

Article

Catching the Green—Diversity of Ruderal Spring Plants Traditionally Consumed in Bulgaria and Their Potential Benefit for Human Health

Teodora Ivanova ^{1,*}, Andrey Marchev ², Mihail Chervenkov ^{1,3}, Yulia Bosseva ¹, Milen Georgiev ², Ekaterina Kozuharova ⁴ and Dessislava Dimitrova ^{1,*}

¹ Department of Plant and Fungal Diversity and Resources, Institute of Biodiversity and Ecosystem Research, Bulgarian Academy of Sciences, 1113 Sofia, Bulgaria

² Laboratory of Metabolomics, Department of Biotechnology, The Stephan Angeloff Institute of Microbiology, Bulgarian Academy of Sciences, 139 Ruski Blvd, 4000 Plovdiv, Bulgaria

³ Faculty of Veterinary Medicine, University of Forestry, 1797 Sofia, Bulgaria

⁴ Department of Pharmacognosy, Faculty of Pharmacy, Medical University-Sofia, 1000 Sofia, Bulgaria

* Correspondence: tai@bio.bas.bg (T.I.); dessidim3010@gmail.com (D.D.)

Abstract: The global climate and societal challenges in the recent years urge us to strengthen food security; thus, the rediscovery of wild foods and foraging practices is also part of the sustainability agenda. Utilization of underappreciated sources such as ruderal plants could be a valuable option, especially for vulnerable parts of the society. We present data on traditional knowledge on spring edible ruderal plant taxa preserved in rural regions of Bulgaria, combining field studies in the period 2017–2022 that were compared to the available recent and historical ethnographic and (ethno)botanical literature. Semi-structured interviews were performed with representatives of 94 households in North and South Bulgaria, focusing on collection practices, used parts, and preparation methods. We list 65 edible ruderals, belonging to 22 plant families, of which 19 appeared only in the literature sources. Unlike in the Mediterranean tradition, edible ruderal plants in Bulgaria were regarded unfavorably, as poverty food. Amaranthaceae and Asteraceae were the most represented families, with 10 taxa each. About half of the taxa were collected for their leaves or whole young herbage that is used as pastry fillings, in stewed, and in cooked dishes. Taxa used in raw salads were mostly from the literature sources. The most diverse utilization was recorded in the southern-most regions of Bulgaria, where immediate tasting of the gathered plants was reported by the participants as the way to collect food plants. The bitter ones or those with an unappealing smell were considered non-edible and were avoided. References about biologically active compounds and potential benefits were collected, classified, and discussed in regard to their potential benefits for human health.

Keywords: wild edible plants; invasive plants; local knowledge; food security; foraging; traditional ecological knowledge



Citation: Ivanova, T.; Marchev, A.; Chervenkov, M.; Bosseva, Y.; Georgiev, M.; Kozuharova, E.; Dimitrova, D. Catching the Green—Diversity of Ruderal Spring Plants Traditionally Consumed in Bulgaria and Their Potential Benefit for Human Health. *Diversity* **2023**, *15*, 435. <https://doi.org/10.3390/d15030435>

Academic Editor: Michael Wink

Received: 20 February 2023

Revised: 7 March 2023

Accepted: 10 March 2023

Published: 15 March 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Diets based on the sustainable utilization of local biodiversity cater not only to the better overall well-being and health of the human population but also to the sustenance of ecological equilibrium and preservation of environment and natural resources [1–5]. Current assessments stress the importance of sustainable plant-based diets for the alleviation of negative impacts of industrial agriculture and for the mitigation of climate change [6–9]. Numerous studies have shown that plant-based foods provide a large spectrum of nutrients and bio-active non-nutrients [10,11], thus contributing to the overall fitness of the human body and reduction of risks of non-communicable diseases. It was shown that the increased number/diversity of consumed species elevates the nutritional adequacy of the diet [12]; however, traditions related to the consumption of both wild and cultivated plants are in

decline, especially in modern urbanized societies (including those in Europe) [13–17]. Ethnobotanical reports have shown that the consumption of leafy vegetables, especially those gathered from the wild, is still popular in the Mediterranean region, the Balkans, and Eastern Europe. Local traditions include the preparation of everyday, as well as festive, dishes; preserves; and medicinal foods. The knowledge about these practices has been preserved and handed down over the generations and is still maintained even in diasporic communities abroad [18–28]. Wild greens are also an important part of the Mediterranean diet (MD), supplying, especially rural communities, consumers with various vitamins, essential fatty acids, minerals, fibers, phenolic and terpenoid compounds, etc. [29–31]. However, in the first decades of the 21st century, some authors have reported on the fading of the traditions for collecting wild vegetables in Central and Eastern Europa, as well as further north, with *Rumex* species being most favored [15,32]. Global climatic and socioeconomic changes in the recent years have brought (back) the importance of wild foods/foraging practices, placing them in the spotlight, as a part of the sustainability agenda that could contribute to a more balanced utilization of biological resources, while also bringing (possible) benefits to human health [33–38].

“To catch the green” (*Da se hvanem za zeleno*, in Bulgarian) is a traditional expression used by Bulgarians that describes the consumption of spring leafy vegetables, which are the first fresh plant food to be consumed after the long winter. This phrase also expresses the hopes of the people for the new season. Indeed, before the industrialization of the agriculture and introduction of modern food preservation (ca. 1950s) leafy greens, either collected from the wild or cultivated in the home gardens, were a source of important nutrients not only towards the end of winter food stocks but also during famines and war times when the procurement of fresh nutritive food rich in vitamins was scarce [39]. The consumption of spring wild greens was also practiced in relation to Easter fasting when they were the only plant food available. Some early ethnographic works even describe communities living in mountainous areas as vegetarians, consuming meat scarcely [40].

Until the beginning of the new millennium, the diet patterns of Bulgarians were close to the recommendations of the MD, which is known to sustain a high life expectancy and healthy body weight, thus reducing the risks related to cardio-metabolic disorders, certain cancers, etc. [41,42]. However, current data for the food preferences of Bulgarians have shown a deviation from the MD, i.e., a reduced intake of plant-based foods and an increase of foods of animal origin [43,44].

Early ethnobotanical reports do not exhibit a systemic interest in the consumption of spring leafy greens (also called “lush greens”, *zlakove* in Bulgarian) by Bulgarians; hence, the information is scarce and scattered throughout the years. It indicates that collection from the wild was and still is unevenly popular around the country, with communities in mountainous areas adhering more frequently to such practices [39,45–48]. Therefore, similarly to other countries, the utilization of wild leafy greens in local culinary practices remains fairly unrecognized. It is due not only to the lack of archaeological evidence (which provides more information about the use of food crops) and the scarcity of ethnobotanical reports but also to the fact that wild greens have never been traded on the market [20,39,46,49,50], contrary to medicinal plants and mushrooms which were collected both for the internal market and for export, this being one of the traditional branches of the Bulgarian economy [51–53]. Their identification, collection, and consumption have remained closed in the house domain opposite to other Mediterranean countries, where such foods are marketed and specific collective terms for their consumption exist, e.g., (ta) *chòrta* in Calabria (Italy), Greece, and Cyprus; *mišanca*, *gruda*, *parapač*, etc., in Dalmatia; or *şxex* in Asia Minor and parts of Cyprus, among others [54–58]. The selection of wild greens for food was often based on their taste in search of palatable flavors [46,59]. The latter implied that the curiosity of the local people in their search for edible wild spring plants matches the unlimited options that exist in the wild. However, this approach presents possible risks related to the ingestion of toxic plants.

The ethnopharmacological application of edible and medicinal plants counters a long list, including treatment of coughs; hay fever and influenza; asthma; bronchitis; rheumatism; gastrointestinal, liver, renal, heart, brain, and skin disorders among many others [60–62]. Therefore, nowadays the interest of society and scientific organizations towards utilization of medicinal and edible plants as potential sources of pharmaceuticals, nutraceuticals, cosmeceuticals, and herbal health-promoting products for human well-being is constantly increasing [63]. However, the prevailing literature focuses more on the known medicinal plants, while the research on the health benefits of wild greens/ruderals used as food lags behind; hence, it needs more comprehensive research. Ruderals are species that occur on sites disturbed by human activities, such as roadsides, construction places surroundings, abandoned lots, railway lines, etc., that were found to be useful spots for foraging edible and medicinal plants (among other functionalities), especially in the urban and peri-urban areas [64–67].

In the current paper, we present the results of a several-year research study on the early spring consumption practices of ruderal plants in Bulgaria and link these results with a literature review on the bioactive compounds that have been registered in different parts of these plants that are known to benefit human health; we also make comments on their potential toxicity and/or adverse effects on the human body.

2. Materials and Methods

2.1. Study Area

Bulgaria occupies the central part of the Balkan Peninsula, Southeast Europe, bordering Romania (Danube River) to the north, Serbia and North Macedonia to the west, Greece and Turkey in Europe to the south, and the Black Sea to the east. The Balkan Mountain range, sprawling from West to the East, separates Bulgaria in northern and southern parts, thus impacting both the temperature and precipitation on both sides of the mountain. To the north, the climate is typical continental, while the southernmost lower parts of Bulgaria exhibit some Mediterranean features [68]. The territory elevation ranges substantially—from sea level to 2925 m. The territory of Bulgaria belongs to several biogeographical regions; therefore, the vascular flora is diverse and comprises 4064 species of spermatophytes affiliated to 921 genera and 159 families [69]. The arable land comprises about 50% of the country's territory, while forests account for 42%. There is an ongoing trend of arable land abandonment due to biophysical and economic factors [70]. Currently (2021), the urban population prevails, being nearly 3 times larger than the rural one [71].

2.2. Field Study and Research Approach

The presented data were collected during field studies in the period 2017–2022. It involved representatives of 94 households from small towns and villages in ten Bulgarian provinces—Blagoevgrad, Haskovo, Pazardzhik, Plovdiv, Sliven, Smolyan, and Sofia in the south; and Lovech, Montana, Pleven, and Vratsa in the north (Figure 1). The selection of participants was random or assisted by formal and informal community leaders (mayors, *chitalishte* (community center) managers, and/or educators). Participants engaged in semi-structured interviews focused on the various aspects of local plant knowledge related to food, spices, and medicines. Information on the consumption of annual and perennial synanthropic plant taxa thriving in disturbed habitats, including roadsides, riverbanks, field boundaries, fallows, and abandoned spaces in/along home yards and gardens, was specifically targeted as the most available foraging option. We also recorded the personal perceptions of the participants on the usage of wild edible plants and their attitudes towards the consumption of ruderals over the years by indirect questions about their current experience compared to the recollections from the past for their diet during the winter–spring period.



Figure 1. Map of the studied provinces (location of Bulgaria in the Balkan Peninsula on the bottom right).

The age of the participants was in the range of 35–85 years, with more than 60% being over 65 years of age. Female respondents prevailed (57.5% females to 42.5% males). Informed consent was verbally obtained from every participant prior to the interview. The guidelines prescribed in the Code of Ethics of the International Society of Ethnobiology [72] were followed during the field study, and their compliance was confirmed by the Scientific Council of the Institute of Biodiversity and Ecosystem Research, Bulgarian Academy of Sciences, acting as independent institutional Ethics Board (Decision No. 6/21/05/21).

Image data and/or reference specimens were collected for identification purposes; herbarium specimens were deposited as vouchers in the Herbarium of the Institute of Biodiversity and Ecosystem Research, Bulgarian Academy of Sciences (SOM). Identification of the plants was carried out at least to the genus rank when collected as juveniles, and later on, with the help of our respondents, who provided photos of the mature plant, the species were determined. We used the *Handbook of Bulgarian Vascular Flora* for the taxonomical identification [73]. Plant names were aligned with the Plant List (2013) [74]. Field data were compared to the available recent and historical ethnographic and (ethno)botanical literature and cited references where accessible [39,46,47,75–81]. Data on traditional Bulgarian food involving wild vegetables were collected from culinary books issued between 1890 and 1952 [82–88].

Information about biologically active compounds and toxicity was retrieved from scientific literature databases—Web of Science, Scopus, and AGRIS (FAO)—as well as published books and reports.

3. Results and Discussion

3.1. Plant Diversity and Consumption Practices

We registered a total of 65 edible ruderal plant taxa to be collected by Bulgarians from the wild during spring months, of which 19 were mentioned only in the literature sources (Figure 2a,b). The plants belonged to 22 families (Table 1). Amaranthaceae and Asteraceae were most represented, with ten taxa each, followed by Brassicaceae with nine and Polygonaceae with four species. The rest of the plant families were had three or less taxa. The leaves (35 species) and young whole herbage (26 species) were the most frequently used parts. However, the young shoots (e.g., *Humulus lupulus* L. and *Rubus* L. spp.), flowers/inflorescences, (e.g., *Robinia pseudoacacia* L. and *Sambucus nigra* L.), and unripe fruits of *Malva* L. spp. and

Table 1. Number of ruderal taxa consumed as spring food in Bulgaria per plant family.

Plant Family	Number of Taxa	Number of Taxa Consumed Raw	Taxa Consumed Only Raw
Amaranthaceae	10	5	
Asteraceae	10	4	* <i>Lapsana communis</i> L. * <i>Sonchus arvensis</i> L. * <i>Cardamine pratensis</i> L. * <i>Cardamine amara</i> L.
Brassicaceae	9	7	* <i>Lepidium campestre</i> (L.) R. Br. * <i>Lepidium latifolium</i> L. * <i>Lepidium perfoliatum</i> L.
Polygonaceae	4	4	<i>Fallopia aubertii</i> (L. Henry) Holub
Urticaceae	3	2	<i>Parietaria officinalis</i> L.
Malvaceae	3	2	<i>Malva sylvestris</i> L.
Apiaceae	3	3	-
Caryophyllaceae	3	-	-
Rosaceae	3	3	-
Boraginaceae	2	-	-
Papaveraceae	2	-	-
Geraniaceae	2	-	-
Ranunculaceae	2	-	-
Plantaginaceae	1	-	-
Portulacaceae	1	1	-
Araceae	1	-	-
Fabaceae	1	1	-
Phytolaccaceae	1	-	-
Cannabaceae	1	-	-
Convolvulaceae	1	-	-
Adoxaceae	1	1	-
Equisetaceae	1	-	-

* Species mentioned only in the literature sources.

The number of the ruderals known to be used in different spring salads was about twice higher (18) compared to those used as snacks; however, only *Rumex acetosa* L. and *Rumex acetosella* L. were found in both categories. In Spain, the number of species used raw was similar to that of those cooked in various ways, and, very often, the snacked plant food turns into salads and dishes once having reached home [89]. Hence, the number of snack-only species was more than twice as high as salad-only ones and about 30% of the raw-consumed species that were used both in salads and as snacks. Shepherding practices, especially when performed by children, were related to a higher number of wild species used for snacking [90]. Thus, it is highly possible that significant traditional knowledge on wild edible plants in Bulgaria has been lost due to the gradual abandonment of pastoral transhumance practices and transition to the sedentary lifestyle of nomad herders reported to be evident since the Balkan Wars onwards (1920s onwards) [91]. The restructuring of agriculture under Communism (1944–1989) into state-owned industrialized farming cost even more, losing knowledge on gathering wild plants for snacking while shepherding and while walking to distant arable plots [92].

The association of consumption of weeds and ruderals with wartimes and poverty at the beginning of the twentieth century also added to their negligence as vegetables [93,94]. It was argued that, among the wild leafy greens consumed in North and Central Europe, those with a large biomass (e.g., *U. dioica*) prevailed, while the inhabitants of South Europe collected more diverse species with a small size, probably in quest of a variety of tastes, as well [95]. In more recent years, however, the use of large amounts of agrochemicals was considered an important factor for the shrinkage of the natural populations of some wild edible species, and this adds to the general reluctance/worry to collect/buy ruderal species growing in/near agricultural lands, roads, etc., for the elevated amounts of toxic pollutants in these environments [15,96,97].

Although many of the *Atriplex* and *Chenopodium* species (Amaranthaceae) that occur in the wild are well-known as leafy greens, the Bulgarian ethnographic literature draws attention mostly to the cultivated *Atriplex hortensis* L. The limited number of publicly acknowledged edible greens is visible also in the reviewed cookbooks from the end of the 19th to the first half of the 20th century, where not more than five species were mentioned, and this tendency did not change in the later years [39,83,84,86–88,98]. The early Bulgarian culinary literature favored few spring leafy greens, especially as a part of the Lenten fare and vegetarian dishes, namely, the leaves of the said *Atriplex hortensis* along with *Rumex acetosa* L., *R. patientia* L., *Urtica dioica* L., and the unripe fruits of *Prunus ceracifera* Ehrh. The common grounds for all of them, except for *R. acetosa*, were home gardens and settlement surroundings, where they were readily available at no cost and collected only for home consumption. Consequently, these vegetables rarely appeared on the market, except for *Rumex patientia*, *R. acetosa*, and *Urtica dioica*, which can be found sometimes together with *Allium ursinum* leaves on farmers' markets in the bigger cities [59].

Contrary to the great awareness of the health benefits of (wild) greens as part of the MD in other countries [30,99], the traditional collection of wild greens in Bulgaria seems to be limited and sometimes ignored even by rural communities. Hence, Bulgarian tradition for the collection and preparation of wild spring vegetables is in a dire situation, and most of the taxa are known only locally in the villages near the southern border both in the eastern and western part of the country. They have not reached broader audiences in Bulgaria, as seniors rarely manage to promulgate them even in their own communities, especially to younger generations [79] (Figure 3).



Figure 3. Gathering of spring wild vegetables alongside village road in SW Bulgaria (Gabrene village, Blagoevgrad Province).

Most of the ruderals recorded as edible during our field work are mentioned in the ethnographic and ethnobotanical sources for Bulgaria. However, during our field studies, we found another 11 taxa that are consumed in South Bulgaria. Eight of the latter are known as edible from other countries around Europe and Aegean Turkey [90,95,100–103], while the remaining three, i.e., *Centaurea cyanus* L., *Erodium cicutarium* (L.) L'Hér, and *Geranium lucidum* L., were found to be used only as pastry fillings (*zelnik*) prepared by Anatolian Bulgarians living close to the Bulgarian–Greek–Turkish border [46]. In the case of *Crepis sancta* (L.) Bornm., our participants consume the young herbage, but the species has no local name, although the knowledge was ascribed to grandmothers.

A marked difference between Mediterranean and Bulgarian traditions in the collection of wild leafy greens is in the knowledge and use of representatives of the Asteraceae family of which, in Bulgaria, we confirmed the consumption of only seven species, a very limited number compared to the Mediterranean area [16,22,23,54,101,104–107]. Additionally, *Taraxacum sect. Taraxacum* F.H.Wigg., *Artemisia vulgaris* L., and *Centaurea cyanus* L. are more popular as medicinal plants rather than food [73,105], and this was valid also for other species, such as *Plantago lanceolata* L. and *Urtica urens* L. The latter is used in traditional spring dishes in Italy, Crete, Belarus, and other countries, while, in Bulgaria, it is not

popular as a food plant, and only *U. dioica* is used on a large scale, and the cultivated one has been marketed in stores in the recent years [22,32,95,108]. Contrary to the Mediterranean tradition to consume wild leafy vegetables as salads on their own and/or with eggs, Bulgarian culinary preparations are more focused on soups, stewing, braising, and shallow frying with wheat flour, rice, and other starchy ingredients, as well as in pastry fillings often with grains/rice that will absorb the moisture of the greens. Interestingly, the usage of flowers of *Papaver* species was not documented, while, in Central and Western Europe, they were a popular colorant for deserts, cheeses, and wines [109,110]. We witnessed some differences in the processing of the greens that are used in the filling for the *zelnik* pastry. In SE Bulgaria, the preparation includes only chopping and mixing of the gathered fresh greens because the ready *zelnik* is expected to have tender but springy bite. Therefore, a mixture of diverse edible greens collected from the wild is preferred to *Lactuca sativa* L. or *Portulaca oleraceae* L. that are grown in the home gardens but considered to have a soft, less crispy texture [46]. Along the same time, in SW Bulgaria, the young parts of the wild greens *Artemisia vulgaris*, *Erigeron annuus* (L.) Pers., *Plantago lanceolata*, and *Doronicum orientale* Hoffm. were thoroughly cooked, and consequently the mixture was used either as pastry filling or directly consumed as a spread on a slice of bread. Similarly, the leaves of *Arum maculatum* L. collected by the Roma community in the vicinity of Sliven (Central Balkan Mts.), were dried and, prior to cooking, boiled with several changes of the water to improve its palatability. However, the Roma people are not aware about the toxicity of the plant and do not relate the preparation technique to its reduction (Figure 4).



Figure 4. Dried *Arum maculatum* (snake dock, *zmiyski lapad* in Bulgaria) from Sliven province used for preparation of beef stews.

3.2. Bioactive Compounds and Potential Health Benefits

Edible ruderal plants, some of which are also traditionally used medicinal plants, are a source of primary nutritional substances (proteins, fats, sugars, vitamins, and minerals), as well as, secondary/specialized metabolites, including phenolic acids, flavonoids, anthocyanins, tannins, terpenoids, alkaloids, steroidal saponins, essential oils, glucosinolates, etc. [111]. The chemical structures of some of the molecules are presented in Figure 5. The phytochemical research usually addresses the plant parts that are used for medicinal purposes. Therefore, in some cases, we do not find published data for those plant parts that are used for food, particularly at the early stages of the plant ontogenesis. However, finding them in other plant parts is an indication that such compounds might be present, even though in small amounts. The preparation method is important, as well. Some of these compounds would break at high temperature; others would be better extracted or vice versa. For example, flavonoids were found to be more thermostable among other polyphenols [112]. Additionally, different cooking techniques were found to affect the content of

phenolic compounds in various ways, e.g., pressure cooking, frying, and steaming being among least destructive [113]. However, these practices are not typical for the Bulgarian traditional cuisine, which favors slow-cooking techniques and baking, which could be found readily also in most Balkan countries [39,114–116]. Fermented leafy greens (e.g., *Atriplex* and *Malva*), as functional food, combine the benefits of the bioactive compounds in the plants with the benefits of microbial origin. This practice was reported by other Bulgarian authors in the past [75], but we have not recorded any of our respondents to ferment spring greens.

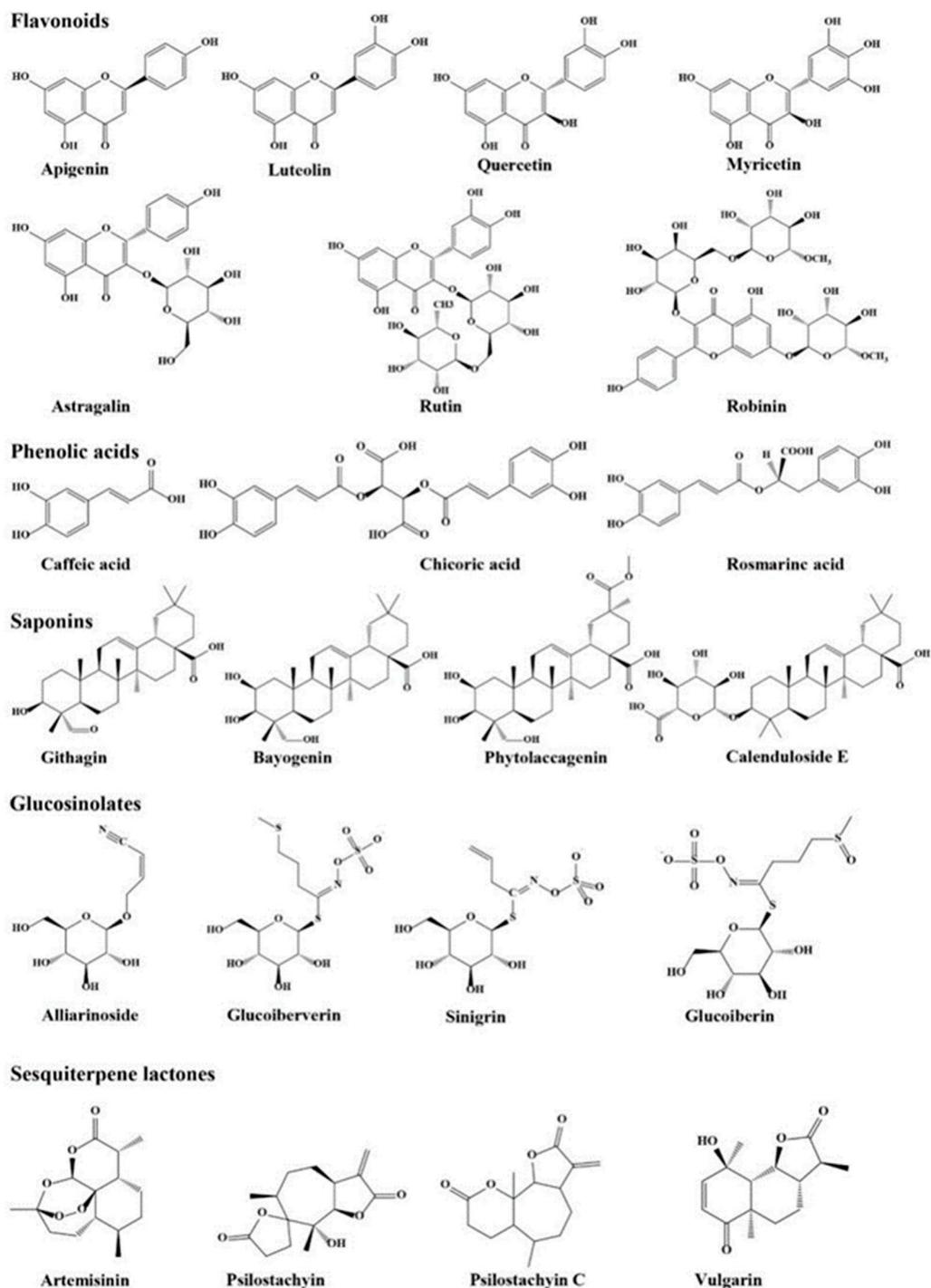


Figure 5. Chemical structures of some plant-derived compounds responsible for the biological activities reported for the taxa covered in the study.

Most commonly found biologically active compounds typical for the consumed taxa belonged to polyphenols—mainly flavonoids and phenolic acids and their derivatives, respectively, 74% and 51% of the taxa (Figure 6). Other important nutritives, such as fatty acids (20%), tocopherols (18%), and carotenoids (11%), were reported. Details on major metabolites and toxic substances in spring edible ruderals consumed traditionally in Bulgaria are presented in Supplementary Table S1. Quercetin, kaempferol, apigenin, and their derivatives and caffeic, p-coumaric, and chlorogenic acids were most frequently reported among the flavonoids and phenolic acids, respectively. Their numerous biological activities are proven by in vitro and in vivo tests and comprise antimicrobial, antioxidant, and wound-healing activities; reduce the risk of myocardial infarction, cancer development, have anti-inflammatory, and immunomodulatory functions; and protect the central nervous system and others [112–115,117].

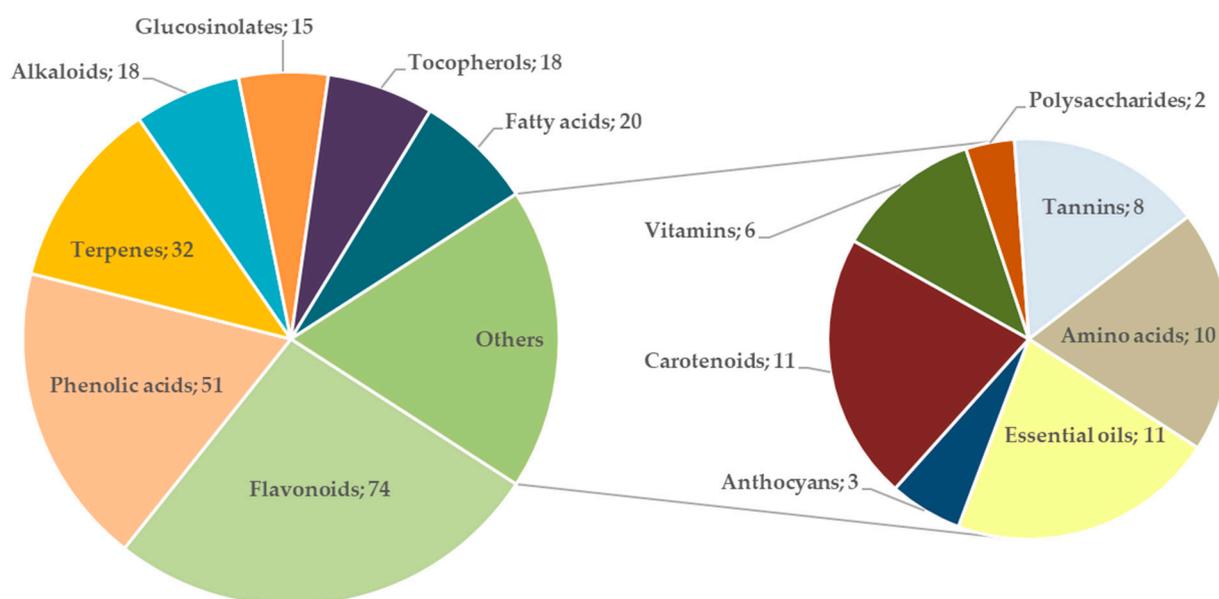


Figure 6. Major groups of compounds (calculated as percentage of the taxa) reported for the screened edible ruderals consumed in Bulgaria.

3.3. Potential Medicinal Benefits

The species from Amaranthaceae and Asteraceae that are the most frequently consumed spring edible ruderals are also frequently used as medicinal plants in treatments of gastrointestinal discomfort, disorders of the nervous system (insomnia, epilepsy, depression, and excessive stress exposure), wound healing, hepatitis, lymphadenitis, and gynecological diseases, among many others [118,119]. The complex applications of these plants were related to the flavonoids, phenolic acids, terpenes, sesquiterpenoid lactones, alkaloids, saponins, etc., contained in them [55,119]. *Artemisia vulgaris* has been frequently used in the treatment of gastrointestinal cancer, and the activity was due to the presence of the flavonoid hispidulin [120]. *Artemisia vulgaris* water extract had very low toxicity (>500 µg/mL) against colorectal cancer (SW-480 cell line), revealing the tissue-specific effect of the plant species [121]. The cytotoxic and anti-proliferative activity of *C. cyanus* towards a large panel of cancer cells was due to the presence of phenolic acids (chlorogenic, caffeic, ferulic, and p-coumaric) and flavonoids (isoquercetin, apigenin, and luteolin) [122].

The methanol extract of *Crepis sancta* aerial parts contains eudesmane-type sesquiterpenoids and different methoxylated flavonols that exhibit antiulcer activity [123]. The high polyphenolic and flavonoid content in the 50% methanol extract of *Chenopodium album* was responsible for its antiarthritic activity. The main effect was correlated with the inhibition of the NF-κB protein [124]. The flavonoid fraction of *C. album* containing mainly rutin and quercetin was found to possess dose-dependent antidiabetic properties [125]. The

water extract of *Sonchus arvensis* had a strong anti-gout effect, expressing anti-inflammatory activity on monosodium urate crystal-induced gout arthritis in Wistar rats [126].

The quenching of reactive oxygen in different species prevents the potential damage of biological macromolecules, such as proteins, lipids, carbohydrates, and nucleic acids, and reduces the risk of causing immunodeficiency syndrome, arteriosclerosis, diabetes, gastric ulcer, cancer, and the aging process [127–130]. Polyphenols, and especially flavonoids, are secondary metabolites that are known to serve various and important roles in the plant organism, such as UV-screening and antioxidant and regulatory functions, thus contributing to the adaptation of the plant organism to the environment and its changes [131–133]. Plants tend to produce more phenolic compounds, such as flavonoids, phenolic acids, and anthocyanins, under various abiotic stresses (cold, drought, heat, etc.) [134]. Hence, the early spring frosts can be considered to be a trigger for the accumulation of secondary metabolites in the wild greens and hence perform their benefits to the consumers. The antioxidant activity of the customary popular *Urtica dioica* was related to the presence of caffeic and p-coumaric acids derivatives, as well as quercetin and kaempferol derivatives in aqueous extracts [135,136]. Similarly, in *Centaurea cyanus* (that is newly recorded as edible plant), the main compounds were found to be chlorogenic, caffeic, ferulic, and p-coumaric acids, as well as iso-quercitrin and coumarin, which presented anti-hemolytic and anti-hypertensive, but not anticancer, activity in vitro of the aqueous extracts [55,137]. On the other hand, *Rumex acetosella* exhibited high radical scavenging activities mainly due to the luteolin and apigenin derivatives [138,139].

The antioxidant activity assessment of *Erodium cicutarium* (L.) L'Hér in water and methanol extracts from four locations in Croatia revealed that the plants from the Plitvice locality showed the highest antioxidant activity according to FRAP, DPPH, and ABTS assays. The observed results were explained by the highest amount of some phenolic acids, such as protocatechuic, 5-O-caffeoylquinic, and p-hydroxybenzoic acid; and the flavonoids hyperoside, narcissin, and quercitrin [140]. We considered that the methanol extract of *Doronicum orientale* Hoffm. could be an alternative source of natural agents in the management of the oxidative stress. The major compounds in this extract were found to be the flavonoids hesperidin, hyperoside, and luteolin-7-glucoside, as well as the phenolic acids chlorogenic, protocatechuic, gallic, and caffeic acids [130]. The flavonoid content in *Ficaria verna* Huds. was much higher in the flowers than the leaves, with the dominant ones being hyperoside (5.0 mg/g dry weight) and quercetin (3.5 mg/g dry weight) [141]. This species could be regarded as a promising natural plant source of antioxidants with a high potential for phytopreparations. The whole herb extract had the highest total phenolics and total flavonoids (mg quercetin/g) content compared to the leaves and flowers extracts only. The total herb extract inhibited approximately 80% of the DPPH radical, which was comparable to the activity of vitamin C and quercetin [142]. The methanol extract of *Ranunculus arvensis* L. might potentially substitute the usage of the synthetic antioxidants butylated hydroxyanisole (BHA) and butylated hydroxytoluene (BHT) and highest total flavonoid content of rutin equivalents dry extract [143]. *Rumex acetosella* exhibited one of the highest radical scavenging activities towards DPPH (31 mg TE/g), correlated with the total flavonoids and phenolic acids content as major compounds that were reported to be the luteolin and apigenin derivatives [111]. The ethyl acetate and acetone extract of *Portulaca oleracea* L. were characterized by the highest flavonoid content (115.49 and 89.65 mg/g quercetin equivalents) compared to the nonpolar extracts, such as n-hexane and chloroform. The same extracts were characterized by the highest antioxidant activity and α -glucosidase inhibitory activity. These results reveal the potential of *P. oleracea* in the management of skin disease, such as age spots and melisma, by controlling the biosynthesis of melanin, as well as in treating diabetes type 2 by reducing the blood sugar [144]. The ethanol extract of *Anchusa officinalis* L. showed the strongest antioxidant and antibacterial activity, which was correlated with the highest concentrations of phenolic compounds (total phenolics of 104.03 mg gallic acid/g extract) compared to other organic extracts [145].

Based on its traditional application for the treatment of pneumonia, the methanol extract of *Anthriscus cerefolium* Hoffm. proved to be effective in inhibiting the biofilm formation of *Staphylococcus aureus* clinical isolate. Its minimum inhibitory concentration (1/2 MIC = 69.88%) was superior to that of streptomycin (1/2 MIC = 55.64%). The extract also inhibited the preformed yeast biofilm of two standard yeast strains (*Candida albicans* and *C. tropicalis*), as well as one clinical isolate, *C. krusei*, at MIC = 5.00 mg/mL, while that of fluconazole was 2.00 mg/mL [146]. Along with its wound-healing potential, *Stellaria media* revealed a strong antibacterial capacity, as well. Due to the differences in the architecture of the bacterial cell wall, the extract had a greater effect towards the Gram-positive (G⁺) *S. aureus* (15.33 mm zone of inhibition) than the Gram-negative (G⁻) *Escherichia coli* (9.66 mm zone of inhibition). Among the individual extract components, vanillic, caffeic, chlorogenic acids, and luteolin possessed antibacterial activity [147]. The ethanol extract of *Anchusa officinalis* showed the strongest antioxidant (53.53% inhibition of DPPH radical) and antibacterial activity, which was correlated with the highest concentrations of phenolic compounds (total phenolics of 104.03 mg gallic acid/g extract) compared to other organic extracts. The ethanol extract has the lowest MIC value, i.e., 3.94 µg/mL, towards *Proteus vulgaris*, *Salmonella enteritidis*, *Enterococcus faecalis*, *Enterococcus faecium*, *Salmonella typhimurium*, and *Candida albicans*. On the other hand, the chloroform extract was the most active (IC₅₀ = 102.28 µg/mL) one towards mouse fibroblast carcinoma [145]. Glucosinolates seem to be characteristic compounds for the family Brassicaceae and identified several species, such as *Alliaria petiolata* (M.Bieb.) Cavara and Grande, *Calepina irregularis* (Asso) Thell, *Cardamine hirsuta* L., *Cardamine pratensis* L., and *Lepidium campestre* (L.) R. Br., and are mainly characterized by their antioxidant, antibacterial, analgesic, anti-inflammatory, and anticancer activity [148,149]. The subcritical CO₂ extract of *Lepidium latifolium* L. containing glucosinolates or their products exhibited antimicrobial properties that were measured through the agar diffusion method, achieving a 20.2, 17.1, 18.3, and 19.3 mm growth inhibition zone for *S. aureus*, *E. coli*, *Kl. pneumoniae*, and *C. albicans*. These values were similar (ranging between 12 and 16 mm) to those of ampicillin at 100 µg/disc [133]. The representatives of *Lepidium* that were found to be consumed in salads in Bulgaria in the past were also found to be a good source of polyunsaturated fatty acids, together with soy and other pulses [150]. Their consumption, along with the consumption of other cruciferous vegetables, has been associated with a reduced risk of cancer (including in organs such as the lungs, stomach, breast, prostate, pancreas, colon, and rectum), and this effect has been assigned to the isothiocyanate content [151]. The in vitro anti-glioblastoma activity of methanol extract of *Anthriscus cerefolium* against the A172 glioblastoma cell line (IC₅₀ = 765.21 µg/mL) was devoted to the presence of 32 phenolic compounds mainly presented by hydroxybenzoic and ferulic acids derivatives, as well as quercetin and luteolin glucosides. The extract has shown to be selective, having no toxicity (IC₅₀ = 800 µg/mL) towards the control human gingival fibroblasts cells HGF-1 [152]. The ethyl acetate and acetone extract of *Portulaca oleracea* were characterized by the highest flavonoid content (115.49 and 89.65 mg/g quercetin equivalents) compared to the nonpolar extracts, such as n-hexane and chloroform. Along with the presence of essential oils and phenolic compounds, *Geranium lucidum* L. contains alkaloids (palmatine, columbamine, pseudo columbamine, and geraniin), which determine its potential similarly to other *Geranium* species to possess anticancer activity [153]. Essential oils are another important substance found in plants. A major part of the oil from *Artemisia vulgaris* extracted from the aerial parts was constituted by monoterpenoids (72%) and sesquiterpenoids (26%). Among the major volatile compounds identified were 1.8-cineole (28.9%), sabinene (13.7%), camphor (13.0%), camphene (9.1%), β-caryophyllene oxide (6.5%), α-, and β-thujone (13.5%). The essential oil is characterized by promising antioxidant, antimicrobial, and anticancer properties. The essential oil of *A. vulgaris*, extracted from the aerial parts and mainly composed of 1.8-cineole and β-thujone, exhibited antibacterial activity against *E. coli*, *S. enteritidis*, *P. aeruginosa*, *K. pneumoniae*, and *S. aureus*, as well as antifungal against *C. albicans* and *Aspergillus niger*. On the other hand, the essential oil extracted from

the roots did not have any antimicrobial activity due to the low level of 1.8-cineole and the lack of β -thujone in the roots. *Artemisia vulgaris* might be a promising source of new anticancer agents, since it induced apoptosis in the HL-60 leukemic cell line. The observed apoptosis was mediated by caspase-dependent pathways, involving caspase-3, -9, and -8, which were initiated by Bcl-2/Bax/Bid-dependent loss of mitochondrial membrane potential, leading to the release of cytochrome c to the cytoplasm to activate the caspase cascade [154]. Three acyclic hydrocarbon monoterpeneoids β -myrcene, cis- β -ocimene (its trans isomer), limonene, α -terpinene, p-cymene, and Δ^3 carene were identified in the essential oil of the leaves of *Chenopodium polyspermum* L. These compounds were mainly responsible for its antimicrobial and especially antifungal activity towards *C. albicans* and *A. niger* [155]. Hops essential oil exhibited potential anticancer, anti-inflammatory, analgesic, and sedative effect on human health. The anticancer activity was attributed to its compounds, such as β -caryophyllene and β -caryophyllene oxide. These compounds have the potential to inhibit the growth and proliferation of various cancer cells, including those of colon, pancreas, breast, cervix, and prostate [156].

The bioactivity-guided investigation of the methanol extract of *Crepis sancta* (L.) Bornm. aerial parts revealed that the extract was very effective in the treatment of ethanol-induced gastric ulcer in male albino rats. Contributing to this activity are the seven isolated methoxyflavonoids, namely kumatakenin, penduletin, pachypodol, chrysoisplentin, jaceidin, casticin, and 3.5.7-tri-*O*-methyl-6-methoxykaempferol [123]. An aqueous extract of *Centaurea cyanus* L. presented anti-hemolytic and anti-hypertensive activity in vitro. The main compounds present in the extract were chlorogenic, caffeic, ferulic, and p-coumaric acids, as well as isoquercitrin and coumarin. However, the extract showed no activity against several cancer cells, such as lung adenocarcinoma (cell line A549), colorectal adenocarcinoma (Caco-2), and human hepatoma carcinoma (HepG2) cells [157]. The 70% ethanol extract of *Stellaria media* (L.) Vill., which is rich in flavonoids, revealed promising wound-healing properties investigated on normal human dermal fibroblasts (NHDFs) [147,158]. The saponin-rich fraction of *Chenopodium bonus-henricus* L. possessed better hepatoprotective activity than the flavonoid-rich fraction. The saponin fraction showed in vitro hepatoprotective and antioxidant activities comparable to those of flavonoid complex silymarin (60 μ g/mL) in a model of metabolic bioactivation induced by CCl₄. The fraction, compared to silymarin, significantly reduced the cellular damage caused by CCl₄ in rat hepatocytes, preserved cell viability and glutathione level, decreased lactate dehydrogenase leakage, and reduced lipid damage [159].

3.4. Toxicity and Community Awareness

Many of the spring greens consumed by Bulgarian communities are reported to contain anti-nutritive compounds (e.g., alkaloids, furanocoumarins, saponins, tannins, oxalates, etc.) that might induce acute toxic effects or affect humans and/or animals after prolonged consumption. Insufficient knowledge of plants and their bioactive composition poses dangers for collectors and buyers at farmers' markets [97,99,160–162]. According to the Toxic Plants–Phytotoxins Database, for 59% of the studied taxa, there is no evidence of toxicity in humans, and only 3% fall into the group of strong toxic plants [163].

Pyrrrolizidine alkaloids that occur in Boraginaceae are not toxic per se but require bioactivation to the toxic dehydropyrrrolizidine alkaloids (so called “pyrroles”), which occurs primarily in the liver [164–166]. Selection through tasting was applied to the representatives of this family which are used in a juvenile state, and they are consumed only after thermal processing, which reduces the effect of the hepatotoxic unsaturated pyrrrolizidine alkaloids. The tasting during the gathering, the stewing of the greens to prepare the pastry filling, and the subsequent baking of the dish and the small amounts of the two taxa (*Symphytum tuberosum* L. and *Anchusa* sp.) used in the green mixtures minimize the risk from their consumption.

The representatives of Ranunculaceae (*Ficaria verna* Huds. and *Ranunculus arvensis* L. [143]) contain the toxic glycoside ranunculin, which, in the case of dermal contact, is converted

to protoanemonin, the toxicity of which may cause third-degree skin burns, leading to dermal–epidermal separation and the formation of bullous lesions. The clinical condition is called phytodermatitis [167]. *Ranunculus arvensis* is traditionally used in the Far East to treat arthritis, asthma, gout, high fever, and psoriasis, but it is highly allergenic in spring during the flowering period [143]. The two species participate in small quantities in the pastry fillings both in South and SW Bulgaria and are collected only after the tasting of the herbage and discarding all bitter and/or flowering plants. Prior to the consumption, the green mass that includes the two species is first stewed, and afterwards the whole dish is baked in the oven [143]. Hence, the negative effect is abolished, as proven by [168] and other authors [168].

The furanocoumarins in many plants may cause a phototoxic reaction when they come in contact with skin that is exposed to UV-A light. This is due to their ability to react with nucleobases in DNA under the influence of UV-A radiation, resulting in crosslinks in DNA [169]. Coumarins and furanocoumarins serve as potent defense compounds in Apiaceae, but, in humans, they can cause mutations or even cancer [170]. *Anthriscus cerefolium* was listed together with *Foeniculum vulgare* Mill. and other species from the family to contain furanocoumarins and other toxic compounds [171]. However, the negative impact could be alleviated when applied in mixtures [172]. Additionally, recent research on herba *Anthrisci cerefolii* showed a decreased proliferation rate of glioblastoma cells while being non-toxic to the control cell line [152]. The cytotoxic effect of *Anthriscus* extracts is due to podophyllotoxin-related lignans, which are currently of interest due to the high availability of these ruderal species [173,174].

Tropane alkaloids are commonly used as anti-colic and spasmolytic drugs (scopolamine) in both digestive- and urinary-tract spastic conditions. Moreover, atropine is commonly used in ophthalmological eyedrops to enlarge pupils, paralyze the accommodation reflex, and enable the ophthalmic examination. Tropane alkaloids or their derivatives (containing tropane core) are potential anticancer agents with potential beneficial effect during the treatment of human multiple myeloma (RPMI 8226), lung carcinoma (A549), breast adenocarcinoma (MDA-MB-231), and mouse skin melanoma (B16-F10) cell lines [175]. On the other hand, the consumption of *Convolvulus arvensis* L, which contains tropane alkaloids, may cause diarrhea, colic, gastrointestinal ulceration, and intestinal thickening and fibrosis, as shown for the roots of this species, mainly when ingested by animals [143,167,176–179]. However, when collected as food in early spring, only young leaves are selected, and afterwards, they undergo thermal treatment as parts of the pastry filling that should alleviate the risk (see above).

While different approaches, e.g., cooking or steeping in water before preparation, to eliminate or at least reduce the amount of toxic compounds are well documented in the Mediterranean area, Bulgarian ethnographical sources claim that “poisonous plants” were taboo in the traditional cuisine [25,39,180]. This could be related also to the rare use of cooked or blanched vegetables as salads and/or snacks in Bulgaria that was confirmed during our field studies where only blanched branch shoots of *Pistacia terebinthus* L. were presented in a salad form. About 41% of the currently listed taxa are reported to be non-toxic, and, in many cases, the reported toxicity for certain species was related to parts which were not consumed, or it is related to alkaloids, saponins, or glucosides that are in relatively small amounts in the seedlings or shoots during the collection time. Another case was the use of the leaves of *Phytolacca americana* L. for the preparation of *sarmi* (green leaves stuffed with a mixture of rice, chopped allium, and spices). The vegetative parts of the plant contain saponins (mainly phytolaccatoxin and phytolaccagenin) in greater amounts than in the fruits and was known to cause stomach and intestinal discomfort if not properly cooked [181]. The first application of this invasive weed was related to the use of its pokeberries for fortification of the wine color and is also reflected in the Bulgarian name of the plant *amerikanski winoboj* (American wine-colorant) [80]. Still, when the participants from the local community in Belasitsa Mt. talked about the use of the leaves of *P. americana* for *sarmi* preparation, they used another colloquial name, *butima* (Figure 7). The leaves

are used either fresh in spring—soaked in hot water boiled after they are stuffed with the filling—or dried and stored for later on. In the latter case, the leaves are again soaked in hot water and then cooked until ready.



Figure 7. *Phytolacca americana* L. is used diversely throughout the season—leaves are used for preparation of *sarmi* (left) and the fruits (right) for color fortification of wines and as ink in the past (Gabrene village, Blagoevgrad Province).

Our respondents from the Central Balkan Mts. used a similar approach when preparing leaves from *Arum maculatum*. Although they were not specifically aware of the toxic effects of the plant, they applied combined drying and continuous boiling of the leaves before consumption to improve their taste. In fact, this practice can be regarded also as traditional knowledge inherited from previous generations that resulted in the reduction of the toxicity of the leaves. A similar approach was found to be effective in the elimination of the oxalates of *Arum palaestinum* Boiss. traditionally consumed in Palestine, thus reducing possible injuries and inflammation of skin and mucosa, and negative impacts on the stomach, intestines, and kidneys [182]. However, the same authors revealed that boiling significantly reduces the total amount of phenolics; hence, the benefits of its consumption are diminished.

While the awareness of common poisonous plants could be regarded as adequate, at least in rural communities under study, anti-nutritional compounds such as oxalates and phytates that are abundant in the popular *Rumex acetosa*, *Urtica dioica*, and some *Chenopodium* species should be considered an important issue, as the first forms insoluble calcium oxalate salts found in more than 50% of patients with renal lithiasis in Bulgaria, and the second reduces the number of the macro- and micronutrients in the food [163,183–186]. Additionally, the gathering of those plants along roadsides in and near arable fields treated with pesticides should be avoided so as to reduce incidental intoxications with other environmental pollutants [97].

When the toxic effects of some early spring greens are discussed, we should bear in mind several facts that stem from the local traditional knowledge and practical needs of our respondents. Firstly, Asteraceae and Amaranthaceae families participate in the green mixtures not only by having the highest number of species but also because they prevail in quantities, as their leaves and young shoots have big biomass. Secondly, the species from the two families that are consumed are not reported to contain toxic compounds. Thirdly, the plant species that contain toxic compounds participate more rarely and in lesser amounts in the green mixtures on the one hand, and, on the other, the mode of preparation always involves thermal treatment, which is reported to reduce the harmful effects.

4. Conclusions

Foraging for spring vegetables is an important part of the traditional ecological knowledge that involves not only various information on local flora but also on biologically active substances, including toxic ones. The preservation of those traditions, together with the promotion of agroecological practices, and hence the reduction of the use of pesticides, could

contribute to food security in the current turbulent socioeconomic situation and ensure the supply of valuable biologically active compounds at minimal or no cost. However, the diminishing of the number of the keepers of such knowledge hampers its intergenerational transfer. Therefore, it is crucial to promote and (re)integrate this knowledge into formal and informal education, especially among children and vulnerable groups (people living in poverty, refugees, homeless, etc.). The inclusion and valorization of this traditional knowledge as a part of local tourism services and the promotion of sustainable practices for nature conservation could encourage the development of new businesses in the rural areas and ensure more diverse local food, as well.

Understanding the traditional food practices related to foraging early spring greens can trigger further research on the chemical compounds in plants, i.e., in different stages of ontogenesis or under different types of processing. The increase of the knowledge about the benefits of plant food can also contribute to the increase of share of the plant-based food in the diet of contemporary people.

Supplementary Materials: The following supporting information can be downloaded at <https://www.mdpi.com/article/10.3390/d15030435/s1>. Supplementary Table S1. Ruderals used as traditional spring food in Bulgaria. References [187–317] are cited in Supplementary Materials.

Author Contributions: Conceptualization, D.D., T.I. and E.K.; methodology, D.D.; formal analysis, T.I., A.M., M.C. and Y.B.; investigation, D.D., T.I., Y.B., A.M., M.C. and M.G.; writing—original draft preparation, T.I. and A.M.; writing—review and editing, D.D., T.I., M.C., Y.B., A.M., E.K. and M.G.; funding acquisition D.D. All authors have read and agreed to the published version of the manuscript.

Funding: This study was carried out in the framework of the National Science Program “Environmental Protection and Reduction of Risks of Adverse Events and Natural Disasters”, approved by the Resolution of the Council of Ministers No. 577/17.08.2018 and supported by the Ministry of Education and Science (MES) of Bulgaria (Agreement No. D01-230/6.12.2018). Part of the field studies were supported under the project DN10/1/2016 “The Garden: Site of Biocultural Diversity and Interdisciplinary Junction”, funded by the National Science Fund.

Institutional Review Board Statement: The guidelines prescribed in the Code of Ethics of the International Society of Ethnobiology were followed during the field study, and their compliance was confirmed by the Scientific Council of the Institute of Biodiversity and Ecosystem Research, Bulgarian Academy of Sciences, acting as independent institutional Ethics Board (Decision No. 6/21/05/21).

Data Availability Statement: Not applicable.

Acknowledgments: The authors are grateful to all participants for the shared knowledge and kind cooperation during the field study. The skillful cooperation of Sofia Kostdinova-Ilkova, National Park Belasitsa Directorate, is gratefully acknowledged. Some of the older culinary books were made available through the valuable collection of Marin Lesev, whose help is greatly appreciated.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Berry, E.M.; Dernini, S.; Burlingame, B.; Meybeck, A.; Conforti, P. Food security and sustainability: Can one exist without the other? *Public Health Nutr.* **2015**, *18*, 2293–2302. [[CrossRef](#)] [[PubMed](#)]
2. Johns, T.; Eyzaguirre, P.B. Linking biodiversity, diet and health in policy and practice. *Proc. Nutr. Soc.* **2006**, *65*, 182–189. [[CrossRef](#)]
3. Burlingame, B.; Dernini, S. Sustainable diets: The Mediterranean diet as an example. *Public Health Nutr.* **2011**, *14*, 2285–2287. [[CrossRef](#)]
4. Ficiyan, A.; Loos, J.; Sievers-Glotzbach, S.; Tschardtke, T. More than Yield: Ecosystem Services of Traditional versus Modern Crop Varieties Revisited. *Sustainability* **2018**, *10*, 2834. [[CrossRef](#)]
5. Truzzi, M.L.; Puviani, M.B.; Tripodi, A.; Toni, S.; Farinetti, A.; Nasi, M.; Mattioli, A.V. Mediterranean Diet as a model of sustainable, resilient and healthy diet. *Prog. Nutr.* **2020**, *22*, 388–394. [[CrossRef](#)]
6. Graça, J.; Godinho, C.A.; Truninger, M. Reducing meat consumption and following plant-based diets: Current evidence and future directions to inform integrated transitions. *Trends Food Sci. Technol.* **2019**, *91*, 380–390. [[CrossRef](#)]
7. Cleveland, D.A.; Gee, Q. Plant-Based Diets for Mitigating Climate Change. In *Vegetarian and Plant-Based Diets in Health and Disease Prevention*; Mariotti, F., Ed.; Academic Press: Cambridge, MA, USA, 2017; pp. 135–156. ISBN 9780128039694.

8. Giampieri, F.; Mazzoni, L.; Cianciosi, D.; Alvarez-Suarez, J.M.; Regolo, L.; Sánchez-González, C.; Capocasa, F.; Xiao, J.; Mezzetti, B.; Battino, M. Organic vs conventional plant-based foods: A review. *Food Chem.* **2022**, *383*, 132352. [[CrossRef](#)] [[PubMed](#)]
9. van Vliet, S.; Kronberg, S.L.; Provenza, F.D. Plant-Based Meats, Human Health, and Climate Change. *Front. Sustain. Food Syst.* **2020**, *4*, 128. [[CrossRef](#)]
10. Samtiya, M.; Aluko, R.E.; Dhewa, T.; Moreno-Rojas, J. Potential Health Benefits of Plant Food-Derived Bioactive Components: An Overview. *Foods* **2021**, *10*, 839. [[CrossRef](#)]
11. Shen, J.; Shan, J.; Zhong, L.; Liang, B.; Zhang, D.; Li, M.; Tang, H. Dietary Phytochemicals that Can Extend Longevity by Regulation of Metabolism. *Plant Foods Hum. Nutr.* **2022**, *77*, 12–19. [[CrossRef](#)]
12. Lachat, C.; Raneri, J.E.; Smith, K.W.; Kolsteren, P.; Van Damme, P.; Verzelen, K.; Penafiel, D.; Vanhove, W.; Kennedy, G.; Hunter, D.; et al. Dietary species richness as a measure of food biodiversity and nutritional quality of diets. *Proc. Natl. Acad. Sci. USA* **2017**, *115*, 127–132. [[CrossRef](#)]
13. Łuczaj, Ł.; Pieroni, A.; Tardío, J.; Pardo-De-Santayana, M.; Sõukand, R.; Svanberg, I.; Kalle, R. Wild food plant use in 21st century Europe: The disappearance of old traditions and the search for new cuisines involving wild edibles. *Acta Soc. Bot. Pol.* **2012**, *81*, 359–370. [[CrossRef](#)]
14. Tardío, J.; Pardo-De-Santayana, M.; Morales, R. Ethnobotanical review of wild edible plants in Spain. *Bot. J. Linn. Soc.* **2006**, *152*, 27–71. [[CrossRef](#)]
15. Łuczaj, Ł. Changes in the utilization of wild green vegetables in Poland since the 19th century: A comparison of four ethnobotanical surveys. *J. Ethnopharmacol.* **2010**, *128*, 395–404. [[CrossRef](#)] [[PubMed](#)]
16. Leonti, M.; Nebel, S.; Rivera, D.; Heinrich, M. Wild Gathered Food Plants in the European Mediterranean: A Comparative Analysis | SpringerLink. *Econ. Bot.* **2006**, *60*, 130–142. [[CrossRef](#)]
17. Hadjichambis, A.C.; Paraskeva-Hadjichambi, D.; Della, A.; Elena Giusti, M.; De Pasquale, C.; Lenzarini, C.; Censorii, E.; Reyes Gonzales-Tejero, M.; Patricia Sanchez-Rojas, C.; Ramiro-Gutierrez, J.M.; et al. Wild and semi-domesticated food plant consumption in seven circum-Mediterranean areas. *Int. J. Food Sci. Nutr.* **2009**, *59*, 383–414. [[CrossRef](#)]
18. Ebert, A.W. The Role of Vegetable Genetic Resources in Nutrition Security and Vegetable Breeding. *Plants* **2020**, *9*, 736. [[CrossRef](#)]
19. Geraci, A.; Amato, F.; Di Noto, G.; Bazan, G.; Schicchi, R. The wild taxa utilized as vegetables in Sicily (Italy): A traditional component of the Mediterranean diet. *J. Ethnobiol. Ethnomedicine* **2018**, *14*, 14. [[CrossRef](#)]
20. Biscotti, N.; Pieroni, A. The hidden Mediterranean diet: Wild vegetables traditionally gathered and consumed in the Gargano area, Apulia, SE Italy. *Acta Soc. Bot. Pol.* **2015**, *84*, 327–338. [[CrossRef](#)]
21. Kalle, R.; Sõukand, R. Historical ethnobotanical review of wild edible plants of Estonia (1770s–1960s). *Acta Soc. Bot. Pol.* **2012**, *81*, 271–281. [[CrossRef](#)]
22. Pieroni, A.; Sulaiman, N.; Sõukand, R. *Chorta* (Wild Greens) in Central Crete: The Bio-Cultural Heritage of a Hidden and Resilient Ingredient of the Mediterranean Diet. *Biology* **2022**, *11*, 673. [[CrossRef](#)]
23. Pieroni, A.; Cattero, V. Wild vegetables do not lie: Comparative gastronomic ethnobotany and ethno-linguistics on the Greek traces of the Mediterranean Diet of southeastern Italy. *Acta Bot. Bras.* **2019**, *33*, 198–211. [[CrossRef](#)]
24. Motti, R. Wild Plants Used as Herbs and Spices in Italy: An Ethnobotanical Review. *Plants* **2021**, *10*, 563. [[CrossRef](#)] [[PubMed](#)]
25. Guarrera, P.M.; Savo, V. Wild food plants used in traditional vegetable mixtures in Italy. *J. Ethnopharmacol.* **2016**, *185*, 202–234. [[CrossRef](#)] [[PubMed](#)]
26. Pieroni, A.; Quave, C.; Giusti, M.E.; Papp, N. “We Are Italians!”: The Hybrid Ethnobotany of a Venetian Diaspora in Eastern Romania. *Hum. Ecol.* **2012**, *40*, 435–451. [[CrossRef](#)]
27. Pasta, S.C.; La Rosa, A.; Garfi, G.; Marcenò, C.; Gristina, A.S.; Carimi, F.; Guarino, R. An Updated Checklist of the Sicilian Native Edible Plants: Preserving the Traditional Ecological Knowledge of Century-Old Agro-Pastoral Landscapes. *Front. Plant Sci.* **2020**, *11*, 388. [[CrossRef](#)]
28. Vitasović-Kosić, I.; Hodak, A.; Łuczaj, Ł.; Marić, M.; Juračak, J. Traditional Ethnobotanical Knowledge of the Central Lika Region (Continental Croatia)—First Record of Edible Use of Fungus *Taphrina pruni*. *Plants* **2022**, *11*, 3133. [[CrossRef](#)]
29. Corrêa, R.C.G.; Di Gioia, F.; Ferreira, I.C.F.R.; Petropoulos, S.A. Wild Greens Used in the Mediterranean Diet. In *The Mediter-ranean Diet*; Preedy, V., Watson, R., Eds.; Academic Press: Cambridge, MA, USA, 2020; pp. 209–228.
30. Chatzopoulou, E.; Carocho, M.; Gioia, F.; Petropoulos, S. The Beneficial Health Effects of Vegetables and Wild Edible Greens: The Case of the Mediterranean Diet and Its Sustainability. *Appl. Sci.* **2020**, *10*, 9144. [[CrossRef](#)]
31. Ozturk, H.I.; Nas, H.; Ekinçi, M.; Turan, M.; Ercisli, S.; Narmanlioglu, H.K.; Yildirim, E.; Assouguem, A.; Almeer, R.; Sayed, A.A.; et al. Antioxidant Activity, Phenolic Composition, and Hormone Content of Wild Edible Vegetables. *Horticulturae* **2022**, *8*, 427. [[CrossRef](#)]
32. Łuczaj, Ł.; Köhler, P.; Piroznicow, E.; Graniszewska, M.; Pieroni, A.; Gervasi, T. Wild Edible Plants of Belarus: From Ros-tafiński’s Questionnaire of 1883 to the Present. *J. Ethnobiol. Ethnomed.* **2013**, *9*, 21. [[CrossRef](#)]
33. Gras, A.; Garnatje, T.; Marín, J.; Parada, M.; Sala, E.; Talavera, M.; Vallès, J. The Power of Wild Plants in Feeding Humanity: A Meta-Analytic Ethnobotanical Approach in the Catalan Linguistic Area. *Foods* **2020**, *10*, 61. [[CrossRef](#)]
34. Sardeshpande, M.; Shackleton, C. Urban foraging: Land management policy, perspectives, and potential. *PLoS ONE* **2020**, *15*, e0230693. [[CrossRef](#)] [[PubMed](#)]
35. Kolosova, V.; Belichenko, O.; Rodionova, A.; Melnikov, D.; Sõukand, R. Foraging in Boreal Forest: Wild Food Plants of the Republic of Karelia, NW Russia. *Foods* **2020**, *9*, 1015. [[CrossRef](#)]

36. Pieroni, A.; Hovsepyan, R.; Manduzai, A.K.; Söukand, R. Wild food plants traditionally gathered in central Armenia: Archaic ingredients or future sustainable foods? *Environ. Dev. Sustain.* **2020**, *23*, 2358–2381. [[CrossRef](#)]
37. Stryamets, N.; Mattalia, G.; Pieroni, A.; Khomyn, I.; Söukand, R. Dining Tables Divided by a Border: The Effect of Socio-Political Scenarios on Local Ecological Knowledge of Romanians Living in Ukrainian and Romanian Bukovina. *Foods* **2021**, *10*, 126. [[CrossRef](#)]
38. Schunko, C.; Brandner, A. Urban nature at the fingertips: Investigating wild food foraging to enable nature interactions of urban dwellers. *AMBIO* **2022**, *51*, 1168–1178. [[CrossRef](#)]
39. Markova, M. *Food and Nutrition: Between Nature and Culture*; Prof. Marin Drinov Academic Publishing House: Sofia, Bulgaria, 2011; ISBN 978-954-322-462-3.
40. Shishkov, S. What Is the Food of the Pomaks. *Ethnographic Notes. Rhodope Prog.* **1908**, *6*, 1–13.
41. Willett, W.C.; Sacks, F.; Trichopoulou, A.; Drescher, G.; Ferro-Luzzi, A.; Helsing, E.; Trichopoulos, D. Mediterranean diet pyramid: A cultural model for healthy eating. *Am. J. Clin. Nutr.* **1995**, *61* (Suppl. 6), 1402S–1406S. [[CrossRef](#)] [[PubMed](#)]
42. Estruch, R.; Ros, E.; Salas-Salvadó, J.; Covas, M.-I.; Corella, D.; Arós, F.; Gómez-Gracia, E.; Ruiz-Gutiérrez, V.; Fiol, M.; Lapetra, J.; et al. Primary Prevention of Cardiovascular Disease with a Mediterranean Diet Supplemented with Extra-Virgin Olive Oil or Nuts. *N. Engl. J. Med.* **2018**, *378*, e34. [[CrossRef](#)]
43. García-Conesa, M.-T.; Philippou, E.; Pafilas, C.; Massaro, M.; Quarta, S.; Andrade, V.; Jorge, R.; Chervenkov, M.; Ivanova, T.; Dimitrova, D.; et al. Exploring the Validity of the 14-Item Mediterranean Diet Adherence Screener (MEDAS): A Cross-National Study in Seven European Countries around the Mediterranean Region. *Nutrients* **2020**, *12*, 2960. [[CrossRef](#)]
44. Quarta, S.; Massaro, M.; Chervenkov, M.; Ivanova, T.; Dimitrova, D.; Jorge, R.; Andrade, V.; Philippou, E.; Zisimou, C.; Maksimova, V.; et al. Persistent Moderate-to-Weak Mediterranean Diet Adherence and Low Scoring for Plant-Based Foods across Several Southern European Countries: Are We Overlooking the Mediterranean Diet Recommendations? *Nutrients* **2021**, *13*, 1432. [[CrossRef](#)] [[PubMed](#)]
45. Kitanov, B. Materials on Utilization of Wild Plants in National Economy. *Bull. Bot. Inst. Bulg. Acad. Sci.* **1953**, *3*, 257–260.
46. Ivanova, T.; Bosseva, Y.; Ganeva-Raycheva, V.; Dimitrova, D. Ethnobotanical Knowledge on Edible Plants Used in Zelnik Pastries from Haskovo Province (Southeast Bulgaria). *Phytol. Balc.* **2018**, *24*, 389–395.
47. Nedelcheva, A. An ethnobotanical study of wild edible plants in Bulgaria. *EurAsian J. Biosci.* **2013**, *7*, 77–94. [[CrossRef](#)]
48. Pieroni, A.; Nedelcheva, A.; Dogan, Y. Local knowledge of medicinal plants and wild food plants among Tatars and Romanians in Dobruja (South-East Romania). *Genet. Resour. Crop. Evol.* **2014**, *62*, 605–620. [[CrossRef](#)]
49. Kreuz, A.; Marinova, E. Archaeobotanical evidence of crop growing and diet within the areas of the Karanovo and the Linear Pottery Cultures: A quantitative and qualitative approach. *Veg. Hist. Archaeobotany* **2017**, *26*, 639–657. [[CrossRef](#)]
50. Hrisrova, I.; Atanassova, J.; Marinova, E. Plant economy and vegetation of the Iron Age in Bulgaria: Archaeobotanical evidence from pit deposits. *Archaeol. Anthr. Sci.* **2017**, *9*, 1481–1494. [[CrossRef](#)]
51. Kathe, W.; Honnef, S.; Heym, A. *Medicinal and Aromatic Plants in Albania, Bosnia-Herzegovina, Bulgaria, Croatia and Romania*; Bundesamt für Naturschutz: Bonn, Germany, 2003.
52. Evstatieva, L.; Hardalova, R.; Stoyanova, K. Medicinal Plants in Bulgaria: Diversity, Legislation, Conservation and Trade. *Phytol. Balc.* **2007**, *13*, 415–427.
53. Kozuharova, E.; Getov, I.N. Herbal Medicinal Products Registrations in the EU and the Implications for the Bulgarian Medicinal Plant Resources. *Comptes Rendus L'Académie Bulg. Des Sci.* **2012**, *65*, 1527–1534.
54. Nebel, S.; Pieroni, A.; Heinrich, M. Ta chörta: Wild edible greens used in the Graecanic area in Calabria, Southern Italy. *Appetite* **2006**, *47*, 333–342. [[CrossRef](#)]
55. Mikropoulou, E.V.; Vougiannopoulou, K.; Kalpoutzakis, E.; Sklirou, A.D.; Skaperda, Z.; Houriet, J.; Wolfender, J.-L.; Trougakos, I.P.; Kouretas, D.; Halabalaki, M.; et al. Phytochemical Composition of the Decoctions of Greek Edible Greens (Chörta) and Evaluation of Antioxidant and Cytotoxic Properties. *Molecules* **2018**, *23*, 1541. [[CrossRef](#)]
56. Redžić, S.; Barudanović, S.; Pilipović, S. Wild Mushrooms and Lichens Used as Human Food for Survival in War Conditions; Podrinje—Zepa Region (Bosnia and Herzegovina, W. Balkan) on JSTOR. *Hum. Ecol. Rev.* **2010**, *17*, 175–187.
57. Łuczaj, Ł.; Zovkokončić, M.; Miličević, T.; Dolina, K.; Pandža, M. Wild vegetable mixes sold in the markets of Dalmatia (southern Croatia). *J. Ethnobiol. Ethnomed.* **2013**, *9*, 2. [[CrossRef](#)]
58. Pieroni, A.; Sulaiman, N.; Polesny, Z.; Söukand, R. From *şxex* to *Chorta*: The Adaptation of Maronite Foraging Customs to the Greek Ones in Kormakitis, Northern Cyprus. *Plants* **2022**, *11*, 2693. [[CrossRef](#)] [[PubMed](#)]
59. Dogan, Y.; Nedelcheva, A. Wild Plants from Open Markets on Both Sides of the Bulgarian-Turkish Border. *Indian J. Tradit. Knowl.* **2015**, *14*, 351–358.
60. Badalamenti, N.; Sottile, F.; Bruno, M. Ethnobotany, Phytochemistry, Biological, and Nutritional Properties of Genus *Crepis*—A Review. *Plants* **2022**, *11*, 519. [[CrossRef](#)]
61. Torrens, F.; Castellano, G. Ethnobotanical Studies of Medicinal Plants: Underutilized Wild Edible Plants, Food, and Medicine. In *Molecular Chemistry and Biomolecular Engineering*; Pogliani, L., Torrens, F., Haghi, A.K., Eds.; Apple Academic Press: Palm Bay, FL, USA, 2019; pp. 63–71. ISBN 9780429060649.
62. Patel, B.; Sharma, S.; Nair, N.; Majeed, J.; Goyal, R.K.; Dhobi, M. Therapeutic opportunities of edible antiviral plants for COVID-19. *Mol. Cell. Biochem.* **2021**, *476*, 2345–2364. [[CrossRef](#)]

63. Süntar, I. Importance of ethnopharmacological studies in drug discovery: Role of medicinal plants. *Phytochem. Rev.* **2019**, *19*, 1199–1209. [CrossRef]
64. Holzner, W. Concepts, Categories and Characteristics of Weeds. In *Biology and Ecology of Weeds*; Holzner, W., Numata, M., Eds.; Springer: The Hague, The Netherlands, 1982; pp. 3–20.
65. Fischer, L.K.; Brinkmeyer, D.; Karle, S.J.; Cremer, K.; Huttner, E.; Seebauer, M.; Nowikow, U.; Schütze, B.; Voigt, P.; Völker, S.; et al. Biodiverse edible schools: Linking healthy food, school gardens and local urban biodiversity. *Urban For. Urban Green.* **2019**, *40*, 35–43. [CrossRef]
66. Rupperecht, C.D.; Byrne, J.A.; Garden, J.G.; Hero, J.-M. Informal urban green space: A trilingual systematic review of its role for biodiversity and trends in the literature. *Urban For. Urban Green.* **2015**, *14*, 883–908. [CrossRef]
67. Toffolo, C.; Gentili, R.; Banfi, E.; Montagnani, C.; Caronni, S.; Citterio, S.; Galasso, G. Urban plant assemblages by land use type in Milan: Floristic, ecological and functional diversities and refugium role of railway areas. *Urban For. Urban Green.* **2021**, *62*, 127175. [CrossRef]
68. Bocheva, L.; Trifonova, L.; Marinova, T.; Malcheva, K. Climate Profile of Bulgaria in the Period 1988–2016 and Brief Climatic Assessment of 2017. *Bulg. J. Meteorol. Hydrol.* **2017**, *22*, 2–15.
69. Petrova, A.; Vladimirov, V. Recent Progress in Floristic and Taxonomic Studies in Bulgaria. *Bot. Serb.* **2018**, *42*, 35–69. [CrossRef]
70. European Environment Agency. *The European Environment—State and Outlook 2020*; Publications Office of the European Union: Luxembourg City, Luxembourg, 2019; ISBN 978-92-9480-090-9.
71. Dancheva, A.; Dimitrova, D.; Jordanova, E.; Cheshmedzhieva, G.; Nikolova, G.; Gergova, M.; Kolev, M.; Panagonova, S.; Petkova, R.; Filipovich, S.; et al. *Statistical Reference Book 2022*; National Statistical Institute: Sofia, Bulgaria, 2022.
72. International Society of Ethnobiology ISE Code of Ethics. Available online: <http://ethnobiology.net/code-of-ethics/> (accessed on 21 June 2021).
73. Delipavlov, D.; Cheshmedzhiev, I.; Popova, M.; Terziyski, D.; Kovachev, I. *Handbook of Bulgarian Vascular Flora*; Delipavlov, D., Cheshmedzhiev, I., Eds.; Academic Publishing House of Agricultural University-Plovdiv: Plovdiv, Bulgaria, 2003.
74. Royal Botanic Gardens Kew Garden; Missouri Botanical Garden The Plantlist Database. Available online: <http://www.theplantlist.org> (accessed on 26 May 2022).
75. Söukand, R.; Pieroni, A.; Biró, M.; Dénes, A.; Dogan, Y.; Hajdari, A.; Kalle, R.; Reade, B.; Mustafa, B.; Nedelcheva, A.; et al. An ethnobotanical perspective on traditional fermented plant foods and beverages in Eastern Europe. *J. Ethnopharmacol.* **2015**, *170*, 284–296. [CrossRef] [PubMed]
76. Boycheva, P.; Marinova, V. Ethnobotanical Study of Medicinal Plants for Culinary Purposes in the North Black Sea Coast, Varna Region (Bulgaria). *Annu. L'université Sofia "St. Kliment Ohridski" Fac. Biol.* **2018**, *103*, 172–182.
77. Georgiev, M. *Bulgarian Folk Medicine*, 2nd ed.; Vasileva, M., Georgiev, M., Georgieva, I., Penchev, V., Popov, R., Simeonova, G., Troeva, E., Tsaneva, E., Eds.; Prof. Marin Drinov Academic Publishing House: Sofia, Bulgaria, 2013; ISBN 978-954-322-542-8.
78. Stojanov, N.; Kitanov, B. *Wild Useful Plants in Bulgaria*; Bulgarian Academy of Sciences: Sofia, Bulgaria, 1960.
79. Ivanova, T.; Bosseva, Y.; Chervenkov, M.; Dimitrova, D. Enough to Feed Ourselves!—Food Plants in Bulgarian Rural Home Gardens. *Plants* **2021**, *10*, 2520. [CrossRef] [PubMed]
80. Petrova, A.; Vladimirov, V.; Georgiev, V. *Invasive Alien Species of Vascular Plants in Bulgaria*; NeoPrint: Sofia, Bulgaria, 2013; ISBN 9789549746303.
81. Dogan, Y.; Nedelcheva, A.; Łuczaj, Ł.; Drăgulescu, C.; Stefkov, G.; Maglajlić, A.; Ferrier, J.; Papp, N.; Hajdari, A.; Mustafa, B.; et al. Of the importance of a leaf: The ethnobotany of sarma in Turkey and the Balkans. *J. Ethnobiol. Ethnomedicine* **2015**, *11*, 1–15. [CrossRef]
82. Hakanova, A. *Lenten and Vegetarian Dishes*; Women Newspaper—Sofia: Sofia, Bulgaria, 1939.
83. Hakanova, A. *Bulgarian Folk Dishes*; Women Newspaper—Sofia: Sofia, Bulgaria, 1939.
84. Cholcheva, P. *1000 Well-Tried Cooking Recipes*; Women Newspaper—Sofia: Sofia, Bulgaria, 1952.
85. *Women Library New Cookbook*; Women Newspaper—Sofia: Sofia, Bulgaria, 1930.
86. *Women Library 500 Cooking Recipes*; Women Newspaper—Sofia: Sofia, Bulgaria, 1927.
87. Slaveykov, P. *Cookbook or How to Make Any Kind of Dish*, 5th ed.; Millenium: Sofia, Bulgaria, 1870; ISBN 978-954-515-299-3.
88. *Bulgarian Almanac Home Cookbook*; Reprint (2020); Bulgarian History: Sofia, Bulgaria, 1895; ISBN 978-619-7496-48-2.
89. Tardío, J.; Pardo-De-Santayana, M. Ethnobotanical Analysis of Wild Fruits and Vegetables Traditionally Consumed in Spain. In *Mediterranean Wild Edible Plants: Ethnobotany and Food Composition Tables*; Springer: New York, NY, USA, 2016; pp. 57–79. ISBN 9781493933297.
90. Abbet, C.; Mayor, R.; Roguet, D.; Spichiger, R.; Hamburger, M.; Potterat, O. Ethnobotanical survey on wild alpine food plants in Lower and Central Valais (Switzerland). *J. Ethnopharmacol.* **2014**, *151*, 624–634. [CrossRef]
91. Brunnbauer, U. *Gebirgsesellschaften Auf Dem Balkan: Wirtschaft Und Familienstrukturen Im Rhodopengebirge (19/20. Jahrhundert)*; Böhlau Verlag: Wien, Austria, 2004; ISBN 9783205771463.
92. Yarkov, D.; Stankov, K.; Stankov, I. Historical Review of the Development of Bulgarian Livestock Production. *Bulg. J. Agric. Sci.* **2022**, *28*, 564–578.
93. Stranski, I. Wild and Cultivated Plants in Bulgaria. Names, Distribution, Utilization. In *Plants in the Folk Customs and Songs*; Dinev, L., Ilchev, S., Levenson, E., Eds.; Bulgarian Academy of Sciences: Sofia, Bulgaria, 1963.

94. Łuczaj, Ł.J.; Kujawska, M. Botanists and their childhood memories: An underutilized expert source in ethnobotanical research. *Bot. J. Linn. Soc.* **2012**, *168*, 334–343. [[CrossRef](#)]
95. Łuczaj, Ł.; Pieroni, A. Nutritional Ethnobotany in Europe: From Emergency Foods to Healthy Folk Cuisines and Contemporary Foraging Trends. In *Mediterranean Wild Edible Plants: Ethnobotany and Food Composition Tables*; de Cortes Sánchez-Mata, M., Tardío, J., Eds.; Springer: New York, NY, USA, 2016; pp. 33–56. ISBN 9781493933297.
96. Ugulu, I.; Unver, M.C.; Dogan, Y. Potentially toxic metal accumulation and human health risk from consuming wild *Urtica urens* sold on the open markets of Izmir. *Euro-Mediterranean J. Environ. Integr.* **2019**, *4*, 36. [[CrossRef](#)]
97. Stark, P.B.; Miller, D.; Carlson, T.J.; de Vasquez, K.R. Open-source food: Nutrition, toxicology, and availability of wild edible greens in the East Bay. *PLoS ONE* **2019**, *14*, e0202450. [[CrossRef](#)]
98. Petrov, L.; Dzhelepov, N.; Jordanov, E.; Uzunova, S. *Bulgarian National Cuisine*; Zemizdat: Sofia, Bulgaria, 1978.
99. Baldi, A.; Bruschi, P.; Campeggi, S.; Egea, T.; Rivera, D.; Obón, C.; Lenzi, A. The Renaissance of Wild Food Plants: Insights from Tuscany (Italy). *Foods* **2022**, *11*, 300. [[CrossRef](#)]
100. Smith, J.; Jehlička, P. Stories around food, politics and change in Poland and the Czech Republic. *Trans. Inst. Br. Geogr.* **2007**, *32*, 395–410. [[CrossRef](#)]
101. Dogan, Y. Traditionally used wild edible greens in the Aegean Region of Turkey. *Acta Soc. Bot. Pol.* **2012**, *81*, 329–342. [[CrossRef](#)]
102. Bianco, V.; Santamaria, P.; Elia, A. Nutritional value and nitrate content in edible wild species used in southern Italy. *Acta Hort.* **1998**, *46*, 71–87. [[CrossRef](#)]
103. Łuczaj, Ł.; Dolina, K. A hundred years of change in wild vegetable use in southern Herzegovina. *J. Ethnopharmacol.* **2015**, *166*, 297–304. [[CrossRef](#)]
104. Pieroni, A.; Nebel, S.; Santoro, R.F.; Heinrich, M. Food for two seasons: Culinary uses of non-cultivated local vegetables and mushrooms in a south Italian village. *Int. J. Food Sci. Nutr.* **2009**, *56*, 245–272. [[CrossRef](#)]
105. Pardo-De-Santayana, M.; Tardío, J.; Morales, R. The gathering and consumption of wild edible plants in the Campoo (Cantabria, Spain). *Int. J. Food Sci. Nutr.* **2009**, *56*, 529–542. [[CrossRef](#)]
106. Turner, N.J.; Łuczaj, Ł.J.; Migliorini, P.; Pieroni, A.; Dreon, A.L.; Sacchetti, L.E.; Paoletti, M.G. Edible and Tended Wild Plants, Traditional Ecological Knowledge and Agroecology. *Crit. Rev. Plant Sci.* **2011**, *30*, 198–225. [[CrossRef](#)]
107. Tardío, J.; De Cortes Sánchez-Mata, M.; Morales, R.; Molina, M.; García-Herrera, P.; Morales, P.; Díez-Marqués, C.; Fernández-Ruiz, V.; Cámara, M.; Pardo-De-Santayana, M.; et al. Ethnobotanical and Food Composition Monographs of Selected Mediterranean Wild Edible Plants. In *Mediterranean Wild Edible Plants: Ethnobotany and Food Composition Tables*; Springer: New York, NY, USA, 2016; pp. 273–470. ISBN 9781493933297.
108. Motti, R.; Bonanomi, G.; Lanzotti, V.; Sacchi, R. The Contribution of Wild Edible Plants to the Mediterranean Diet: An Ethnobotanical Case Study along the Coast of Campania (Southern Italy). *Econ. Bot.* **2020**, *74*, 249–272. [[CrossRef](#)]
109. Pinke, G.; Kapcsándi, V.; Czúcz, B. Iconic Arable Weeds: The Significance of Corn Poppy (*Papaver rhoeas*), Cornflower (*Centaurea cyanus*), and Field Larkspur (*Delphinium consolida*) in Hungarian Ethnobotanical and Cultural Heritage. *Plants* **2022**, *12*, 84. [[CrossRef](#)] [[PubMed](#)]
110. Toscano, S.; Rizzo, V.; Muratore, G.; Romano, D. Edible Wild Flowers: An Innovative but Ancient Food. *Proceedings* **2020**, *70*, 32. [[CrossRef](#)]
111. Feduraev, P.; Skrypnik, L.; Nebreeva, S.; Dzhobadze, G.; Vatagina, A.; Kalinina, E.; Pungin, A.; Maslennikov, P.; Riabova, A.; Krol, O.; et al. Variability of Phenolic Compound Accumulation and Antioxidant Activity in Wild Plants of Some *Rumex* Species (*Polygonaceae*). *Antioxidants* **2022**, *11*, 311. [[CrossRef](#)] [[PubMed](#)]
112. Im, M.H.; Park, Y.-S.; Leontowicz, H.; Leontowicz, M.; Namiesnik, J.; Ham, K.-S.; Kang, S.-G.; Najman, K.; Gorinstein, S. The thermostability, bioactive compounds and antioxidant activity of some vegetables subjected to different durations of boiling: Investigation in vitro. *LWT-Food Sci. Technol.* **2011**, *44*, 92–99. [[CrossRef](#)]
113. Murador, D.; Braga, A.R.; Da Cunha, D.; De Rosso, V. Alterations in phenolic compound levels and antioxidant activity in response to cooking technique effects: A meta-analytic investigation. *Crit. Rev. Food Sci. Nutr.* **2017**, *58*, 169–177. [[CrossRef](#)]
114. Vuković, A.J.; Terzić, A. Gastronomy and Regional Identity: Balkan versus National Cuisine. In *Gastronomy for Tourism Development*; Peštek, A., Kukanja, M., Renko, S., Eds.; Emerald Publishing Limited: Bingley, UK, 2020; pp. 1–25. ISBN 978-1-78973-756-1.
115. Jasar, D.; Curcic, B.; Kubelka-Sabit, K.; Filipovski, V.; Kakurinov, V. Health Issues and Nutrition in the Balkans. *J. Hyg. Eng. Des.* **2021**, *35*, 96–105.
116. Gostin, A.-I. Traditional Balkan Foods in a Global Context: An Introduction. In *Nutritional and Health Aspects of Food in the Balkans*; Academic Press: Cambridge, MA, USA, 2021; pp. 1–8.
117. Watson, R.R.; Preedy, V.R.; Zibadi, S. (Eds.) *Polyphenols: Mechanisms of Action in Human Health and Disease*; Elsevier: Amsterdam, Netherlands, 2018; ISBN 9780128130063.
118. Zheng, Y.; Choi, Y.-H.; Lee, J.-H.; Lee, S.-Y.; Kang, I.-J. Anti-Obesity Effect of *Erigeron annuus* (L.) Pers. Extract Containing Phenolic Acids. *Foods* **2021**, *10*, 1266. [[CrossRef](#)]
119. Trifan, A.; Zengin, G.; Sinan, K.I.; Sieniawska, E.; Sawicki, R.; Maciejewska-Turska, M.; Skalikca-Woźniak, K.; Luca, S.V. Unveiling the Phytochemical Profile and Biological Potential of Five Species. *Antioxidants* **2022**, *11*, 1017. [[CrossRef](#)]
120. Guo, W.; Cao, P.; Wang, X.; Hu, M.; Feng, Y. Medicinal Plants for the Treatment of Gastrointestinal Cancers from the Metabolomics Perspective. *Front. Pharmacol.* **2022**, *13*, 909755. [[CrossRef](#)]

121. Jakovljević, M.R.; Milutinović, M.; Djurdjević, P.; Todorović, Ž.; Stanković, M.; Milošević-Djordjević, O. Cytotoxic and apoptotic activity of acetone and aqueous *Artemisia vulgaris* L. and *Artemisia alba* Turra extracts on colorectal cancer cells. *Eur. J. Integr. Med.* **2023**, *57*, 102204. [[CrossRef](#)]
122. Fernandes, L.; Ramalhosa, E.; Pereira, J.A.; Saraiva, J.A.; Casal, S. Borage, Camellia, Centaurea and Pansies: Nutritional, Fatty Acids, Free Sugars, Vitamin E, Carotenoids and Organic Acids Characterization. *Food Res. Int.* **2020**, *132*, 109070. [[CrossRef](#)] [[PubMed](#)]
123. Ebada, S.S.; El-Kashef, D.H.; Müller, W.E.; Proksch, P. Cytotoxic eudesmane sesquiterpenes from *Crepis sancta*. *Phytochem. Lett.* **2019**, *33*, 46–48. [[CrossRef](#)]
124. Arora, S.K.; Itankar, P.R.; Verma, P.R.; Bharne, A.P.; Kokare, D.M. Involvement of NF κ B in the antirheumatic potential of *Chenopodium album* L., aerial parts extracts. *J. Ethnopharmacol.* **2014**, *155*, 222–229. [[CrossRef](#)]
125. Choudhary, N.; Prabhakar, P.K.; Khatik, G.L.; Chamakuri, S.R.; Tewari, D.; Suttee, A. Evaluation of Acute toxicity, In-vitro, In-vivo Antidiabetic Potential of the Flavonoid Fraction of the plant *Chenopodium album* L. *Pharmacogn. J.* **2021**, *13*, 765–779. [[CrossRef](#)]
126. Parisa, N.; Hidayat, R.; Maritska, Z.; Prananjaya, B.A. Evaluation of the anti-gout effect of *Sonchus Arvensis* on monosodium urate crystal-induced gout arthritis via anti-inflammatory action—An in vivo study. *Med. Pharm. Rep.* **2021**, *94*, 358. [[CrossRef](#)]
127. Neha, K.; Haider, R.; Pathak, A.; Yar, M.S. Medicinal prospects of antioxidants: A review. *Eur. J. Med. Chem.* **2019**, *178*, 687–704. [[CrossRef](#)]
128. Hano, C.; Tungmunnithum, D. Plant Polyphenols, More than Just Simple Natural Antioxidants: Oxidative Stress, Aging and Age-Related Diseases. *Medicines* **2020**, *7*, 26. [[CrossRef](#)] [[PubMed](#)]
129. Forman, H.J.; Zhang, H. Targeting oxidative stress in disease: Promise and limitations of antioxidant therapy. *Nat. Rev. Drug Discov.* **2021**, *20*, 689–709. [[CrossRef](#)]
130. Arumugam, R.; Sarikurkcu, C.; Ozer, M.S. Comparison of methanolic extracts of *Doronicum orientale* and *Echium angustifolium* in terms of chemical composition and antioxidant activities. *Biocatal. Agric. Biotechnol.* **2021**, *33*, 101984. [[CrossRef](#)]
131. Di Ferdinando, M.; Brunetti, C.; Agati, G.; Tattini, M. Multiple functions of polyphenols in plants inhabiting unfavorable Mediterranean areas. *Environ. Exp. Bot.* **2014**, *103*, 107–116. [[CrossRef](#)]
132. Bautista, I.; Boscaiu, M.; Lidón, A.; Llinares, J.V.; Lull, C.; Donat, M.P.; Mayoral, O.; Vicente, O. Environmentally induced changes in antioxidant phenolic compounds levels in wild plants. *Acta Physiol. Plant.* **2015**, *38*, 9. [[CrossRef](#)]
133. Laoué, J.; Fernandez, C.; Ormeño, E. Plant Flavonoids in Mediterranean Species: A Focus on Flavonols as Protective Metabolites under Climate Stress. *Plants* **2022**, *11*, 172. [[CrossRef](#)] [[PubMed](#)]
134. Sharma, A.; Shahzad, B.; Rehman, A.; Bhardwaj, R.; Landi, M.; Zheng, B. Response of Phenylpropanoid Pathway and the Role of Polyphenols in Plants under Abiotic Stress. *Molecules* **2019**, *24*, 2452. [[CrossRef](#)] [[PubMed](#)]
135. Brahmi-Chendouh, N.; Piccolella, S.; Nigro, E.; Hamri-Zeghichi, S.; Madani, K.; Daniele, A.; Pacifico, S. *Urtica dioica* L. leaf chemical composition: A never-ending disclosure by means of HR-MS/MS techniques. *J. Pharm. Biomed. Anal.* **2021**, *195*, 113892. [[CrossRef](#)]
136. Carvalho, A.R.; Costa, G.; Figueirinha, A.; Liberal, J.; Prior, J.A.; Lopes, M.C.; Cruz, M.T.; Batista, M.T. *Urtica* spp.: Phenolic composition, safety, antioxidant and anti-inflammatory activities. *Food Res. Int.* **2017**, *99*, 485–494. [[CrossRef](#)]
137. Muccilli, V.; Kubik, J.; Waszak, Ł.; Adamczuk, G.; Humeniuk, E.; Iwan, M.; Adamczuk, K.; Michalczyk, M.; Korga-Plewko, A.; Józefczyk, A. Phytochemical Analysis and Anti-Cancer Properties of Extracts of *Centaurea castriferae* Borbás & Waisb Genus of *Centaurea* L. *Molecules* **2022**, *27*, 7537. [[CrossRef](#)]
138. Trichopoulou, A.; Vasilopoulou, E.; Hollman, P.; Chamalides, C.; Foufa, E.; Kaloudis, T.; Kromhout, D.; Miskaki, P.; Petrochilou, I.; Poulima, E.; et al. Nutritional Composition and Flavonoid Content of Edible Wild Greens and Green Pies: A Potential Rich Source of Antioxidant Nutrients in the Mediterranean Diet. *Food Chem.* **2000**, *70*, 319–323. [[CrossRef](#)]
139. Ceylan, S.; Cetin, S.; Camadan, Y.; Saral, O.; Ozsen, O.; Tutus, A. Antibacterial and antioxidant activities of traditional medicinal plants from the Erzurum region of Turkey. *Ir. J. Med. Sci.* **2019**, *188*, 1303–1309. [[CrossRef](#)]
140. Bilić, V.L.; Gašić, U.; Milojković-Opsenica, D.; Nemet, I.; Rončević, S.; Kosalec, I.; Rodriguez, J.V. First Extensive Polyphenolic Profile of *Erodium Cicutarium* with Novel Insights to Elemental Composition and Antioxidant Activity. *Chem. Biodivers.* **2020**, *17*, e2000280. [[CrossRef](#)]
141. Tomczyk, M.; Gudej, J.; Sochacki, M. Flavonoids from *Ficaria verna* Huds. *Z. Fur Nat.-Sect. C.* **2002**, *57*, 440–444. [[CrossRef](#)]
142. Karpiuk, V.; Konechna, R. Total phenolic and flavonoid content, antioxidant activity of *Ficaria verna*. *Sci. J. Pol. Univ.* **2021**, *46*, 229–234. [[CrossRef](#)]
143. Bhatti, M.Z.; Ali, A.; Ahmad, A.; Saeed, A.; Malik, S.A. Antioxidant and phytochemical analysis of *Ranunculus arvensis* L. extracts. *BMC Res. Notes* **2017**, *8*, 279. [[CrossRef](#)]
144. Chen, W.-C.; Wang, S.-W.; Li, C.-W.; Lin, H.-R.; Yang, C.-S.; Chu, Y.-C.; Lee, T.-H.; Chen, J.-J. Comparison of Various Solvent Extracts and Major Bioactive Components from *Portulaca oleracea* for Antioxidant, Anti-Tyrosinase, and Anti- α -Glucosidase Activities. *Antioxidants* **2022**, *11*, 398. [[CrossRef](#)] [[PubMed](#)]
145. Boskovic, I.; Đukić, D.A.; Mašković, P.; Mandić, L.; Perovic, S. Phytochemical composition and antimicrobial, antioxidant and cytotoxic activities of *Anchusa officinalis* L. extracts. *Biologia* **2018**, *73*, 1035–1041. [[CrossRef](#)]
146. Brown, A.W.; Stegelmeier, B.L.; Colegate, S.M.; Gardner, D.R.; Panter, K.E.; Knoppel, E.L.; Hall, J.O. The comparative toxicity of a reduced, crude comfrey (*Symphytum officinale*) alkaloid extract and the pure, comfrey-derived pyrrolizidine alkaloids, lycopsamine and intermedine in chicks (*Gallus gallus domesticus*). *J. Appl. Toxicol.* **2016**, *36*, 716–725. [[CrossRef](#)] [[PubMed](#)]

147. Miere, F.; Teușdea, A.C.; Laslo, V.; Cavalu, S.; Fritea, L.; Dobjanschi, L.; Zdrinca, M.; Zdrinca, M.; Ganea, M.; Pașc, P.; et al. Evaluation of In Vitro Wound-Healing Potential, Antioxidant Capacity, and Antimicrobial Activity of *Stellaria Media* (L.) Vill. *Appl. Sci.* **2021**, *11*, 11526. [[CrossRef](#)]
148. Rahman, M.; Khatun, A.; Liu, L.; Barkla, B.J. Brassicaceae Mustards: Traditional and Agronomic Uses in Australia and New Zealand. *Molecules* **2018**, *23*, 231. [[CrossRef](#)] [[PubMed](#)]
149. Narzary, H.; Basumatary, S. Amino Acid Profiles and Anti-Nutritional Contents of Traditionally Consumed Six Wild Vegetables. *Curr. Chem. Lett.* **2019**, *8*, 137–144. [[CrossRef](#)]
150. Saini, R.K.; Prasad, P.; Sreedhar, R.V.; Naidu, K.A.; Shang, X.; Keum, Y.-S. Omega-3 Polyunsaturated Fatty Acids (PUFAs): Emerging Plant and Microbial Sources, Oxidative Stability, Bioavailability, and Health Benefits—A Review. *Antioxidants* **2021**, *10*, 1627. [[CrossRef](#)]
151. Zekić, M.; Radonić, A.; Marijanović, Z. Glucosinolate Profiling of *Calepina irregularis*. *Nat. Prod. Commun.* **2016**, *11*, 1329–1332. [[CrossRef](#)]
152. Stojković, D.; Drakulić, D.; Schwirtlich, M.; Rajčević, N.; Stevanović, M.; Soković, M.; Gašić, U. Extract of *Herba Anthriscis cerefolii*: Chemical Profiling and Insights into Its Anti-Glioblastoma and Antimicrobial Mechanism of Actions. *Pharmaceuticals* **2021**, *14*, 55. [[CrossRef](#)]
153. Sergazy, S.; Vetrova, A.; Orhan, I.E.; Deniz, F.S.S.; Kahraman, A.; Zhang, J.-Y.; Aljofan, M. Antiproliferative and cytotoxic activity of Geraniaceae plant extracts against five tumor cell lines. *Futur. Sci. OA* **2022**, *8*, 0109. [[CrossRef](#)]
154. Ekiert, H.; Pajor, J.; Klin, P.; Rzepiela, A.; Ślesak, H.; Szopa, A. Significance of *Artemisia vulgaris* L. (Common Mugwort) in the History of Medicine and Its Possible Contemporary Applications Substantiated by Phytochemical and Pharmacological Studies. *Molecules* **2020**, *25*, 4415. [[CrossRef](#)]
155. Kokanova-Nedialkova, Z.; Nedialkov, P.T.; Nedialkov, P.T.; Nikolov, S.D. The Genus *Chenopodium*: Phytochemistry, Ethnopharmacology and Pharmacology. *Pharmacogn. Rev.* **2009**, *3*, 280–306.
156. Korpelainen, H.; Pietiläinen, M. Hop (*Humulus lupulus* L.): Traditional and Present Use, and Future Potential. *Econ. Bot.* **2021**, *75*, 302–322. [[CrossRef](#)]
157. Escher, G.B.; Santos, J.S.; Rosso, N.D.; Marques, M.B.; Azevedo, L.; do Carmo, M.A.V.; Daguier, H.; Molognoni, L.; do Prado-Silva, L.; Sant’Ana, A.S.; et al. Chemical study, antioxidant, anti-hypertensive, and cytotoxic/cytoprotective activities of *Centaurea cyanus* L. petals aqueous extract. *Food Chem. Toxicol.* **2018**, *118*, 439–453. [[CrossRef](#)] [[PubMed](#)]
158. Oladeji, O.S.; Oyebamiji, A.K. *Stellaria media* (L.) Vill.-A plant with immense therapeutic potentials: Phytochemistry and pharmacology. *Heliyon* **2020**, *6*, e04150. [[CrossRef](#)] [[PubMed](#)]
159. Kokanova-Nedialkova, Z.; Nedialkov, P.T.; Kokanova-Nedialkova, Z.; Nedialkov, P. Antioxidant Properties of 6-Methoxyflavonol Glycosides from the Aerial Parts of *Chenopodium Bonus-Henricus* L. *Artic. Bulg. Chem. Com-Munications* **2017**, *49*, 253–258.
160. Luczaj, L. Ethnobotanical Review of Wild Edible Plants of Slovakia. *Acta Soc. Bot. Pol.* **2012**, *81*, 198–225. [[CrossRef](#)]
161. Gupta, S.; Lakshmi, A.J.; Manjunath, M.; Prakash, J. Analysis of nutrient and antinutrient content of underutilized green leafy vegetables. *LWT-Food Sci. Technol.* **2005**, *38*, 339–345. [[CrossRef](#)]
162. Acık, D.Y.; Yilmaz, M.; Sahin, H.H.; Sayiner, Z.; Koruk, I.; Tiryaki, O.; Okan, V.; Pehlivan, M. Management of *Chenopodium polyspermum* toxicity with plasma exchange and hemodialysis. *J. Clin. Apher.* **2012**, *27*, 278–281. [[CrossRef](#)]
163. Günthardt, B.F.; Hollender, J.; Hungerbühler, K.; Scheringer, M.; Bucheli, T.D. Comprehensive Toxic Plants-Phytotoxins Database and Its Application in Assessing Aquatic Micropollution Potential. *J. Agric. Food Chem.* **2018**, *66*, 7577–7588. [[CrossRef](#)]
164. Stegelmeier, B.L.; Davis, T.Z.; Clayton, M.J. Plants Containing Urinary Tract, Gastrointestinal, or Miscellaneous Toxins that Affect Livestock. *Veter-Clin. N. Am. Food Anim. Pract.* **2020**, *36*, 701–713. [[CrossRef](#)] [[PubMed](#)]
165. Kopp, T.; Abdel-Tawab, M.; Mizaikoff, B. Extracting and Analyzing Pyrrolizidine Alkaloids in Medicinal Plants: A Review. *Toxins* **2020**, *12*, 320. [[CrossRef](#)] [[PubMed](#)]
166. El-Shazly, A.; Wink, M. Diversity of Pyrrolizidine Alkaloids in the Boraginaceae Structures, Distribution, and Biological Properties. *Diversity* **2014**, *6*, 188–282. [[CrossRef](#)]
167. Akbulut, S.; Semur, H.; Kose, O.; Ozhasenekler, A.; Celiktaş, M.; Basbug, M.; Yagmur, Y. Phytocontact dermatitis due to *Ranunculus arvensis* mimicking burn injury: Report of three cases and literature review. *Int. J. Emerg. Med.* **2011**, *4*, 7. [[CrossRef](#)]
168. Kocak, A.O.; Saritemur, M.; Atac, K.; Guclu, S.; Ozlu, I. A rare chemical burn due to *Ranunculus arvensis*: Three case reports. *Ann. Saudi Med.* **2016**, *36*, 89–91. [[CrossRef](#)] [[PubMed](#)]
169. Bender, N.; Chiu, Y. Photosensitivity. In *Nelson Textbook of Pediatrics*; Kliegman, R., Geme, J., Eds.; Elsevier: Amsterdam, The Netherlands, 2020; Volume 1, pp. 3496–3501. ISBN 9780323529501.
170. Wink, M. Modes of Action of Herbal Medicines and Plant Secondary Metabolites. *Medicines* **2015**, *2*, 251–286. [[CrossRef](#)]
171. Munjal, K.; Amin, S.A.; Mir, S.R.; Gauttam, V.K.; Gupta, S. Furanocoumarins and Lectins as Food Toxins. In *Plant-Derived Bioactives: Chemistry and Mode of Action*; Swamy, M.K., Ed.; Springer: Singapore, 2020; pp. 67–80. ISBN 9789811523618.
172. Levorato, S.; Dominici, L.; Fatigoni, C.; Zadra, C.; Pagiotti, R.; Moretti, M.; Villarini, M. In vitro toxicity evaluation of estragole-containing preparations derived from *Foeniculum vulgare* Mill. (fennel) on HepG2 cells. *Food Chem. Toxicol.* **2018**, *111*, 616–622. [[CrossRef](#)]
173. Koulman, A.; Kubbinga, M.E.; Batterman, S.; Woerdenbag, H.J.; Pras, N.; Woolley, J.G.; Quax, W.J. A Phytochemical Study of Lignans in Whole Plants and Cell Suspension Cultures of *Anthriscus Sylvestris*. *Planta Med.* **2003**, *69*, 733–738. [[CrossRef](#)]

174. Orčić, D.; Berežni, S.; Škorić, D.; Mimica-Dukić, N. Comprehensive study of *Anthriscus sylvestris* lignans. *Phytochemistry* **2021**, *192*, 112958. [\[CrossRef\]](#)
175. Piechowska, K.; Mizerska-Kowalska, M.; Zdzisińska, B.; Cytarska, J.; Baranowska-Łączkowska, A.; Jaroch, K.; Łuczykowski, K.; Płaziński, W.; Bojko, B.; Kruszewski, S.; et al. Tropinone-Derived Alkaloids as Potent Anticancer Agents: Synthesis, Tyrosinase Inhibition, Mechanism of Action, DFT Calculation, and Molecular Docking Studies. *Int. J. Mol. Sci.* **2020**, *21*, 9050. [\[CrossRef\]](#)
176. Todd, F.G.; Stermitz, F.R.; Schultheis, P.; Knight, A.P.; Traub-Dargatz, J. Tropane Alkaloids and Toxicity of *Convolvulus Arvensis*. *Phytochemistry* **1995**, *39*, 301–303. [\[CrossRef\]](#)
177. Shaghghi, A.; Alirezalu, A.; Nazarianpour, E.; Sonboli, A.; Nejad-Ebrahimi, S. Opioid alkaloids profiling and antioxidant capacity of *Papaver* species from Iran. *Ind. Crop. Prod.* **2019**, *142*, 111870. [\[CrossRef\]](#)
178. Grauso, L.; de Falco, B.; Motti, R.; Lanzotti, V. Corn poppy, *Papaver rhoeas* L.: A critical review of its botany, phytochemistry and pharmacology. *Phytochem. Rev.* **2020**, *20*, 227–248. [\[CrossRef\]](#)
179. Wencai, Y.; Ji, M.; Shouxun, Z. Bioactive Amino Acids from *Agrostemma Githago* L. with the Effects of Increasing Production of Wheat. *J. China Pharm. Univ.* **1997**, *28*, 65–68.
180. Alaca, K.; Okumuş, E.; Bakkalbaşı, E.; Javidipour, I. Phytochemicals and antioxidant activities of twelve edible wild plants from Eastern Anatolia, Turkey. *Food Sci. Technol.* **2021**, *42*, 18021. [\[CrossRef\]](#)
181. Woolum, J.A.; Akpunonu, P.; Johnson, M.; Webb, A.N. Human exposures to *Phytolacca americana* in Kentucky. *Toxicol.* **2022**, *220*, 106962. [\[CrossRef\]](#)
182. Qneibi, M.; Jaradat, N.; Zaid, A.N.Z.A.N.; Abu-Khalaf, N.; Natsheh, A.-R.; Hussen, F. Evaluation of Taste, Total Phenols and Antioxidant for Fresh, Roasted, Shade dried and Boiled leaves of Edible *Arum palaestinum* Bioss. *Marmara Pharm. J.* **2018**, *22*, 52–58. [\[CrossRef\]](#)
183. Haliński, A.; Bhatti, K.H.; Boeri, L.; Cloutier, J.; Davidoff, K.; Elqady, A.; Fryad, G.; Gadelmoula, M.; Hui, H.; Petkova, K.; et al. Stone composition of renal stone formers from different global regions. *Arch. Ital. Urol. E Androl.* **2021**, *93*, 307–312. [\[CrossRef\]](#)
184. Marais, J.P. Nitrate and Oxalates. In *Handbook of Plant and Fungal Toxicants*; D’Mello, J.P.F., Ed.; CRC Press: Boca Raton, FL, USA, 2020; pp. 205–218. ISBN 9780429281952.
185. Mahlangeni, N.T.; Moodley, R.; Jonnalagadda, S.B. The distribution of macronutrients, anti-nutrients and essential elements in nettles, *Laportea peduncularis* var. *peduncularis* (River nettle) and *Urtica dioica* (Stinging nettle). *J. Environ. Sci. Health Part B* **2015**, *51*, 160–169. [\[CrossRef\]](#)
186. Bello, O.M.; Fasinu, P.S.; Bello, O.E.; Ogbesejana, A.B.; Adetunji, C.O.; Dada, A.O.; Ibitoye, O.S.; Aloko, S.; Oguntoye, O.S. Wild vegetable *Rumex acetosa* Linn.: Its ethnobotany, pharmacology and phytochemistry—A review. *S. Afr. J. Bot.* **2019**, *125*, 149–160. [\[CrossRef\]](#)
187. Dawidowicz, A.L.; Wianowska, D.; Baraniak, B. The Antioxidant Properties of Alcoholic Extracts from *Sambucus Nigra* L. (Antioxidant Properties of Extracts). *LWT-Food Sci. Technol.* **2006**, *39*, 308–315. [\[CrossRef\]](#)
188. Młynarczyk, K.; Walkowiak-Tomczak, D.; Łysiak, G.P. Bioactive Properties of *Sambucus Nigra* L. as a Functional Ingredient for Food and Pharmaceutical Industry. *J. Funct. Foods* **2018**, *40*, 377–390. [\[CrossRef\]](#)
189. Mocanu, M.L.; Amarie, S. Elderberries-A Source of Bioactive Compounds with Antiviral Action. *Plants* **2022**, *11*, 740. [\[CrossRef\]](#) [\[PubMed\]](#)
190. Pacifico, S.; D’Abrosca, B.; Golino, A.; Mastellone, C.; Piccolella, S.; Fiorentino, A.; Monaco, P. Antioxidant Evaluation of Polyhydroxylated Nerolidols from Redroot Pigweed (*Amaranthus Retroflexus*) Leaves. *LWT-Food Sci. Technol.* **2008**, *41*, 1665–1671. [\[CrossRef\]](#)
191. Conforti, E.; Marrelli, M.; Carmela, C.; Menichini, F.; Valentina, P.; Uzunov, D.; Statti, G.A.; Duez, P.; Menichini, F. Bioactive Phytonutrients (Omega Fatty Acids, Tocopherols, Polyphenols), in Vitro Inhibition of Nitric Oxide Production and Free Radical Scavenging Activity of Non-Cultivated Mediterranean Vegetables. *Food Chem.* **2011**, *129*, 1413–1419. [\[CrossRef\]](#)
192. Kongdang, P.; Dukaew, N.; Pruksakorn, D.; Koonrunsesomboon, N. Biochemistry of *Amaranthus* Polyphenols and Their Potential Benefits on Gut Ecosystem: A Comprehensive Review of the Literature. *J. Ethnopharmacol.* **2021**, *281*, 114547. [\[CrossRef\]](#)
193. Bylka, W.; Stobiecki, M.; Frański, R. Sulphated Flavonoid Glycosides from Leaves of *Atriplex Hortensis*. *Acta Physiol. Plant* **2001**, *23*, 285–290. [\[CrossRef\]](#)
194. Bueno, M.; Lendínez, M.L.; Aparicio, C.; Cordovilla, M.P. Germination and Growth of *Atriplex Prostrata* and *Plantago Coronopus*: Two Strategies to Survive in Saline Habitats. *Flora* **2017**, *227*, 56–63. [\[CrossRef\]](#)
195. Rizk, A.M. *The Phytochemistry of the Flora of Qatar*; Scientific and Applied Research Centre, University of Qatar: Doha, Qatar, 1986; ISBN 9780855462246.
196. Mynarski, A.; Dawiec, E.; Grabowska, K.; Pietrzak, W.; Nowak, R.; Podolak, I. Qualitative and Quantitative Determination of Flavonoids in Different Organs of *Atriplex Nitens* Schkuhr and Evaluation of Anti-Hyaluronidase Activity. *Planta Med.* **2019**, *85*, P215. [\[CrossRef\]](#)
197. Van Niekerk, W.A.; Sparks, C.F.; Rethman, N.F.G.; Coertze, R.J. Mineral Composition of Certain *Atriplex* Species and *Cassia Sturtii* | South African Journal of Animal Science. *S. Afr. J. Anim. Sci.* **2004**, *34*, 94387. [\[CrossRef\]](#)
198. Ali, B.; Musaddiq, S.; Iqbal, S.; Rehman, T.; Shafiq, N.; Hussain, A. The Therapeutic Properties, Ethno Pharmacology and Phytochemistry of *Atriplex* Species: A Review. *Pak. J. Biochem. Biotechnol.* **2021**, *2*, 49–64. [\[CrossRef\]](#)
199. Usman, L.A.; Hamid, A.A.; Muhammad, N.O.; Olawore, N.O.; Edewor, T.I.; Saliu, B.K. Chemical Constituents and Anti-Inflammatory Activity of Leaf Essential Oil of Nigerian Grown *Chenopodium Album* L. *EXCLI J.* **2010**, *9*, 186.

200. Laghari, A.H.; Memon, S.; Nelofar, A.; Khan, K.M.; Yasmin, A. Determination of Free Phenolic Acids and Antioxidant Activity of Methanolic Extracts Obtained from Fruits and Leaves of *Chenopodium Album*. *Food Chem.* **2011**, *126*, 1850–1855. [[CrossRef](#)] [[PubMed](#)]
201. Chamkhi, I.; Charfi, S.; El Hachlafi, N.; Mechchate, H.; Guaouguauou, F.E.; El Omari, N.; Bakrim, S.; Balahbib, A.; Zengin, G.; Bouyahya, A. Genetic Diversity, Antimicrobial, Nutritional, and Phytochemical Properties of *Chenopodium Album*: A Comprehensive Review. *Food Res. Int.* **2022**, *154*, 110979. [[CrossRef](#)]
202. Hanganu, D.; Olah, N.; Vlase, L.; Mărculescu, A.; Pinte, A. Chemical Research of Carotenoids from *Chenopodium Bonus Henricus* L. (Chenopodiaceae). *Farmacologia* **2012**, *60*, 840–849.
203. Kokanova-Nedialkova, Z.; Nedialkov, P.; Kondeva-Burdina, M. Ultra-High-Performance Liquid Chromatography—High-Resolution Mass Spectrometry Profiling and Hepatoprotective Activity of Purified Saponin and Flavonoid Fractions from the Aerial Parts of Wild Spinach (*Chenopodium Bonus-Henricus* L.). *Z. Fur Nat.-Sect. C* **2021**, *76*, 261–271. [[CrossRef](#)]
204. Iwashina, T. Flavonoid Properties in Plant Families Synthesizing Betalain Pigments (Review). *Nat. Prod. Commun.* **2015**, *10*, 1103–1114. [[CrossRef](#)]
205. Nowak, R.; Szewczyk, K.; Gawlik-Dziki, U.; Rzymowska, J.; Komsta, Ł. Antioxidative and Cytotoxic Potential of Some *Chenopodium* L. Species Growing in Poland. *Saudi J. Biol. Sci.* **2016**, *23*, 15–23. [[CrossRef](#)] [[PubMed](#)]
206. Fejes, S.; Lemberkovics, É.; Balázs, A.; Apáti, P.; Kristó, T.S.; Szoke, É.; Kéry, Á.; Blázovics, A. Antioxidant Activity of Different Compounds from *Anthriscus Cerefolium* L. (Hoffm.). *Acta Hort.* **2004**, *597*, 191–198. [[CrossRef](#)]
207. Chizzola, R. Composition of the Essential Oils from *Anthriscus Cerefolium* Var. *Trichocarpa* and *A. Caucalis* Growing Wild in the Urban Area of Vienna (Austria). *Nat. Prod. Commun.* **2011**, *6*, 1147–1150. [[CrossRef](#)] [[PubMed](#)]
208. Slimstad, R.; Rathe, B.A.; Aesoy, R.; Diaz, A.E.C.; Herfindal, L.; Fossen, T. A Novel Bicyclic Lactone and Other Polyphenols from the Commercially Important Vegetable *Anthriscus Cerefolium*. *Sci. Rep.* **2022**, *12*, 7805. [[CrossRef](#)] [[PubMed](#)]
209. Kokkalou, E.; Stefanou, E. The Volatiles of *Chaerophyllum bulbosum* L. ssp. *Bulbosum* Growing Wild in Greece. *Pharm. Acta Helv.* **1989**, *64*, 133–134.
210. Stamenković, J.G.; Đorđević, A.S.; Stojanović, G.S.; Mitić, V.D.; Petrović, G.M. Phytochemical Analysis of Volatiles and Biological Activities of *Chaerophyllum Bulbosum* L. Essential Oils. *J. Serb. Chem. Soc.* **2021**, *86*, 257–267. [[CrossRef](#)]
211. Molo, Z.; Tel-Çayan, G.; Deveci, E.; Öztürk, M.; Duru, M.E. Insight into Isolation and Characterization of Compounds of *Chaerophyllum Bulbosum* Aerial Part with Antioxidant, Anticholinesterase, Anti-Urease, Anti-Tyrosinase, and Anti-Diabetic Activities. *Food Biosci.* **2021**, *42*, 101201. [[CrossRef](#)]
212. Senatore, F.; Oliviero, F.; Scandolera, E.; Tagliatalata-Scafati, O.; Roscigno, G.; Zaccardelli, M.; De Falco, E. Chemical Composition, Antimicrobial and Antioxidant Activities of Anethole-Rich Oil from Leaves of Selected Varieties of Fennel [*Foeniculum Vulgare* Mill. Ssp. *Vulgare* Var. *Azoricum* (Mill.) Thell]. *Fitoterapia* **2013**, *90*, 214–219. [[CrossRef](#)]
213. Barros, L.; Carvalho, A.M.; Ferreira, I.C.F.R. The Nutritional Composition of Fennel (*Foeniculum Vulgare*): Shoots, Leaves, Stems and Inflorescences. *LWT-Food Sci. Technol.* **2010**, *43*, 814–818. [[CrossRef](#)]
214. Comlekcioglu, N.; Çolak, S.; Aygan, A. A Study on the Bioactivity of Plant Extracts Obtained from *Arum Maculatum* Leaves by Different Extraction Techniques. *Croat. J. Food Technol.* **2021**, *16*, 41–46. [[CrossRef](#)]
215. Farahmandfar, R.; Esmaeilzadeh Kenari, R.; Asnaashari, M.; Shahrapour, D.; Bakhshandeh, T. Bioactive Compounds, Antioxidant and Antimicrobial Activities of *Arum Maculatum* Leaves Extracts as Affected by Various Solvents and Extraction Methods. *Food Sci. Nutr.* **2019**, *7*, 465–475. [[CrossRef](#)] [[PubMed](#)]
216. Kozuharova, E.; Naychov, Z.; Kochmarov, V.; Benbassat, N.; Gibernau, M.; Momekov, G. The Potential of *Arum* Spp. as a Cure for Hemorrhoids: Chemistry, Bioactivities, and Application. *Adv. Tradit. Med.* **2020**, *20*, 133–141. [[CrossRef](#)]
217. Melguizo-Melguizo, D.; Diaz-de-Cerio, E.; Quirantes-Piné, R.; Švarc-Gajić, J.; Segura-Carretero, A. The Potential of *Artemisia Vulgaris* Leaves as a Source of Antioxidant Phenolic Compounds. *J. Funct. Foods* **2014**, *10*, 192–200. [[CrossRef](#)]
218. Tigno, X.T.; de Guzman, F.; Flora, A.M.T.V. Phytochemical Analysis and Hemodynamic Actions of *Artemisia vulgaris* L. *Clin. Hemorheol. Microcirc.* **2000**, *23*, 167–175. [[PubMed](#)]
219. Thangjam, N.M.; Taijong, J.; Kumar, A. Phytochemical and Pharmacological Activities of Methanol Extract of *Artemisia vulgaris* L. Leaves. *Clin. Phytoscience* **2020**, *6*, 72. [[CrossRef](#)]
220. Sharonova, N.; Nikitin, E.; Terenzhev, D.; Lyubina, A.; Amerhanova, S.; Bushmeleva, K.; Rakhmaeva, A.; Fitsev, I.; Sinyashin, K. Comparative Assessment of the Phytochemical Composition and Biological Activity of Extracts of Flowering Plants of *Centaurea Cyanus* L., *Centaurea Jacea* L. and *Centaurea Scabiosa* L. *Plants* **2021**, *10*, 1279. [[CrossRef](#)] [[PubMed](#)]
221. Litvinenko, V.I.; Bubenichikova, V.N. Phytochemical Study of *Centaurea Cyanus*. *Chem. Nat. Compd.* **1988**, *24*, 672–674. [[CrossRef](#)]
222. Fernandes, L.; Pereira, J.A.; Saraiva, J.A.; Ramalhosa, E.; Casal, S. Phytochemical Characterization of *Borago Officinalis* L. and *Centaurea Cyanus* L. during Flower Development. *Food Res. Int.* **2019**, *123*, 771–778. [[CrossRef](#)]
223. Ebada, S.S.; Al-Jawabri, N.A.; Youssef, F.S.; Albohy, A.; Aldalaien, S.M.; Disi, A.M.; Proksch, P. In Vivo Antiulcer Activity, Phytochemical Exploration, and Molecular Modelling of the Polyphenolic-Rich Fraction of *Crepis Sancta* Extract. *Inflammopharmacology* **2019**, *28*, 321–331. [[CrossRef](#)]
224. Boskabadi, J.; Askari, Z.; Zakariaei, Z.; Fakhar, M.; Tabaripour, R. Mild-to-Severe Poisoning Due to *Conium Maculatum* as Toxic Herb: A Case Series. *Clin. Case Rep.* **2021**, *9*, e04509. [[CrossRef](#)]
225. Badalamenti, N.; Modica, A.; Iardi, V.; Bruno, M. Chemical Constituents and Biological Properties of Genus *Doronicum* (Asteraceae). *Chem. Biodivers.* **2021**, *18*, e2100631. [[CrossRef](#)] [[PubMed](#)]

226. Jeong, C.-H.; Nam, E.-K.; Shim, K.-H. Chemical Components in Different Parts of *Erigeron Annuus*. *J. Korean Soc. Food Sci. Nutr.* **2005**, *34*, 857–861. [[CrossRef](#)]
227. Poljuha, D.; Sladonja, B.; Šola, I.; Šenica, M.; Uzelac, M.; Veberič, R.; Hudina, M.; Famuyide, I.M.; Eloff, J.N.; Mikulic-Petkovsek, M. LC–DAD–MS Phenolic Characterisation of Six Invasive Plant Species in Croatia and Determination of Their Antimicrobial and Cytotoxic Activity. *Plants* **2022**, *11*, 596. [[CrossRef](#)]
228. Fontanel, D.; Galtier, C.; Viel, C.; Gueiffier, A. Caffeoyl Quinic and Tartaric Acids and Flavonoids from *Lapsana Communis* L. Subsp. *Communis* (Asteraceae). *Z. Fur Nat.-Sect. C* **1998**, *53*, 1090–1092. [[CrossRef](#)]
229. Seal, T. Quantitative HPLC Analysis of Phenolic Acids, Flavonoids and Ascorbic Acid in Four Different Solvent Extracts of Two Wild Edible Leaves, *Sonchus Arvensis* and *Oenanthe Linearis* of North-Eastern Region in India. *J. Appl. Pharm. Sci.* **2016**, *6*, 157–166. [[CrossRef](#)]
230. Itam, A.; Ismail, A.M.S.A.M.; Ismail, Z. Antioxidant and Antiangiogenic Properties, and Gas Chromatographic-Time of Flight Analysis of *Sonchus Arvensis* Leaves Extracts. *J. Chem. Soc. Pak.* **2015**, *37*, 1239.
231. De Paula Filho, G.X.; Barreira, T.F.; Pinheiro-Sant’Ana, H.M. Chemical Composition and Nutritional Value of Three *Sonchus* Species. *Int. J. Food Sci.* **2022**, *2022*, 4181656. [[CrossRef](#)]
232. Jimoh, F.; Adedapo, A.A.; Afolayan, A.J. Comparison of the Nutritive Value, Antioxidant and Antibacterial Activities of *Sonchus Asper* and *Sonchus Oleraceus*. *Rec. Nat. Prod.* **2011**, *5*, 29–42.
233. Wang, L.; Xu, M.L.; Liu, J.; Wang, Y.; Hu, J.H.; Wang, M.H. *Sonchus Asper* Extract Inhibits LPS-Induced Oxidative Stress and pro-Inflammatory Cytokine Production in RAW264.7 Macrophages. *Nutr. Res. Pract.* **2015**, *9*, 579–585. [[CrossRef](#)]
234. Biel, W.; Jaroszewska, A.; Łysoń, E.; Telesiński, A. The Chemical Composition and Antioxidant Properties of Common Dandelion Leaves Compared with Sea Buckthorn. *Can. J. Plant Sci.* **2017**, *97*, 1165–1174. [[CrossRef](#)]
235. Hussain, F.H.; Hama Saeed Hussain, F.; Ahamad, J.; Shafiq Osw, P. A Comprehensive Review on Pharmacognostical and Pharmacological Characters of *Anchusa Azurea*. *Adv. Med. Dent. Health Sci.* **2019**, *2*, 33–37. [[CrossRef](#)]
236. Kuruüzüm-Uz, A.; Güvenalp, Z.; Kazaz, C.; Salih, B.; Demirezer, L.Ö. Four New Triterpenes from *Anchusa Azurea* Var. *Azurea*. *Helv. Chim. Acta* **2010**, *93*, 457–465. [[CrossRef](#)]
237. Baghiani, A.; Naouel, B.; Trabsa, H.; Aouachria, S.; Boussoulim, N.; Arrar, L.; Boumerfeg, S. In Vivo Free Radical Scavenging, Antihemolytic Activity and Antibacterial Effects of *Anchusa Azurea* Extracts. *Int. J. Med. Med. Sci.* **2013**, *46*, 2051–5731.
238. Paun, G.; Neagu, E.; Albu, C.; Savin, S.; Radu, G.L. In Vitro Evaluation of Antidiabetic and Anti-Inflammatory Activities of Polyphenolic-Rich Extracts from *Anchusa Officinalis* and *Melilotus Officinalis*. *ACS Omega* **2020**, *5*, 13014–13022. [[CrossRef](#)]
239. Salehi, B.; Sharopov, F.; Tumer, T.B.; Ozleyen, A.; Rodríguez-Pérez, C.; Ezzat, S.M.; Azzini, E.; Hosseinabadi, T.; Butnariu, M.; Sarac, I.; et al. *Symphytum* Species: A Comprehensive Review on Chemical Composition, Food Applications and Phytopharmacology. *Molecules* **2019**, *24*, 2272. [[CrossRef](#)] [[PubMed](#)]
240. Haribal, M.; Renwick, J.A.A. Isovitexin 6''-O-β-d-Glucopyranoside: A Feeding Deterrent to *Pieris Napi* Oleracea from *Alliaria Petiolata*. *Phytochemistry* **1998**, *47*, 1237–1240. [[CrossRef](#)]
241. Cipollini, D.; Gruner, B. Cyanide in the Chemical Arsenal of Garlic Mustard, *Alliaria Petiolata*. *J. Chem. Ecol.* **2007**, *33*, 85–94. [[CrossRef](#)]
242. Frisch, T.; Agerbirk, N.; Davis, S.; Cipollini, D.; Olsen, C.E.; Motawia, M.S.; Bjarnholt, N.; Møller, B.L. Glucosinolate-Related Glucosides in *Alliaria Petiolata*: Sources of Variation in the Plant and Different Metabolism in an Adapted Specialist Herbivore, *Pieris Rapae*. *J. Chem. Ecol.* **2014**, *40*, 1063–1079. [[CrossRef](#)]
243. Kumarasamy, Y.; Byres, M.; Cox, P.J.; Delazar, A.; Jaspars, M.; Nahar, L.; Shoeb, M.; Sarker, S.D. Isolation, Structure Elucidation, and Biological Activity of Flavone 6-C-Glycosides from *Alliaria Petiolata*. *Chem. Nat. Compd.* **2004**, *40*, 122–128. [[CrossRef](#)]
244. Al-Snafi, A.E.; Al-Snafi, A.E. The Chemical Constituents and Pharmacological Effects of *Capsella Bursa-Pastoris*-A Review. *Int. J. Pharmacol. Toxicol.* **2015**, *5*, 76–81.
245. Duke, J.; Ayensu, E. *Medicinal Plants of China*; Reference Publications: Algonac, MO, USA, 1985; ISBN 9780917256202.
246. Pehlivan, M.; Akgul, H.; Yayla, F. The Some Nutrient and Trace Elements Content of Wild Plants Using as Ethno Botanical and Grown in the Gaziantep Region. *J. Appl. Pharm. Sci.* **2013**, *3*, 143–145. [[CrossRef](#)]
247. Song, N.; Xu, W.; Guan, H.; Liu, X.; Wang, Y.; Nie, X. Several Flavonoids from *Capsella Bursa-Pastoris* (L.) Medic. *Asian J. Tradit. Med.* **2007**, *2*, 218–222.
248. Kubínová, R.; Špačková, V.; Švajdlenka, E.; Lučivjanská, K. Antioxidant Activity of Extracts and HPLC Analysis of Flavonoids from *Capsella Bursa-Pastoris* (L.) Medik. *Ceska. Slov. Farn.* **2013**, *62*, 174–176. [[PubMed](#)]
249. Riaz, I.; Bibi, Y.; Ahmad, N.; Nisa, S.; Qayyum, A. Evaluation of Nutritional, Phytochemical, Antioxidant and Cytotoxic Potential of *Capsella Bursa-Pastoris*, a Wild Vegetable from Potohar Region of Pakistan. *Kuwait J. Sci.* **2021**, *48*. [[CrossRef](#)]
250. Montaut, S.; Bleeker, R.S. *Cardamine* sp.—A Review on Its Chemical and Biological Profiles. *Chem. Biodivers.* **2011**, *8*, 955–975. [[CrossRef](#)]
251. Bakhtiari, M.; Glauser, G.; Defosse, E.; Rasmann, S. Ecological Convergence of Secondary Phytochemicals along Elevational Gradients. *New Phytol.* **2021**, *229*, 1755–1767. [[CrossRef](#)]
252. Kejariwal, M.; Rodrigues, D.; Gowda, H. Study of Growth, Secondary Metabolites and Glucosinolate Content in *Cardamine Hirsuta* v/s *Brassica Juncea* (Indian Mustard). *J. Harmon. Res. Appl. Sci.* **2018**, *6*, 20–27. [[CrossRef](#)]

253. Agerbirk, N.; Hansen, C.C.; Olsen, C.E.; Kiefer, C.; Hauser, T.P.; Christensen, S.; Jensen, K.R.; Ørgaard, M.; Pattison, D.I.; Lange, C.B.A.; et al. Glucosinolate Profiles and Phylogeny in Barbarea Compared to Other Tribe Cardamineae (Brassicaceae) and Reseda (Resedaceae), Based on a Library of Ion Trap HPLC-MS/MS Data of Reference Desulfoglucosinolates. *Phytochemistry* **2021**, *185*, 112658. [[CrossRef](#)]
254. Bandara, M.; Savidov, N.; Driedger, D. Evaluation of Field Pepperweed (*Lepidium Campestre* L.) as a Source for Glucoraphanin Production. *Acta Hort.* **2008**, *765*, 165–172. [[CrossRef](#)]
255. Yusifova, D.Y.; Movsumov, I.S.; Garaev, E.A.; Mahiou-Leddiet, V.; Mabrouki, F.; Herbette, G.; Baghdikian, B.; Ollivier, E. Biologically Active Compounds from *Lepidium Campestre* and Pulp from Lemon-Juice Production. *Chem. Nat. Compd.* **2015**, *51*, 964–965. [[CrossRef](#)]
256. Kaur, T.; Hussain, K.; Koul, S.; Vishwakarma, R.; Vyas, D. Evaluation of Nutritional and Antioxidant Status of *Lepidium Latifolium* Linn.: A Novel Phytofood from Ladakh. *PLoS ONE* **2013**, *8*, e69112. [[CrossRef](#)]
257. Azimkhanova, B.B.; Ustenova, G.O.; Sharipov, K.O.; Rakhimov, K.D.; Sayakova, G.M.; Jumagazyeva, A.B.; Flisyuk, E.V.; Gemejiyeva, N.G. Chemical Composition and Antimicrobial Activity of Subcritical CO₂ Extract of *Lepidium Latifolium* L. (Brassicaceae). *Int. J. Biomater.* **2021**, *2021*, 4389967. [[CrossRef](#)] [[PubMed](#)]
258. Mirzaee, F.; Mohammadi, H.; Azarpeik, S.; Amiri, F.T.; Shahani, S. Attenuation of Liver Mitochondrial Oxidative Damage by the Extract and Desulfo Glucosinolate Fraction of *Lepidium perfoliatum* L. Seeds. *S. Afr. J. Bot.* **2021**, *138*, 377–385. [[CrossRef](#)]
259. Khare, C.P. *Indian Medicinal Plants: An Illustrated Dictionary*; Springer: New York, NY, USA, 2007; ISBN 978-0-387-70637-5.
260. Garcia-Herrera, P.; Sánchez-Mata, M.C.; Cámara, M.; Tardío, J.; Olmedilla-Alonso, B. Carotenoid Content of Wild Edible Young Shoots Traditionally Consumed in Spain (*Asparagus Acutifolius* L., *Humulus Lupulus* L., *Bryonia Dioica* Jacq. and *Tamus Communis* L.). *J. Sci. Food Agric.* **2014**, *94*, 1914–1916. [[CrossRef](#)]
261. Maietti, A.; Brighenti, V.; Bonetti, G.; Tedeschi, P.; Prencipe, F.P.; Benvenuti, S.; Brandolini, V.; Pellati, F. Metabolite Profiling of Flavonols and in Vitro Antioxidant Activity of Young Shoots of Wild *Humulus lupulus* L. (Hop). *J. Pharm. Biomed. Anal.* **2017**, *142*, 28–34. [[CrossRef](#)]
262. Abiko, Y.; Paudel, D.; Uehara, O. Hops Components and Oral Health. *J. Funct. Foods* **2022**, *92*, 105035. [[CrossRef](#)]
263. Weise, C.; Schrot, A.; Wuerger, L.T.D.; Adolf, J.; Gilabert-Oriol, R.; Sama, S.; Melzig, M.F.; Weng, A. An Unusual Type I Ribosome-Inactivating Protein from *Agrostemma githago* L. *Sci. Rep.* **2020**, *10*, 15377. [[CrossRef](#)] [[PubMed](#)]
264. Bohlooli, S.; Bohlooli, S.; Aslanian, R.; Nouri, F.; Teimourzadeh, A. Aqueous Extract of *Agrostemma Githago* Seed Inhibits Caspase-3 and Induces Cell-Cycle Arrest at G1 Phase in AGS Cell Line. *J. Ethnopharmacol.* **2015**, *175*, 295–300. [[CrossRef](#)]
265. Boukhira, S.; Boust, D.; El Mansouri, L.; Nordine, A.; Hamsas El Youbi, A.; Daoudi, A. Phytochemical Studies, Antioxidant Activity and Protective Effect on DNA Damage and Deoxyribose of *Silene Vulgaris* Extract from Morocco Medicinal Plants View Project Activités Apoptotiques et Antiprolifératives Es Extraits Des Plantes View Project Smahane Boukhira. *Artic. Int. J. Pharmacogn. Phytochem. Res.* **2015**, *7*, 1172–1178.
266. Zengin, G.; Mahomoodally, M.F.; Aktumsek, A.; Ceylan, R.; Uysal, S.; Mocan, A.; Yilmaz, M.A.; Picot-Allain, C.M.N.; Ćirić, A.; Glamočlija, J.; et al. Functional Constituents of Six Wild Edible *Silene* Species: A Focus on Their Phytochemical Profiles and Bioactive Properties. *Food Biosci.* **2018**, *23*, 75–82. [[CrossRef](#)]
267. Hu, Y.M.; Wang, H.; Ye, W.C.; Qian, L. New Triterpenoid from *Stellaria media* (L.) Cyr. *Nat. Prod. Res.* **2010**, *23*, 1274–1278. [[CrossRef](#)] [[PubMed](#)]
268. Sharma, A.; Sharma, V.; Kumawat, T.K.; Seth, R. A Review on Antidermatophytic Efficiency of Plant Essential Oils. *Int. J. Pure Appl. Biosci.* **2014**, *2*, 265–278.
269. Mojab, F.; Kamalinejad, M.; Ghaderi, N.; Vahidipour, H.R. Phytochemical Screening of Some Species of Iranian Plants. *Iran. J. Pharm. Res.* **2003**, *2*, 77–82. [[CrossRef](#)]
270. Saleem, U.; Zaib, S.; Khalid, S.; Anwar, F.; Akhtar, M.; Ahmad, B. Chemical Characterization, Docking Studies, Anti-Arthritic Activity and Acute Oral Toxicity of *Convolvulus arvensis* L. Leaves. *Asian Pac. J. Trop Biomed.* **2020**, *10*, 442. [[CrossRef](#)]
271. Menemen, Y.; Williams, C.; Jury, S. Flavonoid Patterns in *Convolvulus* L., (Convolvulaceae) Species from Morocco. *Pak. J. Bot.* **2002**, *34*, 291–295.
272. Khaliq, A.; Taviano, F.; Acquaviva, R.; Malfa, G.A.; Khaliq, H.A.; Ortiz, S.; Alhouayek, M.; Muccioli, G.G.; Quetin-Leclercq, J. Dereplication and Quantification of Major Compounds of *Convolvulus Arvensis* L. Extracts and Assessment of Their Effect on LPS-Activated J774 Macrophages. *Molecules* **2022**, *27*, 963. [[CrossRef](#)]
273. Syrchina, A.I.; Gorokhova, V.G.; Tyukavkina, N.A.; Babkin, V.A.; Voronkov, M.G. Flavonoid Glycosides of Spore-Bearing Stems of *Equisetum Arvense*. *Chem. Nat. Compd.* **1980**, *16*, 245–248. [[CrossRef](#)]
274. Boeing, T.; Tafarelo Moreno, K.G.; Gasparotto Junior, A.; Mota Da Silva, L.; De Souza, P. Phytochemistry and Pharmacology of the Genus *Equisetum* (Equisetaceae): A Narrative Review of the Species with Therapeutic Potential for Kidney Diseases. *Evid.-Based Complement. Altern. Med.* **2021**, *2021*, 6658434. [[CrossRef](#)]
275. Stankov, S.; Fidan, H.; Ivanova, T.; Stoyanova, A.; Damyanova, S.; Desyk, M. Chemical Composition and Application of Flowers of False Acacia (*Robinia pseudoacacia* L.). *Ukr. Food J.* **2018**, *7*, 577–588. [[CrossRef](#)]
276. Fecka, I.; Cisowski, W. Tannins and Flavonoids from the *Erodium Cicutarium* Herb. *Z. Fur Nat.-Sect. B* **2005**, *60*, 555–560. [[CrossRef](#)]
277. Radulović, N.; Dekić, M.; Stojanović-Radić, Z.; Palić, R. Volatile Constituents of *Erodium cicutarium* (L.) L' Hérit. (Geraniaceae). *Cent. Eur. J. Biol.* **2009**, *4*, 404–410. [[CrossRef](#)]

278. Stojanović-Radić, Z.; Čomić, L.; Radulović, N.; Dekić, M.; Randelović, V.; Stefanović, O. Chemical Composition and Antimicrobial Activity of Erodium Species: *E. ciconium* L., *E. cicutarium* L., and *E. absinthoides* Willd. (Geraniaceae). *Chem. Pap.* **2010**, *64*, 368–377. [[CrossRef](#)]
279. Eilers, E.J. Intra-Individual and Intraspecific Terpenoid Diversity in Erodium Cicutarium. *Plants* **2021**, *10*, 1574. [[CrossRef](#)]
280. Qahtan Mostafa Al-Smail, M.; Adnan Hussein, F. Isolation and Identification of Alkaloids and Glycosides Active Compound from Geranium Lucidum and Geranium Purpureum. *J. Phys. Conf. Ser.* **2019**, *1294*, 062040. [[CrossRef](#)]
281. Ilić, M.D.; Marčetić, M.D.; Zlatković, B.K.; Lakušić, B.S.; Kovačević, N.N.; Drobac, M.M. Chemical Composition of Volatiles of Eight *Geranium* L. Species from Vlasina Plateau (South Eastern Serbia). *Chem. Biodivers.* **2020**, *17*, e1900544. [[CrossRef](#)]
282. Fahamiya, N.; Shiffa, M.; Aslam, M. A Comprehensive Review on *Althaea Rosea* Linn. *Indo Am. J. Pharm. Res.* **2016**, *6*, 6888–6894.
283. Akhi, M.A. *An In-Vitro Study on Antioxidant Properties of Alcea Rosea Leaves*; Brac University: Dhaka, Bangladesh, 2020.
284. Abdel-salam, N.A.; Ghazy, N.M.; Sallam, S.M.; Radwan, M.M.; Wanas, A.S.; ElSohly, M.A.; El-Demellawy, M.A.; Abdel-Rahman, N.M.; Piacente, S.; Shenouda, M.L. Flavonoids of *Alcea Rosea* L. and Their Immune Stimulant, Antioxidant and Cytotoxic Activities on Hepatocellular Carcinoma HepG-2 Cell Line. *Nat. Prod. Res.* **2017**, *32*, 702–706. [[CrossRef](#)]
285. Mousavi, S.M.; Hashemi, S.A.; Behbudi, G.; Mazraedoost, S.; Omidifar, N.; Gholami, A.; Chiang, W.H.; Babapoor, A.; Pynadathu Rumjit, N. A Review on Health Benefits of *Malva Sylvestris* L. Nutritional Compounds for Metabolites, Antioxidants, and Anti-Inflammatory, Anticancer, and Antimicrobial Applications. *Evid.-Based Complement. Altern. Med.* **2021**, *2021*, 5548404. [[CrossRef](#)]
286. Unsal, C.; Sariyan, G.; Mat, A.; Oktayoglu, E.; Ozhatay, N. Distribution of Alkaloids in the Samples of *Papaver Dubium* Subsp. *Lecoqii* Var. *Lecoqii* from Turkey: A Potential Source for Thebaine Türkiye Geofitlerinin Kültüre Alınması Yeni Tür ve Çeşitlerin İlgili Sektörlere Kazandırılması View Project The Flora of Yıldız Mountains (Kırklareli) Biosphere Project View Project. *Biochem. Syst. Ecol.* **2006**, *34*, 170–173. [[CrossRef](#)]
287. Turan, M.; Kordali, S.; Zengin, H.; Dursun, A.; Sezen, Y. Macro and Micro Mineral Content of Some Wild Edible Leaves Consumed in Eastern Anatolia. *Acta Agric. Scand. Sect. B—Soil Plant Sci.* **2003**, *53*, 129–137. [[CrossRef](#)]
288. Ünsal, Ç.; Özbek, B.; Saryar, G.; Mat, A. Antimicrobial Activity of Four Annual Papaver Species Growing in Turkey. *Pharm. Biol.* **2009**, *47*, 4–6. [[CrossRef](#)]
289. Akrou, A.; El Jani, H.; Zammouri, T.; Mighri, H.; Neffati, M. Phytochemical Screening and Mineral Contents of Annual Plants Growing Wild in the Southern of Tunisia. *J. Phytol.* **2010**, *2*, 034–040.
290. Ali Hijazi, M.; Aboul-Ela, M.; Bouhadir, K.; Fatfat, M.; Khalife, H.; Ellakany, A.; Gali-Muhtasib, H. Cytotoxic Activity of Alkaloids from *Papaver Rhoeas* Growing in Lebanon. *Rec. Nat. Prod.* **2017**, *11*, 211–216.
291. Maurizi, A.; De Michele, A.; Ranfa, A.; Ricci, A.; Roscini, V.; Coli, R.; Bodesmo, M.; Buruni, G. Bioactive Compounds and Antioxidant Characterization of Three Edible Wild Plants Traditionally Consumed in the Umbria Region (Central Italy): *Bunias Erucago* L. (Corn Rocket), *Lactuca Perennis* L. (Mountain Lettuce) and *Papaver Rhoeas* L. (Poppy). *J. Appl. Bot. Food Qual.* **2015**, *88*, 109–114. [[CrossRef](#)]
292. Vardavas, C.I.; Majchrzak, D.; Wagner, K.H.; Elmadfa, I.; Kafatos, A. Lipid Concentrations of Wild Edible Greens in Crete. *Food Chem.* **2006**, *99*, 822–834. [[CrossRef](#)]
293. Marinaş, I.C.; Oprea, E.; Geană, E.; Mihaela Luntraru, C.; Elena Gîrd, C.; Chifiriuc, M.-C. Chemical Composition, Antimicrobial and Antioxidant Activity of *Phytolacca Americana* L. Fruits and Leaves Extracts. *Farmacologia* **2021**, *69*, 883–889. [[CrossRef](#)]
294. Proestos, C.; Boziaris, I.S.; Nychas, G.J.E.; Komaitis, M. Analysis of Flavonoids and Phenolic Acids in Greek Aromatic Plants: Investigation of Their Antioxidant Capacity and Antimicrobial Activity. *Food Chem.* **2006**, *95*, 664–671. [[CrossRef](#)]
295. Kim, Y.O.; Johnson, J.D.; Lee, E.J. Phytotoxic Effects and Chemical Analysis of Leaf Extracts from Three Phytolaccaceae Species in South Korea. *J. Chem. Ecol.* **2005**, *31*, 1175–1186. [[CrossRef](#)]
296. Saleri, F.D.; Chen, G.; Li, X.; Guo, M. Comparative Analysis of Saponins from Different Phytolaccaceae Species and Their Antiproliferative Activities. *Molecules* **2017**, *22*, 1077. [[CrossRef](#)]
297. Beara, I.N.; Lesjak, M.M.; Orčić, D.Z.; Simin, N.D.; Četojević-Simin, D.D.; Božin, B.N.; Mimica-Dukić, N.M. Comparative Analysis of Phenolic Profile, Antioxidant, Anti-Inflammatory and Cytotoxic Activity of Two Closely-Related Plantain Species: *Plantago altissima* L. and *Plantago lanceolata* L. *LWT-Food Sci. Technol.* **2012**, *47*, 64–70. [[CrossRef](#)]
298. Nichita, C.; Neagu, G.; Cucu, A.; Vulturescu, V.; Bertesteanu, Ş.V.G. Antioxidative Properties of *Plantago Lanceolata* L. Extracts Evaluated by Chemiluminescence Method. *AgroLife Sci. J.* **2016**, *5*, 95–102.
299. Grigore, A.; Bubueanu, C.; Pirvu, L.; Ionita, L.; Toba, G. *Plantago Lanceolata* L. Crops - Source of Valuable Raw Material for Various Industrial Applications. *Sci. Pap. -Ser. A Agron.* **2015**, *58*, 207–214.
300. Guil-Guerrero, J.L. Nutritional Composition of *Plantago* Species (*P. major* L., *P. lanceolata* L., and *P. media* L.). *Ecol. Food Nutr.* **2010**, *40*, 481–495. [[CrossRef](#)]
301. Nadgórska-Socha, A.; Kandziora-Ciupa, M.; Trześcki, M.; Barczyk, G. Air Pollution Tolerance Index and Heavy Metal Bioaccumulation in Selected Plant Species from Urban Biotopes. *Chemosphere* **2017**, *183*, 471–482. [[CrossRef](#)]
302. Lukova, P.K.; Karcheva-Bahchevanska, D.P.; Nikolova, M.M.; Iliev, I.N.; Mladenov, R.D. Comparison of Structure and Antioxidant Activity of Polysaccharides Extracted from the Leaves of *Plantago major* L., *P. media* L. and *P. lanceolata* L. *Bulg. Chem. Commun.* **2017**, *49*, 282–288.
303. Olaru, O.T.; Anghel, A.; Istudor, V.; Ancuceanu, R.; Dinu, M. Contributions to the Pharmacognostical and Phytobiological Study of *Fallopia Aubertii* (L. Henry) Holub. (Polygonaceae). *Farmacologia* **2013**, *61*, 991–999.

304. Zhang, F.-C.; Meng, Z.; Yin, D.; Su, L.; Chen, X. Study on the Differences in Chemical Compositions of Various Medicinal Parts of *Rumex Patientia* in Different Growth Years by High Performance Liquid Chromatography (HPLC). *Afr. J. Biotechnol.* **2009**, *10*, 15084–15088.
305. Demirel, S.; Fen, Ü.; Dergisi, B.E.; Kaya, E.; Akbaş, P.; Ceyhan, G.; Karabekmez Erdem, T.; Alkan, H.; Sütçü, K. Determination the Fatty Acid Composition of the *Rumex Patientia* L. Leaves and in Vitro Antimicrobial Activity of Their Different Extracts. *Süleyman Demirel Univ. J. Nat. Appl. Sci.* **2020**, *24*, 362–367. [[CrossRef](#)]
306. Petropoulos, S.A.; Fernandes, Â.; Dias, M.I.; Vasilakoglou, I.B.; Petrotos, K.; Barros, L.; Ferreira, I.C.F.R. Nutritional Value, Chemical Composition and Cytotoxic Properties of Common Purslane (*Portulaca Oleracea* L.) in Relation to Harvesting Stage and Plant Part. *Antioxidant* **2019**, *8*, 293. [[CrossRef](#)]
307. Bishop, R.R.; Kubiak-Martens, L.; Warren, G.M.; Church, M.J. Getting to the Root of the Problem: New Evidence for the Use of Plant Root Foods in Mesolithic Hunter-Gatherer Subsistence in Europe. *Veg. Hist. Archaeobot.* **2022**, 1–19. [[CrossRef](#)]
308. Treutter, D.; Wang, D.; Farag, M.A.; Baires, G.D.A.; Rühmann, S.; Neumüller, M. Diversity of Phenolic Profiles in the Fruit Skin of *Prunus Domestica* Plums and Related Species. *J. Agric. Food Chem.* **2012**, *60*, 12011–12019. [[CrossRef](#)]
309. Liu, W.; Nan, G.; Nisar, M.F.; Wan, C. Chemical Constituents and Health Benefits of Four Chinese Plum Species. *J. Food Qual.* **2020**, *2020*, 8842506. [[CrossRef](#)]
310. Drogoudi, P.; Pantelidis, G. Phenotypic Variation and Peel Contribution to Fruit Antioxidant Contents in European and Japanese Plums. *Plants* **2022**, *11*, 1338. [[CrossRef](#)] [[PubMed](#)]
311. Krauze-Baranowska, M.; Głód, D.; Kula, M.; Majdan, M.; Hałasa, R.; Matkowski, A.; Kozłowska, W.; Kawiak, A. Chemical Composition and Biological Activity of *Rubus Idaeus* Shoots—A Traditional Herbal Remedy of Eastern Europe. *BMC Complement Altern. Med.* **2014**, *14*, 480. [[CrossRef](#)] [[PubMed](#)]
312. Assafiri, O.; Abdallah, H.; El-Dakdouki, M. Antibacterial Effect and Phytochemical Analysis of the Shoot System of *Rubus Canescens* DC. Growing in Lebanon. *BAU J.-Sci. Technol.* **2020**, *2*, 9. [[CrossRef](#)]
313. Staszowska-Karkut, M.; Materska, M. Phenolic Composition, Mineral Content, and Beneficial Bioactivities of Leaf Extracts from Black Currant (*Ribes nigrum* L.), Raspberry (*Rubus idaeus*), and Aronia (*Aronia melanocarpa*). *Nutrients* **2020**, *12*, 463. [[CrossRef](#)]
314. Echavarría, A.; D'armas Regnault, H.; Nubia, L.; Matute, L.; Jaramillo, C.; Rojas-De-Astudillo, L.; Benítez, R. Evaluation of Antioxidant Capacity and Secondary Metabolites of Sixteen Medicinal Plants Extracts. *Ciencia UNEMI* **2016**, *9*, 29–35. [[CrossRef](#)]
315. Piluzza, G.; Bullitta, S. Correlations between Phenolic Content and Antioxidant Properties in Twenty-Four Plant Species of Traditional Ethnoveterinary Use in the Mediterranean Area. *Pharm. Biol.* **2011**, *49*, 240–247. [[CrossRef](#)]
316. Repajić, M.; Cegledi, E.; Zorić, Z.; Pedisić, S.; Garofulić, I.E.; Radman, S.; Palčić, I.; Dragović-Uzelac, V. Bioactive Compounds in Wild Nettle (*Urtica dioica* L.) Leaves and Stalks: Polyphenols and Pigments upon Seasonal and Habitat Variations. *Foods* **2021**, *10*, 190. [[CrossRef](#)]
317. Mzid, M.; Khedir, S.B.; Salem, M.B.; Regaieg, W.; Rebai, T. Antioxidant and Antimicrobial Activities of Ethanol and Aqueous Extracts from *Urtica Urens*. *Pharm. Biol.* **2017**, *55*, 775–781. [[CrossRef](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.