

Review

Pharmaceutical Applications of Biomass Polymers: Review of Current Research and Perspectives

Cornelia Bejenaru ^{1,†}, Antonia Radu ¹, Adina-Elena Segneanu ^{2,*}, Andrei Biță ³, Maria Viorica Ciocîlteu ⁴, George Dan Mogoșanu ³, Ionela Amalia Bradu ^{2,†}, Titus Vlase ^{2,5}, Gabriela Vlase ^{2,5} and Ludovic Everard Bejenaru ³

- ¹ Department of Pharmaceutical Botany, Faculty of Pharmacy, University of Medicine and Pharmacy of Craiova, 2 Petru Rareș Street, 200349 Craiova, Dolj, Romania; cornelia.bejenaru@umfcv.ro (C.B.); antonia.radu@umfcv.ro (A.R.)
- ² Institute for Advanced Environmental Research, West University of Timișoara (ICAM–WUT), 4 Oituz Street, 300086 Timișoara, Timiș, Romania; ionela.bradu@e-uvv.ro (I.A.B.); titus.vlase@e-uvv.ro (T.V.); gabriela.vlase@e-uvv.ro (G.V.)
- ³ Department of Pharmacognosy & Phytotherapy, Faculty of Pharmacy, University of Medicine and Pharmacy of Craiova, 2 Petru Rareș Street, 200349 Craiova, Dolj, Romania; andreibita@gmail.com (A.B.); george.mogosanu@umfcv.ro (G.D.M.); ludovic.bejenaru@umfcv.ro (L.E.B.)
- ⁴ Department of Analytical Chemistry, Faculty of Pharmacy, University of Medicine and Pharmacy of Craiova, 2 Petru Rareș Street, 200349 Craiova, Dolj, Romania; maria.ciocilteu@umfcv.ro
- ⁵ Research Center for Thermal Analyzes in Environmental Problems, West University of Timișoara, 16 Johann Heinrich Pestalozzi Street, 300115 Timișoara, Timiș, Romania
- * Correspondence: adina.segneanu@e-uvv.ro
- † These authors contributed equally to this work.

Abstract: Polymers derived from natural biomass have emerged as a valuable resource in the field of biomedicine due to their versatility. Polysaccharides, peptides, proteins, and lignin have demonstrated promising results in various applications, including drug delivery design. However, several challenges need to be addressed to realize the full potential of these polymers. The current paper provides a comprehensive overview of the latest research and perspectives in this area, with a particular focus on developing effective methods and efficient drug delivery systems. This review aims to offer insights into the opportunities and challenges associated with the use of natural polymers in biomedicine and to provide a roadmap for future research in this field.

Keywords: biomass; polymers; biodegradability; biocompatibility; pharmaceutical applications



Citation: Bejenaru, C.; Radu, A.; Segneanu, A.-E.; Biță, A.; Ciocîlteu, M.V.; Mogoșanu, G.D.; Bradu, I.A.; Vlase, T.; Vlase, G.; Bejenaru, L.E. Pharmaceutical Applications of Biomass Polymers: Review of Current Research and Perspectives. *Polymers* **2024**, *16*, 1182. <https://doi.org/10.3390/polym16091182>

Academic Editors: Alberto Romero García, Yadong Tang and Lu Jiang

Received: 8 March 2024

Revised: 15 April 2024

Accepted: 19 April 2024

Published: 23 April 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

The shift toward environmental sustainability has catalyzed a marked transition toward natural biopolymers, moving away from the prevalent use of synthetic polymers across various sectors, including the pharmaceutical industry. Unlike their synthetic counterparts, biopolymers—synthesized through microbial, chemical, or natural processes—offer a promising alternative due to their comparable performance, versatility, and the potential for enhanced functionalities. These functionalities make biopolymers a critical asset in pharmaceutical, environmental, and medical applications, promising to significantly mitigate the issue of plastic pollution. Despite their vast potential, the adoption of biopolymers faces challenges, including high costs and inefficiencies in synthesis and processing, which must be overcome to realize their full potential [1–4].

Biomaterials, designed for direct interaction with biological systems, are at the forefront of medical innovation, enabling groundbreaking medical interventions through bio-compatible materials capable of performing specific functions. The success of these materials, especially in tissue engineering (TE) and drug delivery systems (DDSs), is intricately linked to their physical, chemical, and biological properties, necessitating meticulous

customization to elicit desired responses from host systems. The use of polymers, characterized by their diverse and degradable properties, facilitates the creation of biomaterials that can disintegrate into low-molecular-weight products, either to be re-absorbed or excreted by the body, thereby enhancing their applicability and safety [5–8].

Natural polymers, categorized into proteins, polysaccharides, polyesters, lipids, or lignins (complex aromatic polymers), depending on their chemical structures, are utilized for biomedical purposes through copolymerization, merging polysaccharides like chitosan, starch, and cellulose with other polymers, and employing average protein-based biopolymers such as gelatin, collagen, and albumin for the creation of drug delivery nanomolecules due to their advantageous properties, such as minimal toxicity, narrowness, biodegradability, and prolonged stability [9–11].

Furthermore, biomass polymers' inherent biocompatibility and functional chemical structures are used to develop nanomaterials for a broad range of biomedical applications, ensuring their efficient clearance from the body and eliminating the need for surgical retrieval. Degradable polymers undergo hydrolytic cleavage (enzymatic or nonenzymatic), producing soluble degradation products and enhanced properties like bioavailability, stability, and controlled release for applications in TE (e.g., cartilage scaffolds) and prosthetic implants. Refinement of the physical, chemical, and biological attributes of these polymers is achieved through strategies like blending, cross-linking, and forming interpenetrating polymer networks (IPNs), while recent efforts focus on synthesizing macromolecular biomaterials with optimized thermal and mechanical properties using chemical modification on their nanoconstructs. Innovative techniques, including physicochemical cross-linking, polyion complexes (PICs), layer-by-layer assembly, and nanoparticle (NP) coatings, have paved the way for developing sophisticated multiphase polymer systems tailored for specific biomedical uses [12–15].

Biopolymer composites enhanced with metals, natural fibers, and metal oxides represent a cutting-edge area of research, offering improved adsorptive, mechanical, and thermal properties. These advancements are supported by an array of characterization techniques, underscoring the ongoing efforts to develop cost-effective and performance-optimized biocomposites [16–20].

Natural biopolymers, like cellulose, starch, chitosan, and pectin, and synthetic biomass polymers such as polylactic acid (PLA), polycaprolactone (PCL), and polyglycolic acid (PGA), play crucial roles in various applications, including pharmaceutical preparations, purification membranes, hydrogels, prosthetics, drug delivery, and bone tissue engineering (BTE) (Figure 1).

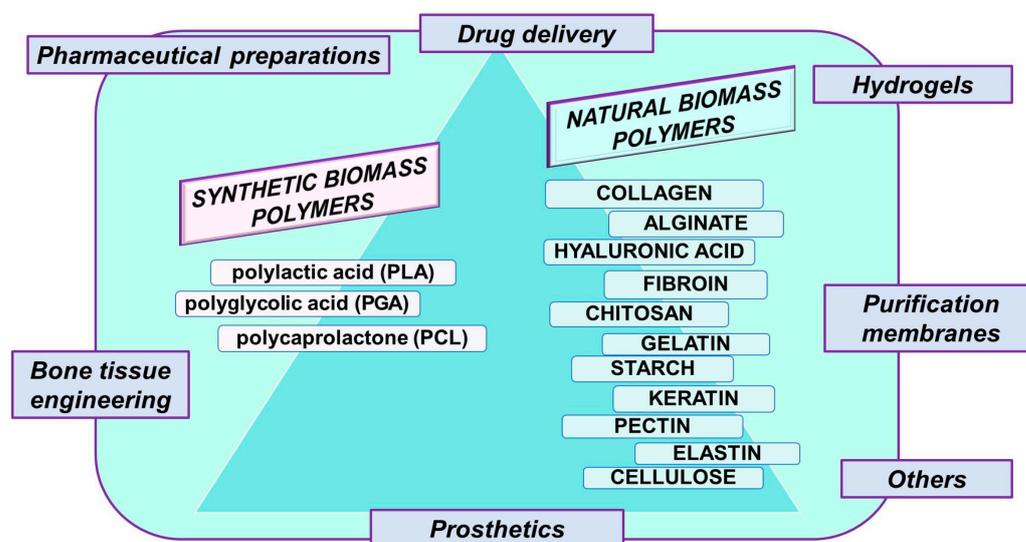


Figure 1. Schematic representation of principal biomass polymers and some of their main applications.

These biopolymers demonstrate unique properties, with most of them being biodegradable, biocompatible, bioactive, or safe, filling the gap left by synthetic polymers. Enhancing the shelf life of food products, edible biopolymer packaging films derived from agricultural waste show nutraceutical, antimicrobial, and antioxidant properties. Furthermore, biopolymer particles generate gel-like structures in emulsion-based products, increasing texture, consistency, and stability [21–24].

2. Biopolymers

Biopolymers, originating from living organisms and composed of monomeric units such as amino acids, saccharides, and nucleic acids, are gaining prominence in medical and pharmaceutical industries, due to their key attributes: eco-friendliness, sustainability, biodegradability, non-toxicity, renewability, and compatibility. Biopolymers derived from natural biomass, exhibiting inherent biodegradability, are directly obtained from the natural environment and possess considerable economic value. Commercially, a myriad of biopolymers serves diverse purposes in biomedical devices, hygiene items, agriculture, and the food industry. Despite their advantages, the fabrication of biopolymers faces a major drawback in substantial financial expenditures, prompting ongoing research into cost-minimization methods, involving purification procedures, high-yielding microorganisms, and substrate selection [1,25,26].

In our study, we specifically focused on biopolymers extracted from biomass, sourced directly from food waste, organic, and other bio-based industries, emphasizing the use of polysaccharides, proteins, and lignin. These biopolymers, derived from natural and renewable sources, hold intrinsic properties that are pivotal for numerous applications within the pharmaceutical sector. Polysaccharides, with their diverse structural features and functionalities; proteins, known for their enzymatic activity and binding capabilities; and lignin, with its complex aromatic structure, offer a broad spectrum of applications ranging from DDSs to TE [27]. The choice of these biopolymers underscores our commitment to exploring sustainable and biocompatible materials that align with the principles of green chemistry and environmental stewardship, while harnessing their natural efficacy for innovative pharmaceutical applications.

Biopolymers are considered economic resources from diverse natural origins including plants, algae, microorganisms, animals, and agricultural residues. Green wastes can come from agricultural plant sources (cotton, tapioca, maize, bananas, cassava, potatoes, wheat, rice, and maize) or wood residues. In contrast, animal biopolymers mainly come from mammals (cattle, pigs) or marine sources (shrimps, lobsters, fish, sponges) [28–31]. Microbial origins (like fungi, algae, and yeasts) and vegetable oils extracted from meadow foam, fish, castor bean, linseed, tung, jojoba, rapeseed, safflower, sunflower, corn, and soybean may serve as rich sources of possible monomers, or co-monomers [32–34].

Biopolymers are categorized based on their biodegradability, origin, thermal response, and composition. The biodegradable and non-biodegradable distinction is evident, along with their classification as derived from natural origins or fossil fuels. The thermal condition response classifies them into thermosets, thermoplastics, and elastomers, while their composition leads to groups such as composites, laminates, and blends. Among the prominent systematization standards is the provenance of raw materials, resulting in natural, synthetic, and microbial biopolymers [1,35–38].

Naturally sourced biomass polymers (Figure 2a–c) encompass proteins (e.g., collagen, soy protein) and polysaccharides (e.g., chitosan, cellulose). Chemically synthesized polymers comprise PLA and petroleum-based polymers, such as polyethylene, PCL, and polyglutamic acid. Biopolymers from microbial generation, like polyhydroxyalkanoates (PHAs), bacterial cellulose, and gellan, serve diverse applications in medical, agro-industrial, and environmental sectors [1,35–38].

In the following sections, starting from recent studies, we selectively characterized various examples of the most relevant biopolymers used in the pharmaceutical domain.

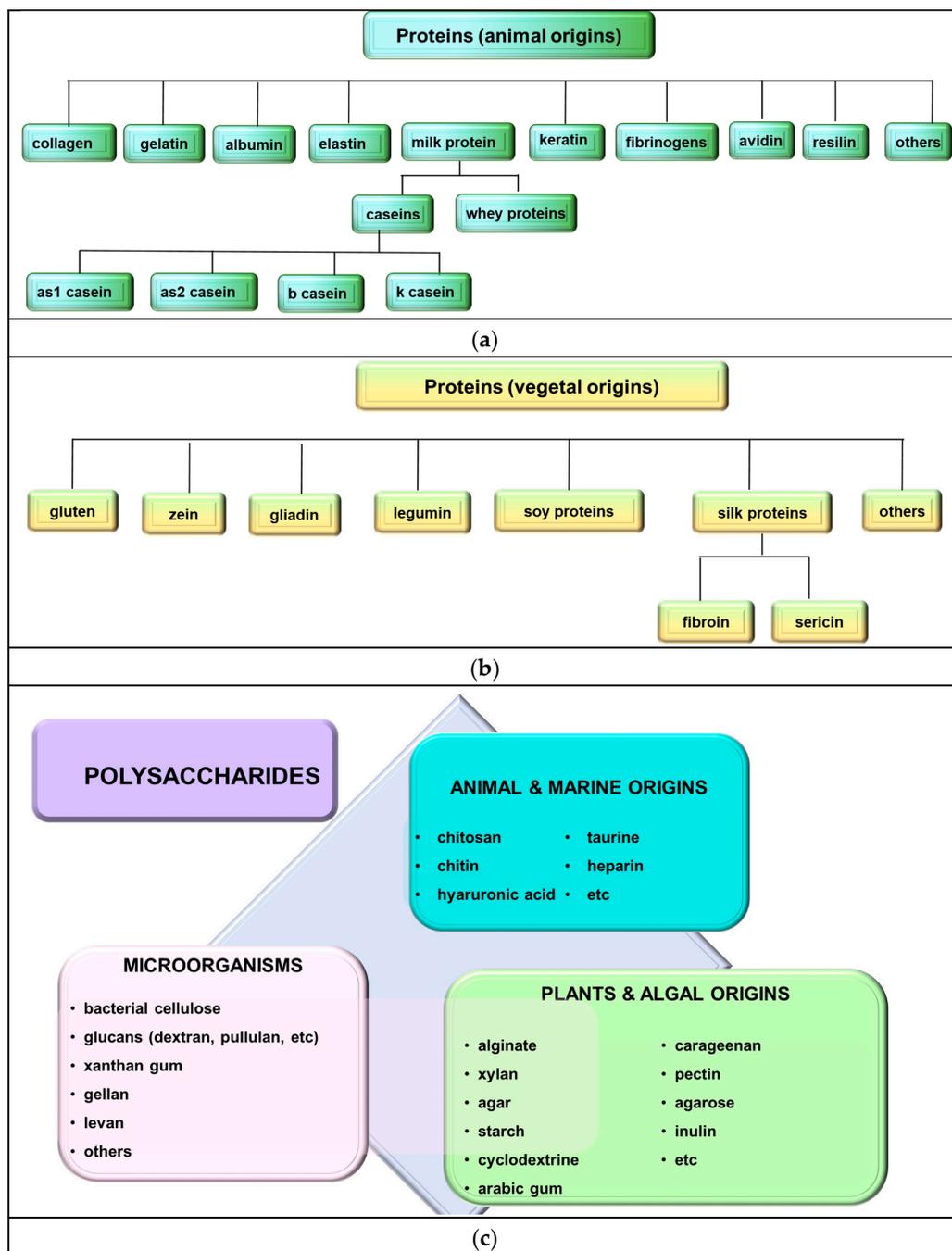


Figure 2. Naturally biomass polymers: proteins (a,b) and polysaccharides (c).

2.1. Polysaccharides

Polysaccharides, natural polymers with functional hydroxyl, amino, and carboxylic groups, have garnered significant attention due to their inert, biocompatible, non-toxic, and cost-effective nature, coupled with excellent water stability. These versatile entities can be easily cross-linked, derivatized, or transformed into multiphase polymer systems such as polyblends, IPNs, graft, and block copolymers, while charged polysaccharides contribute to the formation of valuable PICs [39,40]. This discussion focuses on the key biomedical polysaccharides.

2.1.1. Homoglycans

Starch

Starch is a natural polysaccharide that comprises linear chains of amylose and branched amylopectin segments, primarily derived from α -glucose units and prominently present in cereal grains, fruits, roots, and legumes. Amylose features glucose units linked by α -(1,4) bonds, while amylopectin includes α -(1,4) and α -(1,6) linkages (Figure 3).

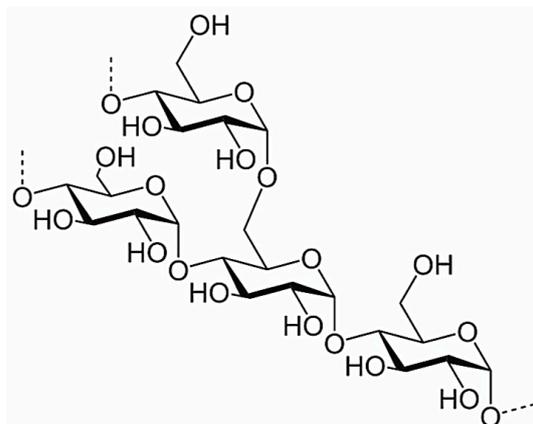


Figure 3. Starch—chemical structure.

Properties of starch vary based on the plant source and its degree of maturity. Chemically altered starch, along with its physical mixtures or IPNs, serve as significant biomaterials in BTE, exhibiting enhanced characteristics. Nevertheless, starch's inherent brittleness, premature degradation before reaching its melting temperature, inferior mechanical features, and challenging processability constrain its application as a standalone material [41–44].

Dextran

This bacterial homopolysaccharide encompasses glucans formed through the polymerization of α -D-glucopyranosyl moieties of sucrose catalyzed by dextransucrase enzyme. The main chain features glucose segments connected by α -(1,6) bonds, while branches exhibit α -(1,4), α -(1,3), and α -(1,2) units (Figure 4).

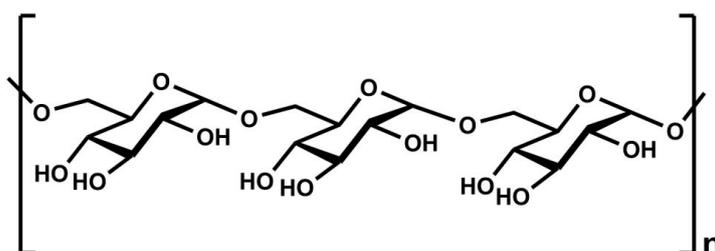


Figure 4. Dextran—chemical structure.

Properties like branching degree, molecular weight, and other attributes vary with the engaged microorganism. Dextran, with its remarkable rheological nature and plasma-volume-enlarging potency, undergoes chemical alterations, introducing thiol, (meth)acrylate, aldehyde, and phenol groups. As a biocompatible polysaccharide with no toxicity, dextran finds extensive applications in pharmaceutical and biomedical domains as an antithrombotic and bio-adhesive agent, in protein/drug delivery or tissue-engineered scaffolds. Injectable hydrogels are proposed as site-specific, trackable chemotherapeutic devices. Despite these advantages, challenges include high costs and limited availability [45–48].

Cyclodextrins

Cyclodextrins are oligosaccharides formed by the enzymatic linkage of glucose units (α -D-glucopyranose) through α -(1,4) bonds, resulting in the production of α -cyclodextrin (six units), β -cyclodextrin (seven units), and γ -cyclodextrin (eight units) (Figure 5a–c).

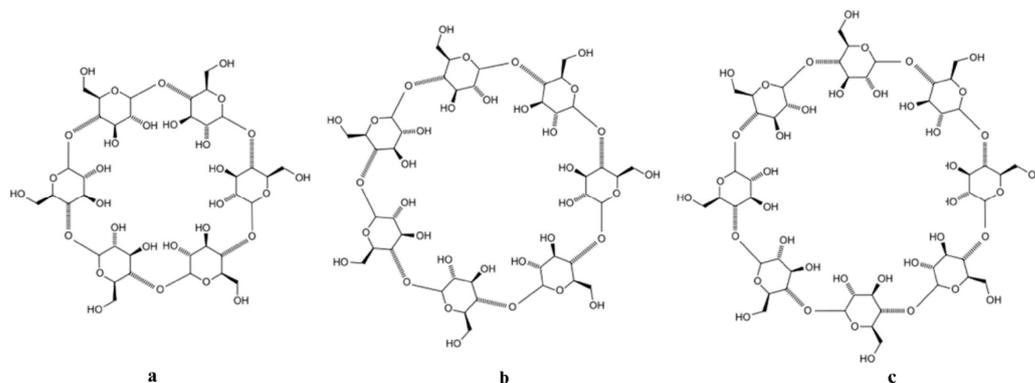


Figure 5. Chemical structure of α -cyclodextrin (a), β -cyclodextrin (b) and γ -cyclodextrin (c).

Characterized by a distinctive truncated cone-like structure, cyclodextrins feature an internal non-polar cavity with polar hydroxyl groups on the surface. This configuration allows hydrophobic substances, including drugs, to be encapsulated through hydrophobic interactions, forming host–guest supramolecular complexes driven by van der Waals and dipole–dipole interactions. The chemistry and applications of cyclodextrins have been subjected to comprehensive scrutiny across various fields. These inclusion complexes are amenable to derivatization and appropriate chemical alterations [49,50].

Cellulose

The most abundant natural polysaccharide, cellulose, consists of β -D-glucopyranose units linked by β -(1,4) glycosidic bonds (Figure 6), offering significant potential as an advanced polymeric material.

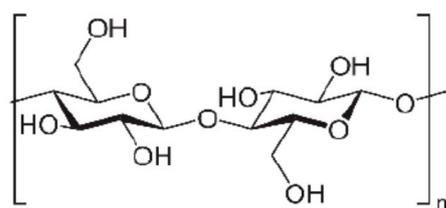


Figure 6. Cellulose—chemical structure.

Cellulose and its derivatives are versatile precursors; cellulose is generally well tolerated by the human body and other living organisms (in particular, cellulose ethers or esters), has low toxicity, and is a cost-effective material. Ionic liquids (ILs) and deep eutectic solvents overcome this issue. The derivatization of cellulose produces environmentally friendly materials, such as methylcellulose, cellulose acetate, hydroxypropyl cellulose, cellulose nitrate, and carboxymethylcellulose. Cellulose nanomaterials, including nanocrystals, bacterial nanocellulose (BNC), and nanofibrils, have been extensively researched. Nevertheless, cellulose has limitations such as low crease resistance, potential antigenicity, and lack of thermoplasticity [51–53].

Chitin and Chitosan

Chitin, a prominent constituent of sea crustacean shells, stands as the second-most abundant biomacromolecule utilized across various industries such as pharmaceuticals, textiles, food, and agriculture. Exhibiting biocompatibility, non-toxicity, biodegradability,

and mucoadhesive properties, chitin can be effortlessly extracted and chemically altered to yield diverse biomaterials (Figure 7a).

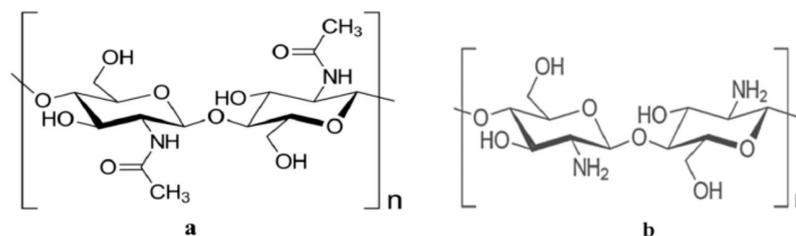


Figure 7. Chemical structure of chitin (a) and chitosan (b).

Chitosan, extracted from crabs and fungal cell walls, undergoes commercial production through the deacetylation of chitin. The degree of deacetylation, impurity composition, and molar mass distribution are based on the natural source and preparation method. As a cationic linear copolymer polysaccharide, chitosan is composed of β-(1→4) connected 2-amino-2-deoxy-D-glucose (D-glucosamine) and 2-acetamido-2-deoxy-D-glucose (N-acetyl-D-glucosamine) segments through glycosidic bonds (Figure 7b). The polymer's primary amino groups confer a positive charge on its surface, promoting inter- and intramolecular hydrogen bonding. Additionally, chitosan exhibits antimicrobial activity against viruses, fungi, and bacteria, rendering it valuable in the biomedical domain. However, its drawback lies in reduced solubility at physiological pH. Despite the variability in synthesis procedures, short-term human testing has shown no signs of allergic reactions [54–58].

2.1.2. Heteroglycans

Alginate

Alginate, a water-soluble anionic polymer comprising α-L-guluronic acid (G) and β-D-mannuronic acid (M) residues connected by 1,4-glycosidic bonds, is biodegradable, biocompatible, and exhibits no toxicity (Figure 8).

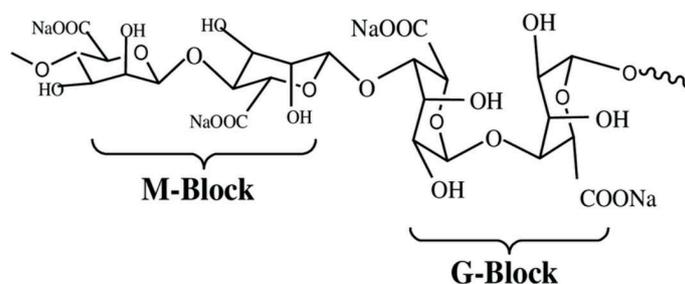


Figure 8. Sodium alginate—chemical structure.

Derived economically from marine brown algae, alginate finds diverse biomedical applications, serving as three-dimensional (3D) scaffolding materials in forms such as foams, microcapsules, sponges, and hydrogels for TE. Physical or chemical alteration enhances alginate's properties, allowing the precise tuning of cell affinity, mechanical strength, and gelation through combinations with other biomaterials, ligand immobilization, and cross-linking. Despite its sensitivity to hydrolysis in acidic environments and challenges in fabrication due to reduced solubility, alginate-based materials have undergone clinical investigations, demonstrating potential benefits such as managing hypertension and advancements in the food industry [59–62].

Agarose

Agarose, an uncharged polysaccharide derived primarily from certain marine red algae, is a key component of agar. It is soluble in hot water, ILs, and polar non-aqueous solvents. From a structural standpoint, agarose is a linear and neutral polysaccharide compris-

ing alternating (1,3)- β -D-galactopyranose and (1,4)-linked 3,6-anhydro- α -L-galactopyranose units (Figure 9).

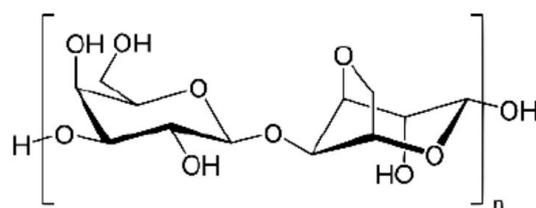


Figure 9. Agarose—chemical structure.

Its solution forms gels upon cooling below ~ 40 °C, with flexible fiber chains capable of curling into helix structures, creating powerful gels with prominent hysteresis. Being non-toxic and biocompatible, agarose is commonly employed as a gelling agent in various applications, including chromatography techniques, nucleic acid electrophoresis, cell culture media, tissue culture overlays, and gel plates. Its exceptional properties, including mechanical resilience and reduced gelling temperature, make it suitable for applications like bio-ink, where gelation forms a 3D network of agarose fibers, disintegrating above 85 °C [63–66].

Carrageenans

Carrageenans, a family of linear sulfated polysaccharides extracted from red algae (*Rhodophyta*), known as Irish moss, exhibit extensive and very flexible molecules capable of forming helical structures, resulting in viscous solutions or elastic gels. Comprising alternate segments of β -D-galactose and 3,6-anhydro- α -D-galactose connected by α -(1,3) and β -(1,4) glycosidic bonds, carrageenans yield three main types—kappa (κ -1 sulfate group/disaccharide), iota (ι -2 sulfate groups/disaccharide), and lambda (λ -3 sulfate groups/disaccharide)—depending on the extraction method and algae source (Figure 10).

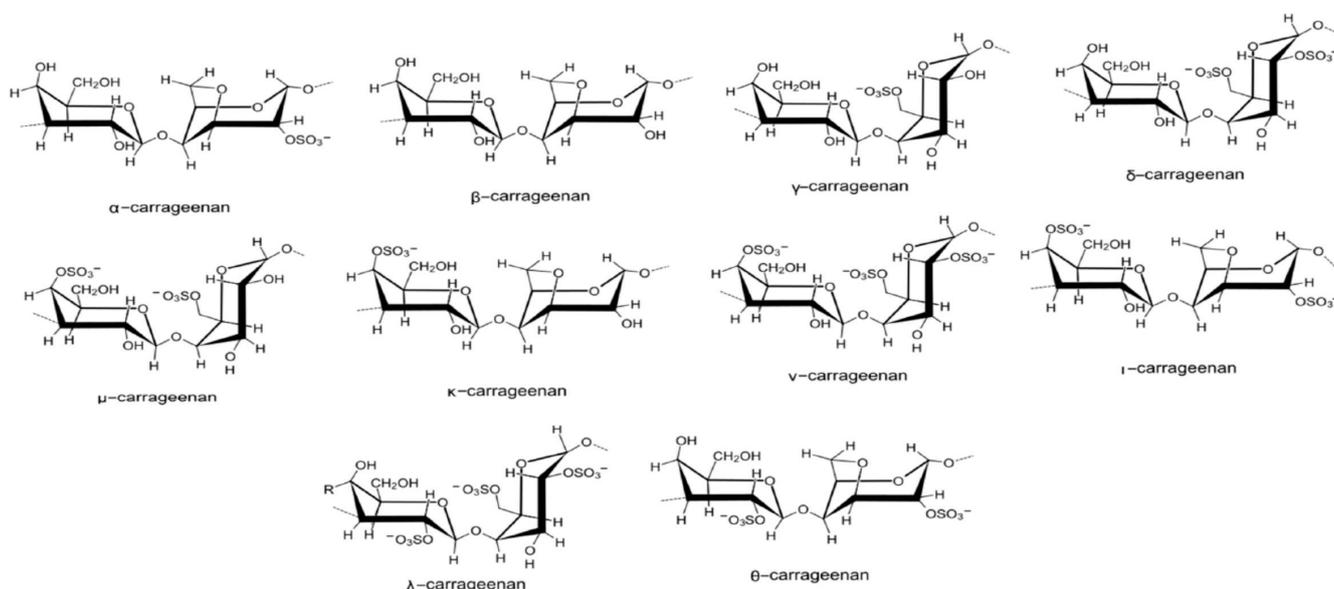


Figure 10. Chemical structures of main carrageenans.

Beyond their applications in pharmaceutical, cosmetic, and food industries for colloid stabilization, thickening, protein binding, and gelling, carrageenans also influence plant growth stimulation and serve as pathogen resistance generators, offering crop protection. Despite their versatile properties, their reduced gel strength and anticoagulant effect remain as notable disadvantages [67–70].

Pectins

Pectins, polysaccharides found in the cell walls of superior plants, feature structures composed of D-galacturonic acid segments linked by α -(1,4) bonds, creating a linear chain framework with interrupted extensively branched regions. Variations in composition depend on the botanical origin (Figure 11).

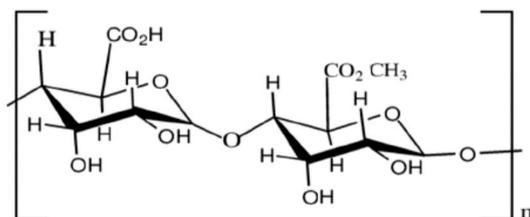


Figure 11. Chemical structure of pectins.

Notably, pectins exhibit limitations such as a reduced water-vapor barrier and poor mechanical characteristics [71].

Arabic Gum

Comprising a complex combination of glycoproteins and polysaccharides (Figure 12), prominently featuring arabinose and galactose, Arabic gum is a water-soluble neutral polymer widely employed as a thickener, stabilizer, and emulsifier in the pharmaceutical, cosmetics, and food industries.

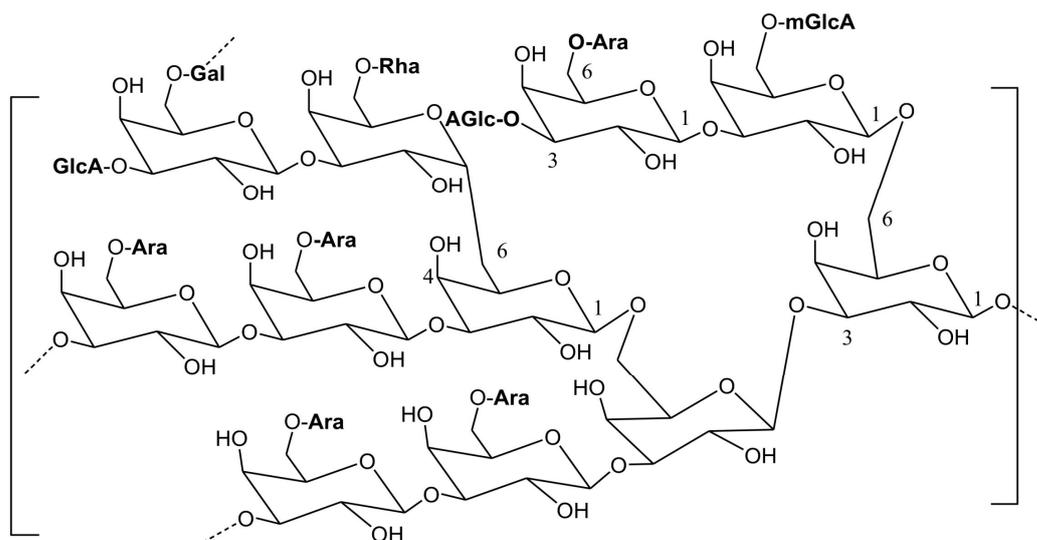


Figure 12. Chemical structure of Arabic gum.

Beyond its conventional uses, Arabic gum serves as a versatile excipient, contributing to the development of nanoscale scaffolds for drug delivery and biomedical practice. Strategies include cross-linking to form hydrogels, combining with other polymers, creating drug conjugates, and attaching to NPs, showcasing its potential biomedical implementation [72–75].

Guar Gum

Guar gum is a water-soluble polysaccharide with a high molecular weight, extracted from the seeds of *Cyamopsis tetragonolobus*. It consists of a primary chain of D-mannopyranose residues linked by β -(1,4) glycosidic bonds, connected to D-galactopyranose residues through α -(1,6) glycosidic bonds (Figure 13).

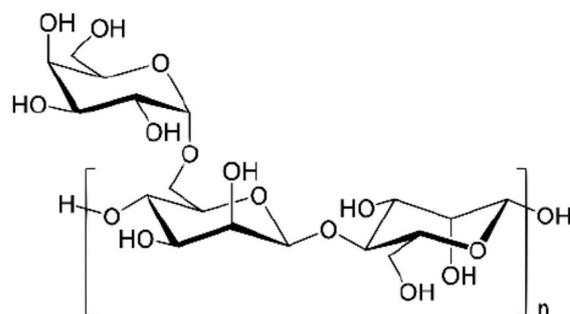


Figure 13. Chemical structure of guar gum.

Known for its emulsifying, thickening, and stabilizing properties, guar gum finds applications in the food, pharmaceutical, and cosmetic industries. Its cold-water solubility is influenced by the galactose/mannose molar ratio. Modified through functionalization (carboxymethylation, hydroxyalkylation, or esterification), guar gum is tailored for biomedical applications, enhancing its mechanical features and reducing aqueous solubility. Guar gum and its derivatives are particularly suitable for oral drug delivery due to their heightened stability across a large pH range [76–79].

Inulin

Inulin is a natural, inexpensive polysaccharide composed of fructose chains joined by β -(2-1) bonds with a glucose terminal unit (Figure 14).

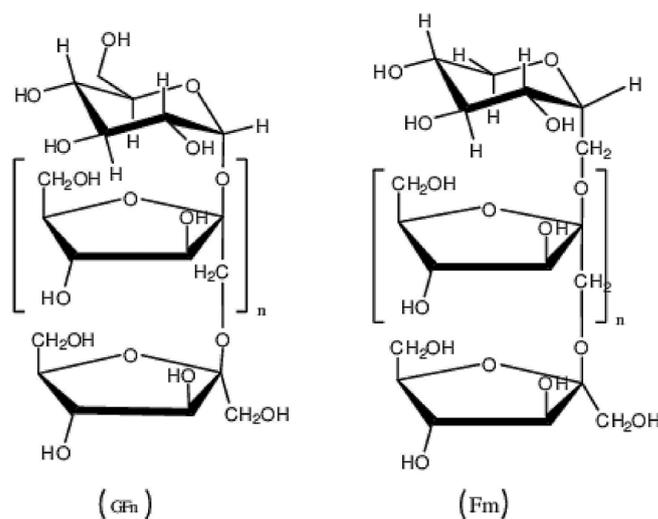


Figure 14. Chemical structure of inulin.

The applications of this biopolymer are multiple, especially in prebiotics and nutraceuticals. The latest research reported various potential biomedical applications: the development of target delivery systems (stable against the action under low pH and the action of specific enzymes such as pepsin and lipase) for colon cancer strategies, nanocarriers with antitumor and antioxidant activities, increasing calcium absorption, and others [80].

Glycosaminoglycans

Hyaluronic Acid

Hyaluronan or hyaluronic acid (HA) is a non-sulfated glycosaminoglycan with a linear structure, consisting of disaccharide repeat segments of β -1,4-D-glucuronic acid and β -1,3-N-acetyl-D-glucosamine connected by β -1,4-glycosidic bonds (Figure 15).

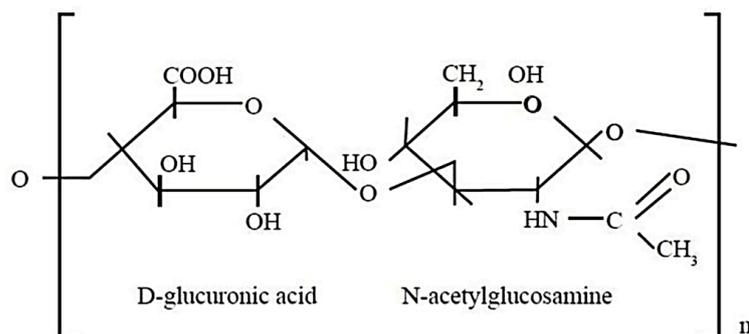


Figure 15. Chemical structure of hyaluronic acid.

Predominantly found in the extracellular matrix of vertebrate soft connective tissues, HA plays a crucial role in tissues like the umbilical cord, synovial fluid, skin, and vitreous humor. Commercially sourced from rooster combs or bacterial fermentation, it is an anionic polysaccharide with the ability to absorb a significant amount of water, serving as a lubricant in native extracellular matrixes and influencing connective tissue viscoelasticity. HA has the potential for chemical alteration through processes such as cross-linking and grafting. In numerous tumor and inflammation conditions, cluster of differentiation (CD)44 and CD168 serve as major ligands for HA. It also plays pivotal roles in biological processes such as cell proliferation, tumor invasion, tissue homeostasis, angiogenesis, and matrix organization through its interactions with cells. Despite its susceptibility to accelerated degradation, HA is extensively used in applications such as drug delivery, TE, and cutaneous rejuvenation. Challenges lie in the brittleness and aqueous solubility of HA hydrogels, leading to the development of useful biomaterials like derivatized HA and IPNs/PICs of HA, albeit facing issues of increased cost and inferior mechanical features [81–85].

Chondroitin

Chondroitin sulfate, a primary component of hyaline cartilage in cartilage and at the bone calcification location, is a sulfated glycosaminoglycan with recurrent disaccharide segments of β-1,4-linked-D-glucuronic acid and β-1,3-linked N-acetyl galactosamine, featuring certain sulfated positions (Figure 16). The two major chondroitin sulfates vary in sulfate positions at 4 or 6.

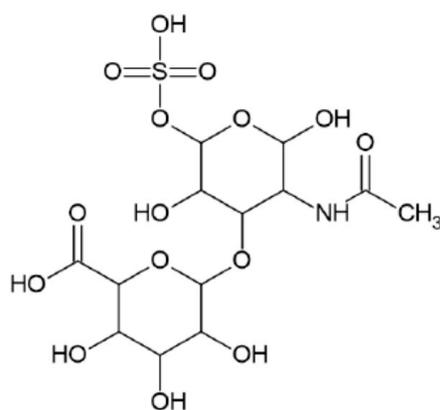


Figure 16. Chemical structure of chondroitin sulfate.

With polar carboxyl and hydroxyl groups, the polymer exhibits covalent/electrostatic interactions with other materials. Possessing antithrombosis, negative immunogenic, anticoagulant, antioxidant, and antiatherosclerosis actions, chondroitin sulfate serves as a valuable biomaterial. Widely employed in osteoarthritis treatment, it can target CD44 receptors on tumor cells, making it applicable for cancer management [86–91].

2.2. Proteins

2.2.1. Collagen

Collagen is the oldest protein structure identified in dinosaur fossils. Approximately 30% of the total animal protein is represented by collagen, which is indispensable in maintaining the biological integrity of the connective tissues. Currently, there are 29 types of collagens, characteristic of different tissues of the human body. Figure 17 shows the chemical structure of collagen I (α chain).

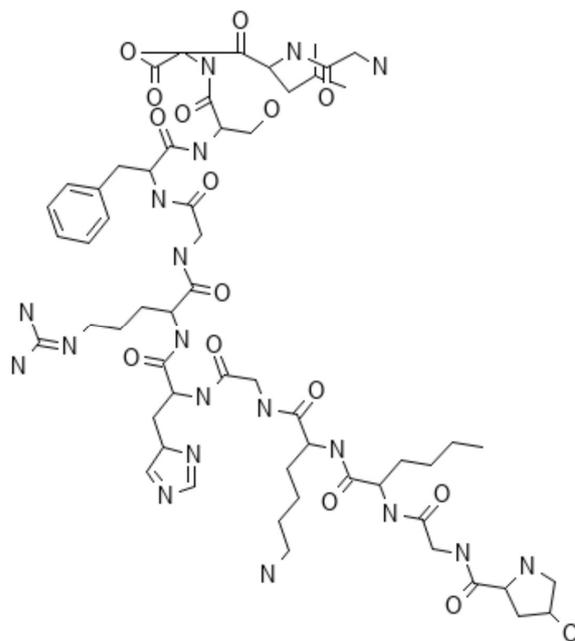


Figure 17. Chemical structure of collagen I (α chain).

The predominant sources of collagen, mainly from bovine origin due to favorable biocompatibility and low immunogenicity, with potential alternatives from marine organisms, are commonly utilized despite associated challenges such as difficulty in sterilization, susceptibility to bacterial contamination, batch variability, and immunogenicity, necessitating ongoing research into the extraction, purification, and industrial-scale production of modified recombinant collagen [92,93]. Common collagen extraction methods involve solubilization in neutral saline, acidic solutions, and acidic solutions with enzymes, albeit at high costs due to requisite chemical treatments for bond elimination, crucial for yield optimization in research-oriented collagen production. Marine collagen, while offering biological safety without disease transmission risks, exhibits lower stability attributed to a lower denaturation temperature compared to mammalian collagen [94,95].

Collagen's amino acid composition, varying across species, influences its physical and chemical properties, thermal stability, solution viscosity, and cross-linking potential, enabling its utilization in wound healing, ophthalmic treatment, drug delivery, and genetic engineering. Key considerations for employing collagen in biomaterial matrices include thermal stability, mechanical resistance, and specific biomolecular interactions. Its excellent biocompatibility and biodegradability render it ideal for various medical implants, such as porous sponges, membranes, and surgical threads, as well as cell culture substrates. While collagen-based supports often incorporate synthetic components for enhanced mechanical strength, they serve diverse purposes, including drug delivery, TE, and epithelial barrier formation to promote tissue regeneration. Despite collagen's inherent biological advantages, its mechanical properties and structural stability may require enhancement through cross-linking treatments, allowing for tailored matrix modifications without compromising cellular responses. Combining natural and synthetic polymers further expands the potential of collagen-based systems to address multifaceted biomedical needs [96–101].

Both naturally derived and recombinant forms of collagen hold significant value as biomaterials, widely utilized in diverse fields such as TE and cosmetic surgery. Recognized by regulatory authorities like the U.S. Food and Drug Administration (FDA), collagen's versatility extends to its incorporation into composite materials with hydroxyapatite and tricalcium phosphate as a biodegradable synthetic bone graft alternative and its use in various drug and gene delivery applications. To conclude, collagen's adaptable utility underscores its indispensable role in biomedical advancements and therapeutic interventions [102–106].

2.2.2. Gelatin

Gelatin, a naturally occurring biopolymer derived from collagen, is abundant in connective tissues, skin, and bones, finding extensive utility across the food, pharmaceutical, and cosmetic industries. Resulting from the partial hydrolysis of collagen, gelatin comprises a heterogeneous ensemble of peptides and proteins, characterized by its hydrophilic nature due to the presence of numerous amino and hydroxyl groups, facilitating water absorption and gel formation (Figure 18).

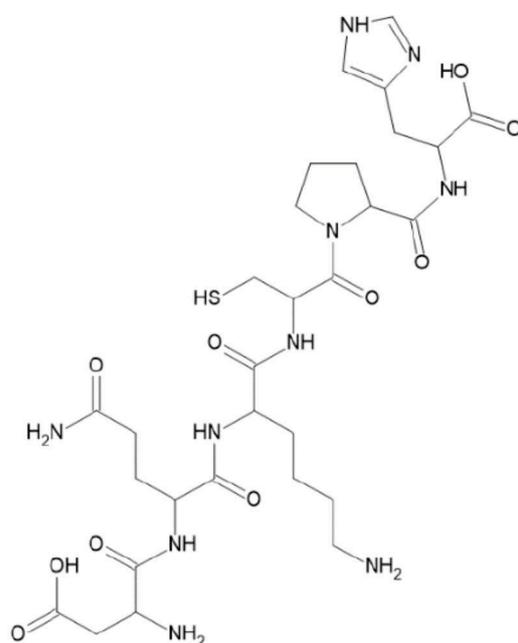


Figure 18. Chemical structure of gelatin.

Its biocompatibility, biodegradability, and capacity for forming stable hydrogels have positioned gelatin as a prominent candidate for DDSs, particularly due to its thermo-reversible properties, enabling gel formation at physiological temperatures suitable for injectable drug delivery applications. Furthermore, gelatin's versatility allows for facile modification to achieve specific drug release profiles through chemical cross-linking or blending with other polymers, thus tailoring mechanical and release properties as needed for diverse therapeutic applications. Additionally, gelatin's inherent bioactivity supports cell adhesion, rendering it suitable for TE endeavors, with the integration of bioactive molecules enhancing its potential for regenerative medicine and targeted drug delivery. Nevertheless, challenges such as potential immunogenicity and rapid *in vivo* degradation necessitate strategies such as cross-linking and polymer blending to address these limitations and ensure the stability and sustained release of encapsulated drugs [107–111].

Cross-linking serves as a pivotal mechanism in bolstering the stability of gelatin structures, averting premature degradation and upholding the integrity of DDSs. Through cross-linking, the porosity and mesh size of gelatin matrices are modulated, thereby influencing the diffusion kinetics of drugs and affording precise control over release mechanisms. This flexibility enables the creation of tailored release profiles, encompassing sustained,

controlled, and stimuli-responsive release patterns. Researchers adeptly manipulate the hydrophilicity, mechanical strength, and degradation rate of gelatin matrices by judiciously selecting cross-linking agents and methodologies [112–114].

Blending gelatin with other polymers represents a prevalent strategy aimed at augmenting the properties and performance of gelatin-based materials, particularly within DDSs. The selection of polymers for blending hinges upon the desired characteristics of the resultant composite material. Notably, poly(lactic-co-glycolic acid) (PLGA) is frequently amalgamated with gelatin to bolster mechanical strength and regulate degradation rates. Conversely, polyethylene glycol (PEG), another hydrophilic polymer, serves to enhance the water solubility and stability of gelatin-based materials, concurrently mitigating protein adsorption, thereby offering potential benefits in specific biomedical applications. Furthermore, the blending of gelatin with chitosan finds extensive exploration in crafting wound dressings, TE scaffolds, and DDSs [115–117].

2.2.3. Silk Protein

Insects, such as silkworms and spiders, generate silk, the most robust natural protein fiber, known for its exceptional mechanical features, including flexibility, increased tensile strength, biodegradability, resistance to compression, and reduced immunogenicity, making it biomedically significant. Silkworm silk consists mainly of fibroin (Figure 19a) and sericin (Figure 19b), with fibroin exhibiting histocompatibility, hydrophobicity, minimal immunogenicity, non-toxicity, and insolubility.

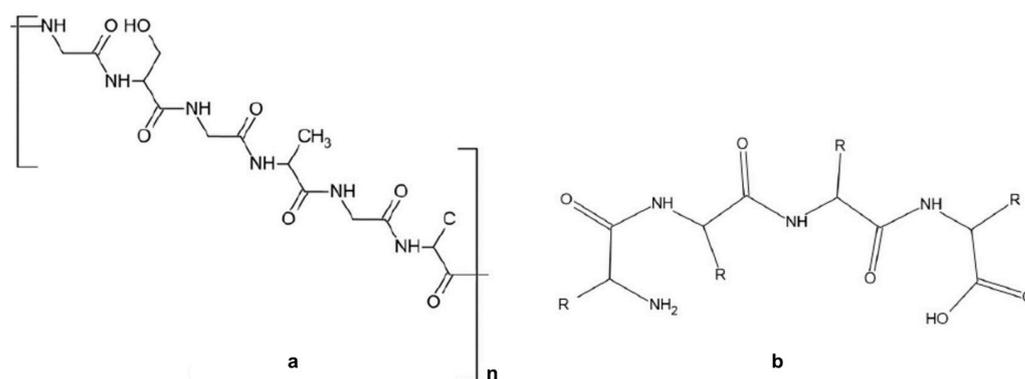


Figure 19. Chemical structure of fibroin (a) and sericin (b).

Derived from silkworm cocoons, fibroin, composed of amino acids (alanine, serine, and glycine), forms various structures like NPs, fibers, gels, hydrogels, scaffolds, and membranes. Its biodegradability and biocompatibility, along with its mechanical strength and malleability, position it as a promising candidate for the drug delivery domain. Sericin, a water-soluble hydrophilic protein, acts as a glue, offering intrinsic antioxidant and anticancer properties in its NP form [118–122].

2.2.4. Albumin

A protein existing in both animal and plant physiological fluids/tissues, albumin plays crucial roles such as maintaining osmotic pressure, neutralizing free radicals, and connecting and transporting numerous substances like drugs and hormones in the circulatory system. It acts as an interface between cells and scaffold materials like collagen, facilitating their integration in TE. Serum albumin, a biodegradable, stable, and non-toxic protein, significantly influences pharmacokinetics and drug distribution/metabolism through drug attachment. Consequently, albumins have surfaced as prospective drug carriers, finding implementation in biosensors, contrast agents, theranostics, and implants for various conditions. The structure and functional groups of albumins (Figure 20) enable the linking and capping of inorganic NPs, increasing compatibility and bioavailability, with low toxicity and selective bioaccumulation [123,124].

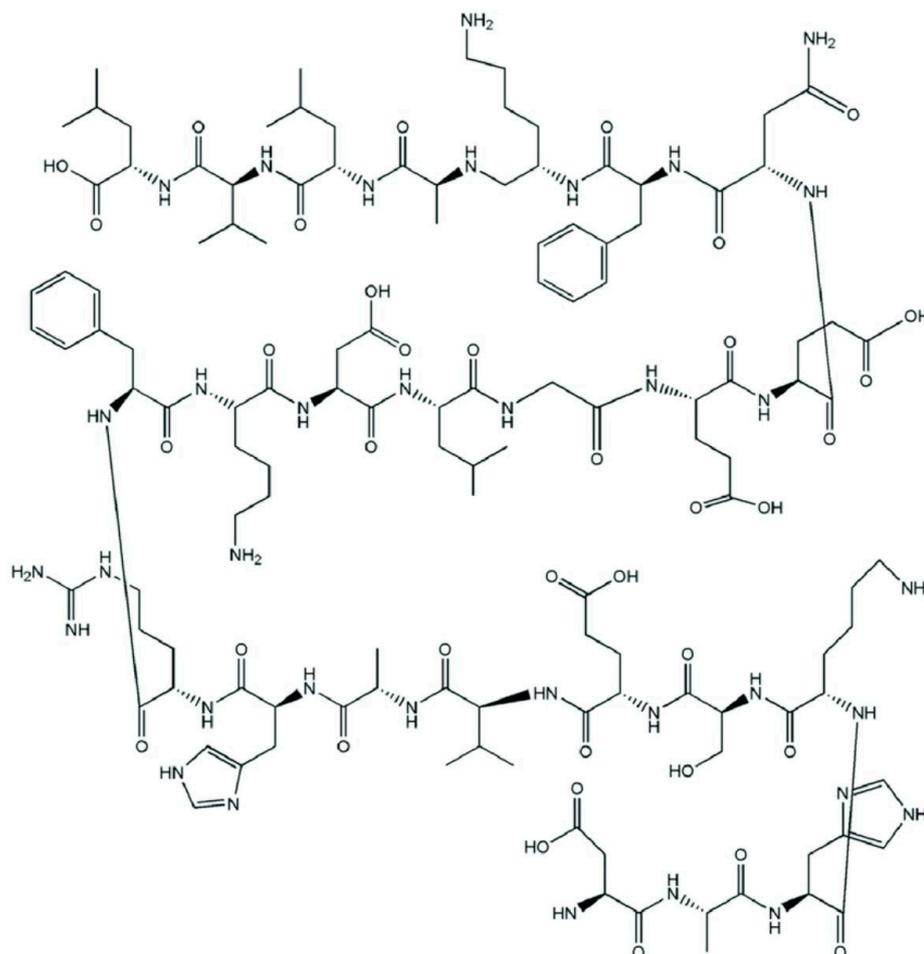


Figure 20. Chemical structure of albumin (serum albumin).

Bovine Serum Albumin

Bovine serum albumin (BSA) plays a pivotal role in fetal bovine serum (FBS) used in vaccine production, necessitating its accurate detection for compliance with regulatory standards. Molecularly imprinted polymers (MIPs), inspired by Fischer's lock and key theory, have emerged as promising artificial receptors for protein detection, offering high selectivity and affinity. Their stability, simplicity in preparation, and adaptability for diverse applications, including vaccine production and clinical diagnostics, position MIPs as valuable tools in biomedical and laboratory settings, facilitating sensitive assays and detection methodologies [125–128].

Human serum albumin (HSA) and BSA are extensively studied major serum proteins, with HSA constituting about 60% of human blood serum, playing a multifunctional role in transporting various substances and influencing their solubility and distribution in the body through its heart-shaped globular structure containing three main domains and two binding sites, while BSA, sharing structural similarities with HSA, is characterized by its acidic nature and negatively charged hydrophobic cavities. Both albumins have applications in biological and medical fields, including the preparation of albumin NPs such as Abraxane for treating metastatic breast cancer [129–133].

Molecular imprinting technology (MIT), recognized for its utility in crafting selective artificial receptors for sensing target molecules, operates by creating specific cavities within a polymer matrix through a three-step process involving the arrangement of functional monomers around the template molecule, polymerization in the presence of cross-linker monomers, and template removal. MIT employs two main strategies, covalent and non-covalent imprinting, with covalent imprinting forming well-designed cavities but requiring

harsh conditions for template removal, while non-covalent imprinting offers a milder approach but may exhibit potential reversibility in complex formation, prompting the introduction of semi-covalent imprinting, which combines the advantages of both methods, with an effective modification employing a “sacrificial spacer” to enhance precision in molecular imprinting [134,135]. Despite advancements in molecular imprinting, persistent challenges exist in developing artificial materials for detecting biological macromolecules like BSA, primarily due to their molecular instability, conformational flexibility, large size, diverse functional groups, and the need for mild imprinting conditions associated with proteins, which often require aqueous environments for stability, prompting the exploration of alternative techniques such as surface molecular imprinting technology (SMIT), epitope-mediated imprinting, micro-contact imprinting, imprinted ILs, and imprinted hydrogels to improve the efficiency of prepared MIPs for selective BSA detection [136].

SMIT has been employed to develop BSA-MIPs, offering benefits such as the homogeneous distribution of binding sites, improved mass transfer, and enhanced adsorption dependency by creating molecular recognition sites on the support substrate surface, notably making the created molecular recognition sites readily accessible to protein molecules [137–141].

ILs have enabled the development of sensitive electrochemical sensors for detecting BSA, with chitosan/IL–graphene-modified electrodes and MIPs showing promising results. Molecularly imprinted hydrogels, responsive to environmental stimuli, offer a dynamic platform for selective protein recognition, particularly with temperature-sensitive components. Utilizing sodium alginate and thermo-sensitive polymers, high-toughness hydrogel films were prepared, exhibiting enhanced BSA adsorption capabilities. Additionally, surface-imprinted materials incorporating hollow magnetite microspheres demonstrated specific BSA recognition, highlighting their potential for bioseparation and biosensor development [142–145].

Other Proteins

Zein

Zein is an amphiphilic protein group consisting of α , β , γ , and δ zein in various proportions, with a predominant proportion being α -zein (about 80%) followed by δ -zein. Zein represents about 50% of the whole protein content in corn (*Zea mays*). The unique properties of zein, such as its solubility, are due to its high proportion of hydrophobic, neutral amino acids like alanine, leucine, and proline, as well as the presence of polar amino acids such as glutamic acid (approximately 20% of its total amino acid content). Its unique physical properties, such as its high thermal and water stability and an isoelectric point (about 6.8) very close to the physiological pH value, are due to the presence of numerous and varied types of functional groups (amines, amides, hydroxyls, carboxylates, and phenols). Zein is used extensively in edible film preparation for the biomedical area and food industries. Recent studies reported the development of new nano- and micromaterials for target drug delivery, imaging, theranostics, and TE [146–148].

Legumin

Legumin is a vegetable protein that contains numerous sulfur-amino acids, with a structure (Figure 21) similar to casein.

It is abundant in soybean seeds, beans, peas, lentils, vetches, and hemp. In recent years, several advanced materials based on legumin have been used in nutraceuticals and biomedical applications [149,150].

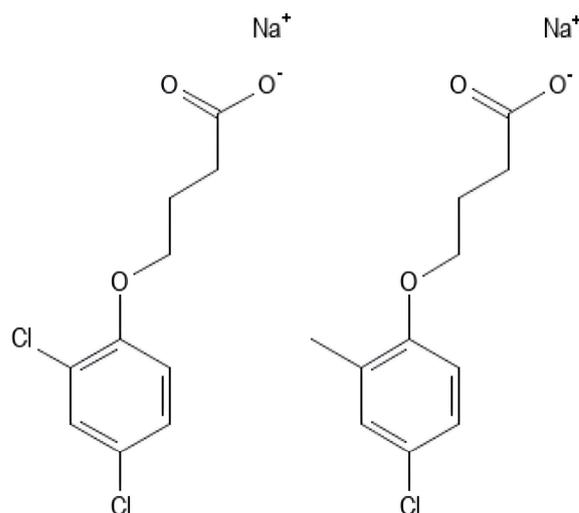


Figure 21. Chemical structure of legumin.

Gliadin

Gliadin proteins (classified as α , β , γ , and ω gliadins) are found in wheat. Among the most alluring properties, from the point of view of the application potential in the biomedical field, are their low water solubility at ordinary pH values (because of its chemical structure (Figure 22) consisting of single-chain polypeptides linked by intramolecular disulfide bonds), high biocompatibility, non-toxicity, and biodegradability [151,152].

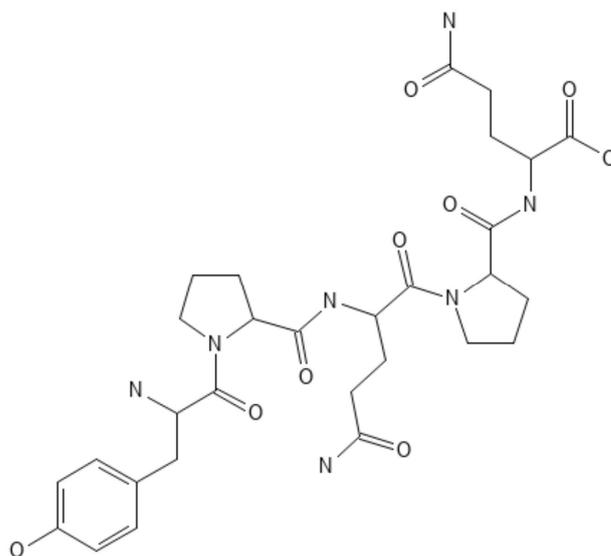


Figure 22. Chemical structure of gliadin.

Recent studies have exploited these characteristics of gliadin for the development of new oral and local DDSs for gastrointestinal (GI) diseases, breast tumors, etc. [151–153].

Avidin

Avidin is a basic, homogeneous glycoprotein consisting of tetrameric biotin-binding protein and about 10% carbohydrate moieties (4–5 mannose and 3 N-acetylglucosamine residues) derived from egg whites.

Avidin is extremely water-soluble and shows high stability in a wide range of pHs and temperatures. Therefore, this biopolymer has gained multiple applications in bio-chemical assays, diagnosis, drug delivery, etc. In recent studies, it was employed for its versatile functionality to obtain new nanomaterials for nano-DDSs and diagnosis [154].

2.3. Lignin

Lignin, comprising up to 35% of lignocellulosic biomass, is the second-most abundant biopolymer, after cellulose (Figure 23) [155–158].

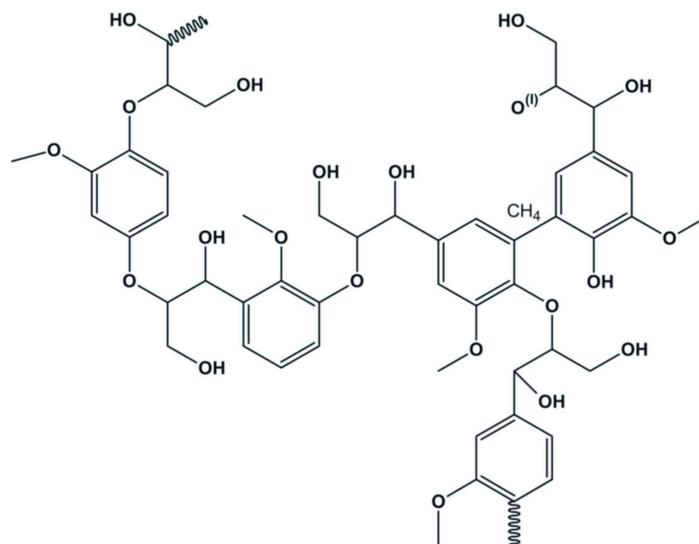


Figure 23. Chemical structure of lignin.

Lignin is considered toxic due to its complex, recalcitrant, and xenobiotic nature, which makes it resistant to enzymatic and microbial degradation. This resistance stems from lignin's complex aromatic structure, which is not readily broken down by most microorganisms. As a result, lignin and its derivatives can accumulate in the environment, posing a challenge for biological systems and microbial communities that are unable to process these compounds efficiently [159]. The main source of toxic lignin comes from lignocellulosic biomass, which is a complex and abundant group of organic materials composed primarily of cellulose, hemicellulose, and lignin. Lignocellulosic biomass is found in agricultural residues, forestry waste, certain grasses, and other plant materials. Among these components, lignin is particularly notable for its complex, amorphous, and recalcitrant nature, which makes it resistant to degradation by enzymes and microorganisms. The paper and pulp industries are significant contributors to lignin generation, as lignin is separated from cellulose during the process of paper production, leading to substantial amounts of lignin by-products. This lignin by-product is often considered waste, although it has the potential for conversion into valuable bioproducts through various biochemical and thermochemical processes. The study conducted by Mohammad and Bhukya (2022) [159] delves into this challenge, presenting a novel biotransformation approach that leverages the capabilities of *Pseudomonas putida* KT2440. This bacterium exhibits remarkable tolerance to high concentrations of lignin and its aromatic derivatives, converting these toxic compounds into eco-friendly biopolymers. The key to transforming lignin into biocompatible materials lies in the acclimatization process and the strategic addition of glucose, which significantly enhances the degradation capability of the strain. This breakthrough underscores a dual benefit: detoxifying the environmental menace posed by lignin and its derivatives while simultaneously synthesizing valuable biopolymers [159].

Lignin exhibits antioxidant, antibacterial, and anti-ultraviolet activities attributed to its unique polyphenolic structure. Nonetheless, the heterogeneity of lignin derived from varied sources and extraction methods poses a significant challenge to its use in the biomedical field [160,161].

In recent times, research studies have been focused on the chemical modification of lignin through techniques such as alkylation, esterification, phenolation, etherification, and urethanization. This approach enhances lignin's solubility, thermal stability, and reactivity while reducing its brittleness, thereby enabling the development of advanced nanomaterials

such as lignin microcapsules, self-assembling NPs, lignin-based complex micelles, lignin-based carbon dots, and biosensors for a wide range of applications, including drug delivery, gene delivery, biosensors, bioimaging, TE, and dietary supplements [162].

2.4. Shellac

Shellac is a composite macromolecule (a long-chain polyester type of resin) consisting of inter- and intra-esters of polyhydroxy carboxylic acids (aliphatic long-chain hydroxy acids and sesquiterpene acids). Unique features of this biopolymer, namely its thermoplasticity, non-toxicity, and water stability at neutral to acidic pHs, determined its use in the medicine and food industries. The latest research reported the development of various tailored DDSs based on shellac [163].

3. Chemical Modifications of Biopolymers

3.1. Cross-Linking

Cross-linking involves the formation of a network in polymer solutions, enhancing mechanical features and viscoelastic behavior. The unstable bonds, produced through physical or chemical cross-linking, can be disintegrated under physiological conditions. Chemical cross-linking, utilizing covalent agents, enhances mechanical stability but may impact polymer integrity and increase toxicity. The ionic gelation procedure involves interactions between polymers with opposite charges and cross-linking agents with complementary charges. In contrast, polyelectrolyte complexation relies only on electrostatic interactions among positively or negatively charged polyions, without the use of cross-linking agents. Innovative dual cross-linking combines both physical and chemical factors, reducing toxicity and improving stability. Interfacial cross-linking allows nanocapsule preparation without additional agents. Alginate readily cross-links via ionic interactions with calcium ions, forming gels employed for encapsulating bioactive molecules. Cationic polysaccharides like chitosan can be cross-linked with glycerol-phosphate disodium salt [164–168].

3.2. Functionalization and Conjugation

Biopolymer conjugation refers to the covalent attachment or linking of two or more biopolymers through specific chemical reactions or cross-linking mechanisms. The new resulting polymer structure has enhanced properties or functionalities. Biodegradable polymers, especially polysaccharides, possess diverse functional groups that can undergo covalent modifications with hydrophobic or hydrophilic substances, enhancing their suitability for biomedical approaches. Chemical conjugation, such as the PEGylation of polysaccharides or proteins ($-OH$ groups of PEG react with $-COOH$, $-NH_2$, or $-SH$ groups on the target molecule), can modify the physical properties and solution behavior for specific utilization (Table 1). Various strategies of functionalization, including etherification, esterification, and enzymatic modifications, yield polysaccharide derivatives with improved biological, chemical, and physical features. Reactive functional groups introduced by phosphorylation, sulfation, acylation, and alkylation significantly impact inherent hallmarks. Additionally, enzymatic modifications, involving glycosylation, oxidation, and molecular weight depletion, have been designed for diverse pharmaceutical employments [169–174].

Table 1. Examples of enhanced biopolymer properties for pharmaceutical applications through chemical modifications.

| Biopolymers | Chemical Modifications | Enhanced Properties |
|-----------------|------------------------|---|
| Polysaccharides | Chitin and chitosan | Improved mechanical strength and stability; prolonged release time; enhanced interaction with other molecules, water absorption capacity, and resistance to enzymatic degradation; and increased surface activity |
| | Cellulose | |
| | Hyaluronic acid | |
| | Alginate | |
| | Pectins | |

Table 1. Cont.

| Biopolymers | Chemical Modifications | Enhanced Properties |
|-------------|------------------------|---|
| Proteins | Collagen | Glutaraldehyde cross-linking, carbodiimide cross-linking, glycosylation, hydroxylation, PEGylation [172], acetylation |
| | Gelatin | PEGylation, hydroxylation, glycosylation, acetylation [173], cross-linking (glutaraldehyde [113] or transglutaminase [174]) |
| | Albumin | Site-specific PEGylation [175], drug conjugation [176] |

IPNs: interpenetrating polymer networks; PEG: polyethylene glycol.

3.3. Interpenetrating Polymer Networks

IPNs consist of two or more incompatible polymers synthesized together, with one system polymerized in the presence of another. For instance, an aqueous solution containing a water-soluble polymer and a vinyl/acryl monomer can be polymerized to form intertwined polymer chains. Unlike polymer blends, IPNs expand but do not dissolve in solvents, reducing flow/creep conduct. They rely on physical forces like electrostatic and hydrogen bonding, making them suitable as vehicles for DDSs and scaffolds for TE. IPN hydrogels are innovative biomaterials for drug delivery, with polysaccharide-based IPNs, particularly using chitosan and alginates, offering a unique enlargement ability, mechanical strength, and specificity [175–180].

3.4. Graft Copolymers

Grafting serves as a versatile strategy to enhance the compatibility between synthetic and natural polymers, particularly in the chemical alteration of polysaccharides. Various polysaccharides, like cellulose, HA, chitosan, and starch, have been successfully employed in grafting processes. The “grafting through/on/from” approaches enable the incorporation of hydrophilic or hydrophobic polymeric moieties onto the polysaccharide backbone, with microwave irradiation emerging as an efficient method, offering improved attributes in terms of flame resistance, water repellence, thermal stability, and opposition to acid–base aggression. Polysaccharide-based graft copolymers, especially those with amphiphilic characteristics, find relevance in the biomedical domain, showcasing potential as biomaterials or conveyances for DDSs [181–186].

3.5. Block Copolymers

Biodegradable block copolymers have garnered significant attention in medical and pharmaceutical studies on account of their customizable biodegradability, biocompatibility, and self-assembly characteristics. These polymers serve as effective vehicles for DDSs, forming drug-loaded NPs that undergo degradation in biological circumstances and are subsequently evacuated via the renal system. The precise control over the structure of block copolymers, achieved through advancements in polymerization techniques like atom transfer radical polymerization (ATRP) and reversible addition–fragmentation chain-transfer (RAFT) processes, coupled with modern nanoaggregate interpretation methods, has heightened their relevance in various biomedical applications [187–189].

3.6. Polyion Complexes

Polyelectrolytes, water-soluble charged polymers, undergo dissociation in aqueous solutions, resulting in the formation of a macroion and counterion. The conduct of polyelectrolytes in solution is significantly affected by the existence of salts, as well as pH and

temperature variations. Natural polyelectrolytes find extensive applications in various industries. Charged polymers interrelate with oppositely charged ones, forming soluble PICs or insoluble coacervates, which have biological significance. Examples include PICs of oppositely charged polysaccharides like chitosan and alginates, and those involving charged proteins/polysaccharides and oppositely charged small molecules or polymers. Dilute solutions of polyelectrolytes, when mixed with oppositely charged substances, can spontaneously form a new phase via powerful electrostatic interactions. Nevertheless, a comprehensive interpretation of the physical status (solid/liquid-like) is essential. The presence of supporting electrolytes significantly affects the origination and qualities of these complexes [187,190–193].

4. Pharmaceutical and Biomedical Applications of Biopolymers

The applications of biomass polymers are economically, socially, and environmentally sustainable, finding utility across various domains. Ongoing extensive research further explores their potential in diverse fields. Their inherent biocompatibility, biodegradability, and minimal immune response induction position them as promising candidates for applications in TE, as well as in drug and gene delivery systems (Figure 24) [194].

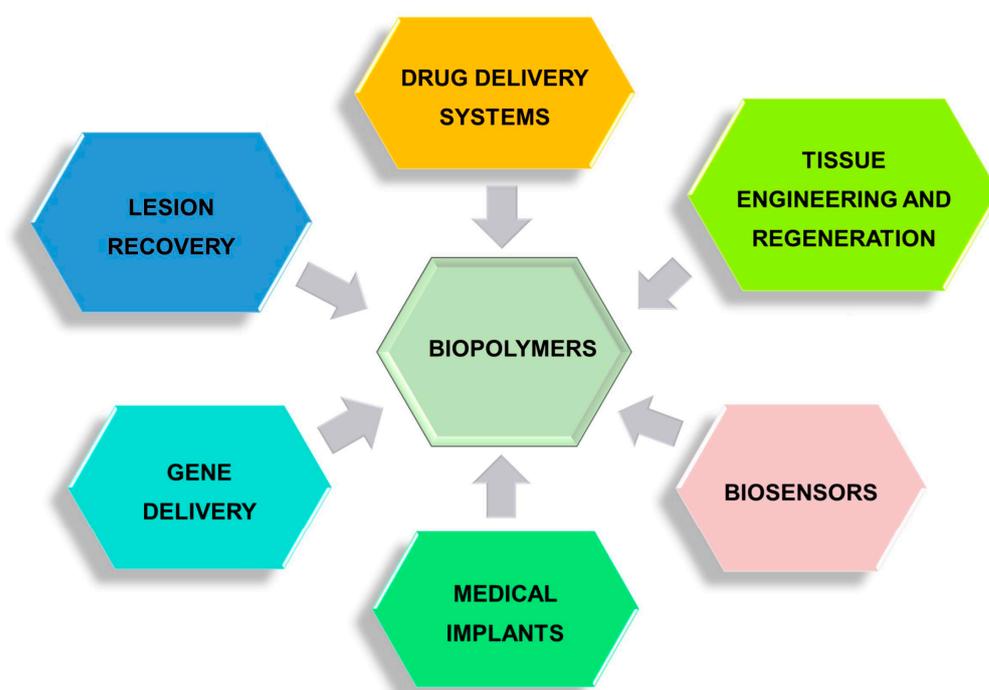


Figure 24. Main biomedical applications of biopolymers.

Current advancements in biopolymers have garnered attention across numerous domains due to their improved features and facile commercialization. Typical biopolymers, including elastin, silk, chitosan, keratin, and collagen, have been strategically mixed with synthetic polymers to amplify their actions as composites. The escalating demand for biodegradable natural polymers is particularly notable in the production of packaging film materials, with applications spanning medical, pharmaceutical, and food industries. This contemporary trend emphasizes a burgeoning focus on biopolymers, especially the synergistic blend of synthetic and natural polymers in composite materials. In the medical and pharmaceutical sectors, these polymers play pivotal roles in gene therapy, BTE, and cell-based transplantation, contributing to the development of products such as implantable medical devices, 3D scaffolds, artificial skin, wound dressing materials, and dialysis systems (Table 2) [25,38,195,196].

Table 2. Examples of biopolymers and their pharmaceutical/biomedical applications.

| Biopolymers | Applications | References |
|--|-------------------------------------|---------------|
| Chitosan Fibroin Starch Gelatin Cellulose Bacterial nanocellulose Collagen Biopolymer composites Elastin-like polypeptides Albumin microspheres | Drug delivery systems | [197–209] |
| Polyethylene imine Poly(L-lysine) Albumin Gelatin Chitosan | Gene delivery | [202,210,211] |
| Hyaluronic acid Cellulose Chitosan Alginate | Lesion recovery | [212–220] |
| Chitosan nanoparticles | Targeted diagnosis | [221–227] |
| Silk Gelatin Collagen Chitosan Hyaluronic acid Alginate Polyurethanes Polyphosphazenes Polyanhydrides Polyesters Polyhydroxyalkanoates Acrylate polymers polyblends | Tissue engineering and regeneration | [228–233] |
| Chitosan-based films | Biosensors | [234–238] |
| Chitosan Polylactic acid Gelatin Collagen Polyhydroxyalkanoates Polyhydroxybutyrate | Medical implants | [203,239–241] |

4.1. Drug Delivery Systems

DDSs encompass the conveyance of natural compounds, genes, or synthetic pharmaceutical drugs to the accurate location without inducing negative effects on biological systems. DDSs necessitate comprehensive considerations, including high drug loading, cellular uptake, programmed target specificity, clearance, metabolism, pharmacokinetics, toxicity, and excretion. An ideal system should enhance drug efficiency and enable controlled release from a biocompatible nanocarrier, promoting patient compliance. Passive accumulation at the target site, facilitated by the enhanced permeability and retention (EPR) effect, contributes significantly to DDS efficacy. Traditional DDSs, characterized by immediate release and potential toxicity, often require frequent administration for therapeutic levels. To address these limitations and enhance pharmacokinetics, second- and third-generation DDSs explore modified particle surfaces for improved stealth effects, utilizing hydrophilic blocks like PEG to reduce plasma protein adsorption and rapid clearance. Stealth effects, influenced by factors such as size, shape, and core composition, increase

blood circulation and accumulation in highly vascularized areas. Despite these advancements, the clinical translation of formulations based on biodegradable polymers remains limited [231,232].

Biopolymers have become integral in pharmaceutical applications, serving as accurate DDSs with diverse structures for various physiological and medical needs. Structural, protective, and reserve polysaccharides exhibit capability in constructing conjugates with lipids and proteins, facilitating drug transport. Common biopolymers like chitosan, fibroin, starch, gelatin, cellulose, and collagen are harnessed for drug delivery through suspensions, employing methods such as freeze-drying, microemulsion, electrospraying, and supercritical fluid extraction. Biopolymer composites, labeled as excipient materials, are gaining attention in the pharmaceutical industry for drug delivery due to their renewable characteristic, biodegradability, endurance, and reduced toxicity. The focus on targeted drug delivery systems (TDDSs) using polymeric DDSs is increasing, exploring avenues like elastin-like polypeptides (ELPs) for intra-articular delivery and albumin microspheres for controlled drug release. BNC shows promise in delivering proteins with maintained integrity and activity, emphasizing controlled release kinetics, biocompatibility, and hydrophilicity. The abundance of naturally available biopolymers facilitates the cost-effective development of hydrogels and nanogels through various cross-linking polymerization techniques, offering potential applications in cancer treatment [204–208]. Various nanomaterials, encompassing organic polymers and inorganic compounds, have been explored as transportation for DDSs, including liposomes, dendrimers, polymersomes, NPs, nanogels, polymer micelles, nanofibers, nanocapsules, and nanocomposites. These nanocarriers play a crucial role in drug delivery, but precise adjustments in shape, size, porosity, and polydispersity, as well as in surface charge and characteristics, are necessary for their specific applications [233–241]. Drug loading and encapsulation efficiencies (DLE and DEE) are crucial parameters in the representation of DDSs. The encapsulation efficiency, drug release profile, and overall performance of polymeric NPs or self-assembled nanoaggregates are influenced by factors like shape, size, surface features, charge, stimuli responsiveness, and polymer biodegradation kinetics. The optimization of these parameters is essential for achieving regulated drug delivery to target sites with optimal doses. Experimental research and theoretical modeling are conducted to configure NPs with specific control over drug discharge, offering expansive and promising applications in clinical medicine. An optimal NP-based delivery system should exhibit an elevated loading capacity, accomplished through drug incorporation during NP preparation or post-incubation diffusion. Understanding drug release mechanisms, influenced by desorption, diffusion, and erosion of the NP matrix, is vital for tailoring drug delivery kinetics. The kinetics of drug delivery are influenced by the biodegradation, diffusion, solubility, and loading effectiveness of the matrix materials. For biodegradable polymers, drug dissolution, swelling, erosion, and diffusion may occur at the same time, inducing zero-order release kinetics, while NP size influences the release pattern [242,243]. The regular course of drug administration via oral ingestion faces challenges with hydrophobic drugs exhibiting reduced bioavailability and protein-based drugs being exposed to enzymatic disintegration in the GI tract. To address this, innovative DDSs are engineered for controlled and targeted release, encapsulating or solubilizing drugs using nanosized elements like inorganic NPs, dendrimers, polymersomes, polymer micelles, liposomes, solid lipid dispersions, and mesoporous materials. Amphiphilic block and graft biomass polymers play a crucial role in these systems, forming nanostructures with enhanced drug-loading ability. Thermo- and pH-responsive polymers are commonly engaged, reacting to the acidic pH of tumor cells and increased concentrations of glutathione (GSH) triphosphate, making them suitable for targeted drug delivery. Biomass polymers, through controlled degradation into biocompatible by-products, offer constant discharge at targeted locations within therapeutic concentration ranges. The provocation lies in designing multiple-functionalized DDSs to ensure on-request, manageable drug dispensation under various external stimuli [196,244–248].

4.2. Gene Delivery

Gene therapy is a promising approach to address various metabolic, neoplastic, cardiovascular, neurological, and genetic disorders. It utilizes deoxyribonucleic acid (DNA) and ribonucleic acid (RNA)—therapeutic gene molecules—to modify mutated, absent, or abnormal genes. To overcome challenges like the brittle characteristic of therapeutic genes and biological impediments, non-viral vectors, including lipid-based nano-assemblies and cationic polyelectrolytes, are developed for gene delivery. Polyplexes formed by cationic polyelectrolytes like polyethylene imines and poly(L-lysine) show potential, but a careful consideration of polymer design, degradability, and toxicity is crucial. Non-toxic protein-based vectors, such as albumin and gelatin, are widely used due to their biocompatibility and biodegradability. Cationic polysaccharides, such as chitosan, though limited by solubility, are explored, with chemical alterations to enhance gene complexation [209–211].

4.3. Lesion Recovery

Wound healing is an intricate biological and cellular mechanism, including phases of inflammation, hemostasis, proliferation, and remodeling triggered by tissue severance. Cutaneous injuries create vulnerabilities for pathogenic bacteria, leading to virulence factor production that hinders tissue integrity, often associated with biofilm formation. Various dressings are employed for severe wounds, possessing high absorption capacity, wound visibility, pain-free removal, and non-allergenic properties. Biomass polymers are utilized in wound dressing formulations such as hydrogels, films, hydrocolloids, membranes, and foams. Polysaccharides like HA, cellulose, chitosan, and alginate stand out as adaptable biomacromolecules due to their increased chelation ability, non-toxicity, biodegradability, biocompatibility, multifunctional groups, and simple chemical alteration, making them effective in the management of cutaneous infections [212–220].

4.4. Targeted Diagnosis

Targeted therapy involves the use of ligand-functionalized NPs to accurately recognize receptors overexpressed in malignant tumor cells, enabling tumor-selective DDSs. Various ligands, including antibodies, aptamers, transferrin, peptides, and folic acid, have been explored to enhance the specificity of DDSs. NPs derived from biomass polymers, particularly chitosan, show promise in anti-tumor targeting due to their ability to promote cellular uptake and adhesion to mucosal surfaces. This targeted approach aims to improve drug release directly to cancer cells, enhancing therapeutic efficacy [221–227].

4.5. Tissue Engineering and Regeneration

TE integrates regulations of engineering and medical sciences to elaborate biological tissue replacements for improving, maintaining, or restoring function. The engineered tissue can be developed in vivo or in vitro and transplanted, serving diagnostic purposes as well. Scaffolds, valuable in this particular field, rely on biodegradable polymers, such as biomass polymers, from natural sources. Perfectly, scaffolds should be mechanically powerful, biocompatible, biodegradable without toxic byproducts, possess an appropriate surface morphology for cell interaction, and sustain cell attachment, proliferation, and differentiation. Proteins like silk, gelatin, and collagen, along with polysaccharides like chitosan, HA, and alginate, are universal natural scaffold materials. Nevertheless, due to the intricate structure of biomass polymers and concerns like immunogenicity, synthetic polymers like polyurethanes, polyphosphazenes, polyanhydrides, and polyesters gain significance. In BTE, the challenge lies in regenerating bone deficiencies caused by tumors, fractures, or trauma, where polymers, ceramic materials, and metals serve as scaffolds to encourage new tissue formation. Inorganic materials like titanium and steel and degradable biomass polymers show promise, with PHA composites, acrylate polymer polyblends, and magnesium-based compounds demonstrating excellent mechanical and cell adhesion features for orthopedic applications [228–233].

4.6. Biosensors

Advanced diagnostic instruments often rely on automated analyzers, but their maintenance is both expensive and time-consuming. The demand for more accelerated, compact, and cost-effective devices in laboratory testing has led to the rise in biosensors. These diagnostic tools recognize precise biochemicals by utilizing immobilized biomolecules, such as receptors, antibodies, or enzymes, on electrodes, offering elevated specificity, maneuverability, utilizer accessibility, and rapid response duration. Biosensors, incorporating biological constituents and transducers, can monitor medical conditions through the examination of clinical samples or real-time physiological modifications inside the human organism. Chitosan-based films have been instrumental in enhancing the sensitivity of biosensors, particularly in applications like cholesterol detection [234–238].

4.7. Medical Implants

Various biopolymers, like chitosan and PLA, have found extensive use in pharmaceutical purposes, due to their biocompatibility and biodegradability when employed as implantable medical devices. For example, chitosan serves as implants in surgery (e.g., nerve regeneration), cardiology (e.g., heart valves), and ophthalmology (e.g., contact lenses). Composites of chitosan are applied for tissue regeneration, bioartificial livers, and bone scaffolds, whereas HA implants aid in tissue maturation in otolaryngology. Gelatin serves multiple purposes, such as 3D biomatrices in dermatology, bone replacement in orthopedics, and grafts in cardiology. Collagen, a ubiquitous biopolymer in mammals, is utilized as cardiovascular implants, bone marrow, and bone scaffolds, while PHAs are employed in nerve, vascular, and esophageal implants. Moreover, polyhydroxybutyrate (PHB) serves as cell culture scaffolds and surgical implants. Biopolymers derived from biomass are designed via techniques such as leaching, freeze-drying, 3D bioprinting, electrospinning, and casting, serving as medical implants, such as barrier membranes and stents, as well as carriers in cell, gene, drug, and growth factor delivery systems [203,239–241].

4.8. Challenges

Using biomass polymers in pharmaceutical applications has several challenges that researchers must overcome. Although the vast majority of them possess desirable properties such as biocompatibility and biodegradability, their use comes with a set of limitations. Proteins (collagen, gelatin, albumin) can potentially trigger immune responses in the body, leading to allergic reactions or even rejection in the case of their use as carrier systems [249,250]. There are some good strategies to minimize immunogenic responses, such as purification techniques or surface modification, but also in this case, it must be considered that traditional purification methods may affect protein properties and integrity [251,252]. The same challenge of maintaining structural integrity is encountered when applying the processes of sterilization techniques (heat, irradiation) [253]. An alternative sterilization that preserves protein functionality while eliminating microbial contaminants for maintaining product quality and safety was developed [254,255]. Other challenges in the case of protein biopolymers were the ethical or cultural objections that involve the use of animal-derived proteins in pharmaceutical products, particularly those sourced from pigs or cows [256]. Marine or plant-based sources are good alternatives to solve consumer preferences and address ethical concerns [256].

Polymer stability [257] is another important parameter in pharmaceutical applications. Maintaining structure during processing and formulation can be challenging. For example, collagen may undergo denaturation or degradation, affecting its stability and performance as a drug carrier. Stabilization techniques, such as cross-linking or encapsulation within protective matrices such as those discussed above, are frequently applied [258].

The cost of sourcing high-quality biopolymers and any additional processing steps can influence the overall affordability of biopolymer-based pharmaceuticals. For example, lignin extraction methods, such as those from paper and pulp industries or lignocellulose biorefineries, may not be optimized for pharmaceutical-grade lignin production [259,260].

Developing efficient and sustainable extraction processes, as well as exploring alternative lignin sources, can decrease the overall price. Therefore, researchers must improve the characteristics of biopolymers to increase their functionalities and pharmaceutical applicability.

5. Conclusions and Perspectives

The investigation into the domain of biomass polymers for pharmaceutical applications reveals an exciting pathway toward the creation of innovative, sustainable healthcare technologies. The sophisticated chemical modifications applied to these biopolymers—such as cross-linking, functionalization, and conjugation, along with the engineering of complex structures like IPNs, grafts, and block copolymers, and PICs—mark a significant step forward in enhancing their versatility and compatibility with biological systems. This progress lays the groundwork for their widespread application in a variety of pharmaceutical and biomedical contexts, including drug and gene delivery, lesion healing, precision diagnosis, TE, biosensing, and the development of medical implants. This exploration underscores the transformative potential of biomass polymers in medicine and pharmaceutical science, offering novel approaches to address enduring challenges in these fields.

Looking forward, several pivotal areas warrant further investigation to fully harness the capabilities of biomass polymers. The development of advanced DDSs that offer controlled release and biodegradation kinetics tailored to physiological conditions is of paramount importance. Optimizing gene delivery vectors to achieve high efficiency with minimal cytotoxicity, expanding research on the integration of biomass polymers with living tissues for regeneration and TE, and advancing biosensor technologies for the sensitive detection of disease biomarkers represent critical frontiers in this domain. Furthermore, the evaluation of biomass-polymer-based medical implants *in vivo* to assess their long-term biocompatibility and functionality will be crucial for their clinical application.

Moreover, tackling the challenges of sustainable biomass sourcing and the scalable production of these polymers will ensure their accessibility and economic feasibility. Encouraging interdisciplinary collaboration will also be instrumental in pioneering new biomass polymer formulations tailored for specific medical applications. Through dedicated research and collaborative innovation, the promising future of biomass polymers in enhancing pharmaceutical and biomedical solutions can be fully realized, steering us toward a future where these sustainable and effective technologies become a cornerstone of healthcare advancements.

Author Contributions: Conceptualization, C.B., A.R. and G.D.M.; methodology, A.-E.S., A.B., M.V.C., I.A.B., T.V. and G.V.; writing—original draft preparation, C.B., A.R., G.D.M. and L.E.B.; writing—review and editing, C.B., A.R. and G.D.M.; supervision, C.B., A.-E.S. and L.E.B.; funding acquisition, C.B. and A.-E.S. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data described in the manuscript will be made publicly and freely available without restriction at https://docs.google.com/document/d/1Lz89uNfXp_TYbX6vJFAr0_JiolruP1Tk/edit?usp=sharing&ouid=106104952021876684289&rtopof=true&sd=true (accessed on 5 March 2024).

Acknowledgments: This work was supported by a grant from the European Research Executive Agency, Topic: HORIZON-MSCA-2022-SE-01-01, Type of action: HORIZON TMA MSCA Staff Exchanges, Project: 101131420—PHENOCYCLES.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. George, A.; Sanjay, M.R.; Srisuk, R.; Parameswaranpillai, J.; Siengchin, S. A comprehensive review on chemical properties and applications of biopolymers and their composites. *Int. J. Biol. Macromol.* **2020**, *154*, 329–338. [[CrossRef](#)] [[PubMed](#)]
2. López, A.S.; Ramos, M.P.; Herrero, R.; Vilariño, J.M.L. Synthesis of magnetic green nanoparticle—Molecular imprinted polymers with emerging contaminants templates. *J. Environ. Chem. Eng.* **2020**, *8*, 103889. [[CrossRef](#)]
3. Hassan, M.S.; Bai, J.; Dou, D.-Q. Biopolymers; definition, classification and applications. *Egypt. J. Chem.* **2019**, *62*, 1725–1737. [[CrossRef](#)]
4. Rao, M.G.; Bharathi, P.; Akila, R.M. A comprehensive review on biopolymers. *Sci. Rev. Chem. Commun.* **2014**, *4*, 61–68.
5. Merlettini, A. Micro-Nanostructured Polymeric Materials with Specific Functionalities for Advanced Biomedical Applications. Ph.D. Thesis, Università di Bologna, Bologna, Italy, 2019.
6. Prajapati, S.K.; Jain, A.; Jain, A.; Jain, S. Biodegradable polymers and constructs: A novel approach in drug delivery. *Eur. Polym. J.* **2019**, *120*, 109191. [[CrossRef](#)]
7. Pushpamalar, J.; Veeramachineni, A.K.; Owh, C.; Loh, X.J. Biodegradable polysaccharides for controlled drug delivery. *ChemPlusChem* **2016**, *81*, 504–514. [[CrossRef](#)]
8. Song, R.; Murphy, M.; Li, C.; Ting, K.; Soo, C.; Zheng, Z. Current development of biodegradable polymeric materials for biomedical applications. *Drug Des. Devel Ther.* **2018**, *12*, 3117–3145. [[CrossRef](#)] [[PubMed](#)]
9. Rong, S.Y.; Mubarak, N.M.; Tanjung, F.A. Structure–property relationship of cellulose nanowhiskers reinforced chitosan biocomposite films. *J. Environ. Chem. Eng.* **2017**, *5*, 6132–6136. [[CrossRef](#)]
10. Aravamudhan, A.; Ramos, D.M.; Nada, A.A.; Kumbar, S.G. Chapter 4—Natural polymers. In *Natural and Synthetic Biomedical Polymers*, 1st ed.; Kumbar, S.G., Kaurencin, C.T., Deng, M., Eds.; Elsevier Science: Burlington, MA, USA, 2014; pp. 67–89.
11. Soltani, R.; Pishnamazi, M.; Pelalak, R.; Rezakazemi, M.; Marjani, A.; Dinari, M.; Sarkar, S.M.; Shirazian, S. Preparation of COOH-KCC-1/polyamide 6 composite by in situ ring-opening polymerization: Synthesis, characterization, and Cd(II) adsorption study. *J. Environ. Chem. Eng.* **2021**, *9*, 104683. [[CrossRef](#)]
12. Samrot, A.V.; Sean, T.C.; Kudaiyappan, T.; Bisvarah, U.; Mirarmandi, A.; Faradjeva, E.; Abubakar, A.; Ali, H.H.; Angalene, J.L.A.; Suresh Kumar, S. Production, characterization and application of nanocarriers made of polysaccharides, proteins, bio-polyesters and other biopolymers: A review. *Int. J. Biol. Macromol.* **2020**, *165*, 3088–3105. [[CrossRef](#)]
13. Manavitehrani, I.; Fathi, A.; Badr, H.; Daly, S.; Negahi Shirazi, A.; Dehghani, F. Biomedical applications of biodegradable polyesters. *Polymers* **2016**, *8*, 20. [[CrossRef](#)] [[PubMed](#)]
14. Tu, Y.; Peng, F.; André, A.A.M.; Men, Y.; Srinivas, M.; Wilson, D.A. Biodegradable hybrid stomatocyte nanomotors for drug delivery. *ACS Nano* **2017**, *11*, 1957–1963. [[CrossRef](#)] [[PubMed](#)]
15. Chen, C.K.; Huang, P.K.; Law, W.C.; Chu, C.H.; Chen, N.T.; Lo, L.W. Biodegradable polymers for gene-delivery applications. *Int. J. Nanomed.* **2020**, *15*, 2131–2150. [[CrossRef](#)] [[PubMed](#)]
16. Ilyas, R.A.; Sapuan, S.M.; Kadier, A.; Krishnan, S.; Atikah, M.S.N.; Ibrahim, R.; Nazrin, A.; Syafiq, R.; Misri, S.; Huzaifah, M.R.M.; et al. Chapter 7—Mechanical testing of sugar palm fiber reinforced sugar palm biopolymer composites. In *Advanced Processing, Properties, and Applications of Starch and Other Bio-Based Polymers*, 1st ed.; Al-Oqla, F.M., Sapuan, S.M., Eds.; Elsevier: Amsterdam, The Netherlands, 2020; pp. 89–110.
17. Abidi, N.; Cabrales, L.; Haigler, C.H. Changes in the cell wall and cellulose content of developing cotton fibers investigated by FTIR spectroscopy. *Carbohydr. Polym.* **2014**, *100*, 9–16. [[CrossRef](#)] [[PubMed](#)]
18. Ansari, M.; Alam, A.; Bera, R.; Hassan, A.; Goswami, S.; Das, N. Synthesis, characterization and adsorption studies of a novel triptycene based hydroxyl azo-nanoporous polymer for environmental remediation. *J. Environ. Chem. Eng.* **2020**, *8*, 103558. [[CrossRef](#)]
19. Eltz, F.Z.; Vebber, M.C.; Aguzzoli, C.; Machado, G.; da Silva Crespo, J.; Giovanela, M. Preparation, characterization and application of polymeric thin films containing silver and copper nanoparticles with bactericidal activity. *J. Environ. Chem. Eng.* **2020**, *8*, 103745. [[CrossRef](#)]
20. Mohammed, L.; Ansari, M.N.M.; Pua, G.; Jawaid, M.; Islam, M.S. A review on natural fiber reinforced polymer composite and its applications. *Int. J. Polym. Sci.* **2015**, *2015*, 243947. [[CrossRef](#)]
21. Udayakumar, G.P.; Kirthikaa, G.B.; Muthusamy, S.; Ramakrishnan, B.; Sivarajasekar, N. Comparison and evaluation of electrospun nanofiber membrane for the clarification of grape juice. In *Sustainable Development in Energy and Environment. Select Proceedings of ICSDEE 2019*; Sivasubramanian, V., Pugazhendhi, A., Moorthy, I.G., Eds.; Springer Proceedings in Energy (SPE) Book Series; Springer Nature: Singapore, 2020; pp. 77–92.
22. Al Battashi, H.; Al-Kindi, S.; Gupta, V.K.; Sivakumar, N. Polyhydroxyalkanoate (PHA) production using volatile fatty acids derived from the anaerobic digestion of waste paper. *J. Polym. Environ.* **2020**, *29*, 250–259. [[CrossRef](#)]
23. Amini, M.; Yousefi-Massumabad, H.; Younesi, H.; Abyar, H.; Bahramifar, N. Production of the polyhydroxyalkanoate biopolymer by *Cupriavidus necator* using beer brewery wastewater containing maltose as a primary carbon source. *J. Environ. Chem. Eng.* **2020**, *8*, 103588. [[CrossRef](#)]
24. Al-Battashi, H.; Annamalai, N.; Al-Kindi, S.; Nair, A.S.; Al-Bahry, S.; Verma, J.P.; Sivakumar, N. Production of bioplastic (poly-3-hydroxybutyrate) using waste paper as a feedstock: Optimization of enzymatic hydrolysis and fermentation employing *Burkholderia sacchari*. *J. Clean. Prod.* **2019**, *214*, 236–247. [[CrossRef](#)]

25. Udayakumar, G.P.; Muthusamy, S.; Selvaganesh, B.; Sivarajasekar, N.; Rambabu, K.; Banat, F.; Sivamani, S.; Sivakumar, N.; Hosseini-Bandegharaei, A.; Show, P.K. Biopolymers and composites: Properties, characterization and their applications in food, medical and pharmaceutical industries. *J. Environ. Chem. Eng.* **2021**, *9*, 105322. [[CrossRef](#)]
26. Francis, R.; Sasikumar, S.; Gopalan, G.P. Chapter 2—Synthesis, structure, and properties of biopolymers (natural and synthetic). In *Polymer Composite*, 1st ed.; Thomas, S., Joseph, K., Malhotra, S.K., Goda, K., Sreekala, M.S., Eds.; Wiley-VCH Verlag GmbH & Co. KGaA: Weinheim, Germany, 2013; Volume 3, pp. 11–107.
27. Jha, A.; Kumar, A. Biobased technologies for the efficient extraction of biopolymers from waste biomass. *Bioprocess Biosyst. Eng.* **2019**, *42*, 1893–1901. [[CrossRef](#)] [[PubMed](#)]
28. Baranwal, J.; Barse, B.; Fais, A.; Delogu, G.L.; Kumar, A. Biopolymer: A sustainable material for food and medical applications. *Polymers* **2022**, *14*, 983. [[CrossRef](#)] [[PubMed](#)]
29. Pothan, L.A.; Oommen, Z.; Thomas, S. Dynamic mechanical analysis of banana fiber reinforced polyester composites. *Compos. Sci. Technol.* **2003**, *63*, 283–293. [[CrossRef](#)]
30. Abotbina, W.; Sapuan, S.M.; Ilyas, R.A.; Sultan, M.T.H.; Alkbir, M.F.M.; Sulaiman, S.; Harussani, M.M.; Bayraktar, E. Recent developments in cassava (*Manihot esculenta*) based biocomposites and their potential industrial applications: A comprehensive review. *Materials* **2022**, *15*, 6992. [[CrossRef](#)] [[PubMed](#)]
31. Iliou, K.; Kikionis, S.; Ioannou, E.; Roussis, V. Marine biopolymers as bioactive functional ingredients of electrospun nanofibrous scaffolds for biomedical applications. *Mar. Drugs* **2022**, *20*, 314. [[CrossRef](#)] [[PubMed](#)]
32. Flaris, V.; Singh, G. Recent developments in biopolymers. *J. Vinyl Addit. Technol.* **2009**, *15*, 1–11. [[CrossRef](#)]
33. Devadas, V.V.; Khoo, K.S.; Chia, W.Y.; Chew, K.W.; Munawaroh, H.S.H.; Lam, M.K.; Lim, J.W.; Ho, Y.C.; Lee, K.T.; Show, P.L. Algae biopolymer towards sustainable circular economy. *Bioresour. Technol.* **2021**, *325*, 124702. [[CrossRef](#)] [[PubMed](#)]
34. Sharma, V.; Kundu, P.P. Addition polymers from natural oils—A review. *Prog. Polym. Sci.* **2006**, *31*, 983–1008. [[CrossRef](#)]
35. Awe, O.W.; Zhao, Y.; Nzihou, A.; Minh, D.P.; Lyczko, N. A review of biogas utilisation, purification and upgrading technologies. *Waste Biomass Valoriz.* **2017**, *8*, 267–283. [[CrossRef](#)]
36. Lelkes, P.I.; Har-El, Y.H.; Marcinkiewicz, C.; Lazarovic, P.; Baharlou, S.M.; Gerstenhaber, J.A. Soy-Derived Bioactive Peptides for Use in Compositions and Methods for Wound Healing, Tissue Engineering, and Regenerative Medicine. Patent WO 2017/015488 A1, 26 January 2017.
37. Othman, S.H. Bio-nanocomposite materials for food packaging applications: Types of biopolymer and nano-sized filler. *Agric. Agric. Sci. Procedia* **2014**, *2*, 296–303. [[CrossRef](#)]
38. Chaabouni, E.; Gassara, F.; Brar, S.K. Biopolymers synthesis and application. In *Biotransformation Waste Biomass into High Value Biochemicals*, 1st ed.; Brar, S.K., Dhillon, G.S., Soccol, C.R., Eds.; Springer: New York, NY, USA, 2014; pp. 415–443.
39. Atanase, L.I.; Desbrieres, J.; Riess, G. Micellization of synthetic and polysaccharides-based graft copolymers in aqueous media. *Prog. Polym. Sci.* **2017**, *73*, 32–60. [[CrossRef](#)]
40. Atanase, L.I. Micellar drug delivery systems based on natural biopolymers. *Polymers* **2021**, *13*, 477. [[CrossRef](#)] [[PubMed](#)]
41. Sakeer, K.; Ispas-Szabo, P.; Benyerbah, N.; Mateescu, M.A. Ampholytic starch excipients for high loaded drug formulations: Mechanistic insights. *Int. J. Pharm.* **2018**, *535*, 201–216. [[CrossRef](#)] [[PubMed](#)]
42. Kaur, L.; Singh, J.; Liu, Q. Chapter 5—Starch—A potential biomaterial for biomedical applications. In *Nanomaterials Nanosystems for Biomedical Applications*, 1st ed.; Mozafari, M.R., Ed.; Springer: Dordrecht, The Netherlands, 2007; pp. 83–98.
43. Lu, D.R.; Xiao, C.M.; Xu, S.J. Starch-based completely biodegradable polymer materials. *Express Polym. Lett.* **2009**, *3*, 366–375. [[CrossRef](#)]
44. Xiao, H.; Yang, T.; Lin, Q.; Liu, G.-Q.; Zhang, L.; Yu, F.; Chen, Y. Acetylated starch nanocrystals: Preparation and antitumor drug delivery study. *Int. J. Biol. Macromol.* **2016**, *89*, 456–464. [[CrossRef](#)] [[PubMed](#)]
45. Chen, F.; Huang, G.; Huang, H. Preparation and application of dextran and its derivatives as carriers. *Int. J. Biol. Macromol.* **2020**, *145*, 827–834. [[CrossRef](#)] [[PubMed](#)]
46. Banerjee, A.; Bandopadhyay, R. Use of dextran nanoparticle: A paradigm shift in bacterial exopolysaccharide based biomedical applications. *Int. J. Biol. Macromol.* **2016**, *87*, 295–301. [[CrossRef](#)] [[PubMed](#)]
47. Wang, C.; Xiong, S.; You, J.; Guan, W.; Xu, G.; Dou, H. Dextran-based coacervate nanodroplets as potential gene carriers for efficient cancer therapy. *Carbohydr. Polym.* **2020**, *231*, 115687.
48. Huang, S.; Huang, G. Design and application of dextran carrier. *J. Drug Deliv. Sci. Technol.* **2020**, *55*, 101392. [[CrossRef](#)]
49. Rakmai, J.; Cheirsilp, B. Continuous production of β -cyclodextrin by cyclodextrin glycosyltransferase immobilized in mixed gel beads: Comparative study in continuous stirred tank reactor and packed bed reactor. *Biochem. Eng. J.* **2016**, *105*, 107–113. [[CrossRef](#)]
50. Mejia-Ariza, R.; Graña-Suárez, L.; Verboom, W.; Huskens, J. Cyclodextrin-based supramolecular nanoparticles for biomedical applications. *J. Mater. Chem. B* **2017**, *5*, 36–52. [[CrossRef](#)] [[PubMed](#)]
51. Liu, W.; Liu, R.; Li, Y.; Kang, H.; Shen, D.; Wu, M.; Huang, Y. Self-assembly of ethyl cellulose-graft-polystyrene copolymers in acetone. *Polymer* **2009**, *50*, 211–217. [[CrossRef](#)]
52. Wang, D.; Tan, J.; Kang, H.; Ma, L.; Jin, X.; Liu, R.; Huang, Y. Synthesis, self-assembly and drug release behaviors of pH-responsive copolymers ethyl cellulose-graft-PDEAEMA through ATRP. *Carbohydr. Polym.* **2011**, *84*, 195–202. [[CrossRef](#)]
53. Kamel, R.; El-Wakil, N.A.; Dufresne, A.; Elkasabgy, N.A. Nanocellulose: From an agricultural waste to a valuable pharmaceutical ingredient. *Int. J. Biol. Macromol.* **2020**, *163*, 1579–1590. [[CrossRef](#)] [[PubMed](#)]

54. Merlusca, I.P.; Ibanescu, C.; Tuchilus, C.; Danu, M.; Atanase, L.I.; Popa, I.M. Characterization of neomycin-loaded xanthan-chitosan hydrogels for topical applications. *Cellul. Chem. Technol.* **2019**, *53*, 709–719. [[CrossRef](#)]
55. Rata, D.M.; Cadinoiu, A.N.; Popa, M.; Atanase, L.I.; Daraba, O.M.; Popescu, I.; Romila, L.E.; Ichim, D.L. Biocomposite hydrogels for the treatment of bacterial infections: Physicochemical characterization and in vitro assessment. *Pharmaceutics* **2021**, *13*, 2079. [[CrossRef](#)] [[PubMed](#)]
56. Ahsan, S.M.; Thomas, M.; Reddy, K.K.; Sooraparaju, S.G.; Asthana, A.; Bhatnagar, I. Chitosan as biomaterial in drug delivery and tissue engineering. *Int. J. Biol. Macromol.* **2018**, *110*, 97–109. [[CrossRef](#)] [[PubMed](#)]
57. Paul, P.; Kolesinska, B.; Sujka, W. Chitosan and its derivatives—Biomaterials with diverse biological activity for manifold applications. *Mini Rev. Med. Chem.* **2019**, *19*, 737–750. [[CrossRef](#)] [[PubMed](#)]
58. Ahmad, S.I.; Ahmad, R.; Khan, M.S.; Kant, R.; Shahid, S.; Gautam, L.; Hasan, G.M.; Hassan, M.I. Chitin and its derivatives: Structural properties and biomedical applications. *Int. J. Biol. Macromol.* **2020**, *164*, 526–539. [[CrossRef](#)] [[PubMed](#)]
59. Venkatesan, J.; Nithya, R.; Sudha, P.N.; Kim, S.-K. Role of alginate in bone tissue engineering. *Adv. Food Nutr. Res.* **2014**, *73*, 45–57. [[PubMed](#)]
60. Sosnik, A. Alginate particles as platform for drug delivery by the oral route: State-of-the-art. *ISRN Pharm.* **2014**, *2014*, 926157. [[CrossRef](#)] [[PubMed](#)]
61. Severino, P.; Oliveira, D.; Chen, M.; Souto, E.B. Chapter 13—Advanced applications of alginates in biomedical. In *Applications of Advanced Green Materials*, 1st ed.; Ahmed, S., Ed.; Woodhead Publishing in Materials; Woodhead Publishing—Elsevier: Kidlington, UK, 2021; pp. 321–337.
62. Mollah, M.Z.I.; Zahid, H.M.; Mahal, Z.; Faruque, M.R.I.; Khandaker, M.U. The usages and potential uses of alginate for healthcare applications. *Front. Mol. Biosci.* **2021**, *8*, 719972. [[CrossRef](#)] [[PubMed](#)]
63. Pokusaev, B.G.; Karlov, S.P.; Nekrasov, D.A.; Zakharov, N.S.; Khrantsov, D.P.; Reznik, V.V.; Vyazmin, A.V.; Shumova, N.V. Agarose gels with bioresorbable additives: The kinetics of the formation, structure, some properties. *Chem. Eng. Trans.* **2019**, *74*, 1171–1176.
64. Salati, M.A.; Khazai, J.; Tahmuri, A.M.; Samadi, A.; Taghizadeh, A.; Taghizadeh, M.; Zarrintaj, P.; Ramsey, J.D.; Habibzadeh, S.; Seidi, F.; et al. Agarose-based biomaterials: Opportunities and challenges in cartilage tissue engineering. *Polymers* **2020**, *12*, 1150. [[CrossRef](#)] [[PubMed](#)]
65. Zarrintaj, P.; Manouchehri, S.; Ahmadi, Z.; Saeb, M.R.; Urbanska, A.M.; Kaplan, D.L.; Mozafari, M. Agarose-based biomaterials for tissue engineering. *Carbohydr. Polym.* **2018**, *187*, 66–84. [[CrossRef](#)]
66. López-Marcial, G.R.; Zeng, A.Y.; Osuna, C.; Dennis, J.; García, J.M.; O’Connell, G.D. Agarose-based hydrogels as suitable bioprinting materials for tissue engineering. *ACS Biomater. Sci. Eng.* **2018**, *4*, 3610–3616. [[CrossRef](#)] [[PubMed](#)]
67. Liu, J.; Zhan, X.; Wan, J.; Wang, Y.; Wang, C. Review for carrageenan-based pharmaceutical biomaterials: Favourable physical features versus adverse biological effects. *Carbohydr. Polym.* **2015**, *121*, 27–36. [[CrossRef](#)]
68. Yegappan, R.; Selvaprithiviraj, V.; Amirthalingam, S.; Jayakumar, R. Carrageenan based hydrogels for drug delivery, tissue engineering and wound healing. *Carbohydr. Polym.* **2018**, *198*, 385–400. [[CrossRef](#)] [[PubMed](#)]
69. Qureshi, D.; Nayak, S.K.; Maji, S.; Kim, D.; Banerjee, I.; Pal, K. Carrageenan: A wonder polymer from marine algae for potential drug delivery applications. *Curr. Pharm. Des.* **2019**, *25*, 1172–1186. [[CrossRef](#)] [[PubMed](#)]
70. Rode, M.P.; Batti Angulski, A.B.; Gomes, F.A.; da Silva, M.M.; Jeremias, T.D.S.; de Carvalho, R.G.; Lucif Vieira, D.G.; Oliveira, L.F.C.; Fernandes Maia, L.; Trentin, A.G.; et al. Carrageenan hydrogel as a scaffold for skin-derived multipotent stromal cells delivery. *J. Biomater. Appl.* **2018**, *33*, 422–434. [[CrossRef](#)] [[PubMed](#)]
71. Munarin, F.; Tanzi, M.C.; Petrini, P. Advances in biomedical applications of pectin gels. *Int. J. Biol. Macromol.* **2012**, *51*, 681–689. [[CrossRef](#)] [[PubMed](#)]
72. Alkarib, S.Y.; MohamedElhassan, D.E.; Nur, A.O. Evaluation of gum Arabic as a film coating former for immediate release oral tablet formulation. *World J. Pharm. Pharmaceut. Sci.* **2016**, *5*, 32–41.
73. Li, M.; Li, H.; Li, X.; Zhu, H.; Xu, Z.; Liu, L.; Ma, J.; Zhang, M. A bioinspired alginate–gum Arabic hydrogel with micro-/nanoscale structures for controlled drug release in chronic wound healing. *ACS Appl. Mater. Interfaces* **2017**, *9*, 22160–22175. [[CrossRef](#)] [[PubMed](#)]
74. Gamal-Eldeen, A.M.; Moustafa, D.; El-Daly, S.M.; El-Hussieny, E.A.; Saleh, S.; Khoobchandani, M.; Bacon, K.L.; Gupta, S.; Katti, K.; Shukla, R.; et al. Photothermal therapy mediated by gum Arabic-conjugated gold nanoparticles suppresses liver preneoplastic lesions in mice. *J. Photochem. Photobiol. B* **2016**, *163*, 47–56. [[CrossRef](#)] [[PubMed](#)]
75. Gerola, A.P.; Rubira, A.F.; Muniz, E.C.; Valente, A.J.M. Gum Arabic: A remarkable biopolymer for food and biomedical applications. In *Engineering Technology and Industrial Chemistry with Applications*, 1st ed.; Haghi, R.K., Torrens, F., Eds.; Apple Academic Press: New York, NY, USA, 2019; pp. 281–312.
76. Seeli, D.S.; Prabakaran, M. Guar gum oleate-graft-poly(methacrylic acid) hydrogel as a colon-specific controlled drug delivery carrier. *Carbohydr. Polym.* **2017**, *158*, 51–57. [[CrossRef](#)]
77. Chen, F.; Yang, Y.; He, J.; Bu, T.; He, X.; He, K.; Xiang, C.; Yu, Z.; Wu, H. The gelation of hydroxypropyl guar gum by nano-ZrO₂. *Polym. Adv. Technol.* **2018**, *29*, 587–593. [[CrossRef](#)]
78. George, A.; Shah, P.A.; Shrivastav, P.S. Guar gum: Versatile natural polymer for drug delivery applications. *Eur. Polym. J.* **2019**, *112*, 722–735. [[CrossRef](#)]
79. Amjed, N.; Zeshan, M.; Farooq, A.; Naz, S. Applications of guar gum polysaccharide for pharmaceutical drug delivery: A review. *Int. J. Biol. Macromol.* **2024**, *257*, 128390. [[CrossRef](#)] [[PubMed](#)]

80. Gupta, N.; Jangid, A.K.; Pooja, D.; Kulhari, H. Inulin: A novel and stretchy polysaccharide tool for biomedical and nutritional applications. *Int. J. Biol. Macromol.* **2019**, *132*, 852–863. [[CrossRef](#)]
81. Huang, G.; Huang, H. Hyaluronic acid-based biopharmaceutical delivery and tumor-targeted drug delivery system. *J. Control. Release* **2018**, *278*, 122–126. [[CrossRef](#)] [[PubMed](#)]
82. Tiwari, S.; Bahadur, P. Modified hyaluronic acid based materials for biomedical applications. *Int. J. Biol. Macromol.* **2019**, *121*, 556–571. [[CrossRef](#)] [[PubMed](#)]
83. Li, C.; Cao, Z.; Li, W.; Liu, R.; Chen, Y.; Song, Y.; Liu, G.; Song, Z.; Liu, Z.; Lu, C.; et al. A review on the wide range applications of hyaluronic acid as a promising rejuvenating biomacromolecule in the treatments of bone related diseases. *Int. J. Biol. Macromol.* **2020**, *165*, 1264–1275. [[CrossRef](#)] [[PubMed](#)]
84. Vasvani, S.; Kulkarni, P.; Rawtani, D. Hyaluronic acid: A review on its biology, aspects of drug delivery, route of administrations and a special emphasis on its approved marketed products and recent clinical studies. *Int. J. Biol. Macromol.* **2020**, *151*, 1012–1029. [[CrossRef](#)] [[PubMed](#)]
85. Zhai, P.; Peng, X.; Li, B.; Liu, Y.; Sun, H.; Li, X. The application of hyaluronic acid in bone regeneration. *Int. J. Biol. Macromol.* **2020**, *151*, 1224–1239. [[CrossRef](#)] [[PubMed](#)]
86. Khan, A.R.; Yang, X.; Du, X.; Yang, H.; Liu, Y.; Khan, A.Q.; Zhai, G. Chondroitin sulfate derived theranostic and therapeutic nanocarriers for tumor-targeted drug delivery. *Carbohydr. Polym.* **2020**, *233*, 115837. [[CrossRef](#)] [[PubMed](#)]
87. Wang, L.; Wang, X.; Wu, H.; Liu, R. Overview on biological activities and molecular characteristics of sulfated polysaccharides from marine green algae in recent years. *Mar. Drugs* **2014**, *12*, 4984–5020. [[CrossRef](#)] [[PubMed](#)]
88. Varghese, O.P.; Liu, J.; Sundaram, K.; Hilborn, J.; Oommen, O.P. Chondroitin sulfate derived theranostic nanoparticles for targeted drug delivery. *Biomater. Sci.* **2016**, *4*, 1310–1313. [[CrossRef](#)] [[PubMed](#)]
89. Kwon, H.J.; Han, Y. Chondroitin sulfate-based biomaterials for tissue engineering. *Turk. J. Biol.* **2016**, *40*, 290–299. [[CrossRef](#)]
90. Pal, D.; Saha, S. Chondroitin: A natural biomarker with immense biomedical applications. *RSC Adv.* **2019**, *9*, 28061–28077. [[CrossRef](#)] [[PubMed](#)]
91. Venkatesan, J.; Anil, S.; Rao, S.; Bhatnagar, I.; Kim, S.-K. Sulfated polysaccharides from macroalgae for bone tissue regeneration. *Curr. Pharm. Des.* **2019**, *25*, 1200–1209. [[CrossRef](#)] [[PubMed](#)]
92. Terzi, A.; Gallo, N.; Bettini, S.; Sibillano, T.; Altamura, D.; Madaghiele, M.; De Caro, L.; Valli, L.; Salvatore, L.; Sannino, A.; et al. Sub- and supramolecular X-ray characterization of engineered tissues from equine tendon, bovine dermis, and fish skin type-I collagen. *Macromol. Biosci.* **2020**, *20*, e2000017. [[CrossRef](#)] [[PubMed](#)]
93. Sorushanova, A.; Delgado, L.M.; Wu, Z.; Shologu, N.; Kshirsagar, A.; Raghunath, R.; Mullen, A.M.; Bayon, Y.; Pandit, A.; Raghunath, M.; et al. The collagen suprafamily: From biosynthesis to advanced biomaterial development. *Adv. Mater.* **2019**, *31*, e1801651. [[CrossRef](#)] [[PubMed](#)]
94. Sharma, S.; Rai, V.K.; Narang, R.K.; Markandeywar, T.S. Collagen-based formulations for wound healing: A literature review. *Life Sci.* **2022**, *290*, 120096. [[CrossRef](#)] [[PubMed](#)]
95. Geahchan, S.; Baharlouei, P.; Rahman, A. Marine collagen: A promising biomaterial for wound healing, skin anti-aging, and bone regeneration. *Mar. Drugs.* **2022**, *20*, 61. [[CrossRef](#)] [[PubMed](#)]
96. Martínez-Puig, D.; Costa-Larrión, E.; Rubio-Rodríguez, N.; Gálvez-Martín, P. Collagen supplementation for joint health: The link between composition and scientific knowledge. *Nutrients* **2023**, *15*, 1332. [[CrossRef](#)] [[PubMed](#)]
97. Davison-Kotler, E.; Marshall, W.S.; García-Gareta, E. Sources of collagen for biomaterials in skin wound healing. *Bioengineering* **2019**, *6*, 56. [[CrossRef](#)] [[PubMed](#)]
98. Bax, D.V.; Smalley, H.E.; Farndale, R.W.; Best, S.M.; Cameron, R.E. Cellular response to collagen–elastin composite materials. *Acta Biomater.* **2019**, *86*, 158–170. [[CrossRef](#)] [[PubMed](#)]
99. Marin, M.M.; Ianchis, R.; Leu Alexa, R.; Gifu, I.C.; Kaya, M.G.A.; Savu, D.I.; Popescu, R.C.; Alexandrescu, E.; Ninciuleanu, C.M.; Preda, S.; et al. Development of new collagen/clay composite biomaterials. *Int. J. Mol. Sci.* **2021**, *23*, 401. [[CrossRef](#)] [[PubMed](#)]
100. Kaczmarek-Szczepańska, B.; Polkowska, I.; Małek, M.; Kluczyński, J.; Paździor-Czapula, K.; Wekwejt, M.; Michno, A.; Ronowska, A.; Pałubicka, A.; Nowicka, B.; et al. The characterization of collagen-based scaffolds modified with phenolic acids for tissue engineering application. *Sci. Rep.* **2023**, *13*, 9966. [[CrossRef](#)] [[PubMed](#)]
101. Meng, D.; Lei, X.; Li, Y.; Kong, Y.; Huang, D.; Zhang, G. Three-dimensional polyvinyl alcohol scaffolds modified with collagen for HepG2 cell culture. *J. Biomater. Appl.* **2020**, *35*, 459–470. [[CrossRef](#)] [[PubMed](#)]
102. Parenteau-Bareil, R.; Gauvin, R.; Berthod, F. Collagen-based biomaterials for tissue engineering applications. *Materials* **2010**, *3*, 1863–1887. [[CrossRef](#)]
103. Ferreira, A.M.; Gentile, P.; Chiono, V.; Ciardelli, G. Collagen for bone tissue regeneration. *Acta Biomater.* **2012**, *8*, 3191–3200. [[CrossRef](#)] [[PubMed](#)]
104. Chattopadhyay, S.; Raines, R.T. Collagen-based biomaterials for wound healing. *Biopolymers* **2014**, *101*, 821–833. [[CrossRef](#)] [[PubMed](#)]
105. Khan, R.; Khan, M.H. Use of collagen as a biomaterial: An update. *J. Indian Soc. Periodontol.* **2013**, *17*, 539–542. [[CrossRef](#)] [[PubMed](#)]
106. Sahiner, M.; Alpaslan, D.; Bitlisli, B.O. Collagen-based hydrogel films as drug-delivery devices with antimicrobial properties. *Polym. Bull.* **2014**, *71*, 3017–3033. [[CrossRef](#)]

107. Hoque, M.E.; Nuge, T.; Yeow, T.K.; Nordin, N.; Prasad, R.G.S.V. Gelatin based scaffolds for tissue engineering—A review. *Polym. Res. J.* **2015**, *9*, 15–32.
108. Saraogi, G.K.; Gupta, P.; Gupta, U.D.; Jain, N.K.; Agrawal, G.P. Gelatin nanocarriers as potential vectors for effective management of tuberculosis. *Int. J. Pharm.* **2010**, *385*, 143–149. [[CrossRef](#)] [[PubMed](#)]
109. Rose, J.B.; Pacelli, S.; El Haj, A.J.; Dua, H.S.; Hopkinson, A.; White, L.J.; Rose, F.R.A.J. Gelatin-based materials in ocular tissue engineering. *Materials* **2014**, *7*, 3106–3135. [[CrossRef](#)] [[PubMed](#)]
110. Yasmin, R.; Shah, M.; Khan, S.A.; Ali, R. Gelatin nanoparticles: A potential candidate for medical applications. *Nanotechnol. Rev.* **2017**, *6*, 191–207. [[CrossRef](#)]
111. Foox, M.; Zilberman, M. Drug delivery from gelatin-based systems. *Expert Opin. Drug Deliv.* **2015**, *12*, 1547–1563. [[CrossRef](#)] [[PubMed](#)]
112. Nouri-Felekor, M.; Khakbiz, M.; Nezafati, N.; Mohammadi, J.; Eslaminejad, M.B. Comparative analysis and properties evaluation of gelatin microspheres crosslinked with glutaraldehyde and 3-glycidoxypopyltrimethoxysilane as drug delivery systems for the antibiotic vancomycin. *Int. J. Pharm.* **2019**, *557*, 208–220. [[CrossRef](#)] [[PubMed](#)]
113. Campiglio, C.E.; Contessi Negrini, N.; Farè, S.; Draghi, L. Cross-linking strategies for electrospun gelatin scaffolds. *Materials* **2019**, *12*, 2476. [[CrossRef](#)] [[PubMed](#)]
114. De Colli, M.; Massimi, M.; Barbetta, A.; Di Rosario, B.L.; Nardecchia, S.; Conti Devirgiliis, L.; Dentini, M. A biomimetic porous hydrogel of gelatin and glycosaminoglycans cross-linked with transglutaminase and its application in the culture of hepatocytes. *Biomed Mater.* **2012**, *7*, 055005. [[CrossRef](#)] [[PubMed](#)]
115. Han, J.; Lazarovici, P.; Pomerantz, C.; Chen, X.; Wei, Y.; Lelkes, P.I. Co-electrospun blends of PLGA, gelatin, and elastin as potential nonthrombogenic scaffolds for vascular tissue engineering. *Biomacromolecules* **2011**, *12*, 399–408. [[CrossRef](#)]
116. Gonçalves, A.S.C.; Rodrigues, C.F.; Fernandes, N.; de Melo-Diogo, D.; Ferreira, P.; Moreira, A.F.; Correia, I.J. IR780 loaded gelatin-PEG coated gold core silica shell nanorods for cancer-targeted photothermal/photodynamic therapy. *Biotechnol. Bioeng.* **2022**, *119*, 644–656. [[CrossRef](#)] [[PubMed](#)]
117. Ranganathan, S.; Balagangadharan, K.; Selvamurugan, N. Chitosan and gelatin-based electrospun fibers for bone tissue engineering. *Int. J. Biol. Macromol.* **2019**, *133*, 354–364. [[CrossRef](#)]
118. Nguyen, T.P.; Nguyen, Q.V.; Nguyen, V.-H.; Le, T.-H.; Huynh, V.Q.N.; Vo, D.-V.N.; Trinh, Q.T.; Kim, S.Y.; Le, Q.V. Silk fibroin-based biomaterials for biomedical applications: A review. *Polymers* **2019**, *11*, 1933. [[CrossRef](#)] [[PubMed](#)]
119. Gianak, O.; Kyzas, G.Z.; Samanidou, V.F.; Deliyanni, E.A. A review for the synthesis of silk fibroin nanoparticles with different techniques and their ability to be used for drug delivery. *Curr. Anal. Chem.* **2019**, *15*, 339–348. [[CrossRef](#)]
120. Pham, D.T.; Saelim, N.; Tiyaboonchai, W. Paclitaxel loaded EDC-crosslinked fibroin nanoparticles: A potential approach for colon cancer treatment. *Drug Deliv. Transl. Res.* **2020**, *10*, 413–424. [[CrossRef](#)] [[PubMed](#)]
121. Totten, J.D.; Wongpinyochit, T.; Carrola, J.; Duarte, I.F.; Seib, F.P. PEGylation-dependent metabolic rewiring of macrophages with silk fibroin nanoparticles. *ACS Appl. Mater. Interfaces* **2019**, *11*, 14515–14525. [[CrossRef](#)] [[PubMed](#)]
122. Crivelli, B.; Bari, E.; Perteghella, S.; Catenacci, L.; Sorrenti, M.; Mocchi, M.; Faragò, S.; Tripodo, G.; Prina-Mello, A.; Torre, M.L. Silk fibroin nanoparticles for celecoxib and curcumin delivery: ROS-scavenging and anti-inflammatory activities in an in vitro model of osteoarthritis. *Eur. J. Pharm. Biopharm.* **2019**, *137*, 37–45. [[CrossRef](#)] [[PubMed](#)]
123. Aiyelabegan, H.T.; Zaidi, S.S.Z.; Fanuel, S.; Eatemadi, A.; Ebadi, M.T.K.; Sadroddiny, E. Albumin-based biomaterial for lung tissue engineering applications. *Int. J. Polym. Mater. Polym. Biomater.* **2016**, *65*, 853–861. [[CrossRef](#)]
124. Tao, C.; Chuah, Y.J.; Xu, C.; Wang, D.-A. Albumin conjugates and assemblies as versatile bio-functional additives and carriers for biomedical applications. *J. Mater. Chem. B* **2019**, *7*, 357–367. [[CrossRef](#)] [[PubMed](#)]
125. Roufegarnejad, L.; Jahanban-Esfahlan, A.; Sajed-Amin, S.; Panahi-Azar, V.; Tabibiazar, M. Molecular interactions of thymol with bovine serum albumin: Spectroscopic and molecular docking studies. *J. Mol. Recognit.* **2018**, *31*, e2704. [[CrossRef](#)] [[PubMed](#)]
126. Andishmand, H.; Roufegari-Nejad, L.; Tabibiazar, M. Nanostructure characterization of bovine serum albumin–resveratrol complex. *Res. Innov. Food Sci. Technol.* **2017**, *6*, 291–300.
127. Ghorbani, M.; Hamishehkar, H.; Tabibiazar, M. BSA/chitosan polyelectrolyte complex: A platform for enhancing the loading and cancer cell-uptake of resveratrol. *Macromol. Res.* **2018**, *26*, 808–813. [[CrossRef](#)]
128. Jahanban-Esfahlan, A.; Ostadrahimi, A.; Jahanban-Esfahlan, R.; Roufegarnejad, L.; Tabibiazar, M.; Amarowicz, R. Recent developments in the detection of bovine serum albumin. *Int. J. Biol. Macromol.* **2019**, *138*, 602–617. [[CrossRef](#)] [[PubMed](#)]
129. Jahanban-Esfahlan, A.; Panahi-Azar, V.; Sajedi, S. Spectroscopic and molecular docking studies on the interaction between N-acetyl cysteine and bovine serum albumin. *Biopolymers* **2015**, *103*, 638–645. [[CrossRef](#)] [[PubMed](#)]
130. Jahanban-Esfahlan, A.; Davaran, S.; Moosavi-Movahedi, A.A.; Dastmalchi, S. Investigating the interaction of juglone (5-hydroxy-1,4-naphthoquinone) with serum albumins using spectroscopic and in silico methods. *J. Iran. Chem. Soc.* **2017**, *14*, 1527–1540. [[CrossRef](#)]
131. Elzoghby, A.O.; Samy, W.M.; Elgindy, N.A. Albumin-based nanoparticles as potential controlled release drug delivery systems. *J. Control. Release* **2012**, *157*, 168–182. [[CrossRef](#)] [[PubMed](#)]
132. Jahanban-Esfahlan, A.; Dastmalchi, S.; Davaran, S. A simple improved desolvation method for the rapid preparation of albumin nanoparticles. *Int. J. Biol. Macromol.* **2016**, *91*, 703–709. [[CrossRef](#)] [[PubMed](#)]
133. Roufegarnejad, L.; Amarowicz, R.; Jahanban-Esfahlan, A. Characterizing the interaction between pyrogallol and human serum albumin by spectroscopic and molecular docking methods. *J. Biomol. Struct. Dyn.* **2019**, *37*, 2766–2775. [[CrossRef](#)] [[PubMed](#)]

134. Whitcombe, M.J.; Chianella, I.; Larcombe, L.; Piletsky, S.A.; Noble, J.; Porter, R.; Horgan, A. The rational development of molecularly imprinted polymer-based sensors for protein detection. *Chem. Soc. Rev.* **2011**, *40*, 1547–1571. [[CrossRef](#)] [[PubMed](#)]
135. Jahanban-Esfahlan, A.; Roufegarinejad, L.; Jahanban-Esfahlan, R.; Tabibiazar, M.; Amarowicz, R. Latest developments in the detection and separation of bovine serum albumin using molecularly imprinted polymers. *Talanta* **2020**, *207*, 120317. [[CrossRef](#)]
136. Gao, R.; Kong, X.; Wang, X.; He, X.; Chen, L.; Zhang, Y. Preparation and characterization of uniformly sized molecularly imprinted polymers functionalized with core-shell magnetic nanoparticles for the recognition and enrichment of protein. *J. Mater. Chem.* **2011**, *21*, 17863–17871. [[CrossRef](#)]
137. Ansari, S. Application of magnetic molecularly imprinted polymer as a versatile and highly selective tool in food and environmental analysis: Recent developments and trends. *TrAC Trends Anal. Chem.* **2017**, *90*, 89–106. [[CrossRef](#)]
138. Sood, A.; Arora, V.; Shah, J.; Kotnala, R.K.; Jain, T.K. Multifunctional gold-coated iron oxide core-shell nanoparticles stabilized using thiolated sodium alginate for biomedical applications. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2017**, *80*, 274–281. [[CrossRef](#)]
139. Guoning, C.; Pengqi, G.; Yan, W.; Lu, W.; Hua, S.; Yunzhe, L.; Wanghui, J.; Chun, C.; Qiang, F. Preparation of molecularly imprinted polymers and application in a biomimetic biotin-avidin-ELISA for the detection of bovine serum albumin. *Talanta* **2019**, *198*, 55–62. [[CrossRef](#)] [[PubMed](#)]
140. Muthusankar, E.; Ragupathy, D. Chitosan based nanocomposite biosensors: A recent review. *Sens. Lett.* **2018**, *16*, 81–91. [[CrossRef](#)]
141. Yang, Z.; Xu, J.; Wang, J.; Zhang, Q.; Zhang, B. Design and preparation of self-driven BSA surface imprinted tubular carbon nanofibers and their specific adsorption performance. *Chem. Eng. J.* **2019**, *373*, 923–934. [[CrossRef](#)]
142. Xia, J.; Cao, X.; Wang, Z.; Yang, M.; Zhang, F.; Lu, B.; Li, F.; Xia, L.; Li, Y.; Xia, Y. Molecularly imprinted electrochemical biosensor based on chitosan/ionic liquid-graphene composites modified electrode for determination of bovine serum albumin. *Sens. Actuators B Chem.* **2016**, *225*, 305–311. [[CrossRef](#)]
143. Qi, M.; Zhao, K.; Bao, Q.; Pan, P.; Zhao, Y.; Yang, Z.; Wang, H.; Wei, J. Adsorption and electrochemical detection of bovine serum albumin imprinted calcium alginate hydrogel membrane. *Polymers* **2019**, *11*, 622. [[CrossRef](#)] [[PubMed](#)]
144. Liu, D.; Zhao, K.; Qi, M.; Li, S.; Xu, G.; Wei, J.; He, X. Preparation of protein molecular imprinted polysiloxane membrane using calcium alginate film as matrix and its application for cell culture. *Polymers* **2018**, *10*, 170. [[CrossRef](#)] [[PubMed](#)]
145. Zhou, J.; Wang, Y.; Ma, Y.; Zhang, B.; Zhang, Q. Surface molecularly imprinted thermo-sensitive polymers based on light-weight hollow magnetic microspheres for specific recognition of BSA. *Appl. Surf. Sci.* **2019**, *486*, 265–273. [[CrossRef](#)]
146. Saha, T.; Hoque, M.E.; Mahbub, T. Chapter 13—Biopolymers for sustainable packaging in food, cosmetics, and pharmaceuticals. In *Advanced Processing, Properties, and Applications of Starch and Other Bio-Based Polymers*, 1st ed.; Al-Oqla, F.M., Sapuan, S.M., Eds.; Elsevier: Amsterdam, The Netherlands, 2020; pp. 197–214.
147. Tortorella, S.; Maturi, M.; Vetri Buratti, V.; Vozzolo, G.; Locatelli, E.; Sambri, L.; Franchini, M.C. Zein as a versatile biopolymer: Different shapes for different biomedical applications. *RSC Adv.* **2021**, *62*, 39004–39026. [[CrossRef](#)]
148. Tivano, F.; Chiono, V. Zein as a renewable material for the preparation of green nanoparticles for drug delivery. *Front. Biomater. Sci.* **2023**, *2*, 1156403. [[CrossRef](#)]
149. Voci, S.; Gagliardi, A.; Fresta, M.; Cosco, D. Antitumor features of vegetal protein-based nanotherapeutics. *Pharmaceutics* **2020**, *12*, 65. [[CrossRef](#)]
150. Fazal, T.; Murtaza, B.N.; Shah, M.; Iqbal, S.; Rehman, M.U.; Jaber, F.; Dera, A.A.; Awwad, N.S.; Ibrahim, H.A. Recent developments in natural biopolymer based drug delivery systems. *RSC Adv.* **2023**, *13*, 23087–23121. [[CrossRef](#)] [[PubMed](#)]
151. Marjanović-Balaban, Ž.; Gojković Cvjetković, V.; Grujić, R. Gliadin proteins from wheat flour: The optimal determination conditions by ELISA. *Foods Raw Mater.* **2021**, *9*, 364–370. [[CrossRef](#)]
152. Hong, S.; Choi, D.W.; Kim, H.N.; Park, C.G.; Lee, W.; Park, H.H. Protein-based nanoparticles as drug delivery systems. *Pharmaceutics* **2020**, *12*, 604. [[CrossRef](#)] [[PubMed](#)]
153. Prakash, S.; Manish; Bansal, P.; Kumar, A.; Saxena, V.; Kumar, V.; Katiyar, D. Overview and *in-silico* pharmacological profiling of Gliadin: A potential biomaterial. *Mater. Today Proc.* **2022**, *62*, 276–282. [[CrossRef](#)]
154. Jain, A.; Cheng, K. The principles and applications of avidin-based nanoparticles in drug delivery and diagnosis. *J. Control. Release* **2017**, *245*, 27–40. [[CrossRef](#)] [[PubMed](#)]
155. Frangville, C.; Rutkevičius, M.; Richter, A.P.; Velev, O.D.; Stoyanov, S.D.; Paunov, V.N. Fabrication of environmentally biodegradable lignin nanoparticles. *ChemPhysChem* **2012**, *13*, 4235–4243. [[CrossRef](#)] [[PubMed](#)]
156. Dai, L.; Liu, R.; Hu, L.-Q.; Zou, Z.-F.; Si, C.-L. Lignin nanoparticle as a novel green carrier for the efficient delivery of resveratrol. *ACS Sustain. Chem. Eng.* **2017**, *5*, 8241–8249. [[CrossRef](#)]
157. Terzioğlu, P.; Parin, F.N.; Sicak, Y. Lignin composites for biomedical applications: Status, challenges and perspectives. In *Lignin: Biosynthesis and Transformation for Industrial Applications*, 1st ed.; Sharma, S., Kumar, A., Eds.; Springer Series on Polymer and Composite Materials; Springer: Cham, Switzerland, 2020; pp. 253–273.
158. Iravani, S. Biomedical applications of lignin-based nanoparticles. In *Nanoparticles and their Biomedical Applications*, 1st ed.; Shukla, A.K., Ed.; Springer Nature: Singapore, 2020; pp. 217–224.
159. Mohammad, S.H.; Bhukya, B. Biotransformation of toxic lignin and aromatic compounds of lignocellulosic feedstock into eco-friendly biopolymers by *Pseudomonas putida* KT2440. *Bioresour. Technol.* **2022**, *363*, 128001. [[CrossRef](#)] [[PubMed](#)]
160. Macarie, C.A.; Segneanu, A.E.; Balcu, I.; Pop, R.; Burtica, G.; Ungurean, M.; Grozescu, I. Use of alkaline lyophilization process for lignocellulosic biomass pretreatment. *Dig. J. Nanomater. Biostruct.* **2012**, *7*, 1577–1586.

161. Balcu, I.; Segneanu, A.E.; Mirica, M.C.; Iorga, M.I.; Macarie, C.; Martagiu, R. Microwaves effect over biomass hydrolysis. *Environ. Eng. Manag. J.* **2009**, *8*, 741–746.
162. Sugiarto, S.; Leow, Y.; Tan, C.L.; Wang, G.; Kai, D. How far is Lignin from being a biomedical material? *Bioact. Mater.* **2021**, *8*, 71–94. [[CrossRef](#)] [[PubMed](#)]
163. Thombare, N.; Kumar, S.; Kumari, U.; Sakare, P.; Yogi, R.K.; Prasad, N.; Sharma, K.K. Shellac as a multifunctional biopolymer: A review on properties, applications and future potential. *Int. J. Biol. Macromol.* **2022**, *215*, 203–223. [[CrossRef](#)] [[PubMed](#)]
164. Lin, M.; Dai, Y.; Xia, F.; Zhang, X. Advances in non-covalent crosslinked polymer micelles for biomedical applications. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2021**, *119*, 111626. [[CrossRef](#)] [[PubMed](#)]
165. Tincu Iurciuc, C.E.; Bouhadiba, B.; Atanase, L.I.; Stan, C.S.; Popa, M.; Ochiuz, L. An accessible method to improve the stability and reusability of porcine pancreatic α -amylase via immobilization in gellan-based hydrogel particles obtained by ionic cross-linking with Mg^{2+} ions. *Molecules* **2023**, *28*, 4695. [[CrossRef](#)] [[PubMed](#)]
166. Iurciuc-Tincu, C.E.; Atanase, L.I.; Ochiuz, L.; Jérôme, C.; Sol, V.; Martin, P.; Popa, M. Curcumin-loaded polysaccharides-based complex particles obtained by polyelectrolyte complexation and ionic gelation. I—Particles obtaining and characterization. *Int. J. Biol. Macromol.* **2020**, *147*, 629–642. [[CrossRef](#)] [[PubMed](#)]
167. Suflet, D.M.; Popescu, I.; Pelin, I.M.; Ichim, D.L.; Daraba, O.M.; Constantin, M.; Fundueanu, G. Dual cross-linked chitosan/PVA hydrogels containing silver nanoparticles with antimicrobial properties. *Pharmaceutics* **2021**, *13*, 1461. [[CrossRef](#)] [[PubMed](#)]
168. Dellali, K.Z.; Dellali, M.; Rață, D.M.; Cadinoiu, A.N.; Atanase, L.I.; Popa, M.; Spataru, M.C.; Solcan, C. Assessment of physico-chemical and in vivo biological properties of polymeric nanocapsules based on chitosan and poly(*N*-vinyl pyrrolidone-*alt*-itaconic anhydride). *Polymers* **2022**, *14*, 1811. [[CrossRef](#)] [[PubMed](#)]
169. Tian, H.; Tang, Z.; Zhuang, X.; Chen, X.; Jing, X. Biodegradable synthetic polymers: Preparation functionalization and biomedical application. *Prog. Polym. Sci.* **2012**, *37*, 237–280. [[CrossRef](#)]
170. Rossi, F.; van Griensven, M. Polymer functionalization as a powerful tool to improve scaffold performances. *Tissue Eng. Part A* **2014**, *20*, 2043–2051. [[CrossRef](#)] [[PubMed](#)]
171. Xiao, Y.; Chinoy, Z.S.; Pecastaings, G.; Bathany, K.; Garanger, E.; Lecommandoux, S. Design of polysaccharide-*b*-elastin-like polypeptide bioconjugates and their thermoresponsive self-assembly. *Biomacromolecules* **2020**, *21*, 114–125. [[CrossRef](#)] [[PubMed](#)]
172. Thakor, P.; Bhavana, V.; Sharma, R.; Srivastava, S.; Singh, S.B.; Mehra, N.K. Polymer–drug conjugates: Recent advances and future perspectives. *Drug Discov. Today* **2020**, *25*, 1718–1726. [[CrossRef](#)] [[PubMed](#)]
173. Zhou, T.; Zhu, Y.; Li, X.; Liu, X.; Yeung, K.W.K.; Wu, S.; Wang, X.; Cui, Z.; Yang, X.; Chu, P.K. Surface functionalization of biomaterials by radical polymerization. *Prog. Mater. Sci.* **2016**, *83*, 191–235. [[CrossRef](#)]
174. Schatz, C.; Lecommandoux, S. Polysaccharide-containing block copolymers: Synthesis, properties and applications of an emerging family of glycoconjugates. *Macromol. Rapid Commun.* **2010**, *31*, 1664–1684. [[CrossRef](#)] [[PubMed](#)]
175. Lohani, A.; Singh, G.; Bhattacharya, S.S.; Verma, A. Interpenetrating polymer networks as innovative drug delivery systems. *J. Drug Deliv.* **2014**, *2014*, 583612. [[CrossRef](#)] [[PubMed](#)]
176. Farooq, U.; Teuwen, J.; Dransfeld, C. Toughening of epoxy systems with interpenetrating polymer network (IPN): A review. *Polymers* **2020**, *12*, 1908. [[CrossRef](#)]
177. Zoratto, N.; Matricardi, P. Chapter 4—Semi-IPNs and IPN-based hydrogels. In *Polymeric Gels: Characterization, Properties and Biomedical Applications*, 1st ed.; Pal, K., Banerjee, I., Eds.; Woodhead Publishing Series in Biomaterials; Woodhead Publishing—Elsevier: Kidlington, UK, 2018; pp. 91–124.
178. Ganguly, S.; Maity, P.P.; Mondal, S.; Das, P.; Bhawal, P.; Dhara, S.; Das, N.C. Polysaccharide and poly(methacrylic acid) based biodegradable elastomeric biocompatible semi-IPN hydrogel for controlled drug delivery. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2018**, *92*, 34–51. [[CrossRef](#)] [[PubMed](#)]
179. Zou, Z.; Zhang, B.; Nie, X.; Cheng, Y.; Hu, Z.; Liao, M.; Li, S. A sodium alginate-based sustained-release IPN hydrogel and its applications. *RSC Adv.* **2020**, *10*, 39722–39730. [[CrossRef](#)] [[PubMed](#)]
180. Thakur, V.K. *Biopolymer Grafting: Synthesis and Properties*, 1st ed.; Elsevier: Amsterdam, The Netherlands, 2017; pp. 1–580.
181. Nishimura, T.; Shishi, S.; Sasaki, Y.; Akiyoshi, K. Thermoresponsive polysaccharide graft polymer vesicles with tunable size and structural memory. *J. Am. Chem. Soc.* **2020**, *142*, 11784–11790. [[CrossRef](#)] [[PubMed](#)]
182. Noreen, A.; Zia, K.M.; Tabasum, S.; Khalid, S.; Shareef, R. A review on grafting of hydroxyethylcellulose for versatile applications. *Int. J. Biol. Macromol.* **2020**, *150*, 289–303. [[CrossRef](#)] [[PubMed](#)]
183. Segneanu, A.E.; Macarie, C.; Ungureanu, M.; Balcu, I.; Gherman, V.; Grozescu, I. Comparative study on enzymatic hydrolysis of cellulose. *Digest J. Nanomater. Biostruct.* **2013**, *8*, 1061–1068.
184. Carvalho, L.T.; Moraes, R.M.; Alves, G.M.; Lacerda, T.M.; Santos, J.C.; Santos, A.M.; Medeiros, S.F. Synthesis of amphiphilic pullulan-*graft*-poly(ϵ -caprolactone) via click chemistry. *Int. J. Biol. Macromol.* **2020**, *145*, 701–711. [[CrossRef](#)]
185. Kumar, D.; Gihar, S.; Shrivash, M.K.; Kumar, P.; Kundu, P.P. A review on the synthesis of graft copolymers of chitosan and their potential applications. *Int. J. Biol. Macromol.* **2020**, *163*, 2097–2112. [[CrossRef](#)]
186. Liu, J.; Yong, H.; Liu, Y.; Bai, R. Recent advances in the preparation, structural characteristics, biological properties and applications of gallic acid grafted polysaccharides. *Int. J. Biol. Macromol.* **2020**, *156*, 1539–1555. [[CrossRef](#)] [[PubMed](#)]
187. Sun, J.; Li, Z. Polyion complexes via electrostatic interaction of oppositely charged block copolymers. *Macromolecules* **2020**, *53*, 8737–8740. [[CrossRef](#)]

188. Zhang, Q.; Ko, N.R.; Oh, J.K. Recent advances in stimuli-responsive degradable block copolymer micelles: Synthesis and controlled drug delivery applications. *Chem. Commun.* **2012**, *48*, 7542–7552. [[CrossRef](#)] [[PubMed](#)]
189. Soni, V.; Pandey, V.; Asati, S.; Gour, V.; Tekade, R.K. Chapter 11—Biodegradable block copolymers and their applications for drug delivery. In *Basic Fundamentals of Drug Delivery*, 1st ed.; Tekade, R.K., Ed.; Academic Press–Elsevier: Cambridge, MA, USA, 2019; pp. 401–447.
190. Ishihara, M.; Kishimoto, S.; Nakamura, S.; Sato, Y.; Hattori, H. Polyelectrolyte complexes of natural polymers and their biomedical applications. *Polymers* **2019**, *11*, 672. [[CrossRef](#)] [[PubMed](#)]
191. Wu, D.; Zhu, L.; Li, Y.; Zhang, X.; Xu, S.; Yang, G.; Delair, T. Chitosan-based colloidal polyelectrolyte complexes for drug delivery: A review. *Carbohydr. Polym.* **2020**, *238*, 116126. [[CrossRef](#)] [[PubMed](#)]
192. Zhao, L.; Skwarczynski, M.; Toth, I. Polyelectrolyte-based platforms for the delivery of peptides and proteins. *ACS Biomater. Sci. Eng.* **2019**, *5*, 4937–4950. [[CrossRef](#)]
193. Timilsena, Y.P.; Akanbi, T.O.; Khalid, N.; Adhikari, B.; Barrow, C.J. Complex coacervation: Principles, mechanisms and applications in microencapsulation. *Int. J. Biol. Macromol.* **2019**, *121*, 1276–1286. [[CrossRef](#)] [[PubMed](#)]
194. Kuperkar, K.; Atanase, L.I.; Bahadur, A.; Crivei, I.C.; Bahadur, P. Degradable polymeric bio(nano)materials and their biomedical applications: A comprehensive overview and recent updates. *Polymers* **2024**, *16*, 206. [[CrossRef](#)]
195. Sionkowska, A. Current research on the blends of natural and synthetic polymers as new biomaterials: Review. *Prog. Polym. Sci.* **2011**, *36*, 1254–1276. [[CrossRef](#)]
196. Park, S.B.; Lih, E.; Park, K.S.; Joung, Y.K.; Han, D.K. Biopolymer-based functional composites for medical applications. *Prog. Polym. Sci.* **2017**, *68*, 77–105. [[CrossRef](#)]
197. Jacob, J.; Haponiuk, J.T.; Thomas, S.; Gopi, S. Biopolymer based nanomaterials in drug delivery systems: A review. *Mater. Today Chem.* **2018**, *9*, 43–55. [[CrossRef](#)]
198. Singh, A.V. Biopolymers in drug delivery: A review. *Pharmacol. Online* **2011**, *1*, 666–674.
199. Gopi, S.; Amalraj, A.; Sukumaran, N.P.; Haponiuk, J.T.; Thomas, S. Biopolymers and their composites for drug delivery: A brief review. *Macromol. Symp.* **2018**, *380*, 1800114. [[CrossRef](#)]
200. Müller, A.; Ni, Z.; Hessler, N.; Wesarg, F.; Müller, F.A.; Kralisch, D.; Fischer, D. The biopolymer bacterial nanocellulose as drug delivery system: Investigation of drug loading and release using the model protein albumin. *J. Pharm. Sci.* **2013**, *102*, 579–592. [[CrossRef](#)] [[PubMed](#)]
201. Das, D.; Pal, S. Modified biopolymer-dextrin based crosslinked hydrogels: Application in controlled drug delivery. *RSC Adv.* **2015**, *5*, 25014–25050. [[CrossRef](#)]
202. Revia, R.A.; Stephen, Z.R.; Zhang, M. Theranostic nanoparticles for RNA-based cancer treatment. *Acc. Chem. Res.* **2019**, *52*, 1496–1506. [[CrossRef](#)] [[PubMed](#)]
203. Chandakavathe, B.N.; Kulkarni, R.G.; Dhadde, S.B. Grafting of natural polymers and gums for drug delivery applications: A perspective review. *Crit. Rev. Ther. Drug Carrier Syst.* **2022**, *39*, 45–83. [[CrossRef](#)] [[PubMed](#)]
204. Pathak, K.; Misra, S.K.; Sehgal, A.; Singh, S.; Bungau, S.; Najda, A.; Gruszecki, R.; Behl, T. Biomedical applications of quaternized chitosan. *Polymers* **2021**, *13*, 2514. [[CrossRef](#)] [[PubMed](#)]
205. Verhoef, J.J.; Anchordoquy, T.J. Questioning the use of PEGylation for drug delivery. *Drug Deliv. Transl. Res.* **2013**, *3*, 499–503. [[CrossRef](#)] [[PubMed](#)]
206. Ahmady, A.; Abu Samah, N.H. A review: Gelatine as a bioadhesive material for medical and pharmaceutical applications. *Int. J. Pharm.* **2021**, *608*, 121037. [[CrossRef](#)] [[PubMed](#)]
207. Yang, G.; Xiao, Z.; Ren, X.; Long, H.; Qian, H.; Ma, K.; Guo, Y. Enzymatically crosslinked gelatin hydrogel promotes the proliferation of adipose tissue-derived stromal cells. *PeerJ* **2016**, *4*, e2497. [[CrossRef](#)]
208. Dozier, J.K.; Distefano, M.D. Site-specific PEGylation of therapeutic proteins. *Int. J. Mol. Sci.* **2015**, *16*, 25831–25864. [[CrossRef](#)]
209. Zong, Y.; Wu, J.; Shen, K. Nanoparticle albumin-bound paclitaxel as neoadjuvant chemotherapy of breast cancer: A systematic review and meta-analysis. *Oncotarget* **2017**, *8*, 17360–17372. [[CrossRef](#)]
210. Kim, H.J.; Kim, A.; Miyata, K.; Kataoka, K. Recent progress in development of siRNA delivery vehicles for cancer therapy. *Adv. Drug Deliv. Rev.* **2016**, *104*, 61–77. [[CrossRef](#)]
211. Kim, K.; Chen, W.C.W.; Heo, Y.; Wang, Y. Polycations and their biomedical applications. *Prog. Polym. Sci.* **2016**, *60*, 18–50. [[CrossRef](#)]
212. Debele, T.A.; Su, W.P. Polysaccharide and protein-based functional wound dressing materials and applications. *Int. J. Polym. Mater. Polym. Biomater.* **2022**, *71*, 87–108. [[CrossRef](#)]
213. Xu, R.; Fang, Y.; Zhang, Z.; Cao, Y.; Yan, Y.; Gan, L.; Xu, J.; Zhou, G. Recent advances in biodegradable and biocompatible synthetic polymers used in skin wound healing. *Materials* **2023**, *16*, 5459. [[CrossRef](#)]
214. Zhang, H.; Lin, X.; Cao, X.; Wang, Y.; Wang, J.; Zhao, Y. Developing natural polymers for skin wound healing. *Bioact. Mater.* **2024**, *33*, 355–376. [[CrossRef](#)]
215. Pandian, M.; Reshma, G.; Arthi, C.; Másson, M.; Rangasamy, J. Biodegradable polymeric scaffolds and hydrogels in the treatment of chronic and infectious wound healing. *Eur. Polym. J.* **2023**, *198*, 112390. [[CrossRef](#)]
216. Prete, S.; Dattilo, M.; Patitucci, F.; Pezzi, G.; Parisi, O.I.; Puoci, F. Natural and synthetic polymeric biomaterials for application in wound management. *J. Funct. Biomater.* **2023**, *14*, 455. [[CrossRef](#)]

217. Nguyen, H.M.; Le, T.T.N.; Nguyen, A.T.; Le, H.N.T.; Pham, T.T. Biomedical materials for wound dressing: Recent advances and applications. *RSC Adv.* **2023**, *13*, 5509–5528. [[CrossRef](#)]
218. Alven, S.; Peter, S.; Mbese, Z.; Aderibigbe, B.A. Polymer-based wound dressing materials loaded with bioactive agents: Potential materials for the treatment of diabetic wounds. *Polymers* **2022**, *14*, 724. [[CrossRef](#)]
219. Zhou, W.; Yu, X.; Zhang, Z.; Zou, X.; Song, H.; Zheng, W. Preparation and evaluation of luteolin-loaded PLA-based shape memory gastroretentive drug delivery systems. *Int. J. Pharm.* **2024**, *650*, 123670. [[CrossRef](#)]
220. Bajracharya, R.; Song, J.G.; Patil, B.R.; Lee, S.H.; Noh, H.-M.; Kim, D.-H.; Kim, G.L.; Seo, S.H.; Park, J.W.; Jeong, S.H.; et al. Functional ligands for improving anticancer drug therapy: Current status and applications to drug delivery systems. *Drug Deliv.* **2022**, *29*, 1959–1970. [[CrossRef](#)]
221. Rață, D.M.; Cadinoiu, A.N.; Atanase, L.I.; Bacaita, S.E.; Mihalache, C.; Daraba, O.-M.; Gherghel, D.; Popa, M. “In vitro” behaviour of aptamer-functionalized polymeric nanocapsules loaded with 5-fluorouracil for targeted therapy. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2019**, *103*, 109828. [[CrossRef](#)]
222. Rata, D.M.; Cadinoiu, A.N.; Atanase, L.I.; Popa, M.; Mihai, C.-T.; Solcan, C.; Ochiuz, L.; Vochita, G. Topical formulations containing aptamer-functionalized nanocapsules loaded with 5-fluorouracil—An innovative concept for the skin cancer therapy. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2021**, *119*, 111591. [[CrossRef](#)]
223. Pedram Rad, Z.; Mokhtari, J.; Abbasi, M. Biopolymer based three-dimensional biomimetic micro/nanofibers scaffolds with porous structures via tailored charge repulsions for skin tissue regeneration. *Polym. Adv. Technol.* **2021**, *32*, 3535–3548. [[CrossRef](#)]
224. Samiei, M.; Fathi, M.; Barar, J.; Fathi, N.; Amiryaghoubi, N.; Omidi, Y. Bioactive hydrogel-based scaffolds for the regeneration of dental pulp tissue. *J. Drug Deliv. Sci. Technol.* **2021**, *64*, 102600. [[CrossRef](#)]
225. Lavanya, K.; Chandran, S.V.; Balagangadharan, K.; Selvamurugan, N. Temperature- and pH-responsive chitosan-based injectable hydrogels for bone tissue engineering. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2020**, *111*, 110862. [[CrossRef](#)]
226. Tang, G.; Tan, Z.; Zeng, W.; Wang, X.; Shi, C.; Liu, Y.; He, H.; Chen, R.; Ye, X. Recent advances of chitosan-based injectable hydrogels for bone and dental tissue regeneration. *Front. Bioeng. Biotechnol.* **2020**, *8*, 587658. [[CrossRef](#)]
227. Rahmani, F.; Larbi Bouamrane, O.; Ben Bouabdallah, A.; Atanase, L.I.; Hellal, A.; Apintiliese, A.N. Biomimetic hydroxyapatite crystals growth on phosphorylated chitosan films by in vitro mineralization used as dental substitute materials. *Polymers* **2023**, *15*, 2470. [[CrossRef](#)]
228. Mohankumar, P.; Ajayan, J.; Mohanraj, T.; Yasodharan, R. Recent developments in biosensors for healthcare and biomedical applications: A review. *Measurement* **2021**, *16*, 108293. [[CrossRef](#)]
229. Gamboa, J.; Paulo-Mirasol, S.; Estrany, F.; Torras, J. Recent progress in biomedical sensors based on conducting polymer hydrogels. *ACS Appl. Bio Mater.* **2023**, *6*, 1720–1741. [[CrossRef](#)]
230. Jadoun, S.; Rathore, D.S. Polymer-based biosensors for medical applications. In *Handbook of Polymers in Medicine*, 1st ed.; Mozafari, M., Chauhan, N.P.S., Eds.; Woodhead Publishing Series in Biomaterials; Woodhead Publishing–Elsevier: Kidlington, UK, 2023; pp. 635–652.
231. Wen, P.; Ke, W.; Dirisala, A.; Toh, K.; Tanaka, M.; Li, J. Stealth and pseudo-stealth nanocarriers. *Adv. Drug Deliv. Rev.* **2023**, *198*, 114895. [[CrossRef](#)]
232. Ezike, T.C.; Okpala, U.S.; Onoja, U.L.; Nwike, C.P.; Ezeako, E.C.; Okpara, O.J.; Okoroafor, C.C.; Eze, S.C.; Kalu, O.L.; Odoh, E.C.; et al. Advances in drug delivery systems, challenges and future directions. *Heliyon* **2023**, *9*, e17488. [[CrossRef](#)]
233. Vargason, A.M.; Anselmo, A.C.; Mitragotri, S. The evolution of commercial drug delivery technologies. *Nat. Biomed. Eng.* **2021**, *5*, 951–967. [[CrossRef](#)]
234. Adepu, S.; Ramakrishna, S. Controlled drug delivery systems: Current status and future directions. *Molecules* **2021**, *26*, 5905. [[CrossRef](#)]
235. Mansour, A.; Romani, M.; Acharya, A.B.; Rahman, B.; Verron, E.; Badran, Z. Drug delivery systems in regenerative medicine: An updated review. *Pharmaceutics* **2023**, *15*, 695. [[CrossRef](#)]
236. Jeong, W.Y.; Kwon, M.; Choi, H.E.; Kim, K.S. Recent advances in transdermal drug delivery systems: A review. *Biomater. Res.* **2021**, *25*, 24. [[CrossRef](#)]
237. Liu, G.; Yang, L.; Chen, G.; Xu, F.; Yang, F.; Yu, H.; Li, L.; Dong, X.; Han, J.; Cao, C.; et al. A review on drug delivery system for tumor therapy. *Front. Pharmacol.* **2021**, *12*, 735446. [[CrossRef](#)]
238. Ahmad, W.; Khan, T.; Basit, I.; Imran, J. A comprehensive review on targeted drug delivery system. *Asian J. Pharm. Res.* **2022**, *12*, 335–340. [[CrossRef](#)]
239. Prakash, S. Nano-based drug delivery system for therapeutics: A comprehensive review. *Biomed. Phys. Eng. Express* **2023**, *9*, 052002. [[CrossRef](#)]
240. Bakhrushina, E.O.; Demina, N.B. Implants as targeted drug delivery systems (Review). *Pharm. Chem. J.* **2022**, *56*, 396–402. [[CrossRef](#)]
241. Mohammed, M.A.; Syeda, J.T.M.; Wasan, K.M.; Wasan, E.K. An overview of chitosan nanoparticles and its application in non-parenteral drug delivery. *Pharmaceutics* **2017**, *9*, 53. [[CrossRef](#)]
242. Kamaly, N.; Yameen, B.; Wu, J.; Farokhzad, O.C. Degradable controlled-release polymers and polymeric nanoparticles: Mechanisms of controlling drug release. *Chem. Rev.* **2016**, *116*, 2602–2663. [[CrossRef](#)]

243. Fortuni, B.; Inose, T.; Ricci, M.; Fujita, Y.; Van Zundert, I.; Masuhara, A.; Fron, E.; Mizuno, H.; Latterini, L.; Rocha, S.; et al. Polymeric engineering of nanoparticles for highly efficient multifunctional drug delivery systems. *Sci. Rep.* **2019**, *9*, 2666. [[CrossRef](#)]
244. Kuperkar, K.; Patel, D.; Atanase, L.I.; Bahadur, P. Amphiphilic block copolymers: Their structures, and self-assembly to polymeric micelles and polymersomes as drug delivery vehicles. *Polymers* **2022**, *14*, 4702. [[CrossRef](#)]
245. De, A.; Nayak, A.K.; Kundu, A.; Das, B.; Samanta, A. Chapter 7—Gum Arabic-based nanomaterials in drug delivery and biomedical applications. In *Biopolymer-Based Nanomaterials in Drug Delivery and Biomedical Applications*, 1st ed.; Bera, H., Mobaswar, C.M., Saha, S., Eds.; Academic Press—Elsevier: Cambridge, MA, USA, 2021; pp. 165–182.
246. Bratek-Skicki, A. Towards a new class of stimuli-responsive polymer-based materials—Recent advances and challenges. *Appl. Surf. Sci. Adv.* **2021**, *4*, 100068. [[CrossRef](#)]
247. Chatterjee, S.; Hui, P.C.-L. Review of stimuli-responsive polymers in drug delivery and textile application. *Molecules* **2019**, *24*, 2547. [[CrossRef](#)]
248. Zhu, M.; Whittaker, A.K.; Han, F.Y.; Smith, M.T. Journey to the market: The evolution of biodegradable drug delivery systems. *Appl. Sci.* **2022**, *12*, 935. [[CrossRef](#)]
249. Warrington, K.J.; Nair, U.; Carbone, L.D.; Kang, A.H.; Postlethwaite, A.E. Characterisation of the immune response to type I collagen in scleroderma. *Arthritis Res. Ther.* **2006**, *8*, R136. [[CrossRef](#)]
250. Dingman, R.; Balu-Iyer, S.V. Immunogenicity of protein pharmaceuticals. *J. Pharm. Sci.* **2019**, *108*, 1637–1654. [[CrossRef](#)]
251. Kuten Pella, O.; Hornyák, I.; Horváthy, D.; Fodor, E.; Nehrer, S.; Lacza, Z. Albumin as a biomaterial and therapeutic agent in regenerative medicine. *Int. J. Mol. Sci.* **2022**, *23*, 10557. [[CrossRef](#)]
252. Lee, C.H. A simple outline of methods for protein isolation and purification. *Endocrinol. Metab.* **2017**, *32*, 18–22. [[CrossRef](#)]
253. Amaecha, B.T.; Higham, S.M.; Edgar, W.M. Effect of sterilisation methods on the structural integrity of artificial enamel caries for intra-oral cariogenicity tests. *J. Dent.* **1999**, *27*, 313–316. [[CrossRef](#)]
254. Meyer, M.; Prade, I.; Leppchen-Fröhlich, K.; Felix, A.; Herdegen, V.; Haseneder, R.; Repke, J.U. Sterilisation of collagen materials using hydrogen peroxide doted supercritical carbon dioxide and its effects on the materials properties. *J. Supercrit. Fluids* **2015**, *102*, 32–39. [[CrossRef](#)]
255. Dai, Z.; Ronholm, J.; Tian, Y.; Sethi, B.; Cao, X. Sterilization techniques for biodegradable scaffolds in tissue engineering applications. *J. Tissue Eng.* **2016**, *7*, 1–13. [[CrossRef](#)]
256. Prakash, A.; Soni, H.; Mishra, A.; Sarma, P. Are your capsules vegetarian or nonvegetarian: An ethical and scientific justification. *Indian J. Pharmacol.* **2017**, *49*, 401–404.
257. Feier, A.M.; Portan, D.; Manu, D.R.; Kostopoulos, V.; Kotrotsos, A.; Strnad, G.; Dobreanu, M.; Salcudean, A.; Bataga, T. Primary MSCs for personalized medicine: Ethical challenges, isolation and biocompatibility evaluation of 3D electrospun and printed scaffolds. *Biomedicines* **2022**, *10*, 1563. [[CrossRef](#)]
258. Gheorghita, R.; Anchidin-Norocel, L.; Filip, R.; Dimian, M.; Covasa, M. Applications of biopolymers for drugs and probiotics delivery. *Polymers* **2021**, *13*, 2729. [[CrossRef](#)]
259. Opreș, O.; Mormile, C.; Lung, I.; Stegarescu, A.; Soran, M.L.; Soran, A. An overview of biopolymers for drug delivery applications. *Appl. Sci.* **2024**, *14*, 1383. [[CrossRef](#)]
260. Mandal, D.D.; Singh, G.; Majumdar, S.; Chanda, P. Challenges in developing strategies for the valorization of lignin—A major pollutant of the paper mill industry. *Environ. Sci. Pollut. Res. Int.* **2023**, *30*, 11119–11140. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.