



- 2 Supplemental Information
- 3

Isolation, Structure Determination, and Synthesis of Cyclic Tetraglutamic Acids from the Box
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#### 39 Section S1. Synthesis of DLLL cyclic tetraglutamic acid cnidarin 4B (2)

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#### Scheme S1. Preparation of cnidarin 4B (2). 41



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44 5-Benzyl 1-(tert-butyl) ((R)-5-(tert-butoxy)-4-((tert-butoxycarbonyl)amino)-5-oxopentanoyl)-L-glutamate 45 (S2)

46 To a solution of S1 (1.03 g, 3.4 mmol) in dichloromethane (15 mL), DMAP (200 mg, 1.7 mmol), 47 EDCI (1.3 g, 6.8 mmol) and 10 (1 g, 3.4 mmol) were added, and the reaction mixture was stirred at 48 room temperature for 2 hr. H2O was added, and the organic layer was extracted with 49 dichloromethane. The extract was dried over MgSO4, filtered, and concentrated in vacuo. The residue 50 was purified by column chromatography to give S2 (1.6 g, 2.76 mmol) in 81% yield.

51

52 5-Benzyl 1-(tert-butyl) ((R)-4-amino-5-(tert-butoxy)-5-oxopentanoyl)-L-glutamate (S3)

53 To a solution of S2 (0.9 g, 1.55 mmol) in CH<sub>3</sub>CN-H<sub>2</sub>O (50:1), BiCl<sub>3</sub> was added portionwise (974.4 54 mg, 3.1 mmol) to selectively deprotect the Boc group. NaHCO<sub>3</sub> was added, and the reaction mixture 55 was filtered through a Celite pad to give S3 (0.83 g, 1.73 mmol) in 63% yield.

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57
       23-Benzyl 11,16,21,6-tetra-tert-butyl (6R,11S,16R,21R)-2,2-dimethyl-4,9,14,19-tetraoxo-3-oxa-5,10,15,20-
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58 tetraazatricosane-6,11,16,21,23-pentacarboxylate (S4)

59 To a solution of amine 14 (732 mg, 1.53 mmol) in dichloromethane, EDCI (585 mg, 3.06 mmol), 60 DMAP (93 mg, 0.765 mmol) and S3 (749 mg, 1.53 mmol) were added, and the mixture was stirred at 61 room temperature for 2 hr. H2O was added to the reaction mixture, and the organic layer was 62 extracted with dichloromethane. The extracts were dried over MgSO4, filtered, and concentrated in 63 vacuo. The residue was purified by column chromatography to give S4 (1.18 g, 1.24 mmol) in 81% 64 vield.

## 66 2.1,5.4-Anhydro( $\gamma$ -L-glutamyl- $\gamma$ -L-glutamyl- $\gamma$ -D-glutamyl-L-glutamic acid) (cnidarin 4B, 2)

67 To a solution of S4 (1.18 g, 1.24 mmol) in CH<sub>3</sub>CN-H<sub>2</sub>O (50:1), BiCl<sub>3</sub> was added portionwise (782 68 mg, 2.48 mmol) to selectively deprotect the Boc group. NaHCO<sub>3</sub> was added and the reaction mixture 69 was filtered through a Celite pad, and filtrates were concentrated in vacuo. The reaction product was 70 further purified by column chromatography to gave the corresponding linear LLDL-tetraglutamate 71 (390 mg, 0.459 mmol) in 37% yield:  $[\alpha]_D^{25}$  +2.9° (c 0.68, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.34 (S, 72 5H), 7.15 (s, 1H), 7.13 (s, 1H), 5.10 (s, 2H), 4.51-4.38 (m, 3H), 3.71-3.65 (m, 1H), 2.61-2.41 (m, 4H), 2.4-73 2.24 (m, 4H), 2.23-2.11 (m, 4H), 2.05-1.89 (m, 4H), 1.51-1.39 (m, 36H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 74 172.9, 172.5, 172.3, 172.2, 171.1, 136.6, 128.5, 128.2, 82.2, 82.0, 66.4, 52.4, 52.1, 32.4, 30.4, 28.2, 27.9, 27.2 75 ppm; HRMS-ESI *m/z* 871.4708 [M+Na]<sup>+</sup> (calcd for C<sub>43</sub>H<sub>68</sub>N<sub>4</sub>O<sub>13</sub>Na<sup>+</sup> *m/z* 871.4680, Δ +2.8 mmu. See 76 Figures S34 and S35 for 1H and 13C NMR spectra. To this product (192 mg, 0.22 mmol) in THF, Pd/C 77 (20 mg) was added, and H<sub>2</sub> was exposed to the mixture to remove the benzyl group. The reaction 78 mixture was filtered through a Celite pad and concentrated in vacuo. The product concentrate (161 79 mg, 0.212 mmol) was subjected to macrocylization in the presence of MNBA (109 mg, 0.318 mmol), 80 DMAP (2.6 mg, 0.0212 mmol), and Et<sub>3</sub>N (176. 3 mL, 1.272 mmol) to give the protected cyclic LLDL-81 glutamic acid. Finally, the Boc group was deprotected using TFA to give cnidarin 4B (2) (78 mg, 82 0.152 mmol) in 69% yield in three steps. [α]<sup>25</sup><sub>D</sub> -15° (*c* 0.94, MeOH); <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O) δ 4.42-83 4.28 (m, 4H), 2.52-2.32 (m, 8H), 2.29-2.11 (m, 4H), 2.09-1.89 (m, 4H); <sup>13</sup>C NMR (75 MHz, D<sub>2</sub>O) & 175.9, 84 175.7, 175.6, 175.4, 53.6, 53.1, 52.7, 52.1, 32.7, 32.3, 31.8, 31.2, 26.7, 26.5, 26.2 ppm; HRMS-ESI *m*/*z* 85 539.1542 [M+Na]<sup>+</sup> (calcd for C<sub>20</sub>H<sub>28</sub>N<sub>4</sub>O<sub>12</sub>Na<sup>+</sup> m/z 539.1601,  $\Delta$  -5.9 mmu). See Figures S36 and S37 for 86 <sup>1</sup>H and <sup>13</sup>C NMR spectra.

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### 89 Section S2. Synthesis of DLDL cyclic tetraglutamic acid iso-cnidarin 4A (4)

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#### 91 Scheme S2. Preparation iso-cnidarin 4A (4).

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97 (S)-5-(Benzyloxy)-4-((R)-5-(benzyloxy)-4-((tert-butoxycarbonyl)amino)-5-oxopentanamido)-5-oxopentanoic98 acid (S5)

99 To a solution of 5 (1.808 g, 5.36 mmol) and N-hydroxysucinimide (0.864 g, 7.50 mmol) in 100 dichloromethane (26.8 mL) at room temperature EDCI (1.28 g, 6.70 mmol) was added. The reaction 101 was monitored by TLC and after 18 hr was diluted with dichloromethane (100 mL) and washed 3 × 102 with saturated KH<sub>2</sub>PO<sub>4</sub>. The combined aqueous layers were back extracted with dichloromethane, 103 and the combined organic layers were washed with saturated NaCl, dried over MgSO4 and 104 concentrated, providing the crude NHS ester (2.52 g, quant) as a white powder, deemed suitable for 105 use in the subsequent reaction. To a solution of the crude NHS ester (1.566 g, 3.61 mmol) and 106 triethylamine (0.704 mL, 0.511g, 5.05 mmol) in dichloromethane (36 mL) at room temperature, 1-107 benzyl L-glutamic acid (0.984 g, 4.15 mmol) was added. The reaction was monitored by TLC and after 108 22 hr was diluted with dichloromethane (114 mL) and washed 4 × with saturated KH<sub>2</sub>PO<sub>4</sub>. The 109 combined aqueous layers were back extracted with dichloromethane and the combined organic 110 layers were washed with saturated NaCl, dried over MgSO4 and concentrated. The crude product 111 was purified by flash chromatography on silica gel, (2% to 4% methanol in dichloromethane with 112 0.5% acetic acid acid) to provide S5 (1.91 g, 95% (two step yield)) as a white powder: mp 89 - 91 °C; 113 Rf = 0.4 (5% methanol in dichloromethane with 0.5% acetic acid); <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>OD) δ 7.36114 7.26 (m, 10H), 5.13 (s, 2H), 5.15 (s, 2H), 4.44 (dd, J = 6.0, 9.0, 1H), 4.15 (dd, J = 3.0, 9.0, 1H), 2.38-2.27 (m, 115 4H), 2.19-2.02 (m, 2H), 1.98-1.81 (m, 2H), 1.41 (s, 9H); <sup>13</sup>C NMR (300 MHz, CD<sub>3</sub>OD) δ 174.8, 173.7, 116 172.4, 171.7,156.9, 136.0, 136.0, 128.4, 128.3, 128.1, 128.1, 128.0, 79.6, 66.9, 66.8, 53.7, 52.2, 31.9, 30.0, 27.7, 117 27.4, 26.5; IR 3362 (br), 2968 (m), 1722 (m), 1654 (s), 1519 (m), 1451 (s) cm<sup>-1</sup>; TOF-MS *m*/z 579.2306 118 [M+Na]<sup>+</sup> (C<sub>29</sub>H<sub>36</sub>N<sub>2</sub>O<sub>9</sub>Na<sup>+</sup> requires 579.2319, Δ -1.3 mmu). See Figures S20 and S21 for <sup>1</sup>H and <sup>13</sup>C NMR 119 spectra.

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 $121 \qquad (6R,11S,16R)-6,11,16-Tris((benzyloxycarbonyl)-2,2-dimethyl-4,9,14-trioxo-3-oxa-5,10,15-triazanonadecan-10,12-triazanonadecan$ 

122 19-oic acid (**S6**)

123 To a solution of S5 (2.641 g, 4.75 mmol) and N-hydroxysucinimide (0.765 g, 6.64 mmol) in 124 dichloromethane (24 mL) at room temperature, EDCI (1.14 g, 5.93 mmol) was added. The reaction 125 was monitored by TLC and after 18 hr was diluted with dichloromethane (76 mL) and washed 4 × 126 with saturated KH2PO4. The combined aqueous layers were back extracted with dichloromethane 127 and the combined organic layers were washed with saturated NaCl, dried over MgSO4 and 128 concentrated, providing the crude NHS ester (3.19 g, quant) as a white foam deemed suitable for use 129 in the subsequent reaction. To a solution of the crude NHS ester (1.48 g, 2.27 mmol) and 130 triethylamine (0.443 mL, 0.321 g, 3.17 mmol) in dichloromethane (22.7 mL) at room temperature, 1-131 benzyl D-glutamate (0.619 g, 2.61 mmol) was added. The reaction was monitored by TLC and after 132 28 hr was diluted with dichloromethane (177 mL) and washed 4 × with saturated KH<sub>2</sub>PO<sub>4</sub>. The 133 combined aqueous layers were back extracted with dichloromethane and the combined organic 134 layers were washed with saturated NaCl, dried over MgSO4 and concentrated. The crude product 135 was purified by flash chromatography on silica gel, (5% methanol in dichloromethane with 0.5% 136 acetic acid) to provide S6 (1.62 g, 93% (two step yield)) as a white foam: Rf = 0.33 (5% methanol in 137 dichloromethane with 0.5% acetic acid); <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>OD) & 7.38-7.25 (m, 15H), 5.17-5.12 138 (m, 6H), 4.52-4.39 (m, 2H), 4.17 (dd, J = 6.0, 9.0, 1H), 2.40-2.26 (m, 6H), 2.21-2.03 (m, 3H), 2.01-1.82 (m, 2H), 2.01-1.82 (139 3H), 1.41 (s, 9H); <sup>13</sup>C NMR (300 MHz, CD<sub>3</sub>OD) & 174.8, 173.7, 173.6, 172.4, 171.7, 171.6, 156.8, 136.0, 140 136.0, 128.35, 128.3, 128.3, 128.1, 128.0, 128.0, 128.0, 79.6, 66.9, 66.8, 53.7, 52.5, 52.3, 31.9, 31.8, 30.0, 27.7, 141 27.4, 27.2, 26.5; IR 3321 (br), 3046 (m), 2953 (m), 2356 (s), 1737 (s), 1654 (s), 1529 (s) cm<sup>-1</sup>; TOF-MS *m/z* 142 798.3242 [M+Na]+ (C<sub>41</sub>H<sub>49</sub>N<sub>3</sub>O<sub>12</sub>Na+ requires 798.3214,  $\Delta$  +2.8 mmu). See Figures S22 and S23 for <sup>1</sup>H 143 and <sup>13</sup>C NMR spectra.

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145 (6S,11R,16S,21R)-6,11,16,21-Tetrakis((benzyloxy)carbonyl)-2,2-dimethyl-4,9,14,19-tetraoxo-3-oxa-

146 *5,10,15,20-tetraazatetracosan-24-oic acid* (**S7**)

147 To a solution of S6 (1.489 g, 1.92 mmol) and N-hydroxysucinimide (0.309 g, 2.69 mmol) in 148 dichloromethane (19.2 mL), EDCI (0.460 g, 2.40 mmol) was added. The reaction was monitored by 149 TLC and after 27 hr was diluted with dichloromethane (181 mL) and washed 4 × with saturated 150 KH<sub>2</sub>PO<sub>4</sub>. The combined aqueous layers were back extracted with dichloromethane and the combined 151 organic layers were washed with saturated NaCl, dried over MgSO4 and concentrated, providing the 152 crude NHS ester (1.73 g, quant) as a white foam deemed suitable for use in the subsequent reaction. 153 To a solution of the crude (1.537 g, 1.76 mmol) and triethylamine (0.343 mL, 0.249 g, 2.46 mmol) in 154 dichloromethane (17.6 mL) at room temperature, 1-benzyl D-glutamate (0.480 g, 2.02 mmol) was 155 added. The reaction was monitored by TLC and after 22 hr was diluted with dichloromethane (183 156 mL) and washed 4 × with saturated KH2PO4. The combined aqueous layers were back extracted with 157 dichloromethane and the combined organic layers were washed with saturated NaCl, dried over 158 MgSO4 and concentrated. The crude product was further purified by flash chromatography on silica 159 gel (eluting with 4% methanol in dichloromethane and 0.5% acetic acid) to provide S7 (1.60 g, 91% 160 (two step yield)) as a white foam: Rf = 0.35 (5% methanol in dichloromethane with 0.5% acetic acid); 161 <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>OD) δ 7.40-7.26 (m, 20H), 5.18-5.13 (m, 8H), 4.51-4.39 (m, 3H), 4.18 (dd, J = 162 3.0,6.0, 1H), 2.41-2.26 (m, 8H), 2.23-2.06 (m, 4H), 2.03-1.84 (m, 4H), 1.43 (s, 9H); <sup>13</sup>C NMR (300 MHz, 163 CD<sub>3</sub>OD) & 174.8, 173.7, 173.6, 172.4, 171.8, 171.7, 171.6, 156.8, 136.0, 135.9, 128.4, 128.4, 128.1, 128.0, 164 128.0, 128.0, 79.7, 66.9, 66.8, 53.8, 52.5, 52.4, 52.3, 31.9, 31.8, 31.8, 30.0, 27.7, 27.4, 27.3, 27.1, 26.5; IR 3321 165 (br), 3061 (m), 2968 (m), 1737 (s), 1654 (s), 1529 (s) cm<sup>-1</sup>; TOF-MS *m/z* 1017.4101 [M+Na]<sup>+</sup> 166 (C53H62N4O15Na<sup>+</sup> requires 1017.4109,  $\Delta$  -0.8 mmu). See Figures S24 and S25 for <sup>1</sup>H and <sup>13</sup>C NMR 167 spectra.

168

169 Tetrabenzyl (2S,7R,12S,17R)-5,10,15,20-tetraoxo-1,6,11,16-tetraazacycloicosane-2,7,12,17-tetracarboxylate
 170 (S8)

171 To a solution of S7 (0.495 g, 0.498 mmol) in dichloromethane (25 mL) at 0 °C, a pre-cooled 1:1 172 solution of trifluoroacetic acid and dichloromethane (25 mL) was added portionwise; 5.0 mL were 173 added every 5 min (5 X) for a total of 25 mL over 25 min. The reaction was monitored TLC by 174 conducting "mini workups" (drying under nitrogen gas followed by reconstituting with 175 dichloromethane,  $3 \times in$  total). After 1.5 hr the solvent was removed by rotary-evaporation, and the 176 crude was reconstituted in dichloromethane followed again by evaporation (3 × in total). The crude 177 was purified by flash chromatography on silica gel (4% to 10% methanol in dichloromethane) to 178 provide the free ammonium trifluoroacetate (0.355 g, 71%) as a brown foam: Rf = 0.24 (8% methanol 179 in dichloromethane); <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>OD) δ7.41-7.22 (m, 20H), 5.245 (d, J=3.0, 2H), 5.15-5.10 180 (m, 6H), 4.49-4.36 (m, 3H), 4.11 (t, J=12.0, 1H), 2.45 (t, J=15.0, 2H), 2.37-2.24 (m, 6H), 2.23-2.02 (m, 5H), 181 2.00-1.81 (m, 3H); <sup>13</sup>C NMR (300 MHz, CD<sub>3</sub>OD) δ 174.8, 173.6, 173.5, 172.9, 171.7, 171.6, 186.8, 135.9, 182 135.9, 135.1, 128.6, 128.5, 128.4, 128.4, 128.3, 128.2, 128.1, 128.01, 128.0, 68.2, 67.0, 66.9, 66.9, 52.5, 52.3 183 31.7, 30.8, 29.9, 27.1, 26.5, 25.9; IR 3310 (br), 3046 (m), 2947 (m), 1737 (s), 1659 (s), 1540 (s) cm<sup>-1</sup>. To a 184 solution of the free ammonium trifluoroacetate (0.151 g, 0.149 mmol) and triethylamine (6.36 µL, 4.61 185 mg, 0.456 mmol) in dichloromethane (152 mL), EDCI (0.0874 g, 0.456 mmol) was added. The reaction 186 was monitored by TLC and after 17 hr was diluted with dichloromethane (75 mL) and washed 4 × 187 with saturated KH<sub>2</sub>PO<sub>4</sub>. The combined aqueous layers were back extracted with dichloromethane, 188 and the combined organic layers were washed with saturated NaCl, dried over MgSO4 and 189 concentrated. The crude product was purified by recrystallization according to the following 190 procedure. The crude sample of was dissolved in warm dichloromethane, filtered through cotton in 191 a glass funnel followed by the addition of a small volume of hexane. The solution was placed in a 192 fume hood overnight and then in a freezer for 4 days providing fine crystals. The suspension was 193 centrifuged, and the supernatant removed followed by 2 hexane-wash / centrifugation cycles 194 providing S8 (0.052 g, 40%) as a white powder: mp 248 - 250 °C; 1H NMR (300 MHz, CDCl3) 87.38-195 7.27 (m, 20H), 7.17 (d, J=6.0, 4H), 5.19-5.08 (m, 8H), 4.70-4.60 (m, 4H), 2.48-2.35 (m, 4H), 2.26-2.05 (m, 196 12H); <sup>13</sup>C NMR (300 MHz, CDCl<sub>3</sub>) & 173.1, 173.1, 171.0, 135.4, 128.8, 128.6, 128.3, 67.5, 53.4, 53.3, 33.3, 197 27.6, 27.5; IR 3290 (br), 3072 (m), 2937 (m), 1732 (s), 1649 (s), 1550 (s) cm<sup>-1</sup>; TOF-MS m/z 899.3461 198  $[M+Na]^{+}$  (C<sub>48</sub>H<sub>52</sub>N<sub>4</sub>O<sub>12</sub>Na<sup>+</sup> requires 899.3479,  $\Delta$  -1.8 mmu). See Figures S26 and S27 for <sup>1</sup>H and <sup>13</sup>C NMR 199 spectra.

### 201 2.1,5.4-Anhydro( $\gamma$ -L-glutamyl- $\gamma$ -D-glutamyl- $\gamma$ -L-glutamyl-D-glutamic acid) (iso-cnidarin 4A, 4)

202 Above a solution of **S8** (0.0101 g, 0.0115 mmol) and 10% Pd/C (~2 mg) in ethanol (5.0 mL) at room 203 temperature, hydrogen gas was maintained at ambient pressure for 21 hr. The reaction was then 204 filtered through celite followed by methanol washing (3 × 1 mL), methanol/water 1:1 (3 × 1 mL) then 205 water (3  $\times$  1 mL). The crude product was purified by HPLC using a Waters Atlantis dC18 (10  $\times$  250 206 mm, 10 µm particle size) column (mobile phase A: water/formic acid (1000:1); mobile phase B: 207 acetonitrile/formic acid (1000:1); gradient: 2.5% B, 0-7 min, 2.5–100% B, 7-27 min, 100% B, 27-35 min) 208 to provide **4** (0.042 g, 71%) as a white solid: decomp = 277 - 283 °C; <sup>1</sup>H NMR (500 MHz, D<sub>2</sub>O) δ4.295 209 (dd, J=5.0,10, 4H), 2.49-2.40 (m, 4H), 2.37-2.28 (m, 4H), 2.19-2.10 (m, 4H), 2.06-1.95 (m, 4H); <sup>13</sup>C NMR 210 (500 MHz, D<sub>2</sub>O) δ 178.3, 177.9, 55.8, 34.9, 29.0; IR 3312 (br), 2358 (m), 1644 (m), 1594 (s), 1410 (br) cm<sup>-1</sup>; 211 TOF-MS *m*/*z* 539.1602 [M+Na]<sup>+</sup> (C<sub>20</sub>H<sub>28</sub>N<sub>4</sub>O<sub>12</sub>Na<sup>+</sup> requires 539.1601, Δ +0.1 mmu). See Figures S28 and 212 S29 for <sup>1</sup>H and <sup>13</sup>C NMR spectra. 213



Section S3. Chromatography and chemical analyses of cnidarin 4A (1)

**Figure S1**. LC-DAD-MS of *Alatina alata* venom. Chromatograms shown from top to bottom panel are: photo diode array detector (PDA) UV absorption (200-600 nm), MS base peak (*m*/*z* 100-1000), extracted ion chromatograms for *m*/*z* 517 ion and MS/MS product ions for *m*/*z* 517. Extract was analyzed on a Waters Atlantis dC18 (3.0 x 250 mm column, 5 µm particle size) column (mobile phase A: water/formic acid (1000:1); mobile phase B: acetonitrile/formic acid (1000:1); gradient: 0% B, 0-10 minutes, 0–50% B, 10-20 minutes, 50-100% B, 20-23 minutes, 100% B, 23-25 minutes). The peak at 18.6 min in extracted ion ([M+H]<sup>+</sup> *m*/*z* 517) and MS/MS chromatograms (bottom 2 panels) was a mixture resolved with LC conditions described in Figure S2.





**Figure S2.** Extracted  $[M+H]^+$  ion (*m*/*z* 517) and MS/MS chromatograms of crude cnidarin 4A (1) (see peak at 18.6 min in Figure S1) from *Alatina alata* venom. Sample was reconstituted in 1% formic acid analyzed on a Waters Atlantis dC18 column (3.0 x 250 mm, 5 µm particle size) (mobile phase A: water/formic acid (1000:1); mobile phase B: acetonitrile/formic acid (1000:1); gradient: 2.5% B, 0-7 minutes, 2.5–100% B, 7-27 minutes, 100% B, 27-35 minutes). Compound **1** eluted at 9.4 min.

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Figure S3. Capillary <sup>1</sup>H nuclear magnetic resonance (NMR) spectrum (500 MHz, D<sub>2</sub>O/H<sub>2</sub>O)
 of crude cnidarin 4A (1). Spectral data indicate multiple glutamic acids.



**Figure S4.** LC-MS of FDLA-labeled products of the compound **1** hydrolysate, Lglutamic acid, and DL-glutamic acid. Chromatograms are summed extracted ion chromatograms of *m*/*z* of 442 and 883 ions, which correspond to protonated glutamic acid-FDLA monomer and dimer ions. Products were analyzed using a Phenomenex Luna C18(2) (2.0 x 250 mm, 3 μm particle size) column (mobile phase A: water/formic acid (1000:1); mobile phase B: acetonitrile/formic acid (1000:1); gradient: 20-80% B, 0-30 minutes, 80% B, 30-35 minutes).



Figure S5. HPLC with PDA detection (200-600 nm) of FDLA-labeled products of compound
1 hydrolysate. (UV chromatograms from Figure S4). Products were analyzed on a
Phenomenex Luna C18(2) (2.0 × 250 mm, 3 µm particle size) column (mobile phase A:
water/formic acid (1000:1); mobile phase B: acetonitrile/formic acid (1000:1); gradient: 2080% B, 0-30 min, 80% B, 30-35 min).

255

256**Table S1.** Ratio of glutamic acid-FDLA derivatives of hydrolysate257of compound 1 using HPLC with MS and UV detection.

Detector	relative concentration		
Detector	D-glutamic acid	L-glutamic acid	
UV	0.86	1.0	
MS	0.99	1.0	

Ratios were calculated from extracted ion chromatograms, see Figures S4 and S5.

259 260



m/z

**Figure S6.** High resolution time-of-flight mass spectrometry (TOF-MS-ESI positive mode) of compound **1** from *Alatina alata* venom.



**Figure S7.** LC-DAD-MS of *Alatina alata* tentacle extract without nematocysts (panels from top to bottom: photo diode array UV detector, MS base peak, extracted ion chromatograms for the  $[M+H]^+$  ion, *m*/*z* 517, and MS/MS product ions for *m*/*z* 517).

![](_page_12_Figure_1.jpeg)

![](_page_12_Figure_2.jpeg)

**Figure S8.** Calibration curve for MS/MS detection of cnidarin 4A (1) (product ion of m/z 517 fragmentation). Synthesized **1** was analyzed at semi log concentrations in triplicate with a fixed volume injection (0.343 – 343 ng on column). LC-MS conditions: Waters Atlantis dC18 column (3.0 × 250 mm, 5 µm particle size); mobile phase A: water/formic acid (1000:1); mobile phase B: acetonitrile/formic acid (1000:1); gradient: 2.5% B, 0-7 min, 2.5–100% B, 7-27 min, 100% B, 27-35 min).

![](_page_13_Figure_1.jpeg)

![](_page_13_Figure_2.jpeg)

![](_page_13_Figure_3.jpeg)

Figure S9. Cytotoxicity of doxorubicin, synthetic cnidarin 4A (1) and *iso*-cnidarin 4A (4)
against HEK-293 cells. Doxorubicin is a positive control. Methods described in article.

![](_page_13_Figure_5.jpeg)

Table S2. Hemolysis of red blood cells by cnidarin 4A (1)	1
normalized to 1% TritonX-100 hemolysis.	

treatment	% hemoglobin	Standard deviation
vehicle	2.63	0.31
24 µM cmpd <b>1</b>	3.74	1.04
121 μM cmpd <b>1</b>	2.86	0.20
TritonX-100, 1%	100	0.28

![](_page_13_Figure_9.jpeg)

Treatments were tested in triplicate.

![](_page_14_Figure_1.jpeg)

![](_page_14_Figure_2.jpeg)

**Figure S10.** <sup>1</sup>H NMR (CD<sub>3</sub>OD) spectrum for **6**.

![](_page_14_Figure_6.jpeg)

**Figure S11.** <sup>13</sup>C NMR (CD<sub>3</sub>OD) spectrum for **6**.

![](_page_15_Figure_1.jpeg)

![](_page_15_Figure_2.jpeg)

Figure S12. <sup>1</sup>H NMR (CD<sub>3</sub>OD) spectrum for 7.

![](_page_15_Figure_4.jpeg)

304 **Figure S13.** <sup>13</sup>C NMR (CD<sub>3</sub>OD) spectrum for 7.

![](_page_16_Figure_1.jpeg)

![](_page_16_Figure_2.jpeg)

Figure S14. <sup>1</sup>H NMR (CD<sub>3</sub>OD) spectrum for 8.

![](_page_16_Figure_4.jpeg)

308 Figure S15. <sup>13</sup>C NMR (CD<sub>3</sub>OD) spectrum for 8.

![](_page_17_Figure_1.jpeg)

![](_page_17_Figure_2.jpeg)

**Figure S16.** <sup>1</sup>H NMR (CDCl<sub>3</sub>) spectrum for **9**.

![](_page_17_Figure_4.jpeg)

**Figure S17.** <sup>13</sup>C NMR (CDCl<sub>3</sub>) spectrum for **9**.

![](_page_18_Figure_1.jpeg)

![](_page_18_Figure_2.jpeg)

316 **Figure S18.** <sup>1</sup>H NMR (D<sub>2</sub>O) spectrum for **1**.

![](_page_18_Figure_4.jpeg)

318 Figure S19. <sup>13</sup>C NMR (D<sub>2</sub>O) spectrum for 1.

# 320 Section S6. NMR spectra of synthetic *iso*-cnidarin 4A (4) and intermediates S5-

- **S8.**

![](_page_19_Figure_4.jpeg)

**Figure S20.** <sup>1</sup>H NMR (CD<sub>3</sub>OD) spectrum for **S5**.

![](_page_19_Figure_6.jpeg)

**Figure S21.** <sup>13</sup>C NMR (CD<sub>3</sub>OD) spectrum for **S5**.

![](_page_20_Figure_1.jpeg)

![](_page_20_Figure_2.jpeg)

Figure S22. <sup>1</sup>H NMR (CD<sub>3</sub>OD) spectrum for S6.

![](_page_20_Figure_4.jpeg)

331 **Figure S23.** <sup>13</sup>C NMR (CD<sub>3</sub>OD) spectrum for **S6**.

332

![](_page_21_Figure_1.jpeg)

![](_page_22_Figure_1.jpeg)

![](_page_22_Figure_2.jpeg)

Figure S26. <sup>1</sup>H NMR (CDCl<sub>3</sub>) spectrum for S8.

![](_page_22_Figure_4.jpeg)

![](_page_22_Figure_5.jpeg)

Figure S27. <sup>13</sup>C NMR (CDCl<sub>3</sub>) spectrum for S8.

![](_page_23_Figure_1.jpeg)

![](_page_23_Figure_2.jpeg)

Figure S28. <sup>1</sup>H NMR (D<sub>2</sub>O) spectrum for 4.

![](_page_23_Figure_4.jpeg)

![](_page_23_Figure_5.jpeg)

348

![](_page_24_Figure_2.jpeg)

#### Section S7. NMR spectra of synthetic cnidarin 4C (3) and intermediates. 350

![](_page_24_Figure_4.jpeg)

353

![](_page_24_Figure_6.jpeg)

![](_page_24_Figure_7.jpeg)

![](_page_25_Figure_1.jpeg)

![](_page_25_Figure_2.jpeg)

 $357 \qquad \mbox{Figure S32. $^1$H NMR (D_2O) spectrum for 3.}$ 

![](_page_25_Figure_4.jpeg)

**Figure S33.** <sup>13</sup>C NMR (D<sub>2</sub>O) spectrum for **3**; 1,4-dioxane added as internal standard.

![](_page_26_Figure_2.jpeg)

361 Section S8. NMR spectra of synthetic cnidarin 4B (2) and intermediates.

362 363

Figure S34. <sup>1</sup>H NMR (CDCl<sub>3</sub>) spectrum for linear LLDL-glutamic acid with free amine.

![](_page_26_Figure_6.jpeg)

![](_page_26_Figure_7.jpeg)

365 366

**Figure S35.** <sup>13</sup>C NMR (CDCl<sub>3</sub>) spectrum for linear LLDL-glutamic acid with free amine.

![](_page_27_Figure_2.jpeg)

![](_page_27_Figure_3.jpeg)

368

Figure S36. <sup>1</sup>H NMR (D<sub>2</sub>O) spectrum for 2.

![](_page_27_Figure_6.jpeg)

![](_page_27_Figure_7.jpeg)

Figure S37. <sup>13</sup>C NMR (D<sub>2</sub>O) spectrum for **2**; 1,4-dioxane added as internal standard.

![](_page_28_Figure_1.jpeg)

374 Figure S38. The <sup>1</sup>H NMR (800 MHz, D<sub>2</sub>O with 10 μL HCOOH) spectra of isolated cnidarins 4A (1,

- 375 bottom), 4B (**2**, middle), 4C (**3**, top).
- 376