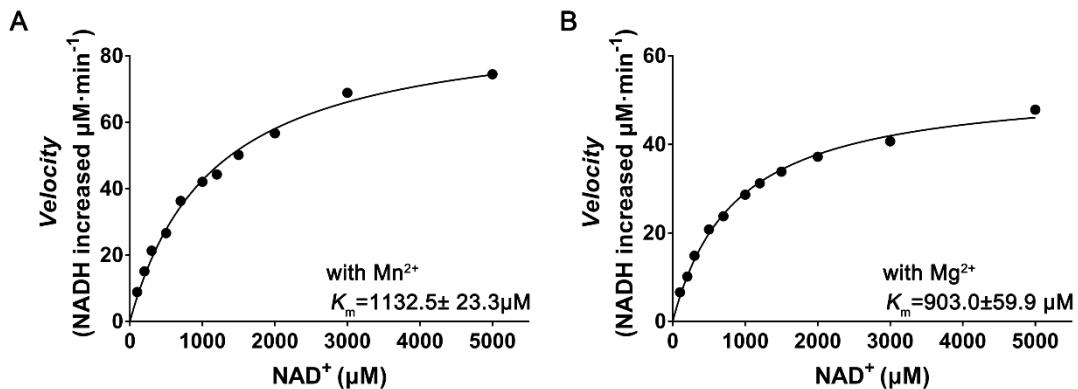




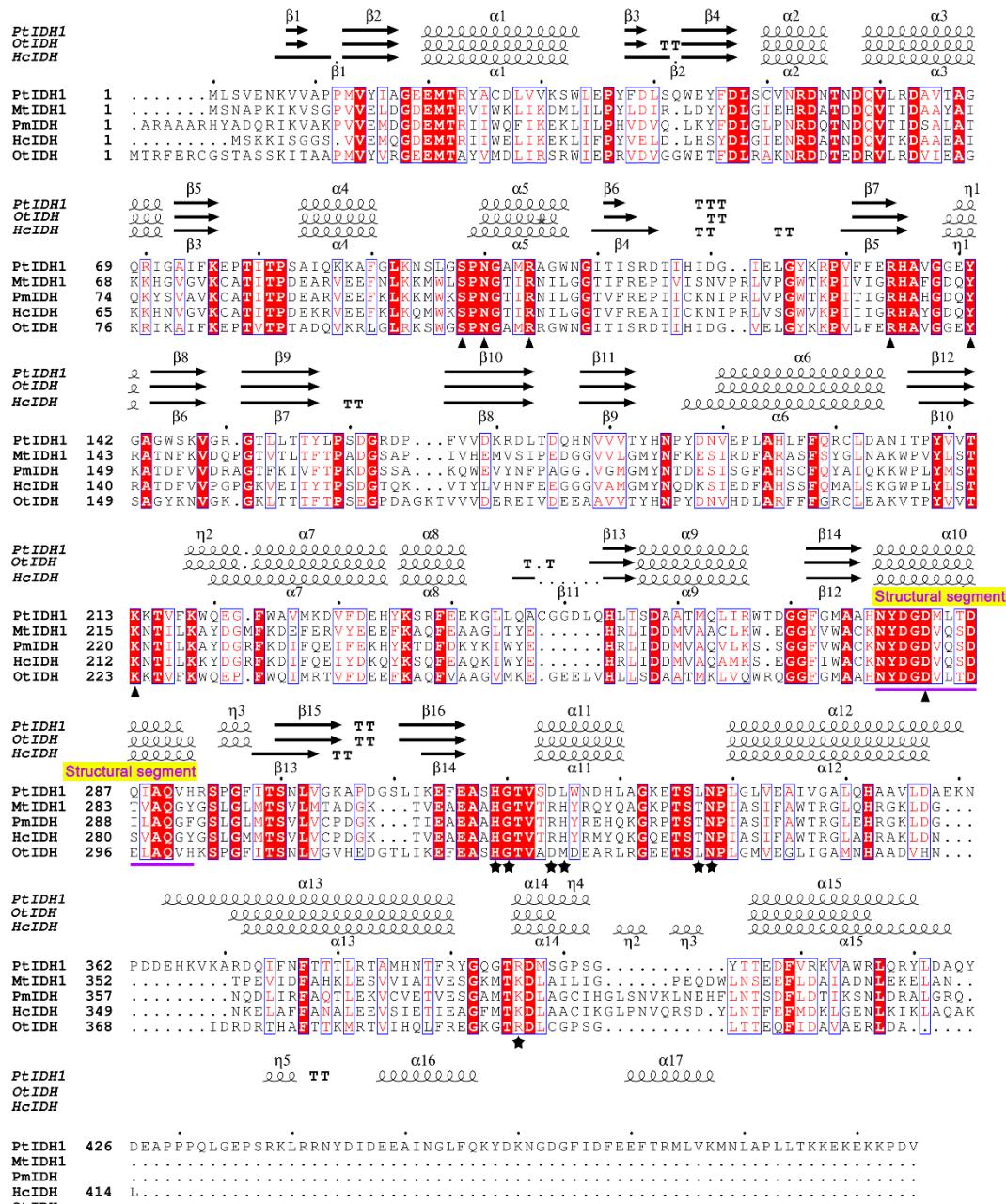
1      **Biochemical Characterization and Crystal Structure**  
2      **of a Novel NAD<sup>+</sup>-Dependent Isocitrate**  
3      **Dehydrogenase from *Phaeodactylum tricornutum***

4      Shi-Ping Huang, Lu-Chun Zhou, Bin Wen, Peng Wang and Guo-Ping Zhu

5      Supplementary Materials

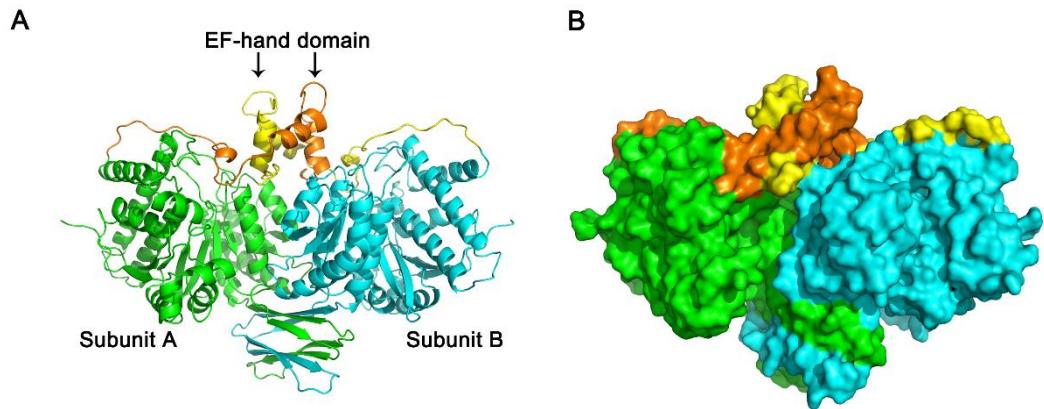


7      **Figure S1.** Kinetic analyses of the PtIDH1. The  $K_m$  values of PtIDH1 for NAD<sup>+</sup> were  $1132.5 \pm 23.3 \mu\text{M}$   
8      with Mn<sup>2+</sup> (A) and  $902.5 \pm 59.9 \mu\text{M}$  with Mg<sup>2+</sup> (B), respectively.



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**Figure S2.** Structure-based multiple sequence alignment of five Type II IDHs. The five IDHs for sequence alignment are PtIDH1, *O. tauri* NAD-IDH (OtIDH), human cytosolic NADP-IDH (HcIDH), porcine mitochondrial NADP-IDH (PmIDH) and *M. tuberculosis* NADP-IDH (MtIDH). The secondary structures of PtIDH1, OtIDH (PDB entry: 6IXN) and HcIDH (PDB entry: 1T0L) are placed above the alignment. The substrate and metal ion binding conserved amino acid residues of the IDH family are indicated by triangles. The residues that directly or indirectly interact with NAD(P)<sup>+</sup> are indicated by stars.



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18 **Figure S3.** Overall structure of PtIDH1 showed by cartoon (A) and surface (B), respectively. The small  
19 domain, clasp domain and EF-hand domain forming the dimeric interface.

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Coordinating ligand	Ca <sup>2+</sup> -binding EF-hand loop position											
	1 X (sc)		2 Y (sc)		3 Z (sc)		6 -Y (bb)		8 -X (sc+w)		10 -Z (sc2)	
	D 100%	K 29%	D 76%	G 56%	D 52%	G 96%	T 23%	I 68%	D 32%	F 23%	E 29%	E 92%
<b>Most common</b>			AQTVI SER	N	KRN	SN	FKQT ER	VL	STENGQ	YATL EN	DKAP N	D
PtIDH1-EF	D <sub>459</sub>	K	N	G	D	G	F	I	D	F	E	E <sub>470</sub>
M4A-EF	A	-	A	-	A	-	A	-	-	-	-	-
M6A-EF	A	-	A	-	A	-	A	-	A	-	-	A

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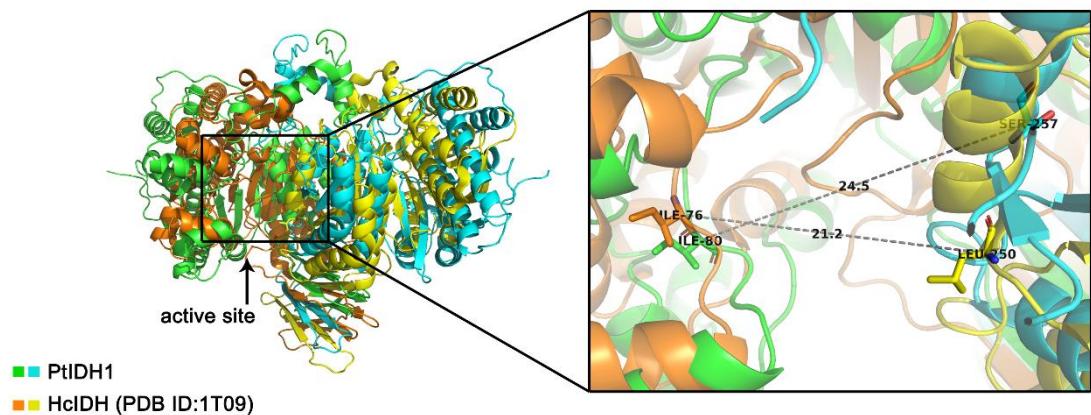
22 **Figure S4.** The sequence of EF-hand domain Ca<sup>2+</sup> binding loop. The Ca<sup>2+</sup> ligands are indicated with  
23 both the linear and coordination positions: 1(X), 3(Y), 5(Z), 7(-Y), 9(-X) and 12(-Z). The percentage of  
24 common residues (%) and other frequently observed residues are shown [1]. Side chain, sc; backbone,  
25 bb; water molecule, w. The sequence of PtIDH1-EF-hand loop also shown in the figure. M4A-EF: the  
26 EF-hand domain of PtIDH1 carries four-site mutant (D459A/D461A/D463A/F465A). M6A-EF: the EF-  
27 hand domain of PtIDH1 carries six-site mutant (D459A/D461A/D463A/F465A/D467A/E470A).

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**Figure S5.** Overlay of the dimers of PtIDH1 (green&cyan) and HcIDH (orange&yellow). The enlarged view (right) highlights the different sizes of active sites between PtIDH1 and HcIDH. The distance between the C<sub>α</sub>-atoms of Ile80 and Ser257 in PtIDH1 and Ile76 and Leu250 in HcIDH are indicated.

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**Table S1.** The kinetic parameters of eukaryotic IDHs toward isocitrate.

Enzyme	$S_{0.5}$ ( $\mu\text{M}$ )	Hill coefficient ( $n_H$ )
PtIDH1 ( $\text{Mg}^{2+}$ )	$608.1 \pm 4.0$	$1.36 \pm 0.03$
PtIDH1 ( $\text{Mn}^{2+}$ )	$84.9 \pm 5.2$	$1.43 \pm 0.14$
<i>O. tauri</i> NAD-IDH ( $\text{Mg}^{2+}$ ) [2]	8.1	1.91
<i>O. tauri</i> NAD-IDH ( $\text{Mn}^{2+}$ ) [2]	6.6	1.83
<i>C. reinhardtii</i> NAD-IDH ( $\text{Mn}^{2+}$ ) [3]	370	1.82
<i>S. cerevisiae</i> NAD-IDH ( $\text{Mg}^{2+}$ ) [4]	530	3.1
Pea NAD-IDH ( $\text{Mg}^{2+}$ ) [5]	300	3.1
Potato NAD-IDH ( $\text{Mg}^{2+}$ ) [6]	690	2.5

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Data are the mean  $\pm$  SD of at least three independent measurements.

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**Table S2.** X-ray data collection and structure refinement statistics.

PtIDH1_Apo	
Data collection	
X-ray source (Beam line)	SSRF (BL18U1)
Wavelength (Å)	0.97930
Space group	P 2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>
Cell dimensions	
a, b, c (Å)	71.97, 128.55, 124.30
α, β, γ (°)	90.00, 90.00, 90.00
Resolution (Å)	47.94-2.71 (2.86-2.71)
R <sub>merge</sub> (%) <sup>a</sup>	9.20 (71.60)
I/σI	18.60 (3.80)
Completeness (%)	100.0 (100.0)
No. of observed reflections	422213 (59008)
No. of unique reflections	31982 (4568)
Mosaicity (°)	0.10
Multiplicity	13.20 (12.90)
Refinement	
Resolution (Å)	47.04-2.80 (2.90-2.80)
No. of reflections working/test set	30800/1599
R <sub>work</sub> /R <sub>free</sub> <sup>b</sup>	0.227/0.283
No. of non-hydrogen atoms (protein/water)	7664/26
Mean B-factor (Å <sup>2</sup> )	
overall	73.88
chain A/B/S	73.01/74.87/54.97
RMSD from ideal geometry	
Bond length (Å)	0.01
Bond angles (°)	1.84
Ramachandran plot	
Core (%)	84.0
Allowed (%)	13.6
Generously allowed (%)	1.2
Disallowed (%)	1.2
PDB ID	6LKZ

The values in parentheses indicate the highest resolution shell. <sup>a</sup> $R_{\text{merge}} = \frac{\sum_{hkl} \sum_i |I_{hkl,i} - \langle I_{hkl} \rangle|}{\sum_{hkl} \sum_i I_{hkl,i}}$ . <sup>b</sup> $R_{\text{work}} = \frac{\sum ||F_{\text{obs}}| - |F_{\text{calc}}||}{\sum |F_{\text{obs}}|}$ , where  $F_{\text{obs}}$  and  $F_{\text{calc}}$  are the observed and calculated structure factor amplitudes.  $R_{\text{free}}$  was calculated same as  $R_{\text{work}}$ , using a randomly selected 5% reflections excluded from refinement.

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**Table S3.** Primers for amplification of wild-type and mutant PtIDH1 genes.

Name <sup>a</sup>	Sequences (5'-3') <sup>b</sup>
PtIDH1_S	GGAATT <u>CCATATGTCCTCCTGTCAACACTCCGAATC</u>
PtIDH1-53_S	GGAATT <u>CCATATGCTCAGTGTGAGAATAAAGTCGTG</u>
PtIDH1-EF_As	<u>CCGCTCGAGCGCGTCGAGGTATCGTTGTAACGCC</u>
PtIDH1_As	<u>CCGCTCGAGCACGTCCGGCTTCTCTCCTTTCTTC</u>
PtIDH1_M4A_S	GTAT <u>gccAAGgctGGTgccGGTgctATTGACTTGAAAG</u>
PtIDH1_M4A_As	CTTCAAAGTCAAT <u>agcACCggcACCagcCTTggcATAC</u>
PtIDH1_M6A_S	GT <u>gccGGTgctATTgccTTTGAAGcaTTCACTAGAATGC</u>
PtIDH1_M6A_As	GCATTCTAGTGAAT <u>gcTTCAAAggcAATagcACCggcAC</u>
Mutant_As	CTTCCTTTCAATATTATTGAAGCATTATCAGG

44      <sup>a</sup> "S" and "As": indicate the sense (S) and antisense (As) primers of the corresponding genes. <sup>b</sup>45      Underlined bases indicate the restriction sites. CATATG, NdeI; CTCGAG, XhoI. Underlined and bold  
46      lower-case bases indicate the mutation site.47      **References**

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