

SUPPLEMENTARY INFORMATION

Figure S1. Plasmids used in this work.

Name	Backbone	Insert sequence
p_mCherry_ EGFPn_intei nN	pEF1 α -mCherry-C1 (Clontech, Catalog No. 631972)	<p>ATGGTGAGCAAGGGCGAGGAGGATAACATGGCCATCATCAAGGAGTTCATGCGCT TCAAGGTGCACATGGAGGGCTCCGTGAACGGCCACGAGTTCGAGATCGAGGGCGA GGGCGAGGGCCGCCCTACGAGGGCACCCAGACCGCCAAGCTGAAGGTGACCAA GGGTGGCCCCCTGCCCTTCGCTGGGACATCCTGTCCCCTCAGTTCATGTACGGCTC CAAGGCCTACGTGAAGCACCCCGCCGACATCCCCGACTACTTGAAGCTGCCTTCC CCGAGGGCTTCAAGTGGGAGCGCGTGATGAACTTCGAGGACGGCGCGTGGTGAC CGTGACCCAGGACTCCTCCCTGCAGGACGGCGAGTTCATCTACAAGGTGAAGCTGC GCGGCACCAACTTCCCCTCCGACGGCCCCGTAATGCAGAAGAAGACCATGGGCTG GGAGGCCTCCTCCGAGCGGATGTACCCCGAGGACGGCGCCCTGAAGGGCGAGATC AAGCAGAGGGCTGAAGCTGAAGGACGGCGGCCACTACGACGCTGAGGTCAAGACC ACCTACAAGGCCAAGAAGCCCGTGCAGCTGCCCGGCGCCTACAACGTCAACATCA AGTTGGACATCACCTCCCACAACGAGGACTACACCATCGTGAACAGTACGAACG CGCCGAGGGCCGCCACTCCACCGCGGCATGGACGAGCTGTACAAGTCCGGAGCG GGCAGCGGGCGGGCAGCGGCAATTTCTGGCAGCATGAGCAAAGGAGAAGAA CTGTTTACGGAGTGGTGCCAATTCTCGTGGAACTGGATGGCGATGTTAATGGCA CAAATTTCTGTGACAGGAGAAGGTGAAGGTGATGCCACAATTGAAAGCTCACT CTGAAATTCATCTGCACCACTGGAAAACCTCCCTGTGCCATGGCCAACACTGGTCAC TACCTTAACCTACGCGTGCAGTGCTTTTCAAGATATCCAGATCATATGAAAAGGC ATGACTTTTTCAAATCAGCAATGCCCGAGGGGTATGTGCAGGAGCGAACGATTCT TTCAAAGATGATGGGAAATACAAGACCCGCGCTGTTGTTAAGTTGAAGGAGACG ATAAAGGCATTGAATGCAGCGTGTTTAGCATTGATAGCAACGGCATTGTGTATACC CAGCCGATTGCGCAGTGGCATCATCGCGGCAAACAGGAAGTGTTTGAATATTGCCT GGAAGATGGCAGCATTATTAAGCGACCAAAGATCATAAATTTATGACCCAGGAT GGCAAATGCTGCCGATTGATGAAATTTTGAACAGGAACTGGATCTGCTGCAGGT GAAAGCCTGCCGAATAAG</p>
p_inteinC_E GFPC_SBF	pCMV-GFP (Addgene #11153)	<p>ATGGTGAGCAAGGGCGAGGAGGATAACATGGCCATCATCAAGGAGTTCATGCGCT TCAAGGTGCACATGGAGGGCTCCGTGAACGGCCACGAGTTCGAGATCGAGGGCGA GGGCGAGGGCCGCCCTACGAGGGCACCCAGACCGCCAAGCTGAAGGTGACCAA GGGTGGCCCCCTGCCCTTCGCTGGGACATCCTGTCCCCTCAGTTCATGTACGGCTC CAAGGCCTACGTGAAGCACCCCGCCGACATCCCCGACTACTTGAAGCTGCCTTCC CCGAGGGCTTCAAGTGGGAGCGCGTGATGAACTTCGAGGACGGCGCGTGGTGAC CGTGACCCAGGACTCCTCCCTGCAGGACGGCGAGTTCATCTACAAGGTGAAGCTGC GCGGCACCAACTTCCCCTCCGACGGCCCCGTAATGCAGAAGAAGACCATGGGCTG GGAGGCCTCCTCCGAGCGGATGTACCCCGAGGACGGCGCCCTGAAGGGCGAGATC AAGCAGAGGGCTGAAGCTGAAGGACGGCGGCCACTACGACGCTGAGGTCAAGACC ACCTACAAGGCCAAGAAGCCCGTGCAGCTGCCCGGCGCCTACAACGTCAACATCA AGTTGGACATCACCTCCCACAACGAGGACTACACCATCGTGAACAGTACGAACG CGCCGAGGGCCGCCACTCCACCGCGGCATGGACGAGCTGTACAAGTCCGGAGCG GGCAGCGGGCGGGCAGCGGCAATTTCTGGCAGCATGAGCAAAGGAGAAGAA CTGTTTACGGAGTGGTGCCAATTCTCGTGGAACTGGATGGCGATGTTAATGGCA CAAATTTCTGTGACAGGAGAAGGTGAAGGTGATGCCACAATTGAAAGCTCACT CTGAAATTCATCTGCACCACTGGAAAACCTCCCTGTGCCATGGCCAACACTGGTCAC TACCTTAACCTACGCGTGCAGTGCTTTTCAAGATATCCAGATCATATGAAAAGGC ATGACTTTTTCAAATCAGCAATGCCCGAGGGGTATGTGCAGGAGCGAACGATTCT TTCAAAGATGATGGGAAATACAAGACCCGCGCTGTTGTTAAGTTGAAGGAGACG</p>

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p_mCherry_ pEF1 α -mCherry-C1
EGFP1- (Clontech,
10_inteinN Catalog No. 631972)

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p_inteinC_E pCMV-GFP (Addgene
GFP11_SBF #11153)
P

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p_EGFP_int pCMV-GFP (Addgene
einN #11153)

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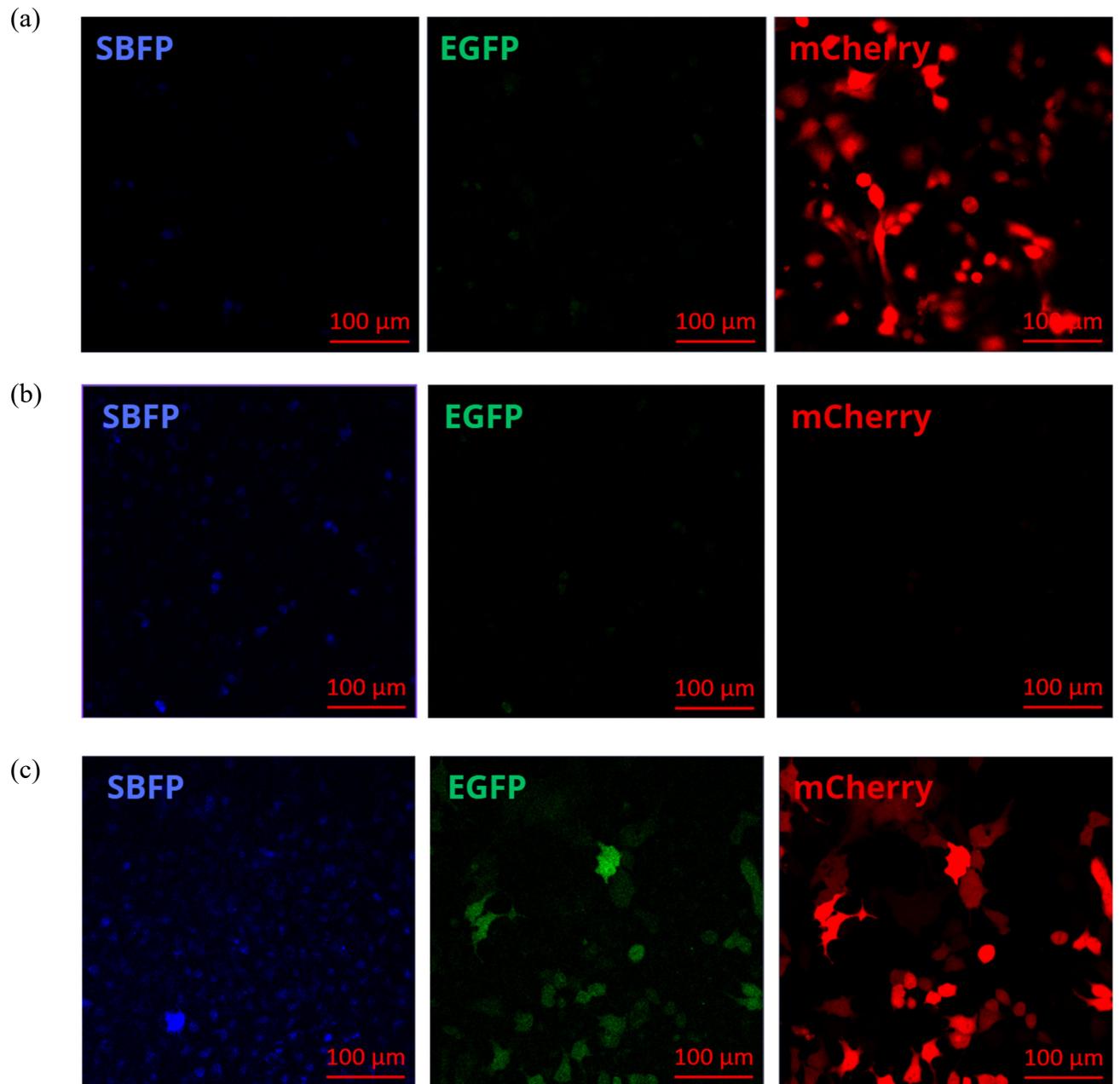


Figure S2. Representative confocal images of HeLa cells treated either with (a) p_egfp1-10_inteinN, (b) p_inteinC_egfp11, or (c) both plasmids simultaneously.

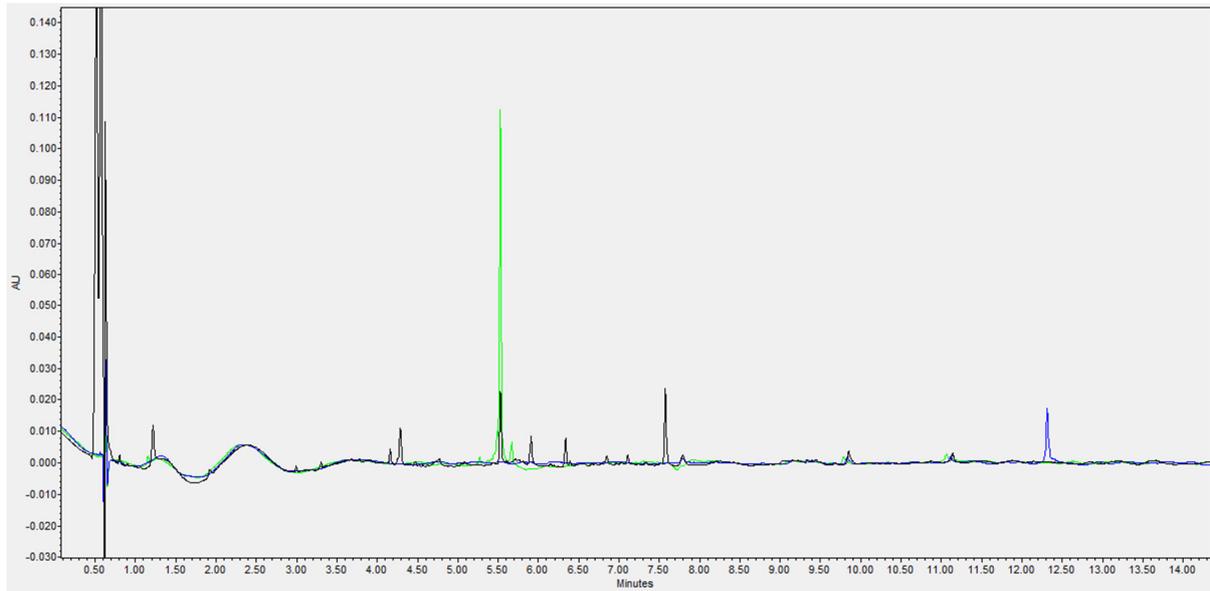


Figure S3. UPLC image. The green line is the intein^C peptide, the blue line is the NF55-XK(Rhod)XPra peptide, and the black line is the purified click reaction mixture. Note the reduction in the amount of intein^C, which was initially in excess compared to NF55-XK(Rhod)XPra, and the disappearance of NF55-XK(Rhod)XPra from the reaction mixture. Furthermore, the emergence of a new compound (NF55-XK(Rhod)X–intein^C conjugate) can be observed.

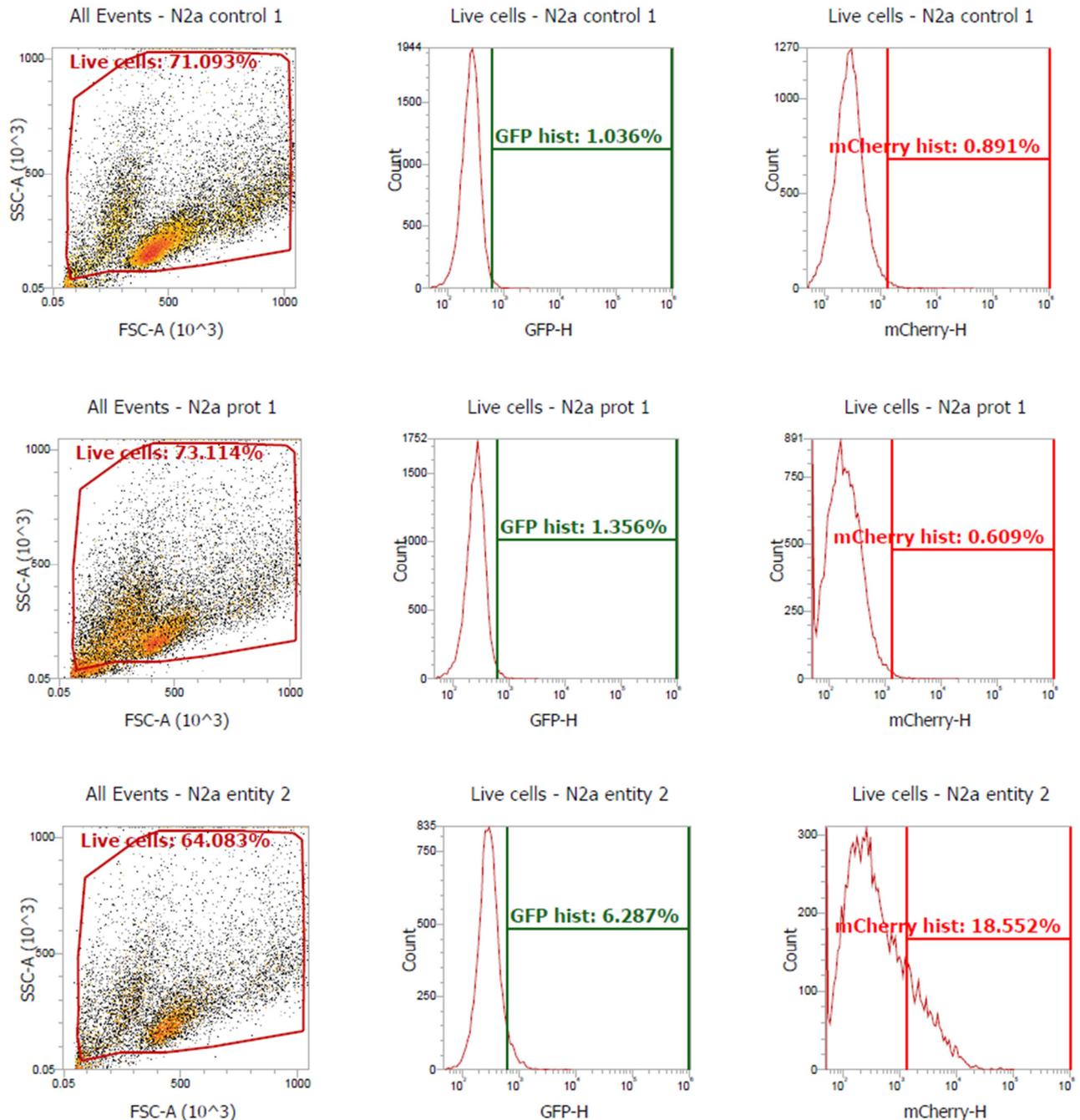


Figure S4. Representative flow cytometry plots of Neuro 2a cells transduction efficacy assay. Flow cytometry was performed using an Attune™ NxT Flow Cytometer equipped with a 488 nm argon laser. The viable cell population was determined from a plot of forward-scattered (FSC) vs. side-scattered (SSC) light. Attune™ NxT Software 3.2.1 software was used to analyze a minimum of 10,000 events per sample. The area scaling factor was 1.28, with a threshold of SSC 0.3, FSC 220, SSC 360, BL1/GFP 460, and RL1/APC/Cy5 520. GFP+ and mCherry show gated events with a signal intensity over the threshold of ~1% of untreated cells.

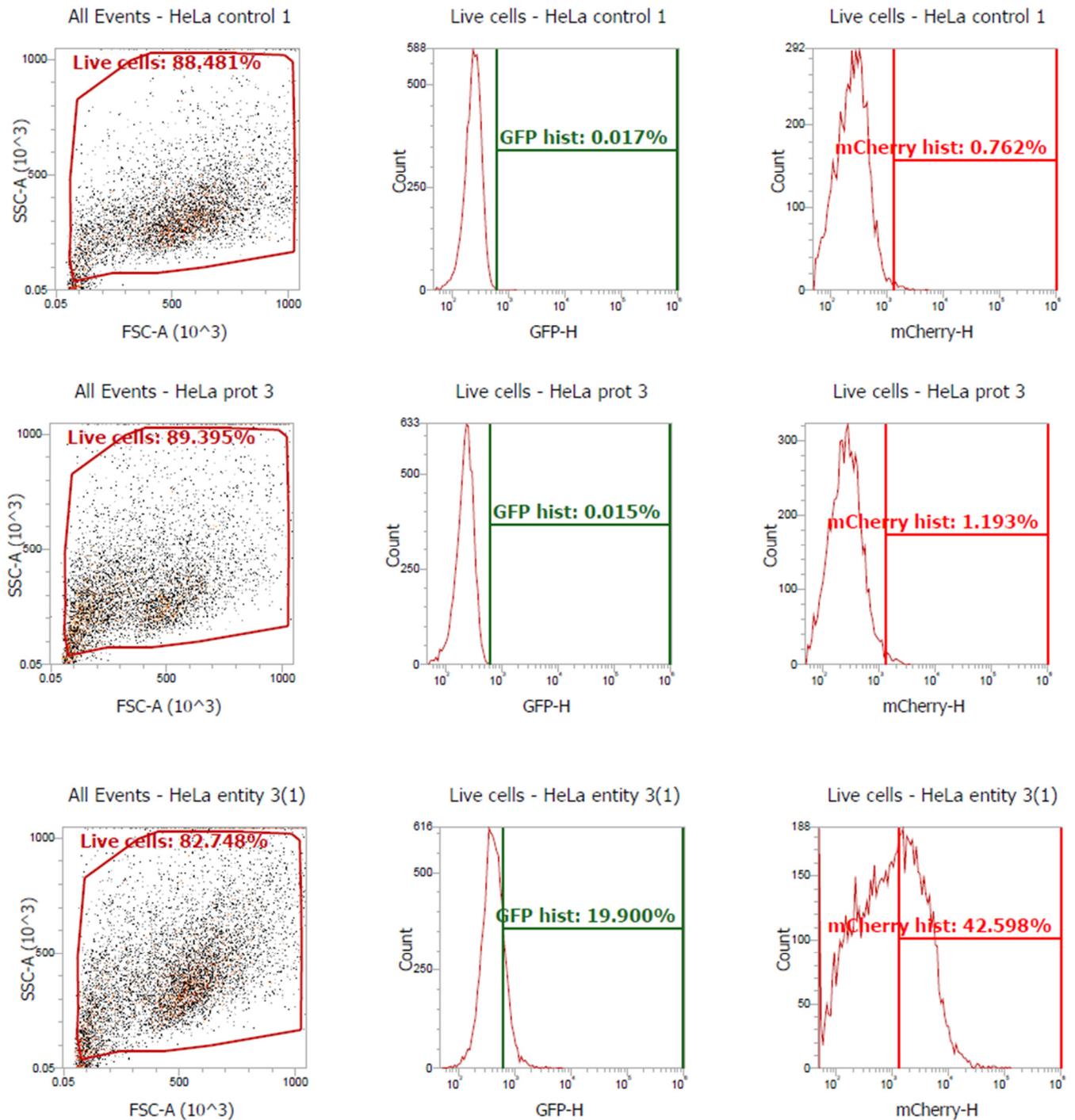


Figure S5. Representative flow cytometry plots of HeLa cells transduction efficacy assay. Flow cytometry was performed using an Attune™ NxT Flow Cytometer equipped with a 488 nm argon laser. The viable cell population was determined from a plot of forward-scattered (FSC) vs. side-scattered (SSC) light. Attune™ NxT Software 3.2.1 software was used to analyze a minimum of 10,000 events per sample. The area scaling factor was 1.28, with a threshold of SSC 0.3, FSC 220, SSC 360, BL1/GFP 460, and RL1/APC/Cy5 520. GFP+ and mCherry show gated events with signal a intensity over the threshold of ~1% of untreated cells.

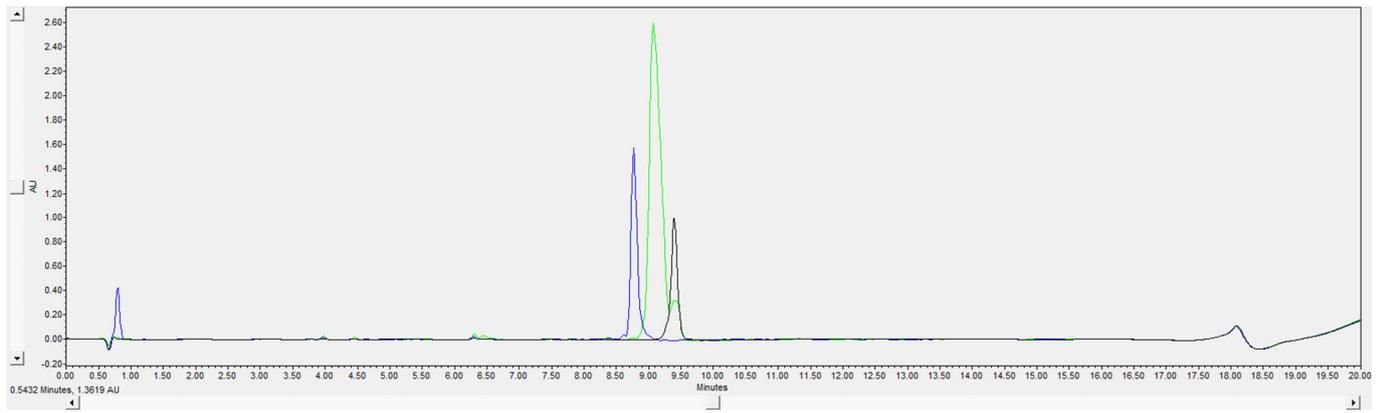


Figure S6. UPLC image. The blue line is the EGFP–intein^N protein, the green line is the reaction mixture of EGFP–intein^N protein and intein^C-activated NF55 peptide, and the black line is the intein^C-activated NF55 peptide. In the reaction mixture, no protein or peptide peaks are registered.

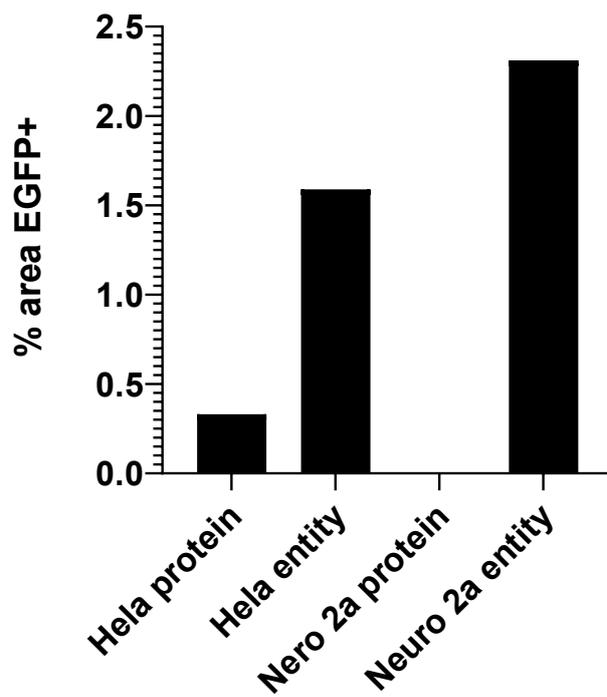


Figure S7. Confocal image analysis showing average percentage of EGFP-positive area. Analysis is performed with ImageJ (Version 1.53a), National Institute of Health, Bethesda, Maryland, USA.