



Anticancer Potential and Molecular Targets of Pristimerin in Human Malignancies

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Abstract: The growing global burden of malignant tumors with increasing incidence and mortality rates underscores the urgent need for more effective and less toxic therapeutic options. Herbal compounds are being increasingly studied for their potential to meet these needs due to their reduced side effects and significant efficacy. Pristimerin (PS), a triterpenoid from the quinone formamide class derived from the Celastraceae and Hippocrateaceae families, has emerged as a potent anticancer agent. It exhibits broad-spectrum anti-tumor activity across various cancers such as breast, pancreatic, prostate, glioblastoma, colorectal, cervical, and lung cancers. PS modulates several key cellular processes, including apoptosis, autophagy, cell migration and invasion, angiogenesis, and resistance to chemotherapy, targeting crucial signaling pathways such as those involving NF-κB, p53, and STAT3, among others. The main objective of this review is to provide a comprehensive synthesis of the current literature on PS, emphasizing its mechanisms of action and molecular targets with the utmost clarity. It discusses the comparative advantages of PS over current cancer therapies and explores the implications for future research and clinical applications. By delineating the specific pathways and targets affected by PS, this review seeks to offer valuable insights and directions for future research in this field. The information gathered in this review could pave the way for the successful development of PS into a clinically applicable anticancer therapy.

Keywords: Pristimerin; apoptosis; autophagy; reactive oxygen species; signaling pathways

1. Introduction

Cancer is a significant global health issue that has attracted attention due to its devastating impact on humans worldwide. Chemotherapy, radiotherapy, immunotherapy, genetic and targeted treatment, hormone therapy, and surgery represent subsets of the multifaceted approaches that have been employed in the pursuit of cancer treatment. Among the myriad therapeutic alternatives at our disposal, chemotherapy stands out as a widely recognized modality that exhibits remarkable efficacy in the management of neoplastic growths [1]. Nevertheless, it is crucial to recognize the possible drawbacks linked to chemotherapy, including the development of multidrug resistance (MDR) in cancer cells and the occurrence of severe drug-induced adverse effects. Henceforth, it is imperative to foster the development of more efficacious treatment modalities [2,3].

For centuries, traditional Chinese medicine (TCM) has been seen as a potential remedy for the challenges that cancer poses [4]. This choice is primarily driven by the remarkably



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Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). diminished likelihood of encountering adverse effects and an elevated probability of achieving triumphant outcomes. Numerous naturally occurring compounds discovered within the realm of TCM have been scientifically validated for their remarkable anticancer attributes. Among these compounds are resveratrol, dioscin, berberine, curcumin, baicalein, wogonin, silibinin, quercetin, celastrol, and PS [1,4–9].

Triterpenoids are a class of essential compounds found in plants as specialized metabolites. Quinone methide triterpenoids, a specific group of compounds containing a quinone ring, have been of particular interest due to their cytotoxic and anti-tumoral activities [10]. Pristimerin (PS), 20α-3-hydroxy-2-oxo-24-nor-friedela-1-10,3,5,7-tetraen-carboxylicacid-29methylester, with the molecular formula $C_{30}H_{40}O_4$ (Figure 1), a quinonemethide triterpenoid compound of indigenous origin, is taxonomically categorized within the families Celastraceae and Hippocrateaceae. In 1951, Bhatnagar and Divekar achieved the commendable feat of isolating PS from botanical specimens of Pristimerae indica and P. grahami. Consequently, in 1954, diligent efforts were made by Kulkarni and Shah to successfully ascertain its precise molecular structure, thus solidifying its scientific identity [11]. Pristimerin has various pharmacological properties including anticancer, antioxidant, anti-inflammatory, antibacterial, antimalarial, and insecticidal actions [11]. Recent pharmacokinetic prediction studies employing molecular docking and simulation methodologies indicate that PS exhibited a substantial binding energy of -10.9 kcal/mol when interacting with pri-miR-378a [12]. The molecular dynamics simulation provided evidence of a stable connection between the PS and miRNA complex [12]. PS has the potential to regulate cancer-causing miRNAs, making it a promising candidate for cancer prevention and treatment due to its precise control of miRNA activity. PS's therapeutic value has drawn the interest of an increasing number of researchers who have made important contributions to the field. These characteristics have been extensively reported in various scientific studies [1,11].

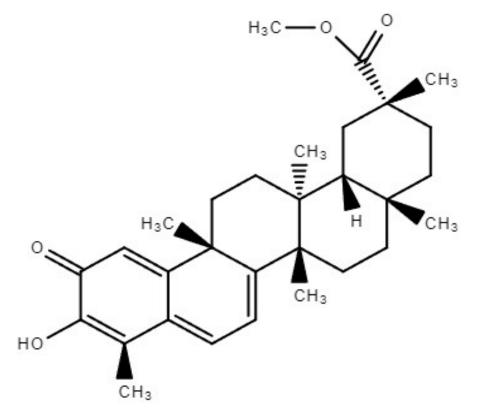


Figure 1. Chemical structure of Pristimerin.

PS demonstrates remarkable pharmacological efficacy in combating tumors through its capacity to selectively target and modulate a wide array of signal transduction pathways and molecular targets. These pathways encompass a myriad of intricate molecular interactions, such as the phosphatidylinositol 3-kinase (PI3K)/AKT pathway, the nuclear factor-kappaB (NF-kB) pathway, the reactive oxygen species (ROS)/mitogen-activated protein kinase (MAPK) pathway, the sonic hedgehog (Shh)/glioma-associated oncogene homolog 1 (Gli1) pathway, and the hypoxia-inducible factor 1a (HIF-1a)/sphingosine kinase 1 (SPHK1) pathway. Moreover, it significantly impacts the complex interaction between proteasome functioning and telomerase activity. PS exerts its anticancer effects via the induction of G1 phase arrest, apoptosis initiation, and autophagy facilitation in cancer cells. Furthermore, it induces inhibitory effects on the tumor cells through the modulation of cellular adhesion, reduction in cancer stem cell (CSC) activity, inhibition of the epithelial–mesenchymal transition (EMT), and inhibition of angiogenesis [11,13].

Several investigations have demonstrated that treating cancer cells with PS can reverse chemoresistance. Multiple studies have also shown that PS can increase the therapeutic effectiveness of various chemotherapeutic agents [14–16]. The primary aim of this review is to systematically consolidate the existing knowledge on the anticancer potential of PS, detailing how it interacts with and influences various molecular pathways to inhibit cancer progression. This review aims to provide a comprehensive resource for both researchers and clinicians, offering a clearer understanding of the multifunctional roles of PS in cancer therapy. By highlighting well-established and newly discovered mechanisms of PS, this synthesis encourages further research into its clinical applications and the development of novel therapeutic strategies that may incorporate or enhance the efficacy of PS.

This paper introduces several new scientific aspects of PS research. It examines the less explored molecular interactions of PS, particularly its ability to modulate proteasome functionality and telomerase activity. These areas are seldom the primary focus of cancer therapy studies but have shown promise in preliminary investigations. Additionally, this review discusses how PS has the potential to overcome chemoresistance, which is a major hurdle in current cancer treatment patterns. By elucidating the molecular basis for PS's ability to reverse resistance mechanisms, this work contributes to the frontier of cancer research. It suggests innovative approaches for integrating PS into existing treatment regimens, which not only deepens our understanding of PS's therapeutic scope but also establishes a foundational framework for future translational and clinical research in oncology.

2. Induction of Apoptosis

Apoptosis is a genetically regulated form of programmed cell death essential for eliminating unnecessary, damaged, or infected cells, thus maintaining cellular homeostasis [17]. The two main categories of apoptosis are the extrinsic death receptor system and the intrinsic mitochondrial pathway [18].

The death receptor pathway involves the activation of death receptors, such as Fas and tumor necrosis factor (TNF) receptor-1. This process results in the formation of a deathinducing signaling complex composed of the adaptor protein, the Fas-associated death domain, and initiator caspases, specifically caspase-8. In the context of the mitochondrial pathway, increased mitochondrial permeability results in the release of cytochrome-c, Apaf-1, and other apoptotic components into the cytoplasm. This event is followed by caspase activation and subsequent cellular death [1,11].

Several reports have documented that PS induces cell death both in vitro and in vivo (Tables 1 and 2) via the activation of mitochondrial signaling, caspase activation (both extrinsic and intrinsic), and PARP cleavage [5,19–22]. In addition, PS has been shown to induce apoptosis via the production of reactive oxygen species (ROS), resulting in significant cytotoxic effects across a diverse range of cancers [5,13,23–26].

| Cancer | Cell Lines | Mechanism of Action/s | PS Alone or/in Combination with Other Drug | References |
|---------------------------------|--|--|--|------------|
| Breast cancer | MDA-MB-231 | Increased expression of RGS4 that suppressed migration and invasion | PS (0–3 μM) | [27] |
| | ADR-resistant MCF-7/ADR | Induced apoptosis via suppression of AKT pathway | PS (0–10 μM) | [16] |
| | MDA-MB-231 | Enhanced autophagy induction | PS (5 μM) and paclitaxel (24 μM) | [15] |
| | MDA-MB-231, MDA-MB-468 | Decreased levels of phosphorylated ASK1 and JNK | PS (0–0.6 μM) | [28] |
| | MCF-7, MDA-MB-231,4T1 | Induced apoptosis, upregulated expression of miR-542-5p while downregulating DUB3 levels | PS (0–4 μM) | [29] |
| | epidermal growth factor receptor 2 (HER2)-positive SKBR3 | Reduced fatty acid synthase and altered AKT, MAPK, and mTOR pathways | PS (0–10 μM) | [30] |
| Cervical cancer | HeLa | Activated ROS-dependent JNK, Bax, and PARP-1 | PS | [25] |
| | HeLa, CasKi, SiHa | Induced ROS-dependent cell death | PS | [25] |
| Colorectal cancer | HCT116, HT-29 | Inhibited Wnt/β-catenin signaling pathway | PS | [31] |
| | AOM/DSS model of colitis-associated colorectal cancer | Suppression of AKT and FOXO3 signaling | PS | [32] |
| Leukemia | HL-60 | Induced apoptosis | PS | [26] |
| | K562 | ROS-JNK-induced autophagy. | PS (0–0.8 μM) | [33] |
| Lung cancer | NCI-H1299 | Targeted Shh/Gli1 signaling pathway | PS (0–500 nM) | [34] |
| | NCI-H1299 | Decreased the rate of migration and invasion | PS (0–500 nM) | [34] |
| Ovarian cancer | OVCAR-5, MDAH- 2774, SK-OV-3 | Inhibited AKT/NF-k B/mTOR pathway | PS (0–10 μM) | [19] |
| Pancreatic cancer | AsPC-1, BxPC-3, PANC-1 | Abrogated Chk1/53BP1-mediated DNA repair, suppressed NF-κB activity | PS (200 nM) + gemcitabine (500 nM) | [14,35] |
| | MiaPaCa-2, Panc-1 | Inhibited hTERT via suppressing transcription factors | PS (0–5 μM) | [36] |
| | MiaPaCa-2, Panc-1 | Induced inhibition of AKT/NF-кB/mTOR pathway | PS (5 μM) + proteasome inhibitors | [20] |
| Prostate | LNCaP, PC-3 | Decreased level of survivin and Bcl-2, and Inhibited hTERT mRNA expression levels. | PS (0–10 μM) | [24,37,38] |
| | PC-3, LNCaP, C4-2B | Induced apoptosis | PS (0–5 μM) | [39] |
| Oral squamous cell carcinoma | CAL-27, SCC-25 | G1 arrest via inhibition of MAPK/Erk1/2 and AKT pathway | PS (0–1 μM) | [40] |

Table 1. Anticancer activity of Pristimerin (PS) in in vitro models.

| Cancer | Animal Models | Mechanism of Action/s | PS Alone or/in Combination with Other Drug | References |
|-------------------|---|--|---|------------|
| Breast cancer | Breast cancer xenograft model | The inhibition of proteasomal activity and inhibition of tumor migration and invasion | PS (1 mg/kg) | [41] |
| | MDA-MB-231 tumor xenografts in nude mice | The stimulation of ROS/ASK1/JNK-mediated apoptosis and autophagy. Suppression of tumor growth | PS (0.5 mg/kg) | [28] |
| | Human breast cancer xenograft model | Suppression of VEGF, tumor growth, and angiogenesis | PS (3 mg/kg) | [42] |
| Cervical cancer | Tumor xenografts on nude mice | Apoptosis via mitochondrial signaling and activation of pro-apoptotic protein Bax | PS + Taxol | [43] |
| Colorectal cancer | Human colorectal cancer xenograft model | Inhibited tumor growth via targeting PI3K/AKT/mTOR pathway | PS (1 mg/kg) | [23] |
| | Human colorectal cancer xenograft model | Inhibited NF-кB signaling pathway | PS (1 mg/kg) | [44] |
| | Tumor xenograft in nude mice | Apoptosis through activation of the ROS/ER stress/JNK pathway | PS | [45] |
| Leukemia | Imatinib-resistant Bcr-Abl-T315I xenografts in mice | Suppressed TNF α -induced NFKB, as well as inhibition of the Bcr-Abl expression | PS (1 mg/kg) | [46] |
| Lung cancer | Human lung tumors xenograft model | Anticancer activity via targeting miR-23a/AKT/GSK3b pathway | PS (0.8 mg/kg) + cisplatin (2 mg/kg) | [47] |
| | Lung tissue samples from patients | Exerted anticancer activities through EphB4/CDC42/N-WASP pathway. | PS (0–8 μM) | [48] |
| Prostate cancer | Intra-tibial injection mouse model | Suppressed stem cell activity and angiogenesis via VEGF inhibition | PS (1.6 μM) | [49] |

Table 2. Anticancer activity of Pristimerin (PS) in in vivo models.

3. Induction of Autophagy

Autophagy is a critical process involved in the destruction of impaired organelles and aged proteins, as well as in the preservation of cellular homeostasis [50]. Autophagy in cancer biology serves a dual function, encompassing both tumor promotion and inhibition. The induction of autophagy in response to diverse cellular stressors plays a crucial role in regulating cell death, therefore offering a promising avenue for developing novel anticancer therapeutics [11,50]. ATG proteins encoded by autophagy-related genes (ATGs) help build the autophagosome [51]. The LC3 conversion (from LC3-I to LC3-II), which is recognized as a hallmark of autophagy [52], occurs when ATG4B combines with ATG7 to produce LC3II, which is formed by the conjugation of LC3-I and phosphatidylethanolamine. In cancer related to esophageal and cholangiocarcinoma, PS enhanced the ratio of LC3-II/LC3-I and increased the accumulation of autophagosomes [53]. Multiple in vitro investigations have demonstrated an upregulation of Beclin-1, ATG7, LC3-II, and p62 expression levels in malignancies associated with breast cancer (BC) and chronic myeloid leukemia (CML) [28,33,54]. Furthermore, the concurrent administration of PS and paclitaxel enhanced extracellular signal-related kinase (ERK)-mediated autophagic cell death. This was evidenced by an increase in the degradation of p62 and the expression of beclin1 [15]. In contrast, the study conducted by Zhang et al. [47] showed that PS had inhibitory effects on autophagy in lung cancer A549 and NCI-H446 cells. This was achieved by downregulating LC3BII and beclin1, ultimately enhancing the apoptotic response induced by cisplatin.

4. Inhibition of Cell Migration, Invasion and Metastasis

The metastasis of cancer encompasses a range of intricate cellular and molecular events, including cancer cell invasion, migration, and the establishment of metastatic colonies in clinical settings [11,55].

4.1. Epithelial–Mesenchymal Transition (EMT)

The inhibition of the epithelial-mesenchymal transition (EMT) involves the transformation of epithelial cells into mesenchymal cells, thereby enhancing the migratory and invasive capabilities of tumor cells [56,57]. Previous studies have demonstrated that treatment with PS significantly reduces the expression of key EMT markers including N-cadherin, fibronectin, vimentin, ZEB1, and Snail in both prostate and lung cancer cells, confirming its role in modulating EMT pathways [58,59]. Furthermore, the research by Shu et al. revealed that PS inhibits EMT in trophoblast cells via the miR-542-5p/EGFR signaling axis [60]. This effect of PS extends to suppressing proliferation, migration, and the EMT in trophoblast cells, while simultaneously inducing apoptosis. Notably, the downregulation of miR-542-5p, AGO2, and EGFR expression in these cells contributes to this phenomenon. Moreover, the co-treatment of PS with miR-542-5p silencing exhibited a synergistic effect, highlighting the potential of PS as a therapeutic agent for preventing embryo implantation in cases of ectopic pregnancy. In studies involving MDA-MB-231 triple-negative breast cancer cells, PS was shown to inhibit cell proliferation and revert EMT by upregulating E-cadherin and downregulating N-cadherin [61]. Additionally, PS suppresses the expression of integrin β 3 mRNA and protein, which are crucial components of the heterodimeric transmembrane receptor associated with the EMT. Experimental data indicate that silencing integrin β 3 enhances the anti-EMT effects of PS, whereas overexpressing this receptor diminishes its efficacy. In xenograft models, PS treatment led to the notable suppression of tumor growth, underscoring its potential utility in cancer therapy.

4.2. Cell Adhesion and Cytoskeleton Inhibition

Matrix metalloproteinases (MMPs) play a pivotal role in carcinoma development by modulating the tumor microenvironment. Notably, the proteins MMP2 and MMP9, which are crucial for facilitating invasion and metastasis, are downregulated by PS in a dose-dependent manner. This effect was specifically observed in EC9706 and EC109 cell lines, which are models for esophageal cancer, indicating the targeted action of PS on these enzymes [62,63]. Furthermore, PS has demonstrated significant efficacy in reducing lung cancer cell viability, migration, invasion, and capillary formation. These effects are mediated through the downregulation of the EphB4/CDC42/N-WASP signaling pathway. This pathway is critical for orchestrating mitochondrial-mediated intrinsic apoptosis and endoplasmic reticulum (ER) stress, thereby impeding cancer cell survival and spread [48]. Additional research involving the NCI-H1299 lung cancer cell line has shown that PS not only suppresses cell proliferation but also induces apoptosis. Alongside these effects, there was a notable reduction in the migration and invasion capabilities of H1299 cells, further supporting the potential of PS as a multifaceted therapeutic agent in the treatment of lung cancer [58].

5. Cancer Stem Cells (CSCs)

The inhibition of cancer stem cells (CSCs) is pivotal in understanding and managing carcinogenesis, tumor metastasis, and resistance to chemotherapy. Specific CSC markers such as CD44, aldehyde dehydrogenase (ALDH), and CD133 are integral to the processes of carcinogenesis, self-renewal, and therapy resistance. Additionally, transcription factors like NANOG, SOX-2, and OCT-4 have been implicated in the processes of metastasis and invasion, highlighting their roles in tumor progression and cellular invasiveness [64,65]. However, a significant challenge in CSC research is the identification of a precise and specific panel of CSC markers, as current markers often lack the specificity required for effective targeting and analysis. The natural compound PS has shown promising results in

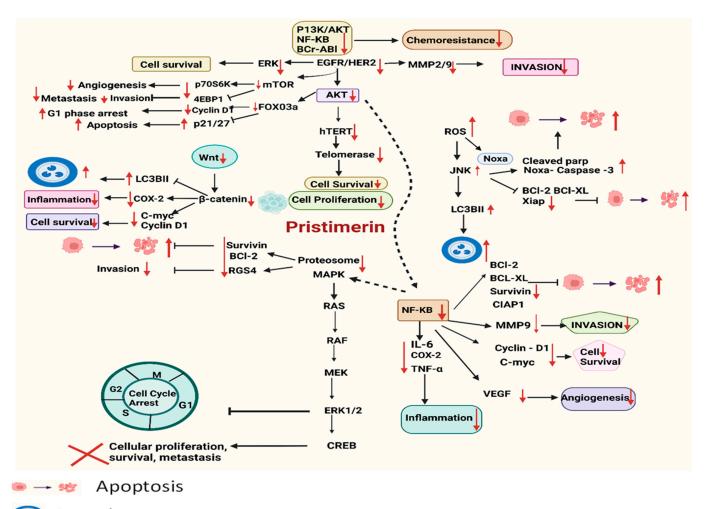
reducing the proportion of ALDH+ cells and inhibiting tumorsphere formation, suggesting its potential in targeting cellular mechanisms essential for CSC maintenance. In the context of uveal melanoma (UM), treatment with PS was found to decrease the levels of stemnessassociated proteins such as Slug and Sox2, although the levels of Nanog and KLF4 remained unchanged, indicating selective effects on stem cell-associated pathways [64]. In prostate cancer, using the PC-3 cell line as a model, PS administration resulted in a reduction in the levels of CD44, CD133, and other stemness factors including KLF4, OCT4, and AGO2. This indicates its broad-spectrum activity against various markers associated with stemness [64]. Additionally, the efficacy of PS in eradicating CSCs in esophageal squamous cell carcinoma (ESCC) has been linked to its modulation of the NF-KB signaling pathway, further underscoring its therapeutic potential [64]. PS also demonstrates a potent anticancer effect against breast cancer stem cells, inducing cell death through both apoptosis and incomplete autophagy. The underlying mechanisms involve the inhibition of thioredoxin-1, activation of the ASK1 and JNK signaling pathways, and generation of reactive oxygen species (ROS), all of which are crucial to the PS-induced cell death process [28,54]. These findings support the role of PS as a multifaceted agent capable of targeting and modifying the behavior of cancer stem cells across different cancer types.

6. Angiogenesis Inhibition

Angiogenesis, the process of new blood vessel formation, is critical for various physiological functions but can contribute detrimentally to cancer development when aberrantly activated. In parallel, the formation of new lymphatic vessels, known as lymphangiogenesis, also plays a significant role in cancer progression [66]. Consequently, therapies that target these processes, particularly anti-angiogenic treatments, are crucial in the fight against cancer. Recent research has highlighted the efficacy of PS in inhibiting angiogenesis triggered by vascular endothelial growth factor (VEGF) in human umbilical vascular endothelial cells (HUVECs). This inhibition has been observed both in vitro and in vivo, marking PS as a potent inhibitor of angiogenic signaling pathways [67]. Further studies have shown that in hypoxic conditions, PS effectively suppresses the expression and phosphorylation of key regulatory proteins such as HIF-1 α , SPHK-1, and AKT/GSK-3 β in PC-3 prostate cancer cells. This suppression leads to a consequential decrease in VEGF levels and angiogenesis, underlining the potential of PS in targeted cancer therapies [68]. Additionally, PS has demonstrated significant anticancer activity in uveal melanoma cells by inhibiting TNFα-induced p65 translocation, IκBα phosphorylation, and NF-κB-dependent gene expression. These molecular interventions contribute to the reduced cell viability and diminished clonogenic, migration, and invasion capabilities of uveal melanoma cells. Remarkably, when PS is combined with vinblastine, a primary therapeutic agent, a synergistic effect is observed, enhancing the therapeutic outcome. Moreover, PS induces apoptosis in these cells, providing a multi-faceted approach to combat uveal melanoma [69].

7. PS and Targeted Pathways

Over the past several decades, extensive research has revealed that many chronic health issues stem from the dysregulation of genes primarily involved in cell cycle control, particularly in oncology. This dysregulation often leads to unchecked cellular proliferation and subsequent metastasis. PS, a naturally occurring compound, has been shown to impact a wide array of biological processes by interacting with multiple signaling pathways. Comprehensive in vitro and in vivo studies have highlighted PS's effectiveness in inhibiting cell proliferation and promoting apoptotic mechanisms, and their detailed results are presented in Table 1 and illustrated in Figure 2.



📀 Autophagy

Figure 2. Schematic presentation of PS's anticancer mechanisms of action targeting various signaling cascades via upregulation and downregulation of different regulatory proteins.

The following sections provide a thorough overview of the anticancer activities attributed to PS, exploring its mechanisms of action and the specific signaling pathways it targets. These investigations have identified PS as a potent agent against a variety of cancer types, where it interrupts critical pathways involved in tumor growth and survival. Each cancer type responds differently, showcasing the adaptability and broad potential of PS as a key component in targeted cancer therapy.

7.1. PI3K/AKT Pathway

Phosphatidylinositol 3-kinase (PI3K/AKT/mTOR pathway cascade), or the PI3K/AKT pathway, is the most frequently altered pathway in humans for cancer development. This pathway plays a crucial role in several cellular processes, including cell cycle regulation, cell survival, metabolism, motility, angiogenesis, chemoresistance, and genomic stability [70]. The phosphorylation of Akt can trigger the subsequent phosphorylation of NF-kB, mTOR, and FoxO3a, which play a crucial role in cell growth, proliferation, survival, and angiogenesis [1]. Numerous research has yielded empirical data supporting the inhibitory efficacy of PS on the PI3K/AKT signaling pathway. In addition, PS treatment reduced the levels of mTOR-regulated phosphorylated S6K1 and phosphorylated 4E-BP1 [19,23,32,54].

Multiple biological processes are known to be linked to the PI3K/AKT pathway. Inhibiting AKT signaling reduces anti-apoptotic proteins while increasing pro-apoptotic

molecules [71]. Previous studies have provided evidence indicating that PS can inhibit the transcription of FoxO3a-target genes, including cyclinD1 and Bcl-xL, while concurrently promoting the overexpression of the p21 and p27 genes [32]. PS also decreased angiogenesis by targeting many signaling pathways, including VEGF-induced AKT, ERK1/2, mTOR, and ribosomal protein S6 kinase [67]. These findings indicate that PS suppresses cell migration, invasion, and metastasis via affecting the PI3K/AKT pathway.

7.2. ROS Generation and MAPK Pathway

The three main subfamilies of mitogen-activated protein kinases (MAPKs) are extracellular signal-regulated kinases (ERKs), p38, and stress-activated protein kinases (JNK) [72]. According to published reports, PS increased phosphorylated JNK and p38 in K562 leukemic cells and BC while decreasing phosphorylated ERK [28,73]. Numerous research studies have shown that the activation of JNK is closely linked to the initiation of apoptosis in many cancers [25,28,33].

Reports were published that PS treatment leads to an increase in reactive oxygen species (ROS) generation, which in turn results in heightened mitochondrial permeability and a reduction in the mitochondrial membrane potential, which leads to cell death [21,23,24,28,71,74]. Moreover, it has been shown that increased levels of reactive oxygen species (ROS) have been associated with the buildup of unfolded proteins and endoplasmic reticulum (ER) stress, eventually resulting in cellular apoptosis [45]. ROS generated by PS triggers JNK activation. Numerous publications have shown that PS has the ability to trigger cellular apoptosis (Table 2) via the activation of the ROS/JNK signaling pathway [25,63,75,76]. According to recent research in colorectal cancer (CRC) cells, JNK inhibition [25,63,75,76] results in caspase suppression, which in turn causes apoptosis [45]. Furthermore, PS has been found to induce cell cycle arrest in the G1 phase, as well as apoptosis and autophagy [28]. These cellular responses were initiated by the ROS/ASK1/JNK signaling cascades.

7.3. NF-*kB* Pathway

The NF-kB transcription factors have been widely recognized for their role in the context of cancer [77]. The activation of NF-kB leads to the upregulation of NF-kB-dependent genes [78], and several other signaling pathways are thought to play important roles in this process. PS has been shown in multiple investigations to effectively suppress TNF-a or LPS-induced p-IKK, p-IkB α , and p65 translocation [35,44,46,63,69,79].

PS has significantly reduced the expression of NF-kB-dependent genes (those involved in apoptosis, invasion, and angiogenesis) and cytokines [19,20,32,35,44,46,63,69]. The inhibition of the AKT/mTOR pathway and NF-kB has also been shown to reduce tumor angiogenesis in osteosarcoma cells [80], which may improve the efficacy of anticancer medicines and radiation.

7.4. HIF-1a/SPHK-1 Pathway

Hypoxia, a common feature in advanced solid tumors, has been identified as a significant factor in the advancement of metastasis and the emergence of treatment resistance. The activation of hypoxia-inducible transcription factors facilitates the occurrence of this phenomenon [81,82]. The overexpression of the transcription factor known as hypoxiainducible factor 1α (HIF- 1α) in human cancers can be attributed to hypoxic conditions within the tumor. This event subsequently stimulates angiogenesis and augments the survival capabilities and proliferation of cancerous cells [83].

The inhibitory effects of PS have been seen in the context of hypoxia-induced sphere and colony growth. PS has been demonstrated to have an inhibitory effect on the activity of the cancer stem cell markers CD44, KLF4, OCT4, and AGO2. Similarly, other studies reported that PS had an inhibitory effect on many epithelial–mesenchymal transition (EMT) markers, including N-cadherin, fibronectin, vimentin, and ZEB1 [59,68]. SPHK1 stabilizes HIF-1a via a downstream pathway involving AKT and GSK-3b. The enzymatic activity of SPHK-1 increases in hypoxia, whereas ROS regulates its control. Several studies have shown that PS can potentially decrease the expression of HIF-1 α via the inhibition of the SPHK-1 pathway. Further, sphingosine kinase-1 (SPHK-1) inhibition inhibited VEGF synthesis and cyclinD1 and CDK4 expression [1,68].

7.5. Ubiquitin–Proteosome Pathway

The ubiquitin–proteasome system coordinates physiological activities, including growth, death, cell cycle control, DNA repair, and antigen presentation [1]. The 26S eukaryotic proteasome comprises two 19S regulatory particles and one 20S catalytic particle. There are three catalytic sites on the 20S core particle: chymotrypsin-like (b5), trypsin-like (b2), and PGPH-like or caspase-like (b1) [84].

Different studies have shown that the conjugated ketone carbon (C6) of PS and the Nterminal threonine of the proteasomal b5 subunit have inhibitory effects on chymotrypsinlike activity. The inhibition results in polyubiquitination and subsequent modifications in the levels of Bax, p27, and IkB α , ultimately leading to the induction of cell death [39,43,85]. The suppression of Chk1 by PS has been found to cause a defect in DNA repair [86]. This could potentially broaden the use of olaparib in tumors that are proficient in BRCA but have TP53 mutations [86]. This suggests that PS can be used in combination with PARP inhibitor-based therapy.

Cancer cells exhibit increased expression of survivin, a protein that inhibits apoptosis and promotes cell survival by obstructing the mechanism of programmed cell death [1]. Several studies have shown that PS effectively decreases the levels of survivin by promoting its degradation via the ubiquitin–proteasome pathway, ultimately leading to apoptosis [37]. PS triggers apoptosis in prostate cancer cells via the generation of reactive oxygen species and the degradation of ubiquitin–proteasomes [24]. Additionally, PS decreases the invasiveness of breast cancer cells via enhancing RGS4 and suppressing proteasomal activity [41].

7.6. Wnt/β-Catenin Pathway

Wnt proteins function as critical mediators in a variety of important biological processes. The aberrant stimulation of the pathway mediated by Wnt/ β -catenin can result in various illnesses, including cancers [1]. PS has shown the ability to modulate the Wnt/ β catenin pathway in MCF-7 breast cancer cells. This modulation is achieved by the targeting and downregulation of LRP6 expression and phosphorylation, resulting in a decrease in the total β -catenin levels. Additionally, PS treatment leads to an elevation in LC3-II levels, suggesting their potential involvement in the regulation of autophagy [54]. PS has also been seen to downregulate Wnt target genes, including cyclinD1, c-Myc, β -catenin, and cox-2, in HT-29 and HCT116 CRC cells [31].

Dishevelled (Dvl), a key component of Wnt/ β -catenin signaling, increases β -catenin via phosphorylating/inactivating GSK3b and increasing AKT-Axin-GSK3b complex interaction [87]. Reports have shown that the degradation of β -catenin by GSK3b activation may cause PS-induced CRC cell death [31]. PS treatment decreased the p-AKT levels in MCF-7 breast cancer cells, suggesting that the PI3K/AKT pathway suppressed the Wnt/ β -catenin pathway in breast cancer [54].

7.7. Shh/Gli1 Pathway

The sonic hedgehog (Shh)/glioma-associated oncogene homolog 1 (Gli1) signaling pathway is known to be aberrantly activated in various human neoplasms, playing a critical role in tumor development and progression. This pathway is divided into two branches: the canonical and the non-canonical pathways. In the canonical Shh pathway, tumor-produced Shh ligands negate the suppressive action of the Patched (PTCH) receptor on the Smoothened (SMO) protein. This de-repression of SMO initiates a cascade of molecular events that ultimately activates Gli1, a transcription factor responsible for modulating gene expression that affects numerous cellular processes critical to tumorigenesis [1]. A

significant study by Lei et al. demonstrated that PS effectively disrupts this pathway. PS was shown to inhibit the nuclear localization of Gli1 in endothelial and pericyte cells within the tumor microenvironment. By preventing Gli1 from entering the nucleus, PS effectively shuts down the SHH/Gli1 signaling pathway, leading to a reduction in its ability to promote angiogenesis. This action results in the suppression of downstream signaling involved in the growth and maintenance of NCI-H1299 xenograft tumors, illustrating a potent antitumor mechanism of PS [34]. Through such targeted interference, PS holds promise as a therapeutic agent capable of mitigating the angiogenic and oncogenic potentials of the SHH/Gli1 pathway in cancer treatment.

7.8. Telomerase

Mammalian telomeres, which consist of a repetitive nucleotide sequence (TTAGGG), play a pivotal role in cellular aging and cancer biology. These sequences are capped and protected by a protein complex known as shelterin. This complex is critical for maintaining telomere integrity by preventing harmful end-to-end chromosomal fusions during the formation of the T-loop, a structure essential for telomere protection [1]. During cell division, these TTAGGG repeat sequences are progressively shortened, a process intrinsically linked to cellular aging known as senescence. This shortening mechanism serves as a natural limit to cell proliferation, thus acting as a barrier against the characteristic of uncontrolled cell growth in cancer. Notably, there is a strong correlation between the increased activity of the enzyme telomerase, which elongates telomeres, and the progression of cancer, underscoring the enzyme's role in tumorigenesis [88]. Recent studies have highlighted the efficacy of PS in combating cancer by targeting telomere dynamics. PS has been shown to inhibit the expression and activity of human telomerase reverse transcriptase (hTERT), the catalytic subunit of telomerase, in both pancreatic ductal adenocarcinoma and prostate cancer cells. This inhibition is mediated through the downregulation of several key transcription factors, including Sp1, c-Myc, NF-kB, and STAT-3, which are crucial for hTERT transcriptional activation. Furthermore, PS impacts the PI3K/AKT pathway, a significant regulator of cell survival and proliferation. It inhibits the phosphorylation of AKT and subsequently hTERT, leading to a reduction in telomerase activity. This effect contributes to telomere shortening and promotes the induction of senescence, thus impeding the proliferation of cancer cells [36,38]. By disrupting these critical cellular pathways, PS not only impedes cancer progression but also has potential as a therapeutic agent in oncology.

8. Role of PS in Overcoming Multidrug Resistance and Enhancing Chemotherapeutic Efficacy

Multidrug resistance (MDR) represents a significant challenge in cancer therapy, as cancer cells can develop resistance to a variety of chemotherapeutic agents, each with distinct molecular structures and mechanisms of action. One of the key players in MDR is the ATP-binding cassette (ABC) transporter family, particularly ABCB1 (P-glycoprotein, Pgp), which functions as a potent drug efflux pump. The elevation of ABCB1, along with the multidrug resistance-associated protein 1 (MRP1/ABCC1) and the breast cancer resistance protein [89–91]. PS has shown potential in overcoming ABCB1-mediated chemotherapeutic drug resistance. In human oral epidermoid carcinoma cells (KBv200), PS has been found to disrupt ABCB1 stability independently of its mRNA expression, highlighting a novel approach to mitigating MDR [92]. Additionally, PS has demonstrated efficacy against imatinib-resistant chronic myeloid leukemia (CML) cells both in vitro and in vivo by inhibiting NF-κB and Bcr-Abl, key factors in the development of resistance [46]. Moreover, PS has been linked to the reversal of MDR in multidrug-resistant MCF-7/ADR breast cancer cells through its association with the AKT signaling pathway, emphasizing its role in broader MDR contexts [16].

The integration of natural products with conventional chemotherapeutics has emerged as a promising strategy to enhance anticancer efficacy [1]. The integration of natural products with conventional chemotherapeutics has emerged as a promising strategy to enhance anticancer efficacy. For instance, PS has shown synergistic effects with 5-fluorouracil (5-FU) in ESCC [63], and when combined with paclitaxel (taxol), it has enhanced anti-tumor activities through ROS-mediated mitochondrial dysfunction in cervical cancer cells [43]. Together, they inhibit ERK1/2 [15], triggering autophagy in human BC cells. Additionally, the combination of PS and cisplatin in A549 and NCI-H446 lung cancer cells has been effective in inducing apoptosis by inhibiting the AKT/GSK3 β and miRNA-23a signaling pathways [47]. In pancreatic cancer, PS augments the cytotoxic effects of gencitabine by blocking the NF- κ B activation that is typically induced by gencitabine treatment [14,35]. Furthermore, the co-administration of PS and bortezomib has shown cytotoxic effects against myeloma, illustrating the broad potential of PS in enhancing the effectiveness of multiple chemotherapeutic regimens [85].

9. Conclusions and Future Perspectives

The potential of Chinese herbal therapy, particularly PS, in cancer treatment is being increasingly recognized. PS's effectiveness against cancer is mediated through several mechanisms, including the induction of autophagy, regulation of inflammation-related tumorigenesis, enhancement of chemosensitivity, and modulation of the tumor microenvironment and immune response. Its ability to inhibit cell proliferation, trigger apoptosis, prevent tumor metastasis and invasion, and suppress angiogenesis positions PS as a promising anticancer agent.

Despite the promising anticancer effects demonstrated by PS, significant research gaps remain that need to be addressed to advance PS from laboratory research to clinical applications. The extraction process of PS is inefficient, limiting its bioavailability and hindering further development. There is a critical need to understand the pharmacokinetics and pharmacodynamics of PS in diverse population groups. Detailed studies are required to explore how PS is metabolized and distributed in the human body, and how these processes vary by genetic makeup, age, and comorbid conditions. This understanding will be vital for optimizing dosages and minimizing potential side effects in clinical settings.

Furthermore, while PS has shown potential in modulating several cancer-related pathways, the exact molecular interactions and the potential for off-target effects remain underexplored. Future investigations should focus on the specificity of PS towards its molecular targets and its interaction with other cellular components in the tumor microenvironment. Such studies could elucidate the unintended impacts of PS, thereby refining its therapeutic profile. Additionally, comprehensive in vivo studies and clinical trials are needed to validate the efficacy and safety of PS observed in vitro. These studies should aim to establish robust clinical protocols and assess long-term outcomes to fully harness PS's potential as a part of integrated cancer therapy. The integration of advanced technologies like CRISPR gene editing and AI-driven predictive models could further delineate the role of PS in cancer therapy, providing deeper insights and accelerating its clinical translation.

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List of Abbreviations

| ABCB1 | ATP-Binding Cassette Subfamily B member |
|--------|---|
| AIF | Apoptosis-Inducing Factor |
| ATG | autophagy-related gene |
| BC | breast cancer |
| CAC | colitis-associated colorectal cancer |
| CML | chronic myeloid leukemia |
| CRC | colorectal cancer |
| CSCs | cancer stem cells |
| DVL | Dishevelled |
| EMT | epithelial-mesenchymal transition |
| ESCC | esophageal squamous cell carcinoma |
| HCC | hepatocellular carcinoma |
| HIF1-α | Hypoxia-Inducing Factor1—α |
| Htert | human telomerase reverse transcriptase |
| HUVEC | human umbilical vein endothelial cell |
| JNK | c-Jun NH(2)-terminal kinase |
| MDR | multidrug resistance |
| MMP | matrix metalloproteinase |
| NAC | N-Acetyl Cystine |
| NF-kB | nuclear factor-kB |
| PS | Pristimerin |
| ROS | reactive oxygen species |
| TCM | traditional Chinese medicine |
| TME | tumor microenvironment |
| TNF | tumor necrosis factor |
| UM | uveal melanoma |
| VEGF | vascular endothelial growth factor |
| | |

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References

- Chen, R.Z.; Yang, F.; Zhang, M.; Sun, Z.G.; Zhang, N. Cellular and Molecular Mechanisms of Pristimerin in Cancer Therapy: Recent Advances. *Front. Oncol.* 2021, *11*, 671548. [CrossRef] [PubMed]
- Bukowski, K.; Kciuk, M.; Kontek, R. Mechanisms of Multidrug Resistance in Cancer Chemotherapy. Int. J. Mol. Sci. 2020, 21, 3233. [CrossRef]
- Zhang, Q.Y.; Wang, F.X.; Jia, K.K.; Kong, L.D. Natural Product Interventions for Chemotherapy and Radiotherapy-Induced Side Effects. Front. Pharmacol. 2018, 9, 1253. [CrossRef]
- Xiang, Y.; Guo, Z.; Zhu, P.; Chen, J.; Huang, Y. Traditional Chinese medicine as a cancer treatment: Modern perspectives of ancient but advanced science. *Cancer Med.* 2019, *8*, 1958–1975. [CrossRef] [PubMed]
- Al-Tamimi, M.; Khan, A.Q.; Anver, R.; Ahmad, F.; Mateo, J.M.; Raza, S.S.; Alam, M.; Buddenkotte, J.; Steinhoff, M.; Uddin, S. Pristimerin mediated anticancer effects and sensitization of human skin cancer cells through modulation of MAPK signaling pathways. *Biomed. Pharmacother.* 2022, 156, 113950. [CrossRef]
- 6. Rauf, A.; Abu-Izneid, T.; Khalil, A.A.; Imran, M.; Shah, Z.A.; Emran, T.B.; Mitra, S.; Khan, Z.; Alhumaydhi, F.A.; Aljohani, A.S.M.; et al. Berberine as a Potential Anticancer Agent: A Comprehensive Review. *Molecules* **2021**, *26*, 7368. [CrossRef]
- Tomeh, M.A.; Hadianamrei, R.; Zhao, X. A Review of Curcumin and Its Derivatives as Anticancer Agents. Int. J. Mol. Sci. 2019, 20, 1033. [CrossRef]
- 8. Xiao, Q.; Zhu, W.; Feng, W.; Lee, S.S.; Leung, A.W.; Shen, J.; Gao, L.; Xu, C. A Review of Resveratrol as a Potent Chemoprotective and Synergistic Agent in Cancer Chemotherapy. *Front. Pharmacol.* **2018**, *9*, 1534. [CrossRef] [PubMed]
- Rauf, A.; Imran, M.; Khan, I.A.; Ur-Rehman, M.; Gilani, S.A.; Mehmood, Z.; Mubarak, M.S. Anticancer potential of quercetin: A comprehensive review. *Phytother. Res.* 2018, 32, 2109–2130. [CrossRef]
- 10. Huong, D.T.L.; Son, N.T. Pristimerin: Natural Occurrence, Biosynthesis, Pharmacology, and Pharmacokinetics. *Rev. Bras. Farmacogn.* **2024**, *34*, 467–480. [CrossRef]
- 11. Li, J.J.; Yan, Y.Y.; Sun, H.M.; Liu, Y.; Su, C.Y.; Chen, H.B.; Zhang, J.Y. Anti-Cancer Effects of Pristimerin and the Mechanisms: A Critical Review. *Front. Pharmacol.* **2019**, *10*, 746. [CrossRef]
- Mathur, A.; Singh, A.; Hussain, Y.; Mishra, A.; Meena, A.; Mishra, N.; Luqman, S. Regulating pri/pre-microRNA up/down expressed in cancer proliferation, angiogenesis and metastasis using selected potent triterpenoids. *Int. J. Biol. Macromol.* 2024, 257 *Pt* 1, 127945. [CrossRef]

- 13. Yousef, B.A.; Hassan, H.M.; Zhang, L.Y.; Jiang, Z.Z. Anticancer Potential and Molecular Targets of Pristimerin: A Mini- Review. *Curr. Cancer Drug Targets* 2017, *17*, 100–108. [CrossRef]
- 14. Jiang, Z.; Zhao, Y.; Zhao, Y.; Liu, Y.; Tao, L. Pristimerin synergizes with gemcitabine through abrogating Chk1/53BP1-mediated DNA repair in pancreatic cancer cells. *Food Chem. Toxicol.* **2021**, *147*, 111919. [CrossRef]
- 15. Lee, Y.; Na, J.; Lee, M.S.; Cha, E.Y.; Sul, J.Y.; Park, J.B.; Lee, J.S. Combination of pristimerin and paclitaxel additively induces autophagy in human breast cancer cells via ERK1/2 regulation. *Mol. Med. Rep.* **2018**, *18*, 4281–4288. [CrossRef] [PubMed]
- 16. Xie, G.; Yu, X.; Liang, H.; Chen, J.; Tang, X.; Wu, S.; Liao, C. Pristimerin overcomes adriamycin resistance in breast cancer cells through suppressing Akt signaling. *Oncol. Lett.* **2016**, *11*, 3111–3116. [CrossRef]
- 17. Ke, B.; Tian, M.; Li, J.; Liu, B.; He, G. Targeting Programmed Cell Death Using Small-Molecule Compounds to Improve Potential Cancer Therapy. *Med. Res. Rev.* 2016, *36*, 983–1035. [CrossRef] [PubMed]
- 18. Elmore, S. Apoptosis: A review of programmed cell death. Toxicol. Pathol. 2007, 35, 495–516. [CrossRef] [PubMed]
- 19. Gao, X.; Liu, Y.; Deeb, D.; Arbab, A.S.; Gautam, S.C. Anticancer activity of pristimerin in ovarian carcinoma cells is mediated through the inhibition of prosurvival Akt/NF-kappaB/mTOR signaling. *J. Exp. Ther. Oncol.* **2014**, *10*, 275–283.
- Deeb, D.; Gao, X.; Liu, Y.B.; Pindolia, K.; Gautam, S.C. Pristimerin, a quinonemethide triterpenoid, induces apoptosis in pancreatic cancer cells through the inhibition of pro-survival Akt/NF-kappaB/mTOR signaling proteins and anti-apoptotic Bcl-2. *Int. J. Oncol.* 2014, 44, 1707–1715. [CrossRef]
- 21. Yan, Y.Y.; Bai, J.P.; Xie, Y.; Yu, J.Z.; Ma, C.G. The triterpenoid pristimerin induces U87 glioma cell apoptosis through reactive oxygen species-mediated mitochondrial dysfunction. *Oncol. Lett.* **2013**, *5*, 242–248. [CrossRef] [PubMed]
- Downard, J.S.; Kim, S.H.; Kil, K.S. Localization of the cis-acting regulatory DNA sequences of the Myxococcus xanthus tps and ops genes. J. Bacteriol. 1988, 170, 4931–4938. [CrossRef] [PubMed]
- Yousef, B.A.; Hassan, H.M.; Guerram, M.; Hamdi, A.M.; Wang, B.; Zhang, L.Y.; Jiang, Z.Z. Pristimerin inhibits proliferation, migration and invasion, and induces apoptosis in HCT-116 colorectal cancer cells. *Biomed. Pharmacother.* 2016, 79, 112–119. [CrossRef] [PubMed]
- Liu, Y.B.; Gao, X.; Deeb, D.; Arbab, A.S.; Gautam, S.C. Pristimerin Induces Apoptosis in Prostate Cancer Cells by Down-regulating Bcl-2 through ROS-dependent Ubiquitin-proteasomal Degradation Pathway. J. Carcinog. Mutagen. 2013, (Suppl. 6), 005. [CrossRef] [PubMed]
- Byun, J.Y.; Kim, M.J.; Eum, D.Y.; Yoon, C.H.; Seo, W.D.; Park, K.H.; Hyun, J.W.; Lee, Y.S.; Lee, J.S.; Yoon, M.Y.; et al. Reactive oxygen species-dependent activation of Bax and poly(ADP-ribose) polymerase-1 is required for mitochondrial cell death induced by triterpenoid pristimerin in human cervical cancer cells. *Mol. Pharmacol.* 2009, *76*, 734–744. [CrossRef] [PubMed]
- Costa, P.M.; Ferreira, P.M.; Bolzani, V.d.S.; Furlan, M.; de Freitas Formenton Macedo Dos Santos, V.A.; Corsino, J.; de Moraes, M.O.; Costa-Lotufo, L.V.; Montenegro, R.C.; Pessoa, C. Antiproliferative activity of pristimerin isolated from Maytenus ilicifolia (Celastraceae) in human HL-60 cells. *Toxicol. In Vitro* 2008, 22, 854–863. [CrossRef] [PubMed]
- Wu, C.C.; Chan, M.L.; Chen, W.Y.; Tsai, C.Y.; Chang, F.R.; Wu, Y.C. Pristimerin induces caspase-dependent apoptosis in MDA-MB-231 cells via direct effects on mitochondria. *Mol. Cancer Ther.* 2005, *4*, 1277–1285. [CrossRef] [PubMed]
- Zhao, Q.; Liu, Y.; Zhong, J.; Bi, Y.; Liu, Y.; Ren, Z.; Li, X.; Jia, J.; Yu, M.; Yu, X. Pristimerin induces apoptosis and autophagy via activation of ROS/ASK1/JNK pathway in human breast cancer in vitro and in vivo. *Cell Death Discov.* 2019, *5*, 125. [CrossRef] [PubMed]
- 29. Cheng, S.; Zhang, Z.; Hu, C.; Xing, N.; Xia, Y.; Pang, B. Pristimerin Suppressed Breast Cancer Progression via miR-542-5p/DUB3 Axis. Onco Targets Ther. 2020, 13, 6651–6660. [CrossRef]
- 30. Lee, J.S.; Yoon, I.S.; Lee, M.S.; Cha, E.Y.; Thuong, P.T.; Diep, T.T.; Kim, J.R. Anticancer activity of pristimerin in epidermal growth factor receptor 2-positive SKBR3 human breast cancer cells. *Biol. Pharm. Bull.* **2013**, *36*, 316–325. [CrossRef]
- Zhao, Q.; Bi, Y.; Zhong, J.; Ren, Z.; Liu, Y.; Jia, J.; Yu, M.; Tan, Y.; Zhang, Q.; Yu, X. Pristimerin suppresses colorectal cancer through inhibiting inflammatory responses and Wnt/beta-catenin signaling. *Toxicol. Appl. Pharmacol.* 2020, 386, 114813. [CrossRef] [PubMed]
- 32. Park, J.H.; Kim, J.K. Pristimerin, a naturally occurring triterpenoid, attenuates tumorigenesis in experimental colitis-associated colon cancer. *Phytomedicine* **2018**, *42*, 164–171. [CrossRef] [PubMed]
- 33. Liu, Y.; Ren, Z.; Li, X.; Zhong, J.; Bi, Y.; Li, R.; Zhao, Q.; Yu, X. Pristimerin Induces Autophagy-Mediated Cell Death in K562 Cells through the ROS/JNK Signaling Pathway. *Chem. Biodivers.* **2019**, *16*, e1900325. [CrossRef] [PubMed]
- 34. Lei, X.; Zhong, Y.; Huang, L.; Li, S.; Fu, J.; Zhang, L.; Zhang, Y.; Deng, Q.; Yu, X. Identification of a novel tumor angiogenesis inhibitor targeting Shh/Gli1 signaling pathway in Non-small cell lung cancer. *Cell Death Dis.* **2020**, *11*, 232. [CrossRef] [PubMed]
- 35. Wang, Y.; Zhou, Y.; Zhou, H.; Jia, G.; Liu, J.; Han, B.; Cheng, Z.; Jiang, H.; Pan, S.; Sun, B. Pristimerin causes G1 arrest, induces apoptosis, and enhances the chemosensitivity to gemcitabine in pancreatic cancer cells. *PLoS ONE* 2012, 7, e43826. [CrossRef] [PubMed]
- 36. Deeb, D.; Gao, X.; Liu, Y.; Pindolia, K.; Gautam, S.C. Inhibition of hTERT/telomerase contributes to the antitumor activity of pristimerin in pancreatic ductal adenocarcinoma cells. *Oncol. Rep.* **2015**, *34*, 518–524. [CrossRef]
- Liu, Y.B.; Gao, X.; Deeb, D.; Brigolin, C.; Zhang, Y.; Shaw, J.; Pindolia, K.; Gautam, S.C. Ubiquitin-proteasomal degradation of antiapoptotic survivin facilitates induction of apoptosis in prostate cancer cells by pristimerin. *Int. J. Oncol.* 2014, 45, 1735–1741. [CrossRef]

- 38. Liu, Y.B.; Gao, X.; Deeb, D.; Pindolia, K.; Gautam, S.C. Role of telomerase in anticancer activity of pristimerin in prostate cancer cells. *J. Exp. Ther. Oncol.* **2015**, *11*, 41–49.
- 39. Yang, H.; Landis-Piwowar, K.R.; Lu, D.; Yuan, P.; Li, L.; Reddy, G.P.; Yuan, X.; Dou, Q.P. Pristimerin induces apoptosis by targeting the proteasome in prostate cancer cells. *J. Cell Biochem.* **2008**, *103*, 234–244. [CrossRef]
- 40. Wu, H.; Li, L.; Ai, Z.; Yin, J.; Chen, L. Pristimerin induces apoptosis of oral squamous cell carcinoma cells via G(1) phase arrest and MAPK/Erk1/2 and Akt signaling inhibition. *Oncol. Lett.* **2019**, *17*, 3017–3025. [CrossRef]
- 41. Mu, X.M.; Shi, W.; Sun, L.X.; Li, H.; Wang, Y.R.; Jiang, Z.Z.; Zhang, L.Y. Pristimerin inhibits breast cancer cell migration by upregulating regulator of G protein signaling 4 expression. *Asian Pac. J. Cancer Prev.* **2012**, *13*, 1097–1104. [CrossRef] [PubMed]
- 42. Deng, Q.; Bai, S.; Gao, W.; Tong, L. Pristimerin inhibits angiogenesis in adjuvant-induced arthritic rats by suppressing VEGFR2 signaling pathways. *Int. Immunopharmacol.* **2015**, *29*, 302–313. [CrossRef] [PubMed]
- Eum, D.Y.; Byun, J.Y.; Yoon, C.H.; Seo, W.D.; Park, K.H.; Lee, J.H.; Chung, H.Y.; An, S.; Suh, Y.; Kim, M.J.; et al. Triterpenoid pristimerin synergizes with taxol to induce cervical cancer cell death through reactive oxygen species-mediated mitochondrial dysfunction. *Anticancer Drugs* 2011, 22, 763–773. [CrossRef]
- Yousef, B.A.; Hassan, H.M.; Zhang, L.Y.; Jiang, Z.Z. Pristimerin exhibits in vitro and in vivo anticancer activities through inhibition of nuclear factor-small ka, CyrillicB signaling pathway in colorectal cancer cells. *Phytomedicine* 2018, 40, 140–147. [CrossRef] [PubMed]
- Zhao, Q.; Bi, Y.; Guo, J.; Liu, Y.; Zhong, J.; Liu, Y.; Pan, L.; Guo, Y.; Tan, Y.; Yu, X. Effect of pristimerin on apoptosis through activation of ROS/endoplasmic reticulum (ER) stress-mediated noxa in colorectal cancer. *Phytomedicine* 2021, *80*, 153399. [CrossRef] [PubMed]
- Lu, Z.; Jin, Y.; Chen, C.; Li, J.; Cao, Q.; Pan, J. Pristimerin induces apoptosis in imatinib-resistant chronic myelogenous leukemia cells harboring T315I mutation by blocking NF-kappaB signaling and depleting Bcr-Abl. *Mol. Cancer* 2010, *9*, 112, Erratum in *Mol. Cancer* 2021, 20, 172. [CrossRef]
- Zhang, Y.; Wang, J.; Hui, B.; Sun, W.; Li, B.; Shi, F.; Che, S.; Chai, L.; Song, L. Pristimerin enhances the effect of cisplatin by inhibiting the miR-23a/Akt/GSK3beta signaling pathway and suppressing autophagy in lung cancer cells. *Int. J. Mol. Med.* 2019, 43, 1382–1394. [CrossRef] [PubMed]
- Tang, Y.; Lei, Y.; Huang, S.; Li, Z.; Chen, X.; Luo, H.; Cheng, C.; Chen, J.; Zou, X.; Chen, X. Pristimerin Exacerbates Cellular Injury in Conditionally Reprogrammed Patient-Derived Lung Adenocarcinoma Cells by Aggravating Mitochondrial Impairment and Endoplasmic Reticulum Stress through EphB4/CDC42/N-WASP Signaling. Oxid. Med. Cell. Longev. 2020, 2020, 7409853. [CrossRef] [PubMed]
- 49. Huang, S.; He, P.; Peng, X.; Li, J.; Xu, D.; Tang, Y. Pristimerin Inhibits Prostate Cancer Bone Metastasis by Targeting PC-3 Stem Cell Characteristics and VEGF-Induced Vasculogenesis of BM-EPCs. *Cell. Physiol. Biochem.* **2015**, *37*, 253–268. [CrossRef]
- 50. Yun, C.W.; Lee, S.H. The Roles of Autophagy in Cancer. Int. J. Mol. Sci. 2018, 19, 3466. [CrossRef]
- 51. Mizushima, N.; Yoshimori, T.; Ohsumi, Y. The role of Atg proteins in autophagosome formation. *Annu. Rev. Cell Dev. Biol.* **2011**, 27, 107–132. [CrossRef] [PubMed]
- 52. Levy, J.M.M.; Towers, C.G.; Thorburn, A. Targeting autophagy in cancer. Nat. Rev. Cancer 2017, 17, 528–542. [CrossRef] [PubMed]
- 53. Huang, P.; Sun, L.Y.; Zhang, Y.Q. A Hopeful Natural Product, Pristimerin, Induces Apoptosis, Cell Cycle Arrest, and Autophagy in Esophageal Cancer Cells. *Anal. Cell. Pathol.* **2019**, 2019, 6127169. [CrossRef] [PubMed]
- 54. Cevatemre, B.; Erkisa, M.; Aztopal, N.; Karakas, D.; Alper, P.; Tsimplouli, C.; Sereti, E.; Dimas, K.; Armutak, E.I.I.; Gurevin, E.G.; et al. A promising natural product, pristimerin, results in cytotoxicity against breast cancer stem cells in vitro and xenografts in vivo through apoptosis and an incomplete autopaghy in breast cancer. *Pharmacol. Res.* 2018, 129, 500–514. [CrossRef] [PubMed]
- 55. Valastyan, S.; Weinberg, R.A. Tumor metastasis: Molecular insights and evolving paradigms. *Cell* **2011**, *147*, 275–292. [CrossRef] [PubMed]
- 56. Bakir, B.; Chiarella, A.M.; Pitarresi, J.R.; Rustgi, A.K. EMT, MET, Plasticity, and Tumor Metastasis. *Trends Cell Biol.* 2020, 30, 764–776. [CrossRef] [PubMed]
- 57. Mittal, V. Epithelial Mesenchymal Transition in Tumor Metastasis. Annu. Rev. Pathol. 2018, 13, 395–412. [CrossRef] [PubMed]
- 58. Li, J.; Guo, Q.; Lei, X.; Zhang, L.; Su, C.; Liu, Y.; Zhou, W.; Chen, H.; Wang, H.; Wang, F.; et al. Pristimerin induces apoptosis and inhibits proliferation, migration in H1299 Lung Cancer Cells. J. Cancer 2020, 11, 6348–6355. [CrossRef] [PubMed]
- Zuo, J.; Guo, Y.; Peng, X.; Tang, Y.; Zhang, X.; He, P.; Li, S.; Wa, Q.; Li, J.; Huang, S.; et al. Inhibitory action of pristimerin on hypoxia-mediated metastasis involves stem cell characteristics and EMT in PC-3 prostate cancer cells. *Oncol. Rep.* 2015, 33, 1388–1394. [CrossRef]
- 60. Shu, C.; Yu, X.; Cheng, S.; Jing, J.; Hu, C.; Pang, B. Pristimerin Suppresses Trophoblast Cell Epithelial-Mesenchymal Transition via miR-542-5p/EGFR Axis. *Drug Des. Dev. Ther.* **2020**, *14*, 4659–4670. [CrossRef]
- Liu, S.; Dong, Y.; Wang, Y.; Hu, P.; Wang, J.; Wang, R.Y. Pristimerin exerts antitumor activity against MDA-MB-231 triple-negative breast cancer cells by reversing of epithelial-mesenchymal transition via downregulation of integrin beta3. *Biomed. J.* 2021, 44 (Suppl. 1), S84–S92. [CrossRef] [PubMed]
- 62. Kessenbrock, K.; Plaks, V.; Werb, Z. Matrix metalloproteinases: Regulators of the tumor microenvironment. *Cell* **2010**, 141, 52–67. [CrossRef] [PubMed]

- 63. Tu, Y.; Tan, F.; Zhou, J.; Pan, J. Pristimerin targeting NF-kappaB pathway inhibits proliferation, migration, and invasion in esophageal squamous cell carcinoma cells. *Cell Biochem. Funct.* **2018**, *36*, 228–240. [CrossRef] [PubMed]
- 64. Chae, Y.C.; Kim, J.H. Cancer stem cell metabolism: Target for cancer therapy. BMB Rep. 2018, 51, 319–326. [CrossRef]
- 65. Singh, P.; Augustine, D.; Rao, R.S.; Patil, S.; Awan, K.H.; Sowmya, S.V.; Haragannavar, V.C.; Prasad, K. Role of cancer stem cells in head-and-neck squamous cell carcinoma—A systematic review. *J. Carcinog.* **2021**, *20*, 12. [CrossRef] [PubMed]
- 66. Nishida, N.; Yano, H.; Nishida, T.; Kamura, T.; Kojiro, M. Angiogenesis in cancer. *Vasc. Health Risk Manag.* 2006, 2, 213–219. [CrossRef] [PubMed]
- 67. Mu, X.; Shi, W.; Sun, L.; Li, H.; Jiang, Z.; Zhang, L. Pristimerin, a triterpenoid, inhibits tumor angiogenesis by targeting VEGFR2 activation. *Molecules* **2012**, *17*, 6854–6868. [CrossRef]
- Lee, S.O.; Kim, J.S.; Lee, M.S.; Lee, H.J. Anti-cancer effect of pristimerin by inhibition of HIF-1alpha involves the SPHK-1 pathway in hypoxic prostate cancer cells. *BMC Cancer* 2016, *16*, 701. [CrossRef] [PubMed]
- 69. Zhang, B.; Zhang, J.; Pan, J. Pristimerin effectively inhibits the malignant phenotypes of uveal melanoma cells by targeting NF-kappaB pathway. *Int. J. Oncol.* 2017, *51*, 887–898. [CrossRef]
- Mabuchi, S.; Kuroda, H.; Takahashi, R.; Sasano, T. The PI3K/AKT/mTOR pathway as a therapeutic target in ovarian cancer. *Gynecol. Oncol.* 2015, 137, 173–179. [CrossRef]
- 71. Yan, F.; Liao, R.; Silva, M.; Li, S.; Jiang, Y.; Peng, T.; Lazarovici, P.; Zheng, W. Pristimerin-induced uveal melanoma cell death via inhibiting PI3K/Akt/FoxO3a signalling pathway. *J. Cell. Mol. Med.* **2020**, *24*, 6208–6219. [CrossRef]
- Rezatabar, S.; Karimian, A.; Rameshknia, V.; Parsian, H.; Majidinia, M.; Kopi, T.A.; Bishayee, A.; Sadeghinia, A.; Yousefi, M.; Monirialamdari, M.; et al. RAS/MAPK signaling functions in oxidative stress, DNA damage response and cancer progression. *J. Cell. Physiol.* 2019, 234, 14951–14965. [CrossRef]
- 73. Huang, H.D.; Yao, B.G. Analysis of 1,698 cases of elderly gastric disease diagnosed by gastroscope. *Chin. Med. J.* **1987**, 100, 685–687. [PubMed]
- 74. Guo, Y.; Zhang, W.; Yan, Y.Y.; Ma, C.G.; Wang, X.; Wang, C.; Zhao, J.L. Triterpenoid pristimerin induced HepG2 cells apoptosis through ROS-mediated mitochondrial dysfunction. *J. BUON* **2013**, *18*, 477–485. [PubMed]
- 75. Hayashi, D.; Shirai, T.; Terauchi, R.; Tsuchida, S.; Mizoshiri, N.; Mori, Y.; Arai, Y.; Mazda, O.; Kubo, T. Pristimerin inhibits the proliferation of HT1080 fibrosarcoma cells by inducing apoptosis. *Oncol. Lett.* **2020**, *19*, 2963–2970. [CrossRef] [PubMed]
- 76. Zhao, H.; Wang, C.; Lu, B.; Zhou, Z.; Jin, Y.; Wang, Z.; Zheng, L.; Liu, K.; Luo, T.; Zhu, D.; et al. Pristimerin triggers AIF-dependent programmed necrosis in glioma cells via activation of JNK. *Cancer Lett.* **2016**, *374*, 136–148. [CrossRef] [PubMed]
- 77. Oeckinghaus, A.; Hayden, M.S.; Ghosh, S. Crosstalk in NF-kappaB signaling pathways. *Nat. Immunol.* **2011**, *12*, 695–708. [CrossRef] [PubMed]
- Yadav, V.R.; Prasad, S.; Sung, B.; Kannappan, R.; Aggarwal, B.B. Targeting inflammatory pathways by triterpenoids for prevention and treatment of cancer. *Toxins* 2010, 2, 2428–2466. [CrossRef] [PubMed]
- Rosa, A.; Ballarino, M.; Sorrentino, A.; Sthandier, O.; De Angelis, F.G.; Marchioni, M.; Masella, B.; Guarini, A.; Fatica, A.; Peschle, C.; et al. The interplay between the master transcription factor PU.1 and miR-424 regulates human monocyte/macrophage differentiation. *Proc. Natl. Acad. Sci. USA* 2007, 104, 19849–19854. [CrossRef]
- 80. Mori, Y.; Shirai, T.; Terauchi, R.; Tsuchida, S.; Mizoshiri, N.; Hayashi, D.; Arai, Y.; Kishida, T.; Mazda, O.; Kubo, T. Antitumor effects of pristimerin on human osteosarcoma cells in vitro and in vivo. *Onco Targets Ther.* **2017**, *10*, 5703–5710. [CrossRef]
- 81. Rankin, E.B.; Giaccia, A.J. Hypoxic control of metastasis. Science 2016, 352, 175–180. [CrossRef] [PubMed]
- 82. Vaupel, P. The role of hypoxia-induced factors in tumor progression. Oncologist 2004, 9 (Suppl. 5), 10–17. [CrossRef] [PubMed]
- 83. Wei, W.; Wu, S.; Wang, X.; Sun, C.K.; Yang, X.; Yan, X.; Chua, M.S.; So, S. Novel celastrol derivatives inhibit the growth of hepatocellular carcinoma patient-derived xenografts. *Oncotarget* **2014**, *5*, 5819–5831. [CrossRef] [PubMed]
- Mofers, A.; Pellegrini, P.; Linder, S.; D'Arcy, P. Proteasome-associated deubiquitinases and cancer. *Cancer Metastasis Rev.* 2017, 36, 635–653. [CrossRef] [PubMed]
- Tiedemann, R.E.; Schmidt, J.; Keats, J.J.; Shi, C.X.; Zhu, Y.X.; Palmer, S.E.; Mao, X.; Schimmer, A.D.; Stewart, A.K. Identification of a potent natural triterpenoid inhibitor of proteosome chymotrypsin-like activity and NF-kappaB with antimyeloma activity in vitro and in vivo. *Blood* 2009, *113*, 4027–4037. [CrossRef] [PubMed]
- 86. Tao, L.; Xia, X.; Kong, S.; Wang, T.; Fan, F.; Wang, W. Natural pentacyclic triterpenoid from Pristimerin sensitizes p53-deficient tumor to PARP inhibitor by ubiquitination of Chk1. *Pharmacol. Res.* **2024**, 201, 107091. [CrossRef] [PubMed]
- 87. Iijima, T.; Taira, N. Modification by manganese ions and verapamil of the responses of the atrioventricular node to norepinephrine. *Eur. J. Pharmacol.* **1976**, *37*, 55–62. [CrossRef] [PubMed]
- Jafri, M.A.; Ansari, S.A.; Alqahtani, M.H.; Shay, J.W. Roles of telomeres and telomerase in cancer, and advances in telomerasetargeted therapies. *Genome Med.* 2016, 8, 69. [CrossRef] [PubMed]
- 89. Wu, Q.; Yang, Z.; Nie, Y.; Shi, Y.; Fan, D. Multi-drug resistance in cancer chemotherapeutics: Mechanisms and lab approaches. *Cancer Lett.* **2014**, 347, 159–166. [CrossRef]
- 90. Dewanjee, S.; Dua, T.K.; Bhattacharjee, N.; Das, A.; Gangopadhyay, M.; Khanra, R.; Joardar, S.; Riaz, M.; Feo, V.; Zia-Ul-Haq, M. Natural Products as Alternative Choices for P-Glycoprotein (P-gp) Inhibition. *Molecules* **2017**, *22*, 871. [CrossRef]

- 91. Choi, Y.H.; Yu, A.M. ABC transporters in multidrug resistance and pharmacokinetics, and strategies for drug development. *Curr. Pharm. Des.* **2014**, *20*, 793–807. [CrossRef] [PubMed]
- 92. Yan, Y.Y.; Wang, F.; Zhao, X.Q.; Wang, X.K.; Chen, Y.F.; Liu, H.; Xie, Y.; Fu, L.W. Degradation of P-glycoprotein by pristimerin contributes to overcoming ABCB1-mediated chemotherapeutic drug resistance in vitro. *Oncol. Rep.* 2017, 37, 31–40. [CrossRef] [PubMed]

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