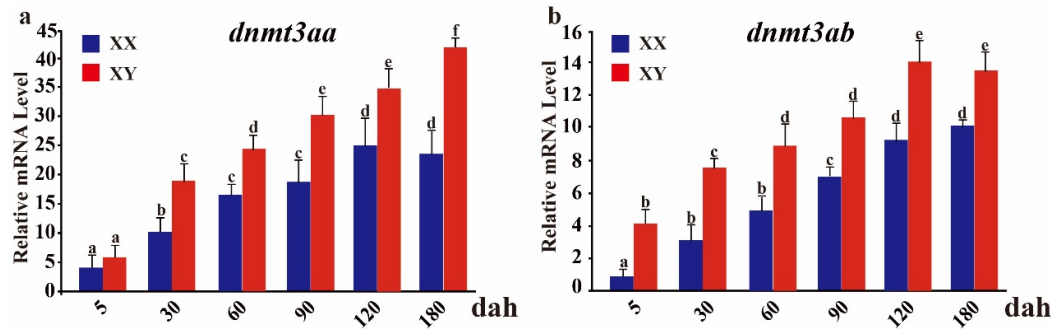
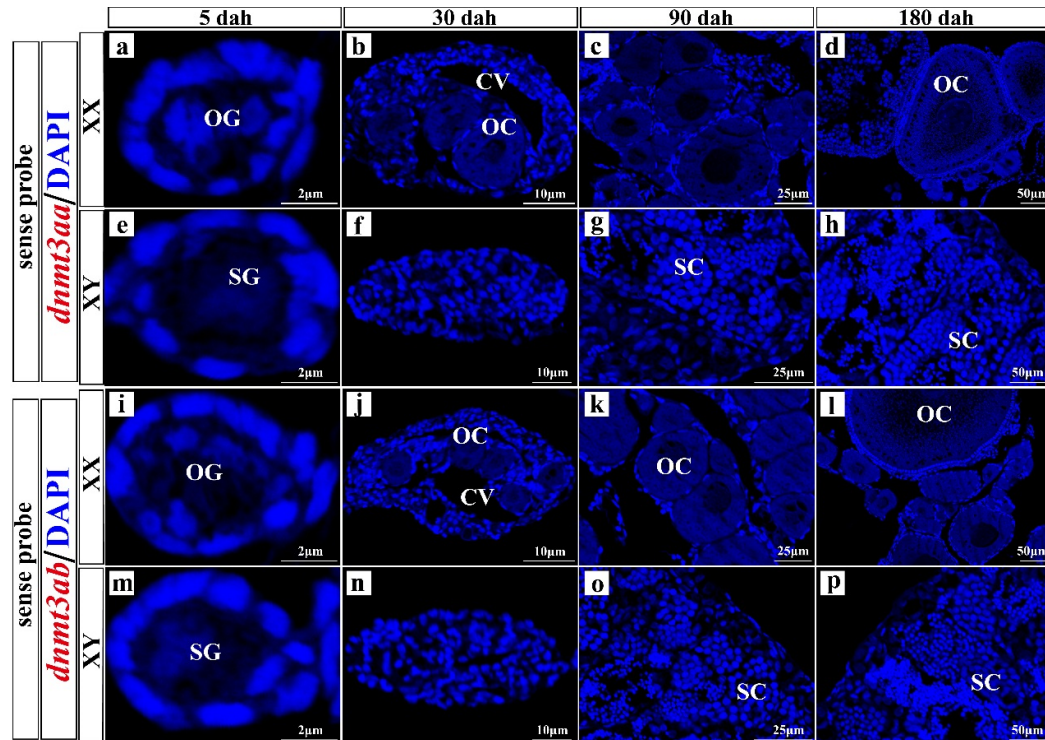


Supplementary Figure S1



Supplementary Figure S1. (a-b) Ontogenetic expression of *dnmt3aa* and *dnmt3ab* by qRT-PCR. Results were presented as mean \pm SD. *gapdh* was selected as the internal control. Different letters above the error bar indicate statistical differences at $p < 0.05$, as determined by one-way ANOVA followed by Tukey's post-hoc test. dah, day after hatching.

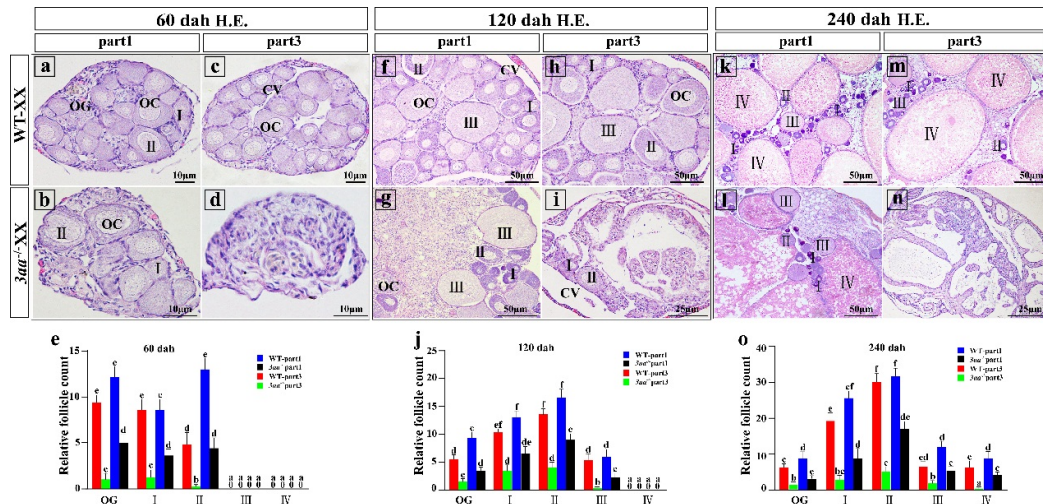
Supplementary Figure S2



Supplementary Figure S2. (a-p) Cellular localization of *dnmt3aa* and *dnmt3ab* in tilapia testes and ovaries at different developmental stages by FISH. No signal for *dnmt3aa* and *dnmt3ab*

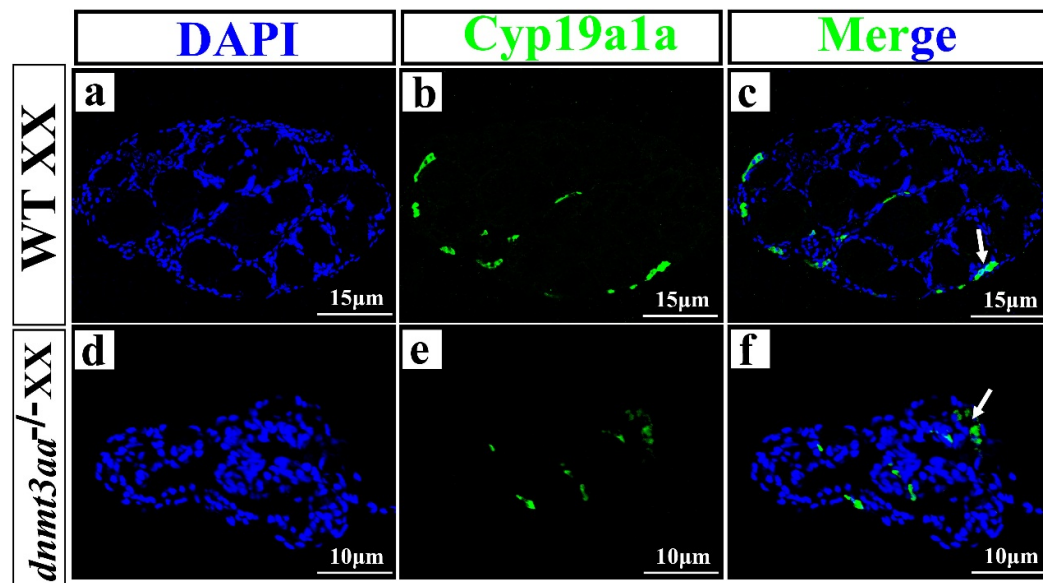
were detectable in the ovaries and testes using the sense probe. dah, day after hatching.

Supplementary Figure S3



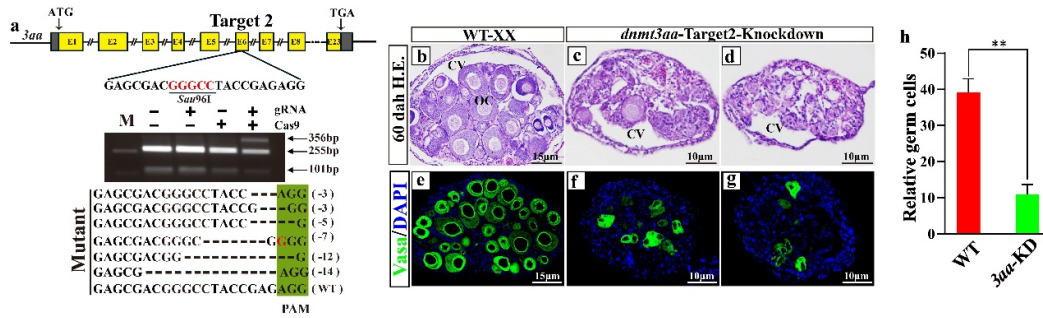
Supplementary Figure S3. (a-d, f-i, k-n) Histological observation WT and *dnmt3aa*^{-/-} ovaries of part1 and part3 at 60, 120 and 240 dah. (e, j, o) Statistical analysis the number of WT and *dnmt3aa*^{-/-} part1 and part3 ovarian follicles at 60, 120 and 240 dah. Results were presented as mean \pm SD. Different letters above the error bar indicate statistical differences at $p < 0.05$ as determined by one-way ANOVA followed by Tukey's post-hoc test. dah, day after hatching.

Supplementary Figure S4



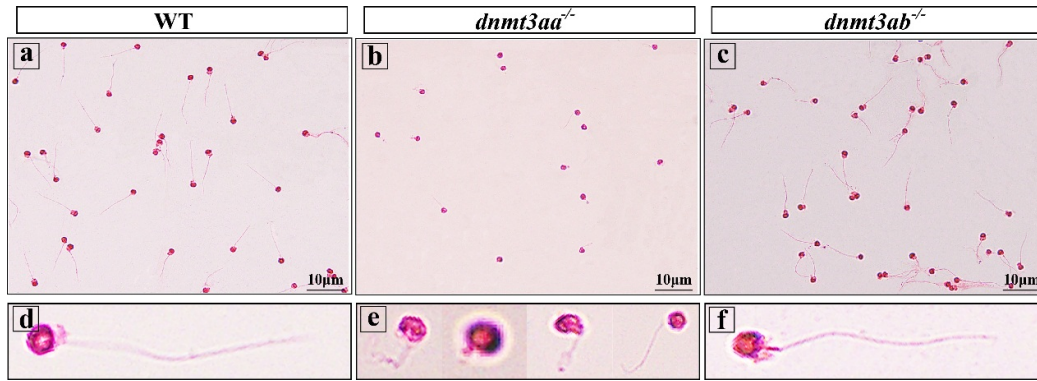
Supplementary Figure S4. Expression analysis of *cyp19a1a* (female pathway key gene) in WT (a-c) and *dnmt3aa*^{-/-} ovaries (d-f). Green fluorescence represents the Cyp19a1a signals (white arrows). Blue fluorescence represents the DAPI signals.

Supplementary Figure S5



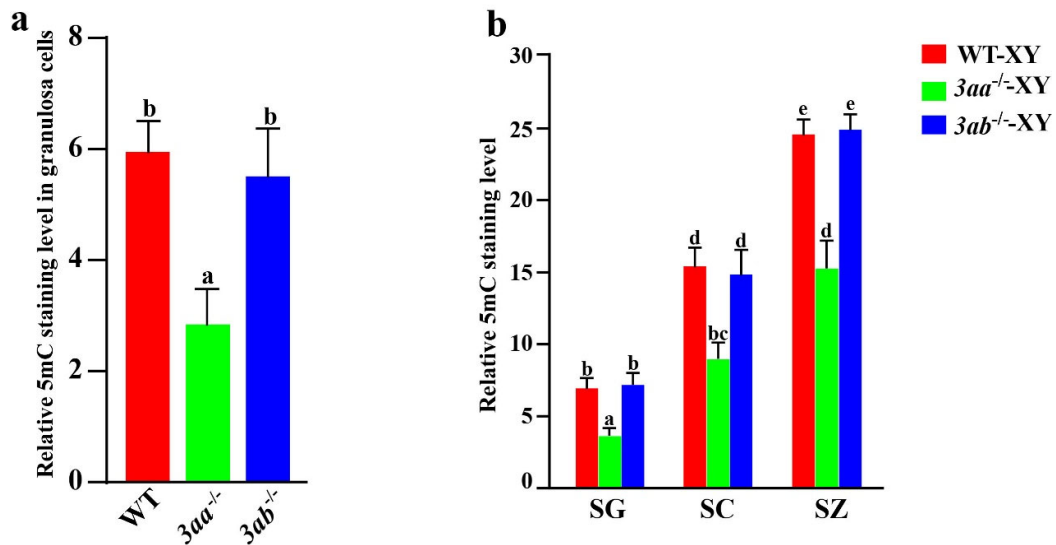
Supplementary Figure S5. Knockdown *dnmt3aa* by CRISPR/Cas9 in another target site. (a) Gene structures of *dnmt3aa* showing the second target site and the *Sau* 96I restriction site. The Cas9 mRNA and gRNA were added as indicated. Sanger sequencing results from the uncleaved bands were listed. The PAM was marked in light green. (b-d) Histological analysis ovaries of WT and *dnmt3aa* highly efficient knockdown females. (e-g) Analysis of Vasa (germ cells marker) expression in WT and *dnmt3aa* knockdown ovaries by IF. Green fluorescence represents the Vasa signals. Blue fluorescence represents the DAPI signals. (h) Statistical analysis the germ cells number of WT and *dnmt3aa* knockdown fish ($n = 5$). Results were presented as mean \pm SD. ** $p < 0.01$.

Supplementary Figure S6



Supplementary Figure S6. Papanicolaou staining analysis of WT, *dnmt3aa*^{-/-} and *dnmt3ab*^{-/-} sperms. d, e, f are the amplification of a, b and c, respectively.

Supplementary Figure S7



Supplementary Figure S7. (a) Statistical analysis relative 5-mC staining level of granulosa cells in WT, *dnmt3aa*^{-/-} and *dnmt3ab*^{-/-} ovaries at 120 dah. (b) Statistical analysis relative 5-mC staining level of WT, *dnmt3aa*^{-/-} and *dnmt3ab*^{-/-} spermatogonia, spermatocytes and spermatozoa in testes at 120 dah. Results were presented as mean ± SD. Different letters above the error bar indicate statistical differences at $p < 0.05$ as determined by one-way ANOVA followed by Tukey's post-hoc test. SG, spermatogonia; SC, spermatocytes; SZ, spermatozoa. dah, day after hatching.

Supplementary Table S1. All primer sequences used in this study.

Primer name	Sequence (5'-3')	Purpose
gRNA- <i>dnmt3aa</i> -target 1-F	TAATACGACTCACTATAGGCAGACCA	CRISPR/Cas9
	GGACAGTCCAGGTTTTAGAGCTAGAA	
gRNA- <i>dnmt3aa</i> -target 2-F	TAATACGACTCACTATAGGAGCGA	
	CGGGCCTACCGAGGTTTTAGAGCTAGAA	
gRNA- <i>dnmt3ab</i> -target 1-F	TAATACGACTCACTATAGGAGAGAA	CRISPR/Cas9
	CAATACTGGACTGTTTTAGAGCTAGAA	
gRNA-R	AGCACCGACTCGGTGCCAC	
<i>dnmt3aa</i> -KO-T1-detect-F	CCTGAAGAGCTTCTTAGCCACA	Mutant screening
<i>dnmt3aa</i> -KO-T1-detect-R	ATGCATGTTTAACAAACCTCTGC	
<i>dnmt3ab</i> -KO-detect-F	TGTCTCATTGTCCACTCAACCA	
<i>dnmt3ab</i> -KO-detect-R	TGTTGAATCAGACCTAATCACTGC	
<i>dnmt3aa</i> -KO-T2-detect-F	AACCTGCCAGCATGTACTCG	Mutant screening
<i>dnmt3aa</i> -KO-T2-detect-R	TGTTGTTGTGAAGGGGGAGG	
<i>dnmt3aa</i> -RT-PCR-F	CAGTCCAGAGGAGGGG	
<i>dnmt3aa</i> -RT-PCR-R	TCTCCCCTGGCGACTGGCTCGG	
<i>dnmt3ab</i> -RT-PCR-F	TGGACTCGGACCTGAT	RT-PCR
<i>dnmt3ab</i> -RT-PCR-R	CCTCCGCACGGCAAGAGGAAGG	
<i>gapdh</i> -F	AAGCTCATTTCTGTGTAT	
<i>gapdh</i> -R	CCTTTGCTGATTTCTTG	
<i>dnmt3aa</i> -ISH-F	CCCAGACACCAGAGAACGAC	ISH
<i>dnmt3aa</i> -ISH-R	TGTTGTTGTGAAGGGGGAGG	
<i>dnmt3ab</i> -ISH-F	TCCACCAAAGCTTTACCCCC	
<i>dnmt3ab</i> -ISH-R	CCTGTTTCATGCCAGGAAGGT	
<i>dnmt1</i> -qPCR-F	TGGCTCCCACGTTGATGAC	qRT-PCR
<i>dnmt1</i> -qPCR-R	AATTTGCCTTGCTCCTCCGT	
<i>dnmt3aa</i> -qPCR-F	TTGAGCCGGCCAAGTTGTAT	

<i>dnmt3aa</i> -qPCR-R	CATGCCGACAGTGATGGAGT	
<i>dnmt3ab</i> -qPCR-F	TCCACCAAAGCTTTACCCCC	
<i>dnmt3ab</i> -qPCR-R	CGGACGATACCCACAGTGAT	
<i>dnmt3bb.1</i> -qPCR-F	AATGAGAACAGCCCCCTGAC	
<i>dnmt3bb.1</i> -qPCR-R	CGCTCCTGAAGACTTGTCGG	
<i>dnmt3ba</i> -qPCR-F	CGAAAGAGGACGACAACCGT	
<i>dnmt3ba</i> -qPCR-R	GTTCATGCCAGGCAGGTTTC	
<i>dnmt3bb.2</i> -qPCR-F	CTTTACCTGAACCGGGCACA	
<i>dnmt3bb.2</i> -qPCR-R	TATTCCCTGGAACGCACAGG	
<i>baxa</i> -qPCR-F	TGCATCAGATTCACGATGAGTT	
<i>baxa</i> -qPCR-R	ACGAGTCGGCATGCAAAGTA	
<i>baxb</i> -qPCR-F	TGGCAATAAAGCAGTGACGA	
<i>baxb</i> -qPCR-R	CCTCTCTTGTTGGGACAAAGT	
<i>caspase3a</i> -qPCR-F	GGAAAACAATCAGCGGGCTC	
<i>caspase3a</i> -qPCR-R	CGTCAGTACCGTTTCGCTGA	
<i>caspase3b</i> -qPCR-F	ACTGTGGCTCAGATGAAAAAGC	
<i>caspase3b</i> -qPCR-R	GACCCTTGCAAGTGGTTTCCT	
<i>caspase8</i> -qPCR-F	CCGCAACAGCAGTTCACATT	
<i>caspase8</i> -qPCR-R	TCAGGAAGAGGGGTGGGATT	
<i>caspase9</i> -qPCR-F	TTCCTAGTAAGCTATCGCCTGA	
<i>caspase9</i> -qPCR-R	CTATGTTGGAGCCCTTGCGA	
AMH-F5	ATGGCTCCGAGACCTTGACTG	Genetic sex
AMH-R3	CAGAAATGTAGACGCCAGGTAT	identification
