

Article

Selenium Alleviates the Adverse Effect of Drought in Oilseed Crops Camelina (*Camelina sativa* L.) and Canola (*Brassica napus* L.)

Zahoor Ahmad ^{1,2}, Shazia Anjum ², Milan Skalicky ³ , Ejaz Ahmad Waraich ⁴, Rana Muhammad Sabir Tariq ⁵, Muhammad Ashar Ayub ⁶, Akbar Hossain ⁷ , Mohamed M. Hassan ⁸ , Marian Brestic ^{4,9} , Mohammad Sohiful Islam ¹⁰, Muhammad Habib-Ur-Rahman ^{11,12} , Allah Wasaya ¹³ , Muhammad Aamir Iqbal ¹⁴  and Ayman EL Sabagh ^{15,*}

- ¹ Department of Botany, University of Central Punjab, Bahawalpur Campus, Bahawalpur 63100, Pakistan; zahoorahmadbwp@gmail.com
- ² Cholistan Institute of Desert Studies, The Islamia University of Bahawalpur, Bahawalpur 63100, Pakistan; shahzia.anjum@iub.edu.pk
- ³ Department of Botany and Plant Physiology, Faculty of Agrobiolgy, Food and Natural Resources, Czech University of Life Sciences Prague, 165 00 Prague, Czech Republic; skalicky@af.czu.cz
- ⁴ Department of Agronomy, University of Agriculture Faisalabad, Punjab 38000, Pakistan; uaf_ewarraich@yahoo.com (E.A.W.); marian.brestic@uniag.sk (M.B.)
- ⁵ Department of Agriculture and Agribusiness Management, University of Karachi, Karachi 75270, Pakistan; sabir_tariq@yahoo.com
- ⁶ Institute of Soil and Environmental Sciences, University of Agriculture, Faisalabad 78000, Pakistan; muhammadasharayub@gmail.com
- ⁷ Department of Agronomy, Bangladesh Wheat and Maize Research Institute, Dinajpur 5200, Bangladesh; akbarhossainwrc@gmail.com
- ⁸ Department of Biology, College of Science, Taif University, P.O. Box 11099, Taif 21944, Saudi Arabia; m.khyate@tu.edu.sa
- ⁹ Department of Plant Physiology, Slovak University of Agriculture, Nitra, 949 01 Nitra, Slovakia
- ¹⁰ Department of Agronomy, Hajee Mohammad Danesh Science and Technology University, Dinajpur 5200, Bangladesh; shahid_sohana@yahoo.com
- ¹¹ Department of Crop Science, Institute of Crop Science and Resource Conservation (INRES), University Bonn, 53115 Bonn, Germany; mhabibur@uni-bonn.de
- ¹² Department of Agronomy, MNS-University of Agriculture-Multan-Pakistan, Punjab 66000, Pakistan
- ¹³ Department of Agronomy, Bahadur Sub-Campus Layyah, College of Agriculture, Bahauddin Zakariya University, Punjab 31200, Pakistan; wasayauaf@gmail.com
- ¹⁴ Department of Agronomy, Faculty of Agriculture, University of Poonch Rawalakot, Rawalakot 12350, Pakistan; aamir1801@yahoo.com
- ¹⁵ Department of Agronomy, Faculty of Agriculture, University of Kafrelsheikh, Kafr El-Sheikh 33516, Egypt
- * Correspondence: ayman.elsabagh@agr.kfs.edu.eg



Citation: Ahmad, Z.; Anjum, S.; Skalicky, M.; Waraich, E.A.; Muhammad Sabir Tariq, R.; Ayub, M.A.; Hossain, A.; Hassan, M.M.; Brestic, M.; Sohiful Islam, M.; et al. Selenium Alleviates the Adverse Effect of Drought in Oilseed Crops Camelina (*Camelina sativa* L.) and Canola (*Brassica napus* L.). *Molecules* **2021**, *26*, 1699. <https://doi.org/10.3390/molecules26061699>

Academic Editors: Marcello Iriti, Agnieszka Saeid and Witold Kwapinski

Received: 31 January 2021

Accepted: 16 March 2021

Published: 18 March 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: Drought poses a serious threat to oilseed crops by lowering yield and crop failures under prolonged spells. A multi-year field investigation was conducted to enhance the drought tolerance in four genotypes of Camelina and canola by selenium (Se) application. The principal aim of the research was to optimize the crop yield by eliciting the physio-biochemical attributes by alleviating the adverse effects of drought stress. Both crops were cultivated under control (normal irrigation) and drought stress (skipping irrigation at stages i.e., vegetative and reproductive) conditions. Four different treatments of Se viz., seed priming with Se (75 μ M), foliar application of Se (7.06 μ M), foliar application of Se + Seed priming with Se (7.06 μ M and 75 μ M, respectively) and control (without Se), were implemented at the vegetative and reproductive stages of both crops. Sodium selenite (Na_2SeO_3), an inorganic compound was used as Se sources for both seed priming and foliar application. Data regarding physiochemical, antioxidants, and yield components were recorded as response variables at crop maturity. Results indicated that WP, OP, TP, proline, TSS, TFAA, TPr, TS, total chlorophyll contents, osmoprotectant (GB, anthocyanin, TPC, and flavonoids), antioxidants (APX, SOD, POD, and CAT), and yield components (number of branches per plant, thousand seed weight, seed, and biological yields were significantly improved by foliar Se + priming Se in both crops under drought stress. Moreover, this treatment was also helpful in boosting yield attributes

under irrigated (non-stress) conditions. Camelina genotypes responded better to Se application as seed priming and foliar spray than canola for both years. It has concluded that Se application (either foliar or priming) can potentially alleviate adverse effects of drought stress in camelina and canola by eliciting various physio-biochemicals attributes under drought stress. Furthermore, Se application was also helpful for crop health under irrigated condition.

Keywords: drought; antioxidants; osmoprotectant; yield; selenium; camelina; canola

1. Introduction

Water scarcity in changing climate scenarios has become a serious threat to sustainable crop production globally, and especially in developing countries of South Asia such as Pakistan [1]. Agricultural droughts pose a significant threat to crop failure leading to food shortages [2–4]. Water stress negatively affects the physiological and biochemical activities of the plant, including photosynthesis rate, osmotic potential, turgor pressure along with serious damages to the cellular membranes [5]. Agricultural yield can be maximized by evaluating and mitigating the impacts of drought on crops [6]. Numerous techniques have been used to protect against cellular and oxidative damages and to regulate enzymatic activities in crop plants under water stress conditions [7]. Selenium (Se) as basal or foliar fertilization is a current technology used to protect crop plants from drought stress. Selenite (IV, VI) are the major two species present in soil and can be used efficiently by plants. It has been reported that the application of Se increased proline accumulation but did not affect the water uptake capacity, nor did it have a negative impact on plant biomass under drought [8]. In foliar application, Se ions easily diffuse to epidermal cells as they are transported by xylem and phloem, hence becoming part of the plant body [9]. There are various studies carried out about the foliar application of Se on cereals [9], oilseed crops [8], carrots roots [10], onion and garlic bulbs [11,12], broccoli [13], etc., resulting in a significant improvement in the growth and yield of crops. Thus, Se application has the potential to increase the nutritional value of the crop, the grain yield as well as enable plants to have abiotic tolerance. Se improves plant growth by accumulating starch in the chloroplast [14]. Besides, Se can also regulate the activities of several antioxidant enzymes and metabolites providing oxidative stress tolerance to plants [15]. However, the toxicity or benefits of Se are highly dependent on the applied concentration [16,17]. Although, Se is not essential for plants as in the case of humans and animals, but numerous studies showed the beneficial effects of Se in plant health [18].

Furthermore, Se is a beneficial element that acts as an antioxidant and anti-senescent, it is also involved in active defense against abiotic and biotic stressors. It has been reported to be closely associated with higher antioxidant activity [19,20]. In plants, Se has chemical properties similar to sulfur (S) being present in sulfur amino acids such as selenomethionine (SeMet) and selenocysteine (SeCys). The Se is uptakes as selenite and transported via S-transporters leading to its transformation into SeCys and SeMet, which are usable forms in plants [21]. In an investigation, Klognerová et al. [22] reported that Se fortified oilseed rape plants have SeMet as a stable compound stored in tissues, while the same compound was predominately found in defatted oilseed rape meal [23]. The Se is involved in counteracting various types of abiotic stress experienced by plants such as cold, drought, heavy metal pollutants, and intense light [20,24,25]. However, higher concentrations (>1 ppm) can be toxic to plants, causing extreme physiological disturbances [25]. Exogenous application of Se can also help to uptake more plant nutrients as reported in wild plants [26], and various cereals [27,28].

Among oilseed crops, canola (*Brassica napus*) is held in special esteem worldwide as the second most important source of oil [29] with ~40–50% oil and 40% proteins (in rapeseed meal) which is of dietary importance for humans and animals [8]. Canola is considered as the third oilseed plant in the world after palm and soybean [30], and besides

oil content, the rich chemistry of phenols, flavonoids, vitamins, and carotenoids make it to treat a good consumable oilseed crop [8]. The leaves of this plant are used as animal feed due to containing a higher concentration of protein and less fibrous [31]. Oil extracted from oilseed rape contains a very high concentration of C18 fatty acids (such as oleic acid and linoleic acid (C18:3, ~10% *v/v*)) making it a fatty acid-rich diet source [32].

Camelina (*Camelina sativa*) is also an oilseed crop cultivated in Pakistan, belonging to the Brassicaceae family. The camellia seed contains almost 35–45% oil, and the plants complete their life cycle within 60–90 days [33]. It is known that the growth and productivity of crops are reduced under drought stress due to reduced uptake of nutrients and physiological functions. Under such conditions, foliar application nutrients can improve plant growth and yield. Brassica species have been reported to accumulate Se affecting their physiology. Therefore, the present field research was carried out to study the effects of foliar-applied Se on water relations, physiochemical, osmoprotectant, and antioxidants activities of camelina and canola genotypes under drought stress condition.

2. Materials and Methods

2.1. Research Location, Duration and Plant Materials

This study was carried out in the 2016 and 2017 growing seasons in the field area of the Cholistan Institute of Desert Studies (CIDS), The Islamia University of Bahawalpur, Pakistan. Healthy, high-yielding seeds of camelina genotypes viz. Canadian camelina, and Australian camelina, and canola genotypes viz. UAF Canola (developed by the University of Agriculture Faisalabad), and AARI canola (developed by Ayub Agricultural Research Institute) were used as plant materials.

2.2. Treatments, Design and Experimentation

Four varieties of both oilseed (camelina and canola, two varieties each) were used in this experiment. The seeds were pre-sanitized with 3% H₂O₂ for 10 minutes, followed by vigorous washing with distilled water 10 times. Sodium selenite (Na₂SeO₃), an inorganic compound, was used as Se source, both for seed priming and foliar application. This salt is a colourless solid. The pentahydrate Na₂SeO₃(H₂O)₅ is the most common water-soluble selenium compound. For the seed priming with selenium, 5.1 mg L⁻¹ Se solution was prepared in distilled water to be used as a stock solution for priming solution preparation. Four levels of Se were applied, T₁ = Seed-priming (with 75 μM Se solution), T₂ = foliar application of 7.06 μM Se solution, T₃ = foliar Se + Seed priming (7.06 μM and 75 μM), and T₄ = control receiving no Se supplementation. The healthy seeds were sown in the field following a randomized complete block design (RCBD). The research field was divided into two distinct patches representing normal and water-stressed conditions. Water stress was achieved by skipping irrigations in the vegetative and reproductive stages for both years in both oilseed crops. The irrigation requirement was calculated keeping in mind the daily requirement of the crop and the field capacity of the soil. Seed priming was done pre-sowing while foliar application of Se was done on 10-day-old seedlings for the whole growing season with the gap of 25 days.

2.3. Data Recording

2.3.1. Water Potential Determination

A pressure chamber (Scholander type) was used to record water potential on the third fully matured leaf from the top tip of the plant. The leaves were collected and frozen at -20 °C following the method adopted by [34]. The osmotic potential was determined, and the difference method was used to get an idea of the water pressure potential inside the leaves of the plant as follows:

$$(\psi_p) = (\psi_w) - (\psi_s) \quad (1)$$

where (ψ_p), turgor potential; (ψ_s), osmotic potential, and (ψ_w), water potential.

2.3.2. Biochemical Parameter Assay

Soluble Proteins, Amino Acids, Sugars, and Proline

The biochemical parameters under study, total soluble sugars (TSS), total soluble proteins (TSP), total free amino acid (TFAA) contents, and proline contents of leaves were measured following standard protocols [35–39].

Total Soluble Protein (TSP)

Total soluble proteins were determined using the procedures described by [35]:

- Reagents: For the preparation of phosphate buffer (0.2 M) solution, the following chemicals were used; a one-molar solution of $\text{NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$ (156.01 g L^{-1}) was prepared as the stock and a one-molar solution of Di-sodium hydrogen phosphate ($\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$, 177.99 g L^{-1}) was prepared as the stock.
- Copper Reagents: Solution A: Na_2CO_3 2.0 g, NaOH 0.2 g and Sodium potassium tartrate 1.0 g. All three chemicals were dissolved in distilled water and the volume was made to 100 mL. Solution B: $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ solution: 0.5 g $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ was dissolved in 100 mL distilled water. Solution C: Fifty mL of solution A and 1.0 mL of solution B were mixed to prepare the alkaline solution. This solution was always prepared fresh.
- Folin phenol reagent: One hundred g of sodium tungstate and 25 g of sodium molybdate were dissolved in 700 mL of distilled water. Fifty mL of 85% orthophosphoric acid and 100 mL of HCl were added and the mixture was refluxed for 10 h. Then 150 g of lithium sulfate was added along with 50 mL of distilled water. A few drops of Br_2 were also added.

The mixture was boiled without a condenser for 15 min to remove extra Br_2 . The mixture was then cooled and diluted to 1000 mL. Standard Bovine Serum Albumin (BSA) solution ($1 \mu\text{g mL}^{-1}$). Ten mg of Bovine serum albumin (BSA) was dissolved in 10.0 mL of distilled water.

Extraction: Fresh leaf material (0.5 g) was chopped in 10 mL of phosphate buffer (0.2 M) of pH 7.0 and was ground. The ground leaf material was centrifuged at 5000 g for 5 min. The supernatant was used for protein determination.

Procedure: One mL of the leaf extract from each treatment was taken in a test tube. The blank contained 1 mL of phosphate buffer (pH 7.0). One mL of solution C was added to each test tube. The reagents in the test tube were thoroughly mixed and allowed to stand for 10 min at room temperature. Then 0.5 mL of Folin-Phenol reagent (1:1 diluted) was added, mixed well, and incubated for 30 min. at room temperature. The optical density (OD) was read at 620 nm on a spectrophotometer (Hitachi, 220, Tokyo, Japan).

Total Free Amino Acids (TFA)

Total free amino acids were determined according to [35–37]. Fresh plant leaves (0.5 g) were chopped and extracted with phosphate buffer (0.2 M) having pH 7.0. Took 1 mL of the extract in 25 mL test tube, added 1 mL of pyridine (10%) and 1 mL of ninhydrin (2%) solution in each test tube. Ninhydrin solution was prepared by dissolving 2 g ninhydrin in 100 mL distilled water. The test tubes with a sample mixture, heated in boiling water bath for about 30 min. The volume of each test tube was made up to 50 mL with distilled water. The optical density of the colored solution at 570 nm was recorded using a spectrophotometer. Developed a standard curve with Leucine and calculated free amino acids using the formulae given below:

$$\begin{aligned} & \text{Total amino acids} \\ & = \frac{\text{Graph reading of sample} \times \text{Volume of factor} \times \text{Dilution factor}}{\text{Weight of fresh tissue} \times 1000} \end{aligned} \quad (2)$$

Total Soluble Sugars (TSS)

Total soluble sugars were determined according to the method of [38,39].

Reagents: Anthrone reagent was prepared by dissolving 150 mg of anthrone in 72% H₂SO₄ solution. This reagent was freshly prepared whenever needed.

Extraction: Dried plant material was ground well in a micromill and the material was sieved through a 1 mm sieve of micromill. Plant material (0.1 g) was extracted in 80% ethanol solution. The extract was incubated for 6 h at 60 °C. This extract was used for the estimation of total soluble sugars.

Procedure: Plant extract was taken in 25 mL test tubes and 6 mL anthrone reagent was added to each tube, heated in boiling water bath for 10 min. The test tubes were ice-cooled for 10 min. and incubated for 20 min. at room temperature (25 °C). Optical density was read at 625 nm on a spectrophotometer (Hitachi, 220, Tokyo, Japan). The concentration of soluble sugars was calculated from the standard curve developed by using the above method.

Proline Determination

The proline was determined according to the method of [39]. Fresh leaf material of 0.5 g was grounded and dissolved in 10 mL of 3% sulfosalicylic acid. The sample material was filtered by using Whatman No. 2 filter paper. Two mL of the filtrate was taken in a test tube and reacted with 2 mL acid ninhydrin solution. Acid ninhydrin solution was prepared by dissolving 1.25 g ninhydrine in 30 mL of glacial acetic acid and 20 mL of 6 M orthophosphoric acid.

Two mL of glacial acetic acid was added in the test tube and kept for 1 h at 100 °C. After terminating the reaction in an ice bath, the reaction mixture was extracted with 10 mL toluene. The continuous air stream was passed vigorously for 1–2 minutes in the reaction mixture. The chromophore containing toluene was aspirated from the aqueous phase, warmed at room temperature and the absorbance was noted at 520 nm on a spectrophotometer. Toluene was used as a blank. The proline concentration was calculated by using a standard curve and calculated on fresh weight basis as follows: $\text{Mmol proline g}^{-1} \text{ fresh weight} = (\text{g proline mL}^{-1} \times \text{mL of toluene} / 115.5) / (\text{wt. of sample} / 5)$.

Osmoprotectants and Total Phenolic

For the determination of osmoprotectants and total phenolic contents, fresh plant leaves were grounded, and oven-dried followed by soaking in 10 mL methanol, water, and HCL solution (79: 20: 1 *v/v*), and the mixture was shaken for 72 h at 4 °C in dark. The above procedure extracted the essence of plant leaves, this mixture was centrifuged at 5000 × *g* and its absorbance were measured at 530 and 657 nm following the protocol of Ribera-Fonseca et al. [40] using the following equations:

$$\text{Corrected } A_{530} = A_{530} - 1/3 A_{657} \quad (3)$$

where A₅₃₀ reading was obtained at 530 nm, similarly A₆₅₇ obtained reading at 657 nm.

For the determination of Anthocyanins, and flavonoid content protocols of Zhao et al. [41] and Wang et al. [42] were used, respectively. For these, 0.5 mL extract volume was mixed with 0.5 mL of 2% AlCl₃ solution in ethanol at room temperature. The absorbance of the solution mixture was calculated at 520 nm and flavonoid content determination was done making relation to equivalent Quercetin as a reference to the standard curve.

Glycine betaine (another osmoprotectant) was determination was done following the method proposed by Salama et al. [43]. For this purpose, 1 g fresh preserved plant leaves were grounded in 10 mL double deionized water and 1 mL mixture was spiked with 1 mL 2 N HCL for acidification of media. Half (0.5) mL of this acidified solution was taken in test tubes containing 0.2 mL of potassium triiodide (I₃K), and the mixture was kept in an ice bath for 90 min. In this mixture, 1 mL of chilled and double deionized water and 1, 2 dichloroethane were introduced and steamed air was passed from the mixture for 1–2 min to completely dissolve solvents and break the double layer. The double beam spectrophotometer (Hitachi-150-20, Tokyo, Japan) was used for the determination of absorbance and ultimately GB contents were determined.

Soluble phenols determination was done following Mohd-Rosni et al. [36] method, 1 g leaf sample was grounded in 10 mL 80% acetone solution and the mixture was centrifuged at 4000 rpm of 10 min. Then, 20 μ L of centrifuged material was loaded with 1.58 mL distilled water, 100 μ L of Folin-Ciocalteu reagent, and 300 μ L of sodium carbonate, and the mixture was kept at 40 °C in a hot bath for 30 min. Mixture absorbance was checked at 765 nm and a calibration curve was used for phenol concentration determination.

Assay of Antioxidant Enzyme Extraction

Antioxidant enzymes (APX, SOD, POD, and CAT) were determined following the Spectrophotometric technique as reported by [44]. The APX and SOD were determined using the methodology adopted by [45], CAT and POD were measured using the method described by [46].

Catalase (CAT)

Catalase activity was assayed by measuring the conversion rate of hydrogen peroxide to water and oxygen molecules. The activity was assayed in 3 mL reaction solution comprising 50 mM phosphate buffer with 7.0 pH, 5.9 mM of H₂O₂ and 0.1 mL enzyme extract. The catalase activity was determined by decline in absorbance at 240 nm after every 20 s due to consumption of H₂O₂. Absorbance change of 0.01 units min⁻¹ was defined as one unit catalase activity.

Peroxidase (POX)

The activity of POD was determined by measuring the peroxidation of hydrogen peroxide with guaiacol as an electron donor [44]. The reaction solution for POD consists of 50 mM phosphate buffer with pH 5, 20 mM of guaiacol, 40 mM of H₂O₂ and 0.1 mL enzyme extract. The increase in the absorbance due to the formation of tetraguaiacol at 470 nm was assayed after every 20 s. One unit of the enzyme was considered as the amount of the enzyme that was responsible for the increase in OD value of 0.01 in 1 min. The enzyme activity was determined and expressed as unit's min⁻¹ g⁻¹ fresh weight basis.

Ascorbate Peroxidase

Ascorbate peroxidase (APX) activity was measured by monitoring the decrease in absorbance of ascorbic acid at 290 nm (extinction coefficient 2.8 mM cm⁻¹) in a 1 mL reaction mixture containing 50 mM phosphate buffer (pH 7.6), 0.1 mM Na-EDTA, 12 mM H₂O₂, 0.25 mM ascorbic acid and the sample extract as described by [44–46].

SPAD Value

A pre-calibrated handheld SPAD meter (SPAD-502) was used to measure the SPAD values.

Yield and Yield-Related Components

For biomass and growth parameter determination, the number of branched per plant, 100-grain weight, fresh and dry weights of tissue were recorded in tons per hectare.

2.4. Statistical Analysis

The recorded data were arranged and statistically analyzed using an ANOVA test (Table 1) at a 5% probability level followed by a pairwise comparison test using the least significant difference (LSD) test with the help of MSTAT-C software [47].

Table 1. Analysis of variance (ANOVA) of growth, water relations, biochemical, osmoprotectants, antioxidant, and yield parameters of camelina and canola under drought stress by foliar-applied Selenium.

Traits	Treatments	Varieties	Treatment × Varieties
Total Chlorophyll Contents (SPAD)	***	**	**
Water Potential (−MPa)	**	**	**
Osmotic Potential (−MPa)	**	**	**
Turgor Pressure (MPa)	***	**	*
Total Proline (μmol/g DW)	***	**	NS
Total Soluble Sugars (μmol/g FW)	***	**	NS
Total Free Amino Acids (mg/g)	***	**	NS
Total Soluble Proteins (mg/g)	**	*	NS
Total Phenolic Contents (mg/g DW)	***	**	NS
Anthocyanin (mg/g DW)	***	**	NS
Flavonoids (mg/g DW)	***	**	NS
Glycine betaine (μmol/g FW)	***	**	NS
Ascorbic peroxidase (units' min ^{−1} g ^{−1} FW)	***	**	NS
Superoxide Dismutase (units' min ^{−1} g ^{−1} FW)	***	**	NS
Peroxidase (units' min ^{−1} g ^{−1} FW)	***	**	NS
Catalase (units' min ^{−1} g ^{−1} FW)	***	**	NS
Number of branches per plant	**	*	NS
Thousand seed weight (g)	**	*	NS
Seed yield (t/ha)	***	**	NS
Biological yield (t/ha)	**	**	NS

*, ** and *** = Significant at 0.05, 0.01 and 0.001 level, respectively; NS = Non-significant.

3. Results

3.1. Physiological Attributes

All the physiological attributes under study, such as water potential, osmotic potential, turgor pressure, and total chlorophyll contents (SPAD value of both oilseed crops were significantly reduced ($p < 0.01$) due to drought (Table 1). The reductions in all physiological attributes were observed where water deficit was applied at vegetative and reproductive stages. The graphical representation shows that both camelina genotypes performed better than both canola genotypes in terms of all physiological traits in both stages (vegetative and reproductive) with optimal water and limited water supply in both years (Figure 1).

Different treatments such as seed priming, foliar, and foliar plus seed priming with Se performed significantly improved all the physiological parameters of both oilseed crops with mentioned stages (vegetative and reproductive) in both years either with full and limited water supply, as shown in the graphical representation (Figure 1). All physiological characters showed a better improvement in camelina during the two stages (vegetative and reproductive) in both years compared to canola when Se was applied with seed priming plus the foliar application (T3) under limited water supply, as well as regular irrigation (Figure 1). However, the interactive effects were found to be non-significant for both years (Table 1).

3.2. Biochemical Attributes

All biochemical associations attributes such as total proline, total free amino acid, total soluble protein, and total soluble sugar were reduced significantly ($p < 0.01$) in the mentioned stages (vegetative and reproductive) for both varieties of camelina and canola in both years in the treatment with limited water application as presented in Table 1. The reduction in all biochemical attributes was noticed in limited water conditions (drought) water at both stages of canola and camelina in both years. The graphical representation shows that both genotypes of camelina performed better than canola genotypes in all biochemical characters under full application of water and limited water in both years (Figure 2). Different treatments such as seed priming, foliar, and foliar plus seed priming with Se performed significantly better by improving all the biochemical parameters of both crops in both stages and for both years either under full water provision and limited water conditions, as presented in Figure 2. In camelina, all the biochemical features evaluated

showed better positive effects for both stages during both years compared to canola when foliar (T3) was applied with seed priming carried out under limited and regular irrigation (Figure 2). The interactions were not found significant as given in Table 1.

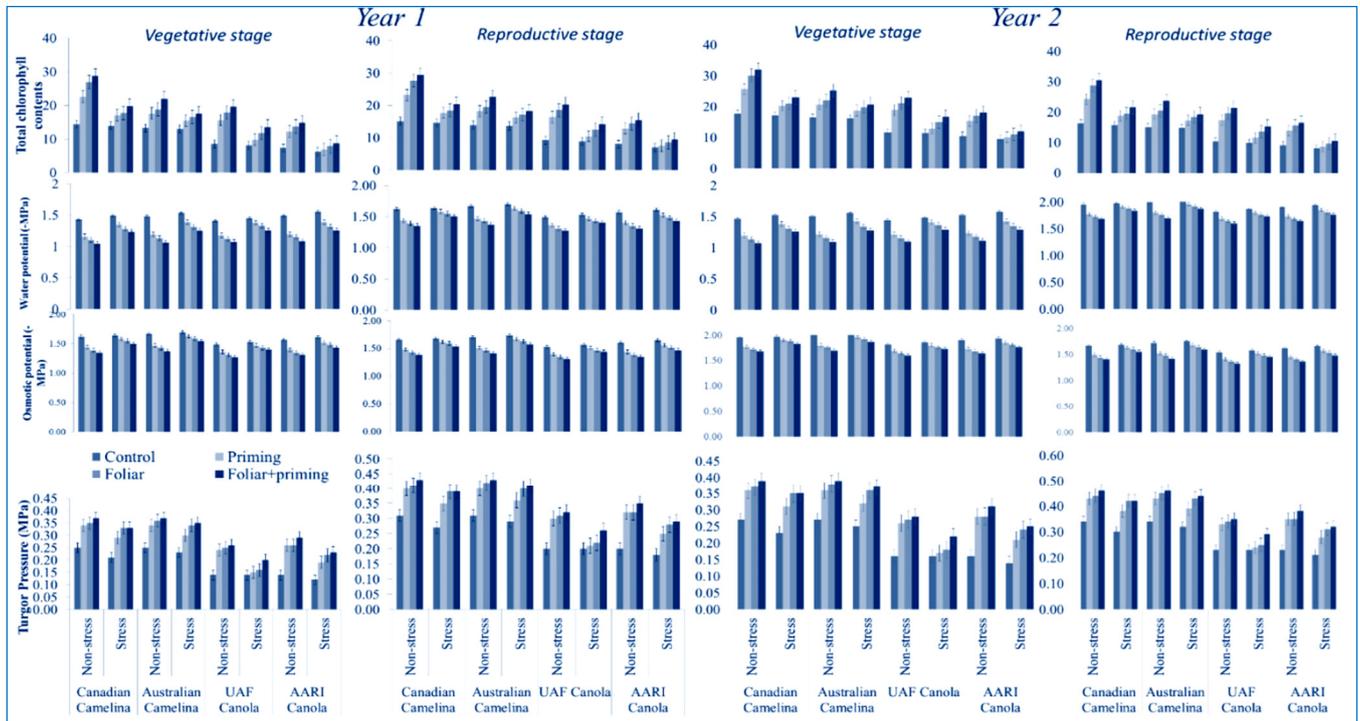


Figure 1. Impact of Se application (seed priming and foliar) on physiological attributes at both stages (vegetative and reproductive) of oilseed crops (camelina and canola) subjected to water deficit in the year 2016 and 2017 (mean values \pm S.E).

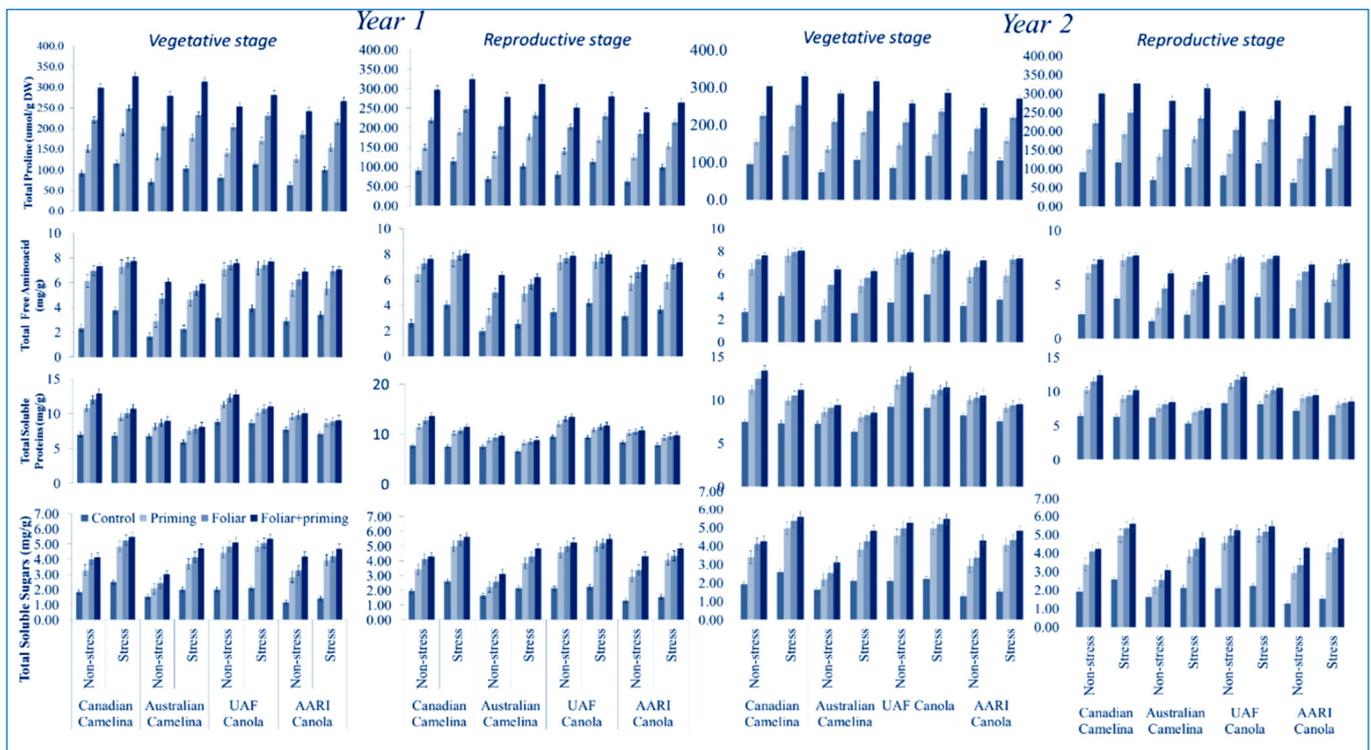


Figure 2. Impact of Se application (seed priming and foliar) on biochemical attributes at both stages (vegetative and reproductive) of oilseed crops (camelina and canola) subjected to water deficit in the year 2016 and 2017 (mean values \pm S.E).

3.2.1. Assay of Antioxidants

Different antioxidant attributes such as APX, SOD, POD, and CAT were significantly reduced ($p < 0.01$) in the mentioned stages (vegetative and reproductive) in both varieties of camelina and canola in both years in case of limited water application as presented in Table 1. The reduction in all antioxidant attributes was observed in the treatments with limited application of water, this response was not detected in the treatments with adequate water supply. All studied antioxidants, both genotypes of camelina performed better than both canola genotypes at both stages under partial and full application of water during both the years (Figure 3). Different treatments such as seed priming, foliar, and foliar plus seed priming with Se improved all the antioxidant parameters of both oilseed crops during both years either under full application of water and limited water conditions (Figure 3). All evaluated antioxidant elements showed better improvement in camelina in both critical stages during both years compared to canola when Se was applied as seed priming plus foliar application for the treatments carried out under limited and regular irrigation (Figure 3). The interactions were not found significant (Table 1).

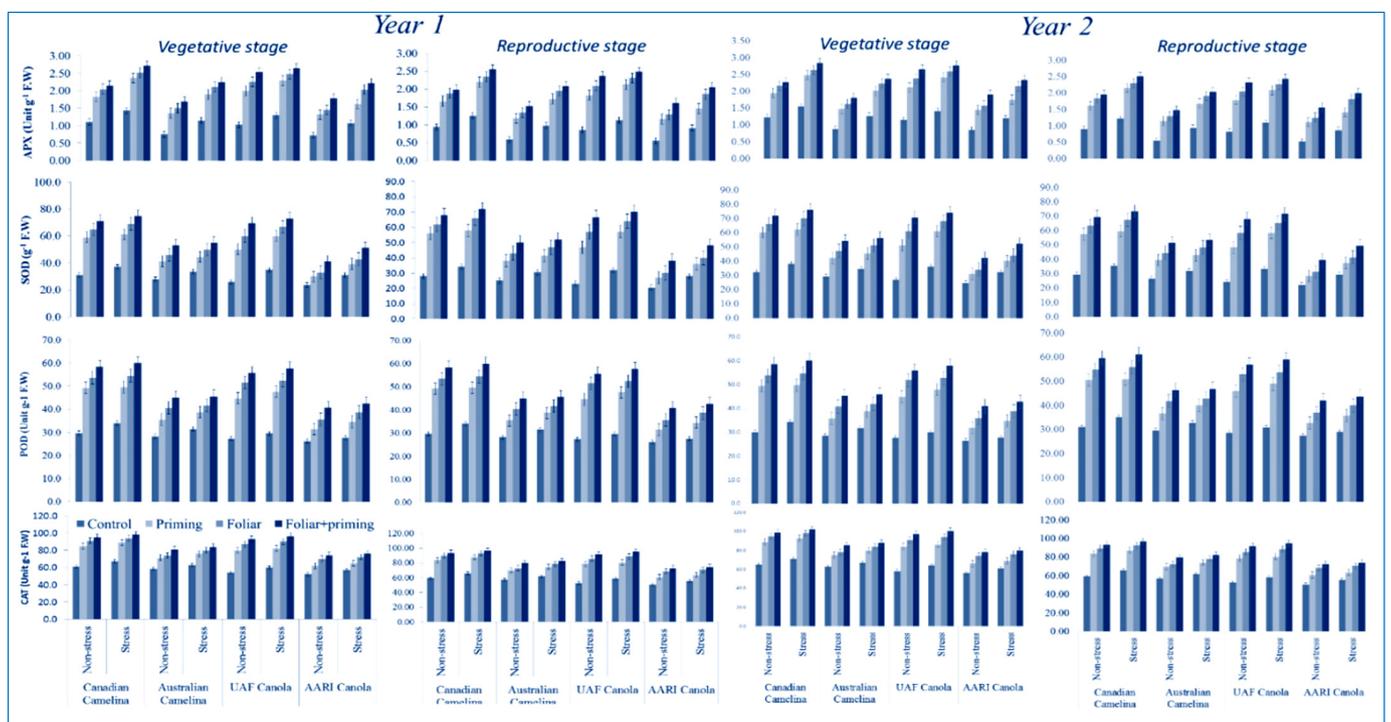


Figure 3. Impact of Se application (seed priming and foliar) on antioxidant activities at both stages (vegetative and reproductive) of oilseed crops (camelina and canola) subjected to water deficit in the year 2016–2017 (mean values \pm S.E).

3.2.2. Osmoprotectants Characters

Osmoprotectant association attributes such as anthocyanin, flavonoid, GB, and TPC were significantly reduced ($p < 0.01$) in the vegetative and reproductive phases in both varieties of camelina and canola during both years with limited application of water (Table 1). The reduction in all osmoprotectant attributes was significantly greater with the limited application of water than with the full application of water in both canola and camelina stages during both years. Camelina genotypes performed better than the canola genotypes on all osmoprotectant traits in both stages under full and limited water application during both years (Figure 4). Different treatments such as seed priming, foliar and foliar plus seed priming with Se performed significantly well in improving all the osmoprotectant parameters of the crops in both stages during both years either with full and limited application of water (Figure 4). Camelina showed better improvements in all the osmoprotectant characters in both stages and years compared to canola when Se

was applied with seed priming plus foliar application under limited as well as the full application of water (Figure 4). The interactions were not found significant as given in Table 1.

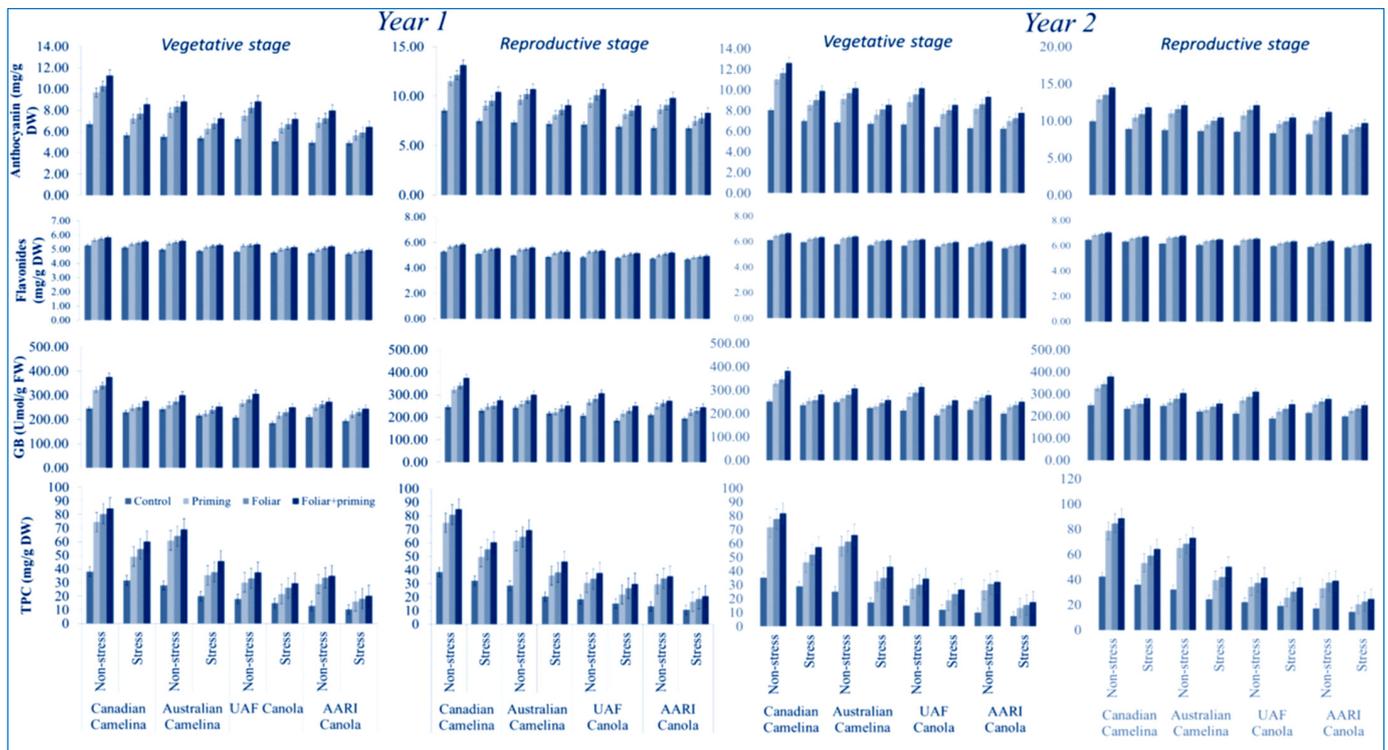


Figure 4. Impact of Se application (seed priming and foliar) on osmoprotectants at both stages (vegetative and reproductive) of oilseed crops (camelina and canola) subjected to water deficit in the year 2016–2017 (mean values \pm S.E.).

3.3. Yield Attributes

Yield attributes such as the number of branches per plant, thousand seed weight, seed and biological yields of both crops were significantly reduced ($P < 0.01$) due to limited water application during both years (Table 1). The data revealed that both camelina genotypes performed better than canola genotypes in terms of all the evaluated yield traits under full and limited water supply during both years (Figure 5). Different treatments such as seed priming, foliar and foliar together with the combined application of Se remained significantly effective in improving all yield parameters of both crops (Figure 5). Yield characters showed better improvement in camelina crop at both stages during both years than canola, in plants where Se was applied with seed priming plus foliar application under lower as well as the full application of water (Figure 5). The interactions were not found significant (Table 1).

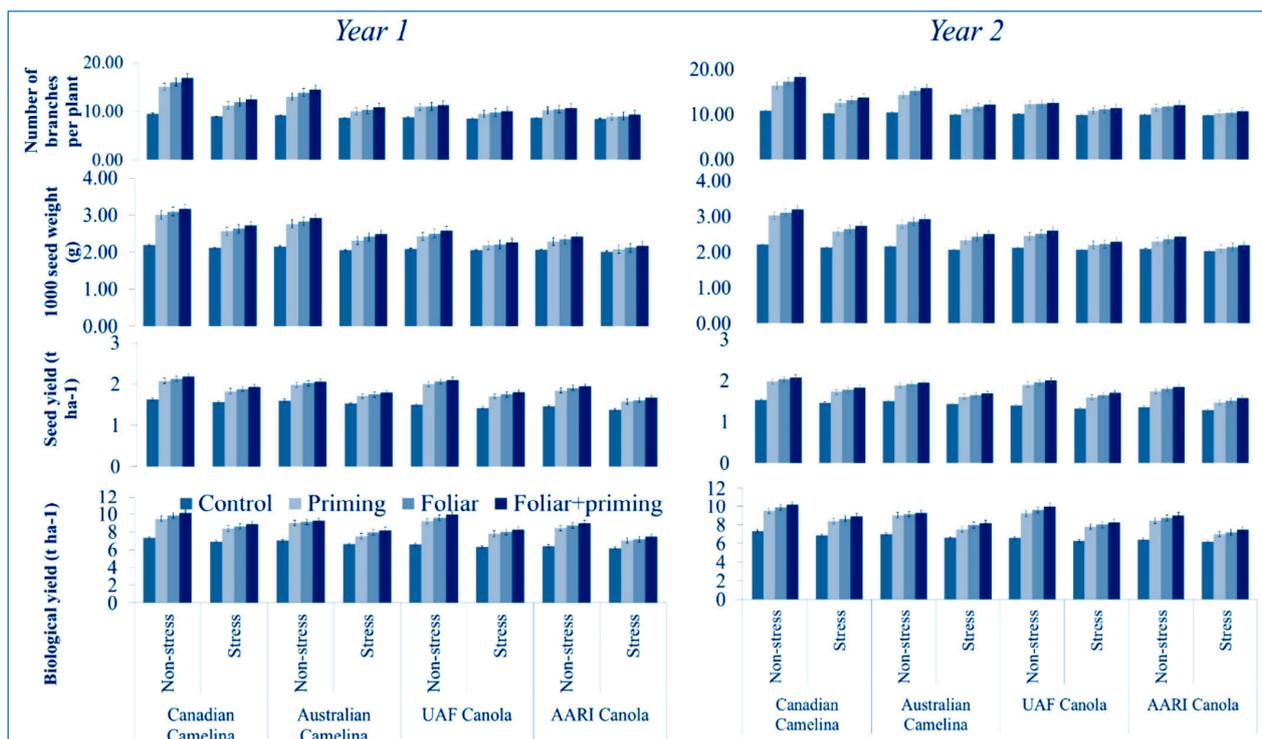


Figure 5. Impact of Se application (seed priming and foliar) on yield and yield components at both stages (vegetative and reproductive) of oilseed crops (camelina and canola) subjected to water deficit in the year 2016–2017 (mean values \pm S.E).

4. Discussion

Both oilseed crops (camelina and canola) exhibited significant improvements in osmoregulation parameters like water-potential (Ψ_w), turgor-pressure, and osmotic potential under both control and water-limited conditions, when treated with Se either as seed priming or foliar application at both stages (vegetative and reproductive) tissues, the application of Se significantly enhanced the water uptake by the roots without decreasing the transpiration rate (E) [48]. The results of the present study revealed that the exogenous application of Se increased the Ψ_w and similar findings were reported by [49]. Likewise, a significant rise in biological and economical yield and water use efficiency (WUE) of the maize crop was seen by foliar-applied Se [50]. Similarly, it was observed that Se addition in plant growing media improved the leaf water potential of maize under severe water deficit conditions [51]. However, maize plant anthocyanin and flavonoids color pigments were moderately reduced under drought stress conditions. It is suggested that the desiccation of the leaves affects the synthesis of anthocyanin and flavonoids more than the Chl contents. In addition, the effect of Se on the pigments of corn leaves was observed for anthocyanin and flavonoids.

Anthocyanin is an important compound involved in the protection of cellular machinery against abiotic stress-mediated by oxidative stress [52]. Similarly, flavonoids are antioxidants that act as cellular protectors defending the plants against water stress [50]. Among plant biochemical machinery units, total chlorophyll, chl 'a', and chl 'b' are very important for optimal photosynthetic activity [53]. Drought (or partial water stress) can severely affect the production of chl content thus significantly decrease photosynthetic activity [54,55].

The results of the study revealed that a considerable reduction in total chlorophyll content was observed under drought stress. Previous studies showed that water deficit caused a reduction of Chl content in several oilseed crops such as sunflower [56] and other cereals including wheat [57], and corn [58]. Reactive oxygen species (ROS) effectively lessened the chlorophyll content of the leaves by damaging the chloroplasts [59]. The

results of the present experiment confirmed the findings that Se seed treatment and foliar application resulted in higher chlorophyll content on the leaves of camelina and canola crops under normal and water-deficit conditions. The plants of the camelina and canola crops also sustained the maximum Chl contents due to Se supply at both stages (vegetative and reproductive). In contrast, drought stress significantly declined the Chl contents at later growth stages (vegetative and reproductive stage) in rapeseed plants reported by [60]. Seed priming with Se plus foliar application revealed the maximum increase in foliar Chl content in camelina and canola under both normal and limited water conditions.

The production of plant TSS as well as its involvement in drought stress management is well documented in soybean [61], wheat [62], and rice [61]. Our study also found similar results, reporting a net increase in TSS in camelina and canola upon exogenous application of Se (foliar as well as seed priming). In our work, plants have shown the maximum accumulation in the vegetative stage and a similar observation was made by [63]. Se supplementation to plants helps increase TSS accumulation. An investigation conducted by [49] also has similar observations for wheat crops and reports a significant increase in TSP and TFA upon exogenous application of Se. The exogenous application of Se has helped various crops like soybean [64] as well as trees like pear-jujube [63]. Increase TFA concentration by Se application may be the result of the modulation of amino acid metabolism that controls protein production and NO₃ reductase activity [65]. The accumulation of TFA can help plants withstanding abiotic stress (such as osmotic stress) [66]. Similar results can be inferred from the present investigation added to the indirect beneficial effects of the application of Se.

The application of Se has shown a significant effect in the enhancement of proline accumulation in plant tissue under saline [67], cold [68], and metal stress [69]. In this research, the exogenous application of Se has shown a significant rush in proline contents of camelina and canola under drought. Higher proline contents are an indication of impaired protein biosynthesis [70] and its content is used to assess water stress due to its direct correlation with the extent of stress [56]. Our findings regarding Se mediated enhancement of proline content is in direct agreement with [51] who reported a 2–4 times increased proline concentration in wheat intending to improve tolerance to water stress.

Abiotic stress in the plant disturbs its biochemical machinery and produces dangerous ROS. To counteract this, plants tend to produce some antioxidants (SOD, POD, and CAT) [50] whose contents can be enhanced by applying exogenous amendments. The results of the present study showed an increase in antioxidants activity of camelina and canola plants under drought stress at both in the vegetative and reproductive stages. Previous studies in three canola cultivars indicated that foliar-applied Se has significantly altered enzyme activities to mitigate drought stress, supporting the present conclusion [71]. Among these parameters, the SOD concentration was significantly improved, indicating the plants response to drought stress by regulating the radical scavenging mechanism. Similarly, the application of Se to multiple canola varieties has been reported to trigger excessive amounts of SOD under drought stress, supporting the present findings [72]. Similar findings can be found in published works of [66] and [30] that have shown an increase in the antioxidant activity of wheat and barley under exogenous applied Se, which counteracts oxidative management. The maximum antioxidant activities were recorded in plants where Se was applied by foliar route with Se plus seed priming with Se at vegetative and reproductive stage of camelina and canola under water deficit conditions. An increase in antioxidants activities by exogenous Se supply under drought stress is in line with the findings of [51] who suggested that optimal Se supply is necessary to increase antioxidant activity in water deficit plants. As far as the mode of application is concerned, Se was not as effective as in a single-mode, rather it was helpful when applied synergistically as seed priming + foliar spray. The present conclusion was supported by Habibi [30], who depicted that foliar-applied Se alone is not helpful to combat the drought stress in Canola, indicated by higher accumulation of MDA content.

The application of Se influences the growth of plants but the cited results are not very consistent. It has a growth-promoting effect in soybean [64]. However, in the work conducted by [73], the authors reported a non-significant effect of Se application on soybean yield, similar results were reported for potato by Germ et al. [72]. In our work, exo-applied Se has shown a clear effect to enhance growth and yield of camelina and canola with foliar + seed priming. Between both crops, camelina responded better under water stress. The fact that camelina has better adaptability under drought stress compared to canola has been previously reported [74]. Hence, the role of camelina has become more proficient as an oilseed crop in the changing climate. In the present study, the application of Se as foliar + seed priming treatment (T3) resulted in higher grain yield compared to the control treatment. In previous studies carried out in two canola cultivars in the subtropical dryland zone where Se was applied at 1.5 mg/L and 3 mg/L, the results showed a significant contribution of this element to grain yield under drought stress [8]. This conclusion was also supported by the work published by Curtin et al. [75] which listed the foliar Se application method as more effective than fertilization for the wheat crop, as the foliar application of Se enhances its foliar diffusion making it more available to plants [76]. Exogenous application of Se is beneficial in general to tackle water stress in our study but a very high concentration should be avoided since it can damage the metabolism of the leaves and reduce the growth and plant production [19]. Therefore, it requires comprehensive further investigations.

5. Conclusions

Drought poses a substantial threat to oilseed crops including camelina and canola jeopardizing the food security of a rapidly growing population. However, supplementation of Se could alleviate the adverse effect of these unfavourable conditions through the alteration of the physio-biochemical process of plants. After two years of observation, it is revealed that when Se was used for seed priming and foliar application, water-related data (water potential, osmotic potential, turgor pressure), biochemical parameters (total soluble sugars, total free amino acids, total proteins, total sugars, total chl contents), osmoprotectants (glycine betaine, proline, anthocyanin, total phenolic contents, flavonoids) and antioxidants (ascorbate peroxidase, superoxide dismutase, peroxidase) and catalase improved significantly, ultimately leading to improving the yield and yield components of both crops under drought stress. Among these crops, camelina genotypes responded better to Se than canola during both years. Therefore, it can be concluded that the application of Se, either foliar or seed priming, can alleviate the adverse effect of drought stress in camelina and canola by eliciting various physio-biochemical attributes. Furthermore, oilseed crop varieties respond differently to Se seed priming and foliar application.

Author Contributions: Conceptualization, Z.A. and S.A.; methodology, Z.A., S.A. and E.A.W.; software, M.A.A. and E.A.W.; validation, Z.A. and S.A.; formal analysis, Z.A., S.A. and A.H.; investigation, Z.A., S.A. and E.A.W.; data curation, R.M.S.T. and M.A.A.; writing—original draft preparation, Z.A., M.A.I., S.A. and E.A.W.; writing—review and editing, M.S., M.B., M.S.I., M.A.I., A.W., M.H.-U.-R., A.H., M.M.H. and A.E.S.; funding acquisition, A.H., M.S., M.B., M.M.H. and A.E.S. All authors have read and agreed to the published version of the manuscript.

Funding: This study was supported by an S-grant from the Ministry of Education, Youth, and Sports of the Czech Republic. The authors are highly grateful to Taif University Researchers Supporting Project Number (TURSP-2020/59), Saudi Arabia.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Most of the recorded data are available in all Tables and Figures in the manuscript.

Acknowledgments: This research was supported by the Higher Education Commission (HEC) of Pakistan by providing Startup Research Grant Project (SRGP) during Interim Placement of Fresh PhDs (IPFP) for the period of one year. Also, the authors are highly grateful to Taif University Researchers Supporting Project Number (TURSP-2020/59), Saudi Arabia.

Conflicts of Interest: The authors declare no conflict of interest.

Sample Availability: Samples of the compounds are available from the authors.

Abbreviations

APX	Ascorbate peroxidase
CAT	Catalase
Chl	Chlorophyll
GB	Glycine betaine
OP	Osmotic potential
POD	Peroxidase
Se	Selenium
SOD	Superoxide dismutase
SS	Soluble sugar
TFAA	Total free amino acids
TP	Turgor pressure
TPC	Total phenolic contents
TPr	Total proteins
TSS	Total soluble sugars
WP	water potential

References

- Ahmad, Z.; Waraich, E.A.; Barutçular, C.; Hossain, A.; Erman, M.; ÇİÇ, F.; Gharib, H.; EL Sabagh, A. Enhancing drought tolerance in wheat through improving morpho-physiological and antioxidants activities of plants by the supplementation of foliar silicon. *Phyton Int. J. Exp. Bot.* **2020**, *89*, 529–539.
- Wang, L.; D’Odorico, P.; Evans, J.P.; Eldridge, D.J.; McCabe, M.F.; Caylor, K.K.; King, E.G. Dryland ecohydrology and climate change: Critical issues and technical advances. *Hydrol. Earth Syst. Sci.* **2012**, *16*, 2585–2603. [\[CrossRef\]](#)
- Hao, Z.; Singh, V.P. Drought characterization from a multivariate perspective: A review. *J. Hydrol.* **2015**, *527*, 668–678. [\[CrossRef\]](#)
- Zhang, Q.; Gu, X.; Singh, V.P.; Kong, D.; Chen, X. Spatiotemporal behavior of floods and droughts and their impacts on agriculture in China. *Glob. Planet Chang.* **2015**, *131*, 63–72. [\[CrossRef\]](#)
- Miao, Y.; Zhu, Z.; Guo, Q.; Ma, H.; Zhu, L. Alternate wetting and drying irrigation-mediated changes in the growth, photosynthesis and yield of the medicinal plant *Tulipa edulis*. *Ind. Crops Prod.* **2015**, *66*, 81–88. [\[CrossRef\]](#)
- Madadgar, S.; AghaKouchak, A.; Farahmand, A.; Davis, S.J. Probabilistic estimates of drought impacts on agricultural production. *Geophys. Res. Lett.* **2017**, *44*, 7799–7807. [\[CrossRef\]](#)
- Filgueiras, L.; Silva, R.; Almeida, I.; Vidal, M.; Baldani, J.I.; Meneses, C.H.S.G. *Gluconacetobacter diazotrophicus* mitigates drought stress in *Oryza sativa*. *Plant Soil* **2020**, *451*, 57–73. [\[CrossRef\]](#)
- Mohtashami, R.; Movahhedi, M.D.; Balouchi, H.; Faraji, H. Improving yield, oil content and water productivity of dryland canola by supplementary irrigation and selenium spraying. *Agric. Water Manag.* **2020**, *232*, 106046. [\[CrossRef\]](#)
- Boldrin, P.F.; Faquin, V.; Ramos, S.J.; Boldrin, K.V.F.; Ávila, F.W.; Guilherme, L.R.G. Soil and foliar application of selenium in rice biofortification. *J. Food Compos. Anal.* **2013**, *31*, 238–244. [\[CrossRef\]](#)
- Kápolna, E.; Hillestrøm, P.R.; Laursen, K.H.; Husted, S.; Larsen, E.H. Effect of foliar application of selenium on its uptake and speciation in carrot. *Food Chem.* **2009**, *115*, 1357–1363. [\[CrossRef\]](#)
- Kápolna, E.; Laursen, K.H.; Husted, S.; Larsen, E.H. Bio-fortification and isotopic labelling of Se metabolites in onions and carrots following foliar application of Se and ⁷⁷Se. *Food Chem.* **2012**, *133*, 650–657. [\[CrossRef\]](#)
- Pöldma, P.; Tõnutare, T.; Viitak, A.; Luik, A.; Moor, U. Effect of selenium treatment on mineral nutrition, bulb size, and antioxidant properties of garlic (*Allium sativum* L.). *Agric. Food Chem.* **2011**, *59*, 5498–5503. [\[CrossRef\]](#) [\[PubMed\]](#)
- Šindelářová, K.; Száková, J.; Tremlová, J.; Mestek, O.; Praus, L.; Kaňa, A.; Najmanová, J.T.P. The response of broccoli (*Brassica oleracea* convar. *italica*) varieties on foliar application of selenium: Uptake, translocation, and speciation. *Food Addit. Contam. Part A* **2015**, *32*, 2027–2038. [\[CrossRef\]](#)
- Mozafariyan, M.; Pessarakli, M.; Saghafi, K. Effects of selenium on some morphological and physiological traits of tomato plants grown under hydroponic condition. *J. Plant Nutr.* **2017**, *40*, 139–144. [\[CrossRef\]](#)
- Helaly, M.N.; El-Hoseiny, H.; El-Sheery, N.I.; Rastogi, A.; Kalaji, H.M. Regulation and physiological role of silicon in alleviating drought stress of mango. *Plant Physiol. Biochem.* **2017**, *118*, 31–44. [\[CrossRef\]](#)

16. Hasanuzzaman, M.; Hossain, M.A.; Fujita, M. Selenium-induced up-regulation of the antioxidant defense and methylglyoxal detoxification system reduces salinity-induced damage in rapeseed seedlings. *Biol. Trace Elem. Res.* **2011**, *143*, 1704–1721. [[CrossRef](#)]
17. Hasanuzzaman, M.; Hossain, M.A.; Fujita, M. Exogenous selenium pretreatment protects rapeseed seedlings from cadmium-induced oxidative stress by upregulating antioxidant defense and methylglyoxal detoxification systems. *Biol. Trace Elem. Res.* **2012**, *149*, 248–261. [[CrossRef](#)] [[PubMed](#)]
18. Lyons, G.H.; Genc, Y.; Soole, K.; Stangoulis, J.C.R.; Liu, F.; Graham, R.D. Selenium increases seed production in Brassica. *Plant Soil* **2009**, *318*, 73–80. [[CrossRef](#)]
19. Száková, J.; Praus, L.; Tremlová, J.; Kulhánek, M.; Tlustoš, P. Efficiency of foliar selenium application on oilseed rape (*Brassica napus* L.) as influenced by rainfall and soil characteristics. *Arch. Agron. Soil Sci.* **2017**, *63*, 1240–1254. [[CrossRef](#)]
20. Djanaguiraman, M.; Prasad, P.V.V.; Seppanen, M. Selenium protects sorghum leaves from oxidative damage under high temperature stress by enhancing antioxidant defense system. *Plant Physiol. Biochem.* **2010**, *48*, 999–1007. [[CrossRef](#)]
21. Ellis, D.R.; Salt, D.E. Plants, selenium and human health. *Curr. Opin. Plant Biol.* **2003**, *6*, 273–279. [[CrossRef](#)]
22. Klognerová, K.; Vosmanská, M.; Száková, J.; Mestek, O. Effect of growing conditions on selenium speciation in rapeseed (*Brassica napus*) tissue. *Chem. List* **2015**, *109*, 216–222.
23. Seppänen, M.M.; Kontturi, J.; Heras, I.L.; Madrid, Y.; Cámara, C.; Hartikainen, H. Agronomic biofortification of *Brassica* with selenium-enrichment of SeMet and its identification in *Brassica* seeds and meal. *Plant Soil* **2010**, *337*, 273–283. [[CrossRef](#)]
24. Feng, J.; Shi, Q.; Wang, X.; Wei, M.; Yang, F.; Xu, H. Silicon supplementation ameliorated the inhibition of photosynthesis and nitrate metabolism by cadmium (Cd) toxicity in *Cucumis sativus* L. *Sci. Hort.* **2010**, *123*, 521–530. [[CrossRef](#)]
25. Kaur, N.; Sharma, S.; Kaur, S.; Nayyar, H. Selenium in agriculture: A nutrient or contaminant for crops? *Arch. Agron. Soil Sci.* **2014**, *60*, 1593–1624. [[CrossRef](#)]
26. Drahoňovský, J.; Száková, J.; Mestek, O.; Tremlová, J.; Kaňa, A.; Najmanová, J.; Tlustoš, P. Selenium uptake, transformation and inter-element interactions by selected wildlife plant species after foliar selenate application. *Environ. Exp. Bot.* **2016**, *125*, 12–19. [[CrossRef](#)]
27. Filek, M.; Zembala, M.; Kornaś, A.; Walas, S.; Mrowiec, H.; Hartikainen, H. The uptake and translocation of macro- and microelements in rape and wheat seedlings as affected by selenium supply level. *Plant Soil* **2010**, *336*, 303–312. [[CrossRef](#)]
28. El-Ramady, H.; Abdalla, N.; Taha, H.S.T.; Alshaal, A.; El-Henawy, S.E.D.A.; Faizy, M.S.; Shams, S.M.; Youssef, T.; Shalaby, Y.; Bayoumi, N.; et al. Selenium and NanoSelenium in Plant Nutrition. *Environ. Chem. Lett.* **2016**, *14*, 123–147. [[CrossRef](#)]
29. FAO. *Food Outlook*; Global Information and Early Warning System; Food and Agriculture Organization: Rome, Italy, 2015. [[CrossRef](#)]
30. Habibi, G. Contrastive response of *Brassica napus* L. to exogenous salicylic acid, selenium and silicon supplementation under water stress. *Arch. Biol. Sci.* **2015**, *67*, 397–404. [[CrossRef](#)]
31. Kerr, C.; Walker, E.; Booth, J. Agricultural aspects of rape and other *Brassica* products. *Eur. J. Lipid Sci. Technol.* **2001**, *103*, 441–446. [[CrossRef](#)]
32. Jahangir, M.; Abdel-Farid, I.B.; Simoh, S.; Kim, H.K.; Verpoorte, R.C.Y. Investigation of *Brassica* biochemical status by NMR-based metabolomics. *Acta Hort.* **2012**, *936*, 163–172. [[CrossRef](#)]
33. Murphy, E.J. Camelina (*Camelina sativa*). In *Industrial Oil Crops*; Elsevier Inc.: Amsterdam, The Netherlands, 2016; pp. 207–230.
34. Ahmad, Z.; Waraich, E.A.; Ahmad, R.; Shahbaz, M. Modulation in water relations, chlorophyll contents and antioxidants activity of maize by foliar phosphorus application under drought stress. *Pak. J. Bot.* **2017**, *49*, 11–19.
35. Latef, A.A.A.; Tran, L.-S.P. Impacts of priming with silicon on the growth and tolerance of maize plants to alkaline stress. *Front. Plant Sci.* **2016**, *7*, 243. [[CrossRef](#)]
36. Mohd, R.S.; Faisal, A.; Azwan, A.; Chye, F.Y.; Matanjun, P. Crude proteins, total soluble proteins, total phenolic contents and SDS-PAGE profile of fifteen varieties of seaweed from Semporna, Sabah, Malaysia. *Int. Food Res. J.* **2015**, *22*, 1483–1493.
37. Sood, V.; Rawat, D.; Khanna, R.; Sharma, S.; Gupta, P.K.; Alam, S.; Sarin, S.K. Study of carnitine/acylcarnitine and amino acid profile in children and adults with acute liver failure. *J. Pediatr. Gastroenterol. Nutr.* **2017**, *64*, 869–875. [[CrossRef](#)] [[PubMed](#)]
38. Sinay, H.; Karuwal, R.L. Proline and total soluble sugar content at the vegetative phase of six corn cultivars from Kisar Island Maluku, grown under drought stress conditions. *IJAAR* **2014**, *2*, 77–82.
39. Cao, B.; Li, W.L.; Gao, S.; Xia, J.; Xu, K. Silicon-mediated changes in radial hydraulic conductivity and cell wall stability are involved in silicon-induced drought resistance in tomato. *Protoplasma* **2017**, *254*, 2295–2304. [[CrossRef](#)]
40. Ribera-Fonseca, A.; Rumpel, C.; Mora, M.D.L.L.; Nikolic, M.; Cartes, P. Sodium silicate and calcium silicate differentially affect silicon and aluminium uptake, antioxidant performance and phenolics metabolism of ryegrass in an acid Andisol. *Crop Pasture Sci.* **2018**, *69*, 205–215. [[CrossRef](#)]
41. Zhao, L.; Temelli, F.; Chen, L. Encapsulation of anthocyanin in liposomes using supercritical carbon dioxide: Effects of anthocyanin and sterol concentrations. *J. Funct. Foods* **2017**, *34*, 159–167. [[CrossRef](#)]
42. Wang, J.Y.; Xiong, Y.C.; Li, F.M.; Siddique, K.H.; Turner, N.C. Effects of drought stress on morphophysiological traits, biochemical characteristics, yield, and yield components in different ploidy wheat: A meta-analysis. *Adv. Agron.* **2017**, *143*, 139–173. [[CrossRef](#)]
43. Salama, K.H.A.; Mansour, M.M.F.; Al-Malawi, H.A. Glycinebetaine priming improves salt tolerance of wheat. *Biologia* **2015**, *70*, 1334–1339. [[CrossRef](#)]

44. Tale, A.S.; Haddad, R. Study of silicon effects on antioxidant enzyme activities and osmotic adjustment of wheat under drought stress. *Czech J. Genet. Plant Breed.* **2011**, *47*, 17–27. [[CrossRef](#)]
45. R Core Team. *R: A Language and Environment for Statistical Computing*; The R Foundation for Statistical Computing: Vienna, Austria, 2014; Available online: <http://www.R-project.org/> (accessed on 31 January 2021).
46. Kuznetsov, V. Selenium regulates the water status of plants exposed to drought. *Dokl. Biol. Sci.* **2003**, *390*, 266–268. [[CrossRef](#)] [[PubMed](#)]
47. Nawaz, F.; Ashraf, M.Y.; Ahmad, R.; Waraich, E.A.; Shabbir, R.N.; Bukhari, M.A. Supplemental selenium improves wheat grain yield and quality through alterations in biochemical processes under normal and water deficit conditions. *Food Chem.* **2015**, *175*, 350–357. [[CrossRef](#)] [[PubMed](#)]
48. Ahmad, R.; Waraich, E.A.; Nawaz, F.; Ashraf, M.Y.; Khalid, M. Selenium (Se) improves drought tolerance in crop plants—A myth or fact? *J. Sci. Food Agric.* **2016**, *96*, 372–380. [[CrossRef](#)] [[PubMed](#)]
49. Yao, X.; Chu, J.; Wang, G. Effects of selenium on wheat seedlings under drought stress. *Biol. Trace Elem. Res.* **2009**, *130*, 283–290. [[CrossRef](#)]
50. Ahmad, Z.; Waraich, E.A.; Akhtar, S.; Anjum, S.; Ahmad, T.; Mahboob, W.; Hafeez, O.B.A.; Tapera, T.; Labuschagne, M.; Rizwan, M. Physiological responses of wheat to drought stress and its mitigation approaches. *Acta Physiol. Plant.* **2018**, *40*, 80. [[CrossRef](#)]
51. Nageswara, R.R.C.; Talwar, H.S.; Wright, G.C. Rapid assessment of specific leaf area and leaf nitrogen in peanut (*Arachis hypogaea* L.) using a chlorophyll meter. *J. Agron. Crop Sci.* **2001**, *186*, 175–182. [[CrossRef](#)]
52. Iturbe-Ormaetxe, I.; Escuredo, P.R.; Arrese-Igor, C.; Becana, M. Oxidative damage in pea plants exposed to water deficit or paraquat. *Plant Physiol.* **1998**, *116*, 173–181. [[CrossRef](#)]
53. Waraich, E.A.; Rashid, F.; Ahmad, Z.; Ahmad, R.; Ahmad, M. Foliar applied potassium stimulate drought tolerance in canola under water deficit conditions. *J. Plant Nutr.* **2020**, *43*, 1923–1934. [[CrossRef](#)]
54. Manivannan, P.; Jaleel, C.A.; Sankar, B.; Kishorekumar, A.; Somasundaram, R.; Lakshmanan, G.A.; Panneerselvam, R. Growth, biochemical modifications and proline metabolism in *Helianthus annuus* L. as induced by drought stress. *Colloids Surf. B Biointerfaces* **2007**, *59*, 141–149. [[CrossRef](#)] [[PubMed](#)]
55. Fotovat, R.; Valizadeh, M.; Toorchi, M. Association between water-use efficiency components and total chlorophyll content (SPAD) in wheat (*Triticum aestivum* L.) under well-watered and drought stress conditions. *J. Food Agric. Environ.* **2007**, *5*, 225–227.
56. Khayatnezhad, M.; Gholamin, R.; Jamaati-e-Somarin, S.; Zabihi-e-Mahmoodabad, R. The leaf chlorophyll content and stress resistance relationship considering in corn cultivars (*Zea mays*). *Adv. Environ. Biol.* **2011**, *5*, 118–122.
57. EL Sabagh, A.; Hossain, A.; Barutçular, C.; Khaled, A.A.; Fahad, S.; Anjorin, F.B.; Islam, M.S.; Ratnasekera, D.; Kizilgeçi, F.; Singh, G.; et al. Sustainable maize (*Zea mays* L.) production under drought stress by understanding its adverse effect, survival mechanism and drought tolerance indices. *J. Exp. Biol. Agric. Sci.* **2018**, *6*, 282–295. [[CrossRef](#)]
58. EL Sabagh, A.; Hossain, A.; Barutçular, C.; Gormus, O.; Ahmad, Z.; Hussain, S.; Islam, M.S.; Alharby, H.; Bamagoos, A.; Kumar, N.; et al. Effects of drought stress on the quality of major oilseed crops: Implications and possible mitigation strategies—A review. *Appl. Ecol. Environ. Res.* **2019**, *17*, 4019–4043. [[CrossRef](#)]
59. Ahmadizadeh, M. Physiological and agro-morphological response to drought stress. *Middle East J. Sci. Res.* **2013**, *13*, 998–1009. [[CrossRef](#)]
60. Shabbir, R.N.; Waraich, E.A.; Ali, H.; Nawaz, F.; Ashraf, M.Y.; Ahmad, R.; Awan, M.I.; Ahmad, S.; Irfan, M.; Hussain, S.; et al. Supplemental exogenous NPK application alters biochemical processes to improve yield and drought tolerance in wheat (*Triticum aestivum* L.). *Environ. Sci. Pollut. Res.* **2015**, *23*, 2651–2662. [[CrossRef](#)]
61. Zhao, Y.; Wu, P.; Wang, Y.; Feng, H. Different approaches for selenium biofortification of pear-jujube (*Zizyphus jujuba* M. cv. *Lizao*) and associated effects on fruit quality. *J. Food Agric. Environ.* **2013**, *225*, 1–5.
62. Silva, R.; Filgueiras, L.; Santos, B.; Coelho, M.; Silva, M.; Estrada-Bonilla, G.; Vidal, M.; Baldani, J.I.; Meneses, C. *Gluconacetobacter diazotrophicus* changes the molecular mechanisms of root development in *Oryza sativa* growing under water stress. *Int. J. Mol. Sci.* **2020**, *21*, 333. [[CrossRef](#)]
63. Schiavon, M.; Pilon-Smits, E.A.H. The fascinating facets of plant selenium accumulation—Biochemistry, physiology, evolution and ecology. *New Phytol.* **2017**, *213*, 1582–1596. [[CrossRef](#)] [[PubMed](#)]
64. Yao, X.; Chu, J.; Liang, L.; Geng, W.; Li, J.; Hou, G. Selenium improves recovery of wheat seedlings at rewatering after drought stress. *Russ. J. Plant Physiol.* **2012**, *59*, 701–707. [[CrossRef](#)]
65. Hawrylak-Nowak, B. Beneficial effects of exogenous selenium in cucumber seedlings subjected to salt stress. *Biol. Trace Elem. Res.* **2009**, *132*, 259–269. [[CrossRef](#)] [[PubMed](#)]
66. Abbas, S.M. Low levels of selenium application attenuate low temperature stress in *Sorghum* [*Sorghum bicolor* (L.) *moench.*] seedlings. *Pak. J. Bot.* **2013**, *45*, 1597–1604.
67. Akbulut, M.; Çakir, S. The effects of Se phytotoxicity on the antioxidant systems of leaf tissues in barley (*Hordeum vulgare* L.) seedlings. *Plant Physiol. Biochem.* **2009**, *48*, 160–166. [[CrossRef](#)]
68. Cechin, I.; Corniani, N.; De Fátima, F.T.; Cataneo, A.C. Ultraviolet-B and water stress effects on growth, gas exchange and oxidative stress in sunflower plants. *Radiat. Environ. Biophys.* **2008**, *47*, 405–413. [[CrossRef](#)] [[PubMed](#)]
69. Zahedi, H.; Rad, A.H.S.; Moghadam, H.R.T. Effects of zeolite and selenium applications on some agronomic traits of three *Canola* cultivars under drought stress. *Pesqui. Agropecuária Trop.* **2011**, *41*, 179–185. [[CrossRef](#)]

70. Pazoki, A.R.; Rad, A.H.S.; Habibi, D.; Paknejad, F.; Kobraee, S.; Hadayat, N. Effect of drought stress and selenium spraying on superoxide dismutase activity of winter rapeseed (*Brassica napus* L.) cultivars. *Int. J. Agric. Biosyst. Eng.* **2010**, *4*, 655–658. [[CrossRef](#)]
71. Zhang, L.; Li, Q.; Yang, X.; Xia, Z. Effects of sodium selenite and germination on the sprouting of chickpeas (*Cicer arietinum* L.) and its content of selenium, formononetin and biochanin A in the sprouts. *Biol. Trace Elem. Res.* **2011**, *146*, 376–380. [[CrossRef](#)]
72. Germ, M.; Stibilj, V.; Osvald, J.; Kreft, I. Effect of selenium foliar application on chicory (*Cichorium intybus* L.). *J. Agric. Food Chem.* **2007**, *55*, 795–798. [[CrossRef](#)] [[PubMed](#)]
73. Gao, L.; Caldwell, C.D.; Jiang, Y. Photosynthesis and growth of *Camelina* and *Canola* in response to water deficit and applied nitrogen. *Crop Sci.* **2018**, *58*, 393–401. [[CrossRef](#)]
74. Ahmad, Z.; Waraich, E.A.; Barutçular, C.; Alharby, H.; Bamagoos, A.; Kizilgeci, F.; Öztürk, F.; Hossain, A.; Bayoumi, Y.; EL Sabagh, A. Enhancing drought tolerance in *Camelina sativa* L. and *Canola napus* L. through application of selenium. *Pak. J. Bot.* **2020**, *52*. [[CrossRef](#)]
75. Curtin, D.; Hanson, R.; Van Der Weerden, T.J. Effect of selenium fertiliser formulation and rate of application on selenium concentrations in irrigated and dryland wheat (*Triticum aestivum*). *N. Z. J. Crop Hortic. Sci.* **2008**, *36*, 1–7. [[CrossRef](#)]
76. Proietti, P.; Nasini, L.; Del Buono, D.; D'Amato, R.; Tedeschini, E.; Businelli, D. Selenium protects olive (*Olea europaea* L.) from drought stress. *Sci. Hortic.* **2013**, *164*, 165–171. [[CrossRef](#)]