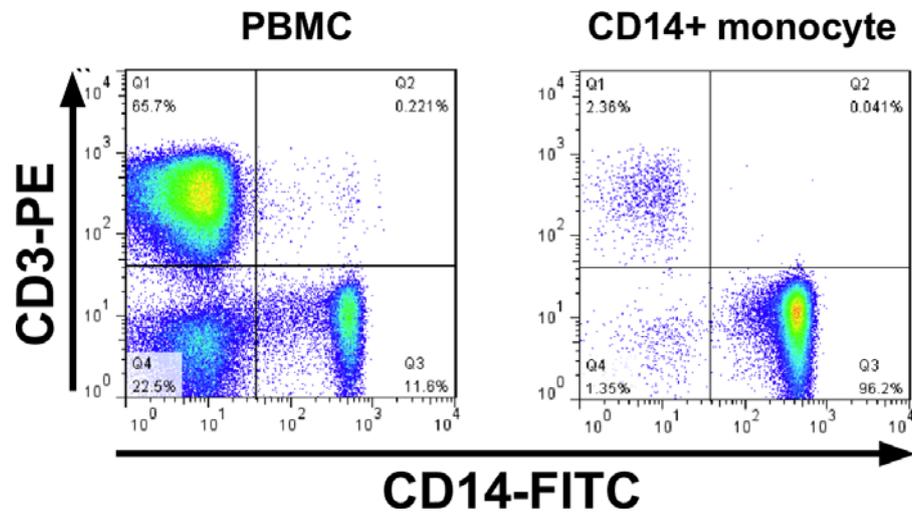
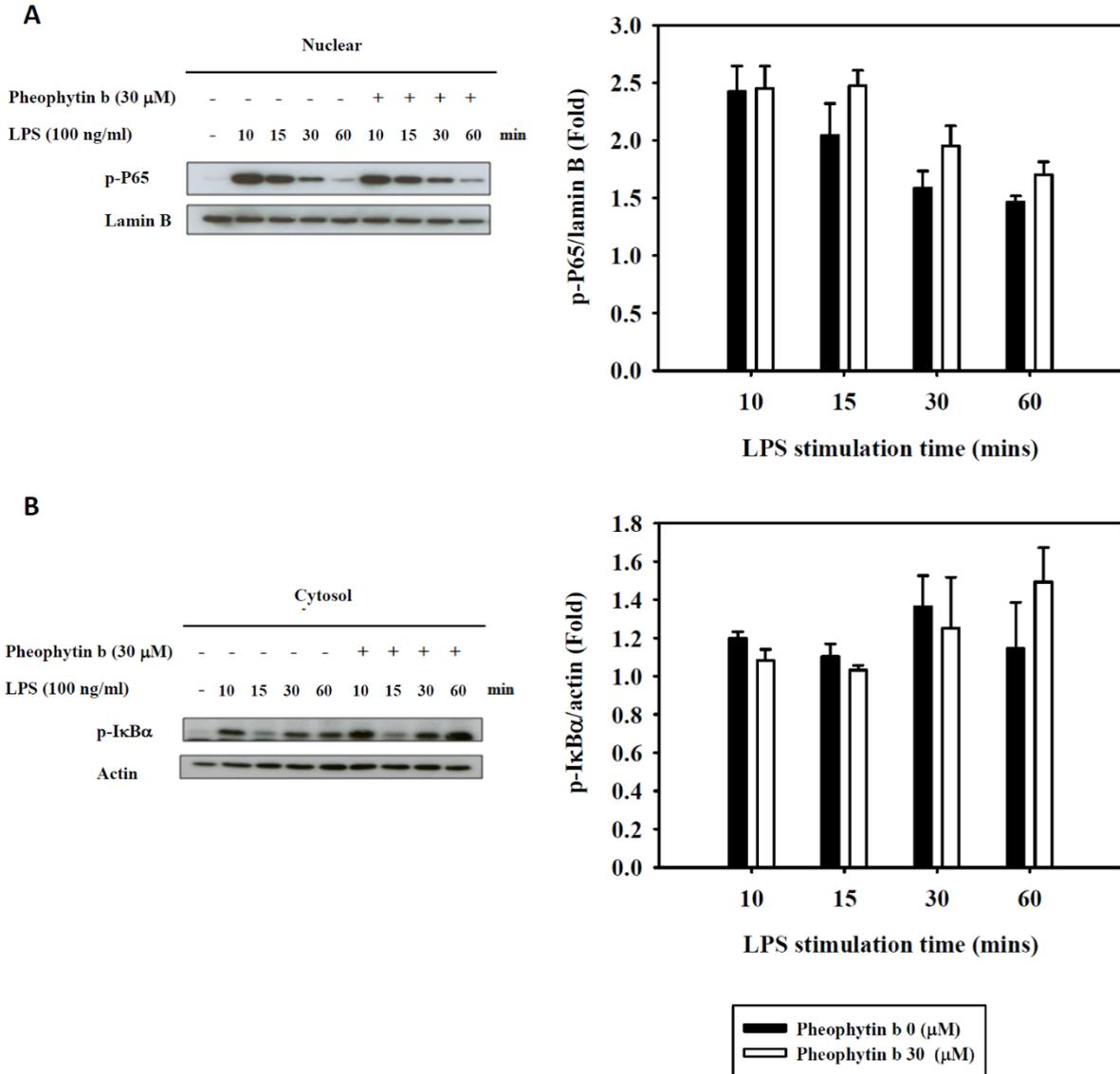


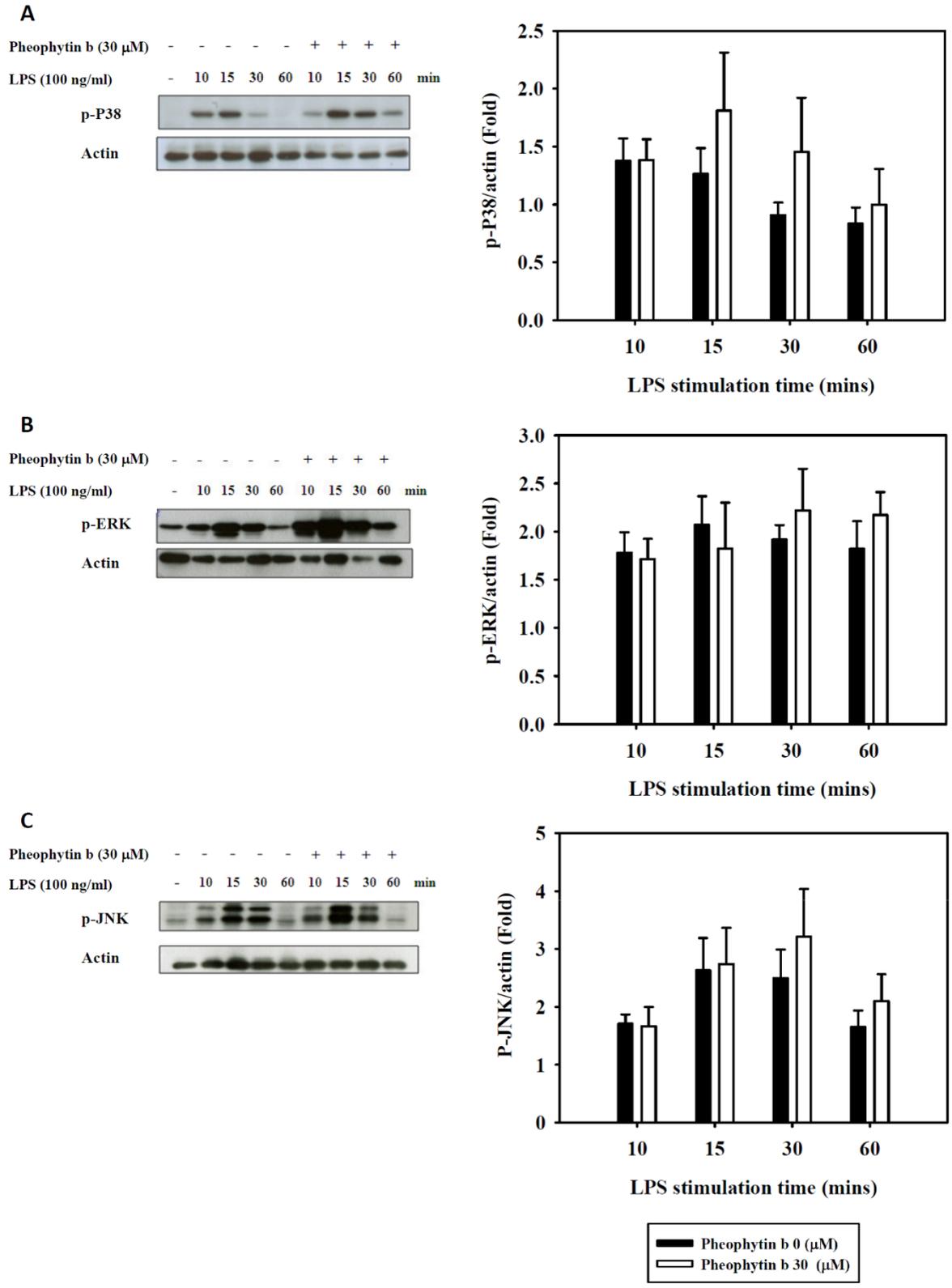
## Supplementary materials



**Figure S1.** The purity of CD14<sup>+</sup> monocytes was evaluated by flow cytometry. CD14<sup>+</sup> monocytes were enriched and isolated from PBMCs using the MiniMACS® Separator and human CD14 MicroBeads kit, according to the manufacturer's instructions. Post-enrichment, CD14<sup>+</sup> cells were labeled with CD14-FITC and CD3-PE and analyzed by flow cytometry. PBMCs were used as a control. After enrichment, CD14<sup>+</sup> positive cells due to >95% of purity of monocytes was noted.



**Figure S2.** There were no significant effects of Pheophytin-b (30 $\mu$ M) on the NF- $\kappa$ B pathway in LPS-stimulated RAW 264.7 cells. Cells were pre-treated with or without 30 $\mu$ M Pheophytin-b for 30 min, and nuclear and cytosolic proteins were harvested at four indicated time points (10, 15, 30 and 60 min) after LPS (100 ng/mL) stimulation. Western blot analysis of phosphorylated p65 levels (A) in the nuclear fraction of LPS-stimulated RAW 264.7 cells and its semi-quantification after normalization to lamin B levels in three different experiments. Western blot analysis of phosphorylated I $\kappa$ B $\alpha$  expression (B) in the cytosolic fraction of LPS-stimulated RAW 264.7 cells and its semi-quantification after normalization to actin levels in three different experiments.



**Figure S3.** There were no significant effects of Pheophytin-b (30 $\mu$ M) on the MAPK signaling pathways in LPS-stimulated RAW 264.7 cells. Cells were pre-treated with or without 30 $\mu$ M Pheophytin-b for 30 min, and total protein was harvested at four indicated time points (10, 15, 30 and 60 min) after LPS (100 ng/mL) stimulation. Western blot analysis of phosphorylated p38 levels (A), phosphorylated ERK levels (B), and phosphorylated JNK expression (C) in LPS-stimulated cells and its semi-quantification after normalization to actin in three different experiments.

