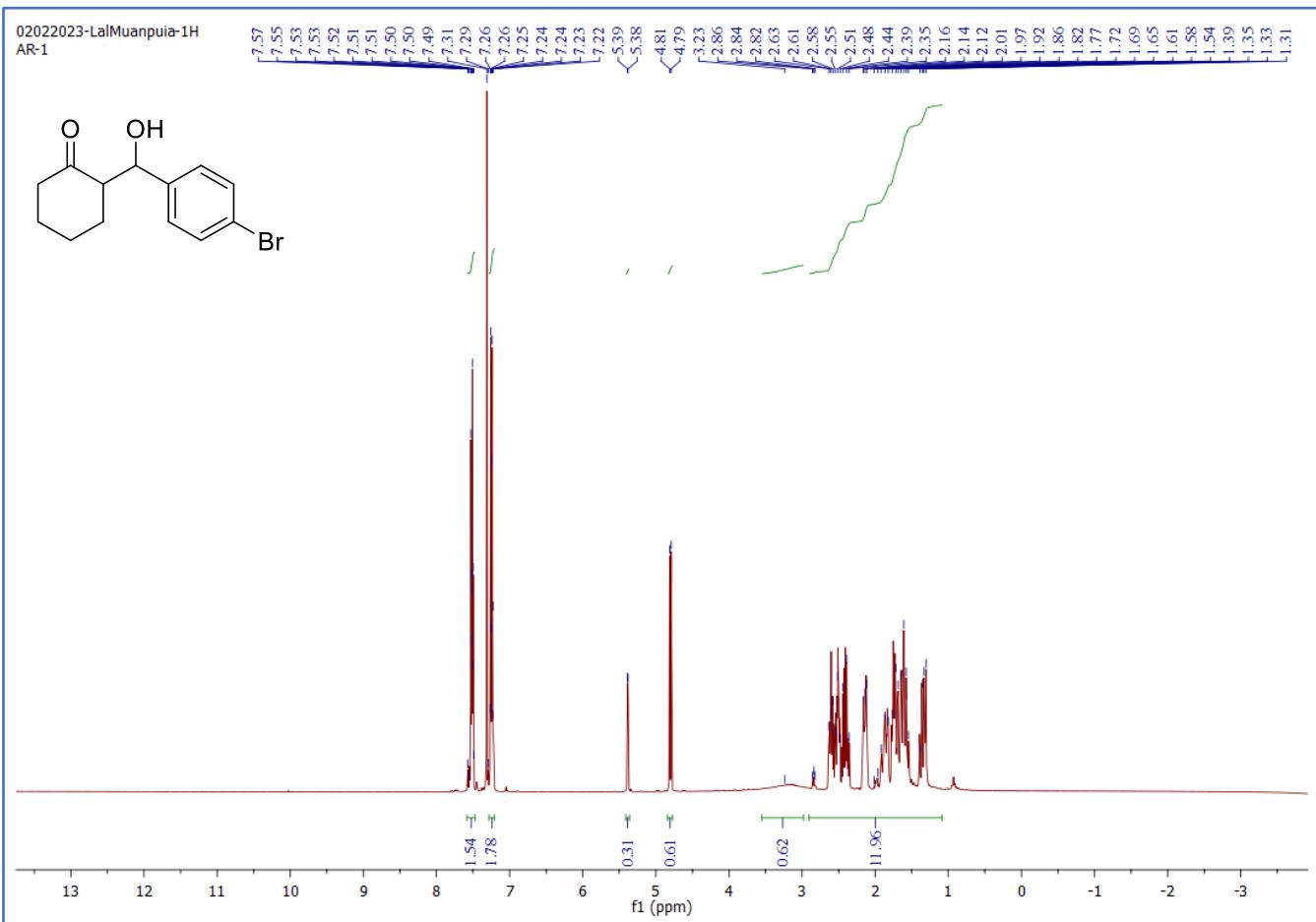


Figure S52: HPLC data of enantioenriched and racemic of **9a**.

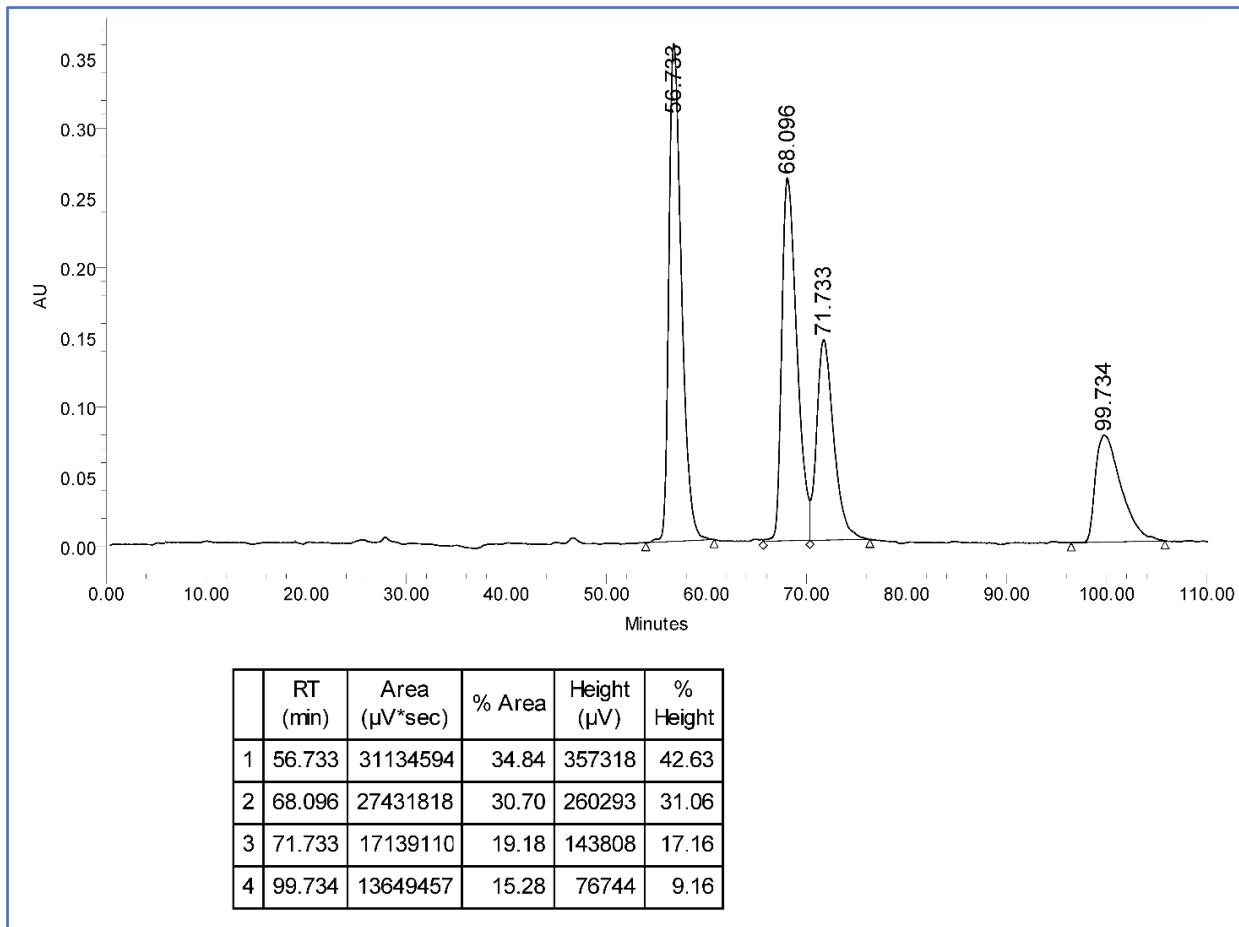
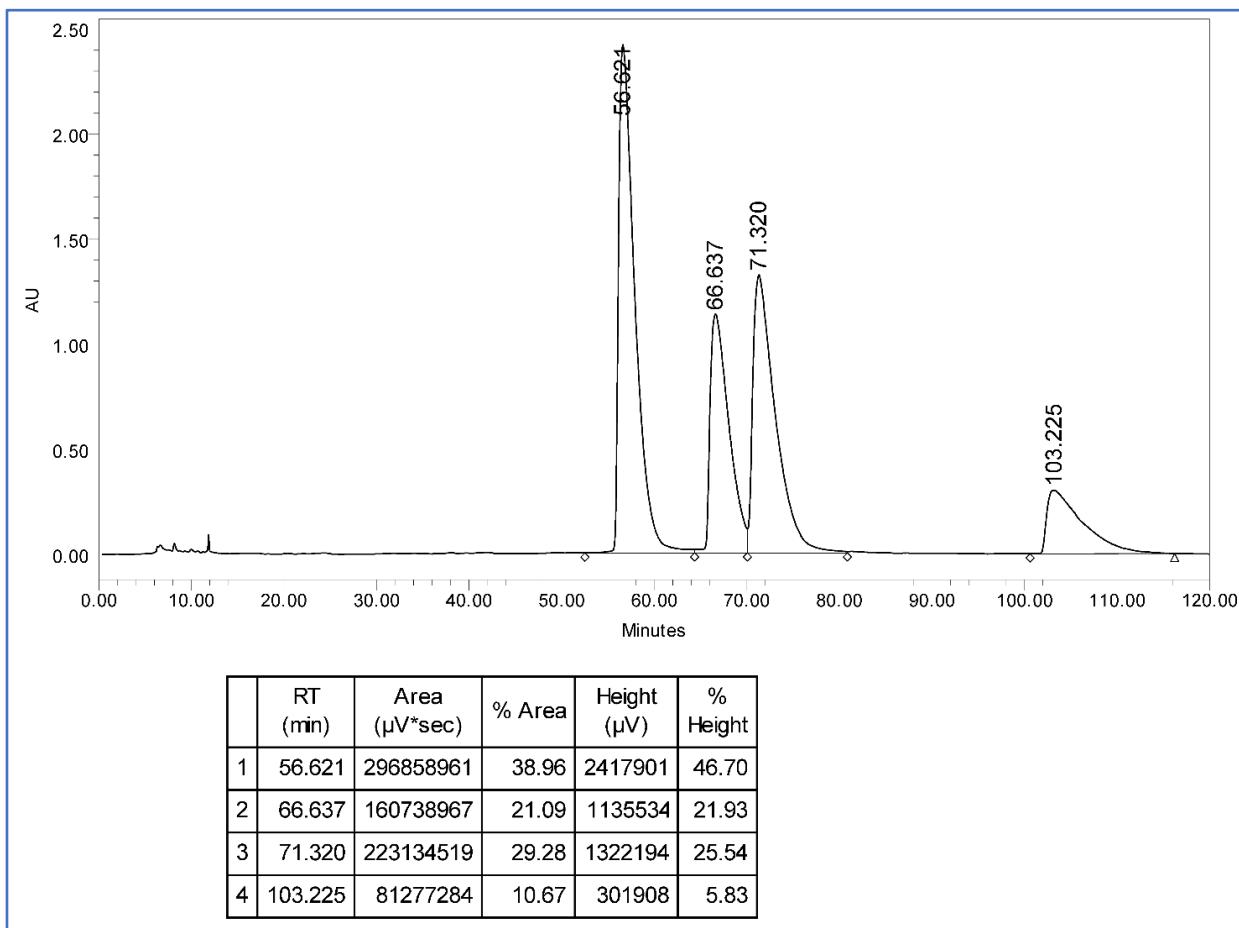


Figure S53: HPLC data of enantioenriched and racemic of **9b**.

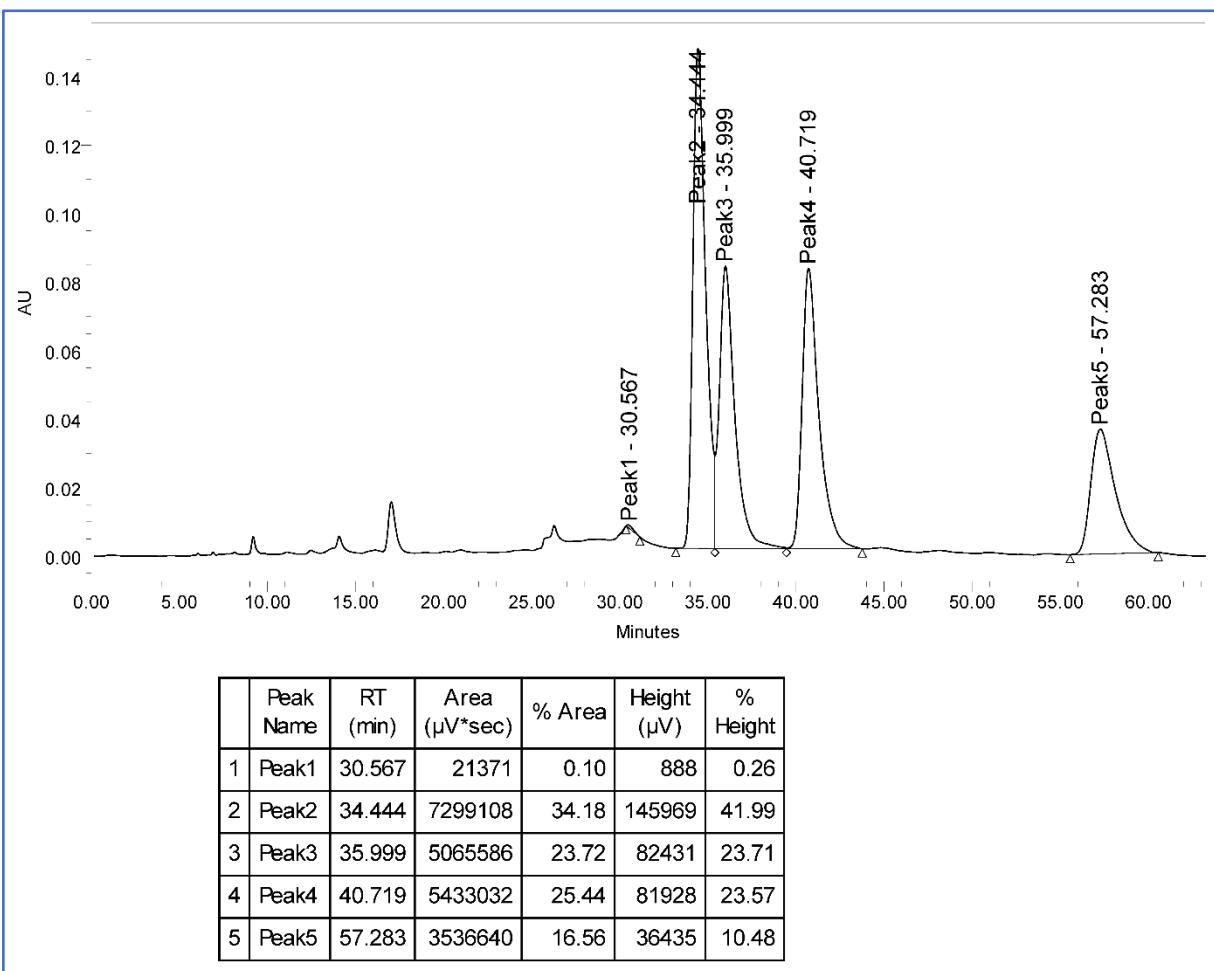
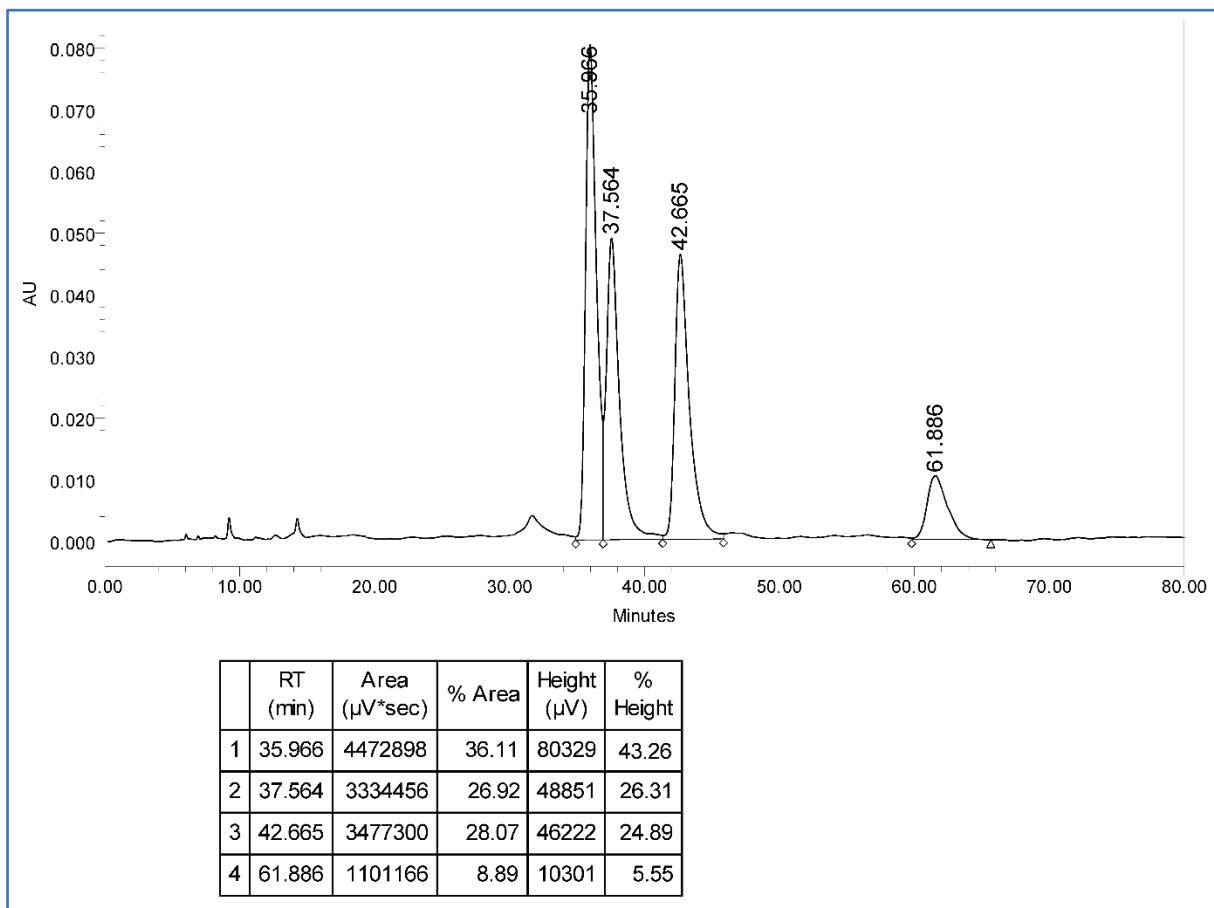


Figure S54: HPLC data of enantioenriched and racemic of **9c**.

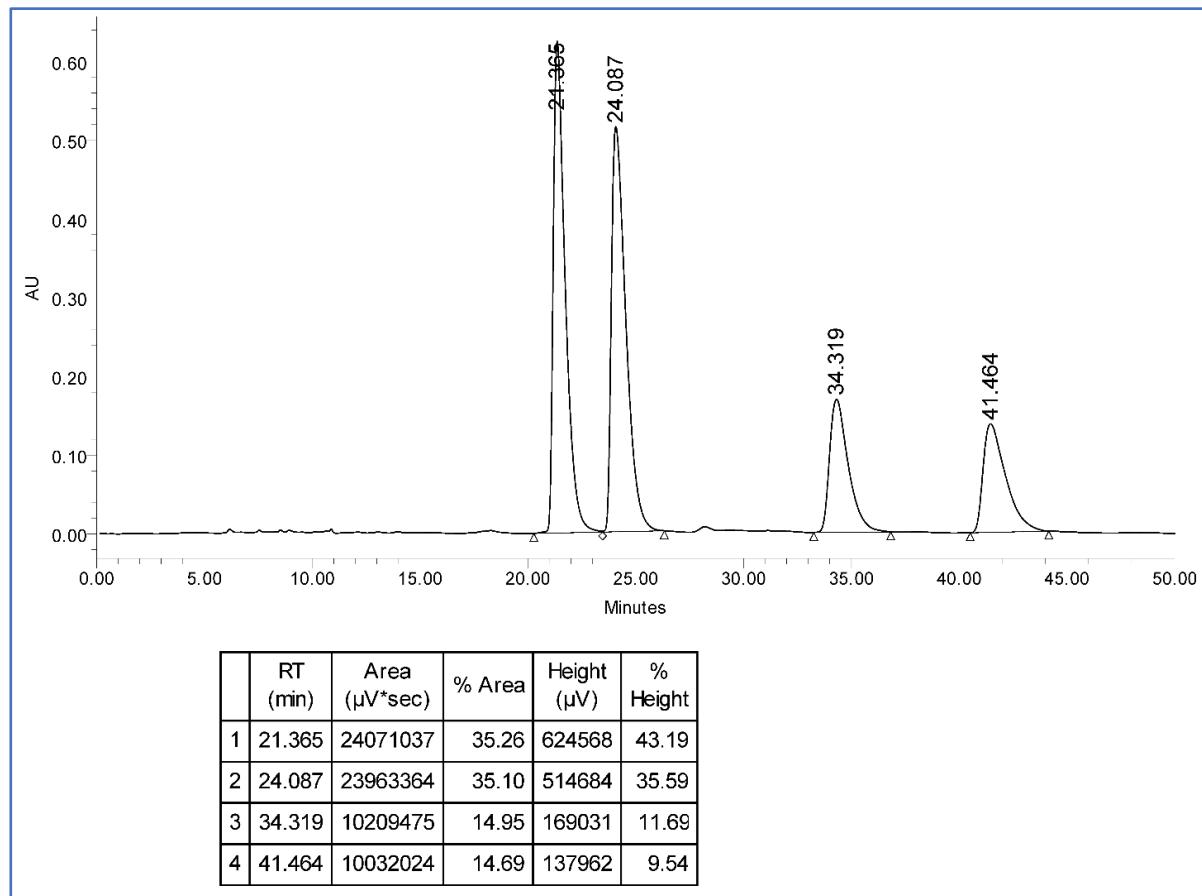
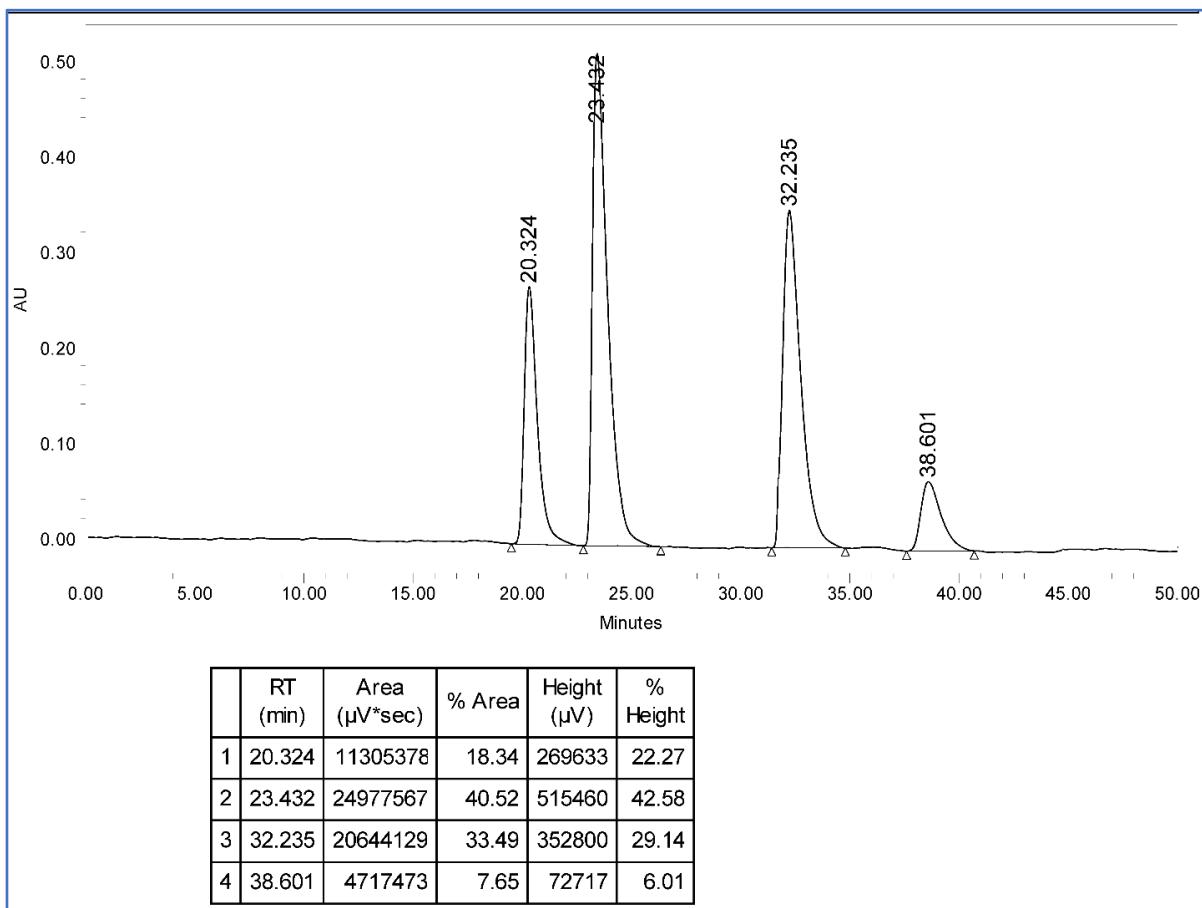


Figure S55: HPLC data of enantioenriched and racemic of **9d**.

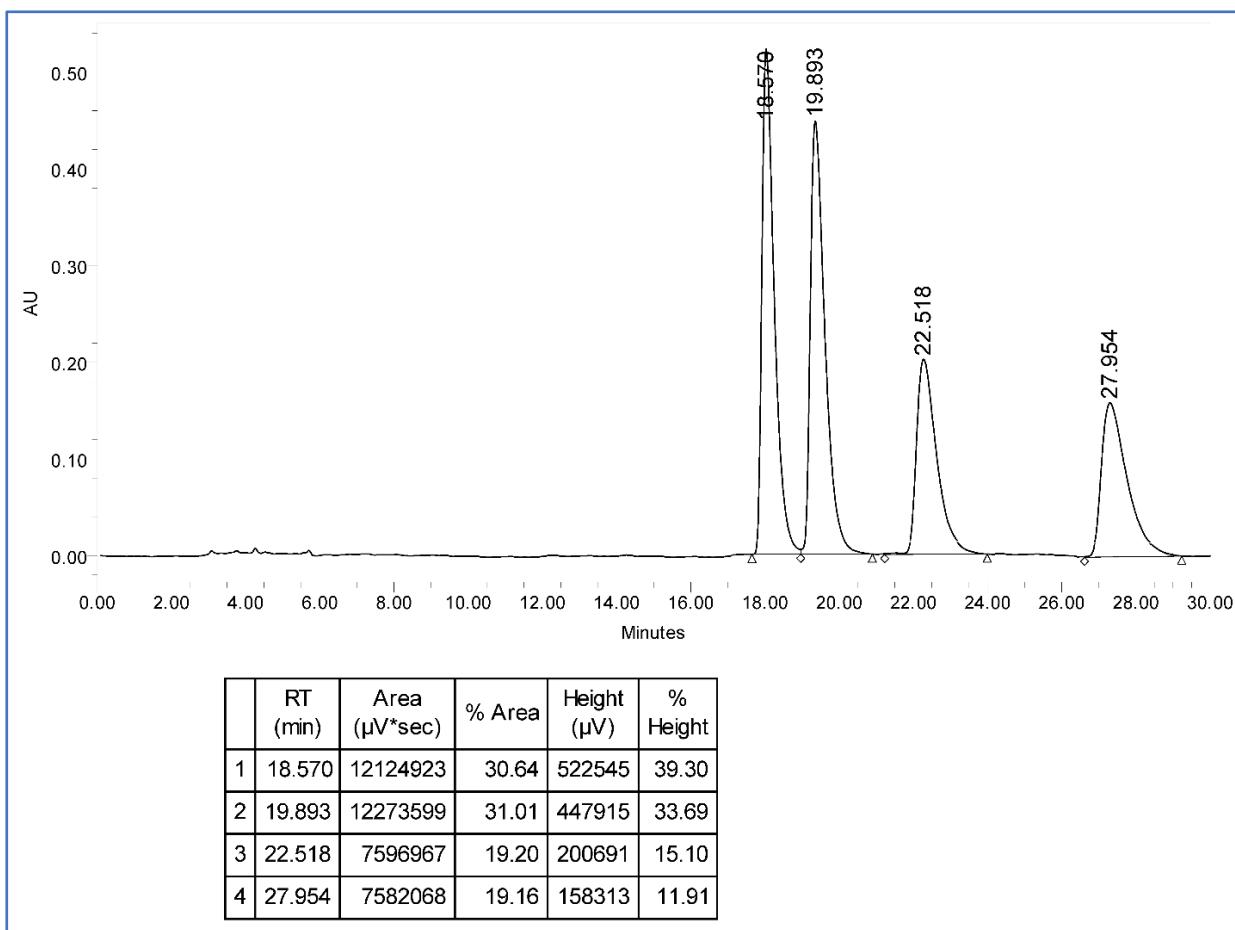
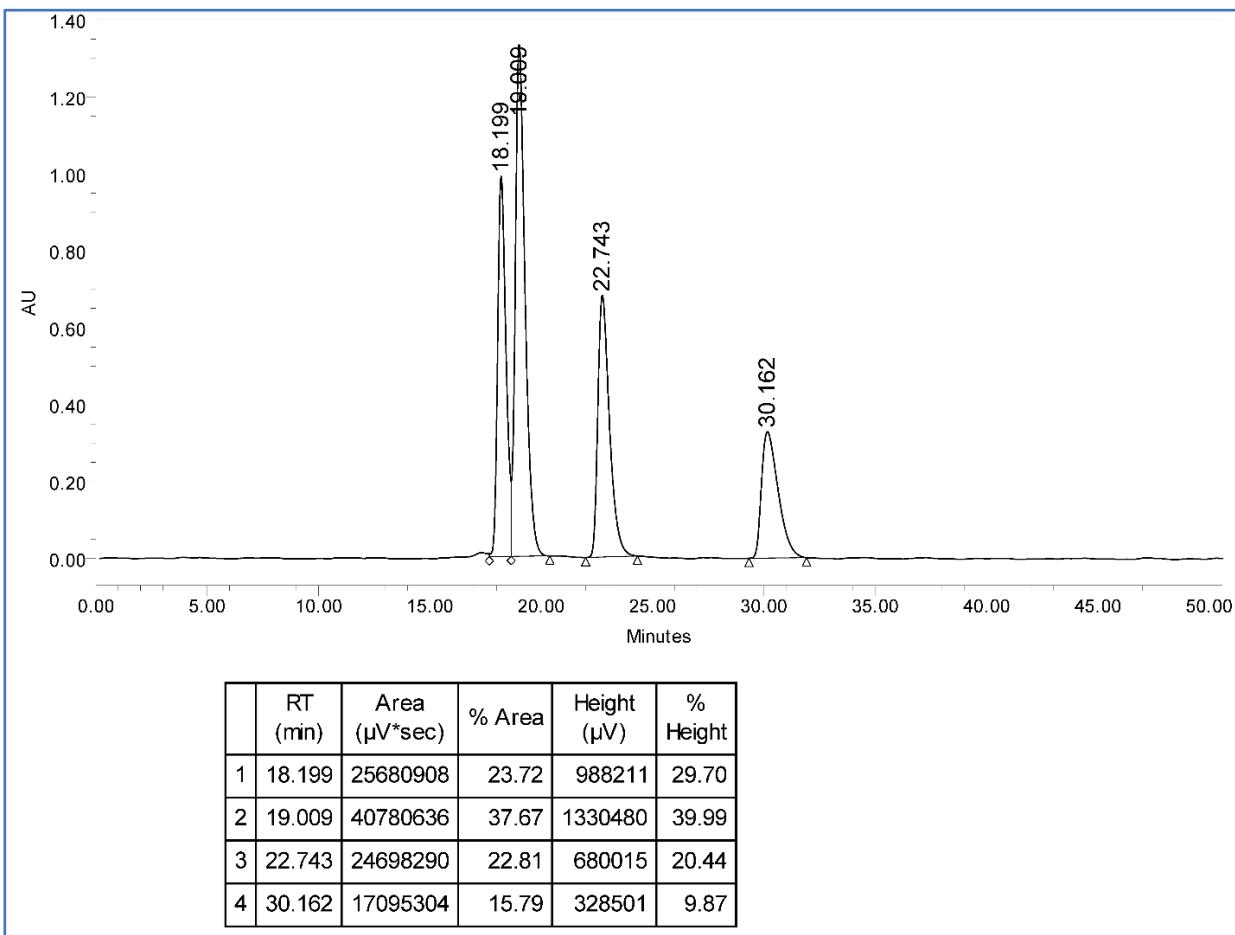


Figure S56: HPLC data of enantioenriched and racemic of **9e**.

