

Review



Use of Medicinal Plants in the Process of Wound Healing: A Literature Review

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Abstract: The literature on the use of medicinal plants in wound healing was comprehensively searched to obtain and assess the data. The data were procured via clinical studies that utilized medicinal plants and their compounds in vitro and in vivo for wound healing. This review collected data from electronic databases, including Google Scholar, PubMed, Science Direct, Web of Science, SciFinder, Thesis, and Scopus, using the search terms "natural products", "wound healing", and "natural compounds", along with the keywords "plants", "extracts", and "phytochemicals". Results from the last decade reveal a total of 62 families and 109 genera of medicinal plants, and their compounds have been studied experimentally both in vivo and in vitro and clinically found to effectively promote healing. This activity is related to the presence of secondary metabolites such as flavonoids, alkaloids, saponins, tannins, terpenoids, and phenolic compounds, which act at different stages through different mechanisms to exert anti-inflammatory, antimicrobial, and antioxidant effects, confirming that the use of medicinal plants could be an adequate alternative to current conventional practices for treating wounds.

Keywords: plant extracts; phytotherapy; treatments; healing; medicinal plants; secondary metabolites

1. Introduction

Statistics derived by the World Health Organization (WHO) have shown that around 80% of the world's population uses traditional medicine for primary healthcare, and 85% of this group utilizes plants. One of the great challenges of modern medicine concerns the healing and treatment of wounds. Studies by the WHO show that around 5 million people die annually because of imperfect wound healing. The use of natural products has shown promise in preventing and treating wounds. This review aims to elucidate the modes of preparation of herbal treatments, their phytochemical contents, and their use in formulations for wounds. Plants show a broad spectrum of bioactive phytochemicals, categorizable into the families of alkaloids, carotenoids, phenolic compounds, steroids, flavonoids, saponins, tannins, and terpenoids. These compounds act at different phases of the healing process through different mechanisms and show anti-inflammatory, antimicrobial, and antioxidant effects, whilst promoting collagen synthesis, cell proliferation, and angiogenesis. The application of natural compounds via new systems can contribute to enhancements in wound treatment.

2. Methods

2.1. Information Sources, Searching, and Selection of Studies

For this review, the period considered was January 2013 to October 2023, and the electronic databases used include Google Scholar, PubMed, Scopus, Science Direct, Web of Science, SciFinder, and Theses. The terms used for the search were "wound healing" (sought



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Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). in the titles and abstracts) and the keywords "plant", "extract", "natural compounds", and "phytochemicals". We also manually searched for references to bioactive phytochemicals that act at different phases of the healing process through different mechanisms and have anti-inflammatory, antimicrobial, antioxidant, and cell proliferation-promoting effects. The full documents were read to verify that they met the inclusion criteria.

2.2. Eligibility Criteria

Inclusion criteria: Studies published in English, including theses, articles, and proceedings from January 2013 to 01 October 2023, with "wound healing" in the title or abstract, addressing experimental or clinical studies.

Exclusion criteria: Newspapers and reviews.

2.3. Results

Following the initial screening, we identified approximately 5000 articles in different databases, citing about 480 different genera of plants with healing activities. Only 22% of these were included in this study, in accordance with the eligibility criteria. This is the first study in the last ten years to address plants and major and/or new compounds with regard to their activity in wound healing. The results obtained from eligible studies reveal a total of 62 families and 109 genera of medicinal plants used for wound treatment that have been discussed in studies from the last 10 years. Their effects, attributed to flavonoids, alkaloids, saponins, and phenolic compounds, which act at different stages through different mechanisms, include anti-inflammatory, antimicrobial, and antioxidant effects.

3. Classification of Wounds

The Healing Society defines wounds as physical lesions resulting from an opening or breaking of the skin that causes disturbances within the anatomy and normal functioning of the skin [1,2]. Wound healing can be a complex process because it entails a series of interdependent and overlapping stages: inflammation (exudative phase), reconstruction (proliferative phase), epithelization (regenerative phase), and maturation [3].

Several factors may affect the healing process, including (a) the presence of a contaminated surface contacting the wound; (b) delays due to the consumption of infected nutrients as a source of energy by white blood cells; (c) associated illnesses, such as diabetes and morbid obesity, which cause hyperglycemia and thus impact the defense mechanisms of the body, impairing the capacities of white blood cells in general, and especially neutrophils; and (d) treatment with radio-chemotherapy, NSAIDs and immunosuppressive drugs [4–6]. Wounds can be classified in various ways based on their etiology, their position, the kind of injury, the associated changes in bodily function, the wound depth, tissue loss, or clinical appearance. Table 1 describes the classification of a wound.

Table 1. Classification of a wound.

Classification	Туре		
Cause	Pathological: resulting from a pathology (pressure ulcer, neoplasia). Surgical or traumatic: resulting from surgery or trauma. Iatrogenic: resulting from procedures or treatment with radiotherapy.		
Evolution	Acute: wounds of easy resolution, rupture of vascularization, and mediate triggering of homeostasis (cuts, scoring, burns). Chronic: long-lasting wounds (deviation from the physiological cicatricial process).		
Presence of infection	Clean: free of microorganisms. Clean-contaminated: lesions less than 6 h between trauma and initial care. Infected: presence of local infectious agent. Contaminated: wounds serviced more than 6 h after trauma.		

Classification	Туре		
Regarding tissue impairment	Stage I: skin integrates with signs of hyperemia, discoloration, or hardening.		
	Stage II: the epidermis and dermis are ruptured, with subcutaneous tissue showing hyperemia,		
	blisters, and a shallow crater.		
	Stage III: total loss of cutaneous tissue, necrosis of the subcutaneous tissue to the muscular fascia.		
	Stage IV: great tissue destruction with necrosis reaching muscles, tendons, and bones.		
Degree of openness	Open: wounds in which the edges of the skin do not touch.		
	Closed: wounds where the edges of the skin are juxtaposed.		

3.1. Healing Process

Wounds can change the physiology of the skin, particularly those that affect the dermal layer. Therefore, tissue lesions can modify the anatomical structure of the skin, and the degree of damage to the tissue is highly dependent upon the healing mechanism. The wound-healing process entails a cascade of cellular and molecular events aimed at restoring the injured area [7].

The healing cascade is an organized sequence of events, and classifications have been applied to it to facilitate our understanding of the dynamic processes it involves that closely determine healing [8,9]. Different authors divide up the healing process in different ways; some consider the initial step to involve inflammation, followed by proliferation and ending with repair in the remodeling stage [7–9] (Figure 1).

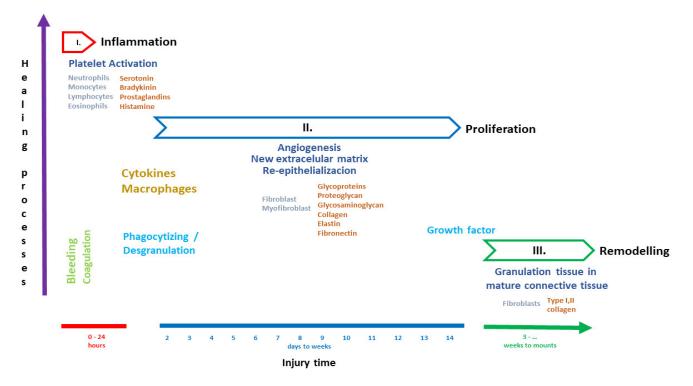


Figure 1. Wound-healing stages.

3.2. Inflammation

Inflammation is a defensive reaction to harmful agents, including microorganisms and damaged cells produced by the body, and promotes biological processes such as vascular responses and systemic reactions intended to reestablish the equilibrium of tissue homeostasis. In the absence of the inflammatory process, infections would develop in an uncontrolled manner; thus, the destructive processes unfolding in organs would continue until a total loss of function. The inflammatory process can be assessed clinically using five classical signs, called the "Cardinal Signals": swelling, heat, redness, pain, and loss of function [10,11].

The major function of the inflammatory response is the conduction of leukocytes to the affected region, which play an important role in defense by phagocytizing or producing substances that destroy microorganisms and necrotic tissues; they can inactivate or degrade antigens. Circulating cells such as neutrophils, monocytes, lymphocytes, and eosinophils reach the site of aggression through the bloodstream, crossing the vessel wall and migrating toward the site of aggression in significant quantities during the first 24 h of injury. The circulating cells are attracted by the notable inflammatory cytokine effects produced by activated platelets, endothelial cells, and the degraded products of the pathogens present in the lesion [12,13].

These chemical mediators can cause the dilation of the arterioles and increase the permeability of capillaries and venules, allowing a greater flow of blood to the damaged area, as well as the exudation (extravasation) of liquids, proteins, and defense cells into the interstitial space. Fluid exudation results in inflammatory edema, whereby the blood gradually becomes more viscous due to the increased density of red blood cells, and the circulation of the small vessels gradually slows, ultimately culminating in blood stasis (small, dilated vessels filled with red blood cells). At the same time, leukocytes migrate through the vascular wall into the interstitial space (called transmigration, diapedesis, or leukocyte emigration); this constitutes the initial (acute) phase of any type of inflammatory response, also referred to as the vascular phenomena [12,14,15].

The recruitment and activation of inflammatory cells, in either acute or chronic inflammation, are consequences of physical changes occurring at the wound site. The first of these changes is the liberation of substances following platelet degranulation. These include thrombin, which prompts the release of distinct growth factors such as platelet-derived growth factors (PDGFs), transforming growth factor- β (TGF- β), epidermal growth factor (EGF), transforming growth factor- α (TGF- α), and endothelial cell growth factor (VEGF). The mediators mentioned beforehand include adhesive glycoproteins such as fibronectin and thrombospondin—important constituents of the extracellular matrix [16].

The inflammatory phase of wound healing includes the activation of vasoactive substances such as serotonin, bradykinin, prostaglandins, and histamine. These can increase the permeability of the endothelium at the lesion site and enhance interstitial fluid perfusion in this area. The increase in permeability facilitates infiltration by immune and repair cells that facilitate the previously described events, while the increase in circulation leads to greater oxygen distribution in the tissue; consequently, the temperature increases at the site of injury. The warm and humid microenvironment thus produced within the wound is essential for the ensuing healing phase. At the end of the inflammatory stage of wound healing, macrophages synthesize distinct growth factors, such as PDGF, TGF- β , fibroblast growth factor (FGF), and VEGF, which stand out as the major cytokines required to stimulate the formation of granulation tissue and thus generate the environment required for the next phases of cell proliferation and repair [1,17].

3.3. Proliferation

The closure of the lesion occurs in the proliferation phase. Angiogenesis occurs as a result of the formation of granulation tissue and is responsible for filling the injured tissue. The new extracellular matrix that will be involved in cell growth and the new blood vessels that will convey oxygen and nutrients indispensable for local cellular metabolism are produced with the aid of fibroblasts. With the progression of the proliferative phase, the provisional matrix changes as a result of the newly formed granulation tissue. Wound epithelialization represents the final stage of the proliferative phase [18,19].

The formation of the extracellular matrix provides a substrate for cell adhesion and regulates the growth, movement, and differentiation of cells within it. The extracellular matrix consists of structural proteins, including collagen and elastin, along with an interstitial matrix composed of adhesive glycoproteins, proteoglycan, and glycosaminoglycan [20,21].

The increase in microvascular permeability, characteristic of the inflammatory process, represents the first stage of this proliferative process; here, cellular elements, along with cytokines, are released, and we also see the formation of the provisional extracellular matrix necessary for the migration and proliferation of endothelial cells [22].

Angiogenesis is an exceedingly important stage in the healing process, during which new blood vessels are formed from preexisting vessels. The new vessels participate in the formation of provisional granulation and the supply of nutrients and oxygen to the growing tissue. On the other hand, vasculogenesis refers to the early stages of vascular development, during which vascular endothelial precursor cells enact the mobilization of endothelial progenitors derived from bone marrow [23].

Fibroblast migration is induced by the PDGF and TGF- β released by macrophages. When fibroblasts reach the wound bed, they proliferate and produce matrix proteins such as fibronectin, collagen, and proteoglycans. These components help build the new extracellular matrix, which supports the further growth of cells essential for the repair process. A crucial interaction takes place between the fibroblasts and extracellular matrix, which regulates the additional synthesis of the components along with tissue remodeling [24].

The process of re-epithelialization in the injured tissue is accelerated by the contraction of the underlying connective tissue, which is responsible for the approximation of the wound's margins. This contraction is induced by myofibroblasts, activated by TGF- β and PDGF; thus, these myofibroblasts play an important role in wound healing, especially for open lesions. When present in open wounds, these cells produce larger amounts of extra-cellular matrix components. However, if abnormalities arise in the physiological process, such as delays, this may cause cicatricial defects due to alterations to the differentiation of fibroblasts in myofibroblasts [25].

3.4. Remodeling

The final stage of wound healing involves the remodeling or maturation of granulation tissue into mature connective tissue or scar tissue. The wound-healing process is most potent during this phase. Wound maturation begins during the third week after the wounding and is characterized by an increase in resistance and a controlled decrease in the amount of collagen. This mechanism is characterized by a balance in the production and destruction of collagen fibers, resulting from the action of an enzyme called collagenase. An imbalance in this relationship can favor the emergence of hypertrophic and keloid scars [1,2].

Finally, the remodeling process consists of the proper deposit of elements previously mentioned, mainly including collagen fibers. This stage involves a change in the type of collagen present and its disposition. Type III collagen, which is initially more abundant in the wound than type I, is more actively degraded over time; in contrast, the production of collagen I by fibroblasts increases, and this causes an increase in tension force and the reduction in the quantity of collagen [3,4].

Numerous factors can aggravate the wound-healing process, specifically as regards the biological events that comprise it. Factors such as advanced age, the patient's nutritional status, and vascular changes can directly alter the healing process. However, diabetes mellitus also drastically alters the process of tissue recovery, interfering at all stages and thus causing serious complications for the patient.

The process of wound healing is complicated when the patient is diabetic. In patients with diabetes, wounds show less revascularization and lower expression of growth factors compared to injuries in non-diabetics, thus impairing healing. These complications can evolve to produce severe consequences, such as a stagnant repair mechanism leading to a loss of tissue function [26–28].

Impaired healing in diabetic patients is characterized by acute inflammation and abnormalities in angiogenesis, entailing difficulties in forming new blood. The proper healing of a wound requires a regulated inflammatory response; however, diabetic wounds show prolonged inflammatory responses. Wounds in DM1 exhibit increased expression of inflammatory cytokines, including tumor necrosis factor alpha (TNF- α) and interleukins IL-1 and IL-6, and decreased IL-10, leading to injury following a prolonged inflammatory phase. This deregulated and prolonged inflammation leads to the wound becoming chronic and unable to be completely healed [29]. These chronic injuries, such as foot ulcers (diabetic foot), lead to high morbidity and increased treatment costs. In addition, foot ulcers substantiate more than 50% of the cases of amputation among diabetics. Increased oxidative stress is one of the leading causes of wound complications in diabetics, causing late scarring. Reducing persistent inflammation and the excretion of free radicals by incorporating an

strategy for improving the healing of diabetic wounds [30]. Factors associated with both angiogenesis and the vasculogenesis process are vital for wound healing, as they play a vital role in the delivery of oxygen, nutrients, and other mediators to the wound site. Thus, they have become therapeutic targets that can improve the healing of damaged wounds in diabetes patients when activated, thus restoring the neovasculogenesis mechanism [31].

anti-inflammatory and antioxidant agent into wound treatment has become an important

4. Medicinal Plants Used for Wound Healing

Preparations made using medical plants (such as extracts) and the active compounds present in some of these plants have been used to accelerate wound healing. The ethnopharmacological approach to investigating medicinal plants consists of combining information acquired from users of medicinal flora (traditional communities) with the results of chemical and pharmacological studies [32].

The application of medicinal plants has always been a part of the evolution of humanity; these plants represent one of the first therapeutic resources to be used by humans, and they still hold great importance for the maintenance of human health. According to the World Health Organization (WHO, 2002) [33], approximately 80% of the population in developing countries use traditional medicine as their primary healthcare, most of which entails using plant extracts or their active compounds. According to the statistics provided by the WHO, medicinal plants, herbal preparations, or derived products are conventionally used in primary care in various countries. The WHO classifies a medicinal plant as a plant species that, when administered to humans, exerts a pharmacological action. The findings of ethnopharmacology, in terms of the therapeutic properties of plants and popular knowledge regarding their usage, have been presented as source material for developing technical scientific knowledge. The accumulation of information regarding the use of natural assets by traditional populations has provided researchers with models for the sustainable use of these resources while also providing directions for the exploitation of the pharmacological properties of certain species. Over the centuries, products of plant origin have been commonly used as the basis of treatments for different diseases by virtue of knowledge transmitted down through generations, and certain plant species can be understood as sources of active molecules [34–36].

In the context of wound healing, the utilization of plants and plant extracts dates to the prehistoric era [37]. Records describe the use of plants and extracts in the form of poultices to stop hemorrhages and to facilitate cicatrization. Other uses have been described in relation to the ingestion of certain plants, which act systemically [36]. Thus, the data collected through the years confirm that the development of modern medicine has only been possible via the inheritance of ancient healing methods and the empirical knowledge pertaining to such practices [37]. Every year, approximately 100 million patients around the world acquire scars resulting from surgical interventions, burns, or tissue ruptures due to accidents of various kinds, which require effective and rapid treatment. These statistics indicate that wound healing is a modern therapeutic challenge [38]. Multiple studies have sought to improve the treatment of wounds by promoting the healing process; nevertheless, the most effective organic and inorganic substances in this regard remain a scientific mystery to this day [39]. Healing involves several complex processes in which different cellular structures are involved. The process begins with an amplified immune

response that prevents wound complications, enacted via chemoattraction, which facilitates the development of other mediators necessary to subsequent phases, such as inflammation, cell proliferation, and re-epithelization, which eventually lead to wound closure [40].

Medicinal plants are significant sources of novel chemical substances with valuable therapeutic effects. Table 2 displays the families and genera of plants utilized for wound healing. A total of 62 families and 109 genera were documented with applicability in wound healing and treatment based on traditional medicine (Table 2). Most of the wound-healing information was collected from recent literature from the last 10 years. The Euphorbiaceae family was the least represented (five members), followed by the Asteraceae family (six members) and the Fabaceae family (eight members). The most commonly used plant parts were cited as leaves (37%), followed by fruits (9%), seeds (8%), roots (8%), aerial parts (7%), flowers (6%), the whole plant (6%) bark (5%), saponins (3%), rhizome (2%) and others. These data also show that medicinal plants are used to treat wounds in many different parts of the world. Different families and genera have been analyzed in this work as regards their components. In 36% of the genera, the major phytochemical compounds found were alkaloids, steroids, flavonoids, saponins, tannins, and terpenes.

Ref. Family Genus Part Used/Type Extraction Compounds Alkaloids, Flavonoids, Glycosides, Justicia flava Leaf/Methanol [41] Tannins Acanthaceae A. paniculata Leaf/10% aqueous extract Diterpenoids [42] Leaf/Ethanol Achyranthes aspera Flavonoids, Saponins, Tepernoids [43] 2,4-dihydroxy-2,5-dimethyl-3(2H)furan-3-one, hexadecanoic acid, A. sessilis Stem and Leaf/Methanol [44] 2-1,2,4-trioxolane,3-phenyl-, Amaranthaceae palmitate-ethyl-, L-glutamic acid. A.triandra Air seed/Petroleum ether Oil ricinoleic acid [45] Root/Dichloromethane and Celosia argentea Terpenoids [46] ethyl acetate Alkaloids, Flavonoids, Polyphenols, Buchanania lanzan S. [47] Root/Petroleum ether Steroids Anacardiaceae Lannea welwitschii Alkaloids, Flavonoids, Glycosides, Leaf/Methanol [41] Hiern Steroids, Tannins Leaf/Ethanol [48] Angelica sinensis n-buthylidenephthalide and proteins Asiaticoside, Madecassic acid, Leaf/Methanol [49] Centella asiatica Madecassoside asiatic acid, Apiaceae Triterpenes, Essential oils Cuminum cyminum Seeds [50] L. striatum Rhizoma Essential oils [51] Alkaloids, Phenols, Proteins, Saponins, Catharanthus roseus Leaf/Aqueous and Methanol [52] Tannins Alkaloids, Flavonoids, Saponins, S. hispidus Leaf and Root [53] Tannins, Apocyanaceae Alkaloids, Flavonoids, Phenolics, Wrightia tinctoria Leaf/Aqueous [54] Saponins, Tannins Saba florida Leaf/Methanol 99.9% Total extract [55]

Table 2. Families and Genus with wound healing activity.

Family	Genus	Part Used/Type Extraction	Compounds	Ref
	Panax ginseng	Panax ginseng saponins (PGS)	Ginsenoside Rb1 (G-Rb1)	[56]
Araliaceae	Panax notoginseng	Panax notoginseng saponins (PNS)	High-glucose (HG-30Mn)	[57]
Asclepiadaceae	Calotropis giganthea	Root Bark	Taraxasteroryl isovalerate, Gigantin, Giganteol, Isogiganteol, α-amyrin-3-amyrin, Taraxasterol	[58]
	Calotropis procera	Root bark/Ethanol	Alkaloids, Flavonoids, Steroids, Tannins	[59]
Asphodelaceae	Aloe vera	Leaf/Acetonic extract	Polymers	[60]
	Achillea millefolium	Aerial parts	Yarrow Oil	[61]
	Arctium lappa	Ground bark/Ethanol	Alkaloids, Flavonoids, Lignans, Phenolic acid, Tannins, Terpenoids	[62]
Asteraceae	Blumea balsamifera	Leaf/Methanol 95%	Flavonoids, Nonvalatile constituents	[63]
Asteruceue	Calendula officinalis	Flowers/Hydroethanol	Rutin, Quercetin-3-O-glucoside	[64,6
	Carthamus tinctorius	Saflowers	Hydroxysaflow yellow A (HSYA)	[66]
	Wedelia trilobata	Leaves/Ethylacetate, Chloroform:Methanol	Kaura-9(11),16-dien-19-oic acid	[67]
	Kigelia africana	Leaves/Roots/Methanol	Flavonoids, Carbohydrates, Sapogenetic glycosides, Saponins, Steroids	[68]
Bignoniaceae	S. campanulata	Leaf/Methnol	Flavonoids, Phenols, Saponins, Steroids	[69]
	Tecoma capensis	Shoots/Hydroalcoholic	Myrecetin	[70]
Boraginaceae	H. indicum	Leaf/Ethanol	Crude extract	[71]
	L. erythrorhizon	Root	Purification of Shikonin	[72
D	Boswelia sacra	Leaf/Methanol	Oil	[73
Burseracea	C. myrrha	Leaf/Methanol	Oleo-gum-resins	[73,7
Cactaceae	O. ficus-indica	Seed/Oil extraction	OFI-SNEDDSs	[75]
Caricaceae	Carica papaya	Papaya fruit extraction	Crude extract	[76
	Cecropia peltata	Leaf	Saponins	[77]
Cecropiaceae	Myrianthus arboreus	Leaves/Ethanol	Alkaloids, Flavonoids, Glycosides, Sterols, Tannins, Terpenoids	[78]
Caprifoliacea	Locinera japonica	Flowers/Ethanol	Chlorogenic acid	[79
	C. mucronatum	Leaf/Ethanol	Procyanidin B2	[80
Combretaceae	Terminalia chebula	Fruit extraction	Anthraquinone, Flavonoids, Sapogenins, Saponins, Steroids, Tannins	[81]
	Terminalia arjuna	Fruit extraction/Methanol	Anthraquinones, Carbohydrates, Flavonol, Glucose sorbitol, Hydrolyzable Tannins	[82

Fam:1.	Conuc	Part Hood/Truna Estrection	Compounds	Ref.
Family	Genus	Part Used/Type Extraction	Compounds	Ker.
Crassulaceae	<i>Bryophylum pinnatum</i> Lam	Leaf/Aqueous	Patulitin-O-deoxy-hexoside-O- hexoside, Quercetin-O-hexoside, Quercetin-O-deoxy-hexoside-O- pentoside	[83]
Cyperacea	Cyperus rotundus L	Aerial part/Methanol	Alkaloids, Phenols	[84]
	Alchornea cordifolia (Schum & Thonn)	Leaf/Ethanol	Quercetin, Hyperin, Guaijaverin	[78]
	Euphoria hirta	Whole plant/Methanol	Alkaloids, Flavonoids, Glycosides, Proteins, Saponins, Tannins	[85]
Euphorbiacea	Jatropha curcas L.	Flowers/Methanol	Alkaloids, Flavonoids, Glycoside, Saponins, Tannins	[86]
	<i>Mallotus oppositifolius</i> (Geiseler)	Leaf/Ethanol	Aspinidiol B, methylene bis-aspidinol, α -tocoferol	[87]
	P. emblica L.	Leaves/Ethanol	Flavonoids, Saponins, Tannins	[88]
	P. muellerianus (Kuntze)	Leaf/Aqueous	Geranin	[89]
	Astragalus membranaceus Sprants	Seeds/Ethanol	Tryptophan, Linoleic acid, Adenine	[90]
	Caesalpinia sappan L.	Wood	Sappachalcone	[91]
	Entada phaseoloides		Total Tannins	[92]
	Glycyrrhiza glabra L.	Root/Ethanol	Glycyrrhiza cream	[93]
Fabaceae	Indigofera enneaphylla L.	Whole plant/Petroleum ether, Ethyl Acetate, Ethanol	Flavonoids, Saponins, Tannins	[94]
	Mimosa pudica L.	Seeds/Ethyl Acetate Root/Petroleum ether	Alkaloids, Glycosides, Phytosterol	[95,90
	Sophora flavescens		Compound, Sophora flavescen lotion	[97]
	Tephrosia purpurea	Aerial plants/Ethanol	Flavonoids, TPF-A 7 peaks	[98]
Fagaceae	Quercus infectoria Oliver	Nutgails/Ethanol	Pharmaceutical formulations	[99]
Ganodermataceae	Ganoderma lucidum	Fruting bodies/Hot water	Polysaccharides 25.1% Ganodermic acid A	[100
Gentianaceae	<i>Anthocleista nobilis</i> G. Don	Stem bark/Ethanol Ethyl Acetate Buthanol n-Hexane	Isovitexin and Isovitexin-2"-O-xyl Isovitexin Apigenin monoglycoside p-Hydroxybenzoic acid, Sarasinside	[101
Ginkgoaceae	Ginkgo biloba L.	Leaf/Aqueous	Myricerin, Quercetin, Kaempferol, Isorhamnitin, Terpenes lactones, Ginkgolic acid	[102
Hypericaceae	Hypericu mysorense	Parts plant/Methanol	Flavonoids, Saponins, Tannins	[103
Iridaceae	Crocus sativus L.	Stigmas/Glycerin/water/Ethanol	Flavonoids, Anthocyanins	[104
	Occimum sanctum L.	Leaf/Water	Essential Oil	[105
Lamiaceae	Rosmarinus officinalis	Aerial parts/Hydrodistillation	Essential Oil	[106
	Salvia miltiorrhiza	Leaf/Hydroethanolic	Flavonoids, Total Phenols	[107
Lauraceae	Cinnamomum cassia		Cinnamon Oils	[74]
			Alkaloids, Flavonoids, Phenols,	[108

Family	Genus	Part Used/Type Extraction	Compounds	Ref
Lycopodiaceae	Lycopodium serratum	Aerial parts/Ethanol	Crude etanol extract	[109
Lythraceae	Lawsonia alba	Leaf/Methanol	Coumarin, Flavonoid, Steroid, Tannin, Terpenoid	[11(
	Lawsonia inermis L.	Leaf/Aqueous	Total Phenols, Total Flavonoids, Total Tannins, Saponins	[111
	Punica granatum L.	Fruit whole	Pomegranate are Tannins, Flavonoids, Punicic acid, Phytoestrogen	[112
	Hibiscus rosa sinensis L.	Flowers/Methanol	Phenolic compounds, Flavonoids, Essential Oils, Anthocyanins	[113
Malvaceae	Malva sylvestris	Flowers/Ethanol:Water (80:20)	Total phenolic, Flavonoids, Anthocyanin	[114
	Thespesia populnea L.	Fruit/Aqueous	Glycosides, Flavonoids, Alkaloids, Phytosterol, Quercetin, Rutin, Lupeal	[11
Martyniaccae	Martynia annua	Leaf/Ethanol	Glycosides, Phenols, Flavonoids, Tannins, Anthocyanins MAF-C 7 peaks	[98
	A. indica A. Juss	Steam bark/Water:Ethanol	Crude	[11
Meliaceae	Carapa guianensis Aubl	Andiroba seed oil	Lauric axid, Myristic, Palmitic acid, Stearic acid, Oleic acid, Linoleica cid, Lignoceric acid, Palmitoleic acid, Heptadecanoic acid, Arachidic acid, Behenic acid	[11]
Mimosaceae	Prosopis cineraria	Leaves/Petroleum ether	Protocatechuic acid, Caffeic acid, Chlorogenic acid, Ferrulic acid	[11
Moraceae	Ficus religiosa L.	Leaves/Methanol	Glycosides, Alkaloids, Tannins, Terpenoids	[119
Moringaceae	Moringa oleífera Lam.	Leaves/Ethanol	Flavonoids, Phenolic acids	[12
Musaceae	Musa sapientum L.	Fruits/Ethanol	Saponins, Flavonoids, Glycosides, Steroids, Alkaloids	[12
Myrsinaceae	Embelia ribes Burn.	Fruits/Petroleum ether	Embelin	[12
Myrtaceae	Eucalyptus globulus	Leaves/Hydrodistillation	1,8-cineole content 72.3%, α-pinone 9.4%	[12
Nymphaeaceae	Nelumbo nucifera	Aerial part/Ethanol	30 peaks Ethanol,2-(-Octadecinyloxy, γ-sitosterol, Hexadecanoic acid	[12
Oleaceae	Jasminum auriculatum Vahl.	Leaves/Petroleum ether	Alkaloids, Carbohydrates, Flavonoids, Phenolic compounds, Saponins, Steroids, Tannins, Tepernoids	[12
	Jasminum grandiflorum L.	Leaves/Methanol	Crude	[12
Orchidaceae	Bletilla striata	Root/Boiled water	Polysaccharide content (65.3%)	[12
Paeoniaceae	Paeonia suffruticosa	Bark root/Alcohol	Flavonoids, Phenolic acid, Polysaccharide, Saponins	[12
Papaveraceae	Argemone mexicana L.	Fruits/Methanol	Alkaloids, Flavonoids, Glycosides, Saponins, Steroids, Tannins, Terpenoids	[12
Papilionaceae	Trigonella foenum-graecum	Aerial part/Methanol	Flavonoids	[13

Family	Genus	Part Used/Type Extraction	Compounds	Ref.
Pedaliaceae	Sesamum indicum	Seed/Ethanol	Sesame Oil	[131]
Plantaginaceae	Plantago	Leaves/Distilled water	Polyphenolic compounds	[132]
Polygonaceae	Rheum officinale	Powders/Ethanol	TMC extracts	[133
Potulacaceae	Portulaca grandiflora	Total plant/Ethanol	Alkaloids, Flavonoids, Saponins, Terpenoids	[134
Phyllanthaceae	<i>Bridelia ferruginea</i> Benth.	Leaves/Methanol Stem barks/Ethyl Acetate	High phenolic content High flavonoids content	[135
Rosaceae	Sanguisorba officinalis		Polysaccharide	[136
Rubiaceae	Morinda citrifolia L.	Leaf	Alkaloids, Coumarins, Flavonoids, Saponins, Tannins, Triterpenes	[137
	Rubia cordifolia L.		100 Compounds bicyclic peptides, terpenes, polysaccharides, Flavonoids, Quinones	[138
Rutaceae	Aegle marmelos L.	Flower/Ethanol 60%	Aegelin, Cineol, Cuminaldheyde, Luvangetin, Eugenol	[139]
	Zanthoxylum bungeanum Maxim	140 constituents of this plant	Alkaloids, Fatty acids, Flavonoids, Tepernoids, Flavonoids	[140]
Salicaceae	Casearia sylvestris	Leaves/Hydroalcoholic	Crude extract	[141
Scrophulariaceae	Rehmannia glutinosa		Polysaccharides	[142
Stemonaceae	Stemona tuberosa		9,10-dihydro-5-methoxy-8-methyl-2,7- phenanthrenediol	[143
Theaceae	Camellia sinensis	Tea leaves/Methanol		[144
Thymelaeaceae	Daphne genkwa Sie.		Diterpenoids/yuanhuapine	[145
Vitaceae	Ampelopsis japonica	Root/Methanol	Catechin, Gallic acid, Kaempferol, Euscaphic acid, Resveratrol, Epicatechin	[146
Zingiberaceae	Curcuma longa Linn	Extracts	Alkaloids, Flavonoids, Phenolic, Saponins, Terpenoids, Steroids	[147

5. Bioactive Phytocompounds with Wound-Healing Properties

In the relevant literature, a variety of studies have addressed different plants with wound-healing properties. These studies have described the pharmacological activities of plants employed in wound healing and their molecular mechanisms to validate their traditional use and development into safe and effective herbal treatments for wounds. Due to the plants' metabolism, secondary metabolites can be considered as bioactive molecules with therapeutic potential of great value in the pharmaceutical, cosmetic, and food industries, as concerns the design and formulation of medicines for different illnesses with less severe side effects [148–152]. The bioactive phytochemical compounds found include secondary metabolites such as alkaloids, essential oils, flavonoids, tannins, terpenoids, saponins, and phenolic compounds [153–156] (Figure 2).

The allocation of these active compounds into different plant parts, as has been widely described, involves the use of different selective solvents to derive complex mixtures of groups of metabolites (Figure 3).

Phenolic acids are the bioactive compounds most widely found in legumes, cereals, vegetables, and fruits. They are also responsible for certain characteristics of foods, such as aroma and astringency, as well as color and flavor [153,155–157] (Figure 4).

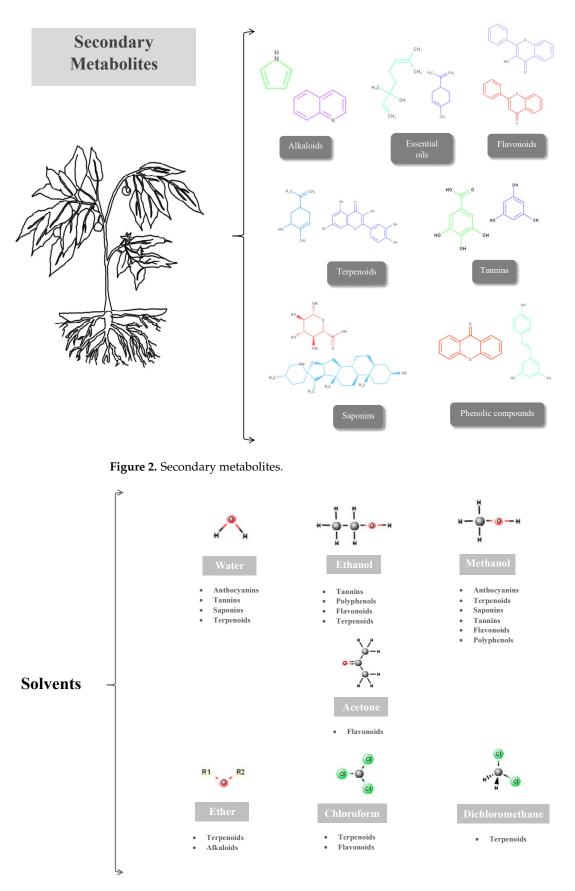


Figure 3. Selective solvents and groups of metabolites.

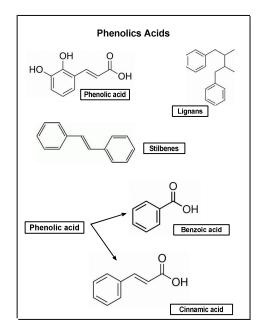


Figure 4. General structure of phenolic acids.

These compounds also play a role in plants' ability to protect themselves against different insults, such as ultraviolet radiation and pathogens [153,156,157]. Importantly, the amounts of phenolic compounds produced by plants can vary according to environmental conditions, genetic factors, and degree of maturation [158]. In the literature, it has been described how phenolic compounds act as anti-inflammatory and antiproliferative agents, antioxidants, transduction modulators, stimulants of collagen production, and antimicrobials, in addition to carrying out other functions [154,156,157]. These compounds can be categorized into hydroxybenzoic acids, such as gallic and vanillic acids, as well as hydroxycinnamic acids such as ferulic and caffeic acids. They have also been shown to have immunomodulatory, antioxidant, hepatoprotective, and anti-inflammatory actions [159] (Figure 5).

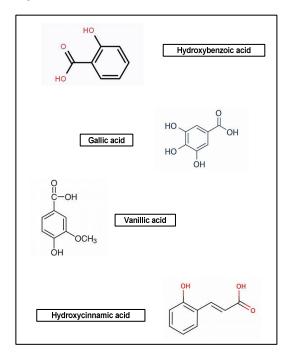
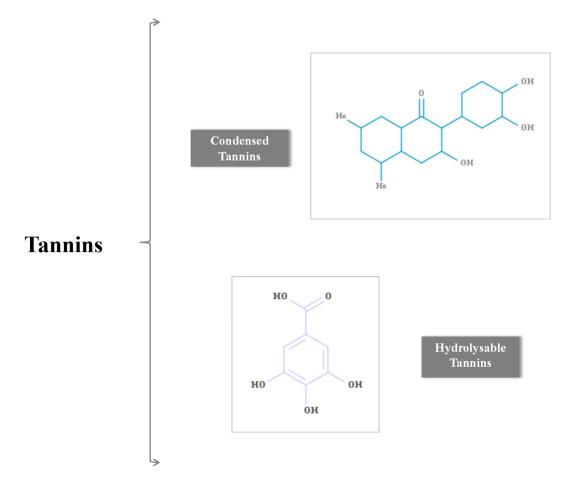
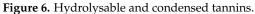


Figure 5. Other phenolic compounds.

Potent antioxidant agents, such as flavonoids, act as reducing agents and protect against radiation [160,161]. These protective effects mean that they can modulate proinflammatory molecules, such as those involved in the healing process [160,162]. The effects of flavonoids in the inflammatory process extend to the treatment of diseases linked to inflammation and processes of which inflammation is a part, such as the healing process and the inhibition of invasion, angiogenesis, and metastasis mechanisms [160].

Tannins are the most complex of the phenolic compounds, categorizable as condensates or hydrolysable. Their role is to protect plants from pathogens through protein complexation, and via their antimutagenic activity, they promote healing through the modulation of different cellular mechanisms and growth factors [153,156,157] (Figure 6).





6. Activity of Bioactive Phytochemicals in Wound Healing

Impaired vascular function, ischemia, superficial debris, and necrosis are the main factors that cause poor immune responses and, consequently, contribute to the development of continued chronic wounds. Excessive bacterial growth and the formation of a biofilm lead to a chronic and self-perpetuating inflammatory state via the modification of aspects of the wound microenvironment, such as its humidity, pH, metalloproteinases, and reactive oxygen species. As many of these microenvironment-related factors as possible must be taken into account to develop beneficial therapeutic strategies [163]. Nature, as described in the literature, is a rich source of therapeutic possibilities. Secondary metabolites can promote the wound-healing process through their pharmacological effects on the body. These compounds include phenolics, alkaloids, and fatty acids, as well as glycosylates and polysaccharides. Such compounds have also been confirmed to have beneficial effects related to their anti-inflammatory, antioxidant, and antibacterial properties, and they promote collagen synthesis and facilitate protective cell regeneration [164–166]. In addition,

these active compounds present low toxicity and good absorption by the skin barrier [164]. The improved efficiency of treatments using natural extracts is related to the establishment of synergy, which enhances the effects of products of natural origin as well as current therapeutic approaches. Various studies have demonstrated that such synergistic interaction is a result of these substances' antibacterial, antioxidant, and anti-inflammatory properties [167]. Active research in this area is currently focused on developing wound treatments able to prevent microorganisms from entering wounds with a bactericidal effect. Recent studies have shown that the use of vegetal extracts and their secondary metabolites has been integrated into diverse treatment modalities, and this has been proven to be effective against both Gram-positive and Gram-negative bacteria [168]. Some have already been selected for use in clinical trials or incorporated into nanoparticles [169]. Studies have shown that natural metabolites can represent beneficial candidates for use in wound healing. One obstacle in developing their clinical use is their poor oral or topical bioavailability.

6.1. Essential Oils

Research has shown that volatile essential oils present a variety of beneficial properties, such as antioxidant, antiviral, anticancer, insecticidal, anti-inflammatory, antiallergic, and antimicrobial effects [168]. These mixtures of lipophilic components are considered safe and biocompatible, although due to their low water solubility, bioavailability, and stability, their therapeutic uses can be limited [169].

6.2. Polyphenols

Polyphenols are considered multifaceted agents due to their beneficial activities, such as antibacterial, anticancer, anti-inflammatory, and antioxidant effects, in addition to their complex wound-healing properties [170]. However, the main problems include their hydrophobicity and poor water solubility, permeability, and bioavailability.

6.3. Flavonoids

As an exemplary flavonoid, quercetin has been harnessed for its antibacterial, antiinflammatory, and antioxidant activities. When converted into quercetin nanofibers, it provides a large porous surface area that can carry many active compounds that facilitate penetration into the skin. Trials conducted with quercetin patches have shown them to have antibacterial activity that combats acne [171]. In other trials, film structures of N-carboxybutyl chitosan (CBC) and agarose were analyzed for their potential utilization in topical membranous wound treatment. Other research has demonstrated the use of polymeric biomaterials loaded with quercetin and thymol. These have been utilized both individually and in the form of mixtures of these two substances, which have antiinflammatory and anesthetic properties. The incorporation of quercetin into semisolid bases such as creams and acid carbomer gels has been proposed to investigate the effects of additives such as propylene glycol and polyethylene glycol on its release and skin retention. With respect to quercetin and chrysin, or quercetin within chitosan nanoparticles, propylene glycol is an absorption accelerator that can also prolong the antioxidant activity [172,173]. Another study has demonstrated that polymeric nanoparticles can enhance antiradical activity, along with chelating quercetin and catechin [174]. Other studies have demonstrated the additional benefits offered by apigenin to the skin via the stimulation of epidermal differentiation, the synthesis and secretion of lipids, and cutaneous antimicrobial production. In vitro studies have demonstrated that hesperidin and naringin obtained from citrus fruits can be used to synthesize stabilized nanoparticles in a green manner [174].

Phytochemicals have been described to enhance the effects of antibiotics due to their low toxicity and anti-infective, anti-inflammatory, and antioxidant properties [175]. They can act as efflux pump inhibitors, preventing biofilm formation or targeting specific bacterial virulence factors [175]. Research confirms that plants from different families can facilitate the healing process and attenuate inflammation [176–178]. Certain compounds, including the flavonoid baicalein and the monoterpene phenol thymol, have an inhibitory effect on

inflammation that has been demonstrated in mixtures of ethanol and can act synergistically, suggesting their use as an alternative treatment to antibiotics [176–178].

7. Mechanisms of Effects of Phytochemicals on Wound-Healing Agents

7.1. Antioxidant Activities of Wound-Healing Agents

Large amounts of energy must be produced for normal cellular activities, which is achieved through mechanisms such as oxidation, resulting in the generation of reactive oxygen species (ROS) and reactive nitrogen species (RNS) [156]. These reactive species possess unpaired electrons in their valence shell and are unstable [154]. Radical species include hydroxyl radicals (OH⁻), nitric oxide radicals (NO), singlet oxygen (¹O₂), and superoxide radicals (O₂⁻). These are produced naturally in the body, but adverse factors such as stress and pollution can increase their levels, causing them to damage molecules such as proteins and DNA, leading to the disintegration of cell membranes. Oxidative stress is strongly linked to the development of chronic diseases and aging [154,157,160].

The use of antioxidants to control the levels of reactive species in the body is recommended. Antioxidants are defined as substances with the capacity to control the oxidation of biomolecules and act in the sequestration of reactive species such as ROS or RNS; some can also chelate metal ions and modulate enzymes related to oxidative stress [154,156]. Such enzymes include catalase (CAT), which catalyzes the degradation of hydrogen peroxide (H₂O₂), and glutathione peroxidase (GPx), which removes hydroperoxides [154].

Non-enzymatic processes involve transferrin, reduced glutathione (GSH), ubiquinol, and melatonin [154,156]. The antioxidant effects of these compounds are related to the presence of phenolic compounds, amino acids, sterols, ascorbic acid, peptides, and phospholipids in their composition [179]. Several studies have shown that antioxidants have anti-inflammatory, vasodilatory, antitumor, antiallergic, antiviral, and cardioprotective activities, among other properties [156,157,179].

During the healing process, excess free radicals are produced at the site of injury. This can be limited by the presence of antioxidants, which prevent some of the damage caused to cells [180]. The antioxidant action of medicinal plants is strongly related to the quantities of bioactive compounds they contain, such as flavonoids, which act as antioxidants and also directly participate in the inflammatory phase, limiting cellular damage due to their effects on prostaglandins and macrophages [181]. Flavonoids are also capable of increasing the resistance of collagen fibers, thus facilitating the process of the contraction and re-epithelization of wounds [180,182].

The healing process restores tissue integrity when an injury occurs [183]. It can be impeded by factors such as diabetes, which causes it to be slower and less efficient, thus potentially causing chronicity [184,185]. Poor healing can lead to the loss of tissue function, the chronification of injuries, and amputation, and it can also produce physical, psychological, social, and economic damage [184]. The treatments used to promote healing include the use of natural products and their derivatives. Some of the medications currently available are not completely effective in treating chronic wounds. For this reason, it is essential to continue research into new substances with more effective healing properties. Flavonoids and tannins have shown antiproliferative properties and are capable of regulating the production of free radicals; they are also involved in limiting inflammatory mechanisms [180]. Further investigations in the pharmaceutical, food, and cosmetic sectors will be essential in addressing the sources of antioxidants and substances that can be used to treat certain conditions, such as chronic wounds and cancer. Following the formation of ROS, the wound-healing process is significantly delayed; however, their formation is limited by the presence of flavonoids, which are responsible for increasing the levels of common antioxidant enzymes. The use of flavonoids in the clinical setting is very limited due to their low bioavailability. An important property of flavonoids obtained from plants is their lipophilicity against Gram-positive bacteria, which is a product of their involvement in the damage done to the respiratory chain and other aspects [186].

7.2. Anti-Inflammatory Properties of Wound-Healing Agents

Flavonoids have also been suggested as a candidate for use in the treatment of a variety of skin lesions, with minimal side effects when administered by topical application due to their lipophilic nature [187]. The many properties exhibited by flavonoids, such as their anti-inflammatory, antimicrobial, and antifibrotic effects, can be understood as a result of their polyhydroxy structure. Among all the structurally different flavonoids, twenty-four have demonstrated the ability to accelerate the healing process, and the most studied are quercetin, epigallocatechin gallate, and naringenin [55] (Figure 7).

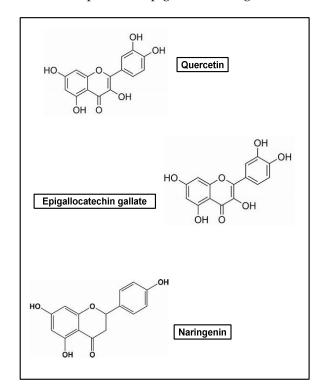


Figure 7. Most widely studied flavonoids.

Numerous studies have also shown that flavonoids are capable of decreasing the levels of inflammatory mediators, such as prostaglandins and leukotriene, and pro-inflammatory cytokines, such as IL-1 β , TNF- α , IL-6, and IFN- γ . They can also increase the production of anti-inflammatory mediators, such as interleukin 10 (IL-10), negatively regulate the expression of nuclear factor kappa B (NF- κ B), and block cyclooxygenase activity.

Prenylated flavonoids are found in plants' roots, bark, seeds, and buds. These are part of a subclass of modified flavonoids with at least one lipophilic side chain of variable length, and they possess favorable biological activities, such as antimicrobial, antifungal, larvicidal, estrogenic, osteogenic, immunosuppressive, anticancer, anti-inflammatory, antioxidant, antiallergic and cytotoxic effects [188]. The group of prenylated flavonoids includes Cprenylated chalcones/dihydrochalcones, flavanones, flavones, flavonols, isoflavones, and, less frequently, O-prenylated forms (Figure 8).

These structures can be replaced, following oxidation, reduction, dehydration, and/or cycling, with 3,3-dimethylallyl, 1,1-dimethylallyl, geranyl, lavandulyl, and farnesyl side chains [188]. Studies have shown the advantages offered by prenyl compared to flavonoids. Prenylated flavonoids have a greater affinity with the cellular membrane and P-glycoprotein inhibitors [189] and show antibacterial and inhibitory or enzyme-enhancing actions, while prenylation causes an increase in lipophilicity and the affinity for biological membranes [188,189].

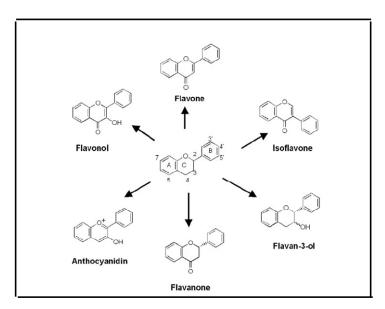


Figure 8. Prenylated flavonoids.

Diplacone, with its 6-geranyl-30,40,5,7-tetrahydroxyflavanone structure, has shown anti-inflammatory properties both in vitro and in vivo, with different mechanisms of action. It can cause reductions in TNF- α and MCP-1 expression and regulates the expression of zinc-finger protein 36, which increases cytokine degradation [190]. Another compound, isobavachalcone, suppresses the production of nitric oxide and negatively regulates inflammation-related enzymes such as iNOS and 15-LOX [191,192]. Licochalcone A is a 5-(2-methylbut-3-en-2-yl) chalcone obtained from licorice roots, and it has been traditionally used to treat inflammatory diseases. It inhibits the activation of transcription factors such as NF- κ B and AP-1; it also suppresses pro-inflammatory cytokines and NO and PGE2 production [193,194]. The main function of Sophoraflavanone G is the inhibition of eicosanoid-forming enzymes [195]. It can also disrupt NF-kB and MAPK signaling pathways [195,196]. Another prenylated chalcone is xanthohumol, which is found in *Humulus lupulus* L. hops and has antistaphylococcal activity [197,198]. Its anti-inflammatory effect is enacted through the inhibition of NO levels due to the suppression of inducible NO synthase, and it inhibits both the activation of NF- κ B [199,200] and the production of the cytokines MCP-1, TNF- α , and IL-12, as well as oxidative stress [201–203].

7.3. Antimicrobial Effects of Wound-Healing Agents

In the previous sections, we mention that flavonoids are widely used as effective therapeutic agents and that numerous in vitro and in vivo studies have confirmed them to have important functions, mainly defensive and regulatory [203]. Regarding their functions as protective agents against microorganisms, flavonoids act directly on bacterial cells, as well as suppressing virulence and the formation of biofilms. They can also act synergistically with antibiotics [204]. These properties have enabled the production and use of semisynthetic or synthetic flavonoids to combat microorganisms [205].

The antibacterial activities of flavonoids and prenylated flavonoids are due to the structure of 2-phenyl-1,4-benzopyrone, which has been suggested to be capable of influencing different cellular processes [204]. Apigenin and quercetin present the ability to inhibit bacterial cell walls by inhibiting D-alanine–D-alanine (D-Ala–D-Ala) ligase, which is crucial to the completion of peptidoglycan precursors [206]. Researchers have indicated that several flavonoids can modify membrane permeability and damage membrane functions. On the other hand, flavanols, flavolans, and green tea catechins have been shown to disturb bacterial cytoplasmic membranes through hydrogen peroxide [207,208]. Another flavone, Artocarpin, obtained from the Moraceae family, with prenyl in position 3

and a (1E)-3-methylbut-1-enyl moiety in position 6, presented remarkable antibacterial activity [197,209–211].

8. Conclusions

Plants are excellent wound healers, and when used in the context of different wound models, they can be employed as part of proper measures to treat wounds and control the healing process. Thus, herbal medicines have gained popularity in several countries. The factors that must be considered in the healing of a lesion are the wound closure rate, epithelialization, tensile strength, histopathology, and granuloma weight. This study discusses how traditional medicines could play important roles in wound healing. Modern knowledge of these bioactive principles can provide alternatives to improve or accelerate wound healing with minimal toxicity. The preliminary evidence and results in the current literature suggest that this is an active area of study. In future studies, factors such as the potential toxicity to human cells, kinetics and speed of healing, wound types, chronicity, timing of application, and dose of therapeutic agent must be considered. The preparation of formulations that include medicinal plants as part of their release and distribution systems for their anti-inflammatory, antioxidant, and wound-healing properties requires further investigation. These proposed studies on natural or synthetic formulations can be achieved by acquiring certain quantities of pure compounds and their extracts for standardization.

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References

- 1. Strodtbeck, F. Physiology of wound healing. Newborn Infant. Nurs. Rev. 2001, 1, 43–52. [CrossRef]
- 2. Young, A.; McNaught, C.E. The physiology of wound healing. *Surgery* **2011**, *29*, 475–479. [CrossRef]
- 3. Singer, A.J.; Clarck, R.A.F. Cutaneous wound healing. N. Engl. J. Med. 1999, 341, 738–746. [CrossRef] [PubMed]
- 4. Broughton, G.; Janis, J.E.; Attinger, C.E. The basic science of wound healing. *Plast. Reconstr. Surg.* 2006, 117 (Suppl. 7), S12–S34. [CrossRef] [PubMed]
- Andrade, M.G.L. Alterações do processo de cicatrização de queimaduras em indivíduos diabéticos. *Rev. Bras. Queimaduras* 2013, 12, 42–48.
- Brem, H.; Tomic-Canic, M. Cellular and molecular basis of wound healing in diabetes. J. Clin. Investig. 2007, 117, 1219–1222. [CrossRef] [PubMed]
- 7. Isaac, C. Processo de cura das feridas: Cicatrização fisiológica. Rev. Med. 2010, 89, 125–131. [CrossRef]
- 8. Mandelbaum, S.H.; Di Santis, E.P.; Mandelbaum, M.H.S.A. Cicatrization: Current concepts and auxiliary resources—Parte I. *An. Bras. Dermatol.* **2003**, *72*, 393–410. [CrossRef]
- 9. Mallefet, P.; Dweck, A.C. Mechanism of Wound Healing Examined. Personal. Care 2008, 9, 75–83.
- 10. Pober, J.S.; Sessa, W.C. Evolving functions of endothelial cells in inflammation. Nat. Rev. Immunol. 2007, 7, 803-815. [CrossRef]
- 11. Kumar, V.; Abbas, A.K.; Aster, J.C. Robbins Patologia Básica, 9th ed.; Elsevier: Rio de Janeiro, Brazil, 2013; p. 928.
- 12. Lamano, T.L.C. Patologia Geral. Inflamação; Universidade de São Paulo: Ribeirão Preto, Brazil, 2008.
- 13. Rodero, M.P.; Khosrotehrani, K. Skin wound healing modulation by macrophages. Int. J. Clin. Exp. Pathol. 2010, 3, 643–653.
- 14. Ross, R.; Odland, G. Human wound repair. II. Inflammatory cells, epithelial-mesenchymal interrelations, and fibrogenesis. *J. Cell Biol.* **1968**, *39*, 152–168. [CrossRef] [PubMed]

- Kim, M.H.; Liu, W.; Borjesson, D.L.; Curry, F.-R.E.; Miller, L.S.; Cheung, A.L.; Liu, F.-T.; Isseroff, R.R.; Simon, S.I. Dynamics of neutrophil infiltration during cutaneous wound healing and infection using fluorescence imaging. *J. Investig. Dermatol.* 2008, 128, 1812–1820. [CrossRef] [PubMed]
- Deppermann, C.; Cherpokova, D.; Nurden, P.; Schulz, J.-N.; Thielmann, I.; Kraft, P.; Vögtle, T.; Kleinschnitz, C.; Dütting, S.; Krohne, G.; et al. Gray platelet syndrome and defective thrombo-inflammation in Nbeal2-deficient mice. *J. Clin. Investig.* 2013, 123, 3331–3342. [CrossRef] [PubMed]
- 17. Mendonça, R.J.; Coutinho-Neto, J. Aspectos celulares da cicatrização. An. Bras. Dermatol. 2009, 84, 257–262. [CrossRef]
- Waldrop, J.; Doughty, D. Wound-healing Physiology. In Acute and Chronic Wounds: Nursing Management; Mosby Inc.: London, UK, 2000; pp. 17–39.
- 19. Enoch, S.; Leaper, D. Basic science of wound healing. Surgery 2007, 26, 31–37.
- 20. Eckes, B.; Nischt, R.; Krieg, T. Cell-matrix interactions in dermal repair and scarring. *Fibrogenesis Tissue Repair.* **2010**, *3*, 4. [CrossRef]
- 21. Reinke, J.M.; Sorg, H. Wound Repair and Regeneration. Eur. Surg. Res. 2012, 49, 35–43. [CrossRef]
- Waldrop, M.A.; Kolb, S.J. Current treatment options in neurology-SMA therapeutics. *Curr. Treat Options Neurol.* 2019, 21, 25. [CrossRef]
- 23. Eming, S.A.; Brachvogel, B.; Odorisio, B.; Koch, M. Regulation of angiogenesis: Wound healing as a model. *Prog. Histochem. Cytochem.* **2007**, *42*, 115–170. [CrossRef]
- 24. Beldon, P. Basic science of wound healing. Surgery 2010, 28, 409–412. [CrossRef]
- 25. Delavary, B.M.; van der Veer, W.M.; van Egmond, M.; Niessen, F.B.; Beelen, R.H. Macrophages in skin injury and repair. *Immunobiology* **2011**, *216*, 753–762. [CrossRef] [PubMed]
- Galiano, R.D.; Michaelis, J.; Dobryansky, M.; Levine, J.P.; Gurtner, G.C. Quantitative and reproducible murine model of excisional wound healing. *Wound Rep. Regen.* 2004, 12, 485–492. [CrossRef]
- 27. Galkowska, H.; Wojewodzka, U.; Olszewski, W.L. Chemokines, cytokines, and growth factors in keratinocytes and dermal endothelial cells in the margin of chronic diabetic foot ulcers. *Wound Rep. Regen.* **2006**, *14*, 558–565. [CrossRef]
- Eo, H.; Lee, H.J.; Lim, Y. Ameliorative Effect of Dietary Genistein on Diabetes Induced Hyper-Inflammation and Oxidative Stress During Early Stage of Wound Healing in Alloxan Induced Diabetic Mice. *Biochem. Biophys. Rev. Commun.* 2016, 478, 1021–1027. [CrossRef] [PubMed]
- 29. Park, J.H.; Kim, S.; Hong, H.S.; Son, Y. Substance P promotes diabetic wound healing by modulating inflammation and restoring cellular activity of mesenchymal stem cells. *Wound Rep. Reg.* 2016, 24, 337–348. [CrossRef] [PubMed]
- Romana-Souza, A.P.; Nascimento, A.P.; Monte-Alto-Costa, A. Propranolol improves cutaneous wound healing in streptozotocininduced diabetic rats. *Eur. J. Pharmacol.* 2009, 611, 77–84. [CrossRef]
- Kant, V.; Gopal, A.; Pathak, N.N.; Kumar, P.; Tandan, S.K.; Kumar, D. Antioxidant and anti-inflammatory potential of curcumin accelerated the cutaneous wound healing in streptozotocin-induced diabetic rats. *Int. Immunopharmacol.* 2014, 20, 322–330. [CrossRef] [PubMed]
- 32. Kant, V.; Gopal, A.; Kumar, D.; Pathak, N.N.; Ram, M.; Jangir, B.L.; Tandan, S.K.; Kumar, D. Curcumin-induced angiogenesis hastens wound healing in diabetic rats. *J. Surg. Res.* 2015, 193, 978–988. [CrossRef]
- 33. WHO Implementation of WHO Traditional Medicine Strategy 2014–2023; Bolletin; WHO: Geneva, Switzerland, 2013; ISBN 9789241506090.
- Alvim, N.A.T.; Ferreira, M.D.A.; Cabral, I.E.; Almeida Filho, A.J.D. O uso de plantas medicinais como recurso terapêutico: Das influências da formação profissional às implicações éticas e legais de sua aplicabilidade como extensão da prática de cuidar realizada pela enfermeira. *Rev. Lat.-Am. Enferm.* 2006, 14, 316–323. [CrossRef] [PubMed]
- 35. Carvalho, A.C.B.; Silveira, D. Drogas vegetais: Uma antiga nova forma de utilização de plantas medicinais. *Brasília Méd.* **2010**, *48*, 219–237.
- 36. Silva, D.M.; Mocelin, K.R. O cuidado de enfermagem ao cliente portador de feridas sob a ótica do cuidado transcultural. *Nursing* **2007**, *9*, 8188.
- Aquino, D.; Silva, R.B.L.D.; Gomes, V.F.; Araújo, E.C.D. Nível de conhecimento sobre riscos e benefícios do uso de plantas medicinais e fitoterápicos de uma comunidade do Recife—PE. *Rev. Enferm. UFPE Online* 2007, 1, 107–110. [CrossRef]
- 38. Sund, B. New Developments in Wound Care; Clínica Report; PJB Publications: New York, NY, USA, 2000; pp. 1–255.
- 39. Wanda, A.; Dorsette, M. Rat models of skin wound healing: A review. Wound Rep. Regen. 2004, 12, 591–599.
- 40. Jurjus, A.; Atiyeh, B.; Abdallah, I.; Jurjus, R.; Hayek, S.; Jaoude, M.; Gerges, A.; Tohme, R. Pharmacological modulation of wound healing in experimental burns. *Burns* **2007**, *3*, 892–907. [CrossRef]
- 41. Agyare, C.; Bempah, S.B.; Boakye, Y.D.; Ayande, P.G.; Adarkwa-Yiadom, M.; Mensah, K.B. Evaluation of antimicrobial and wound healing potential of *Justicia flava* and *Lannea welwitschii. Evid. Based Complement. Altern. Med.* **2013**, 2013, 632927. [CrossRef]
- 42. Al-Bayaty, F.H.; Abdulla, M.A.; Hassan, M.I.A.; Ali, H.M. Effect of *Andrographis paniculata* leaf extract on wound healing in rats. *Nat. Prod. Res.* 2012, *26*, 423–429. [CrossRef]
- Mengie, T.; Mequanente, S.; Nigussie, D.; Legesse, B.; Makonnen, E. Investigation of wound healing and anti-inflammatory activities of 80% methanol leaf extract of *Achyranthes aspera* L. (Amaranthaceae) in rats. *J. Inflamm. Res.* 2021, 14, 1775–1787. [CrossRef]

- Muniandy, K.; Gothai, S.; Sean Tan, W.; Suresh Kumar, S.; Mohd Esa, N.; Chandramohan, G.; Al-Numair, K.; Arulselvan, P. In Vitro Wound Healing Potential of Stem Extract of *Alternanthera sessilis*. *Evid. Based Complement. Altern. Med.* 2018, 2018, 3142073. [CrossRef]
- 45. Kallappa, M.H.; Ganjihal, S.S.; Chavadi, D.V. *Alternanthera triandra* seed oil: A moderate source of ricinoleic acid and its possible industrial utilization. *Ind. Crops Prod.* 2004, 2, 133–136.
- 46. Ofusori, A.E.; Raharjo, Y.; Ofusori, D.A.; Adekunle, V.O. A comparative study of dichloromethane and ethyl acetate root extracts of *Celoisa trigyna*: Phytochemical and wound healing effects analyses. *J. Wound Manag. Res.* **2023**, *19*, 87–95. [CrossRef]
- Pattnaik, A.; Sarkar, R.; Sharma, A.; Yadav, K.K.; Kumar, A.; Roy, P.; Sen, T. Pharmacological studies on *Buchanania lanzan* Spreng.-A focus on wound healing with particular reference to anti-biofilm properties. *Asian Pac. J. Trop. Biomed.* 2013, *3*, 967–974. [CrossRef] [PubMed]
- 48. Hsiao, C.Y.; Hung, C.-Y.; Tsai, T.-H.; Chak, K.-F. A Study of the wound healing mechanism of a traditional Chinese medicine, *Angelica sinensis*, using a proteomic approach. *Evid.-Based Complement. Altern. Med.* **2012**, 2012, 467531. [CrossRef] [PubMed]
- Tanga, B.M.; Bang, S.; Fang, X.; Seo, C.; de Zoysa, M.; Saadeldin, M.I.M.; Lee, S.; Park, S.; Chung, S.-O.; Lee, G.-J.; et al. *Centella* asiatica extract in carboxymethyl cellulose at its optimal concentration improved wound healing in mice model. *Heliyon* 2022, 8, e12031. [CrossRef] [PubMed]
- 50. Gohari, A.R.; Saeidnia, S.A. Review on Phytochemistry of *Cuminum cyminum* seeds and its standards from field to market. *Pharmacogn. J.* **2011**, *3*, 1–5. [CrossRef]
- 51. Wu, J.-G.; Wei, Y.-J.; Ran, X.; Zhang, H.; Nian, H.; Qin, L.-P. Inhibitory effects of essential oil from rhizomes of *Ligusticum chuanxiong* on hypertrophic scarring in the rabbit ear model. *Pharm. Biol.* **2011**, *49*, 764–769. [CrossRef] [PubMed]
- 52. Kabesh, K.; Senthulkumar, P.; Ragunathan, R.; Kumar, R. Phytochemical análisis of *Catharantus roseus* plan extract and its anti-microbial acitivity. *Int. J. Pure App. Biosci.* 2015, *3*, 162–172.
- Agyare, C.; Dwobeng, A.S.; Agyepong, N.; Boakye, Y.D.; Mensah, K.B.; Ayande, P.G.; Adarkwa-Yiradon, M. Antimicrobial, antioxidant, and wound healing properties of *Kigelia africana* (Lam.) Beneth. and *Strophanthus hispidus* DC. *Adv. Pharmacol. Sci.* 2013, 2013, 692613. [CrossRef] [PubMed]
- Meenu, N.C.; Manokari, S.L.; Yogeswari, G.; Duraisami, R. Analysis of phytochemical constituents and antibacterial activity of Wrightia tinctoria: Traditional medicinal plant of India for application on wound dressing materials. Indian J. Trad. Knowl. 2022, 21, 48–54.
- 55. Omale, J.; Victoria, I.A. Excision and incision wound healing potential of *Saba florida* (Benth) leaf extract in *Rattus novergicus*. *Int. J. Pharm. Biomed. Res.* **2010**, *1*, 101–107.
- 56. Namgoong, S.; Lee, H.; Han, S.-K.; Lee, H.-W.; Jeong, S.-H.; Dhong, E.-S. Effect of Panax ginseng extract on the activity of diabetic fibroblasts in vitro. *Int. Wound J.* 2019, *16*, 737–745. [CrossRef]
- 57. Lu, T.; Gao, Y.; Duan, Y.; Cui, C.; Zhang, L.; Mingning, S. *Panax notoginseng* saponins improves healing of high glucose-induced wound through the GSK-3β/β-catenin pathway. *Environ. Toxicol.* **2022**, *37*, 1867–1877.
- 58. Ali-Seyed, M.; Ayesha, S. *Calotropis* A multi-potential plant to human kind: Special focus on its wound healing efficacy. *Biocatal. Agric. Biotechnol.* **2020**, *28*, 101725. [CrossRef]
- 59. Mali, R.P.; Raq, P.S.; Vikle, D.N. Wound healing activity of *Calotropis procera* root bark on diabetic rats. *J. Drug Deliv. Ther.* **2020**, 10, 86–89. [CrossRef]
- Garcia-Orue, I.; Gainza, G.; Gutierrez, F.B.; Aguirre, J.J.; Evora, C.; Pedraz, J.L.; Hernandez, R.M.; Delgado, A.; Igartua, M. Novel nanofibrous dressings containing rhEGF and *Aloe vera* for wound healing applications. *Int. J. Pharm.* 2016, 523, 556–566. [CrossRef] [PubMed]
- Tadić, V.; Arsić, I.; Zvezdanović, J.; Zugić, A.; Cvetković, D.; Pavkov, S. The Estimation of the Traditionally Used Yarrow (*Achillea millefolium* L. Asteraceae) Oil Extracts with Anti-Inflamatory Potential in Topical Application. *J. Ethnopharmacol.* 2017, 199, 138–148. [CrossRef] [PubMed]
- Nascimento, B.A.C.; Gardinassi, L.G.; Silveira, I.M.G.; Galucci, M.G.; Tomre, M.A.; Oliveira, J.F.D.; Moreira, M.R.A.; Meirelles, A.F.G.; Faccioli, L.H.; Tefé-Silva, C.; et al. *Arctium lappa* extracts suppresses inflammation and inhibits melanoma progression. *Medines* 2019, 6, 81. [CrossRef]
- 63. Pang, Y.; Wang, D.; Hu, X.; Wang, H.; Fu, W.; Fan, Z.; Chen, X.; Yu, F. Effect of volatile oil from *Blumea balsamifera* (L.) DC. leaves on wound healing in mice. *J. Tradit. Chin. Med.* **2014**, *34*, 716–724. [CrossRef] [PubMed]
- Dinda, M.; Mazumdar, S.; Das, S.; Ganguly, D.; Dasgupta, U.B.; Dutta, A.; Jana, K.; Karmakar, P. The water fraction of *Calendula* officinalis hydroethanol extract stimulates in vitro and in vivo proliferation of dermal fibroblasts in wound healing. *Phytother. Res.* 2016, 30, 1696–1707. [CrossRef] [PubMed]
- 65. Parente, L.M.; Lino Júnior, R.d.S.; Faustino Tresvenzol, L.M.; Vinaud, M.C.; de Paula, J.R.; Paulo, N.M. Wound healing and anti-inflammatory effect in animal models of *Calendula officinalis* L. growing in Brazil. *Evid.-Based Complement. Altern. Med.* 2012, 2012, 375671. [CrossRef]
- 66. Gao, S.-Q.; Chang, C.; Niu, X.-Q.; Li, L.-J.; Zhang, Y.; Gao, J.-Q. Topical application of Hydroxysafflor yellow A accelerates the wound healing in streptozotocin induced T1DM rats. *Eur. J. Pharmacol.* **2018**, *823*, 72–78. [CrossRef]
- 67. Balekar, N.; Nakpheng, T.; Katkam, N.G.; Srichana, T. *Wedelia trilobata* L. Phytochemical and Pharmacological Reviews. *Chiang Mai J. Sci.* **2014**, *41*, 590–605.

- 68. Agyare, G.; Koffuor, A.; Boamah, V.E.; Adu, F.; Mensah, K.B.; Adu-Amoah, L. Antimicrobial and anti-inflammatory activities of *Pterygota macrocarpa* and *Cola gigantea* (Sterculiaceae). *Evid.-Based Complement. Altern. Med.* **2012**, 2012, 902394. [CrossRef]
- Kuklkarni, M.; Singhai, R.G.; Bhise, K.; Tambe, R. Phytochemical screening, HPTLC studies screening of antioxidants activity of extracts of leaves of *Spanthodea campanulate*. J. Pharmacogn. Phytochem. 2014, 3, 8–13.
- 70. Elshamy, A.I.; Ammar, N.M.; Hassan, H.A.; El-Kashak, W.A.; Al-Rejaie, S.S.; Abd-ElGawad, A.M.; Farrag, A.-R.H. Topical Wound Healing Activity of Myricetin Isolated from *Tecomaria capensis* v. *aurea*. *Molecules* **2020**, 25, 4870. [CrossRef]
- 71. Nakuntwalai, W.; Thungmungmee, S.; Khobjai, W. Factor promoting wound healing: Radical scavenging and anti-inflammatory activity and growth factor promotion of *Heliotropium indicum*. *Int. J. Appl. Pharm.* **2019**, *11*, 44–48.
- 72. Kojoma, M. Cultivation study of *Lithospermum erythrorhizon* to obtain "Shikon" as a purple dye and traditional medicine–root growth and shikonin derivatives content. In Proceedings of the ISHS Acta Horticulturae 1361: XXXI International Horticultural Congress (IHC2022): International Symposium on Natural Colorants from Plants, Angers, France, 14–20 August 2022; Räisänen, R., de la Sayette, A., Eds.; 2022.
- Kavousi, A.; Nikkhah, E.; Tayarani-Najaran, Z.; Javadi, B. Wound Healing Effects and Related Mechanims of Action of Methanol Extracts of *Boswellia sacra* and *Commiphora myrrha* Oleo-Gum Resins on Adult Human Dermal Fibroblasts (HDFa). Available online: https://ssrn.com/id3969804 (accessed on 9 January 2024).
- Yuan, X.; Han, L.; Fu, P.; Zeng, H.; Lv, C.; Chang, W.; Runyon, R.S.; Ishii, M.; Han, L.; Liu, K.; et al. Cinnamaldehyde accelerates wound healing by promoting angiogenesis via up-regulation of PI3K and MAPK signaling pathways. *Lab. Investig.* 2018, 98, 783–798. [CrossRef]
- 75. Koshak, A.; Algandaby, M.N.; Mujallid, M.I.; Abdel-Naim, A.B.; Alhakamy, N.A.; Fahmy, U.A.; Alfarsi, A.; Badr-Eldin, S.M.; Neamatallah, T.; Nasrullah, M.Z.; et al. Wound healing activity of *Opuntia ficus-indica* fixed oil formulated in a self-nanoemulsifying formulation. *Int. J. Nanomed.* 2021, 16, 3889–3905. [CrossRef] [PubMed]
- Fanani Hakim, R.; Dinni, F. Effect of Carica papaya extract toward incised wound healing process in mice (Mus musculus) clinically and histologically. Evid.-Bases Complemment. Altern. Med. 2019, 2019, 8306519. [CrossRef]
- 77. Quillay Davila, M.A.; Arana Arias, Y.A.; Jaramillo Jaramillo, C.G.; Buelle, S.C.; Rojas de Astudillo, L.L.; Jaramillo Alcívar, V. Contenido de saponinas y actividad cicatrizante de *Cecropia peltata y Parthenium hysterophorus*. *Rev. Cuba. Farm.* 2018, 51. Available online: http://www.revfarmacia.sld.cu/index.php/far/article/view/250/147 (accessed on 9 January 2024).
- 78. Agyare, C.; Ansah, A.O.; Ossei, P.P.S.; Apenteng, J.A.; Boakye, Y.D. Wound healing and anti-infective properties of *Myrianthus arboreus* and *Alchornea cordifolia*. *Med. Chem.* **2014**, *4*, 533–539. [CrossRef]
- 79. Chen, W.C.; Liou, S.-S.; Tzeng, T.-F.; Lee, S.-L.; Liu, I.-M. Wound repair and anti-inflammatory potential of *Lonicera japonica* in excision wound-induced rats. *BMC Complement. Altern. Med.* **2012**, *12*, 226. [CrossRef] [PubMed]
- Kisseih, E.; Lechtenber, M.; Petereit, F.; Sendker, J.; Brandt, S.; Agyare, C.; Hensel, A. Phytochemical characterization and in vitro wound healing activity of leaf extracts from *Combretum mucronatum* Schum. & Thonn.: Oligomeric procyanidins as strong inductors of cellular differentiation. *J. Ethnopharmacol.* 2015, 174, 628–636.
- Jokar, A.; Masoomi, F.; Sadeghpour, O.; Nassiri-Toosi, M.; Hamedi, S. Potential therapeutic applications for *Terminalia chebula* in Iranian traditional medicine. J. Trd. Chin. Med. 2016, 36, 250–254.
- 82. Khan, M.U.; Khalilullah, H.; Akhtar, J.; Elhasan, G.O. Terminalia chebula: An ephemeral glance. Int. J. Pharm. Phram. 2015, 7, 40-43.
- Rodrigues Dantas Araujo, E.; Costa da Silva, V.; Figueiredo de Lima, J.B.; Schlamb, J.; Freitas Fernandes-Pedrosa, M.; Silva Junior, A.A.; Rathinasabapathy, T.; Moncada, M.; Esposito, D.; Coelho Bernardo Guerra, G.; et al. Gel formulated with *Bryophyllum pinnatum* leaf extract promotes skin wound healing in vivo by increasing VEGF expressión: A novel potential active ingredient for pharmaceuticals. *Front. Pharmacol.* 2023, *13*, 1104705. [CrossRef] [PubMed]
- Ullah Arshad, M.; Hassan, A. Medical treatment of various diseases through Nagarmotha (*Cyperus rotundus*) plant. *Eur. J. Biol. Med. Sci. Res.* 2022, 10, 26–43. [CrossRef]
- 85. Bigonya, P.; Agrawal, S.; Verma, N.K. Potential wound healing activity of *Euphoria hirta* Linn total flavonoids fractions. *Int. J. Pharm. Sci. Rev. Res.* **2013**, 22, 149–156.
- 86. Nwala, C.O.; Akaninwor, J.O.; Monanu, M.O. Phytochemical screening and wound healing activities of extracts of *Jatropha curcas* leaf formulated in a simple ointment base. *Int. J. Eng. Sci. Invent.* **2013**, *2*, 72–75.
- Kabran, F.A.; Okpekon, T.A.; Roblot, F.; Seon-Meniel, T.; Leblanc, K.; Bories, C.; Champy, P.; Yolou, S.F.; Loiseau, P.M.; Djakoure, L.A.; et al. Bioactive phloroglucinols from *Mallotus oppositifolius*. *Fitoterapia* 2015, 107, 100–104. [CrossRef]
- 88. Ajirni, N.A.; Nazaruddin, A.; Sutriana, D.; Masyitha Isa, M. The effect of ethanol of Malacca leaves (*Phyllanthus emblica*) on the number of fibroblast cells in White rats (*Tattus norvegicus*) burns wound. *Int. J. Trop. Vet. Biomed. Res.* **2020**, *5*, 7–12. [CrossRef]
- 89. Boakye, Y.D.; Agyare, C.; Ayande, P.G.; Asiamah, E.A.; Titloye, N.A. Wound healing activity of geraniin and aqueous leaf extract of *Phyllanthus muellerianus* (Kuntze) Excel (Euphorbiaceae). *Planta Med.* **2014**, *80*, WS20. [CrossRef]
- 90. Jeong, S.Y.; Moon, M.Y.; Ryu, E.K.; Lee, J.S.; Chan, J. Identification of the phytochemical compounds and their type I procollagen induction in *Astragalus membranaceus* Sprouts grown under different light conditions. J. Appl. Pharmacetical. Sci. 2018, 8, 1–7.
- 91. Tewtrakul, S.; Tungcharoen, P.; Sudsai, T.; Karalai, C.; Ponglimanont, C.; Yodsaoue, O. Antiinflammatory and wound healing effects of *Caesalpinia sappan* L. *Phytother. Res.* **2015**, *29*, 850–856. [CrossRef]
- 92. Su, X.; Liu, X.; Wang, S.; Li, B.; Pan, T.; Liu, D.; Wang, F.; Diao, Y.; Li, K. Wound-healing promoting effect of total tannins from *Entada phaseoloides* (L.) Merr. In rats. *Burns* 2017, *43*, 830–838. [CrossRef] [PubMed]

- 93. Malekzadeh, M.; Sandoughdaran, S.; Shandiz, F.H.; Honary, S. The efficacy of Licorice root (*Glycyrrhiza glabra*) and yarrow (*Achillea millefolium*) in preventing radiation dermatitis in patients with breast cancer, a randomized, double-blinded, placebo-controlled clinical trial. *Asian Pac. J. Cancer Care* 2016, 1, 9. [CrossRef]
- 94. Saini, S.; Dhiman, A.; Nanda, S. Traditional Indian Medicinal plants with potential wound healing activity: A review. *Int. J. Pharm. Sci. Res.* 2009, *7*, 1809–1891.
- 95. Kannan, S.; Vijay Jesuraj, S.A.; Jeeva Kumar, E.S.; Saminathan, K.; Suthakaran, R.; Saminathan, K.; Suthakaran, R.; Ravi Kumar, M.; Parimala Devi, B. Wound healing activity of *Mimosa pudica* Linn formulation. *Int. J. PharmTech. Res.* **2009**, *1*, 1554–1558.
- 96. Saini, P.; Kumar Verma, P. Evaluation of the wound healing properties of *Mimosa pudica* Linn. In streptozocin-induced diabetes mellitus in rats. *Int. J. Pharm. Sci. Res.* **2019**, *10*, 661–665.
- 97. Xu, X.; Li, X.; Zhang, L.; Liu, Z.; Pan, Y.; Chen, D.; Bin, D.; Deng, Q.; Sun, Y.; Hoffman, R.M.; et al. Enhancement of wound healing by the Chinese medicine herbal mixture *Sophora flavescens* in a rat model of perianal ulceration. *Vivo* **2017**, *31*, 543–549.
- 98. Lodhi, S.; Jain, A.; Jain, A.P.; Pawar, R.S.; Singhai, A.K. Effects of flavonoids from *Martynia annua* and *Tephora purpurea* on cutaneous wound healing. *Avicenna J. Phytomed.* **2016**, *6*, 578–591.
- 99. Chokpaisam, J.; Chusri, S.; Ammuaikit, T.; Udomuksorn, W.; Voravuthikunchai, S.P. Potential wound healing activity of *Cuercus infectoria* formulation in diabetic rats. *PeerJ* 2017, *5*, e3608. [CrossRef]
- Cheng, P.G.; Phan, C.-W.; Sabaratnam, V.; Abdullah, N.; Abdulla, M.A.; Kuppusamy, U.R. Polysaccharides-rich extract of Ganoderma lucidum (M.A. Curtis:Fr.) P. Karst accelerates wound healing in streptozotocin-induced diabetic rats. Evid.-Based Complement. Altern. Med. 2013, 2013. [CrossRef]
- 101. Ngwoke, K.G.; Akwaqbulam, A.G.; Erhirhie, E.O.; Ajaghaku, D.L.; Chiedu Okoye, F.B.; Okechukwu Esimone, C. Antioxidant, Anti-inflammatory, Analgesic Properties, and Phytochemical Characterization of Stem Bark Extract and Fractions of *Anthocleista nobilis. Pharmacogn. Res.* **2018**, *10*, 81–87. [CrossRef]
- Bardaa, S.; Makni, K.; Boudaouara, O.; Bardaa, T.; Ktari, N.; Hachira, S.; Salah, R.B.; Kallel, R.; Sahnound, Z.; Boufi, S. Development and evaluation of the wound healing effect of a novel topical cream formula based on *Ginkgo biloba* extract on wounds in diabetic rats. *BioMed Res. Int.* 2021, 2021, 6474706. [CrossRef] [PubMed]
- 103. Hariharapura, R.; Srinivasan, R.; Ashok, G.; Dongra, S.H.; Jagani, H.V.; Vijayan, P. Investigation of the antioxidant and hepatoprotective potential *Hypericum mysorense*. *Antioxidants* **2014**, *3*, 526–543. [CrossRef] [PubMed]
- 104. Gigliobianco, M.R.; Cortese, M.; Vargas Peregrina, D.V.; Villa, C.; Lupidi, G.; Pruccoli, L.; Angeloni, C.; Tarozzi, A. Development of new extracts of *Crocus sativus* L. By products from two different Italian regions as new potential active ingredient in cosmetic formulations. *Cosmetics* 2021, 8, 51. [CrossRef]
- 105. Jayapal, V.; Subha, V.; Pradeep, J.; Janardan Salwe, K.; Manimekalai, R.; Tahinamala, R.; Kumar, B.; Rayvathy, B.; Ravi Kumar, S. Evaluation of wound healing potential of the essential oil of *Ocimum sanctum* L. (Thulasi/basil) containing ointment in female Wistar albino rats. *J. Pharmacogn. Phytochem.* 2023, *12*, 189–193. [CrossRef]
- Rašković, A.; Milanović, I.; Pavlović, N.; Ćebović, T.; Vukmirović, S.; Mikov, M. Antioxidant Activity of Rosemary (*Rosmarinus officinalis* L.) Essential Oil and Its Hepatoprotective Potential. BMC Complement. Altern. Med. 2014, 14, 225. [CrossRef] [PubMed]
- 107. Karimzadeh, S.; Farahpour, M.R. Topical application of *Salvia officinalis* hydroethanolic leaf extract improves wound healing process. *Indian J. Exp. Biol.* **2017**, *55*, 98–106.
- 108. Guevara-Vazquez, A.M.; Marin-Tello, C.L. Wound healing activity of Allium cepa L. bulbs in a second-degree burn wound model in Holtzman rats. *Vitae* **2021**, *28*, 345737. [CrossRef]
- Park, J.-Y.; Hyck, K.; Lim, D.-W.; Kim, J.-E.; Park, W.-H.; Park, S.-D. Ethanol Extract of *Lycopodium serratum* Thunb. Attenuates Lipopolysaccharide-Induced C6 Glioma Cells Migration via Matrix Metalloproteinase-9 Expression. *Chin. J. Integr. Med.* 2018, 24, 860–866. [CrossRef]
- 110. Dutta, S.; Pattnak, A.K.; Bersa, S.E. Wound healing potential of methanolic extract and its fraction of *Lawsonia alba* Lam leaves formulated a topical gel. *World J. Pharm. Res.* **2016**, *5*, 1091–1109.
- 111. El Massoudi, S.; Zinedine, A.; Rocha, J.M.; Benidir, M.; Najjari, I.; El Ghadraoui, L.; Benjelloun, M.; Errachidi, F. Phenolic composition and wound healing potential assessment of Moroccan Henna (*Lawsonia inermis*) Aqueous extract. *Cosmetics* **2023**, 10, 92. [CrossRef]
- 112. Lukiswanto, B.S.; Miranti, A.; Sudjarwo, S.A.; Primarizky, H.; Yuniati, W.M. Evaluation of wound healing potential of pomegranate (*Punica granatum*) whole fruit extract on skin burn wound in rats (*Rattus norvegicus*). J. Adv. Vet. Anim. Res. 2019, 6, 202–207. [CrossRef] [PubMed]
- 113. Balla, R.; Kaur, R.; Kaur, B.; Kaur, P. *Hibiscus rosa sinensis* Linn. A phytochemical and pharmacological review. *Int. J. Health Sci.* **2022**, *6*, 5165–5193.
- Almasian, A.; Najafi, F.; Eftekhari, M.; Shams Ardekani, M.R.; Sharifzadeh, M.; Khanavi, M. Polyurethane/carboxymethylcellulose nanofibers containing *Malva sylvestris* extract for healing diabetic wounds: Preparation, characterization, in vitro and in vivo studies. *Mat. Sci. Eng.* 2020, 114, 111039. [CrossRef] [PubMed]
- 115. Asif, A.H.; Mulla, S.M.; Ashariff, A.; Sreeharsha, N.; Meravanige, G.; Shiroorkar, P.N.; Basheeruddin Asdaq, S.M.; Khalid Answer, M.D.; Roopashree, T.S.; Karnatti, R.K. Exploring the topical gel of *Thespesia populnea* leaf extract for in vivo wound healing efficacy. *Pharmacogn. Mag.* **2022**, *18*, 519–523.
- Maan, P.; Singh Yadav, K.; Yadav, N.P. Wound healing activity of *Azadirachta indica* A. Juss stem bark in mice. *Pharmacogn. Mag.* 2017, 13 (Suppl. S2), S316–S320.

- 117. Silva, D.F.; Lima, K.T.; Bastos, G.T.N.; Oliveira, J.A.R.; do Nascimento, L.A.S.; Costa, C.E.F.; Filho, N.R.; Concha, V.O.; Passos, M.F. PCL/Andiroba oil (*Carapa guianensis* Aubl.) hybrid film for wound healing applications. *Polymers* **2021**, *13*, 1591. [CrossRef]
- 118. Yadav, E.; Singh, D.; Yadav, P.; Vrma, A. Antioxidant and anti-inflammatory properties of *Prosopis cineraria* based phenolic rich ointment in wound healing. *Biomed. Pharmacother.* 2018, 108, 1572–1583. [CrossRef]
- 119. Gupta, R.; Gupta, J. Ointment of methanolic extract *Ficus religiosa*: Traditional approach in wound healing in rats. *Int. J. Pharm. Sci. Res.* **2016**, *7*, 5006–5011.
- 120. Susanto, A.; Muhaimina, R.K.; Amalliya, A.; Sutjiatmo, A.B. The effectiveness of ethanolic of *Moringa oleífera* Lam. Gel on the wound healing process of the rat's palate. *J. Int. Dent. Med. Res.* **2019**, *12*, 504–509.
- 121. Kumar, M.; Kumar Gautan, M.; Singh, A.; Kumar Goel, R. Healing effect of *Musa sapientum* var. *Paradisiaca* in diabetic rats with co-occurring gastric ulcer: Cytokines and growth factor by PCR amplification. *BMC Complement. Altern. Med.* 2013, *13*, 305. [CrossRef]
- 122. Shrimali, H.; Kumar Mandal, U.; Nivsarkar, M.; Shrivastawa, N. Fabrication and evaluation of a medicated hydrogel film with embelin from *Embelia ribes* for wound healing activity. *Future J. Pharm. Sci.* **2019**, *5*, 12. [CrossRef]
- 123. Moreira, P.; Sousa, F.J.; Matos, P.; Sousa Brites, G.; Gonçalves, M.J.; Cavaleiro, C.; Figueirinha, A.; Salgueiro, L.; Batista, M.T.; Costa Branco, P.; et al. Chemical composition and effect against skin alterations of bioactive extracts obtained by the hydrodistillation of *Eucalyptus globulus* leaves. *Pharmaceutics* 2022, 14, 561. [CrossRef]
- 124. Khuanekkaphan, M.; Noysang, C.; Khobjai, W. Anti-aging potential and phytochemicals of *Centella asiática, nelumbo nucifera* and *Hibiscus sabdariffa* extracts. J. Adv. Pharm. Technol. Res. **2020**, 11, 174–178. [CrossRef] [PubMed]
- 125. Mittal, A.; Satish, S.S.; Anima, P. Evaluation of wound healing, antioxidant and antimicrobial efficacy of *Jasminum auriculatum* Vahl. leaves. *Avicenna J. Phytomed.* **2015**, *6*, 295–304.
- 126. Chaturvedi, A.P.; Kumar, M.; Tripathi, Y.B. Efficacy of *Jasminum grandiflorum* L. leaf extract on dermal wound healing in rats. *Int. Wound J.* **2013**, *10*, 675–682. [CrossRef] [PubMed]
- 127. Zhang, C.; He, Y.; Chen, Z.; Shi, J.; Qu, Y.; Zhang, J. Effect of Polysaccharides from *Bletilla striata* on the Healing of Dermal Wounds in Mice. *Evid.-Based Complement. Altern. Med.* **2019**, 2019, 9212314. [CrossRef]
- Ekiert, H.; Klimek-Szczykutowicz, M.; Szopa, A. Paeonia suffruticosa (Mountan Peony)—A review of the chemical composition, traditional and profesional use in medicine, position in cosmetics industries and biotechnological studies. Plants 2022, 11, 3379. [CrossRef]
- 129. Ayele, T.M.; Chekol Abebe, E.; Tilahum Muche, Z.; Mekonnen Agidew, M.; Shumet Yimer, Y.; Testaw Addis, G.; Dagnaw Baye, N.; Bogale Kassie, A.; Adela Alemu, M.; Gobezie Yblet, T.; et al. Evaluation of in vivo wound-healing and anti-inflammatory activities of solvent fractions of fruits of *Argemone mexicana* L. (Papaveraceae). *Evid.-Based Complement. Altern. Med.* 2022, 2022, 6154560. [CrossRef]
- Heba, E.; El Aty, A.; Zaazaa, A.M.; Mohamed, S.H.; El Dayem, S.A.; Foda, F. Promising Therapeutic Efficacy of *Trigonella-foenum* graecum and Bone Marrow-Derived Mesenchymal Stem Cells on Skeletal Muscle Atrophy in Experimental Rat Model. *Biointerrface Res. Appl. Chem.* 2023, 13, 133. [CrossRef]
- 131. Somwanshi Sachim, B.; Hiremath Shivanand, N. In vivo evaluation of the wound healing activity of the *Sesamum indicum* L. seed extract in novel ethosomal vesicular system. *J. Drug Del. Therap.* **2018**, *8*, 411–420. [CrossRef]
- 132. Ghanadian, M.; Soltani, R.; Homayouni, A.; Khorvash, F.; Jouabadi, S.M.; Abdollahzadeh, M. The Effect of Plantago Major Hydroalcoholic Extract on the Healing of Diabetic Foot and Pressure Ulcers: A Randomized Open-Label Controlled Clinical Trial. *Int. J. Low Extrem. Wounds* 2022. [CrossRef]
- 133. Yang, W.-T.; Ke, C.-Y.; Wu, W.-T.; Harn, H.-J.; Teng, Y.-H.; Lee, R.-P. Effects of *Angelica dahurica* and *Rheum officinale* extracts on excisional wound healing in rats. *Evid.-Based Complement. Altern. Med.* **2017**, 2017, 1583031. [CrossRef] [PubMed]
- 134. Budiawan, A.; Purwanto, A.; Puradewa, L.; Dwi Cahyani, E.; Endang Purwaningsh, C. Wound healing activity and flavonoid contentes of pursale (*Portulaca grandiflora*) of various varieties. *RSC Adv.* **2023**, *13*, 9871. [CrossRef]
- 135. Mahomoodally, M.F.; Jugreet, S.; Sinan, K.I.; Zengin, G.; Gunes, A.K.; Ramazan, C.; Josef, J.; Zoltan, C.; Angelin, P.; Angeles Flores, G.; et al. Pharmacological potential and chemical characrization of *Bridelia ferruginea* Benth.—A native tropical African Medicinal Plants. *Antibiotics* 2021, 10, 223. [CrossRef]
- 136. Zhang, H.; Chen, J.; Cen, Y. Burn wound healing potential of a polysaccharide from *Sanguisorba officinalis* L. in mice. *Int. J. Biol. Macromol.* **2018**, 112, 862–867. [CrossRef] [PubMed]
- 137. Ly, H.T.; Pham Nguyen, M.T.; Oanh Nguyen, T.K.; Quynh Bui, T.P.; Ke, X.; Le, V.M. Phytochemical analysis and wound healing activity of Noni (*Morinda citrifolia*) leaf extract. J. Herbs. Spices Med. Plants **2020**, 26, 379–393. [CrossRef]
- Wen, M.; Chen, Q.; Chen, W.; Yang, J.; Zhou, X.; Zhang, C.; Wu, A.; Lai, J.; Chen, J.; Mei, Q.; et al. A comprehensive review of *Rubia cordifolia* L.: Traditional uses, phytochemistry, pharmacological activities, and clinical applications. *Front. Pharmacol.* 2022, 13, 965390. [CrossRef]
- 139. Azmil, L.; Shukla, I.; Goutam, A.; Allauddin Rao, C.H.V.; Jawaid, T.; Kamal, M.; Awaad, A.S.; Alqasoumi, S.I. In vitro wound healing activity of 1-hydroxy-5,7-dimethoxy-2-naphthalene-carboxaldehyde (HDNC) and other isolates of *Aegle marmelos* L.: Enhances keratinocytes motility via Wnt/β-catenin and RAS-ERK pathways. *Saudi Pharm. J.* 2019, 27, 532–539. [CrossRef] [PubMed]

- Zhang, M.; Wang, J.; Zhu, L.; Li, T.; Jiang, W.; Peng, W.; Wu, C. Zanthoxylum bungeanum Maxim. (Rutacea): A systematic Review of traditional uses, botany, phytochemistry, pharmacology, pharmacokinetics, and toxicology. Int. J. Mol. Sci. 2017, 18, 2172. [CrossRef] [PubMed]
- Albano, M.N.; da Silveira, M.R.; Danielski, L.G.; Florentino, D.; Petronilho, F.; Piovezan, A.P. Anti-Inflammatory and Antioxidant Properties of Hydroalcoholic Crude Extract from *Casearia sylvestris* Sw. (Salicaceae). J. Ethnopharmacol. 2013, 147, 612–617. [CrossRef]
- 142. Xu, L.; Zhang, W.; Zeng, L.; Jin, J.-O. *Rehmannia glutinosa* polysaccharide induced an anti-cancer effect by activating natural killer cells. *Int. J. Biol. Macromol.* **2017**, *105*, 680–685. [CrossRef]
- 143. Kil, Y.-S.; Park, J.; Han, A.-R.; Woo, H.; Seo, E.-K. A new 9,10-dihydrophenanthrene and cell proliferative 3,4-δ-dehydrotocopherols from *Stemona tuberosa*. *Molecules* **2015**, *20*, 5965–5974. [CrossRef] [PubMed]
- 144. Hajiaghaalipour, F.; Kanthimathi, M.S.; Abdulla, M.A.; Sanusi, J. The effect of *Camellia sinensis* on wound healing potential in an animal model. *Evid.-Based Complement. Altern. Med.* **2013**, 2013. [CrossRef] [PubMed]
- 145. Chen, Y.; Guo, J.; Tang, Y.; Wu, L.; Tao, W.; Qian, Y.; Duan, J.-A. Pharmacokinetic profile and metabolite identification of yuanhuapine, a bioactive component in *Daphne genka* by ultra-high performance liquid chromatography coupled with tandem mass spectrometry. J. Pharm. Biomed. Anal. 2015, 112, 60–69. [CrossRef] [PubMed]
- 146. Liang, J.H.; Lin, H.R.; Yang, C.S.; Liaw, C.C.; Wang, I.C.; Chen, J.J. Bioactive components from *Ampelopsis japonica* with antioxidant, glucosidase, and antiacetylcholinesterase activities. *Antioxidants* **2022**, *11*, 1228. [CrossRef]
- 147. Irham, W.H.; Hardiyanti, R. Wound healing bioactivity of Curcuma longa Linn. Rasayan J. Chem. 2021, 14, 2386–2391. [CrossRef]
- 148. Singh, S.; Gupta, A.; Singh, B.B. Effect of foliage supplementation to *Heteropogon contortus* based diets on nutrients digestibility, gas and metabolites production in sheep and goat inoculums. *Anim. Nutr. Feed Technol.* **2016**, *16*, 439–450. [CrossRef]
- 149. Chandika, P.; Ko, S.-C.; Jung, W.-K. Marine-derived biological macromolecule-based biomaterials for wound healing and skin tissue regeneration. *Macromolecules* 2015, 77, 24–25. [CrossRef] [PubMed]
- 150. Ghitescu, R.E.; Volf, I.; Carausu, C.; Bühlmann, A.M.; Gilca, J.A.; Popa, V.I. Optimization of ultrasound-assisted extraction of polyphenols from spruce wood bark. *Ultrason. Sonochem.* **2015**, *22*, 535–541. [CrossRef] [PubMed]
- 151. Arruda, H.S.; Pereira, G.A.; Pastore, G.M. Optimization of extraction parameters of total phenolics from *Annona crassiflora* Mart. (araticum) fruits using response surface methodology. *Food Anal. Methods* **2017**, *10*, 100–110. [CrossRef]
- 152. George, E.; Kasipandi, M.; Vekataramana, M.; Kumar, K.N.; Allen, J.A.; Parimelazhagan, T.; Gopalan, N. In vitro anti-oxidant and cytotoxic analysis of *Pogostemon mollis* Benth. *Bangladesh J. Pharmacol.* **2016**, *11*, 148–158. [CrossRef]
- Velderrain-Rodríguez, G.; Palafox-Carlos, H.; Wall-Medrano, A.; Ayala-Zavala, J.F.; Chen, C.-Y.O.; Robles-Sánchez, M.; Astiazaran-García, H.; Alvarez-Parrilla, E.; González-Aguilar, G.A. Phenolic compounds: Their journey after intake. *Food Funct.* 2014, *5*, 189–197. [CrossRef] [PubMed]
- Arruda, H.S.; Silva, E.K.; Pereira, E.K.; Angolini, C.F.F.; Eberlin, M.N.; Meireles, M.A.A.; Pastore, G.M. Effects of high-intensity ultrasound process parameters on the phenolic compound recovery from araticum peel. *Ultrason. Sonochem.* 2019, 50, 82–95. [CrossRef] [PubMed]
- 155. Arruda, H.S.; Pereira, G.A.; Pastore, G.M. Brazilian Cerrado fruit araticum (*Annona crassiflora* Mart.) as a potential source of natural antioxidant compounds. *Int. Food Res. J.* 2018, 25, 2005–2012. Available online: http://www.ifrj.upm.edu.my/25(05)2018/(33).pdf (accessed on 9 January 2024).
- 156. Arruda, H.S.; Fernandes, R.V.B.; Botrel, D.A.; Almeida, M.E.F. Frutos do Cerrado: Conhecimento e aceitação de Annona crassiflora Mart. (araticum) e Eugenia dysenterica Mart. (Caigata) por crianças utilizando o paladar e a visão. J. Health Biol. Sci. 2015, 3, 224–230. [CrossRef]
- 157. Chen, M.-X.; Huo, J.-M.; Hu, J.; Xu, Z.-P.; Zhang, X. Amaryllidaceae alkaloids from *Crinum latifolium* with cytotoxic, antimicrobial, antioxidant, and anti-inflammatory activities. *Fitoterapia* **2018**, *130*, 48–53. [CrossRef]
- 158. Formagio, A.S.N.; Vieira, M.C.; Volobuff, C.R.F.; Silva, M.S.; Matos, A.I.; Cardoso, C.A.L.; Carvalho, J.E. In vitro biological screening of the anticholinesterase and antiproliferative activities of medicinal plants belonging to Annonaceae. *Braz. J. Med. Biol. Res.* **2015**, *48*, 308–315. [CrossRef]
- 159. Liu, Y.; Xiao-Hong, G.; Tang, J.; Liu, K. Antioxidant Activities of Hops (*Humulus Lupulus*) and Their Products. J. Am. Soc. Brew. Chem. 2018, 65, 116–121. [CrossRef]
- 160. Zhang, I.; Ravipati, A.S.; Koyyalamudi, S.R.; Jeong, S.C.; Reddy, N.; Smith, P.T.; Bartlet, J.; Shanmugam, K.; Munch, G.; Wu, M.J. Antioxidant and anti-inflammatory activities of selected medicinal plants containing phenolic and flavonoid compounds. J Agric. Food Chem. 2011, 59, 12361–12367. [CrossRef]
- 161. Fattahi, M.; Rahimi, R. Optimization of extraction parameters of phenolic antioxidants from leaves of *Capparis spinosa* using response surface methodology. *Food Anal. Method* **2016**, *9*, 2321–2334. [CrossRef]
- Formagio, A.S.N.; Masetto, T.E.; da Baldivia, D.S.; Vieira, M.C.; Zarate, N.A.H.; Pereira, Z.V. Potencial alelopático de cinco espécies da família Annonaceae. *Braz. J. Bioscien.* 2010, *8*, 349–354.
- 163. Scalise, A.; Bianchi, A.; Tartaglione, C.; Bolletta, E.; Pierangeli, M.; Torresetti, M.; Marazzi, M.; Di Benedetto, G. Microenvironment and Microbiology of Skin Wounds: The Role of Bacterial Biofilms and Related Factors. *Semin. Vasc. Surg.* 2015, 28, 151–159. [CrossRef] [PubMed]
- 164. Ibrahim, N.; Wong, S.K.; Mohamed, I.N.; Mohamed, N.; Chin, K.Y.; Ima-Nirwana, S.; Shuid, A.N. Wound Healing Properties of Selected Natural Products. Int. J. Environ. Res. Public Health 2018, 15, 2360. [CrossRef] [PubMed]

- 165. Tsala, D.E.; Amadou, D.; Habtemariam, S. Natural Wound Healing and Bioactive Natural Products. *Phytopharmacology* **2013**, *4*, 532–560.
- 166. Bittner Fialová, S.; Rendeková, K.; Mucaji, P.; Nagy, M.; Slobodníková, L. Antibacterial Activity of Medicinal Plants and Their Constituents in the Context of Skin and Wound Infections, Considering European Legislation and Folk Medicine A Review. Int. J. Mol. Sci. 2021, 22, 10746. [CrossRef] [PubMed]
- 167. Amparo, T.R.; Seibert, J.B.; de Abreu Vieira, P.M.; Teixeira, F.F.M.; dos Santos, O.D.F.; de Souza, G.H.B. Herbal Medicines to the Treatment of Skin and Soft Tissue Infections: Advantages of the Multi-Targets Action. *Phyther. Res.* **2020**, *34*, 94–103. [CrossRef]
- 168. Simões, D.; Miguel, S.P.; Ribeiro, M.P.; Coutinho, P.; Mendonça, A.G.; Correia, J.J. Recent Advances on Antimicrobial Wound Dressing: A Review. *Eur. J. Pharm. Biopharm.* **2018**, 127, 130–141. [CrossRef]
- Andreu, V.; Mendoza, G.; Arruebo, M.; Irusta, S. Smart Dressings Based on Nanostructured Fibers Containing Natural Origin Antimicrobial, Anti-Inflammatory, and Regenerative Compounds. *Materials* 2015, 8, 5154–5193. [CrossRef]
- Maheshwari, R.K.; Singh, A.K.; Gaddipati, J.; Srimal, R.C. Multiple Biological Activities of Curcumin: A Short Review. *Life Sci.* 2006, 78, 2081–2087. [CrossRef] [PubMed]
- 171. Amer, S.S.; Mamdouh, W.; Nasr, M.; ElShaer, A.; Polycarpou, E.; Abdel-Aziz, R.T.A.; Sammour, O.A. Quercetin Loaded CosmNutraceutical Electrospun Composite Nanofibers for Acne Alleviation: Preparation, Characterization and Experimental Clinical Appraisal. *Int. J. Pharm.* 2022, 612, 121309. [CrossRef] [PubMed]
- 172. Dyja, R.; Jankowski, A. The Effect of Additives on Release and in Vitro Skin Retention of Flavonoids from Emulsion and Gel Semisolid Formulations. *Int. J. Cosmet. Sci.* 2017, 39, 442–449. [CrossRef] [PubMed]
- 173. Roy, P.; Parveen, S.; Ghosh, P.; Ghatak, K.; Dasgupta, S. Flavonoid Loaded Nanoparticles as an Effective Measure to Combat Oxidative Stress in Ribonuclease A. *Biochimie* **2019**, *162*, 185–197. [CrossRef]
- 174. Anwar, A.; Masri, A.; Rao, K.; Rajendran, K.; Khan, N.A.; Shah, M.R.; Siddiqui, R. Antimicrobial Activities of Green Synthesized Gums-Stabilized Nanoparticles Loaded with Flavonoids. *Sci. Rep.* **2019**, *9*, 3122. [CrossRef]
- Gomes, F.; Henriques, M. Control of Bovine Mastitis: Old and Recent Therapeutic Approaches. *Curr. Microbiol.* 2016, 72, 377–382. [CrossRef]
- 176. Mala, L.; Lalouckova, K.; Skrivanova, E. Bacterial Skin Infections in Livestock and Plant-Based Alternativ es to Their Antibiotic Treatment. *Animals* 2021, *11*, 2473. [CrossRef]
- 177. Cheng, W.N.; Han, S.G. Bovine Mastitis: Risk Factors, Therapeutic Strategies, and Alternative Treatments—A Review. AsianAustralas. J. Anim. Sci. 2020, 33, 1699–1713. [CrossRef]
- Mushtaq, S.; Shah, A.M.A.; Lone, S.A.; Hussain, A.; Hassan, Q.P.; Ali, M.N. Bovine Mastitis: An Appraisal of Its Alternative Herbal Cure. *Microb. Pathog.* 2018, 114, 357–361. [CrossRef]
- 179. Tohma, H.; Gulcin, I.; Bursal, E.; Goren, A.; Alwasel, S.H.; Koksal, I. Antioxidant activity and phenolic compounds of ginger (*Zingiber officinale* Rosc.) determined by HPLC-MS/MS. *Food Meas.* **2017**, *11*, 556–566. [CrossRef]
- Moghadam, M.; Salami, M.; Mohammadian, M.; Khodadadi, M.; Emam-Djomeh, Z. Development of antioxidant edible films based on mung bean protein enriched with pomegranate peel. *Food Hydrocoll.* 2020, 104, 105735. [CrossRef]
- 181. Krishnaiah, D.; Sarbatly, R.; Nithyanandam, R. A review of the antioxidant potential of medicinal plant species. *Food Biproducts Process.* **2011**, *89*, 217–233. [CrossRef]
- 182. Ambika, P.P.S.; Chauhan, S.S.M. Activity-guided isolation of antioxidants from the leaves of *Terminalia arjuna*. *Former. Nat. Prod. Lett.* **2014**, *28*, 760–763. [CrossRef]
- 183. Odeh, D.; Orsolic, N.; Berendika, M.; Dikic, D.; Drozdek, S.D.; Balbino, S.; Repajic, M.; Dragovic-Uzelac, V.; Jurcevic, I.L. Antioxidant and anti-atherogenic activities of essential oils from *Myrtus communis* L. and *Laurus nobilis* L. in rat. *Nutrients* 2022, 14, 1465. [CrossRef] [PubMed]
- 184. Machado, A.; Ferreira, S.; da Silva Medeiros, F.; Fujiwara, R.; de Souza Filho, J.; Pimenta, L. Nematicidal activity of *Annona* crassiflora leaf extract on *Caenorhabditis elegans*. *Parasites Vectors* **2015**, *8*, 113. [CrossRef] [PubMed]
- 185. Quílez, A.M.; Fernández-Arche, M.A.; García-Giménez, M.D.; De la Puerta, R. Potential therapeutic applications of the genus *Annona*: Local and traditional uses and pharmacology. *J. Ethnopharm.* **2018**, 225, 244–270. [CrossRef] [PubMed]
- 186. Yuan, G.; Guan, Y.; Yi, H.; Lai, S.; Sun, Y.; Cao, S. Antibacterial Activity and Mechanism of Plant Flavonoids to Gram-Positive Bacteria Predicted from Their Lipophilicities. *Sci. Rep.* **2021**, *11*, 10471. [CrossRef]
- Carvalho, M.T.B.; Araújo-Filho, H.G.; Barreto, A.S.; Quintans-Júnior, L.J.; Quintans, J.S.S.; Barreto, R.S.S. Wound Healing Properties of Flavonoids: A Systematic Review Highlighting the Mechanisms of Action. *Phytomedicine* 2021, 90, 153636. [CrossRef]
- 188. Yang, X.; Jiang, Y.; Yang, J.; He, J.; Sun, J.; Chen, F.; Zhang, M.; Yang, B. Prenylated Flavonoids, Promising Nutraceuticals with Impressive Biological Activities. *Trends Food Sci. Technol.* **2015**, *44*, 93–104. [CrossRef]
- Mukai, R. Prenylation Enhances the Biological Activity of Dietary Flavonoids by Altering Their Bioavailability. *Biosci. Biotechnol. Biochem.* 2018, 82, 207–215. [CrossRef]
- Hošek, J.; Závalová, V.; Šmejkal, K.; Bartoš, M. Effect of Diplacone on Lps-Induced Inflammatory Gene Expression in Macrophages. Folia Biol. 2010, 56, 124–130.
- Shin, H.J.; Shon, D.H.; Youn, H.S. Isobavachalcone Suppresses Expression of Inducible Nitric Oxide Synthase Induced by Toll-like Receptor Agonists. Int. Immunopharmacol. 2013, 15, 38–41. [CrossRef]
- 192. Dzoyem, J.P.; Nkuete, A.H.L.; Ngameni, B.; Eloff, J.N. Anti-Inflammatory and Anticholinesterase Activity of Six Flavonoids Isolated from *Polygonum* and *Dorstenia* Species. *Arch. Pharm. Res.* 2017, 40, 1129–1134. [CrossRef] [PubMed]

- 193. Hu, J.; Liu, J. Licochalcone Attenuates Lipopolysaccharide-Induced Acute Kidney Injury by Inhibiting NF-KB Activation. *Inflammation* **2016**, *39*, 569–574. [CrossRef]
- 194. Jia, T.; Qiao, J.; Guan, D.; Chen, T. Anti-Inflammatory Effects of Licochalcone A on IL-1β-Stimulated Human Osteoarthritis Chondrocytes. *Inflammation* 2017, 40, 1894–1902. [CrossRef]
- Cha, S.M.; Cha, J.D.; Jang, E.J.; Kim, G.U.; Lee, K.Y. Sophoraflavanone G Prevents *Streptococcus mutans* Surface Antigen I/IIInduced Production of NO and PGE2 by Inhibiting MAPK-Mediated Pathways in RAW 264.7 Macrophages. *Arch. Oral Biol.* 2016, 68, 97–104. [CrossRef]
- Wun, Z.Y.; Lin, C.F.; Huang, W.C.; Huang, Y.L.; Xu, P.Y.; Chang, W.T.; Wu, S.J.; Liou, C.J. Anti-Inflammatory Effect of Sophoraflavanone G Isolated from *Sophora flavescens* in Lipopolysaccharide-Stimulated Mouse Macrophages. *Food Chem. Toxicol.* 2013, 62, 253–361. [CrossRef]
- Song, M.; Liu, Y.; Li, T.; Liu, X.; Hao, Z.; Ding, S.; Panichayupakaranant, P.; Zhu, K.; Shen, J. Plant Natural Flavonoids against Multidrug Resistant Pathogens. *Adv. Sci.* 2021, *8*, 2100749. [CrossRef]
- Bogdanova, K.; Röderova, M.; Kolar, M.; Langova, K.; Dusek, M.; Jost, P.; Kubelkova, K.; Bostik, P.; Olsovska, J. Antibiofilm Activity of Bioactive Hop Compounds Humulone, Lupulone and Xanthohumol toward Susceptible and Resistant Staphylococci. *Res. Microbiol.* 2018, 169, 127–134. [CrossRef]
- Hartkorn, A.; Hoffmann, F.; Ajamieh, H.; Vogel, S.; Heilmann, J.; Gerbes, A.L.; Vollmar, A.M.; Zahler, S. Antioxidant Effects of Xanthohumol and Functional Impact on Hepatic Ischemia-Reperfusion Injury. J. Nat. Prod. 2009, 72, 1741–1747. [CrossRef]
- 200. Cho, Y.C.; Kim, H.J.; Kim, Y.J.; Lee, K.Y.; Choi, H.J.; Lee, I.S.; Kang, B.Y. Differential Anti-Inflammatory Pathway by Xanthohumol in IFN-γ and LPS-Activated Macrophages. *Int. Immunopharmacol.* **2008**, *8*, 567–573. [CrossRef]
- 201. Lupinacci, E.; Meijerink, J.; Vincken, J.P.; Gabriele, B.; Gruppen, H.; Witkamp, R.F. Xanthohumol from Hop (*Humulus lupulus* L.) Is an Efficient Inhibitor of Monocyte Chemoattractant Protein-1 and Tumor Necrosis Factor-α Release in LPS-Stimulated RAW 264.7 Mouse Macrophages and U937 Human Monocytes. J. Agric. Food Chem. 2009, 57, 7274–7281. [CrossRef]
- Cho, Y.C.; You, S.K.; Kim, H.J.; Cho, C.W.; Lee, I.S.; Kang, B.Y. Xanthohumol Inhibits IL-12 Production and Reduces Chronic Allergic Contact Dermatitis. *Int. Immunopharmacol.* 2010, 10, 556–561. [CrossRef] [PubMed]
- 203. Negrão, R.; Costa, R.; Duarte, D.; Gomes, T.T.; Coelho, P.; Guimarães, J.T.; Guardão, L.; Azevedo, I.; Soares, R. XanthohumolSupplemented Beer Modulates Angiogenesis and Inflammation in a Skin Wound Healing Model. Involvement of Local Adipocytes. J. Cell. Biochem. 2012, 113, 100–109. [CrossRef] [PubMed]
- Górniak, I.; Bartoszewski, R.; Króliczewski, J. Comprehensive Review of Antimicrobial Activities of Plant Flavonoids. *Phytochem. Rev.* 2019, 18, 241–272. [CrossRef]
- Sarbu, L.G.; Bahrin, L.G.; Babii, C.; Stefan, M.; Birsa, M.L. Synthetic Flavonoids with Antimicrobial Activity: A Review. J. Appl. Microbiol. 2019, 127, 1282–1290. [CrossRef] [PubMed]
- 206. Wu, D.; Kong, Y.; Han, C.; Chen, J.; Hu, L.; Jiang, H.; Shen, X. D-Alanine:D-Alanine Ligase as a New Target for the Flavonoids Quercetin and Apigenin. Int. J. Antimicrob. Agents 2008, 32, 421–426. [CrossRef] [PubMed]
- Sirk, T.W.; Brown, E.F.; Sum, A.K.; Friedman, M. Molecular Dynamics Study on the Biophysical Interactions of Seven Green Tea Catechins with Lipid Bilayers of Cell Membranes. J. Agric. Food Chem. 2008, 56, 7750–7758. [CrossRef]
- Kusuda, M.; Inada, K.; Ogawa, T.O.; Yoshida, T.; Shiota, S.; Tsuchiya, T.; Hatano, T. Polyphenolic Constituent Structures of Zanthoxylum piperitum Fruit and the Antibacterial Effects of Its Polymeric Procyanidin on Methicillin-Resistant Staphylococcus aureus. *Biosci. Biotechnol. Biochem.* 2006, 70, 1423–1431. [CrossRef] [PubMed]
- Chan, E.W.C.; Wong, S.K.; Tangah, J.; Chan, H.T. Chemistry and Pharmacology of Artocarpin: An Isoprenyl Flavone from Artocarpus Species. Syst. Rev. Pharm. 2018, 9, 58–63. [CrossRef]
- Dej-Adisai, S.; Meechai, I.; Puripattanavong, J.; Kummee, S. Antityrosinase and Antimicrobial Activities from Thai Medicinal Plants. Arch. Pharm. Res. 2014, 37, 473–483. [CrossRef] [PubMed]
- Septama, A.W.; Panichayupakaranant, P. Synergistic Effect of Artocarpin on Antibacterial Activity of Some Antibiotics against Methicillin-Resistant Staphylococcus aureus, Pseudomonas aeruginosa, and Escherichia coli. Pharm. Biol. 2016, 54, 686–691. [CrossRef]

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