

SUPPLEMENTAL INFORMATION

FIGURE S1

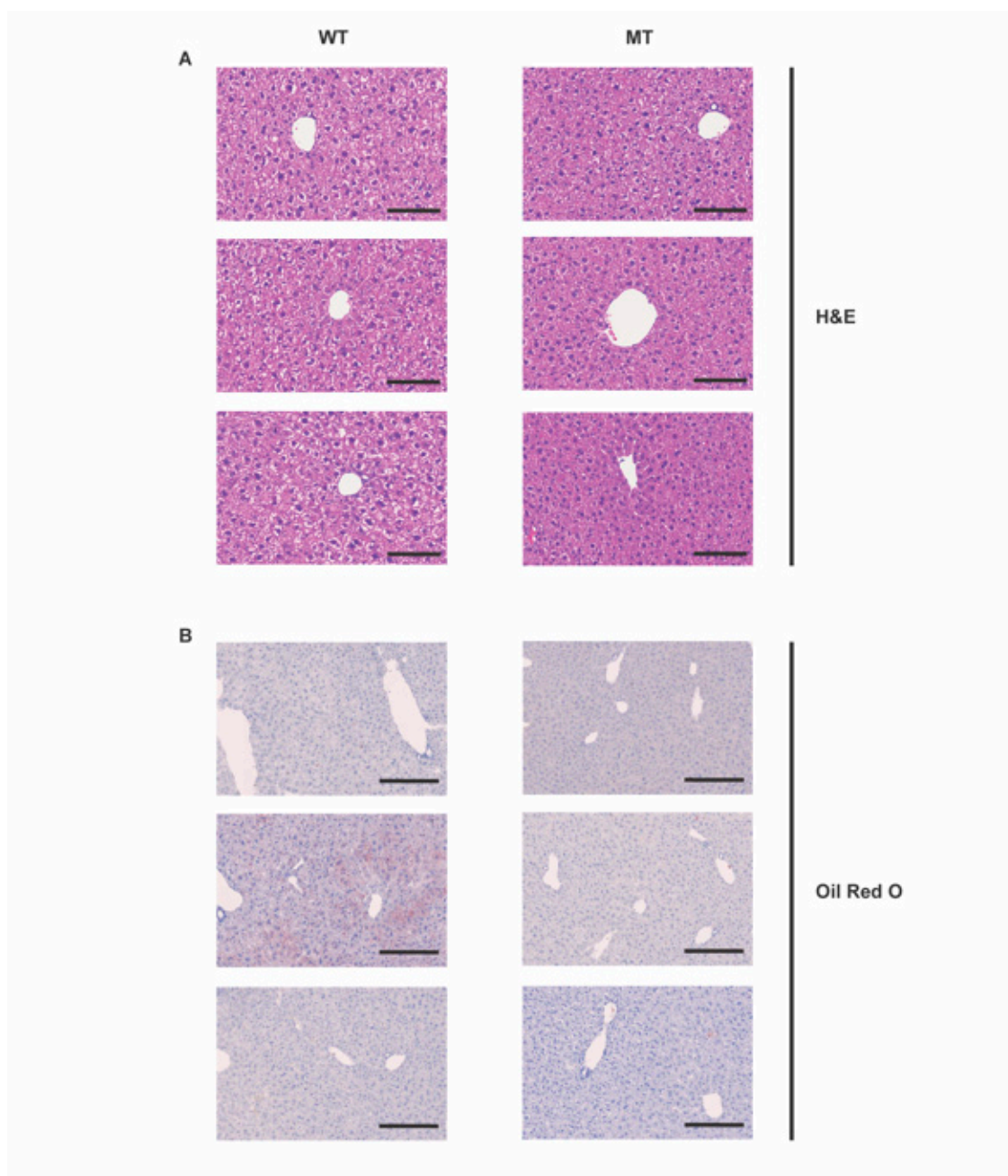


Figure S1. Representative images of liver histological analyses by (A) haematoxylin and eosin (H&E) and (B) oil red O staining. For A, scale bars = 100 μm ; For B, scale bars = 200 μm

FIGURE S2

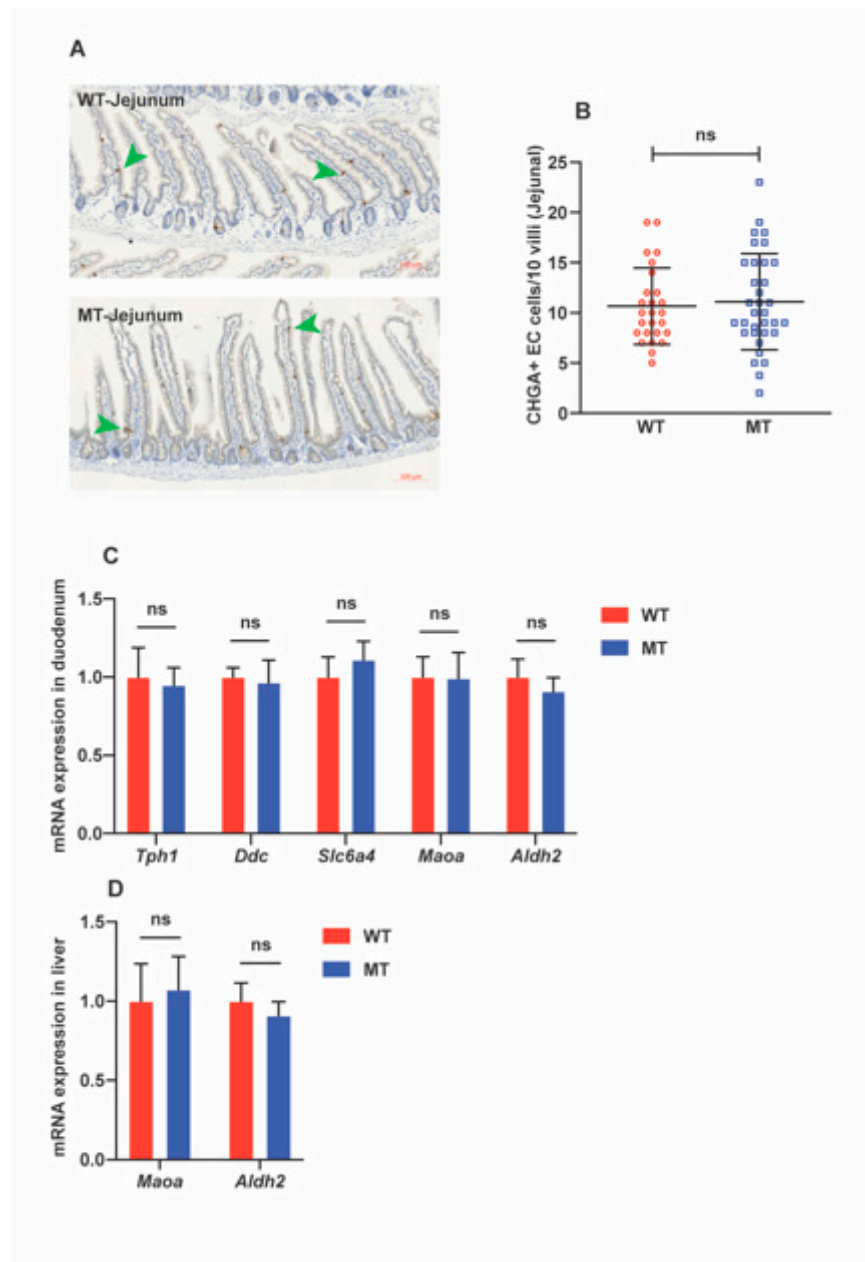


Figure S2. Jejunal serotonin-producing enterochromaffin cell numbers and serotonin metabolism in SI and liver in WT and MT mice. (A) Representative images of jejunal tissue sections stained with anti-chromogranin A (CHGA) antibody for enterochromaffin cells (in brown, indicated by green arrow heads), in jejunum of WT and MT mice, with haematoxylin-counterstained nuclei (in blue). Scale bar = 100 μ m. (B) Quantification of CHGA⁺ enterochromaffin cells (number of cells per 10 villi) in jejunum of WT and MT mice (n=6 per group). (C) mRNA expression of enzymes for serotonin synthesis *Tph1* (tryptophan hydroxylase 1) and *Ddc* (5-hydroxytryptophan decarboxylase), serotonin transporter *Slc6a4*, and enzymes for serotonin metabolism *Maoa* and *Aldh2*, in the duodenum of WT and MT mice

(n=8 per group). **(D)** mRNA expression of enzymes for serotonin metabolism *Maoa* and *Aldh2* in the livers of WT and MT mice (n=8 per group). mRNA levels were normalized to the control gene *Hprt1*. Data graphs show means \pm S.D. *p* values were calculated with the student's *t* test. ns denotes not significant with $p \geq 0.05$ between indicated groups.

FIGURE S3

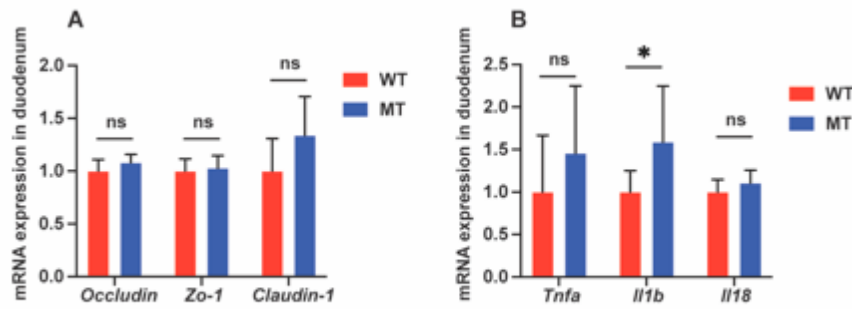


Figure S3. qPCR analyses on **(A)** intestinal tight junction proteins, *Occludin*, *Zo-1* (zonula occludens-1), and *Claudin-1* (n=8 per group); and **(B)** pro-inflammatory cytokines *Tnfa* (tumour necrosis factor alpha), *Il1b* (interleukin 1 beta), *Il18* (interleukin 18), in the duodena of WT and MT mice (n=7-8 per group). mRNA levels were normalized to the control gene *Hprt1*. Data graphs show means \pm S.D. *p* values were calculated with the student's *t* test. Statistical significance is presented as $*p < 0.05$ between indicated groups. ns denotes not significant with $p \geq 0.05$.

FIGURE S4

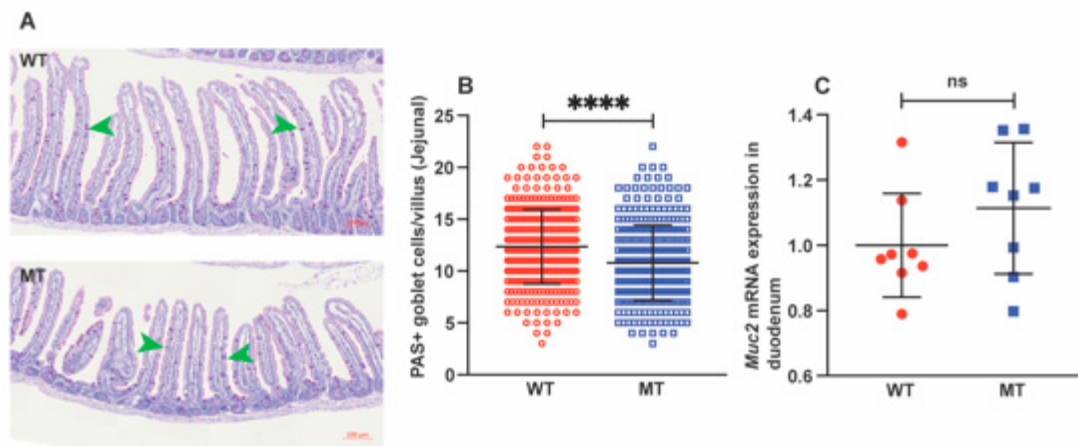


Figure S4. Indole-non-producing *E. coli*-colonized mice contained fewer mucus-producing goblet cells in the small intestines. (A) Representative images of jejunal tissue sections stained with periodic acid-Schiff (PAS) reagent for goblet cells (in deep purple, green arrow heads indicate representative cells), in jejunum of WT and MT mice, with haematoxylin-counterstained nuclei (in blue). Scale bar, 100 μ m. (B) Quantification of goblet cells (number of PAS⁺ cells per villus) in jejunum of WT and MT mice (n=6 per group). (C) mRNA expression of mucin 2 (*Muc2*) in the duodena of WT and MT mice (n=8 per group). mRNA levels were normalized to the control gene *Hprt1*. Data graphs show means \pm S.D. *p* values were calculated with the student's *t* test. Statistical significance is presented as *****p* < 0.0001 between indicated groups. ns denotes not significant with *p* \geq 0.05.

FIGURE S5

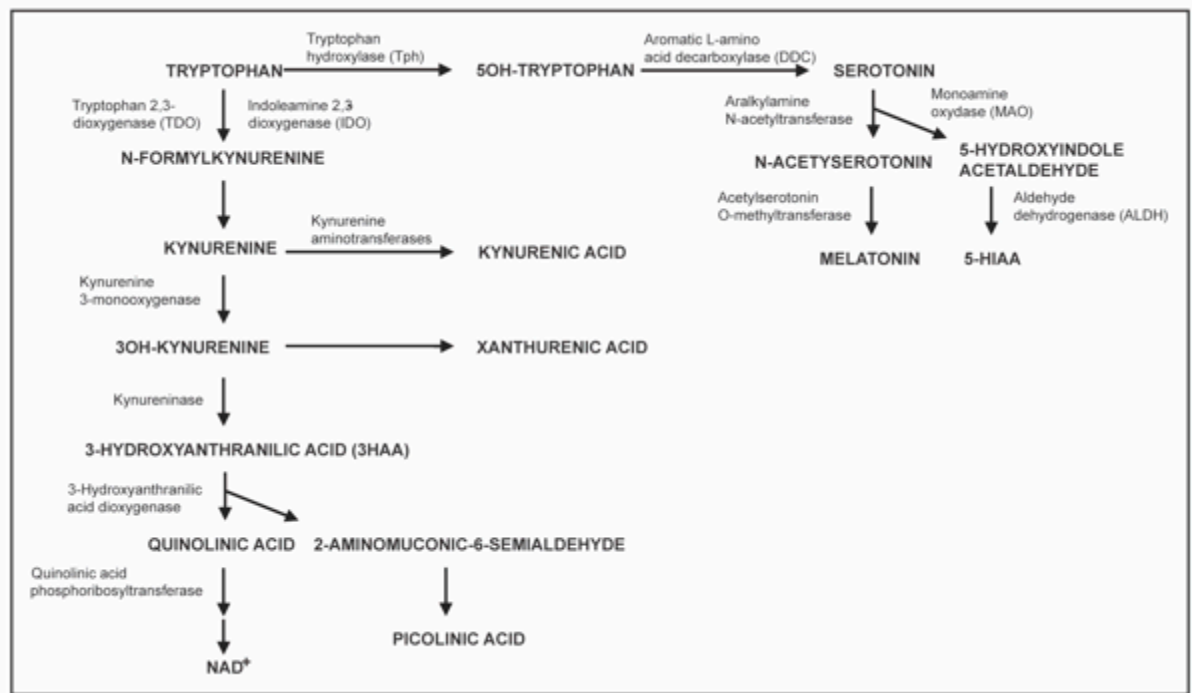


Figure S5. Illustration of tryptophan metabolisms in the mammalian host. Tryptophan can be metabolized in the host via the kynurenine pathway, through which kynurenine and NAD⁺, along with other metabolites, are produced. Alternatively, tryptophan can also be metabolized in the intestines by the enterochromaffin cells, or in the brain, through the serotonin pathway, leading to the production of serotonin. Serotonin can be further metabolized to 5-hydroxyindoleacetic acid (5-HIAA) or be utilized as a substrate for production of melatonin, a hormone regulating the sleep-wake cycle. NAD⁺, nicotinamide adenine dinucleotide (oxidized). Figure is adapted from Schwarcz & Stone, 2017⁴¹. Copyright © 2016 Elsevier Ltd. Adapted with permission.

Table S1. mRNA expression levels of selected genes in liver samples from WT and MT mice. n=7-8 per group. *p* values were calculated with the student's *t* test. Statistical significance is presented as **p* < 0.05, ***p* < 0.01, ****p* < 0.001, and *****p* < 0.0001 between indicated groups. ns denotes no statistical significance with *p* ≥ 0.05.

Functions	Genes	Gene products	Expression in MT mice compared with WT mice	Statistical significance	<i>p</i> values
Metabolism regulators	<i>Pgc1a</i>	Peroxisome proliferator-activated receptor gamma coactivator 1-alpha, a regulator of energy metabolism and mitochondrial biogenesis	-	ns	0.1093
	<i>Sirt1</i>	Sirtuin 1, a regulator of hepatic gluconeogenesis, fatty acid oxidation and cholesterol degradation	↑	**	0.0015
Glucose metabolism regulator	<i>Hnf4a</i>	Hepatocyte nuclear factor 4 alpha	↓	ns	0.1068
TCA cycle enzymes	<i>Pc</i>	Pyruvate carboxylase, converting pyruvate to oxaloacetate	↓	**	0.0046
	<i>Cs</i>	Citrate synthase, converting oxaloacetate to citrate	↓	****	<0.0001
	<i>Aco2</i>	Aconitase, converting citrate to <i>cis</i> -Aconitate	↓	*	0.0251
Lipogenesis enzymes	<i>Acly</i>	ATP citrate lyase, converting citrate to acetyl-CoA	↓	**	0.0096
	<i>Acaca</i>	Acetyl-CoA carboxylase (alpha), converting acetyl-CoA to malonyl-CoA (the rate-limiting step in fatty acid synthesis)	↓	*	0.0229
	<i>Acacb</i>	Acetyl-CoA carboxylase (beta)	↓	*	0.0146
	<i>Fasn</i>	Fatty acid synthase	↓	*	0.0167
	<i>Hmgcs1</i>	Hydroxymethylglutaryl-CoA synthase, second step enzyme of cholesterol synthesis pathway	↓	*	0.0429
	<i>Hmgcr</i>	Hydroxymethylglutaryl-CoA reductase, rate-controlling enzyme of cholesterol synthesis pathway	-	ns	0.2850
	<i>Gyk</i>	Glycerol kinase, an enzyme involved in lipogenesis	-	ns	0.5474
Lipolysis enzymes	<i>Acs11</i>	long-chain fatty-acid-coenzyme A ligase	-	ns	0.3367
	<i>Lipe</i>	Hormone-sensitive lipase	-	ns	0.1510
	<i>Pnpla2</i>	Adipose triglyceride lipase	-	ns	0.1439
Lipid	<i>Ppara</i>	Peroxisome proliferator-	-	ns	0.2821

metabolism regulators		activated receptor alpha, a major regulator of lipid metabolism in the liver			
	<i>Fiaf</i>	Angiopoietin-like 4, a serum hormone regulating lipid metabolism	↑	**	0.0061
Serotonin metabolic enzymes	<i>Maoa</i>	Monoamine oxidase A, enzyme for first step of serotonin metabolism	-	ns	0.5229
	<i>Aldh2</i>	Aldehyde dehydrogenase, enzyme for second step of serotonin metabolism	↓	*	0.0240
Inflammation	<i>Il1b</i>	Interleukin 1B, a proinflammatory cytokine	-	ns	0.1731
	<i>Il18</i>	Interleukin 18, a proinflammatory cytokine	-	ns	0.3366
K ⁺ channel	<i>Kcnj12</i>	Potassium Inwardly Rectifying Channel Subfamily J Member 12	↓	****	<0.0001

Table S2. mRNA expression levels of selected genes in epididymal white adipose tissue (WAT) samples from WT and MT mice. n=6-8 per group. *p* values were calculated with the student's *t* test. Statistical significance is presented as **p* < 0.05, ***p* < 0.01, ****p* < 0.001, and *****p* < 0.0001 between indicated groups. ns denotes no statistical significance with *p* ≥ 0.05.

Functions	Genes	Gene products	Expression in MT mice compared with WT mice	Statistical significance	<i>p</i> values
Metabolic regulators	<i>Pgc1a</i>	Peroxisome proliferator-activated receptor gamma coactivator 1-alpha	-	ns	0.8131
	<i>Pparg</i>	Peroxisome proliferator-activated receptor gamma, a regulator of fat storage and glucose metabolism	-	ns	0.4300
Glucose transporter	<i>Glut4</i>	Glucose transporter type 4	↑	*	0.0174
Glyceroneogenesis enzymes	<i>Pck1</i>	Phosphoenolpyruvate carboxykinase 1, cytosolic, an enzyme for glyceroneogenesis in adipose tissues	↑	*	0.0216
	<i>Pck2</i>	Phosphoenolpyruvate carboxykinase 2, mitochondrial, an enzyme for glyceroneogenesis in adipose tissues	↑	**	0.0027
Lipolysis enzymes	<i>Adrb2</i>	Adrenergic receptor beta-2, regulating lipolysis	↑	*	0.0323
	<i>Adrb3</i>	Adrenergic receptor beta-3, regulating lipolysis	↑	ns	0.1982
	<i>Acsl1</i>	long-chain fatty-acid-coenzyme A ligase	↑	**	0.0341
	<i>Lpl</i>	Lipoprotein lipase	-	ns	0.6162
	<i>Liipe</i>	Hormone-sensitive lipase	-	ns	0.9809
	<i>Pnpla2</i>	Adipose triglyceride lipase	-	ns	0.4106
	<i>Pnpla3</i>	Adiponutrin, a triacylglycerol lipase	-	ns	0.8219
	<i>Fiaf</i>	Angiopoietin-like 4, a serum hormone regulating lipid metabolism	-	ns	0.4988
Lipogenesis enzymes	<i>Scd1</i>	Stearoyl-CoA desaturase-1, catalysing the rate-limiting step in the formation of monounsaturated fatty acids	-	ns	0.9098
	<i>Scd2</i>	Stearoyl-CoA desaturase 2	↓	*	0.0294
	<i>Fasn</i>	Fatty acid synthase	↓	ns	0.9768
	<i>Acaca</i>	Acetyl-CoA carboxylase	-	ns	0.870

		(alpha)			2
	<i>Acacb</i>	Acetyl-CoA carboxylase (beta)	-	ns	0.532 2
Fatty acid transport	<i>Cd36</i>	Fatty acid translocase	-	ns	0.069 7
	<i>Fatp1</i>	Long-chain fatty acid transport protein 1	↑	**	0.002 0
	<i>Fatp4</i>	Long-chain fatty acid transport protein 4	↑	**	0.007 0
Heat production	<i>Ucp1</i>	Thermogenin	-	ns	0.084 8
	<i>Ucp2</i>	Mitochondrial uncoupling protein 2	-	ns	0.656 8
Inflammation	<i>Il1b</i>	Interleukin 1 beta, a proinflammatory cytokine	-	ns	0.157 4
	<i>Il6</i>	Interleukin 6, a pro-inflammatory cytokine and an anti-inflammatory myokine	↓	**	0.009 5
	<i>Il18</i>	Interleukin 18, a proinflammatory cytokine	-	ns	0.245 0
	<i>Tnfa</i>	Tumour necrosis factor, a cytokine for systemic inflammation	-	ns	0.487 6

Table S3. mRNA expression levels of selected genes in hindlimb muscles gastrocnemius (GS) samples from WT and MT mice. n=7-8 per group. *p* values were calculated with the student's *t* test. Statistical significance is presented as **p* < 0.05, ***p* < 0.01, ****p* < 0.001, and *****p* < 0.0001 between indicated groups. ns denotes no statistical significance with *p* ≥ 0.05.

Functions	Genes	Gene products	Expression in MT mice compared with WT mice	Statistical significance	<i>p</i> values
Metabolism regulators	<i>Pgcl1a</i>	Peroxisome proliferator-activated receptor gamma coactivator 1-alpha, a regulator of energy metabolism and mitochondrial biogenesis	-	ns	0.9843
	<i>Pparg</i>	Peroxisome proliferator-activated receptor gamma, a regulator of fat storage and glucose metabolism	-	ns	0.3779
Glucose metabolism and mitochondrial functions	<i>Glut4</i>	Glucose transporter	↓	*	0.0285
	<i>Sdh</i>	succinate dehydrogenase, a TCA cycle and respiratory chain enzyme	↓	ns	0.1879
	<i>Tfam</i>	Mitochondrial transcription factor A	↓	*	0.0456
	<i>Pdk4</i>	Pyruvate dehydrogenase lipoamide kinase isozyme 4, mitochondrial, a regulator of glucose metabolism	-	ns	0.6495
Lipid metabolism	<i>Ppara</i>	Peroxisome proliferator-activated receptor alpha, a major regulator of lipid metabolism in the liver	-	ns	0.6454
	<i>Fiaf</i>	Angiopoietin-like 4, a serum hormone regulating lipid metabolism	-	ns	0.9469
	<i>Srebp2</i>	Sterol regulatory element-binding protein 2, a transcription factor regulating cholesterol homeostasis	↓	*	0.0138
Muscle components	<i>Myhc-1</i>	Myosin heavy chain 1	-	ns	0.5249
	<i>Myhc-2a</i>	Myosin heavy chain 2a	-	ns	0.5336
	<i>Myhc-2b</i>	Myosin heavy chain 2b	-	ns	0.4575
	<i>Myhc-2d</i>	Myosin heavy chain 2d	↓	*	0.0182
	<i>Rapsyn</i>	Receptor-associated protein of the synapse (neuromuscular junction-associated gene)	↓	ns	0.2569
Muscle atrophy	<i>Murf-1</i>	Muscle RING-finger protein-1, a muscle atrophy E3 ubiquitin ligase	↓	*	0.0215
	<i>Atrogin-1</i>	A muscle atrophy E3 ubiquitin ligase	↓	**	0.0055

Myogenesis	<i>Myod</i>	Myoblast determination protein 1, regulating muscle differentiation	↓	n.s.	0.1975
Inflammation	<i>Il6</i>	Interleukin 6, a pro-inflammatory cytokine and an anti-inflammatory myokine	-	ns	0.4321
K ⁺ channel	<i>Kcnj12</i>	Potassium Inwardly Rectifying Channel Subfamily J Member 12	-	ns	0.3598

Table S4. mRNA expression levels of selected genes in small intestine (SI) samples from WT and MT mice. n=7-8 per group. *p* values were calculated with the student's *t* test. Statistical significance is presented as **p* < 0.05, ***p* < 0.01, ****p* < 0.001, and *****p* < 0.0001 between indicated groups. ns denotes no statistical significance with *p* ≥ 0.05.

Functions	Genes	Gene products	Expression in MT mice compared with WT mice	Statistical significance	<i>p</i> values
Gut barrier integrity	<i>Occludin</i>	Occludin, an epithelial cell tight junction protein	-	ns	0.1124
	<i>Zo-1</i>	Zonula occludens-1, an epithelial cell tight junction protein	-	ns	0.6254
	<i>Claudin-1</i>	Claudin-1, an epithelial cell tight junction protein	-	ns	0.0636
Inflammation	<i>Tnfa</i>	Tumour necrosis factor, a cytokine for systemic inflammation	-	ns	0.2304
	<i>Il1b</i>	Interleukin 1 beta, a proinflammatory cytokine	↑	*	0.0443
	<i>Il18</i>	Interleukin 18, a proinflammatory cytokine	-	ns	0.1732
	<i>Cd68</i>	Protein expressed by immune cells	-	ns	0.5714
Serotonin synthesis, transport, and metabolism	<i>Tph1</i>	Tryptophan hydroxylase 1, first-step enzyme in serotonin synthesis	-	ns	0.5121
	<i>Ddc</i>	Aromatic amino acid decarboxylase, second-step enzyme in serotonin synthesis	-	ns	0.5296
	<i>Slc6a4</i>	Serotonin transporter	-	ns	0.0962
	<i>Maoa</i>	Monoamine oxidase A, enzyme for first step of serotonin metabolism	-	ns	0.9137
	<i>Aldh2</i>	Aldehyde dehydrogenase, enzyme for second step of serotonin metabolism	-	ns	0.0934
Active intestinal stem cell markers	<i>Lgr5</i>	Leucine-rich G-protein-coupled receptor 5	↓	***	0.0007
	<i>Olfm4</i>	Olfactomedin 4	-	ns	0.2988
Goblet cell marker	<i>Muc2</i>	Mucin 2	-	ns	0.2306
Wnt signalling pathway (regulating stem cell self-renewal)	<i>Ctnnb1</i>	β-catenin, a protein involved in Wnt signalling, a pathway important for intestinal stem cell renewal	-	ns	0.3537
	<i>Lrp6</i>	Low-density lipoprotein receptor-related protein 6, a receptor in Wnt signalling	-	ns	0.7782
	<i>Fzd7</i>	Frizzled-7, a receptor in Wnt signalling	-	ns	0.2633

Notch signalling (modulating stem cell differentiation)	<i>Hes1</i>	A transcription factor interacts with Notch signalling, modulating secretory lineage differentiation in intestinal epithelium	-	ns	0.2309
Cell proliferation	<i>Mcc</i>	Colorectal mutant cancer protein, a negative regulator of cell cycle progression	↓	*	0.0171
	<i>Pcna</i>	Proliferating cell nuclear antigen	-	ns	0.0587
	<i>cMyc</i>	A transcription factor regulating many pro-proliferation genes	-	ns	0.6450
	<i>Ccnd1</i>	Cyclin D1, a protein required for progression through G1 phase of cell cycle	-	ns	0.4143
	<i>Cdkn1a</i>	Cyclin-dependent kinase (CDK) inhibitor 1, p21Cip1, inhibitor of cyclins/CDKs, including PCNA	-	ns	0.7205
Glucose metabolism	<i>Sirt1</i>	Sirtuin 1, a regulator of hepatic gluconeogenesis, fatty acid oxidation and cholesterol degradation	-	ns	0.7542
	<i>Glut2</i>	Glucose transporter 2	-	ns	0.5875
	<i>Abcg5</i>	Proteins for regulation of sterol absorption and excretion	-	ns	0.7158
	<i>Abcg8</i>		-	ns	0.1150
	<i>Idh2</i>	Isocitrate dehydrogenase [NADP], mitochondrial, a TCA cycle enzyme	↓	**	0.0046
Lipid metabolism	<i>Npc1l1</i>	A protein involved in cholesterol absorption	↓	*	0.0175
	<i>Scarb1</i>	Scavenger receptor class B type 1, a receptor involved in cholesterol transport	-	ns	0.5271
	<i>Fatp4</i>	Long-chain fatty acid transport protein 4	-	ns	0.9533
Neural signalling	<i>Npy</i>	Neuropeptide Y, modulating diverse neurological processes including food intake and fat storage	-	ns	0.2131
K ⁺ channel	<i>Kcnj12</i>	Potassium Inwardly Rectifying Channel Subfamily J Member 12	↓	**	0.004

Supplemental Table 5. DNA Oligo Primer Sequences Used in qPCR.

Gene Name	Sequence (5'→3')	
<i>Abcg5</i>	Forward	AGGGCCTCACATCAACAGAG
	Reverse	GCTGACGCTGTAGGACACAT
<i>Abcg8</i>	Forward	CTGTGGAATGGGACTGTACTTC
	Reverse	GTTGGACTGACCACTGTAGGT
<i>Acaca</i>	Forward	ATGGGCGGAATGGTCTCTTTC
	Reverse	TGGGGACCTTGTCTTCATCAT
<i>Acacb</i>	Forward	CCTTTGGCAACAAGCAAGGTA
	Reverse	AGTCGTACACATAGGTGGTCC
<i>Acly</i>	Forward	CAGCCAAGGCAATTCAGAGC
	Reverse	CTCGACGTTTGATTAAGTGGTCT
<i>Aco2</i>	Forward	ATCGAGCGGGGAAAGACATAC
	Reverse	TGATGGTACAGCCACCTTAGG
<i>Acs11</i>	Forward	TGCCAGAGCTGATTGACATTC
	Reverse	GGCATACCAGAAGGTGGTGAG
<i>Actb</i>	Forward	GGCTGTATTCCCCTCCATCG
	Reverse	CCAGTTGGTAACAATGCCATGT
<i>Adrb2</i>	Forward	GGGAACGACAGCGACTTCTT
	Reverse	GCCAGGACGATAACCGACAT
<i>Adrb3</i>	Forward	CCTTGGGCGAAACTGGTTG
	Reverse	GTTGGTGACAGCTAGGTAGCG
<i>Ahr</i>	Forward	GCCAGGACCAGTGTAGAGC
	Reverse	ATTCAGCGCCTGTAACAAGAA
<i>Ahrr</i>	Forward	GACCCTCCTCAGGTGGTGTT
	Reverse	AATGAAGCAGCGTGTCAAGAA
<i>Aldh2</i>	Forward	GACGCCGTCAGCAGGAAAA
	Reverse	CGCCAATCGGTACAACAGC
<i>Alpi</i>	Forward	AGGATCCATCTGTCCTTTGG
	Reverse	ACGTTGTATGTCTTGGACAG
<i>Atrogin-1</i>	Forward	ATGCACACTGGTGCAGAGAG
	Reverse	TGTAAGCACACAGGCAGGTC

<i>Bmi1</i>	Forward	CCAATGAAGACCGAGGAGAA
	Reverse	TTTCCGATCCAATCTGCTCT
<i>Ccnd1</i>	Forward	GCGTACCCTGACACCAATCTC
	Reverse	CTCCTCTTCGCACTTCTGCTC
<i>Cd36</i>	Forward	TTGTGGGGTTACAAAGATCCA
	Reverse	AGGATAAAACACACCAACTGTGG
<i>Cd68</i>	Forward	TGTCTGATCTTGCTAGGACCG
	Reverse	AGGAGAGTAACGGCCTTTTTG
<i>Cdkn1a</i>	Forward	CCTGGTGATGTCCGACCTG
	Reverse	CCATGAGCGCATCGCAATC
<i>Chga</i>	Forward	CACGGGAGACAGTGAGAGAG
	Reverse	TCCGACTGACCATCATCTTTCT
<i>Claudin-1</i>	Forward	TGGGTTTCATCCTGGCTTCT
	Reverse	TGTATCTGCCCCGGTGCTTT
<i>cMyc</i>	Forward	CCCTATTTTCATCTGCGACGAG
	Reverse	GAGAAGGACGTAGCGACCG
<i>Cpt1a</i>	Forward	CGCACATTACAAGGACATGG
	Reverse	GAAGAGCCGAGTCATGGAAG
<i>Cs</i>	Forward	GGACAATTTTCCAACCAATCTGC
	Reverse	TCGGTTCATTCCCTCTGCATA
<i>Ctnnb1</i>	Forward	ATGGAGCCGGACAGAAAAGC
	Reverse	CTTGCCACTCAGGGAAGGA
<i>Cyplal</i>	Forward	CAATGAGTTTGGGGAGGTTACTG
	Reverse	CCCTTCTCAAATGTCCTGTAGTG
<i>Cyplal2</i>	Forward	AGTACATCTCCTTAGCCCCAG
	Reverse	GGTCCGGGTGGATTCTTCAG
<i>Cyp11b1</i>	Forward	CACCAGCCTTAGTGCAGACAG
	Reverse	GAGGACCACGGTTTCCGTTG
<i>Ddc</i>	Forward	AGCTGGTTGCTTACACATCTG
	Reverse	CGAAGGGACTGCTTTTAGCTT
<i>Fasn</i>	Forward	TGGTGAATTGTCTCCGAAAAGA
	Reverse	CACGTTCATCACGAGGTCATG

<i>Fatp1</i>	Forward	CGCTTTCTGCGTATCGTCTG
	Reverse	GATGCACGGGATCGTGTCT
<i>Fatp4</i>	Forward	CTGTTTGCTTCAATGGTACAGC
	Reverse	CGGAAGGTCCAGTGAGTGTC
<i>Fiaf</i>	Forward	CAATGCCAAATTGCTCCAATT
	Reverse	TGGCCGTGGGCTCAGT
<i>Fzd7</i>	Forward	AGACCCACCTTTCACTGCG
	Reverse	AAGTACATGAGGCCGTTAGCA
<i>Glut2</i>	Forward	TCAGAAGACAAGATCACCGGA
	Reverse	GCTGGTGTGACTGTAAGTGGG
<i>Glut4</i>	Forward	GTGACTGGAACACTGGTCCTA
	Reverse	CCAGCCACGTTGCATTGTAG
<i>Gyk</i>	Forward	GTCAGCAACCAGAGGGAAACC
	Reverse	CCACGGCATTATAGAGAGGCT
<i>Hes1</i>	Forward	GATAGCTCCCGGCATTCCAAG
	Reverse	GCGCGGTATTTCCCCAACA
<i>Hmgcs1</i>	Forward	AAATGCCAGACCTACAGGTGG
	Reverse	ATGCTGCATGTGTGTCCCA
<i>Hmgcr</i>	Forward	AGCTTGCCCGAATTGTATGTG
	Reverse	TCTGTTGTGAACCATGTGACTTC
<i>Hnf4a</i>	Forward	GGTTTAGCCGACAATGTGTGG
	Reverse	TCCCGCTCATTTTGGACAGC
<i>Idh2</i>	Forward	GGAGAAGCCGGTAGTGGAGAT
	Reverse	GGTCTGGTCACGGTTTGGAA
<i>Il1b</i>	Forward	GAAATGCCACCTTTTGACAGTG
	Reverse	TGGATGCTCTCATCAGGACAG
<i>Il6</i>	Forward	CTGCAAGAGACTTCCATCCAG
	Reverse	AGTGGTATAGACAGGTCTGTTGG
<i>Il18</i>	Forward	GACTCTTGCGTCAACTTCAAGG
	Reverse	CAGGCTGTCTTTTGTCAACGA
<i>Ki67</i>	Forward	ATCATTGACCGCTCCTTTAGGT
	Reverse	GCTCGCCTTGATGGTTCCT

<i>Kcnj12</i>		PrimePCR SYBR Green Assay – Bio-Rad. Catalog # 10029101
<i>Lgr4</i>	Forward	GCAGTCTCTACGCTTAGATGC
	Reverse	CACTTCCGTCAAGATGTTGTCA
<i>Lgr5</i>	Forward	ACATTCCCAAGGGAGCGTTC
	Reverse	ATGTGGTTGGCATCTAGGCG
<i>Lipe</i>	Forward	CCAGCCTGAGGGCTTACTG
	Reverse	CTCCATTGACTGTGACATCTCG
<i>Lpl</i>	Forward	TTGCCCTAAGGACCCCTGAA
	Reverse	TTGAAGTGGCAGTTAGACACAG
<i>Lrp6</i>	Forward	TTGTTGCTTTATGCAAACAGACG
	Reverse	GTTCGTTTAATGGCTTCTTCGC
<i>Lyz</i>	Forward	GTCACACTTCCTCGCTTTC
	Reverse	TGGCTTTGCTGACTGACAAG
<i>Maoa</i>	Forward	GCCCAGTATCACAGGCCAC
	Reverse	CGGGCTTCCAGAACCAAGA
<i>Mcc</i>	Forward	CGACGTTGACAGGCACATAGA
	Reverse	GCCAGGTTGGGATACAGATCG
<i>Muc2</i>	Forward	CCGACTTCAACCCAAGTGAT
	Reverse	GAGCAAGGGACTCTGGTCTG
<i>Murf-1</i>	Forward	AGAAGTCGGGGGTCAGGGGACG
	Reverse	GGTCCATGATCACTTCATGGCGGCACGAGG
<i>Myhc-1</i>	Forward	CCAAGGGCCTGAATGAGGAG
	Reverse	GCAAAGGCTCCAGGTCTGAG
<i>Myhc-2a</i>	Forward	AAGCGAAGAGTAAGGCTGTC
	Reverse	GTGATTGCTTGCAAAGGAAC
<i>Myhc-2b</i>	Forward	ACAAGCTGCGGGTGAAGAGC
	Reverse	CAGGACAGTGACAAAGAACG
<i>Myhc-2d</i>	Forward	CCAAGTGCAGGAAAGTGACC
	Reverse	AGGAAGAGACTGACGAGCTC
<i>Myod</i>	Forward	TACAGTGGCGACTCAGATGC
	Reverse	GAGATGCGCTCCACTATGCT
<i>Npc11l</i>	Forward	CGCCCTTCTTTCTACATGGGT

	Reverse	GAATCTGCGCTTACGAGGGAG
<i>Npy</i>	Forward	TACTCCGCTCTGCGACACTA
	Reverse	TCTTCAAGCCTTGTTCTGGG
<i>Occludin</i>	Forward	GCAGCCTTCTGCTTCATCG
	Reverse	CGTCGGGTTCACCTCCATTA
<i>Olfm4</i>	Forward	CAGCCACTTTCCAATTTCACTG
	Reverse	GCTGGACATACTCCTTCACCTTA
<i>Pc</i>	Forward	CAGTGGCTGTCTACTCGGAG
	Reverse	CCGCATCTACACCATTTTCCT
<i>Pck1</i>	Forward	TGACAGACTCGCCCTATGTG
	Reverse	CCCAGTTGTTGACCAAAGGC
<i>Pck2</i>	Forward	ATGGCTGCTATGTACCTCCC
	Reverse	GCGCCACAAAGTCTCGAAC
<i>Pcna</i>	Forward	ACCTCACCAGCATGTCCAA
	Reverse	TCTTGATTGGTGCTTCGAATA
<i>Pdk4</i>	Forward	CCGCTTAGTGAACACTCCTTC
	Reverse	TGACCAGCGTGTCTACAAACT
<i>Pgcl1a</i>	Forward	AAGGGTCCCCATTTGAGA
	Reverse	AGGGTTATCTTGGTTGGCTTT
<i>Pnpla2</i>	Forward	TCCGTGGCTGTCTACTAAAGA
	Reverse	TGGGATATGATGACGTTCTCTCC
<i>Pnpla3</i>	Forward	AGGTCATTTCTGGCAAGGTTC
	Reverse	TCCACGACTTCGTCTTTGGAA
<i>Ppara</i>	Forward	AACATCGAGTGTCTGAATATGTGG
	Reverse	CCGAATAGTTCGCCGAAAGAA
<i>Pparg</i>	Forward	TCGCTGATGCACTGCCTATG
	Reverse	GAGAGGTCCACAGAGCTGATT
<i>Rapsyn</i>	Forward	GGCAGGACCAGACAAAGCAA
	Reverse	CGAGTGAGCTGTTACCAAGCA
<i>Scarb1</i>	Forward	TGTACTGCCTAACATCTTGGTCC
	Reverse	ACTGTGCGGTTCATAAAAGCA
<i>Scd1</i>	Forward	TTCTTGCGATACACTCTGGTGC

	Reverse	CGGGATTGAATGTTCTTGTCGT
<i>Scd2</i>	Forward	GATCTCTGGCGCTTACTCAGC
	Reverse	CTCCCCAGTGGTGAGAACTC
<i>Sdh</i>	Forward	GAAAGGCGGGCAGGCTCATC
	Reverse	CACCACGGCACTCCCCATTTT
<i>Sirt1</i>	Forward	GCTGACGACTTCGACGACG
	Reverse	TCGGTCAACAGGAGGTTGTCT
<i>Slc2a4</i>	Forward	GGACCGGATTCCATCCCAC
	Reverse	TCCCAACCATTGAGAAATGATGC
<i>Slc6a4</i>	Forward	GTCATTGGCTATGCCGTGGA
	Reverse	CACCCATTTTCGGTGGTACTG
<i>Slc25a1</i>	Forward	GAGGCATCGAAATCTGCATCA
	Reverse	GGATGGAGCCGTAGAGCAA
<i>Srebp2</i>	Forward	CCAAAGAAGGAGAGAGGCGG
	Reverse	CGCCAGACTTGTGCATCTTG
<i>Tfam</i>	Forward	GCTCTACACGCCCCTGGTTTCTGG
	Reverse	TCGCTGTAGTGCCTGCTGCTCCTG
<i>Tnfa</i>	Forward	GTCCCCAAAGGGATGAGAAGT
	Reverse	TTTGCTACGACGTGGGCTAC
<i>Tph1</i>	Forward	AACAAAGACCATTCCCTCCGAAAG
	Reverse	TGTAACAGGCTCACATGATTCTC
<i>Ucp1</i>	Forward	AGGCTTCCAGTACCATTAGGT
	Reverse	CTGAGTGAGGCAAAGCTGATTT
<i>Ucp2</i>	Forward	ATGGTTGGTTTCAAGGCCACA
	Reverse	CGGTATCCAGAGGGAAAGTGAT
<i>Vil1</i>	Forward	GGCAACGAGAGCGAGACTTT
	Reverse	GGAGTTTGTCTTCTACGTGCTTCA
<i>Zo-1</i>	Forward	TCACGATCTCCTGACCAACG
	Reverse	GGCTGACGGGTAAATCCACA