



Review

Precision Medicine in Erythropoietin Deficiency and Treatment Resistance: A Novel Approach to Management of Anaemia in Chronic Kidney Disease

Nava Yugavathy ¹, Bashar Mudhaffar Abdullah ², Soo Kun Lim ³, Abdul Halim Bin Abdul Gafor ⁴,
Muh Geot Wong ^{5,6}, Sunita Bavanandan ⁷, Hin Seng Wong ⁸ and Hasniza Zaman Huri ^{1,*}

¹ Department of Clinical Pharmacy and Pharmacy Practice, Faculty of Pharmacy, Universiti Malaya, Kuala Lumpur 50603, Malaysia; navayugavathy90@gmail.com

² Clinical Laboratory Technology Department, Al-Rafidain University College, Baghdad 46036, Iraq; basharmudhaffar22@ruc.edu.iq

³ Department of Medicine, Faculty of Medicine, Universiti Malaya, Kuala Lumpur 50603, Malaysia; limsk@ummc.edu.my

⁴ Faculty of Medicine, National University of Malaysia, Bangi 43600, Malaysia; halimgafor@gmail.com

⁵ Department of Renal Medicine, Royal North Shore Hospital, Sydney, NSW 2065, Australia; mwong@georgeinstitute.org.au

⁶ The George Institute for Global Health, University of New South Wales, Kensington, NSW 2052, Australia

⁷ Department of Nephrology, Hospital Kuala Lumpur, Kuala Lumpur 50586, Malaysia; sbavanandan@gmail.com

⁸ Department of Nephrology, Hospital Selayang, Batu Caves 68100, Malaysia; hinseng@gmail.com

* Correspondence: hasnizazh@um.edu.my; Tel.: +60-3-79674909; Fax: +60-3-79674964



Citation: Yugavathy, N.; Abdullah, B.M.; Lim, S.K.; Abdul Gafor, A.H.B.; Wong, M.G.; Bavanandan, S.; Wong, H.S.; Huri, H.Z. Precision Medicine in Erythropoietin Deficiency and Treatment Resistance: A Novel Approach to Management of Anaemia in Chronic Kidney Disease. *Curr. Issues Mol. Biol.* **2023**, *45*, 6550–6563. <https://doi.org/10.3390/cimb45080413>

Academic Editor: Emiel P.C. van der Vorst

Received: 26 June 2023

Revised: 27 July 2023

Accepted: 27 July 2023

Published: 7 August 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: The study of anaemia is a well-developed discipline where the concepts of precision medicine have, in part, been researched extensively. This review discusses the treatment of erythropoietin (EPO) deficiency anaemia and resistance in cases of chronic kidney disease (CKD). Traditionally, erythropoietin-stimulating agents (ESAs) and iron supplementation have been used to manage anaemia in cases of CKD. However, these treatments pose potential risks, including cardiovascular and thromboembolic events. Newer treatments have emerged to address these risks, such as slow-release and low-dosage intravenous iron, oral iron supplementation, and erythropoietin-iron combination therapy. Another novel approach is the use of hypoxia-inducible factor prolyl hydroxylase inhibitors (HIF-PHIs). This review highlights the need for precision medicine targeting the genetic components of EPO deficiency anaemia in CKD and discusses individual variability in genes such as the erythropoietin gene (EPO), the interleukin- β gene (IL- β), and the hypoxia-inducible factor gene (HIF). Pharmacogenetic testing aims to provide targeted therapies and interventions that are tailored to the specific characteristics of an individual, thus optimising treatment outcomes and minimising resistance and adverse effects. This article concludes by suggesting that receptor modification has the potential to revolutionise the treatment outcomes of patients with erythropoietin deficiency anaemia through the integration of the mentioned approach.

Keywords: precision medicine; erythropoietin deficiency anaemia; HIF-PHI; pharmacogenetics; inflammation

1. Introduction

Anaemia is a common complication of chronic kidney disease (CKD) and can significantly impact the quality of life of affected individuals. The cause of anaemia in patients with deteriorating kidney function is multifactorial. As the most common causes, when iron and nutrient deficiencies are ruled out, EPO deficiency is the most likely diagnosis. However, chronic inflammation, blood loss, and hyperparathyroidism are also taken into account. Treatment for anaemia among patients with CKD has focused on administering

erythropoietin-stimulating agents (ESAs) and iron supplementation. Methoxy polyethylene glycol-epoetin beta (Mircera) and Darbepoetin alfa (Aranesp) are two of the most commonly used ESAs. They function by increasing the production of red blood cells, which helps to alleviate anaemia symptoms. However, these drugs cause an undesirable number of adverse effects in a certain group of patients [1,2].

Recently, new treatments have offered additional options for the management of erythropoietin (EPO) deficiency anaemia in relation to CKD. One approach is modifying conventional iron therapy [3]. Intravenous iron, oral iron, and erythropoietin-iron combination therapy have all been used to improve iron reserves and support red blood cell production. However, oral ferrous (Fe^{2+}) compounds can cause gastrointestinal side effects, and intravenous iron (IV) therapy can be associated with the risk of anaphylaxis [4]. Therefore, newer formulations of IV iron administration with slow release and low dosages are being considered.

Another novel approach is the usage of hypoxia-inducible factor prolyl hydroxylase inhibitors (HIF-PHIs), roxadustat (Evrenzo), and daprodustat (Duvroq). HIF-PHIs increase endogenous erythropoietin production and red blood cell production, which can improve the symptoms of anaemia without posing the risks associated with exogenous erythropoietin. The OLYMPUS and DOLOMITES clinical trials have shown that roxadustat is effective in treating cases of anaemia with various comorbidities, presenting a favourable safety profile [5,6]. HIF-PH is also preferred in certain groups of CKD patients, including those suffering from heart failure [7] and chronic inflammation [8].

One of the greatest unmet needs in relation to EPO deficiency is the identification of causative genetic mutations, which could facilitate personalised medicine. A broad range of genes are involved in EPO deficiency. However, a molecular diagnosis needs to be established in a clinical setting to avoid unnecessary costly and invasive treatments. Of the many genes involved in this mechanism, we focus on the erythropoietin gene (EPO), the interleukin- β gene (IL- β), and the hypoxia-inducible factor gene (HIF).

This review article focuses on the three approaches to treating EPO deficiency anaemia in patients with CKD. We discuss current treatment options, practical advantages and shortcomings, precision molecular medicine targeting genetic components, and emerging alternative therapies.

2. Overview of Pharmacogenetics

Pharmacogenetics focuses on the influence of single-gene polymorphisms on drug responses. This process depends on the variations in how people respond to medicinal therapy, which is a rapidly expanding field in molecular biology and clinical medicine. With the advent of genetic testing, pharmacogenetics, a term that gained popularity in the 1930s, has now been rediscovered [9]. Up to 95% of the variability in treatment responses can be attributed to genetic variables. However, other elements, including age, gender, physiology, pathophysiology, culture, behaviour, and environment, may significantly impact the variations in these parameters. For example, if members of the same family with the same inherited condition respond differently to the same medical treatment, genetic factors are likely to play a role [10].

A single-nucleotide polymorphism (SNP), often known as a genetic variation, is a change in a nucleotide sequence that affects the pharmacokinetic and pharmacodynamic characteristics of medications [11]. Pharmacogenetic tests are designed to identify patients who respond or do not respond to treatment, show interactions with other drugs, experience side effects, or need their dosages adjusted [10]. Pharmacogenetics has become a vital subject due to the concept of individualised medication. In the field of medicine known as “personalised medicine”, treatment choices are based on a patient’s entire dataset, for which their individuality is taken into account. These data consist of genetic, environmental, and quality-of-life information.

3. Impact of Iron Deficiency and Treatment

Chronic kidney disease is largely considered an inflammatory disease that affects haematopoietic function and the corresponding pathway [12]. There are two types of iron deficiency (ID) that disrupt EPO production, namely absolute and relative [13]. Absolute iron deficiency occurs when there is a diminished level of iron in the body due to halted iron absorption or severe blood loss. An affected individual will typically present with decreased iron, ferritin, and transferrin saturation and an increased total iron binding capacity (TIBC). Relative iron deficiency, in contrast, is due to the inefficient use of stored iron, which occurs due to inflammation, genetic errors, or EPO deficiency [4]. ID is also correlated with cardiovascular disease prognosis, for which a patient's iron status is often screened [14]. Campodonico et al. [15] reported that heart failure patients with transferrin saturation (TSAT) < 20% and a ferritin level above 100 ug/L are likely to have the worst outcomes.

Iron supplementation therapy is typically administered in the form of iron-containing oral supplements such as ferric maltol, a non-salt-based form of iron. Ferric maltol has been shown to be more effective than other iron salts, presenting fewer side effects in a phase III trial. Evidence from the 1-year AEGIS-CKD [16] trial has shown that this compound is able to normalise and sustain Hb levels with a much lower rate of gastrointestinal disorder than that of ferrous sulphate [17]. Although this compound has been licensed in the United States (US) and Europe for the treatment of iron deficiency anaemia [18], its efficacy and safety profile are still under assessment with respect to CKD patients. Newer intravenous iron supplementation treatments such as ferric carboxymaltose (FCM) and ferric derisomaltose (FDM) have been formulated to improve safety and reduce hypersensitivity reactions. FCM and FDM are highly concentrated forms of iron that utilise a unique iron-carbohydrate complex and can be administered quickly in a single or a few IV doses [19,20]. However, it is important to note that the administration of IV iron to patients with CKD requires careful monitoring and close collaboration between the patient's nephrologist and haematologist. This is because patients with CKD are susceptible to iron overload, which may lead to undesired side effects [21]. Additionally, patients with advanced CKD may have difficulty excreting excess iron, which can lead to iron overload and further complications. In the case of functional iron deficiency, which is usually caused by the underutilisation of EPO due to hepcidin upregulation, IV iron administration may cause iron overload toxicity and enhance oxidative stress. In an attempt to compare the safety profiles of FCM, FDM, and iron sucrose in terms of hypersensitivity using a robust and reliable method, Pollock and Biggar [22] concluded that the risk of hypersensitivity regarding FDM is relatively lower compared to the other two compounds.

4. Erythropoietin

In a normal kidney, renal EPO-producing cells (REPs) are peritubular interstitial fibroblast-like cells and pericytes of the cortical-medullary region [23] that control EPO gene expression, primarily through the hypoxia-inducible factor (HIF) pathway [24]. EPO acts as the primary hormone regulator of erythrocyte production or erythropoiesis in the bone marrow. It is also synthesised by the liver [25] and the brain; however, the amount synthesised by these tissues alone is insufficient to maintain adequate erythropoiesis [26]. According to Nangaku et al. [27], EPO deficiency may be caused by constant inflammatory cell infiltration. At the cellular level, the production of EPO in REPs is regulated by a number of EPO-producing cells via an "on" or "off" mechanism, which changes explicitly in response to hypoxia and or anaemia. Under normal conditions, REPs possess an EPO-producing ability but do not produce EPO (OFF-REPs). In healthy individuals, when oxygen supplies decrease or oxygen demands rise, OFF-REPs begin to produce EPO through HIF-mediated EPO gene transcription (ON-REPs) [28]. In CKD patients, however, sustained inflammation in renal fibrosis becomes the major contributor to EPO deficiency, as the damaged REPs in fibrotic kidneys transform into myofibroblasts, causing the concomitant repression of their ability to produce EPO [28].

5. Pathophysiology of Inflammation and Linked Genetic Factors

The serum erythropoietin levels in anaemic patients without renal failure but with inflammation due to other factors were found to be lower compared to these levels in similarly anaemic patients without inflammation, demonstrating the relationship between inflammation and impaired erythropoiesis [29,30]. Signalling mediators of pro-inflammatory and pro-fibrotic pathways, such as GATA-2 (binding factor), nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B), and transforming growth factor beta (TGF- β)/Smad, often play a role in the direct and indirect suppression of the EPO-regulatory mechanism [30,31].

The GATA family transcription factor GATA-2 binds to the GATA motif in the vicinity of the transcriptional initiation site of the EPO promoter in the -30 region [32]. Some evidence has shown that the functional activation of the GATA-2 signalling pathway by pro-inflammatory cytokines leads to the impairment of EPO production. Observations from one study revealed that GATA-2 suppressed EPO gene expression in cell cultures. When the TATA element was transfected, EPO gene promoter activity rose 2.5-fold; however, pro-inflammatory cytokines decreased EPO gene promoter activity in cells transfected with pGATAwt relative to control cells [31]. This result suggests that the signal transduction pathways of these cytokines may be modulated by GATA-2, inducing lower EPO production under inflammatory conditions.

NF κ B signalling is also responsible for the negative regulation of EPO gene transcription via competition with co-activators of hypoxia-induced EPO gene expression [33]. The activity of transcription factor NF- κ B is closely related to the processes of activation, proliferation, and cell differentiation into myofibroblasts [28]. Souma et al. [28] revealed that the sustained activation of NF κ B signals in renal erythropoietin-producing cells results in a phenotypic transition. These findings show that although the NF κ B signal is important with regard to repressing erythropoietin production, it is also associated with transforming EPCs into myofibroblasts.

Another major signalling pathway involved in the pathogenesis of fibrosis with the concomitant loss of EPO is the TGF- β /Smad3 pathway [34]. Activated TGF- β functions via Smad-dependent and -independent signalling pathways, which are major pathways in many pathophysiological processes of kidney diseases [35]. It has been well documented that Smad3 is a strong downstream mediator of renal fibrosis in diabetic nephropathy [36], hypertensive nephropathy [37], obstructive kidney disease, and glomerulonephritis [38]. In contrast, Smad2 acts as a reno-protective agent by competing with Smad3 signalling through phosphorylation and nuclear translocation [37].

Elevated levels of pro-inflammatory cytokines and inflammation-related indicators characterise renal fibrosis in CKD and the loss of EPO production [39]. Gene transcription and cytokine release may be affected by cytokine gene polymorphisms, which could modulate the risk of renal fibrosis and EPO deficiency anaemia progression [40]. As shown in Figure 1, Yan and Xu summarised the pathophysiology of the inflammatory cytokines that cause anaemia and EPO resistance [10].

The interleukin-6 (IL-6) gene is found on chromosome 7p21 and consists of five exons and four introns. IL-6 has several polymorphisms in the following promoter regions: -174 G to C and -597 G to A. IL-6 is rapidly expressed in a highly transient manner during inflammation. Mutations in rs2228145 have been associated with renal disease due to inflammation, and they also cause renal fibrosis [41]. A previous study on dialysis patients conducted by Sharples et al. [42] showed the influence of an IL-6 (-174 G/C) polymorphism on ESAs' responsiveness. They observed that there was a significantly higher ESA requirement in the GG and GC ACE genotypes compared with the CC group, which remained an independent association.

Tumour necrosis factor (TNF) is a pro-inflammatory cytokine produced by immune cells and adipocytes. In CKD, there is increased production of TNF as a result of inflammation and oxidative stress. Elevated TNF levels can have multiple effects on the body, including the suppression of EPO production by the kidneys [12]. TNF- α inhibits the

production of EPO in the kidneys by interfering with the transcription of the EPO gene and by promoting the destruction of EPO-producing cells. This inhibitory effect of TNF on EPO production contributes to the development of renal anaemia in CKD patients [43].

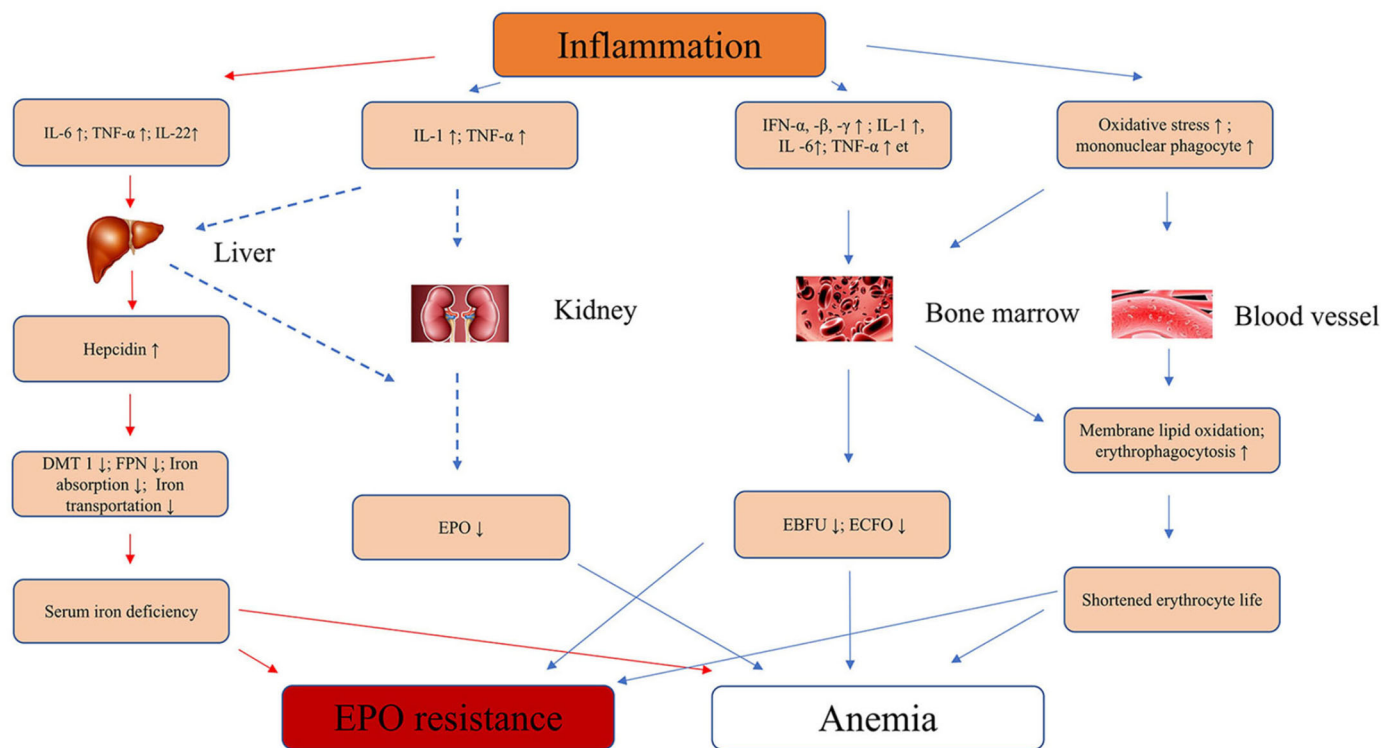


Figure 1. Inflammatory cytokines leading to anaemia. IL-6, interleukin-6; IL-1, interleukin-1; IFN- α , - β , - γ , interferon- α , - β , - γ ; TNF- α , tumour necrosis factor- α ; EBFU, erythroid burst-forming units; ECFO, erythroid colony-forming units; ↑, increase; ↓, decrease; FNP, ferroportin; DMT1, divalent metal transporter 1 ([12] (permission obtained)).

The IL-1 gene cluster on chromosome 2q contains three related genes within a 430-kb region, IL-1 α , IL-1B, and IL-1RN, which encode the pro-inflammatory cytokines IL-1 α and IL-1 β . The IL-1 β -511C/T (rs1143627) single-nucleotide polymorphism (SNP) has been associated with a variety of diseases in which inflammation plays an important role [44]. Jeong et al. [45] reported that the IL-1 β -511CC genotype was significantly associated with lower ERI values among haemodialysis patients. In this study, patients with the IL-1 β -511 TT genotype also showed a significantly higher mean IL-1 β level (0.9 ± 0.6 pg/mL) compared to those with the IL-1 β -511CC genotype (0.3 ± 0.3 pg/mL; $p = 0.02$) [45].

6. Erythropoietin Treatment and Resistance

According to the KDIGO guidelines (2013) [46], it is recommended that erythropoietin-stimulating agent (ESA) treatment should be considered for anaemic patients with CKD who are not on dialysis when their haemoglobin level is <10 g/dL. This treatment must be individualised based on the rate of decrease in haemoglobin levels, prior response to iron therapy, the likely requirement for a transfusion, the risks related to ESA therapy, and the presence of symptoms attributable to anaemia [47]. A third-generation ESA, darbepoetin alpha, which is modified from EPO via the insertion of a large pegylation chain (called the continuous EPO receptor activator) that increases the duration of the drug's effects, has been approved for sale on the market [48]. Although ESAs have been found to improve patients' quality of life, reduced haematopoietic responses to ESA treatment have been associated with an increased risk of adverse outcomes [49,50]. Patients with heart conditions are often associated with higher erythropoietin resistance [49,51]. Safety concerns regarding

ESA therapies have paved the way for the development of a new class of drugs. Table 1 lists the current treatment approaches [8].

Table 1. Summary of trials focusing on newer treatment approaches.

Future Treatment Improvement Focus	Author/Year	Country of Study	Study Population, Sample Size (n)	Results		
Clinical Trials						
Comparison between weekly single dose and divided moderate dose of rHuEPO	Xiuling et al. [52]	China	Haemodialyzed (n = 88)	There was no significant difference in terms of medication safety in both groups		
HIF-PH inhibition according to blood group	Funakoshi [53]	Japan	Haemodialyzed (n = 163)	Treatment Roxadustat Daprodustat	Blood group A O	Efficacy% 47 55
Basic Experiments						
Selective inhibition of PH2	Su et al. [54]	China	In vivo (n = 25)	Ongoing trial		
Ferroptosis as a potential therapeutic target in CKD	Wang et al. [55]	China	Mouse model (n = 24)	1. Dysfunctional iron metabolism is an important contributor to ferroptosis. 2. Ferritinophagy was observed among CKD-afflicted rats. 3. Regulation of iron metabolism and TGF-β1/Smad3 pathway can interfere with the progression of renal damage.		

7. Precision Medicine in EPO Deficiency and Treatment Resistance

There has been a substantial amount of interest in creating a gene therapy approach with which to deliver EPO via a single infusion of the EPO gene and thus guarantee EPO delivery in the long term. In one attempt, investigators managed to establish a hypoxia-regulatory mechanism that was in line with the homeostatic system, thus characterising it as a natural approach. They also revealed that the promotor OBHRE region increased the haematocrit level gradually and safely in a relevant anaemic mouse model [56].

In a different approach, EPO from human cells (hEPO) was manufactured to develop a method whereby EPO could be directly secreted without any further formulation, to reduce the risk of antibody production. This study was carried out by Lippin et al. [57] using a Biopump transduced with clinical-grade Ad-MG/EPO-1 (Ad5 E1/E3 deleted) expressing hEPO, yielding a promising result.

However, it is important to note that gene therapy is still an emerging field and is not yet widely available for clinical use. Clinical trials are currently underway to evaluate the safety and efficacy of gene therapy with respect to various genetic conditions, including EPO deficiency caused by mutations in the EPO gene.

8. Genetic Factors of EPO Deficiency and Treatment Resistance

The human EPO gene is located on chromosome 7q21, which is well known for harbouring increased susceptibility to diabetic nephropathy [58]. Endogenous and recombinant EPO stimulates erythropoiesis by binding to the erythropoietin receptor (EpoR). Messenger RNA (mRNA) alternative splicing can give rise to the soluble form of the receptor (sEpoR), which lacks a transmembrane domain. sEpoR acts as an antagonist of EPO due to its higher affinity for EPO, which can lead to resistance towards ESA. Moreover, in vitro experiments conducted by Tong et al. [59] showed that a single-nucleotide polymorphism from G to T in the EPO promoter (rs1617640) could alter EPO mRNA levels [60]. A predisposition in this promoter region might also contribute to EPO deficiency in pre-dialysis patients and, therefore, needs to be investigated.

Pro-inflammatory genes are widely studied with respect to their role in anaemia associated with chronic diseases and are known to cause ESA resistance [61]. Jeong et al. [45]

reported that the IL-1 β -511CC genotype was significantly associated with lower erythropoietin resistance values in haemodialysis patients. A polymorphism in this region was shown to reduce EPO mRNA expression and erythropoietin secretion in human hepatoma cell lines. According to Nangaku and Eckardt [27], this EPO deficiency may be due to constant inflammatory cell infiltration that leads to the decreased production of EPO. The roles of several pathways in inducing inflammatory conditions in renal fibrosis, which eventually leads to EPO deficiency anaemia, have been identified (Figure 2).

Hypoxia-inducible factor (HIF) was first discovered during the identification of a hypoxia-responsive element (HRE) in the erythropoietin gene in 1991 [62]. Besides regulating oxygen via signalling the EPO gene, it also plays a role in stem cell maintenance, growth factor signalling, epithelial–mesenchymal transition, angiogenesis, and metabolism [63,64]. Under normoxic conditions, HIF is rapidly ubiquitinated and degraded, a process primarily controlled by a family of oxygen-dependent prolyl hydroxylases (PH). However, hypoxia triggers the stabilisation of HIF-1 α , which is then translocated from the cytoplasm to the nucleus and heterodimerises with HIF-1 β , which protects it from Von-Hippel–Lindau (VHL)-mediated proteasomal degradation. The transcriptionally active HIF complex formed via heterodimerisation associates with HRE in the regulatory regions of target genes and binds to transcriptional coactivators to induce EPO gene expression [65,66].



Figure 2. Pleiotropic effects of prolyl hydroxylase (PH) inhibition. HIF, hypoxia-inducible factor. Improved iron [67,68]; improved lipid profile [69]; glucose profile [52]; protective against kidney injury [70]; development of cancer [71], cardiovascular risk [72]; inflammation [73,74].

9. Hypoxia-Inducible Factor Prolyl Hydroxylase (HIF-PH) Inhibitors

HIF-PH inhibitors target the activity of prolyl hydroxylase (PH) enzymes, which play a key role in regulating the stability and activity of HIF [75]. PHs belong to the iron and α -ketoglutarate (α -KG)-dependent dioxygenase superfamily, and they have several identified isoforms, namely PH 1, 2, and 3 [76]. PHs exhibit overlapping but different tissue expression levels, and their subcellular localisation varies [67]. PH 2 is often associated with HIF-1 α in normoxia regulation, and any loss of function relating to PH 2 causes familial erythrocytosis [68]. PH 3, on the other hand, is the regulator of HIF-2 α and is found to be overexpressed in hypoxic conditions [71]. Several recent studies have investigated

the potential use of HIF-PH inhibitors as a treatment for CKD [77–79]. By blocking the activity of these enzymes, HIF-PH inhibitors can increase the activity of HIF and trigger cellular responses to low oxygen levels that regulate and stimulate erythrocyte production, potentially alleviating symptoms of anaemia.

CKD is associated with inflammation, which, in turn, causes functional iron deficiency and ESA hypo-responsiveness. HIF is believed to improve these two factors since it plays a crucial role in the regulation of cellular responses to low oxygen levels and reportedly reduces serum hepcidin levels [79]. Moreover, HIF-PH inhibitors were reported to decrease fibroblast growth factor (FGF)-23 levels in an animal model of CKD [80]. It was also noted by researchers that HIF-1 α and its isoform 2 α play a role in clear cell renal carcinoma (ccRC), but their functional significance is not clear in the developmental stage [81]. An earlier investigation revealed that tumour cells often present the overexpression of HIF-1 α . The overexpression of either isoform was also associated with cardiomyopathy [72]. However, recent evidence shows that these two isoforms have opposite outcomes [82]. The pleiotropic effects (Figure 2) of HIF have also led to the investigation of metabolic changes; for instance, an improvement in glucose tolerance and insulin sensitivity in a mouse model was observed by Sugahara et al. [83]. Both isoforms of HIF have been shown to be expressed in innate immunity [73], while HIF-1 α has been revealed to be expressed in adaptive immunity [74].

In clinical trials, roxadustat (FG-4592) users experienced higher rates of upper respiratory infections, pneumonia, and urinary tract infections than the control group, which may have been related to the immunological regulation brought about by PH inhibition [5]. While it is true that HIF-PHIs have been well tolerated in clinical trials, there may still be some concerns regarding copper toxicity. Nakamura et al. [84] described four cases of individuals with renal anaemia who had elevated serum copper levels. Research on treatment with HIF-PHI medication, whose administration regimen was returned to normal after switching from HIF-PHI to darbepoetin alfa, indicated that HIF-PHI may be linked to elevated serum copper levels. Since a genome-wide analysis of the HIF transcriptome has shown that at least 500–1000 genes are under the control of HIF, stabilising this factor to improve the oxygen-sensing mechanisms might affect other complex pathophysiological characteristics in the long term.

In phase 3 trials, namely OLYMPUS and ROCKIES, Fishbane [5] reported that, regardless of inflammation, roxadustat improved haemoglobin levels in comparison to a placebo and an epotin alfa group. It was also proven to promote intestinal iron absorption and hepcidin reduction [85]. However, extensive clinical trials are required to study the long-term safety profile of roxadustat with respect to CKD inflammatory anaemia.

Daprodustat (GSK-1278863) is another potent HIF-PH 1–3 inhibitor, and it was first approved in Japan for renal anaemia treatment in June 2020 (GlaxoSmithKline) [86]. However, this molecule is still under investigation in many countries (NCT03029247; 205665; ASCEND-BP and NCT03457701; 201771; ASCEND-Fe); the FDA, after some hesitation, finally approved it in February 2023. Several other HIF-PH inhibitors are being developed and studied in order to target several patient cohorts—for example, dialysed and dependent patients.

Vadadustat (AKB-6548), developed by Akebia Therapeutics, inhibits all three PHs, with increased affinity towards PHD3 [75]. Vadadustat (Vafseo[®]) received marketing authorisation from the European Commission and the United Kingdom Medicines and Healthcare Products Regulatory Agency in April 2023 and May 2023, respectively. It is currently being used in 33 nations, including Japan. In the PRO₂TECT trial (NCT02648347 and NCT02680574), this drug did not meet its pre-specified safety endpoint in a non-dialysed patient cohort [87]. However, it showed good efficacy in terms of increasing haemoglobin levels, reducing serum ferritin and hepcidin levels, and improving TIBC [88].

Molidustat (BAY 85-3934) and enarodustat (JTZ-951) are also being tested in phase III clinical trials by different pharmaceutical companies, and they have presented considerable efficacy and safety profiles in phase 2 trials. Several review articles have summarised

the phase 3 clinical trials evaluating the efficacy of HIF stabilisers for the non-dialysis population (pertinent tables can be found in [89,90]). In the SYMPHONY ND trial, no adverse events occurred in the enarodustat arm; embolic and thrombotic events were previously observed in a Chuvash polycythemia patient. Retinal disorders were somewhat frequent. A limitation of this trial was its short duration, i.e., 24 weeks [91]. Although increased serum vascular endothelial growth factor (VEGF) levels were observed in the molidustat (Varenzin-CA1) group, the enarodustat (ENAROY)-treated patient did not exhibit the same trend [92].

10. Future Treatment Approaches

The newest molecules currently being researched are known as Tetrahydropyridin-4-ylpicolinoylglycines, which are intended to inhibit PH-2 [54]. This targeted approach might improve the selective binding and reduce some of the undesirable pleiotropic effects of HIF stabilisers. In the context of personalised medicine, an investigation of the association between an ABO blood group and HIF PH showed that roxadustat had better therapeutic efficacy for the A blood group, while daprodustat was more effective for the O blood group, with regard to haemodialyzed individuals [53]. While recombinant EPO treatment is not entirely disadvantageous for all patients, a more personalised approach to determining the treatment target would be beneficial. For instance, according to Joksimovic Jovic et al. [93], elevated serum ferritin levels and increased catalase activity contribute to ephemeral EPO resistance. Oxidative stress, poor nutrition, chronic inflammation, and vitamin D deficiency increase the risk of long-acting EPO resistance. Kidney cells are prone to iron overload, which causes ferroptosis, i.e., regulated cell death due to iron overload. This aggravates CKD and renal anaemia, leading to irregular iron metabolism [94]. It is believed that anaemia in CKD develops as the number of renal EPO-producing cells and the production of fibroblast-derived erythropoietin are decreased despite the tissue hypoxia caused by anaemia. As a result, EPO deficiency anaemia becomes the major cause of anaemia in CKD [23]. Wang et al. [55] attempted to demonstrate that ferroptosis could be a novel target approach in delaying CKD that eventually leads to renal anaemia.

11. Potential Approach from the Perspective of Drug Design and Development to Addressing Treatment Resistance in Erythropoietin Deficiency Anaemia: Receptor Modification

Receptor modification can occur through a variety of mechanisms, such as changes in a receptor's gene expression, post-translational modifications (such as phosphorylation, glycosylation, or acetylation), or alterations in a receptor's structure or conformation. These modifications can affect a receptor's sensitivity or specificity to certain ligands (molecules that bind to a receptor), its signalling efficiency or duration, or even its ability to interact with other proteins or downstream effectors. For example, the EpoR can be engineered to have a higher affinity for EPO, allowing it to bind to EPO more effectively and stimulate erythropoiesis more efficiently. Alternatively, the EpoR can be modified to have a longer half-life, which would prolong its activity and allow it to stimulate erythropoiesis for a longer period. Computational techniques are being used to model the erythropoietin receptor and perform molecular simulations. These simulations could help to identify potential binding sites, predict conformational changes, and assess the impacts of specific modifications on receptor function [95].

12. Conclusions

EPO deficiency has a complex mechanism that involves many pathways. Comorbidities such as dyslipidaemia, hypertension, and diabetes are also contributing factors in this condition. In addition, concomitant drugs, such as ACE inhibitors, statins, diuretics, anti-platelets, oral diabetic agents, and so on, are also widely investigated with respect to their roles as potential biomarkers. Identified or known pathways are being extensively researched to provide more personalised treatments and achieve good treatment efficiency. For example, drugs that target specific molecular pathways involved in EPO production

or responses are being investigated. These targeted therapies may be identified through genetic testing and personalised treatment plans. The investigation of biomarkers is also on the rise, and genetic contributions to pathophysiology should also be investigated. Some genetic polymorphisms and expression patterns may result in variations in individual responses to ESA treatment. Administering EPO based on the physiological demand could also reduce adverse events and unnecessary costs for patients.

Author Contributions: Conceptualisation, N.Y., B.M.A. and H.Z.H.; writing—original draft preparation, N.Y.; writing—review and editing, B.M.A., H.Z.H., S.K.L., A.H.B.A.G., M.G.W., S.B. and H.S.W.; supervision, H.Z.H. and S.K.L.; project administration, H.Z.H. and S.K.L.; funding acquisition, H.Z.H., S.K.L., A.H.B.A.G., M.G.W., S.B. and H.S.W. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Fundamental Research Grant Scheme (FRGS), FP020-2016.

Conflicts of Interest: The authors have no conflict of interest to declare.

References

1. Bonomini, M.; Del Vecchio, L.; Sirolli, V.; Locatelli, F. New Treatment Approaches for the Anemia of CKD. *Am. J. Kidney Dis.* **2016**, *67*, 133–142. [[CrossRef](#)] [[PubMed](#)]
2. Drüeke, T.B.; Massy, Z.A. Erythropoiesis-Stimulating Agents and Mortality. *J. Am. Soc. Nephrol.* **2019**, *30*, 907–908. [[CrossRef](#)] [[PubMed](#)]
3. Lee, K.-H.; Ho, Y.; Tarng, D.-C. Iron Therapy in Chronic Kidney Disease: Days of Future Past. *Int. J. Mol. Sci.* **2021**, *22*, 1008. [[CrossRef](#)]
4. Batchelor, E.K.; Kapitsinou, P.; Pergola, P.E.; Kovesdy, C.P.; Jalal, D.I. Iron Deficiency in Chronic Kidney Disease: Updates on Pathophysiology, Diagnosis, and Treatment. *J. Am. Soc. Nephrol.* **2020**, *31*, 456–468. [[CrossRef](#)]
5. Fishbane, S.; El-Shahawy, M.A.; Pecoits-Filho, R.; Pham Van, B.; Houser, M.T.; Frison, L.; Little, D.J.; Guzman, N.J.; Pergola, P.E. OLYMPUS: A Phase 3, Randomized, Double-Blind, Placebo-Controlled, International Study of Roxadustat Efficacy in Patients with Non-Dialysis-Dependent (NDD) CKD and Anemia [Abstract TH-OR023]. *J. Am. Soc. Nephrol.* **2019**, *30*, 6.
6. Barratt, J.; Andric, B.; Tataradze, A.; Schömig, M.; Reusch, M.; Valluri, U.; Mariat, C. Roxadustat for the treatment of anaemia in chronic kidney disease patients not on dialysis: A Phase 3, randomized, open-label, active-controlled study (DOLOMITES). *Nephrol. Dial. Transplant.* **2021**, *36*, 1616–1628. [[CrossRef](#)] [[PubMed](#)]
7. Imamura, T.; Ueno, Y.; Kinugawa, K. Impact of Hypoxia-Inducible Factor Prolyl Hydroxylase Inhibitor on Renal Function in Patient with Heart Failure. *J. Cardiovasc. Dev. Dis.* **2021**, *8*, 189. [[CrossRef](#)]
8. Haase, V.H. Hypoxia-inducible factor–prolyl hydroxylase inhibitors in the treatment of anemia of chronic kidney disease. *Kidney Int. Suppl.* **2021**, *11*, 8–25. [[CrossRef](#)]
9. Scott, S.A. Personalizing medicine with clinical pharmacogenetics. *Genet. Med.* **2011**, *13*, 987–995. [[CrossRef](#)]
10. Oates, J.T.; Lopez, D. Pharmacogenetics: An Important Part of Drug Development with A Focus on Its Application. *Int. J. Biomed. Investig.* **2018**, *1*, 111.
11. Lee, H.H.; Ho, R.H. Interindividual and interethnic variability in drug disposition: Polymorphisms in organic anion transporting polypeptide 1B1 (OATP1B1; SLCO1B1). *Br. J. Clin. Pharmacol.* **2017**, *83*, 1176–1184. [[CrossRef](#)]
12. Yan, Z.; Xu, G. A Novel Choice to Correct Inflammation-Induced Anemia in CKD: Oral Hypoxia-Inducible Factor Prolyl Hydroxylase Inhibitor Roxadustat. *Front. Med.* **2020**, *7*, 393. [[CrossRef](#)]
13. Ueda, N.; Takasawa, K. Impact of Inflammation on Ferritin, Hepcidin and the Management of Iron Deficiency Anemia in Chronic Kidney Disease. *Nutrients* **2018**, *10*, 1173. [[CrossRef](#)]
14. Camaschella, C. Iron deficiency. *Blood* **2019**, *133*, 30–39. [[CrossRef](#)] [[PubMed](#)]
15. Campodonico, J.; Nicoli, F.; Motta, I.; Migone De Amicis, M.; Bonomi, A.; Cappellini, M.; Agostoni, P. Prognostic role of transferrin saturation in heart failure patients. *Eur. J. Prev. Cardiol.* **2021**, *28*, 1639–1646. [[CrossRef](#)] [[PubMed](#)]
16. Pergola, P.E.; Kopyt, N.P. Oral Ferric Maltol for the Treatment of Iron-Deficiency Anemia in Patients With CKD: A Randomized Trial and Open-Label Extension. *Am. J. Kidney Dis.* **2021**, *78*, 846–856.e841. [[CrossRef](#)] [[PubMed](#)]
17. Tolkien, Z.; Stecher, L.; Mander, A.P.; Pereira, D.I.A.; Powell, J.J. Ferrous Sulfate Supplementation Causes Significant Gastrointestinal Side-Effects in Adults: A Systematic Review and Meta-Analysis. *PLoS ONE* **2015**, *10*, e0117383. [[CrossRef](#)] [[PubMed](#)]
18. Schmidt, C.; Allen, S.; Kopyt, N.; Pergola, P. Iron Replacement Therapy with Oral Ferric Maltol: Review of the Evidence and Expert Opinion. *J. Clin. Med.* **2021**, *10*, 4448. [[CrossRef](#)]
19. Barish, C.F.; Koch, T.; Butcher, A.; Morris, D.; Bregman, D.B. Safety and Efficacy of Intravenous Ferric Carboxymaltose (750 mg) in the Treatment of Iron Deficiency Anemia: Two Randomized, Controlled Trials. *Anemia* **2012**, *2012*, 172104. [[CrossRef](#)]
20. Charytan, C.; Bernardo, M.V.; Koch, T.A.; Butcher, A.; Morris, D.; Bregman, D.B. Intravenous ferric carboxymaltose versus standard medical care in the treatment of iron deficiency anemia in patients with chronic kidney disease: A randomized, active-controlled, multi-center study. *Nephrol. Dial. Transplant.* **2013**, *28*, 953–964. [[CrossRef](#)]

21. Boots, J.M.M.; Quax, R.A.M. High-Dose Intravenous Iron with Either Ferric Carboxymaltose or Ferric Derisomaltose: A Benefit-Risk Assessment. *Drug Saf.* **2022**, *45*, 1019–1036. [[CrossRef](#)] [[PubMed](#)]
22. Pollock, R.F.; Biggar, P. Indirect methods of comparison of the safety of ferric derisomaltose, iron sucrose and ferric carboxymaltose in the treatment of iron deficiency anemia. *Expert Rev. Hematol.* **2020**, *13*, 187–195. [[CrossRef](#)] [[PubMed](#)]
23. Koury, M.J.; Haase, V.H. Anaemia in kidney disease: Harnessing hypoxia responses for therapy. *Nat. Rev. Nephrol.* **2015**, *11*, 394–410. [[CrossRef](#)] [[PubMed](#)]
24. Stolze, I.; Berchner-Pfannschmidt, U.; Freitag, P.; Wotzlaw, C.; Rössler, J.; Frede, S.; Acker, H.; Fandrey, J. Hypoxia-inducible erythropoietin gene expression in human neuroblastoma cells. *Blood* **2002**, *100*, 2623–2628. [[CrossRef](#)]
25. Koury, M.J.; Bondurant, M.C. Maintenance by erythropoietin of viability and maturation of murine erythroid precursor cells. *J. Cell. Physiol.* **1988**, *137*, 65–74. [[CrossRef](#)]
26. Molineux, G.; Foote, M.A.; Elliott, S.G. *Erythropoietins and Erythropoiesis: Molecular, Cellular, Preclinical, and Clinical Biology*; Birkhäuser: Basel, Switzerland, 2005.
27. Nangaku, M.; Eckardt, K.-U. Pathogenesis of Renal Anemia. *Semin. Nephrol.* **2006**, *26*, 261–268. [[CrossRef](#)]
28. Souma, T.; Yamazaki, S.; Moriguchi, T.; Suzuki, N.; Hirano, I.; Pan, X.; Minegishi, N.; Abe, M.; Kiyomoto, H.; Ito, S.; et al. Plasticity of Renal Erythropoietin-Producing Cells Governs Fibrosis. *J. Am. Soc. Nephrol.* **2013**, *24*, 1599–1616. [[CrossRef](#)]
29. Baer, A.N.; Dessypris, E.N.; Goldwasser, E.; Krantz, S.B. Blunted erythropoietin response to anaemia in rheumatoid arthritis. *Br. J. Haematol.* **1987**, *66*, 559–564. [[CrossRef](#)]
30. Miller, C.B.; Jones, R.J.; Piantadosi, S.; Abeloff, M.D.; Spivak, J.L. Decreased Erythropoietin Response in Patients with the Anemia of Cancer. *N. Engl. J. Med.* **1990**, *322*, 1689–1692. [[CrossRef](#)]
31. La Ferla, K.; Reimann, C.; Jelkmann, W.; Hellwig-Bürgel, T. Inhibition of erythropoietin gene expression signaling involves the transcription factors GATA-2 and NF- κ B. *FASEB J.* **2002**, *16*, 1–17. [[CrossRef](#)]
32. Roach, K.M.; Duffy, S.M.; Coward, W.; Feghali-Bostwick, C.; Wulff, H.; Bradding, P. The K⁺ Channel KCa3.1 as a Novel Target for Idiopathic Pulmonary Fibrosis. *PLoS ONE* **2014**, *8*, e85244. [[CrossRef](#)]
33. Batmunkh, C.; Krajewski, J.; Jelkmann, W.; Hellwig-Bürgel, T. Erythropoietin production: Molecular mechanisms of the antagonistic actions of cyclic adenosine monophosphate and interleukin-1. *FEBS Lett.* **2006**, *580*, 3153–3160. [[CrossRef](#)] [[PubMed](#)]
34. Böttinger, E.P. TGF- β in Renal Injury and Disease. *Semin. Nephrol.* **2007**, *27*, 309–320. [[CrossRef](#)]
35. Derynck, R.; Zhang, Y.E. Smad-dependent and Smad-independent pathways in TGF- β family signalling. *Nature* **2003**, *425*, 577–584. [[CrossRef](#)] [[PubMed](#)]
36. Chung, A.C.K.; Zhang, H.; Kong, Y.-Z.; Tan, J.-J.; Huang, X.R.; Kopp, J.B.; Lan, H.Y. Advanced Glycation End-Products Induce Tubular CTGF via TGF- β -Independent Smad3 Signaling. *J. Am. Soc. Nephrol.* **2010**, *21*, 249–260. [[CrossRef](#)]
37. Wang, W.; Huang, X.R.; Canlas, E.; Oka, K.; Truong, L.D.; Deng, C.; Bhowmick, N.A.; Ju, W.; Bottinger, E.P.; Lan, H.Y. Essential Role of Smad3 in Angiotensin II-Induced Vascular Fibrosis. *Circ. Res.* **2006**, *98*, 1032–1039. [[CrossRef](#)]
38. Huang, X.R.; Chung, A.C.K.; Zhou, L.; Wang, X.J.; Lan, H.Y. Latent TGF- β 1 Protects Against Crescentic Glomerulonephritis. *J. Am. Soc. Nephrol.* **2008**, *19*, 233–242. [[CrossRef](#)]
39. Meng, X.-M. Inflammatory Mediators and Renal Fibrosis. In *Renal Fibrosis: Mechanisms and Therapies*; Liu, B.-C., Lan, H.-Y., Lv, L.-L., Eds.; Springer: Singapore, 2019; pp. 381–406.
40. Petreski, T.; Piko, N.; Ekart, R.; Hojs, R.; Bevc, S. Review on Inflammation Markers in Chronic Kidney Disease. *Biomedicines* **2021**, *9*, 182. [[CrossRef](#)]
41. Zoccali, C.; Mallamaci, F. Innate Immunity System in Patients With Cardiovascular and Kidney Disease. *Circ. Res.* **2023**, *132*, 915–932. [[CrossRef](#)]
42. Sharples, E.J.; Varagunam, M.; Sinnott, P.J.; McCloskey, D.J.; Raftery, M.J.; Yaqoob, M.M. The Effect of Proinflammatory Cytokine Gene and Angiotensin-Converting Enzyme Polymorphisms on Erythropoietin Requirements in Patients on Continuous Ambulatory Peritoneal Dialysis. *Perit. Dial. Int.* **2006**, *26*, 64–68. [[CrossRef](#)]
43. Yadav, P.; Divvi, V.S.S.R.; Shah, T. Assessment of Cytokine (α -TNF) with Erythropoietin and their Correlation in Pulmonary Tuberculosis with Anaemia. *J. Pharm. Res. Int.* **2021**, *33*, 1–9. [[CrossRef](#)]
44. Glas, J.; Török, H.-P.; Schneider, A.; Brännler, G.; Kopp, R.; Albert, E.D.; Stolte, M.; Folwaczny, C. Allele 2 of the Interleukin-1 Receptor Antagonist Gene Is Associated With Early Gastric Cancer. *J. Clin. Oncol.* **2004**, *22*, 4746–4752. [[CrossRef](#)] [[PubMed](#)]
45. Jeong, K.-H.; Lee, T.-W.; Ihm, C.-G.; Lee, S.-H.; Moon, J.-Y. Polymorphisms in two genes, IL-1B and ACE, are associated with erythropoietin resistance in Korean patients on maintenance hemodialysis. *Exp. Mol. Med.* **2008**, *40*, 161–166. [[CrossRef](#)] [[PubMed](#)]
46. KDIGO. KDIGO Clinical Practice Guideline for the Evaluation and Management of Chronic Kidney Disease. *Kidney Int. Suppl.* **2013**, *3*, 5–14.
47. Locatelli, F.; Bárány, P.; Covic, A.; De Francisco, A.; Del Vecchio, L.; Goldsmith, D.; Hörl, W.; London, G.; Vanholder, R.; Van Biesen, W.; et al. Kidney Disease: Improving Global Outcomes guidelines on anaemia management in chronic kidney disease: A European Renal Best Practice position statement. *Nephrol. Dial. Transplant.* **2013**, *28*, 1346–1359. [[CrossRef](#)]
48. Hayat, A.; Haria, D.; Salifu, M.O. Erythropoietin stimulating agents in the management of anemia of chronic kidney disease. *Patient Prefer. Adherence* **2008**, *2*, 195–200.

49. López-Gómez, J.M.; Portolés, J.M.; Aljama, P. Factors that condition the response to erythropoietin in patients on hemodialysis and their relation to mortality: New strategies to prevent cardiovascular risk in chronic kidney disease. *Kidney Int.* **2008**, *74*, S75–S81. [\[CrossRef\]](#)
50. Rossert, J.; Gassmann-Mayer, C.; Frei, D.; McClellan, W. Prevalence and predictors of epoetin hyporesponsiveness in chronic kidney disease patients. *Nephrol. Dial. Transplant.* **2007**, *22*, 794–800. [\[CrossRef\]](#)
51. Guerrero Riscos, M.Á.; Guerrero-Riscos, M.Á.; Montes Delgado, R.; Montes-Delgado, R.; Seda Guzmán, M.; Seda-Guzmán, M.; Praena-Fernández, J.M.; Praena-Fernández, J.M. Erythropoietin resistance and survival in non-dialysis patients with stage 4–5 chronic kidney disease and heart disease. *Nefrología* **2012**, *32*, 343–352.
52. Xiuling, W.; Jianjun, L.; Ying, Y.; Rong, X.; Lu, W.; Xuedong, W.; Fubin, T. Safety of Weekly Single versus Divided Administration of Moderate-dose Erythropoietin in the Treatment of Maintenance Hemodialysis Patients with Renal Anemia. *Chin. Gen. Pract.* **2023**, *26*, 711–717.
53. Funakoshi, S. Difference in Therapeutic Effects between Roxadustat and Daprodustat, HIF-Ph Inhibitors, Depending on the Blood Type in Hemodialysis (HD) Patients. *Blood* **2021**, *138*, 4147. [\[CrossRef\]](#)
54. Su, K.; Li, Z.; Zhang, L.; Fang, S.; Mao, M.; Sun, Z.; Zhang, X. Tetrahydropyridin-4-ylpicolinoylglycines as novel and orally active prolyl hydroxylase 2 (PHD2) inhibitors for the treatment of renal anemia. *Eur. J. Med. Chem.* **2022**, *238*, 114479. [\[CrossRef\]](#) [\[PubMed\]](#)
55. Wang, J.; Wang, Y.; Liu, Y.; Cai, X.; Huang, X.; Fu, W.; Wang, L.; Qiu, L.; Li, J.; Sun, L. Ferroptosis, a new target for treatment of renal injury and fibrosis in a 5/6 nephrectomy-induced CKD rat model. *Cell Death Discov.* **2022**, *8*, 127. [\[CrossRef\]](#) [\[PubMed\]](#)
56. Binley, K.; Askham, Z.; Iqbal, S.; Spearman, H.; Martin, L.; de Alwis, M.; Thrasher, A.J.; Ali, R.R.; Maxwell, P.H.; Kingsman, S.; et al. Long-term reversal of chronic anemia using a hypoxia-regulated erythropoietin gene therapy. *Blood* **2002**, *100*, 2406–2413. [\[CrossRef\]](#)
57. Lippin, Y.; Dranitzki-Elhalel, M.; Brill-Almon, E.; Mei-Zahav, C.; Mizrahi, S.; Liberman, Y.; Iaina, A.; Kaplan, E.; Podjarny, E.; Zeira, E.; et al. Human erythropoietin gene therapy for patients with chronic renal failure. *Blood* **2005**, *106*, 2280–2286. [\[CrossRef\]](#)
58. Iyengar, S.K.; Abboud, H.E.; Goddard, K.A.B.; Saad, M.F.; Adler, S.G.; Arar, N.H.; Bowden, D.W.; Duggirala, R.; Elston, R.C.; Hanson, R.L.; et al. Genome-Wide Scans for Diabetic Nephropathy and Albuminuria in Multiethnic Populations: The Family Investigation of Nephropathy and Diabetes (FIND). *Diabetes* **2007**, *56*, 1577–1585. [\[CrossRef\]](#)
59. Tong, Z.; Yang, Z.; Patel, S.; Chen, H.; Gibbs, D.; Yang, X.; Hau, V.S.; Kaminoh, Y.; Harmon, J.; Pearson, E.; et al. Promoter polymorphism of the erythropoietin gene in severe diabetic eye and kidney complications. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 6998–7003. [\[CrossRef\]](#)
60. Wang, W.; Koka, V.; Lan, H.Y. Transforming growth factor- β and Smad signalling in kidney diseases. *Nephrology* **2005**, *10*, 48–56. [\[CrossRef\]](#)
61. Santos, E.J.F.; Dias, R.S.C.; Brito Lima, J.F.d.; Salgado Filho, N.; Santos, A.M.d. Erythropoietin resistance in patients with chronic kidney disease: Current perspectives. *Int. J. Nephrol. Renov. Dis.* **2020**, *13*, 231–237. [\[CrossRef\]](#)
62. Semenza, G.L.; Neifelt, M.K.; Chi, S.M.; Antonarakis, S.E. Hypoxia-inducible nuclear factors bind to an enhancer element located 3' to the human erythropoietin gene. *Proc. Natl. Acad. Sci. USA* **1991**, *88*, 5680–5684. [\[CrossRef\]](#)
63. Schito, L.; Semenza, G.L. Hypoxia-Inducible Factors: Master Regulators of Cancer Progression. *Trends Cancer* **2016**, *2*, 758–770. [\[CrossRef\]](#)
64. Semenza, G.L. Hypoxia-inducible factors: Mediators of cancer progression and targets for cancer therapy. *Trends Pharmacol. Sci.* **2012**, *33*, 207–214. [\[CrossRef\]](#)
65. Ivan, M.; Kondo, K.; Yang, H.; Kim, W.; Valiando, J.; Ohh, M.; Salic, A.; Asara, J.M.; Lane, W.S.; Kaelin, W.G., Jr. HIF α Targeted for VHL-Mediated Destruction by Proline Hydroxylation: Implications for O₂ Sensing. *Science* **2001**, *292*, 464–468. [\[CrossRef\]](#)
66. Jaakkola, P.; Mole, D.R.; Tian, Y.M.; Wilson, M.I.; Gielbert, J.; Gaskell, S.J.; von Kriegsheim, A.; Hebestreit, H.F.; Mukherji, M.; Schofield, C.J.; et al. Targeting of HIF- α to the von Hippel-Lindau ubiquitylation complex by O₂-regulated prolyl hydroxylation. *Science* **2001**, *292*, 468–472. [\[CrossRef\]](#)
67. Lieb, M.E.; Menzies, K.; Moschella, M.C.; Ni, R.; Taubman, M.B. Mammalian EGLN genes have distinct patterns of mRNA expression and regulation. *Biochem. Cell Biol.* **2002**, *80*, 421–426. [\[CrossRef\]](#)
68. Percy, M.J.; Zhao, Q.; Flores, A.; Harrison, C.; Lappin, T.R.J.; Maxwell, P.H.; McMullin, M.F.; Lee, F.S. A family with erythrocytosis establishes a role for prolyl hydroxylase domain protein 2 in oxygen homeostasis. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 654–659. [\[CrossRef\]](#)
69. Holdstock, L.; Meadowcroft, A.M.; Maier, R.; Johnson, B.M.; Jones, D.; Rastogi, A.; Zeig, S.; Lepore, J.J.; Cobitz, A.R. Four-Week Studies of Oral Hypoxia-Inducible Factor–Prolyl Hydroxylase Inhibitor GSK1278863 for Treatment of Anemia. *J. Am. Soc. Nephrol.* **2016**, *27*, 1234–1244. [\[CrossRef\]](#) [\[PubMed\]](#)
70. Hill, P.; Shukla, D.; Tran, M.G.B.; Aragones, J.; Cook, H.T.; Carmeliet, P.; Maxwell, P.H. Inhibition of Hypoxia Inducible Factor Hydroxylases Protects Against Renal Ischemia-Reperfusion Injury. *J. Am. Soc. Nephrol.* **2008**, *19*, 39–46. [\[CrossRef\]](#) [\[PubMed\]](#)
71. Appelhoff, R.J.; Tian, Y.-M.; Raval, R.R.; Turley, H.; Harris, A.L.; Pugh, C.W.; Ratcliffe, P.J.; Gleadle, J.M. Differential Function of the Prolyl Hydroxylases PHD1, PHD2, and PHD3 in the Regulation of Hypoxia-inducible Factor. *J. Biol. Chem.* **2004**, *279*, 38458–38465. [\[CrossRef\]](#) [\[PubMed\]](#)

72. Moslehi, J.; Minamishima, Y.A.; Shi, J.; Neuberg, D.; Charytan, D.M.; Padera, R.F.; Signoretti, S.; Liao, R.; Kaelin, W.G. Loss of Hypoxia-Inducible Factor Prolyl Hydroxylase Activity in Cardiomyocytes Phenocopies Ischemic Cardiomyopathy. *Circulation* **2010**, *122*, 1004–1016. [[CrossRef](#)]
73. Walmsley, S.R.; Print, C.; Farahi, N.; Peyssonnaud, C.; Johnson, R.S.; Cramer, T.; Sobolewski, A.; Condliffe, A.M.; Cowburn, A.S.; Johnson, N.; et al. Hypoxia-induced neutrophil survival is mediated by HIF-1 α -dependent NF- κ B activity. *J. Exp. Med.* **2005**, *201*, 105–115. [[CrossRef](#)] [[PubMed](#)]
74. Dang, E.V.; Barbi, J.; Yang, H.-Y.; Jinasena, D.; Yu, H.; Zheng, Y.; Bordman, Z.; Fu, J.; Kim, Y.; Yen, H.-R.; et al. Control of TH17/Treg Balance by Hypoxia-Inducible Factor 1. *Cell* **2011**, *146*, 772–784. [[CrossRef](#)] [[PubMed](#)]
75. Sanghani, N.S.; Haase, V.H. Hypoxia-Inducible Factor Activators in Renal Anemia: Current Clinical Experience. *Adv. Chronic Kidney Dis.* **2019**, *26*, 253–266. [[CrossRef](#)] [[PubMed](#)]
76. Ariazi, J.L.; Duffy, K.J.; Adams, D.F.; Fitch, D.M.; Luo, L.; Pappalardi, M.; Biju, M.; DiFilippo, E.H.; Shaw, T.; Wiggall, K.; et al. Discovery and Preclinical Characterization of GSK1278863 (Daprodustat), a Small Molecule Hypoxia Inducible Factor–Prolyl Hydroxylase Inhibitor for Anemia. *J. Pharmacol. Exp. Ther.* **2017**, *363*, 336–347. [[CrossRef](#)]
77. Besarab, A.; Provenzano, R.; Hertel, J.; Zabaneh, R.; Klaus, S.J.; Lee, T.; Leong, R.; Hemmerich, S.; Yu, K.-H.P.; Neff, T.B. Randomized placebo-controlled dose-ranging and pharmacodynamics study of roxadustat (FG-4592) to treat anemia in nondialysis-dependent chronic kidney disease (NDD-CKD) patients. *Nephrol. Dial. Transplant.* **2015**, *30*, 1665–1673. [[CrossRef](#)]
78. Chen, N.; Hao, C.; Liu, B.-C.; Lin, H.; Wang, C.; Xing, C.; Liang, X.; Jiang, G.; Liu, Z.; Li, X.; et al. Roxadustat Treatment for Anemia in Patients Undergoing Long-Term Dialysis. *N. Engl. J. Med.* **2019**, *381*, 1011–1022. [[CrossRef](#)]
79. Chen, N.; Hao, C.; Peng, X.; Lin, H.; Yin, A.; Hao, L.; Tao, Y.; Liang, X.; Liu, Z.; Xing, C.; et al. Roxadustat for Anemia in Patients with Kidney Disease Not Receiving Dialysis. *N. Engl. J. Med.* **2019**, *381*, 1001–1010. [[CrossRef](#)] [[PubMed](#)]
80. Flamme, I.; Ellinghaus, P.; Urrego, D.; Krüger, T. FGF23 expression in rodents is directly induced via erythropoietin after inhibition of hypoxia inducible factor proline hydroxylase. *PLoS ONE* **2017**, *12*, e0186979. [[CrossRef](#)] [[PubMed](#)]
81. Schödel, J.; Grampp, S.; Maher, E.R.; Moch, H.; Ratcliffe, P.J.; Russo, P.; Mole, D.R. Hypoxia, Hypoxia-inducible Transcription Factors, and Renal Cancer. *Eur. Urol.* **2016**, *69*, 646–657. [[CrossRef](#)]
82. Keith, B.; Johnson, R.S.; Simon, M.C. HIF1 α and HIF2 α : Sibling rivalry in hypoxic tumour growth and progression. *Nat. Rev. Cancer* **2012**, *12*, 9–22. [[CrossRef](#)]
83. Sugahara, M.; Tanaka, S.; Tanaka, T.; Saito, H.; Ishimoto, Y.; Wakashima, T.; Ueda, M.; Fukui, K.; Shimizu, A.; Inagi, R.; et al. Prolyl Hydroxylase Domain Inhibitor Protects against Metabolic Disorders and Associated Kidney Disease in Obese Type 2 Diabetic Mice. *J. Am. Soc. Nephrol.* **2020**, *31*, 560–577. [[CrossRef](#)] [[PubMed](#)]
84. Nakamura, H.; Kurihara, S.; Anayama, M.; Makino, Y.; Nagasawa, M. Four Cases of Serum Copper Excess in Patients with Renal Anemia Receiving a Hypoxia-Inducible Factor-Prolyl Hydroxylase Inhibitor: A Possible Safety Concern. *Case Rep. Nephrol. Dial.* **2022**, *12*, 124–131. [[CrossRef](#)] [[PubMed](#)]
85. Chen, N.; Qian, J.; Chen, J.; Yu, X.; Mei, C.; Hao, C.; Jiang, G.; Lin, H.; Zhang, X.; Zuo, L.; et al. Phase 2 studies of oral hypoxia-inducible factor prolyl hydroxylase inhibitor FG-4592 for treatment of anemia in China. *Nephrol. Dial. Transplant.* **2017**, *32*, 1373–1386. [[CrossRef](#)] [[PubMed](#)]
86. Duvroq (Daprodustat): Japanese Prescribing Information. Available online: https://www.pmda.go.jp/PmdaSearch/iyakuDetail/ResultDataSetPDF/340278_39990D4F1024_1_01 (accessed on 14 August 2020).
87. Chertow, G.M.; Pergola, P.E.; Farag, Y.M.K.; Agarwal, R.; Arnold, S.; Bako, G.; Block, G.A.; Burke, S.; Castillo, F.P.; Jardine, A.G.; et al. Vadadustat in Patients with Anemia and Non-Dialysis-Dependent CKD. *N. Engl. J. Med.* **2021**, *384*, 1589–1600. [[CrossRef](#)] [[PubMed](#)]
88. Xiong, L.; Zhang, H.; Guo, Y.; Song, Y.; Tao, Y. Efficacy and Safety of Vadadustat for Anemia in Patients With Chronic Kidney Disease: A Systematic Review and Meta-Analysis. *Front. Pharmacol.* **2022**, *12*, 795214. [[CrossRef](#)]
89. Sugahara, M.; Tanaka, T.; Nangaku, M. Future perspectives of anemia management in chronic kidney disease using hypoxia-inducible factor-prolyl hydroxylase inhibitors. *Pharmacol. Ther.* **2022**, *239*, 108272. [[CrossRef](#)] [[PubMed](#)]
90. Locatelli, F.; Minutolo, R.; De Nicola, L.; Del Vecchio, L. Evolving Strategies in the Treatment of Anaemia in Chronic Kidney Disease: The HIF-Prolyl Hydroxylase Inhibitors. *Drugs* **2022**, *82*, 1565–1589. [[CrossRef](#)]
91. Akizawa, T.; Nangaku, M.; Yamaguchi, T.; Koretomo, R.; Maeda, K.; Miyazawa, Y.; Hirakata, H. A Phase 3 Study of Enarodustat in Anemic Patients with CKD not Requiring Dialysis: The SYMPHONY ND Study. *Kidney Int. Rep.* **2021**, *6*, 1840–1849. [[CrossRef](#)]
92. Akizawa, T.; Yamada, T.; Nobori, K.; Matsuda, Y.; Hayashi, Y.; Hayasaki, T.; Yamamoto, H. Molidustat for Japanese Patients With Renal Anemia Receiving Dialysis. *Kidney Int. Rep.* **2021**, *6*, 2604–2616. [[CrossRef](#)]
93. Joksimovic Jovic, J.; Antic, S.; Nikolic, T.; Andric, K.; Petrovic, D.; Bolevich, S.; Jakovljevic, V. Erythropoietin Resistance Development in Hemodialysis Patients: The Role of Oxidative Stress. *Oxidative Med. Cell. Longev.* **2022**, *2022*, 9598211. [[CrossRef](#)]

-
94. Wang, J.; Liu, Y.; Wang, Y.; Sun, L. The Cross-Link between Ferroptosis and Kidney Diseases. *Oxidative Med. Cell. Longev.* **2021**, *2021*, 6654887. [[CrossRef](#)] [[PubMed](#)]
 95. Mravic, M.; He, L.; Kratochvil, H.; Hu, H.; Nick, S.E.; Bai, W.; Edwards, A.; Jo, H.; Wu, Y.; DiMaio, D.; et al. Designed Transmembrane Proteins Inhibit the Erythropoietin Receptor in a Custom Binding Topology. *bioRxiv* **2023**. [[CrossRef](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.