

Review

Application of Deferoxamine in Tissue Regeneration Attributed to Promoted Angiogenesis

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Abstract: Deferoxamine, an iron chelator used to treat diseases caused by excess iron, has had a Food and Drug Administration-approved status for many years. A large number of studies have confirmed that deferoxamine can reduce inflammatory response and promote angiogenesis. Blood vessels play a crucial role in sustaining vital life by facilitating the delivery of immune cells, oxygen, and nutrients, as well as eliminating waste products generated during cellular metabolism. Dysfunction in blood vessels may contribute significantly to the development of life-threatening diseases. Anti-angiogenesis therapy and pro-angiogenesis/angiogenesis strategies have been frequently recommended for various diseases. Herein, we describe the mechanism by which deferoxamine promotes angiogenesis and summarize its application in chronic wounds, bone repair, and diseases of the respiratory system. Furthermore, we discuss the drug delivery system of deferoxamine for treating various diseases, providing constructive ideas and inspiration for the development of new treatment strategies.

Keywords: deferoxamine; angiogenesis; wound healing; diabetes ulcer; bone repair



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1. Introduction

Iron is an essential trace element in the human body and plays an important role in biological activities such as oxygen transport, oxygen sensing, electron sensing, electron transfer, energy metabolism, and DNA synthesis [1]. However, excessive iron can lead to diseases such as hemochromatosis, thalassemia, myelodysplastic syndrome, aplastic anemia, etc. The iron-chelating drug deferoxamine (DFO) is commonly used in the treatment of such diseases [2]. DFO that has been approved for use by the Food and Drug Administration (FDA) is a natural product extracted from the fermentation liquor of *Streptococcus* spp. [3].

Iron is closely associated with inflammation [4–6]. During inflammation, the degradation of ferroportin increases, resulting in reduced iron excretion and elevated intracellular iron concentrations and ultimately leading to iron toxicity in cells and tissues [6–8]. DFO can bind to unliganded or incompletely liganded iron, rendering the ion inert and preventing its reaction with peroxides which, in turn, mitigates oxidative damage to tissues and alleviates oxidative stress [9,10]. Thus, DFO exhibits potent anti-inflammatory effects as an iron chelator and represents a promising therapeutic approach for mitigating inflammation in various autoimmune and inflammatory disorders [11]. In addition, studies have shown that Fe(II) in the prolyl hydroxylase domain (PHD) catalytic center can be exchanged or chelated by three hydroxamic acid groups of DFO, making PHD enzymes inactive [12]. Because PHD is a hypoxia-inducible factor (HIF) prolyl hydroxylase, it is known to play an important role in oxygen regulation in the physiological network. Hypoxia-inducible

factor-1 α (HIF-1 α) is an oxygen-sensitive molecule [13–16]. The expression of HIF-1 α is up-regulated in hypoxic conditions and subsequently regulates multiple target genes [17–21]. The PHD utilizes O₂ and α -ketoglutarate as substrates to hydroxylate two proline residues of HIF-1 α [22–24]. Then, the Von Hippel–Lindau protein (VHL) swiftly degrades the hydroxylated HIF-1 α [25,26] (Figure 1A). Thus, HIF-1 α -mediated gene transcription is inhibited [22]. DFO is able to activate and stabilize a hypoxic HIF-1 α pathway by rendering PHD inactive [27,28]. Then, upregulated HIF-1 α expression can increase the expression of vascular endothelial growth factor (VEGF, a key signaling molecule in the induction of angiogenesis), platelet-derived growth factor (PDGF), stromal cell-derived factor-1 (SDF-1), and other growth factors, thus stimulating angiogenesis [29–33] (Figure 1B). Numerous studies have demonstrated that DFO, functioning as an iron chelator, can effectively induce the accumulation of HIF-1 α , subsequently leading to a significant promotion in endothelial tube formation, cell proliferation, and migration [34–36].

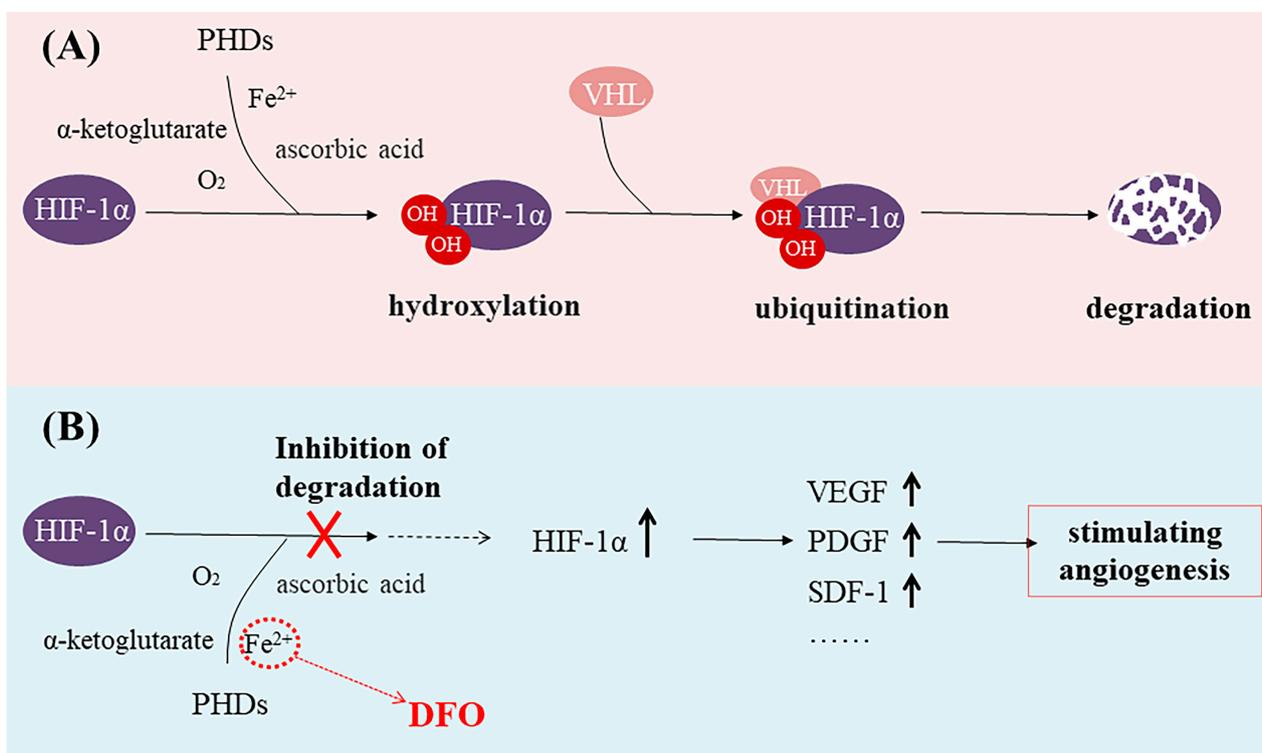


Figure 1. (A) The prolyl hydroxylase domain (PHD) utilizes O₂ and α -ketoglutarate as substrates to hydroxylate two proline residues of hypoxia-inducible factor-1 α (HIF-1 α), leading to the degradation of HIF-1 α . (B) Deferoxamine (DFO) binds to Fe²⁺, makes PHD enzymes inactive, and stabilizes the expression of HIF-1 α .

The blood vasculature is a closed circulatory system and includes networks of arteries, veins, and capillaries [37]. They play a crucial role in sustaining vital life by facilitating the delivery of immune cells, oxygen, and nutrients, as well as eliminating waste products generated during cellular metabolism [38–42]. The endothelial cells (ECs) are enveloped by mural cells to varying degrees to form blood vessels in various circulatory network locations. Endothelial cells (ECs) line the innermost layer of all of these vessels and exhibit a high degree of heterogeneity among different sections of the vasculature. They play an important role in sensing the circulating environment and responding to extrinsic signals [38]. The process of blood vessel development is intricate, with our current understanding indicating that endothelial cells are the earliest differentiated blood vessel cells during embryonic development and play a pivotal role in the formation of blood vessel walls and the establishment of complete blood vessel networks [43–46]. In adult

organisms, ECs rarely proliferate and remain dormant, but they retain the ability to rapidly form new blood vessels in nutrient-deficient, ischemic/hypoxic environments to restore blood flow (providing oxygen and nutrients) in order to support tissue growth and function [47]. VEGF is implicated in multiple steps of vascular EC development [48] and is a key signaling molecule in the induction of angiogenesis. DFO is able to render the PHD inactive to activate and stabilize HIF-1 α in order to increase the expression of VEGF in cells (such as stem cells, human dermal fibroblast cells, and human umbilical vein endothelial cells) [27,28]. VEGF can trigger quiescent ECs to become activated [47] in order to promote cell proliferation and migration and thus promote angiogenesis.

Dysfunction in blood vessels may significantly contribute to the development of life-threatening diseases [49]. Anti-angiogenesis therapy and pro-angiogenesis/angiogenesis strategies have been frequently recommended for various diseases [47]. Chronic, non-healing wounds are a persistent medical problem, and reduced blood vessel growth is a key reason many chronic wounds are difficult to heal [50–52]. Thus, targeted angiogenesis therapy is playing an increasingly important role as a therapeutic strategy for wound healing [50,53,54]. In the skeletal system, the local vascular system is actively involved in bone formation and bone resorption [55,56]. Angiogenesis plays a central role in bone reconstruction by providing oxygen, minerals, nutrients, and growth factors to the injured microenvironment [57,58]. Angiogenesis also plays a pivotal role in the intricate process of fetal lung development and subsequent tissue regeneration following lung transplantation. Based on the role of DFO in promoting angiogenesis, this review discusses the application of DFO in various diseases and provides constructive ideas and enlightenment for the development of more therapeutic strategies for DFO.

2. Chronic Wounds

Wound healing typically moves through four overlapping stages: hemostasis/coagulation, inflammation, proliferation, and maturation/remodeling [59]. Chronic wounds fail to proceed through a normal, orderly, and timely repair sequence, resulting in delayed wound healing or even non-healing wounds [50]. Chronic wounds are classified by their etiology into four categories: arterial, diabetic, pressure, and venous ulcers [59,60]. Chronic wounds are often accompanied by high levels of proinflammatory cytokines, persistent infections, the formation of drug-resistant microbial biofilms, and senescent cells that do not respond to repair stimuli [59]. Over the years, chronic wounds have caused great suffering for patients. Non-healing chronic wounds impose physical, psychological, social, and financial burdens on individuals and the broader health system [61]. Reduced angiogenesis is one of the primary causes of the non-healing nature of chronic wounds [50,62]. During the healing process, angiogenesis is an important behavior in the phase of proliferation. Stimulated by moderate hypoxia, cytokines, and protein hydrolases, endothelial cells are activated to proliferate and migrate toward pro-angiogenic signals (such as VEGF and PDGF) to induce angiogenesis [63]. In addition, pericytes and smooth muscle cells can stabilize neovascularization [50]. The new blood vessel network delivers oxygen and nutrients to the damaged tissue and maintains cell function. It can also provide the wound site with cytokines and other substances necessary to repair the damaged tissue. Therefore, promoting angiogenesis and rebuilding tissue blood flow are promising therapeutic targets of new therapies to promote chronic wound healing [50]. Numerous studies have demonstrated that the promotion of angiogenesis can enhance the healing process of chronic wounds [64–67]. It is worth noting that chronic wounds have a common characteristic: the local deposition of free iron [68]. By chelating iron deposited at the wound site, DFO not only mitigates oxidative stress but also activates the HIF-1 α /VEGF pathway (as mentioned previously), thereby facilitating neovascularization and, ultimately, promoting the healing of chronic wounds [69] (Figure 2).

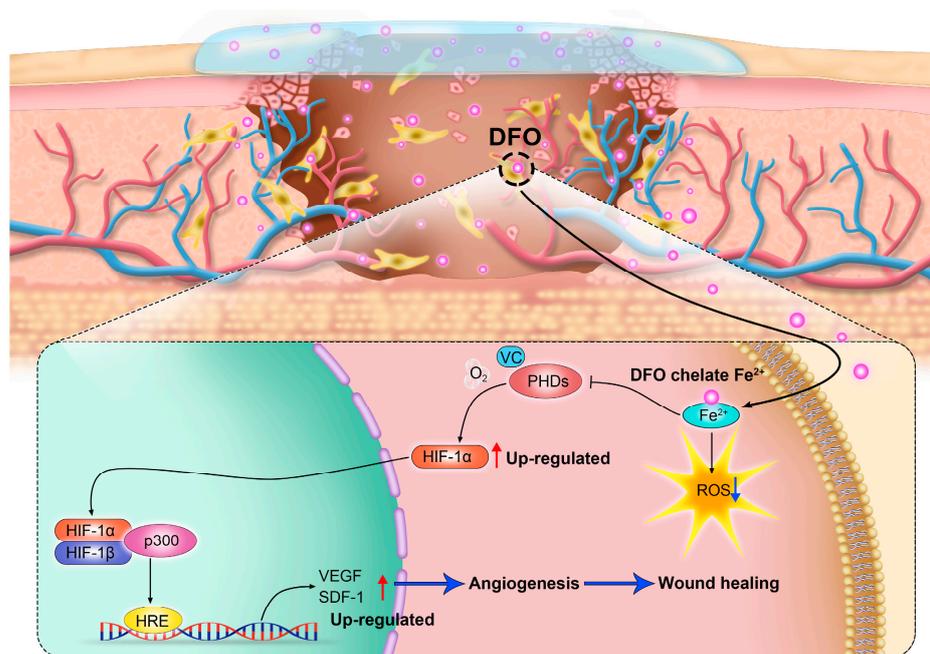


Figure 2. By regulating the hypoxia-inducible factor-1 α (HIF-1 α) signaling pathway, deferoxamine (DFO) promotes angiogenesis and accelerates wound healing.

2.1. Diabetic Wounds

Persistent hyperglycemia has been shown to detrimentally impact vascular function and elevate susceptibility to infection. Therefore, diabetic wounds frequently do not follow the four stages of wound healing and often develop into chronic wounds [70]. Diabetic foot ulcers are a classic chronic wound [59,60]. Increasing evidence suggests that defective angiogenesis significantly contributes to a delay in diabetic wound healing because damaged blood vessels are unable to deliver critical oxygen and nutrients to the wounded tissue [71]. Thus, promoting angiogenesis is crucial in diabetic wound healing. This process depends on the proliferation and migration of endothelial cells in response to cytokines such as VEGF. As mentioned above, DFO can stimulate the HIF-1 α /VEGF pathway to stimulate angiogenesis; therefore, DFO is expected to promote diabetic wound healing.

A recent study investigated the effect of DFO on diabetic wounds. The researchers showed that DFO was able to enhance angiogenesis and accelerate wound healing in diabetic patients by accumulating HIF-1 α and regulating endothelial cell function [36]. Dominik Duscher et al. compared the efficacy of the hydroxylase inhibitor dimethyl oxalate (DMOG) and DFO in ameliorating diabetes-related skin wound healing defects by augmenting HIF-1 α activation both in vitro and in vivo. The findings demonstrated that DFO effectively stabilized HIF-1 α expression in the presence of hypoxia and hyperglycemia, surpassing the impact of DMOG on wound healing and angiogenesis in aged and diabetic mice. These results highlight the significant therapeutic potential of local administration of DFO for diabetic wounds [72].

During the past few years, researchers have worked to use an appropriate approach to enable DFO to perform better in treating diabetic wounds and reducing its side effects. Thus, researchers have taken an interest in utilizing wound dressings loaded with DFO to treat diabetic wounds. Hao Chen et al. utilized DFO-loaded hydrogel nanofibrous scaffolds and a DFO-loaded photo-crosslinked gelatin hydrogel to exploit their potential in promoting diabetic wound healing. The incorporation of DFO into the wound dressing created an optimal microenvironment for cell viability, adhesion, and proliferation. Moreover, the sustained release of DFO significantly enhanced neovascularization. Ultimately, both in vitro and in vivo experiments demonstrated the safety and efficacy of these strategies [73,74]. In addition, the co-delivery of various drugs which have complementary

bioactivity provides a better therapeutic strategy for treating diabetic wounds [75]. Due to the synergistic effect of combining DFO and liposome nanoparticles, drug delivery can be enhanced and maintained, thereby amplifying the therapeutic response. Asif Qayoom et al. developed lecithin-based DFO nanoparticles which exhibit superior potential in treating diabetic wounds compared to using DFO alone [76]. Lingzhi Kong et al. demonstrated the synergistic effect of bioglass (BG) (which has been shown to promote vascular regeneration by modulating the expression of VEGF through the inclusion of Si ions) and DFO in promoting revascularization and developed an injectable hydrogel incorporating both BG and DFO for the treatment of chronic diabetic wounds. The findings revealed that the hydrogel exhibited superior efficacy in enhancing wound healing compared to either BG or DFO alone [3]. Bacterial infection and insufficient angiogenesis are the main factors that hinder the healing of diabetic ulcers. Therefore, antimicrobial and angiogenic treatment strategies are key to treating diabetic ulcer wounds. Shan Gao et al. loaded a microneedle patch with the antibacterial drug tetracycline hydrochloride and DFO at the same time, and the prepared microneedle patch not only had good antibacterial properties but also promoted angiogenesis, thus promoting the healing of diabetic ulcer wounds [77]. The in-depth investigation of DFO in diabetic ulcer treatment underscores the pivotal role of angiogenesis in wound repair, thereby providing valuable insights for advancing wound healing therapies [78,79].

2.2. Burn Wounds

Burn wounds may secrete a large amount of exudate, increasing excessive inflammation and leading to wound infection, scar formation, and even damage to new blood vessels. Eventually, these wounds may progress into chronic non-healing wounds [80]. The combined action of VEGF, PDGF, and other factors could effectively improve cell (cells involved in skin wound healing and inflammation) function, including proliferation, migration, differentiation, collagen remodeling, etc. Therefore, therapeutic strategies that regulate growth factors at the wound site may promote skin tissue regeneration in burn wounds [81]. Angiogenesis provides nutrients and oxygen to damaged tissues, is essential for maintaining normal cell function, and is an important part of tissue regeneration [47]. Oxidative stress and inflammation may mediate cellular damage and tissue destruction, as the burn wound continues to progress after the abatement of the initial insult [82,83]. Intervening in oxidative stress-induced excitation damage can prevent the progression of partial-thickness second-degree burns to a deep partial-thickness burn or of a deep second-degree burn becoming a third-degree burn [82]. Trace metals such as iron and copper may induce vital cellular injuries via lipid peroxidation [84,85]. Amina El Ayadi et al. treated porcine brass comb burn models with the Livionex formulation (LF) lotion (containing ethylenediaminetetraacetic acid as a metal chelator), and the experimental results showed that the application of LF lotion onto burn wounds provided protection oxidative damage and inflammation and prevented subsequent burn wound progression [82]. Therefore, it is imperative to devise therapeutic strategies that promote angiogenesis and prevent excessive inflammation. DFO can inhibit the activity of PHD, upregulate the expression of HIF-1 α , and subsequently stimulate the expression of various growth factors (such as VEGF, PDGF, and SDF-1) [29–33]. In addition, DFO, as an iron-chelating agent, can chelate free iron at the wound site, which is expected to prevent excessive inflammation at the burn wound site and promote wound healing [86,87]. Wu Hongfu et al. developed a hydrogel based on the anti-inflammatory effect of glycyrrhizic acid (GA) and the angiogenic effect of DFO. They demonstrated that the hydrogel effectively reduced pro-inflammatory mediators (TNF- α and IL-6) and upregulated anti-inflammatory mediators (TGF- β 3) while promoting proliferation, migration, and angiogenesis of human umbilical vein endothelial cells (HUVECs). Finally, the evaluation of a deep second-degree burn wound model in rats demonstrated that the synthetic hydrogel expedited burn wound healing, providing substantiation for its potential application in treating burn wounds through anti-inflammatory and angiogenesis-promoting mechanisms [80].

2.3. Leg Ulcers as the Main Complications of SCD

Leg ulcers are the main complications of sickle cell disease (SCD); about 2.5–40% of SCD patients have the risk of developing leg ulcers because of chronic hemolysis and poor angiogenesis. Leg ulcers are often difficult to heal [88]. As early as 1968, DFO was FDA-approved for chelation of the excess iron produced by hemolysis in SCD patients [89,90]. In order to achieve effective and localized delivery of DFO for ulcer treatment, Melanie Rodrigues developed a novel transdermal delivery system for DFO (DFO-TDDS) that utilizes reverse micelles to ensure continuous delivery of DFO to the skin surface. Rodrigues' team initially created excision wounds in a transgenic sickle cell mouse model expressing > 99% human sickle hemoglobin (HbSS-BERK); these were subsequently treated with DFO-TDDS. The findings demonstrated that DFO-TDDS significantly expedited wound healing in HbSS-BERK mice by effectively chelating excessive free iron [91]. Their research makes it possible to translate DFO-TDDS into an effective treatment for patients with sickle cell leg ulcers (SCLUs).

3. Bone Repair

Angiogenesis is critical for bone regeneration [92–96]. Following fractures, a substantial quantity of locally produced angiogenic growth factors stimulates the process of angiogenesis. These vascular networks not only facilitate the supply of oxygen and nutrients [97–101] but also contribute to the recruitment of bone marrow stem cells (BMSCs) for osteoblastic differentiation and provision of essential ions required for subsequent mineralization stages. Thus, they play a pivotal role in bone regeneration [101–103]. Prolyl hydroxylase inhibitors have demonstrated efficacy in activating the HIF-1 α pathway [104–106], thereby effectively promoting angiogenesis (Figure 3). Thus, DFO as a prolyl hydroxylase inhibitor has been proposed for use in bone repair [107–110]. Rui Shi et al. co-encapsulated DFO-loaded NPs and free DFO in nanofibers through coaxial electrospinning and investigated its effects on cell viability, migration, and osteogenic differentiation. The results suggested that DFO maintained cell viability and promoted the migration of human mesenchymal stem cells. Alkaline phosphatase (ALP) activity, calcium deposition, and the expression of osteogenesis-related markers and HIF-1 α were all increased with DFO, indicating that DFO may accelerate bone regeneration [111].

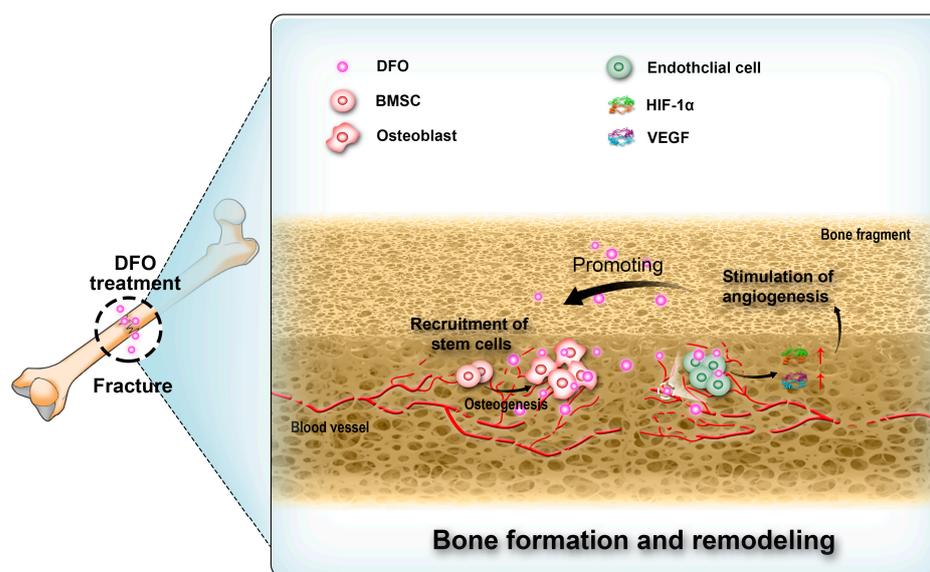


Figure 3. Deferoxamine (DFO) interacts with endothelial cells, bone marrow stem cells (BMSCs), and osteoblasts in the process of bone regeneration.

3.1. Distraction Osteogenesis

Distraction osteogenesis (DO) is a technique to initiate regeneration by using mechanical strain to enhance the biological response of injured tissue. It is a metabolic-dependent reconstruction process that relies heavily on adequate local blood supply [112]. However, when distraction osteogenesis is used for bone repair after radiotherapy, the distraction osteogenesis therapy is ineffective due to the reduction of blood vessels [113]. Researchers have confirmed that DFO can optimize the quality and quantity of the regeneration tissue in the sites of mandibular distraction by augmenting vascularity [114,115]. Moreover, in a DO model featuring radiation-induced impairment of bone healing, angiogenesis, and biomechanical properties, DFO has demonstrated the ability to restore vascularity to the distraction site, thereby counteracting the detrimental effects caused by radiation therapy (XRT) and facilitating bone regeneration [108,116,117]. The findings presented here enhance the potential utility of vascular enhancement as a means to optimize bone regeneration in DO.

3.2. Steroid-Induced Osteonecrosis of the Femoral Head

Steroids can reduce the expression of VEGF and disrupt vascularization [118]. Therefore, addressing angiogenesis is critical for the treatment of steroid-induced osteonecrosis of the femoral head (ONFH). Jia Li et al. first reported that local DFO administration can improve angiogenesis and bone repair in early-stage models of rabbit ONFH, which may be an efficient, economical, and facile method to treat early-stage ONFH [119].

3.3. Bone Defects

In recent years, researchers have made fresh attempts to use DFO treatment in bone repair. Biomimetic materials produced by 3D printing offer a good treatment method for bone transplantation after major defects, and they also make up for the disadvantages of bone autografting [120]. Although the scaffold-based approach has a great therapeutic potential, it relies on the construction of new blood vessels for regeneration; thus, induction of neovascularization at the site of regeneration is crucial [121]. Justin Drager et al. used 3D technology to print biomimetic materials which were transplanted into a rabbit model of bone segmental defect and, through local injection of DFO, increased the formation of blood vessels at the injured site, creating an environment conducive to bone repair [120]. The findings present a novel concept for the design of bone scaffolds with potential for vascularization.

4. Lung and Airway

4.1. Bronchopulmonary Dysplasia

Bronchopulmonary dysplasia (BPD) is a frequent complication in premature infants which seriously affects the health of children. Fetal lungs undergo development in a hypoxic intrauterine environment where HIF-1 α plays a crucial role in promoting normal organ growth and maturation. However, premature exposure to oxygen reduces its expression in preterm infants, hindering alveolar and angiogenic processes while disrupting pulmonary development [122–125]. PHD inhibitors have been shown to promote pulmonary angiogenesis in BPD primate models by increasing HIF-1 α and downstream angiogenic factors (Figure 4) [126,127]. As a PHD inhibitor, DFO has been shown to improve lung development in BPD rats by accumulating HIF-1 α [128]. Yanru Chen et al. verified, in a mice BPD model, that deferoxamine-loaded aerosol particles (DFO@APs) can release DFO in the alveolar interstitium, thus promoting the reconstruction of microvasculature and, ultimately, inducing lung development for treating BPD [129].

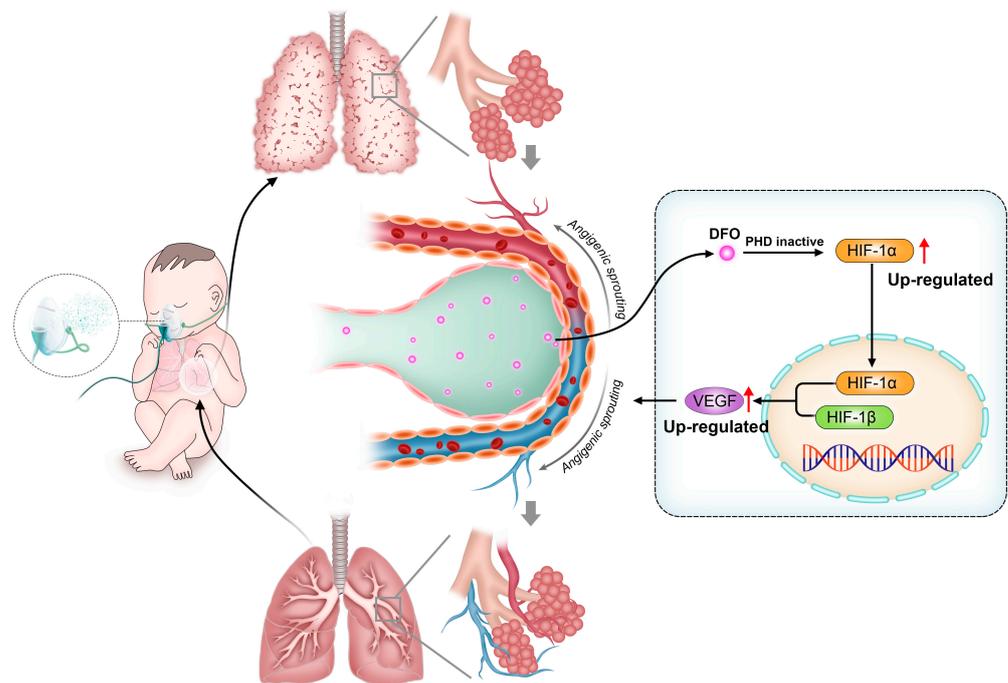


Figure 4. In bronchopulmonary dysplasia (BPD) models, deferoxamine (DFO) promotes angiogenic sprouting by regulating hypoxia-inducible factor-1 α (HIF-1 α) signaling pathways.

4.2. Complications of Lung Transplantation

Lung transplantation is often necessary for the treatment of various end-stage lung diseases; however, the occurrence of donor bronchial ischemia poses a significant risk for the development of airway anastomotic complications, potentially leading to severe postoperative complications and transplant failure. Therefore, it is important to promote microvascular repair and alleviate allograft ischemia and hypoxia [130]. Xinguo Jiang et al. developed a DFO nanoparticle and confirmed its ability to improve mouse orthotopic tracheal transplant model complications by producing angiogenic growth factors and reducing ROS production, suggesting that the use of DFO is an effective strategy to reduce postoperative complications following lung and airway transplantation [131].

5. Spinal Cord Injury

Spinal cord injury (SCI) is a serious traumatic disease. As we know, iron overload, reactive oxygen species accumulation, lipid peroxidation, and glutamate accumulation are all associated with spinal cord injury and are also inducers of ferroptosis (ferroptosis is a regulated form of cell death characterized by iron-dependent phospholipid peroxidation) [132–136]. As an iron death inhibitor, DFO can promote the recovery of spinal cord injury by inhibiting iron death [134,137]. Despite the fact that the therapeutic effect of DFO on SCI has been demonstrated in previous studies, the exact mechanism of action is still controversial [138,139]. Guoqing Tang et al. hypothesized that DFO improves spinal cord compression by promoting angiogenesis and demonstrated, in a moderately compressed SCI rat model, that DFO-induced revascularization via activation of the HIF-1 α /VEGF pathway is a key mechanism for improving prognosis in spinal cord injury [140]. In addition, the influx of erythrocytes caused by hemorrhage during SCI provides abundant iron sources at the site of the injury, and the increase in iron concentration, iron metabolism, and superoxide metabolism promote each other, producing a large number of free radicals, mediating the oxidative stress response that contributes to secondary injury [141,142]. Many scholars have studied secondary injury responses, among which the inflammatory cascade caused by tumor necrosis factor- α (TNF- α) is considered to be the core of the secondary injury method [143]. Hence, controlling the inflammatory response is essential for treating

SCI and preventing further injury. The potential mechanism of DFO in suppressing the inflammatory response following SCI involves chelation of locally produced iron from bleeding, thereby inhibiting TNF- α and interleukin-1 β (IL-1 β) production by macrophages and microglia. This subsequently promotes the polarization of macrophages from M1 to M2 phenotype, ultimately leading to inhibition of secondary SCI injury [144,145]. Taken together, these results show that DFO treatment reduces the development of inflammation and tissue injury associated with spinal cord trauma. This may accelerate the clinical application of DFO in SCI.

6. Others

During recent years, research on DFO promoting angiogenesis has become increasingly popular. Some researchers use DFO to treat traumatic brain injury. DFO can not only chelate excessive iron from bleeding to prevent oxidative damage through the blood–brain barrier, it can also achieve this through accumulating HIF-1 α in order to promote the expression of VEGF, subsequently improving hypoxia tolerance and promoting angiogenesis [146]. DFO has achieved results in the investigation of salivary gland and mammary gland injury reconstruction and in increasing vascularization of islet transplantation due to angiogenesis of DFO [147,148]. In a study of fat transplantation, the experimental results showed that DFO-pretreated adipose fat significantly improved the postoperative weight/volume retention rate, suggesting that DFO promoted angiogenesis in the grafts [149].

7. Drug Delivery System

Since DFO is a low-molecular-weight, water-soluble drug with a short retention time in blood vessels, it is necessary to develop a DFO release system to achieve targeted DFO delivery [150].

The penetration of DFO through the intact cuticle is essential for achieving the objective of preventing and treating diabetic ulcers [65,150]. Hence, Dominik Duscher et al. encapsulated DFO with nonionic surfactants and polymers to form reverse micelles, which were then dispersed within a release-controlling polymer matrix patch. This enabled the delivery of DFO through the hydrophobic stratum corneum, ensuring its targeted delivery to the dermis. The experimental results demonstrated the efficacy of the transdermal drug delivery system in preventing diabetic pressure ulcers and promoting the healing process of existing diabetic wounds [65]. In 2019, Dominik Duscher et al. used state-of-the-art surface micro-texturing technology to develop an enhanced TDDS (eTDDS). Micro-textured surfaces ensure that the patch contacts the wound bed and increases drug release. The results showed that the improved transdermal delivery system not only released DFO continuously but also had a stronger skin penetration ability. Compared with other delivery methods (drip-on aqueous solution and degradable polymer spray application), DFO eTDDS accelerated healing [150]. With the continuous progress of drug delivery systems, in addition to the enhancement of targeted drug penetration [65,150,151], the combination of DFO and local drug delivery systems—which can not only provide therapeutic payloads but also promote wound healing—has attracted widespread attention [152,153]. Electrospinning is an advanced method used for developing wound dressings [154–158]. A wound dressing made from electrospun fibers can maintain a moist environment, absorb wound secretions, or provide adequate oxygen [67,159,160]. These types of porous scaffoldings have a high surface-area-to-volume ratio, use a hydrophilic polymer, and can load drugs and other bioagents as active components. Mohammad Hossein Kazemi et al. used the electrospinning technique to produce a fiber mat loaded with DFO and ciprofloxacin which was verified *in vitro* to promote wound healing [161].

In addition, the combination of DFO and hydrogels with a porous structure [162–164] that can mimic the structure and function of extracellular matrix and promote cell migration, proliferation, and maturation provides a new strategy for the treatment of diabetic ulcers [165–167]. For instance, Haijun Shen et al. developed a biomimetic hydrogel containing copper sulfide (CuS) nanoparticles and deferoxamine. DFO and CuS nanoparticles were

incorporated into a biomimetic hydrogel which mimics the structure and function of the extracellular matrix. This biomimetic hydrogel can promote cell adhesion and migration, be degraded by cell-secreted matrix metalloproteinases (MMPs), and then release DFO and CuS nanoparticles at the wound site, where they can exert their therapeutic effects. Meanwhile, it can stimulate angiogenesis, effectively eradicate drug-resistant bacteria, and facilitate cell adhesion and migration, all of which are pivotal factors for the healing of diabetic ulcers [78]. An increasing number of studies have shown that changing the administration of DFO can effectively promote angiogenesis and tissue reconstruction [168–170]. There is also some evidence that continuous release of DFO or prolongation of the half-life of DFO through the design of a stable drug delivery system can promote cell proliferation and migration and stimulate the formation of blood vessels, providing a theoretical basis for the application of DFO in bone repair [110,171–177]. Yahong Li et al. used zeolitic imidazolate framework-8 (ZIF-8), which can promote osteogenesis and bone regeneration [178], as a carrier to extend the half-life of DFO. This not only prolonged the drug release but also achieved a synergistic enhancement effect in promoting H-type vessels, angiogenesis, and osteogenic coupling. This provides a new therapeutic strategy, which has a better effect on bone repair, for the regeneration of bone defects of critical size [178]. In the treatment of BPD, DFO is transported into the alveolar interstitium by respiratory delivery, thereby promoting microvascular reconstruction and, ultimately, inducing lung development. In summary, advanced drug delivery systems provide a promising strategy for achieving targeted therapy and improving therapeutic efficacy. This section focuses on exploring the therapeutic effects of diverse delivery systems for DFO in various diseases, aiming to optimize the efficacy of DFO (Table 1).

Table 1. Different DFO delivery systems.

Delivery System	Composition	Properties	Model Used	Application	Reference
Transdermal drug delivery system	PVP, polymer ethyl cellulose	This approach combines reverse micelle encapsulation of DFO by nonionic surfactants with dispersion in a degradable slow-release matrix, which allows for the targeted delivery of DFO molecules to the dermis.	db/db mice (pressure ulcer model)	Diabetic pressure ulcer	[65]
Injectable hydrogel	SFNs	It can be administered in a locally targeted and minimally invasive manner, and sustained drug release lasts for 40 days.	Diabetic rats (full-thickness wounds)	Diabetic wound healing	[79]
Biomimetic hydrogel	MMP-degradable peptide, HA, RGD	It mimics the structure and function of the extracellular matrix to promote cell adhesion and migration, is degraded by cell-secreted matrix metalloproteinases (MMPs), and subsequently releases the drug at the wound site.	Diabetic mice (full-thickness wounds)	Diabetic wound healing	[78]
Electrospun mat	SF, Ch, PVA	The substance exhibits low toxicity, possesses hemostatic and antimicrobial properties, enables sustained drug release for a duration of 72 h, and facilitates cell adhesion.		Wound healing	[161]
Microneedle patch	HA, Ch, SF	It can destroy biofilms and deliver drugs at a deeper level.	Diabetic rat (full-thickness wounds)	Wound healing	[77]

Table 1. Cont.

Delivery System	Composition	Properties	Model Used	Application	Reference
Electrospun artificial Periosteum	PCL	It can support cell attachment, proliferation, and migration by mimicking the shape and structure of the extracellular matrix. It can be continuously and slowly released for more than 21 days.		Osteogenesis	[111]
Biomimetically hierarchical scaffold	MnCO nanosheets, gelatin-methacryloyl hydrogel, polylactide/HA matrix	With a well-organized gradient structure, it mimics the cortical and cancellous bone tissues; meanwhile, the hydrogels inside the scaffolds provide the scaffolds with additional extracellular matrix characteristics.	Rat femur defect model	Bone regeneration	[171]
Drug-delivery nanoplatform	ZIF-8	The excellent biocompatibility, high porosity, and adjustable pore size of ZIF-8 make it a suitable carrier for encapsulating DFO to extend the half-life of DFO. Moreover, ZIF-8 itself can promote osteogenesis and bone regeneration.	Cranial defect models of rats	Bone regeneration	[178]
Injectable temperature-sensitive hydrogel	GMs, type I, collagen, fibronectin	GMs possess long-term release characteristics of DFO, hydrogel that allows the material to automatically adapt to the three-dimensional structure of the defect site, and components similar to the extracellular matrix that promote repair-related cells.	Rat femur critical bone defect model	Bone regeneration	[110]
Aerosol particles	lactic-co-glycolic acid, membranes of macrophages	Its optimized size and the shell-core structure endow aerosol particles with Brownian motion and atomization stability, thus enabling the aerosol particles to reach the bronchi and alveoli deeply for effective deposition.	C57BL/6 mice (oxygen-induced BPD model)	Alveolar reconstruction and lung development	[129]

SF: silk fibroin; Ch: chitosan; PVA: polyvinyl alcohol; SFNs: SF nanofibers; HA: hyaluronic acid; MMP: matrix metalloproteinases; PCL: polycaprolactone; MnCO: manganese carbonyl; ZIF-8: zeolitic imidazolate framework-8; PVP: polymer polyvinylpyrrolidone; GMs: gelatin microspheres.

8. Conclusions and Future Perspectives

Deferoxamine can not only be used as an iron chelator to treat iron overload diseases but can also play an indispensable role in the treatment of angiogenesis deficiency diseases. The intrinsic mechanism of deferoxamine in promoting the therapeutic effect of angiogenesis is closely related to the hypoxia-inducible factor-1 α signaling pathway. However, deferoxamine is a drug with a short half-life, small molecular weight, and good water solubility, which limits the durable effect of deferoxamine in angiogenesis. Therefore, in treatment to repair skin and tissue (such as diabetic wounds and burn wounds), researchers coated deferoxamine in various hydrogels or patches and applied them to the repair site. The results showed that the targeted release of deferoxamine can promote endothelial cell proliferation, migration, and angiogenesis, thereby promoting wound healing. In bone regeneration treatment, the combination of biomaterials and deferoxamine can prolong drug release while reducing cytotoxicity. This combination also enriches the function of scaffold materials and plays a role in tissue repair and regeneration, immune regulation, optimization of angiogenesis, and promotion of bone tissue regeneration. In addition, an increasing number of studies have shown that deferoxamine has toxic effects in wound

healing, such as visual toxicity and osteotoxicity [179–182]. Therefore, challenges persist regarding how to control the dosage of deferoxamine and improve the mode of administration. Polyelectrolyte capsules have captured our interest in the context of our current research. These polyelectrolyte capsules are believed to be promising drug delivery systems against cancer and are also utilized in self-healing coatings [183]. They are currently undergoing mass production using automated systems at the first stage. These capsules also constitute a promising platform for deferoxamine drug delivery. In addition, the poly (lactic acid) (PLA) microchamber array (MCA) is a biodegradable and biocompatible controlled drug-release system sensitive to the high-intensity focused ultrasound. The synthesis of such arrays has minimal impact on the drug, preserving the drug's biological properties. This system can open at therapeutic parameters of ultrasound exposure and complete degradation once the drug is released in full [184]. In conclusion, the combination of different drug delivery systems is expected to maximize the potential advantages of deferoxamine in regenerative medicine treatment.

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References

1. Holden, P.; Nair, L.S. Deferoxamine: An Angiogenic and Antioxidant Molecule for Tissue Regeneration. *Tissue Eng. Part B Rev.* **2019**, *25*, 461–470. [[CrossRef](#)]
2. Ikeda, Y.; Tajima, S.; Yoshida, S.; Yamano, N.; Kihira, Y.; Ishizawa, K.; Aihara, K.; Tomita, S.; Tsuchiya, K.; Tamaki, T. Deferoxamine promotes angiogenesis via the activation of vascular endothelial cell function. *Atherosclerosis* **2011**, *215*, 339–347. [[CrossRef](#)]
3. Kong, L.; Wu, Z.; Zhao, H.; Cui, H.; Shen, J.; Chang, J.; Li, H.; He, Y. Bioactive Injectable Hydrogels Containing Desferrioxamine and Bioglass for Diabetic Wound Healing. *ACS Appl. Mater. Interfaces* **2018**, *10*, 30103–30114. [[CrossRef](#)]
4. Roemhild, K.; von Maltzahn, F.; Weiskirchen, R.; Knüchel, R.; von Stillfried, S.; Lammers, T. Iron metabolism: Pathophysiology and pharmacology. *Trends Pharmacol. Sci.* **2021**, *42*, 640–656. [[CrossRef](#)]
5. Deng, L.; He, S.; Guo, N.; Tian, W.; Zhang, W.; Luo, L. Molecular mechanisms of ferroptosis and relevance to inflammation. *Inflamm. Res.* **2023**, *72*, 281–299. [[CrossRef](#)]
6. González-Domínguez, Á.; Visiedo-García, F.M.; Domínguez-Riscart, J.; González-Domínguez, R.; Mateos, R.M.; Lechuga-Sancho, A.M. Iron Metabolism in Obesity and Metabolic Syndrome. *Int. J. Mol. Sci.* **2020**, *21*, 5529. [[CrossRef](#)]
7. Coradduzza, D.; Congiargiu, A.; Chen, Z.; Zinellu, A.; Carru, C.; Medici, S. Ferroptosis and Senescence: A Systematic Review. *Int. J. Mol. Sci.* **2023**, *24*, 3658. [[CrossRef](#)]
8. Lieu, P.T.; Heiskala, M.; Peterson, P.A.; Yang, Y. The roles of iron in health and disease. *Mol. Asp. Med.* **2001**, *22*, 1–87. [[CrossRef](#)]
9. Ouyang, S.; You, J.; Zhi, C.; Li, P.; Lin, X.; Tan, X.; Ma, W.; Li, L.; Xie, W. Ferroptosis: The potential value target in atherosclerosis. *Cell Death Dis.* **2021**, *12*, 782. [[CrossRef](#)]
10. Hatcher, H.C.; Singh, R.N.; Torti, F.M.; Torti, S.V. Synthetic and natural iron chelators: Therapeutic potential and clinical use. *Future Med. Chem.* **2009**, *1*, 1643–1670. [[CrossRef](#)]
11. Di Paola, A.; Tortora, C.; Argenziano, M.; Marrapodi, M.M.; Rossi, F. Emerging Roles of the Iron Chelators in Inflammation. *Int. J. Mol. Sci.* **2022**, *23*, 7977. [[CrossRef](#)] [[PubMed](#)]
12. Keberle, H. The biochemistry of desferrioxamine and its relation to iron metabolism. *Ann. N. Y. Acad. Sci.* **1964**, *119*, 758–768. [[CrossRef](#)] [[PubMed](#)]

13. Fong, G.H.; Takeda, K. Role and regulation of prolyl hydroxylase domain proteins. *Cell Death Differ.* **2008**, *15*, 635–641. [[CrossRef](#)] [[PubMed](#)]
14. Li, W.; Xiang, Z.; Xing, Y.; Li, S.; Shi, S. Mitochondria bridge HIF signaling and ferroptosis blockage in acute kidney injury. *Cell Death Dis.* **2022**, *13*, 308. [[CrossRef](#)] [[PubMed](#)]
15. Wenger, R.H.; Camenisch, G.; Stiehl, D.P.; Katschinski, D.M. HIF prolyl-4-hydroxylase interacting proteins: Consequences for drug targeting. *Curr. Pharm. Des.* **2009**, *15*, 3886–3894. [[CrossRef](#)] [[PubMed](#)]
16. Rabinowitz, M.H. Inhibition of hypoxia-inducible factor prolyl hydroxylase domain oxygen sensors: Tricking the body into mounting orchestrated survival and repair responses. *J. Med. Chem.* **2013**, *56*, 9369–9402. [[CrossRef](#)] [[PubMed](#)]
17. Shukla, S.D.; Walters, E.H.; Simpson, J.L.; Keely, S.; Wark, P.A.B.; O'Toole, R.F.; Hansbro, P.M. Hypoxia-inducible factor and bacterial infections in chronic obstructive pulmonary disease. *Respirology* **2020**, *25*, 53–63. [[CrossRef](#)] [[PubMed](#)]
18. Li, Y.; Sun, R.; Zou, J.; Ying, Y.; Luo, Z. Dual Roles of the AMP-Activated Protein Kinase Pathway in Angiogenesis. *Cells* **2019**, *8*, 752. [[CrossRef](#)] [[PubMed](#)]
19. Bi, J.; Zhou, W.; Tang, Z. Pathogenesis of diabetic complications: Exploring hypoxic niche formation and HIF-1 α activation. *Biomed. Pharmacother.* **2024**, *172*, 116202. [[CrossRef](#)] [[PubMed](#)]
20. Welsh, S.J.; Koh, M.Y.; Powis, G. The hypoxic inducible stress response as a target for cancer drug discovery. *Semin. Oncol.* **2006**, *33*, 486–497. [[CrossRef](#)]
21. Wouters, A.; Boeckx, C.; Vermorken, J.B.; Van den Weyngaert, D.; Peeters, M.; Lardon, F. The intriguing interplay between therapies targeting the epidermal growth factor receptor, the hypoxic microenvironment and hypoxia-inducible factors. *Curr. Pharm. Des.* **2013**, *19*, 907–917. [[CrossRef](#)] [[PubMed](#)]
22. Zhu, Y.; Chang, B.; Pang, Y.; Wang, H.; Zhou, Y. Advances in Hypoxia-Inducible Factor-1 α Stabilizer Deferoxamine in Tissue Engineering. *Tissue Eng. Part B Rev.* **2023**, *29*, 347–357. [[CrossRef](#)] [[PubMed](#)]
23. Heber-Katz, E. Oxygen, Metabolism, and Regeneration: Lessons from Mice. *Trends Mol. Med.* **2017**, *23*, 1024–1036. [[CrossRef](#)] [[PubMed](#)]
24. Kim, S.Y.; Yang, E.G. Recent Advances in Developing Inhibitors for Hypoxia-Inducible Factor Prolyl Hydroxylases and Their Therapeutic Implications. *Molecules* **2015**, *20*, 20551–20568. [[CrossRef](#)] [[PubMed](#)]
25. Carroll, V.A.; Ashcroft, M. Role of hypoxia-inducible factor (HIF)-1 α versus HIF-2 α in the regulation of HIF target genes in response to hypoxia, insulin-like growth factor-I, or loss of von Hippel-Lindau function: Implications for targeting the HIF pathway. *Cancer Res.* **2006**, *66*, 6264–6270. [[CrossRef](#)] [[PubMed](#)]
26. Semenza, G.L. Pharmacologic Targeting of Hypoxia-Inducible Factors. *Annu. Rev. Pharmacol. Toxicol.* **2019**, *59*, 379–403. [[CrossRef](#)] [[PubMed](#)]
27. Weinreb, O.; Mandel, S.; Youdim, M.B.H.; Amit, T. Targeting dysregulation of brain iron homeostasis in Parkinson's disease by iron chelators. *Free Radic. Biol. Med.* **2013**, *62*, 52–64. [[CrossRef](#)]
28. Zanolini, F.; Zanolini, I.; Trentini, M.; Tiengo, E.; Pusceddu, T.; Licastro, D.; Degasperi, M.; Leo, S.; Tremoli, E.; Ferroni, L.; et al. Mitochondrial Metabolism and EV Cargo of Endothelial Cells Is Affected in Presence of EVs Derived from MSCs on Which HIF Is Activated. *Int. J. Mol. Sci.* **2023**, *24*, 6002. [[CrossRef](#)] [[PubMed](#)]
29. Zhu, D.; Wei, W.; Zhang, J.; Zhao, B.; Li, Q.; Jin, P. Mechanism of damage of HIF-1 signaling in chronic diabetic foot ulcers and its related therapeutic perspectives. *Heliyon* **2024**, *10*, e24656. [[CrossRef](#)]
30. Li, G.; Ko, C.N.; Li, D.; Yang, C.; Wang, W.; Yang, G.J.; Di Primo, C.; Wong, V.K.W.; Xiang, Y.; Lin, L.; et al. A small molecule HIF-1 α stabilizer that accelerates diabetic wound healing. *Nat. Commun.* **2021**, *12*, 3363. [[CrossRef](#)]
31. Harris, A.L. Hypoxia—a key regulatory factor in tumour growth. *Nat. Rev. Cancer* **2002**, *2*, 38–47. [[CrossRef](#)] [[PubMed](#)]
32. Yang, C.; Zhong, Z.F.; Wang, S.P.; Vong, C.T.; Yu, B.; Wang, Y.T. HIF-1: Structure, biology and natural modulators. *Chin. J. Nat. Med.* **2021**, *19*, 521–527. [[CrossRef](#)] [[PubMed](#)]
33. Catrina, S.B.; Zheng, X. Disturbed hypoxic responses as a pathogenic mechanism of diabetic foot ulcers. *Diabetes Metab. Res. Rev.* **2016**, *32* (Suppl. S1), 179–185. [[CrossRef](#)] [[PubMed](#)]
34. Wu, Y.; Li, X.; Xie, W.; Jankovic, J.; Le, W.; Pan, T. Neuroprotection of deferoxamine on rotenone-induced injury via accumulation of HIF-1 α and induction of autophagy in SH-SY5Y cells. *Neurochem. Int.* **2010**, *57*, 198–205. [[CrossRef](#)]
35. Martínez-Romero, R.; Martínez-Lara, E.; Aguilar-Quesada, R.; Peralta, A.; Oliver, F.J.; Siles, E. PARP-1 modulates deferoxamine-induced HIF-1 α accumulation through the regulation of nitric oxide and oxidative stress. *J. Cell Biochem.* **2008**, *104*, 2248–2260. [[CrossRef](#)] [[PubMed](#)]
36. Hou, Z.; Nie, C.; Si, Z.; Ma, Y. Deferoxamine enhances neovascularization and accelerates wound healing in diabetic rats via the accumulation of hypoxia-inducible factor-1 α . *Diabetes Res. Clin. Pract.* **2013**, *101*, 62–71. [[CrossRef](#)] [[PubMed](#)]
37. Street, J.; Bao, M.; deGuzman, L.; Bunting, S.; Peale, F.V., Jr.; Ferrara, N.; Steinmetz, H.; Hoeffel, J.; Cleland, J.L.; Daugherty, A.; et al. Vascular endothelial growth factor stimulates bone repair by promoting angiogenesis and bone turnover. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 9656–9661. [[CrossRef](#)] [[PubMed](#)]
38. Marziano, C.; Genet, G.; Hirschi, K.K. Vascular endothelial cell specification in health and disease. *Angiogenesis* **2021**, *24*, 213–236. [[CrossRef](#)]
39. Chi, Z.; Chen, L.; Ye, X.; Liu, A.; Yu, G.; Sun, Y. The vasculature niches required for hematopoiesis. *J. Mol. Med.* **2022**, *100*, 53–61. [[CrossRef](#)]

40. Blanchard, L.; Girard, J.P. High endothelial venules (HEVs) in immunity, inflammation and cancer. *Angiogenesis* **2021**, *24*, 719–753. [[CrossRef](#)]
41. Fujioka, T.; Kaneko, N.; Sawamoto, K. Blood vessels as a scaffold for neuronal migration. *Neurochem. Int.* **2019**, *126*, 69–73. [[CrossRef](#)] [[PubMed](#)]
42. Liu, X.; Zhang, P.; Gu, Y.; Guo, Q.; Liu, Y. Type H vessels: Functions in bone development and diseases. *Front. Cell Dev. Biol.* **2023**, *11*, 1236545. [[CrossRef](#)] [[PubMed](#)]
43. Francis, C.R.; Kushner, E.J. Trafficking in blood vessel development. *Angiogenesis* **2022**, *25*, 291–305. [[CrossRef](#)]
44. Hirschi, K.K.; Rohovsky, S.A.; D'Amore, P.A. PDGF, TGF-beta, and heterotypic cell-cell interactions mediate endothelial cell-induced recruitment of 10T1/2 cells and their differentiation to a smooth muscle fate. *J. Cell Biol.* **1998**, *141*, 805–814. [[CrossRef](#)] [[PubMed](#)]
45. Hirschi, K.K.; Rohovsky, S.A.; Beck, L.H.; Smith, S.R.; D'Amore, P.A. Endothelial cells modulate the proliferation of mural cell precursors via platelet-derived growth factor-BB and heterotypic cell contact. *Circ. Res.* **1999**, *84*, 298–305. [[CrossRef](#)] [[PubMed](#)]
46. Naito, H.; Iba, T.; Takakura, N. Mechanisms of new blood-vessel formation and proliferative heterogeneity of endothelial cells. *Int. Immunol.* **2020**, *32*, 295–305. [[CrossRef](#)] [[PubMed](#)]
47. Eelen, G.; Treppe, L.; Li, X.; Carmeliet, P. Basic and Therapeutic Aspects of Angiogenesis Updated. *Circ. Res.* **2020**, *127*, 310–329. [[CrossRef](#)] [[PubMed](#)]
48. Casie Chetty, S.; Rost, M.S.; Enriquez, J.R.; Schumacher, J.A.; Baltrunaite, K.; Rossi, A.; Stainier, D.Y.; Sumanas, S. Vegf signaling promotes vascular endothelial differentiation by modulating etv2 expression. *Dev. Biol.* **2017**, *424*, 147–161. [[CrossRef](#)] [[PubMed](#)]
49. Rodrigues, M.; Kosaric, N.; Bonham, C.A.; Gurtner, G.C. Wound Healing: A Cellular Perspective. *Physiol. Rev.* **2019**, *99*, 665–706. [[CrossRef](#)]
50. Veith, A.P.; Henderson, K.; Spencer, A.; Sligar, A.D.; Baker, A.B. Therapeutic strategies for enhancing angiogenesis in wound healing. *Adv. Drug Deliv. Rev.* **2019**, *146*, 97–125. [[CrossRef](#)]
51. Shaabani, E.; Sharifiaghdam, M.; Faridi-Majidi, R.; De Smedt, S.C.; Braeckmans, K.; Fraire, J.C. Gene therapy to enhance angiogenesis in chronic wounds. *Mol. Ther. Nucleic Acids* **2022**, *29*, 871–899. [[CrossRef](#)]
52. De Wolde, S.D.; Hulskes, R.H.; Weenink, R.P.; Hollmann, M.W.; Van Hulst, R.A. The Effects of Hyperbaric Oxygenation on Oxidative Stress, Inflammation and Angiogenesis. *Biomolecules* **2021**, *11*, 1210. [[CrossRef](#)]
53. Rai, V.; Moellmer, R.; Agrawal, D.K. Stem Cells and Angiogenesis: Implications and Limitations in Enhancing Chronic Diabetic Foot Ulcer Healing. *Cells* **2022**, *11*, 2287. [[CrossRef](#)] [[PubMed](#)]
54. Cucci, L.M.; Satriano, C.; Marzo, T.; La Mendola, D. Angiogenin and Copper Crossing in Wound Healing. *Int. J. Mol. Sci.* **2021**, *22*, 10704. [[CrossRef](#)]
55. Qin, Q.; Lee, S.; Patel, N.; Walden, K.; Gomez-Salazar, M.; Levi, B.; James, A.W. Neurovascular coupling in bone regeneration. *Exp. Mol. Med.* **2022**, *54*, 1844–1849. [[CrossRef](#)] [[PubMed](#)]
56. Peng, Y.; Wu, S.; Li, Y.; Crane, J.L. Type H blood vessels in bone modeling and remodeling. *Theranostics* **2020**, *10*, 426–436. [[CrossRef](#)]
57. Liu, J.; Yang, L.; Liu, K.; Gao, F. Hydrogel scaffolds in bone regeneration: Their promising roles in angiogenesis. *Front. Pharmacol.* **2023**, *14*, 1050954. [[CrossRef](#)] [[PubMed](#)]
58. Tuckermann, J.; Adams, R.H. The endothelium-bone axis in development, homeostasis and bone and joint disease. *Nat. Rev. Rheumatol.* **2021**, *17*, 608–620. [[CrossRef](#)]
59. Bowers, S.; Franco, E. Chronic Wounds: Evaluation and Management. *Am. Fam. Physician* **2020**, *101*, 159–166.
60. Frykberg, R.G.; Banks, J. Challenges in the Treatment of Chronic Wounds. *Adv. Wound Care* **2015**, *4*, 560–582. [[CrossRef](#)]
61. Zhao, R.; Liang, H.; Clarke, E.; Jackson, C.; Xue, M. Inflammation in Chronic Wounds. *Int. J. Mol. Sci.* **2016**, *17*, 2085. [[CrossRef](#)] [[PubMed](#)]
62. Sharma, A.; Sharma, D.; Zhao, F. Updates on Recent Clinical Assessment of Commercial Chronic Wound Care Products. *Adv. Healthc. Mater.* **2023**, *12*, 2300556. [[CrossRef](#)] [[PubMed](#)]
63. Huang, F.; Lu, X.; Yang, Y.; Yang, Y.; Li, Y.; Kuai, L.; Li, B.; Dong, H.; Shi, J. Microenvironment-Based Diabetic Foot Ulcer Nanomedicine. *Adv. Sci.* **2023**, *10*, e2203308. [[CrossRef](#)] [[PubMed](#)]
64. Zhang, R.; Jiang, G.; Gao, Q.; Wang, X.; Wang, Y.; Xu, X.; Yan, W.; Shen, H. Sprayed copper peroxide nanodots for accelerating wound healing in a multidrug-resistant bacteria infected diabetic ulcer. *Nanoscale* **2021**, *13*, 15937–15951. [[CrossRef](#)] [[PubMed](#)]
65. Duscher, D.; Neofytou, E.; Wong, V.W.; Maan, Z.N.; Rennert, R.C.; Inayathullah, M.; Januszyk, M.; Rodrigues, M.; Malkovskiy, A.V.; Whitmore, A.J.; et al. Transdermal deferoxamine prevents pressure-induced diabetic ulcers. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, 94–99. [[CrossRef](#)] [[PubMed](#)]
66. Chen, H.; Cheng, Y.; Tian, J.; Yang, P.; Zhang, X.; Chen, Y.; Hu, Y.; Wu, J. Dissolved oxygen from microalgae-gel patch promotes chronic wound healing in diabetes. *Sci. Adv.* **2020**, *6*, eaba4311. [[CrossRef](#)]
67. Choi, J.S.; Kim, H.S.; Yoo, H.S. Electrospinning strategies of drug-incorporated nanofibrous mats for wound recovery. *Drug Deliv. Transl. Res.* **2015**, *5*, 137–145. [[CrossRef](#)] [[PubMed](#)]
68. Tchanque-Fossuo, C.N.; Dahle, S.E.; Buchman, S.R.; Isseroff, R.R. Deferoxamine: Potential novel topical therapeutic for chronic wounds. *Br. J. Dermatol.* **2017**, *176*, 1056–1059. [[CrossRef](#)] [[PubMed](#)]

69. Thangarajah, H.; Yao, D.; Chang, E.I.; Shi, Y.; Jazayeri, L.; Vial, I.N.; Galiano, R.D.; Du, X.L.; Grogan, R.; Galvez, M.G.; et al. The molecular basis for impaired hypoxia-induced VEGF expression in diabetic tissues. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 13505–13510. [[CrossRef](#)]
70. Zhang, Y.; Zhu, Y.; Ma, P.; Wu, H.; Xiao, D.; Zhang, Y.; Sui, X.; Zhang, L.; Dong, A. Functional carbohydrate-based hydrogels for diabetic wound therapy. *Carbohydr. Polym.* **2023**, *312*, 120823. [[CrossRef](#)]
71. Wu, H.; Li, F.; Shao, W.; Gao, J.; Ling, D. Promoting Angiogenesis in Oxidative Diabetic Wound Microenvironment Using a Nanozyme-Reinforced Self-Protecting Hydrogel. *ACS Cent. Sci.* **2019**, *5*, 477–485. [[CrossRef](#)]
72. Duscher, D.; Januszyk, M.; Maan, Z.N.; Whittam, A.J.; Hu, M.S.; Walmsley, G.G.; Dong, Y.; Khong, S.M.; Longaker, M.T.; Gurtner, G.C. Comparison of the Hydroxylase Inhibitor Dimethylxalylglycine and the Iron Chelator Deferoxamine in Diabetic and Aged Wound Healing. *Plast. Reconstr. Surg.* **2017**, *139*, 695e–706e. [[CrossRef](#)]
73. Chen, H.; Jia, P.; Kang, H.; Zhang, H.; Liu, Y.; Yang, P.; Yan, Y.; Zuo, G.; Guo, L.; Jiang, M.; et al. Upregulating Hif-1 α by Hydrogel Nanofibrous Scaffolds for Rapidly Recruiting Angiogenesis Relative Cells in Diabetic Wound. *Adv. Healthc. Mater.* **2016**, *5*, 907–918. [[CrossRef](#)]
74. Chen, H.; Guo, L.; Wicks, J.; Ling, C.; Zhao, X.; Yan, Y.; Qi, J.; Cui, W.; Deng, L. Quickly promoting angiogenesis by using a DFO-loaded photo-crosslinked gelatin hydrogel for diabetic skin regeneration. *J. Mater. Chem. B* **2016**, *4*, 3770–3781. [[CrossRef](#)]
75. Gao, S.Q.; Chang, C.; Li, J.J.; Li, Y.; Niu, X.Q.; Zhang, D.P.; Li, L.J.; Gao, J.Q. Co-delivery of deferoxamine and hydroxysafflor yellow A to accelerate diabetic wound healing via enhanced angiogenesis. *Drug Deliv.* **2018**, *25*, 1779–1789. [[CrossRef](#)]
76. Qayoom, A.; Aneesha, V.A.; Anagha, S.; Dar, J.A.; Kumar, P.; Kumar, D. Lecithin-based deferoxamine nanoparticles accelerated cutaneous wound healing in diabetic rats. *Eur. J. Pharmacol.* **2019**, *858*, 172478. [[CrossRef](#)]
77. Gao, S.; Zhang, W.; Zhai, X.; Zhao, X.; Wang, J.; Weng, J.; Li, J.; Chen, X. An antibacterial and proangiogenic double-layer drug-loaded microneedle patch for accelerating diabetic wound healing. *Biomater. Sci.* **2023**, *11*, 533–541. [[CrossRef](#)]
78. Shen, H.; Zhang, C.; Meng, Y.; Qiao, Y.; Ma, Y.; Chen, J.; Wang, X.; Pan, L. Biomimetic Hydrogel Containing Copper Sulfide Nanoparticles and Deferoxamine for Photothermal Therapy of Infected Diabetic Wounds. *Adv. Healthc. Mater.* **2023**, *13*, e2303000. [[CrossRef](#)]
79. Ding, Z.; Zhang, Y.; Guo, P.; Duan, T.; Cheng, W.; Guo, Y.; Zheng, X.; Lu, G.; Lu, Q.; Kaplan, D.L. Injectable Desferrioxamine-Laden Silk Nanofiber Hydrogels for Accelerating Diabetic Wound Healing. *ACS Biomater. Sci. Eng.* **2021**, *7*, 1147–1158. [[CrossRef](#)]
80. Wu, H.; Wang, T.; Liang, Y.; Chen, L.; Li, Z. Self-assembled and dynamic bond crosslinked herb-polysaccharide hydrogel with anti-inflammation and pro-angiogenesis effects for burn wound healing. *Colloids Surf. B Biointerfaces* **2024**, *233*, 113639. [[CrossRef](#)]
81. Shpichka, A.; Butnaru, D.; Bezrukov, E.A.; Sukhanov, R.B.; Atala, A.; Burdukovskii, V.; Zhang, Y.; Timashev, P. Skin tissue regeneration for burn injury. *Stem Cell Res. Ther.* **2019**, *10*, 94. [[CrossRef](#)]
82. El Ayadi, A.; Salsbury, J.R.; Enkhbaatar, P.; Herndon, D.N.; Ansari, N.H. Metal chelation attenuates oxidative stress, inflammation, and vertical burn progression in a porcine brass comb burn model. *Redox Biol.* **2021**, *45*, 102034. [[CrossRef](#)]
83. El Ayadi, A.; Wang, C.Z.; Zhang, M.; Wetzel, M.; Prasai, A.; Finnerty, C.C.; Enkhbaatar, P.; Herndon, D.N.; Ansari, N.H. Metal chelation reduces skin epithelial inflammation and rescues epithelial cells from toxicity due to thermal injury in a rat model. *Burn. Trauma* **2020**, *8*, tkaa024. [[CrossRef](#)]
84. Wang, C.Z.; Ayadi, A.E.; Goswamy, J.; Finnerty, C.C.; Mifflin, R.; Sousse, L.; Enkhbaatar, P.; Papaconstantinou, J.; Herndon, D.N.; Ansari, N.H. Topically applied metal chelator reduces thermal injury progression in a rat model of brass comb burn. *Burns* **2015**, *41*, 1775–1787. [[CrossRef](#)]
85. Welch, K.D.; Davis, T.Z.; Van Eden, M.E.; Aust, S.D. Deleterious iron-mediated oxidation of biomolecules. *Free Radic. Biol. Med.* **2002**, *32*, 577–583. [[CrossRef](#)]
86. Beaufay, F.; Quarles, E.; Franz, A.; Katamanin, O.; Wholey, W.Y.; Jakob, U. Polyphosphate Functions In Vivo as an Iron Chelator and Fenton Reaction Inhibitor. *mBio* **2020**, *11*, e01017-20. [[CrossRef](#)]
87. Żwierzeło, W.; Styburski, D.; Maruszewska, A.; Piorun, K.; Skórka-Majewicz, M.; Czerwińska, M.; Maciejewska, D.; Baranowska-Bosiacka, I.; Krajewski, A.; Gutowska, I. Bioelements in the treatment of burn injuries—The complex review of metabolism and supplementation (copper, selenium, zinc, iron, manganese, chromium and magnesium). *J. Trace Elem. Med. Biol.* **2020**, *62*, 126616. [[CrossRef](#)]
88. Nguyen, V.T.; Nassar, D.; Batteux, F.; Raymond, K.; Tharaux, P.L.; Aractingi, S. Delayed Healing of Sickle Cell Ulcers Is due to Impaired Angiogenesis and CXCL12 Secretion in Skin Wounds. *J. Investig. Dermatol.* **2016**, *136*, 497–506. [[CrossRef](#)]
89. Vichinsky, E.; Onyekwere, O.; Porter, J.; Swerdlow, P.; Eckman, J.; Lane, P.; Files, B.; Hassell, K.; Kelly, P.; Wilson, F.; et al. A randomised comparison of deferasirox versus deferoxamine for the treatment of transfusional iron overload in sickle cell disease. *Br. J. Haematol.* **2007**, *136*, 501–508. [[CrossRef](#)]
90. Franchini, M.; Gandini, G.; Veneri, D.; Aprili, G. Safety and efficacy of subcutaneous bolus injection of deferoxamine in adult patients with iron overload: An update. *Blood* **2004**, *103*, 747–748. [[CrossRef](#)]
91. Rodrigues, M.; Bonham, C.A.; Minniti, C.P.; Gupta, K.; Longaker, M.T.; Gurtner, G.C. Iron Chelation with Transdermal Deferoxamine Accelerates Healing of Murine Sickle Cell Ulcers. *Adv. Wound Care* **2018**, *7*, 323–332. [[CrossRef](#)]
92. Wan, C.; Gilbert, S.R.; Wang, Y.; Cao, X.; Shen, X.; Ramaswamy, G.; Jacobsen, K.A.; Alaql, Z.S.; Eberhardt, A.W.; Gerstenfeld, L.C.; et al. Activation of the hypoxia-inducible factor-1 α pathway accelerates bone regeneration. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 686–691. [[CrossRef](#)]

93. Bosch-Ru  ,  .; D  ez-Tercero, L.; Buitrago, J.O.; Castro, E.; P  rez, R.A. Angiogenic and immunomodulation role of ions for initial stages of bone tissue regeneration. *Acta Biomater.* **2023**, *166*, 14–41. [[CrossRef](#)]
94. Ribeiro, T.P.; Flores, M.; Madureira, S.; Zanutto, F.; Monteiro, F.J.; Laranjeira, M.S. Magnetic Bone Tissue Engineering: Reviewing the Effects of Magnetic Stimulation on Bone Regeneration and Angiogenesis. *Pharmaceutics* **2023**, *15*, 1045. [[CrossRef](#)]
95. Longoni, A.; Li, J.; Lindberg, G.C.J.; Rnjak-Kovacina, J.; Wise, L.M.; Hooper, G.J.; Woodfield, T.B.F.; Kieser, D.C.; Lim, K.S. Strategies for inclusion of growth factors into 3D printed bone grafts. *Essays Biochem.* **2021**, *65*, 569–585. [[CrossRef](#)]
96. Simunovic, F.; Finkenzeller, G. Vascularization Strategies in Bone Tissue Engineering. *Cells* **2021**, *10*, 1749. [[CrossRef](#)]
97. Saul, D.; Khosla, S. Fracture Healing in the Setting of Endocrine Diseases, Aging, and Cellular Senescence. *Endocr. Rev.* **2022**, *43*, 984–1002. [[CrossRef](#)]
98. Rather, H.A.; Jhala, D.; Vasita, R. Dual functional approaches for osteogenesis coupled angiogenesis in bone tissue engineering. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2019**, *103*, 109761. [[CrossRef](#)]
99. Wagner, D.R.; Karnik, S.; Gunderson, Z.J.; Nielsen, J.J.; Fennimore, A.; Promer, H.J.; Lowery, J.W.; Loghmani, M.T.; Low, P.S.; McKinley, T.O.; et al. Dysfunctional stem and progenitor cells impair fracture healing with age. *World J. Stem Cells* **2019**, *11*, 281–296. [[CrossRef](#)]
100. Irfan, D.; Ahmad, I.; Patra, I.; Margiana, R.; Rasulova, M.T.; Sivaraman, R.; Kandeel, M.; Mohammad, H.J.; Al-Qaim, Z.H.; Jawad, M.A.; et al. Stem cell-derived exosomes in bone healing: Focusing on their role in angiogenesis. *Cytotherapy* **2023**, *25*, 353–361. [[CrossRef](#)]
101. Stegen, S.; van Gestel, N.; Carmeliet, G. Bringing new life to damaged bone: The importance of angiogenesis in bone repair and regeneration. *Bone* **2015**, *70*, 19–27. [[CrossRef](#)]
102. Grcevic, D.; Pejda, S.; Matthews, B.G.; Repic, D.; Wang, L.; Li, H.; Kronenberg, M.S.; Jiang, X.; Maye, P.; Adams, D.J.; et al. In Vivo Fate Mapping Identifies Mesenchymal Progenitor Cells. *Stem Cells* **2012**, *30*, 187–196. [[CrossRef](#)]
103. Colnot, C. Cellular and molecular interactions regulating skeletogenesis. *J. Cell Biochem.* **2005**, *95*, 688–697. [[CrossRef](#)]
104. Shen, X.; Wan, C.; Ramaswamy, G.; Mavalli, M.; Wang, Y.; Duvall, C.L.; Deng, L.F.; Guldberg, R.E.; Eberhart, A.; Clemens, T.L.; et al. Prolyl hydroxylase inhibitors increase neoangiogenesis and callus formation following femur fracture in mice. *J. Orthop. Res.* **2009**, *27*, 1298–1305. [[CrossRef](#)]
105. Warnecke, C.; Griethe, W.; Weidemann, A.; J  rgensen, J.S.; Willam, C.; Bachmann, S.; Ivashchenko, Y.; Wagner, I.; Frei, U.; Wiesener, M.; et al. Activation of the hypoxia-inducible factor-pathway and stimulation of angiogenesis by application of prolyl hydroxylase inhibitors. *Faseb J.* **2003**, *17*, 1186–1188. [[CrossRef](#)]
106. Wang, G.L.; Semenza, G.L. Desferrioxamine induces erythropoietin gene expression and hypoxia-inducible factor 1 DNA-binding activity: Implications for models of hypoxia signal transduction. *Blood* **1993**, *82*, 3610–3615. [[CrossRef](#)]
107. Stewart, R.; Goldstein, J.; Eberhardt, A.; Chu, G.T.; Gilbert, S. Increasing vascularity to improve healing of a segmental defect of the rat femur. *J. Orthop. Trauma* **2011**, *25*, 472–476. [[CrossRef](#)]
108. Farberg, A.S.; Jing, X.L.; Monson, L.A.; Donneys, A.; Tchanque-Fossuo, C.N.; Deshpande, S.S.; Buchman, S.R. Deferoxamine reverses radiation induced hypovascularity during bone regeneration and repair in the murine mandible. *Bone* **2012**, *50*, 1184–1187. [[CrossRef](#)]
109. Zhang, J.; Tong, D.; Song, H.; Ruan, R.; Sun, Y.; Lin, Y.; Wang, J.; Hou, L.; Dai, J.; Ding, J.; et al. Osteoimmunity-Regulating Biomimetically Hierarchical Scaffold for Augmented Bone Regeneration. *Adv. Mater.* **2022**, *34*, e2202044. [[CrossRef](#)]
110. Zeng, Y.; Huang, C.; Duan, D.; Lou, A.; Guo, Y.; Xiao, T.; Wei, J.; Liu, S.; Wang, Z.; Yang, Q.; et al. Injectable temperature-sensitive hydrogel system incorporating deferoxamine-loaded microspheres promotes H-type blood vessel-related bone repair of a critical size femoral defect. *Acta Biomater.* **2022**, *153*, 108–123. [[CrossRef](#)]
111. Shi, R.; Zhang, J.; Niu, K.; Li, W.; Jiang, N.; Li, J.; Yu, Q.; Wu, C. Electrospun artificial periosteum loaded with DFO contributes to osteogenesis via the TGF-  1/Smad2 pathway. *Biomater. Sci.* **2021**, *9*, 2090–2102. [[CrossRef](#)] [[PubMed](#)]
112. Ai-Aql, Z.S.; Alagl, A.S.; Graves, D.T.; Gerstenfeld, L.C.; Einhorn, T.A. Molecular mechanisms controlling bone formation during fracture healing and distraction osteogenesis. *J. Dent. Res.* **2008**, *87*, 107–118. [[CrossRef](#)] [[PubMed](#)]
113. Cao, X.; Wu, X.; Frassica, D.; Yu, B.; Pang, L.; Xian, L.; Wan, M.; Lei, W.; Armour, M.; Tryggstad, E.; et al. Irradiation induces bone injury by damaging bone marrow microenvironment for stem cells. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 1609–1614. [[CrossRef](#)]
114. Farberg, A.S.; Sarhaddi, D.; Donneys, A.; Deshpande, S.S.; Buchman, S.R. Deferoxamine enhances bone regeneration in mandibular distraction osteogenesis. *Plast. Reconstr. Surg.* **2014**, *133*, 666–671. [[CrossRef](#)] [[PubMed](#)]
115. Donneys, A.; Farberg, A.S.; Tchanque-Fossuo, C.N.; Deshpande, S.S.; Buchman, S.R. Deferoxamine enhances the vascular response of bone regeneration in mandibular distraction osteogenesis. *Plast. Reconstr. Surg.* **2012**, *129*, 850–856. [[CrossRef](#)]
116. Felice, P.A.; Ahsan, S.; Donneys, A.; Deshpande, S.S.; Nelson, N.S.; Buchman, S.R. Deferoxamine administration delivers translational optimization of distraction osteogenesis in the irradiated mandible. *Plast. Reconstr. Surg.* **2013**, *132*, 542e–548e. [[CrossRef](#)] [[PubMed](#)]
117. Donneys, A.; Weiss, D.M.; Deshpande, S.S.; Ahsan, S.; Tchanque-Fossuo, C.N.; Sarhaddi, D.; Levi, B.; Goldstein, S.A.; Buchman, S.R. Localized deferoxamine injection augments vascularity and improves bony union in pathologic fracture healing after radiotherapy. *Bone* **2013**, *52*, 318–325. [[CrossRef](#)] [[PubMed](#)]
118. Li, X.; Jin, L.; Cui, Q.; Wang, G.J.; Balian, G. Steroid effects on osteogenesis through mesenchymal cell gene expression. *Osteoporos. Int.* **2005**, *16*, 101–108. [[CrossRef](#)] [[PubMed](#)]

119. Li, J.; Fan, L.; Yu, Z.; Dang, X.; Wang, K. The effect of deferoxamine on angiogenesis and bone repair in steroid-induced osteonecrosis of rabbit femoral heads. *Exp. Biol. Med.* **2015**, *240*, 273–280. [[CrossRef](#)]
120. Drager, J.; Ramirez-GarciaLuna, J.L.; Kumar, A.; Gbureck, U.; Harvey, E.J.; Barralet, J.E. Hypoxia Biomimicry to Enhance Monetite Bone Defect Repair. *Tissue Eng. Part A* **2017**, *23*, 1372–1381. [[CrossRef](#)]
121. Boccaccini, A.R.; Kneser, U.; Arkudas, A. Scaffolds for vascularized bone regeneration: Advances and challenges. *Expert Rev. Med. Devices* **2012**, *9*, 457–460. [[CrossRef](#)]
122. van Tuyl, M.; Liu, J.; Wang, J.; Kuliszewski, M.; Tibboel, D.; Post, M. Role of oxygen and vascular development in epithelial branching morphogenesis of the developing mouse lung. *Am. J. Physiol. Lung Cell Mol. Physiol.* **2005**, *288*, L167–L178. [[CrossRef](#)]
123. Caniggia, I.; Mostachfi, H.; Winter, J.; Gassmann, M.; Lye, S.J.; Kuliszewski, M.; Post, M. Hypoxia-inducible factor-1 mediates the biological effects of oxygen on human trophoblast differentiation through TGF β ₃. *J. Clin. Investig.* **2000**, *105*, 577–587. [[CrossRef](#)]
124. Chakraborty, D.; Rumi, M.A.; Soares, M.J. NK cells, hypoxia and trophoblast cell differentiation. *Cell Cycle* **2012**, *11*, 2427–2430. [[CrossRef](#)]
125. Asikainen, T.M.; Ahmad, A.; Schneider, B.K.; Ho, W.B.; Arend, M.; Brenner, M.; Günzler, V.; White, C.W. Stimulation of HIF-1 α , HIF-2 α , and VEGF by prolyl 4-hydroxylase inhibition in human lung endothelial and epithelial cells. *Free Radic. Biol. Med.* **2005**, *38*, 1002–1013. [[CrossRef](#)]
126. Asikainen, T.M.; Waleh, N.S.; Schneider, B.K.; Clyman, R.I.; White, C.W. Enhancement of angiogenic effectors through hypoxia-inducible factor in preterm primate lung in vivo. *Am. J. Physiol. Lung Cell Mol. Physiol.* **2006**, *291*, L588–L595. [[CrossRef](#)]
127. Asikainen, T.M.; Chang, L.Y.; Coalson, J.J.; Schneider, B.K.; Waleh, N.S.; Ikegami, M.; Shannon, J.M.; Winter, V.T.; Grubb, P.; Clyman, R.I.; et al. Improved lung growth and function through hypoxia-inducible factor in primate chronic lung disease of prematurity. *Faseb J.* **2006**, *20*, 1698–1700. [[CrossRef](#)]
128. Choi, C.W.; Lee, J.; Lee, H.J.; Park, H.S.; Chun, Y.S.; Kim, B.I. Deferoxamine Improves Alveolar and Pulmonary Vascular Development by Upregulating Hypoxia-inducible Factor-1 α in a Rat Model of Bronchopulmonary Dysplasia. *J. Korean Med. Sci.* **2015**, *30*, 1295–1301. [[CrossRef](#)]
129. Chen, Y.; Chen, W.; Xiang, X.; Deng, L.; Qian, J.; Cui, W.; Chen, H. Pollen-Inspired Shell-Core Aerosol Particles Capable of Brownian Motion for Pulmonary Vascularization. *Adv. Mater.* **2023**, *35*, e2207744. [[CrossRef](#)] [[PubMed](#)]
130. Weigt, S.S.; Wallace, W.D.; Derhovanessian, A.; Saggarr, R.; Saggarr, R.; Lynch, J.P.; Belperio, J.A. Chronic allograft rejection: Epidemiology, diagnosis, pathogenesis, and treatment. *Semin. Respir. Crit. Care Med.* **2010**, *31*, 189–207. [[CrossRef](#)]
131. Jiang, X.; Malkovskiy, A.V.; Tian, W.; Sung, Y.K.; Sun, W.; Hsu, J.L.; Manickam, S.; Wagh, D.; Joubert, L.M.; Semenza, G.L.; et al. Promotion of airway anastomotic microvascular regeneration and alleviation of airway ischemia by deferoxamine nanoparticles. *Biomaterials* **2014**, *35*, 803–813. [[CrossRef](#)] [[PubMed](#)]
132. Koszyca, B.; Manavis, J.; Cornish, R.J.; Blumbergs, P.C. Patterns of immunocytochemical staining for ferritin and transferrin in the human spinal cord following traumatic injury. *J. Clin. Neurosci.* **2002**, *9*, 298–301. [[CrossRef](#)] [[PubMed](#)]
133. Bai, X.Y.; Liu, X.L.; Deng, Z.Z.; Wei, D.M.; Zhang, D.; Xi, H.L.; Wang, Q.Y.; He, M.Z.; Yang, Y.L. Ferroptosis is a new therapeutic target for spinal cord injury. *Front. Neurosci.* **2023**, *17*, 1136143. [[CrossRef](#)] [[PubMed](#)]
134. Dixon, S.J.; Lemberg, K.M.; Lamprecht, M.R.; Skouta, R.; Zaitsev, E.M.; Gleason, C.E.; Patel, D.N.; Bauer, A.J.; Cantley, A.M.; Yang, W.S.; et al. Ferroptosis: An iron-dependent form of nonapoptotic cell death. *Cell* **2012**, *149*, 1060–1072. [[CrossRef](#)] [[PubMed](#)]
135. Yu, J.; Guo, Y.; Sun, M.; Li, B.; Zhang, Y.; Li, C. Iron is a potential key mediator of glutamate excitotoxicity in spinal cord motor neurons. *Brain Res.* **2009**, *1257*, 102–107. [[CrossRef](#)] [[PubMed](#)]
136. Liang, D.; Minikes, A.M.; Jiang, X. Ferroptosis at the intersection of lipid metabolism and cellular signaling. *Mol. Cell* **2022**, *82*, 2215–2227. [[CrossRef](#)]
137. Yao, X.; Zhang, Y.; Hao, J.; Duan, H.Q.; Zhao, C.X.; Sun, C.; Li, B.; Fan, B.Y.; Wang, X.; Li, W.X.; et al. Deferoxamine promotes recovery of traumatic spinal cord injury by inhibiting ferroptosis. *Neural Regen. Res.* **2019**, *14*, 532–541. [[CrossRef](#)]
138. Sinis, N.; Di Scipio, F.; Schönle, P.; Werdin, F.; Kraus, A.; Koopmanns, G.; Masannek, C.; Hermanns, S.; Danker, T.; Guenther, E.; et al. Local administration of DFO-loaded lipid particles improves recovery after end-to-end reconstruction of rat median nerve. *Restor. Neurol. Neurosci.* **2009**, *27*, 651–662. [[CrossRef](#)] [[PubMed](#)]
139. Liang, Y.; Yang, Q.H.; Yu, X.D.; Jiang, D.M. Additive effect of tetramethylpyrazine and deferoxamine in the treatment of spinal cord injury caused by aortic cross-clamping in rats. *Spinal Cord.* **2011**, *49*, 302–306. [[CrossRef](#)] [[PubMed](#)]
140. Tang, G.; Chen, Y.; Chen, J.; Chen, Z.; Jiang, W. Deferoxamine Ameliorates Compressed Spinal Cord Injury by Promoting Neovascularization in Rats. *J. Mol. Neurosci.* **2020**, *70*, 1437–1444. [[CrossRef](#)]
141. Emerit, J.; Beaumont, C.; Trivin, F. Iron metabolism, free radicals, and oxidative injury. *Biomed. Pharmacother.* **2001**, *55*, 333–339. [[CrossRef](#)]
142. Liu, D.; Liu, J.; Sun, D.; Alcock, N.W.; Wen, J. Spinal cord injury increases iron levels: Catalytic production of hydroxyl radicals. *Free Radic. Biol. Med.* **2003**, *34*, 64–71. [[CrossRef](#)] [[PubMed](#)]
143. Si, Q.; Nakamura, Y.; Kataoka, K. A serum factor enhances production of nitric oxide and tumor necrosis factor- α from cultured microglia. *Exp. Neurol.* **2000**, *162*, 89–97. [[CrossRef](#)] [[PubMed](#)]
144. Kroner, A.; Greenhalgh, A.D.; Zarruk, J.G.; Passos Dos Santos, R.; Gaestel, M.; David, S. TNF and increased intracellular iron alter macrophage polarization to a detrimental M1 phenotype in the injured spinal cord. *Neuron* **2014**, *83*, 1098–1116. [[CrossRef](#)] [[PubMed](#)]

145. Hao, J.; Li, B.; Duan, H.Q.; Zhao, C.X.; Zhang, Y.; Sun, C.; Pan, B.; Liu, C.; Kong, X.H.; Yao, X.; et al. Mechanisms underlying the promotion of functional recovery by deferoxamine after spinal cord injury in rats. *Neural Regen. Res.* **2017**, *12*, 959–968. [[CrossRef](#)]
146. Wang, K.; Jing, Y.; Xu, C.; Zhao, J.; Gong, Q.; Chen, S. HIF-1 α and VEGF Are Involved in Deferoxamine-Ameliorated Traumatic Brain Injury. *J. Surg. Res.* **2020**, *246*, 419–426. [[CrossRef](#)] [[PubMed](#)]
147. Lynn, J.V.; Urlaub, K.M.; Ranganathan, K.; Donneys, A.; Nelson, N.S.; Subramanian, C.; Cohen, M.S.; Buchman, S.R. The Role of Deferoxamine in Irradiated Breast Reconstruction: A Study of Oncologic Safety. *Plast. Reconstr. Surg.* **2019**, *143*, 1666–1676. [[CrossRef](#)]
148. Dassoulas, K.R.; Mericli, A.F.; Wang, J.S.; Lei, S.S.; Kim, T.; Cottler, P.S.; Lin, K.Y. Treatment With Topical Deferoxamine Improves Cutaneous Vascularity and Tissue Pliability in an Irradiated Animal Model of Tissue Expander-Based Breast Reconstruction. *Ann. Plast. Surg.* **2019**, *82*, 104–109. [[CrossRef](#)] [[PubMed](#)]
149. Lin, Y.; Zhang, X.; Li, H.; Mu, D. Deferoxamine Mesylate Improves the Survival Rate of Transplanted Fat by Promoting Angiogenesis. *Aesthet. Surg. J.* **2023**, *43*, 789–798. [[CrossRef](#)]
150. Duscher, D.; Trotsyuk, A.A.; Maan, Z.N.; Kwon, S.H.; Rodrigues, M.; Engel, K.; Stern-Buchbinder, Z.A.; Bonham, C.A.; Barrera, J.; Whittam, A.J.; et al. Optimization of transdermal deferoxamine leads to enhanced efficacy in healing skin wounds. *J. Control. Release* **2019**, *308*, 232–239. [[CrossRef](#)]
151. Schuster, L.; Seifert, O.; Vollmer, S.; Kontermann, R.E.; Schlosshauer, B.; Hartmann, H. Immunoliposomes for Targeted Delivery of an Antifibrotic Drug. *Mol. Pharm.* **2015**, *12*, 3146–3157. [[CrossRef](#)] [[PubMed](#)]
152. Madhukiran, D.; Jha, A.; Kumar, M.; Ajmal, G.; Bonde, G.V.; Mishra, B. Electrospun nanofiber-based drug delivery platform: Advances in diabetic foot ulcer management. *Expert Opin. Drug Deliv.* **2021**, *18*, 25–42. [[CrossRef](#)] [[PubMed](#)]
153. Bhardwaj, H.; Khute, S.; Sahu, R.; Jangde, R.K. Advanced Drug Delivery System for Management of Chronic Diabetes Wound Healing. *Curr. Drug Targets* **2023**, *24*, 1239–1259. [[CrossRef](#)]
154. Tan, G.; Wang, L.; Pan, W.; Chen, K. Polysaccharide Electrospun Nanofibers for Wound Healing Applications. *Int. J. Nanomed.* **2022**, *17*, 3913–3931. [[CrossRef](#)] [[PubMed](#)]
155. Juncos Bombin, A.D.; Dunne, N.J.; McCarthy, H.O. Electrospinning of natural polymers for the production of nanofibres for wound healing applications. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2020**, *114*, 110994. [[CrossRef](#)] [[PubMed](#)]
156. Liu, M.; Duan, X.P.; Li, Y.M.; Yang, D.P.; Long, Y.Z. Electrospun nanofibers for wound healing. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2017**, *76*, 1413–1423. [[CrossRef](#)] [[PubMed](#)]
157. Guo, S.; Wang, P.; Song, P.; Li, N. Electrospinning of botanicals for skin wound healing. *Front. Bioeng. Biotechnol.* **2022**, *10*, 1006129. [[CrossRef](#)]
158. Liu, Y.; Li, C.; Feng, Z.; Han, B.; Yu, D.G.; Wang, K. Advances in the Preparation of Nanofiber Dressings by Electrospinning for Promoting Diabetic Wound Healing. *Biomolecules* **2022**, *12*, 1727. [[CrossRef](#)]
159. Akhmetova, A.; Heinz, A. Electrospinning Proteins for Wound Healing Purposes: Opportunities and Challenges. *Pharmaceutics* **2020**, *13*, 4. [[CrossRef](#)]
160. Akombaetwa, N.; Bwanga, A.; Makoni, P.A.; Witika, B.A. Applications of Electrospun Drug-Eluting Nanofibers in Wound Healing: Current and Future Perspectives. *Polymers* **2022**, *14*, 2931. [[CrossRef](#)]
161. Kazemi, M.H.; Sajadimajid, S.; Gorgin Karaji, Z. In vitro investigation of wound healing performance of PVA/chitosan/silk electrospun mat loaded with deferoxamine and ciprofloxacin. *Int. J. Biol. Macromol.* **2023**, *253*, 126602. [[CrossRef](#)]
162. Zhu, J.; Marchant, R.E. Design properties of hydrogel tissue-engineering scaffolds. *Expert Rev. Med. Devices* **2011**, *8*, 607–626. [[CrossRef](#)] [[PubMed](#)]
163. Chyzy, A.; Plonska-Brzezinska, M.E. Hydrogel Properties and Their Impact on Regenerative Medicine and Tissue Engineering. *Molecules* **2020**, *25*, 5795. [[CrossRef](#)] [[PubMed](#)]
164. Zhang, Y.; Chen, H.; Li, J. Recent advances on gelatin methacrylate hydrogels with controlled microstructures for tissue engineering. *Int. J. Biol. Macromol.* **2022**, *221*, 91–107. [[CrossRef](#)] [[PubMed](#)]
165. Khademhosseini, A.; Langer, R. Microengineered hydrogels for tissue engineering. *Biomaterials* **2007**, *28*, 5087–5092. [[CrossRef](#)] [[PubMed](#)]
166. Xiao, S.; Zhao, T.; Wang, J.; Wang, C.; Du, J.; Ying, L.; Lin, J.; Zhang, C.; Hu, W.; Wang, L.; et al. Gelatin Methacrylate (GelMA)-Based Hydrogels for Cell Transplantation: An Effective Strategy for Tissue Engineering. *Stem Cell Rev. Rep.* **2019**, *15*, 664–679. [[CrossRef](#)] [[PubMed](#)]
167. Li, Y.; Kilian, K.A. Bridging the Gap: From 2D Cell Culture to 3D Microengineered Extracellular Matrices. *Adv. Healthc. Mater.* **2015**, *4*, 2780–2796. [[CrossRef](#)] [[PubMed](#)]
168. Vignesh, S.; Sivashanmugam, A.; Annapoorna, M.; Janarthanan, R.; Subramania, I.; Shantikumar, V.N.; Jayakumar, R. Injectable deferoxamine nanoparticles loaded chitosan-hyaluronic acid coacervate hydrogel for therapeutic angiogenesis. *Colloids Surf. B Biointerfaces* **2018**, *161*, 129–138. [[CrossRef](#)] [[PubMed](#)]
169. Shen, A.H.; Borrelli, M.R.; Adem, S.; Deleon, N.M.D.; Patel, R.A.; Mascharak, S.; Yen, S.J.; Sun, B.Y.; Taylor, W.L.t.; Januszyk, M.; et al. Prophylactic treatment with transdermal deferoxamine mitigates radiation-induced skin fibrosis. *Sci. Rep.* **2020**, *10*, 12346. [[CrossRef](#)]
170. Zhang, J.; Luo, Q.; Hu, Q.; Zhang, T.; Shi, J.; Kong, L.; Fu, D.; Yang, C.; Zhang, Z. An injectable bioactive dressing based on platelet-rich plasma and nanoclay: Sustained release of deferoxamine to accelerate chronic wound healing. *Acta Pharm. Sin. B* **2023**, *13*, 4318–4336. [[CrossRef](#)]

171. Yu, F.X.; Lee, P.S.Y.; Yang, L.; Gao, N.; Zhang, Y.; Ljubimov, A.V.; Yang, E.; Zhou, Q.; Xie, L. The impact of sensory neuropathy and inflammation on epithelial wound healing in diabetic corneas. *Prog. Retin. Eye Res.* **2022**, *89*, 101039. [[CrossRef](#)]
172. Schmidt-Bleek, K.; Kwee, B.J.; Mooney, D.J.; Duda, G.N. Boon and Bane of Inflammation in Bone Tissue Regeneration and Its Link with Angiogenesis. *Tissue Eng. Part. B Rev.* **2015**, *21*, 354–364. [[CrossRef](#)] [[PubMed](#)]
173. Chen, F.M.; Zhang, M.; Wu, Z.F. Toward delivery of multiple growth factors in tissue engineering. *Biomaterials* **2010**, *31*, 6279–6308. [[CrossRef](#)] [[PubMed](#)]
174. Donneys, A.; Yang, Q.; Forrest, M.L.; Nelson, N.S.; Zhang, T.; Ettinger, R.; Ranganathan, K.; Snider, A.; Deshpande, S.S.; Cohen, M.S.; et al. Implantable hyaluronic acid-deferoxamine conjugate prevents nonunions through stimulation of neovascularization. *NPJ Regen. Med.* **2019**, *4*, 11. [[CrossRef](#)]
175. Li, H.; Luo, B.; Wen, W.; Zhou, C.; Tian, L.; Ramakrishna, S. Deferoxamine immobilized poly(D,L-lactide) membrane via polydopamine adhesive coating: The influence on mouse embryo osteoblast precursor cells and human umbilical vein endothelial cells. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2017**, *70*, 701–709. [[CrossRef](#)]
176. Ran, Q.; Yu, Y.; Chen, W.; Shen, X.; Mu, C.; Yuan, Z.; Tao, B.; Hu, Y.; Yang, W.; Cai, K. Deferoxamine loaded titania nanotubes substrates regulate osteogenic and angiogenic differentiation of MSCs via activation of HIF-1 α signaling. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2018**, *91*, 44–54. [[CrossRef](#)] [[PubMed](#)]
177. Jia, P.; Chen, H.; Kang, H.; Qi, J.; Zhao, P.; Jiang, M.; Guo, L.; Zhou, Q.; Qian, N.D.; Zhou, H.B.; et al. Deferoxamine released from poly(lactic-co-glycolic acid) promotes healing of osteoporotic bone defect via enhanced angiogenesis and osteogenesis. *J. Biomed. Mater. Res. A* **2016**, *104*, 2515–2527. [[CrossRef](#)] [[PubMed](#)]
178. Li, Y.; Zhu, J.; Zhang, X.; Li, Y.; Zhang, S.; Yang, L.; Li, R.; Wan, Q.; Pei, X.; Chen, J.; et al. Drug-Delivery Nanoplatfrom with Synergistic Regulation of Angiogenesis-Osteogenesis Coupling for Promoting Vascularized Bone Regeneration. *ACS Appl. Mater. Interfaces* **2023**, *15*, 17543–17561. [[CrossRef](#)] [[PubMed](#)]
179. Chan, Y.L.; Chu, C.W.; Chik, K.W.; Pang, L.M.; Shing, M.K.; Li, C.K. Deferoxamine-induced dysplasia of the knee: Sonographic features and diagnostic performance compared with magnetic resonance imaging. *J. Ultrasound Med.* **2001**, *20*, 723–728. [[CrossRef](#)]
180. Brittenham, G.M. Iron-chelating therapy for transfusional iron overload. *N. Engl. J. Med.* **2011**, *364*, 146–156. [[CrossRef](#)]
181. Miller, S.C.; Pan, H.; Wang, D.; Bowman, B.M.; Kopecková, P.; Kopecek, J. Feasibility of using a bone-targeted, macromolecular delivery system coupled with prostaglandin E(1) to promote bone formation in aged, estrogen-deficient rats. *Pharm. Res.* **2008**, *25*, 2889–2895. [[CrossRef](#)] [[PubMed](#)]
182. Wang, D.; Miller, S.C.; Kopecková, P.; Kopecek, J. Bone-targeting macromolecular therapeutics. *Adv. Drug Deliv. Rev.* **2005**, *57*, 1049–1076. [[CrossRef](#)] [[PubMed](#)]
183. Li, W.; Gai, M.; Rutkowski, S.; He, W.; Meng, S.; Gorin, D.; Dai, L.; He, Q.; Frueh, J. An Automated Device for Layer-by-Layer Coating of Dispersed Superparamagnetic Nanoparticle Templates. *Colloid J.* **2018**, *80*, 648–659. [[CrossRef](#)]
184. Sindeeva, O.A.; Gusliakova, O.I.; Inozemtseva, O.A.; Abdurashitov, A.S.; Brodovskaya, E.P.; Gai, M.; Tuchin, V.V.; Gorin, D.A.; Sukhorukov, G.B. Effect of a Controlled Release of Epinephrine Hydrochloride from PLGA Microchamber Array: In Vivo Studies. *ACS Appl. Mater. Interfaces* **2018**, *10*, 37855–37864. [[CrossRef](#)] [[PubMed](#)]

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